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Brian B. Monson *Harvard University*

Zach Eaton-Rosen University College London

Kush Kapur Harvard University

Einat Liebenthal *Harvard University*

Abraham Brownell Harvard University

See next page for additional authors

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Authors

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Differential Rates of Perinatal Maturation of Human Primary and Nonprimary Auditory Cortex

Brian B. Monson,^{1,2} Zach Eaton-Rosen,³ Kush Kapur,⁴ [©]Einat Liebenthal,⁵ Abraham Brownell,¹ Christopher D. Smyser,^{6,7} Cynthia E. Rogers,^{7,8} Terrie E. Inder,¹ Simon K. Warfield,² and Jeffrey J. Neil⁴

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¹Department of Pediatric Newborn Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, MA 02115, ²Department of Radiology, Boston Children's Hospital, Harvard Medical School, Boston, MA 02115, ³Translational Imaging Group, University College London, London, WC1E 7JE United Kingdom, ⁴Department of Neurology, Boston Children's Hospital, Harvard Medical School, Boston, MA 02115, ⁵Department of Psychiatry, Brigham and Women's Hospital, Harvard Medical School, Boston, MA 02115, ⁶Department of Neurology, Washington University School of Medicine, St. Louis, MO 63130, ⁷Department of Pediatrics, Washington University School of Medicine, St. Louis, MO 63130, and ⁸Department of Psychiatry, Washington University School of Medicine, St. Louis, MO 63130

Abstract

Primary and nonprimary cerebral cortex mature along different timescales; however, the differences between the rates of maturation of primary and nonprimary cortex are unclear. Cortical maturation can be measured through changes in tissue microstructure detectable by diffusion magnetic resonance imaging (MRI). In this study, diffusion tensor imaging (DTI) was used to characterize the maturation of Heschl's gyrus (HG), which contains both primary auditory cortex (pAC) and nonprimary auditory cortex (nAC), in 90 preterm infants between 26 and 42 weeks postmenstrual age (PMA). The preterm infants were in different acoustical environments during their hospitalization: 46 in open ward beds and 44 in single rooms. A control group consisted of 15 term-born infants. Diffusion parameters revealed that (1) changes in cortical microstructure that accompany cortical maturation had largely already occurred in pAC by 28 weeks PMA, and (2) rapid changes were taking place in nAC between 26 and 42 weeks PMA. At term equivalent PMA, diffusion parameters for auditory cortex were different between preterm infants and term control infants, reflecting either delayed maturation or injury. No effect of room type was observed. For the preterm group, disturbed maturation of nonprimary (but not primary) auditory cortex was associated with poorer language performance at age two years.

Key words: audition; cortical development; diffusion tensor imaging; neuroimaging; preterm infants

Significance Statement

Different brain regions mature at different rates, particularly early in development. Knowledge of when specific sensory brain regions are maturing is critical for understanding the susceptibility to external sensory influences (e.g., during critical periods) and potential for vulnerability to injury. Here, we demonstrate in vivo that, during the perinatal period, human primary auditory cortex (pAC) matures earlier than nonprimary auditory cortex (nAC), consistent with accounts of brain development from histology. However, we detect more rapid changes in nAC during this period. Our findings indicate that disruption of nonprimary cortex (but not primary cortex) maturation during this developmental period is associated with poorer childhood language development. Differential developmental timelines may render nonprimary sensory cortex more vulnerable than primary sensory cortex.

Introduction

Humans are altricial mammals with precocial hearing. An abundance of neurobiological evidence clearly dem-

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onstrates that the human auditory system comes online at least as early as 25 weeks postmenstrual age (PMA; Graziani et al., 1968; Starr et al., 1977; Birnholz and

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Benacerraf, 1983; Rotteveel et al., 1987; Hepper and Shahidullah, 1994), some 15 weeks before term birth. By this age, the structural development of the nervous system is sufficient for peripheral auditory input to reach auditory cortex (Weitzman and Graziani, 1968; Rotteveel et al., 1987; Jardri et al., 2008; Mahmoudzadeh et al., 2013). Furthermore, cortical memory traces are forming long before term birth for auditory input arising from acoustic stimuli in the extrauterine environment, including speech and language (DeCasper and Fifer, 1980; Decasper and Spence, 1986; Moon et al., 1993; Mahmoudzadeh et al., 2013; Moon et al., 2013; Partanen et al., 2013). This makes auditory cortex unique among the sensory cortices and therefore of particular interest when evaluating cortical maturation processes in humans.

During development, neuronal genesis and differentiation occur in primary sensory cortical regions before nonprimary and association regions (Conel, 1939; Sidman and Rakic, 1982), indicating that primary sensory cortex matures in advance of nonprimary cortex. However, the differences between the timelines for primary versus nonprimary cortex maturation are unclear. It may be that nonprimary cortex develops with a rate of maturation identical to, but delayed from, that of primary cortex. On the other hand, nonprimary cortex might follow an altogether different rate of maturation from that of primary cortex. This distinction has implications for the timing and severity of disruption and/or injury to developing cortex and the consequences thereof.

Diffusion tensor imaging (DTI) permits tracking of human cortical maturation *in vivo* through the parameters of fractional anisotropy (FA), mean diffusivity (MD), axial diffusivity (AD), and radial diffusivity (RD; McKinstry et al., 2002; Ball et al., 2013; Smyser et al., 2015). FA, which reflects the degree of anisotropy of water molecule displacements in brain tissue, decreases in developing gray matter (GM) as histologic changes disrupt the initial radial organization of the cortical plate (McKinstry et al., 2002). In developing white matter (WM), FA values increase as

B. B. Monson's present address: Department of Speech and Hearing Science, University of Illinois at Urbana-Champaign, 901 South Sixth Street, Champaign, IL 61820

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Correspondence should be addressed to Brian B. Monson at the above address, E-mail: monson@illinois.edu.

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preoligodendroglial ensheathment and myelination inhibit water displacements orthogonal to maturing axons. MD, AD, and RD, which measure the mean, axial, and radial magnitude of water displacements, respectively, decrease in both maturing GM and WM as brain water content decreases and cell density increases. It has been demonstrated that cortical GM tissue matures in synchrony with underlying subplate and WM (Kostović et al., 2014; Smyser et al., 2015), suggesting that diffusion parameters from both GM and adjacent subplate/WM reflect the maturational status of developing cortex.

We used DTI to investigate the maturational timelines of the cortical plate and adjacent subcortical tissue between 26 and 42 weeks PMA in auditory cortex regions in preterm infants. We examined the variation in maturational timeline along the axis of Heschl's gyrus (HG) in the temporal lobe where cortex transitions from primary auditory cortex (pAC, postero-medially) to nonprimary auditory cortex (nAC, antero-laterally; Fig. 1; Morosan et al., 2001; Moerel et al., 2014). We compared the timeline of maturation of pAC versus nAC, hypothesizing that pAC would mature in advance of nAC. We further hypothesized that different acoustic environments during the perinatal period would affect the maturational timeline of auditory cortex. Finally, we hypothesized that disturbed auditory cortex maturation in infancy would be related to poorer language development in childhood. We tested our hypotheses in a cohort of 90 very preterm infants (born <30 weeks' gestation) who underwent diffusion magnetic resonance imaging (MRI) up to four times during their hospital stay and neurodevelopmental follow-up at two years old.

Materials and Methods

Subjects

A total of 136 very preterm infants born before 30 weeks' gestation were recruited from the St. Louis Children's Hospital Neonatal Intensive Care Unit (NICU) between 2007 and 2010. Infants with moderate to severe (Kidokoro et al., 2013) WM injury or severe brain abnormalities were excluded from the analysis (Fig. 2). Infants with any conductive or sensorineural hearing loss (assessed after discharge for infants who failed newborn hearing screening) were likewise excluded. Infants who failed newborn hearing screening but underwent no audiological follow-up were also excluded. Infants underwent MRI one to four times during hospital stay based on their clinical stability to travel to the MRI scanner. Of those meeting selection criteria, 90 subjects had usable MRI data collected at some point during hospital stay, with 56 subjects imaged at multiple timepoints, for a total of 173 images between 26 and 42 weeks PMA. Of the 90 subjects, 57 had data collected at term-equivalent age (37-42 weeks PMA). Infants in the NICU environment were pseudorandomly assigned to either a noisier open bay multi-bed unit (N = 46) or a quieter single patient room (N = 44) based on staffing and bed availability (according to standard clinical practice), but otherwise had access to the same medical care and physicians, as described elsewhere (Pineda et al., 2014). Fifteen healthy term-born control infants were re-

B.B.M. and J.J.N. wrote the paper.

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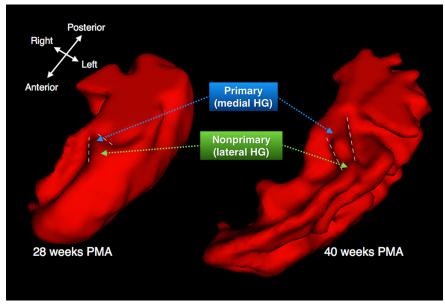


Figure 1. Development of the left hemisphere temporal plane from 28 to 40 weeks PMA. HG (demarcated with white dashed lines) is forming by 28 weeks PMA and has an adult-like appearance by 40 weeks PMA. pAC and nAC are located in medial and lateral HG, respectively.

cruited from the Barnes-Jewish Hospital Newborn Nursery and scanned within the first 4 d of life. Term infants had no history of illicit substance exposure *in utero* and no evidence of acidosis in the first hour of life. No infants had chromosomal abnormalities or congenital infections. Informed written parental consent was obtained for each subject. The study was approved by the Washington University Human Studies Committee.

Participants in the preterm group returned for follow-up assessment at two years old and were assessed with the Bayley Scales of Infant and Toddler Development, 3rd edition (Bayley and Reuner, 2006). Outcomes used for this study were scores from the receptive communication subtest (assessing preverbal behaviors, verbal comprehension and vocabulary), expressive communication subtest (assessing preverbal babbling and gesturing, as well as vocabulary and utterances), and cognitive subtest (assessing sensorimotor development, memory, object relatedness, and concept formation). Of those with useable MRI data at term-equivalent age (N = 57), 46 had behavioral data collected at two years old (for characteristics of these infants, see Table 1). The 11 infants who were not assessed at two years old had a higher average birth weight (1158 vs 935 g, p = 0.03) and a lower average maternal age (24 vs 29 years, p = 0.04) than the other 46 infants, but otherwise did not have significantly different characteristics.

MRI data acquisition and processing

Infants were imaged during natural sleep or quiescence without the use of sedation. Infants wore neonatal earmuffs (Natus Medical) for hearing protection. Heart rate and arterial oxygen saturation were monitored continuously throughout data acquisition. Diffusion MRI data were acquired with a single-shot echo-planar sequence (repetition time/echo time 13,300/112 ms, 1266 Hz/Px bandwidth, 128-mm field of view, voxel size $1.2 \times 1.2 \times$ 1.2 mm³, 48 *b*-directions with multiple amplitudes ranging from 0 to 1200s/mm²) using a 3-T Siemens TIM Trio system with an infant-specific quadrature head coil (Advanced Imaging Research). Other MRI data collected included: rapid gradient echo T1-weighted images (repetition time/echo time 1500/3 ms, voxel size 1 \times 0.7 \times 1 mm³) and fast spin echo T2-weighted images (repetition time/echo time 8500/160 ms, voxel size 1 \times 1 \times 1 mm³). Total data acquisition time was ~60 min.

MD, AD, RD, and FA values were estimated using a weighted linear least square approach, implemented in FSL v5.0.2 (Jenkinson et al., 2012). The FA noise floor was \sim 0.06. (To determine FA noise floor, we randomly selected three infants and sampled a region of cerebrospinal fluid, which is known to have low FA. Each sample consisted of \geq 60 contiguous voxels. The average FA for these samples was 0.062, 0.058, and 0.044.)

Regions of interest (ROIs) were placed manually by an expert rater in native space using T2-weighted (b = 0s/mm²) images and MD parametric maps to identify the cortical GM and adjacent WM of Heschl's gyrus (HG) in the left hemisphere. HG was defined using a previously established method based on anatomic landmarks (Penhune et al., 1996). In cases of HG duplications, only the anterior gyrus was used for analysis. The boundary between cortical GM and WM was determined by using an intensity threshold value based on histograms generated from initial manual selection of low-intensity voxels reliably identified as HG cortical tissue and high-intensity voxels reliably identified as WM. To minimize partial volume effects, any voxels whose intensity values were ambiguous (i.e., were between threshold values for GM and WM), were excluded from analysis. ROIs were subdivided into single-voxel-thick oblique "slices" in equidistant steps along the length of HG. Average FA, MD, AD, and RD were obtained for each tissue type in each ROI slice.

VERY PRETERM COHORT

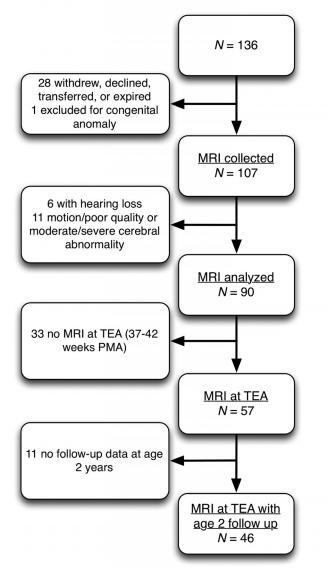


Figure 2. Flowchart detailing retention and follow-up of the very preterm cohort. TEA, term-equivalent age.

On average, the number of slices required to define the length of HG was 13 slices for the earliest ages (26–28 weeks PMA) and 29 slices at term-equivalent age (37–42 weeks PMA).

For region comparison (primary versus nonprimary), each diffusion parameter was averaged over the first three slices for pAC, beginning with the most postero-medial slice and moving laterally. The use of three slices ensured that tissues were well within the putative boundary between pAC (areas Te1.0/1.1) and nAC (area Te1.2), believed to be one-half to two-thirds the length of HG (Rademacher et al., 1993; Penhune et al., 1996; Morosan et al., 2001; Glasser and Van Essen, 2011; Clarke and Morosan, 2012; Marie et al., 2015). For nAC, each parameter was averaged over the most lateral three slices. The intra-rater reliability rating for pAC FA was 0.83, while reliability for all other regions and tissues was >0.88. For group comparisons at term-equivalent age, ROIs were also generated for whole-brain cortical GM and cerebral WM using a preterm-specific automated segmentation algorithm (Cardoso et al., 2013), using tissue class priors from a T2-weighted longitudinal atlas (Kuklisova-Murgasova et al., 2011).

For data visualization in Figures 3, 5, slices were interpolated to 100 points along the length of HG for each dataset. Datasets were then binned into groups based on PMA in weeks [\leq 28 (N = 7), 29–30 (N = 25), 31–32 (N = 20), 33–34 (N = 44), 35–36 (N = 20), 37–38 (N = 42), and 39–42 (N = 15)] and averaged.

Experimental design and statistical analysis

Changes in diffusion parameters over time were modeled as 3-level linear mixed-effects models which allowed for nesting of repeated observations within regions and regions within subjects. The model for the mean response included the main effect of region indicator variable, and linear and quadratic trends of age along with their interactions with the region indicator variable as the fixed effects. In addition, it included random effects of intercept and slope to account for correlations within regions and within time points nested within regions. The final model selection was performed using Akaike Information criterion (AIC). Likelihood ratio test (LRT) for the nested models was also used to test the significance of variance covariance parameters of the ran-

-			
	Open bay ($n = 22$) Mean (SD)	Private room ($n = 24$)	p value
Birth GA (weeks)	26.3 (1.7)	27 (2.1)	0.20
Birthweight (grams)	883 (242)	983 (250)	0.17
Length of stay (days)	92 (18)	91 (25) ´	0.81
PMA at scan (weeks)	37.8 (1.2)	38.3 (1.5)	0.16
Maternal age (years)	29.1 (8.5)	29.8 (7.3)	0.79
Social risk score (out of 5)	1.45 (1.34)	1.30 (1.26)	0.70
	n (%)		
Male	10 (45)	10 (42)	0.80
SGA	1 (5)	1 (4)	0.95
Infection	4 (18)	2 (9)	0.38
BPD	12 (55)	15 (68)	0.35

GA, gestational age; SGA, small for gestational age; BPD, bronchopulmonary dysplasia.

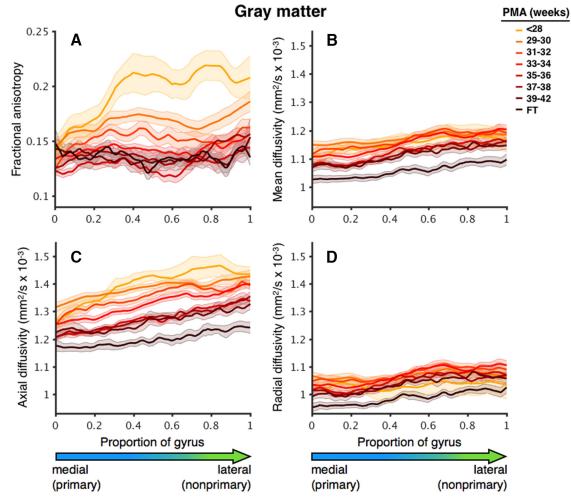


Figure 3. Changes in GM diffusion parameters with increasing age along the length of HG from medial (primary) to lateral (nonprimary). Changes are shown for (A) FA, (B) MD, (C) AD, and (D) RD. Curves are separated by age group (in weeks), indicated by color. Shading indicates ± 1 SE. FT, full-term controls.

dom effects. A reduced 2-level mixed-effects model was fit in cases where the variance components were not found to significantly explain the correlations within regions and/or within time points within regions.

Group (preterm versus full term) and region (primary versus nonprimary) differences in FA, MD, AD, and RD for each tissue type were assessed at term-equivalent age (37–42 weeks PMA) using a single, 2-level linear mixed-effects model for each parameter and tissue type. Each model included a group \times region interaction term and controlled for PMA at the time of scan. Interaction terms were removed from the model if found to be not significant. To determine whether observed differences were specific to auditory cortex, a second model for each analysis controlled for the corresponding whole-brain diffusion parameter values.

Associations between language outcomes (receptive language and expressive language) at two years old and diffusion parameters at term-equivalent age were tested for the preterm group using separate linear regression models for each parameter (FA, MD, AD, and RD), tissue type (GM and WM), and region (primary and nonprimary). Each model controlled for room type since room type was known to affect language outcome in these patients (Pineda et al., 2014). Models in a secondary analysis also controlled for birth destational age and a social risk score based on a 5-point scale, calculated as the sum of binary values indicating the presence or absence of (1) maternal education level lower than high school diploma, (2) African-American race, (3) public insurance, (4) maternal age <19 years, and (5) single-parent household. For each outcome and tissue type, significance values were adjusted for multiple comparisons using Bonferroni correction. As additional controls, we also tested for associations (1) between whole-brain diffusion parameters at term-equivalent age and language outcomes at two years old, and (2) between auditory cortex parameters at term-equivalent age and cognitive composite scores at two years old. Analyses for term-equivalent age data were performed using R (R Core Team, 2016). All other analyses were performed using SAS (SAS Institute Inc.).

Results

Cortical gray matter

At 28 weeks PMA, FA was lower in pAC and higher in nAC, as can be seen in the plot corresponding to <28 weeks

	0		0	0	0	•	0		
		Age ²		Age		Region		Age $ imes$ region	
		Coefficient	p value	Coefficient	p value	Coefficient	p value	Coefficient	p value
GM	FA	0.594	<0.001	-13.163	< 0.001	-20.076	0.013	0.926	0.304
	MD	_	_	-4.603	0.001	-37.863	0.028	-3.533	0.068
	AD	_		-12.055	< 0.001	-94.541	<0.001	-1.729	0.417
	RD	_		-1.333	0.388	-16.440	0.379	-4.457	0.035
WM	FA	_		3.096	< 0.001	24.853	<0.001	-1.481	0.036
	MD	-0.976	0.006	-10.189	0.081	-189.456	<0.001	7.976	0.001
	AD	-0.781	0.044	-11.068	0.092	-164.175	<0.001	5.683	0.033
	RD	-1.060	0.003	-9.959	0.088	-202.240	<0.001	9.130	<0.001

Table 2. Regression coefficients showing	ng effects of age and regi	ion on diffusion parameter changes

Bold values indicate significant effects of region and age \times region interactions. Coefficient values are multiplied by 10³.

PMA in Figure 3A. Note that the FA values are higher in the lateral (nonprimary) portion of the gyrus. FA decreased with increasing PMA with a quadratic trend (Table 2; Fig. 4A). FA decreased more sharply for nAC than for pAC. As a result, FA values for nAC, while significantly higher than pAC values at 30 (p = 0.001) and 35 weeks PMA (p < 0.001), reached similar values by 40 weeks PMA (p = 0.24). An effect of

cortical region (primary versus nonprimary) was observed, with lower FA in pAC and no significant interaction between region and age.

Both MD and AD decreased with increasing PMA, with effects of region (lower in pAC) and no interactions between region and age (Figs. 3B,C, 4B,C; Table 2). There was a region \times age interaction for the decreases in GM RD, with pAC

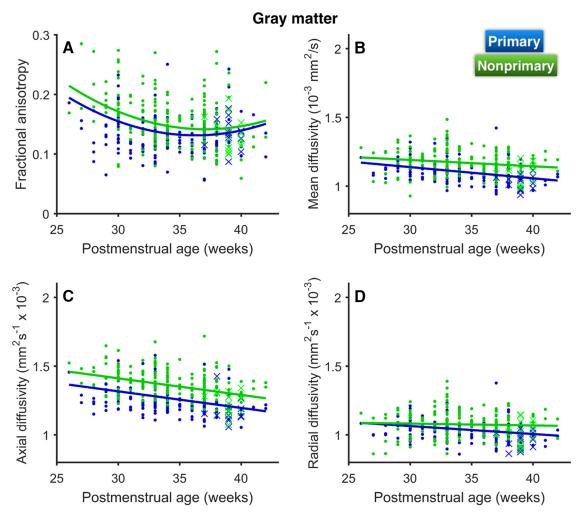


Figure 4. Changes in GM diffusion parameters as a function of PMA for primary (blue) and nonprimary (green) auditory cortex in HG. Changes are shown for (A) FA, (B) MD, (C) AD, and (D) RD. Curves represent results from linear mixed-effects models. Values for full-term controls are denoted by crosses (\times).

Table 3. Regression coefficients showing associations between diffusion parameter values at term-equivalent age and language ability at two years old, controlling for room type

		Receptive	ve		Expressive		
		Coefficient	Corrected p value	R^2	Coefficient	Corrected p value	R^2
pGM	FA	-7.08	>0.99	0.04	10.83	>0.99	0.09
	MD	9.43	>0.99	0.07	9.07	>0.99	0.10
	AD	6.26	>0.99	0.05	9.91	>0.99	0.12
	RD	9.10	>0.99	0.07	6.51	>0.99	0.09
nGM	FA	-10.46	>0.99	0.06	-29.39	0.04	0.23
	MD	12.96	0.10	0.16	10.41	0.68	0.14
	AD	8.69	0.33	0.12	1.63	>0.99	0.08
	RD	11.74	0.14	0.15	13.12	0.17	0.18
pWM	FA	14.06	>0.99	0.06	-2.71	>0.99	0.08
	MD	2.97	>0.99	0.04	9.63	>0.99	0.12
	AD	5.19	>0.99	0.06	7.96	>0.99	0.12
	RD	0.44	>0.99	0.03	7.87	>0.99	0.11
nWM	FA	-12.37	>0.99	0.05	-14.84	>0.99	0.10
	MD	0.26	>0.99	0.03	3.86	>0.99	0.09
	AD	-0.30	>0.99	0.03	2.27	>0.99	0.08
	RD	0.57	>0.99	0.03	4.38	>0.99	0.10

Bold values indicate a significant association. Coefficient values for diffusivity measures are multiplied by 10³. p/nGM, primary/nonprimary GM.

RD declining significantly with age (p < 0.001) and nAC RD showing no linear trend (p = 0.39; Figs. 3D, 4D).

The group comparison at term-equivalent age revealed that preterm infants had higher values for GM MD (p < 0.001), AD (p < 0.001), and RD (p = 0.004) than healthy full-term infants (Fig. 3). No effect of prematurity was observed in FA (p = 0.69). An effect of cortical region was observed for FA (p = 0.04), MD (p < 0.001), AD (p < 0.001), and RD (p < 0.001), and RD (p < 0.001), with nAC values higher than pAC values. No interactions between group and cortical region were observed for GM diffusion measures. The group difference in AD remained significant when controlling for whole-brain GM values (p = 0.04), while group differences in all other parameters were diminished. For the preterm group, we found no effect of room

environment (open bay versus single patient room) on any

GM diffusion measures. For the preterm group, higher FA in nAC at term-equivalent age was associated with poorer expressive language performance at two years old (Tables 3, 4). pAC and whole-brain cortical GM FA showed no such association with expressive language performance (corrected p > 0.9). No associations were observed between GM diffusion parameters and receptive language. (Two associations observed between nAC diffusion parameters and receptive language did not remain significant after correction for multiple comparisons.) No associations were observed between auditory GM diffusion parameters and cognitive composite score.

White matter

FA for HG WM increased with increasing PMA for the entire length of HG (Table 2; Figs. 5A, 6A). While an effect

Table 4. Regression coefficients showing associations between diffusion parameter values at term-equivalent age a	nd
language ability at two years old, controlling for room type, social risk, and birth gestational age	

		Receptive			Expressive		
		Coefficient	Corrected p value	R^2	Coefficient	Corrected p value	R^2
pGM	FA	-0.83	>0.99	0.08	8.66	>0.99	0.09
	MD	9.11	>0.99	0.12	9.07	>0.99	0.11
	AD	7.41	>0.99	0.12	9.55	>0.99	0.12
	RD	8.08	>0.99	0.12	6.85	>0.99	0.10
nGM	FA	-9.10	>0.99	0.10	-32.59	0.03	0.26
	MD	12.11	0.22	0.19	12.85	0.37	0.17
	AD	8.05	0.55	0.16	2.62	>0.99	0.09
	RD	11.15	0.30	0.18	16.23	0.07	0.23
pWM	FA	15.74	>0.99	0.12	-3.71	>0.99	0.08
	MD	1.78	>0.99	0.09	9.53	>0.99	0.13
	AD	4.39	>0.99	0.10	7.80	>0.99	0.12
	RD	-0.50	>0.99	0.08	7.98	>0.99	0.11
nWM	FA	-8.82	>0.99	0.09	-19.28	>0.99	0.11
	MD	-0.06	>0.99	0.08	5.11	>0.99	0.11
	AD	-0.47	>0.99	0.09	3.10	>0.99	0.09
	RD	0.20	>0.99	0.08	5.68	>0.99	0.12

Bold values indicate a significant association. Coefficient values for diffusivity measures are multiplied by 10³. p/nGM, primary/nonprimary GM.

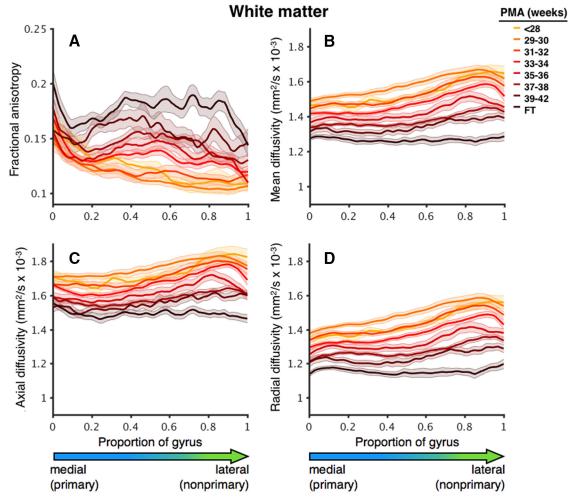


Figure 5. Changes in WM diffusion parameters with increasing age along the length of HG from medial (primary) to lateral (nonprimary). Changes are shown for (A) FA, (B) MD, (C) AD, and (D) RD. Curves are separated by age group (in weeks), indicated by color. Shading indicates ± 1 SE. FT, full-term controls.

of region was observed, with primary WM (pWM) FA values higher than nonprimary WM (nWM) values, more rapid changes were apparent once again in the lateral two-thirds of HG, confirmed by a significant region \times age interaction (Table 2). FA values for the middle portion of HG (from 35% to 90% of full HG length), while initially lower than values for the medial region, ultimately reached values similar to that of the most medial point at term-equivalent age (Fig. 5*A*).

WM MD, AD, and RD exhibited similar behavior to one another (Fig. 5*B–D*). Each decreased with age with a significant quadratic trend and an effect of region, with lower values for PWM (Fig. 6*B–D*; Table 2). Each showed a more rapid decline in nWM than in pWM, confirmed by significant region \times age interactions (Table 2). Each also showed more dramatic changes than those observed for their GM counterparts.

At term-equivalent age, preterm infants had higher values than healthy full-term infants for WM MD (p = 0.004) and RD (p = 0.001), whereas higher values for AD (p = 0.13) and lower values for FA (p = 0.07) did not reach significance (Fig. 5). However, significant interactions between group and cortical region were observed for FA

(p = 0.02), MD (p < 0.001), AD (p = 0.005), and RD (p < 0.001). In each case, posthoc comparisons showed that the deviation of preterm infants from full-term infants was larger and highly significant in nWM (p < 0.001 for all parameters). This analysis also revealed that, for the preterm group, (1) nWM MD, AD, and RD values were significantly higher than those for pWM (p < 0.001 for all parameters), and (2) nWM FA was significantly lower than pWM FA (p = 0.02). Main effects of group were diminished when controlling for whole-brain values, but all interactions remained significant. We found no effect of room environment on any WM diffusion parameters and language outcomes at two years old for the preterm group (Tables 3, 4).

Discussion

Cortical gray matter

The asymptotic decline of FA in maturing GM during the perinatal period has been reported previously (McKinstry et al., 2002; Ball et al., 2013; Smyser et al., 2015). Our results indicate that pAC FA reaches a mature value

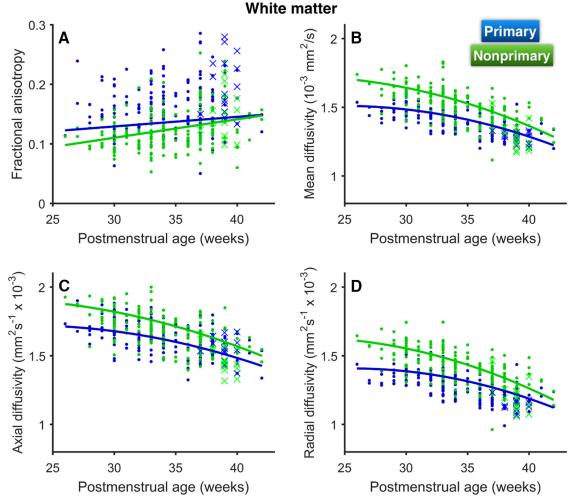


Figure 6. Changes in WM diffusion parameters as a function of PMA for primary (blue) and nonprimary (green) auditory cortex in HG. Changes are shown for (A) FA, (B) MD, (C) AD, and (D) RD. Curves represent results from linear mixed-effects models. Values for full-term controls are denoted by crosses (\times).

earlier than that observed for the rest of the temporal lobe, prefrontal areas, and other cortical regions (Ball et al., 2013; Smyser et al., 2015), indicating that pAC matures relatively early in development. This observation is in accordance with non-human primate diffusion data (Kroenke et al., 2007).

The neuroanatomical bases for changes to FA and diffusivity measures remain a matter of investigation, but it is likely that decreasing GM FA at this stage reflects a variety of processes - myelination of intra-cortical WM, outgrowth of basal dendrites from pyramidal cells, regression of radial glia, and maturation of interneurons - all of which disrupt the initial radial organization of the maturing cortical plate (McKinstry et al., 2002). The sharper decline in AD and shallower decline in RD observed for GM is consistent with this interpretation, as most of the processes listed above, with the possible exception of regression of radial glia, would hinder water displacements in a radial orientation and thereby reduce AD. Our data suggest that the microstructural maturational processes leading to the decline of FA of the developing cortical plate are established to a considerable degree in pAC by 28 weeks PMA. As a result, the decline in FA we detect in pAC after 28 weeks PMA is relatively modest. In contrast, these processes in nAC are in early stages at 28 weeks PMA, and thus we detect a larger decline in FA.

White matter

The perinatal period before term is a time of transition for tissue adjacent to the cortical plate. During this time, the transient subplate zone, the largest compartment of the human neocortical wall, involutes and is gradually replaced by developing WM. In humans, the width of the subplate zone is at its maximum around 22-24 weeks PMA, but shows region dependence: the maximum width is \sim 1 mm in primary visual cortex and can be nearly 5 mm in somatosensory cortex (Kostović and Rakic, 1990). After 24 weeks PMA, the subplate zone width gradually decreases, initially showing marked thinning adjacent to the depths of the sulci, with a thicker band of subplate neurons observed adjacent to the crowns of the gyri (Kostović et al., 2014). We know of no study that has examined the width of the subplate in pAC in humans, but, given our spatial resolution (1.2 mm), it is likely that our WM analysis

captures the changes in diffusion parameters associated with the transition of adjacent tissue from the subplate zone at the earliest time points (26–27 weeks PMA) to maturing WM at the later time points.

We observed region-dependent differences in WM diffusion parameters, with lower FA and higher MD, AD, and RD for nWM. These findings all point to less mature tissue in nWM. At the same time, we also observed that the rates of increase in FA and decrease in MD, AD, and RD were greater for nWM, suggesting that maturing nWM is changing more rapidly than pWM during the perinatal period. We questioned whether the more rapid changes observed in nWM might be a consequence of preterm birth and postnatal events. However, lower FA and higher MD/ AD/RD values for nWM (relative to pWM) were observed at our earliest time points (<28 weeks), very shortly after birth for most of these infants. Since developmental changes in diffusion parameters occur on a relatively slow timescale, it is likely that these early diffusion values reflect those that would be observed in utero.

Finally, there has been some dispute regarding the topographical boundaries of pAC and nAC in HG, including the definition of a medial/lateral border (Da Costa et al., 2011; Clarke and Morosan, 2012; Moerel et al., 2014). While we are unable to resolve this issue, our data show a clear but gradual transition from pAC to nAC along the axis of HG during development. Our results suggest that the lateral two-thirds of HG follows a maturational process distinct from that of the medial third, whereas the putative boundary between pAC and nAC is believed to be at approximately the medial one-half to two-thirds of HG (Rademacher et al., 1993; Penhune et al., 1996; Clarke and Morosan, 2012; Marie et al., 2015).

Vulnerability

Maturation of both auditory GM and WM was disrupted by premature birth. The directions of preterm-birth related differences in GM MD/AD/RD and WM FA/MD/AD/RD are all consistent with a delay in maturation and/or injury. Whereas GM appeared equally affected across HG, nWM showed significantly larger deviations from healthy values than did pWM, and we speculate that the primary region is more resistant to disruption from preterm birth. This resistance may be due to the fact that pAC is more mature at the time of preterm birth, but could also be related to the rapidity of tissue changes during the perinatal period. That is, the phenomenon observed here that nWM matures at a more rapid pace (as defined by changes in diffusion measures) than pWM during this period may render nAC/nWM tissue more vulnerable to disruption or injury due to circumstances consequential to preterm birth. Although we observed no regional effects of prematurity in cortical GM, given the importance of the subplate in establishing thalamocortical connections during development (Ghosh et al., 1990; Kanold et al., 2003; Hoerder-Suabedissen and Molnár, 2015), the regional effects we observed in subplate and WM tissue might induce regionspecific changes in GM as cortex continues to mature beyond 42 weeks PMA.

Contrary to our hypothesis, microstructural maturation of auditory cortical GM and WM appeared unaffected by acoustic differences between an open-bay NICU and single patient rooms. The open-bay NICU environment tends to be noisier, whereas infants in single patient rooms tend to experience more and longer periods of silence (Jobe, 2014). One recent study suggests infants in single patient rooms experience up to three more hours of silence in a 16-h period than infants in an open-bay NICU, with daily average sounds levels being 2-4 dB higher in the openbay environment (Pineda et al., 2017). There is some indication that these differences in room environment affect macrostructural development of nonprimary auditory regions (Pineda et al., 2014). Furthermore, animal studies have revealed that sound deprivation (or lack of an enriched auditory environment) can induce deficits in both structure and function of auditory cortex neurons (McMullen and Glaser, 1988; Bose et al., 2010; Mowery et al., 2015). As we detected no microstructural differences associated with the two room types here, it may be that diffusion measures lack sensitivity to these subtler changes in neural structure, or that the acoustic differences between the room types were not sufficiently large to induce such changes. Alternatively, since drastic changes to sensory experience during the perinatal period appear to have a highly selective impact on only some sensory neural development processes, it may be that the differences between acoustic environments produce neural changes undetectable by diffusion MRI. For example, some evidence suggests that altered perinatal sensory experience might induce differences in size and distribution of synapses, whereas synaptogenesis and overall synaptic density is preserved (Bourgeois et al., 1989).

We found that higher FA in nAC at term-equivalent age was associated with poorer expressive communication ability (but not general cognitive functioning) at two years old for preterm infants. Since higher FA can reflect less mature (or injured) GM, this finding suggests that the disruption or delay of maturation of nAC during the perinatal period might have long-lasting consequences. Although we did not observe a strong relationship with receptive communication ability, nAC is intimately involved in the perception of complex sounds including speech and other vocalizations, and therefore important for learning spoken language communication (Davis and Johnsrude, 2003; Liebenthal et al., 2005; Hickok and Poeppel, 2007; Leaver and Rauschecker, 2010; Poeppel, 2014; Norman-Haignere et al., 2015). Additionally, nAC is one of many regions that exhibit functional activity during both speaking and listening (Awad et al., 2007; Stephens et al., 2010; Hagoort, 2014). Thus, it is biologically plausible that a relationship exists between disrupted maturation of nAC and expressive language development. Since human fetuses are learning several aspects of extrauterine vocal communication and language before term (DeCasper and Fifer, 1980; Decasper and Spence, 1986; Moon et al., 1993; Mahmoudzadeh et al., 2013; Moon et al., 2013; Partanen et al., 2013), it is possible that the disrupted maturation of auditory cortex that we observed here could underlie, at least in part, the auditory process-

ing, speech, and language deficits widely reported for preterm infants (Pasman et al., 1992; Davis et al., 2001; Mikkola et al., 2007; Barre et al., 2011; van Noort-van der Spek et al., 2012; Reidy et al., 2013; Vohr, 2014; Paquette et al., 2015). Whereas the auditory periphery and auditory brainstem appear largely unaffected by preterm birth (Eggermont and Salamy, 1988; Jiang, 1995; Eggermont et al., 1996; Tognola et al., 2005; Jedrzejczak et al., 2007; Jiang et al., 2009; Li et al., 2013), our findings suggest that auditory cognitive deficits in preterm infants originate in auditory cortex.

Our data provide *in vivo* demonstration of an important facet of human cortical maturation: primary sensory cortex develops earlier than nonprimary cortex. We were able to distinguish between pAC and nAC as early as 28 weeks PMA, a time at which the sulcal boundaries of HG are just beginning to appear. We found associations between diffusion parameters and preterm birth, and between diffusion FA and language function. In conclusion, diffusion MRI provides a unique window into cortical maturation in human infants, for whom histology is rarely available.

References

- Awad M, Warren JE, Scott SK, Turkheimer FE, Wise RJ (2007) A common system for the comprehension and production of narrative speech. J Neurosci 27:11455–11464. CrossRef Medline
- Ball G, Srinivasan L, Aljabar P, Counsell SJ, Durighel G, Hajnal JV, Rutherford MA, Edwards AD (2013) Development of cortical microstructure in the preterm human brain. Proc Natl Acad Sci USA 110:9541–9546. CrossRef
- Barre N, Morgan A, Doyle LW, Anderson PJ (2011) Language abilities in children who were very preterm and/or very low birth weight: a meta-analysis. J Pediatr 158:766–774.e1. CrossRef
- Bayley N, Reuner G (2006) Bayley scales of infant and toddler development: Bayley-III: Harcourt Assessment. San Antonio: Psy-chCorp.
- Birnholz JC, Benacerraf BR (1983) The development of human fetal hearing. Science 222:516–518. Medline
- Bose M, Muñoz-Llancao P, Roychowdhury S, Nichols JA, Jakkamsetti V, Porter B, Byrapureddy R, Salgado H, Kilgard MP, Aboitiz F, Dagnino-Subiabre A, Atzori M (2010) Effect of the environment on the dendritic morphology of the rat auditory cortex. Synapse 64: 97–110. CrossRef Medline
- Bourgeois JP, Jastreboff PJ, Rakic P (1989) Synaptogenesis in visual cortex of normal and preterm monkeys: evidence for intrinsic regulation of synaptic overproduction. Proc Natl Acad Sci USA 86:4297–4301. CrossRef
- Cardoso MJ, Melbourne A, Kendall GS, Modat M, Robertson NJ, Marlow N, Ourselin S (2013) AdaPT: an adaptive preterm segmentation algorithm for neonatal brain MRI. Neuroimage 65:97–108. CrossRef Medline
- Clarke S, Morosan P (2012) Architecture, connectivity, and transmitter receptors of human auditory cortex. In: The human auditory cortex, pp 11-38. New York: Springer.
- Conel JLR (1939) The postnatal development of the human cerebral cortex. Vol. 1. The cortex of the newborn. J Anat 73:674.
- Da Costa S, van der Zwaag W, Marques JP, Frackowiak RS, Clarke S, Saenz M (2011) Human primary auditory cortex follows the shape of Heschl's gyrus. J Neurosci 31:14067–14075. CrossRef Medline
- Davis MH, Johnsrude IS (2003) Hierarchical processing in spoken language comprehension. J Neurosci 23:3423–3431. Medline
- Davis NM, Doyle LW, Ford GW, Keir E, Michael J, Rickards AL, Kelly EA, Callanan C (2001) Auditory function at 14 years of age of very-low-birthweight. Dev Med Child Neurol 43:191–196. Medline

- DeCasper AJ, Fifer WP (1980) Of human bonding: newborns prefer their mothers' voices. Science 208:1174–1176. Medline
- Decasper AJ, Spence MJ (1986) Prenatal maternal speech influences newborns perception of speech sounds. Infant Behav Dev 9:133– 150. CrossRef
- Eggermont JJ, Salamy A (1988) Maturational time course for the ABR in preterm and full term infants. Hear Res 33:35–47. Medline
- Eggermont JJ, Brown DK, Ponton CW, Kimberley BP (1996) Comparison of distortion product otoacoustic emission (DPOAE) and auditory brain stem response (ABR) traveling wave delay measurements suggests frequency-specific synapse maturation. Ear Hear 17:386–394. Medline
- Ghosh A, Antonini A, McConnell SK, Shatz CJ (1990) Requirement for subplate neurons in the formation of thalamocortical connections. Nature 347:179–181. CrossRef Medline
- Glasser MF, Van Essen DC (2011) Mapping human cortical areas in vivo based on myelin content as revealed by T1- and T2-weighted MRI. J Neurosci 31:11597–11616. CrossRef Medline
- Graziani LJ, Weitzman ED, Velasco MS (1968) Neurologic maturation and auditory evoked responses in low birth weight infants. Pediatrics 41:483–494. Medline
- Hagoort P (2014) Nodes and networks in the neural architecture for language: Broca's region and beyond. Curr Opin Neurobiol 28: 136–141. CrossRef Medline
- Hepper PG, Shahidullah BS (1994) Development of fetal hearing. Arch Dis Child 71:F81–F87. Medline
- Hickok G, Poeppel D (2007) The cortical organization of speech processing. Nat Rev Neurosci 8:393–402. CrossRef Medline
- Hoerder-Suabedissen A, Molnár Z (2015) Development, evolution and pathology of neocortical subplate neurons. Nat Rev Neurosci 16:133–146. CrossRef Medline
- Jardri R, Pins D, Houfflin-Debarge V, Chaffiotte C, Rocourt N, Pruvo J-P, Steinling M, Delion P, Thomas P (2008) Fetal cortical activation to sound at 33 weeks of gestation: a functional MRI study. Neuroimage 42:10–18. CrossRef Medline
- Jedrzejczak WW, Hatzopoulos S, Martini A, Blinowska KJ (2007) Otoacoustic emissions latency difference between full-term and preterm neonates. Hear Res 231:54–62. CrossRef Medline
- Jenkinson M, Beckmann CF, Behrens TEJ, Woolrich MW, Smith SM (2012) FSL. Neuroimage 62:782–790. CrossRef
- Jiang ZD (1995) Maturation of the auditory brainstem in low risk preterm infants: a comparison with age-matched full term infants up to 6 years. Early Hum Dev 42:49–65. Medline
- Jiang ZD, Brosi DM, Wu YY, Wilkinson AR (2009) Relative maturation of peripheral and central regions of the human brainstem from preterm to term and the influence of preterm birth. Pediatr Res 65:657–662. CrossRef Medline
- Jobe AH (2014) A risk of sensory deprivation in the neonatal intensive care unit. J Pediatr 164:1265–1267. CrossRef Medline
- Kanold PO, Kara P, Reid RC, Shatz CJ (2003) Role of subplate neurons in functional maturation of visual cortical columns. Science 301:521–525. CrossRef Medline
- Kidokoro H, Neil JJ, Inder TE (2013) A new MR assessment tool to define brain abnormalities in very preterm infants at term. AJNR Am J Neuroradiol 34:2208–2214. CrossRef
- Kostović I, Rakic P (1990) Developmental history of the transient subplate zone in the visual and somatosensory cortex of the macaque monkey and human brain. J Comp Neur 297:441–470. CrossRef Medline
- Kostović I, Jovanov-Milošević N, Radoš M, Sedmak G, Benjak V, Kostović-Srzenti M, Vasung L, uljat M, Radoš M, Hüppi P, Judaš M (2014) Perinatal and early postnatal reorganization of the subplate and related cellular compartments in the human cerebral wall as revealed by histological and MRI approaches. Brain Struct Funct 219:231–253. CrossRef Medline
- Kroenke CD, Van Essen DC, Inder TE, Rees S, Bretthorst GL, Neil JJ (2007) Microstructural changes of the baboon cerebral cortex during gestational development reflected in magnetic resonance imaging diffusion anisotropy. J Neurosci 27:12506–12515. Cross-Ref Medline

- Kuklisova-Murgasova M, Aljabar P, Srinivasan L, Counsell SJ, Doria V, Serag A, Gousias IS, Boardman JP, Rutherford MA, Edwards AD, Hajnal JV, Rueckert D (2011) A dynamic 4D probabilistic atlas of the developing brain. Neuroimage 54:2750–2763. CrossRef Medline
- Leaver AM, Rauschecker JP (2010) Cortical representation of natural complex sounds: effects of acoustic features and auditory object category. J Neurosci 30:7604–7612. CrossRef Medline
- Li M, Zhu L, Mai X, Shao J, Lozoff B, Zhao Z (2013) Sex and gestational age effects on auditory brainstem responses in preterm and term infants. Early Hum Dev 89:43–48. CrossRef Medline
- Liebenthal E, Binder JR, Spitzer SM, Possing ET, Medler DA (2005) Neural substrates of phonemic perception. Cereb Cortex 15:1621– 1631. CrossRef Medline
- Mahmoudzadeh M, Dehaene-Lambertz G, Fournier M, Kongolo G, Goudjil S, Dubois J, Grebe R, Wallois F (2013) Syllabic discrimination in premature human infants prior to complete formation of cortical layers. Proc Natl Acad Sci USA 110:4846–4851. CrossRef
- Marie D, Jobard G, Crivello F, Perchey G, Petit L, Mellet E, Joliot M, Zago L, Mazoyer B, Tzourio-Mazoyer N (2015) Descriptive anatomy of Heschl's gyri in 430 healthy volunteers, including 198 left-handers. Brain Struct Funct 220:729–743. CrossRef Medline
- McKinstry RC, Mathur A, Miller JH, Ozcan A, Snyder AZ, Schefft GL, Almli CR, Shiran SI, Conturo TE, Neil JJ (2002) Radial organization of developing preterm human cerebral cortex revealed by noninvasive water diffusion anisotropy MRI. Cereb Cortex 12:1237– 1243. Medline
- McMullen NT, Glaser EM (1988) Auditory cortical responses to neonatal deafening: pyramidal neuron spine loss without changes in growth or orientation. Exp Brain Res 72:195–200. Medline
- Mikkola K, Kushnerenko E, Partanen E, Serenius-Sirve S, Leipälä J, Huotilainen M, Fellman V (2007) Auditory event-related potentials and cognitive function of preterm children at five years of age. Clin Neurophysiol 118:1494–1502. CrossRef Medline
- Moerel M, De Martino F, Formisano E (2014) An anatomical and functional topography of human auditory cortical areas. Front Neurosci 8:225. CrossRef Medline
- Moon C, Cooper RP, Fifer WP (1993) 2-day-olds prefer their native language. Infant Behav Dev 16:495–500. CrossRef
- Moon C, Lagercrantz H, Kuhl PK (2013) Language experienced in utero affects vowel perception after birth: a two-country study. Acta Paediatr 102:156–160. CrossRef Medline
- Morosan P, Rademacher J, Schleicher A, Amunts K, Schormann T, Zilles K (2001) Human primary auditory cortex: cytoarchitectonic subdivisions and mapping into a spatial reference system. Neuroimage 13:684–701. CrossRef Medline
- Mowery TM, Kotak VC, Sanes DH (2015) Transient hearing loss within a critical period causes persistent changes to cellular properties in adult auditory cortex. Cereb Cortex 25:2083–2094. Cross-Ref Medline
- Norman-Haignere S, Kanwisher NG, McDermott JH (2015) Distinct cortical pathways for music and speech revealed by hypothesisfree voxel decomposition. Neuron 88:1281–1296. CrossRef Medline
- Paquette N, Vannasing P, Tremblay J, Lefebvre F, Roy MS, McKerral M, Lepore F, Lassonde M, Gallagher A (2015) Early electrophysiological markers of atypical language processing in prematurely born infants. Neuropsychologia 79:21–32. CrossRef Medline
- Partanen E, Kujala T, Näätänen R, Liitola A, Sambeth A, Huotilainen M (2013) Learning-induced neural plasticity of speech processing before birth. Proc Natl Acad Sci USA 110:15145–15150. CrossRef Medline

- Pasman JW, Rotteveel JJ, de Graaf R, Stegeman DF, Visco YM (1992) The effect of preterm birth on brainstem, middle latency and cortical auditory evoked responses (BMC AERs). Early Hum Dev 31:113–129. Medline
- Penhune VB, Zatorre RJ, MacDonald JD, Evans AC (1996) Interhemispheric anatomical differences in human primary auditory cortex: probabilistic mapping and volume measurement from magnetic resonance scans. Cereb Cortex 6:661–672. Medline
- Pineda R, Durant P, Mathur A, Inder T, Wallendorf M, Schlaggar BL (2017) Auditory exposure in the neonatal intensive care unit: room type and other predictors. J Pediatr 183:56–66.
- Pineda RG, Neil J, Dierker D, Smyser CD, Wallendorf M, Kidokoro H, Reynolds LC, Walker S, Rogers C, Mathur AM, Van Essen DC, Inder T (2014) Alterations in brain structure and neurodevelopmental outcome in preterm infants hospitalized in different neonatal intensive care unit environments. J Pediatr 164:52–60.e52. Cross-Ref
- Poeppel D (2014) The neuroanatomic and neurophysiological infrastructure for speech and language. Curr Opin Neurobiol 28:142– 149. CrossRef Medline
- R Core Team (2016) R: a language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing.
- Rademacher J, Caviness V, Steinmetz H, Galaburda A (1993) Topographical variation of the human primary cortices: implications for neuroimaging, brain mapping, and neurobiology. Cereb Cortex 3:313–329. Medline
- Reidy N, Morgan A, Thompson DK, Inder TE, Doyle LW, Anderson PJ (2013) Impaired language abilities and white matter abnormalities in children born very preterm and/or very low birth weight. J Pediatr 162:719–724. CrossRef Medline
- Rotteveel JJ, de Graaf R, Stegeman DF, Colon EJ, Visco YM (1987) The maturation of the central auditory conduction in preterm infants until three months post term. V. The auditory cortical response (ACR). Hear Res 27:95–110. Medline
- Sidman RL, Rakic P (1982) Development of the human central nervous system. In: Histology and histopathology of the nervous system (Haymaker W, Adams RD, eds), pp 3–145. Springfield: Charles C Thomas.
- Smyser TA, Smyser CD, Rogers CE, Gillespie SK, Inder TE, Neil JJ (2015) Cortical gray and adjacent white matter demonstrate synchronous maturation in very preterm infants. Cereb Cortex 26: 3370–3378.
- Starr A, Amlie R, Martin W, Sanders S (1977) Development of auditory function in newborn infants revealed by auditory brainstem potentials. Pediatrics 60:831–839. Medline
- Stephens GJ, Silbert LJ, Hasson U (2010) Speaker-listener neural coupling underlies successful communication. Proc Natl Acad Sci USA 107:14425–14430. CrossRef Medline
- Tognola G, Parazzini M, de Jager P, Brienesse P, Ravazzani P, Grandori F (2005) Cochlear maturation and otoacoustic emissions in preterm infants: a time-frequency approach. Hear Res 199:71– 80. CrossRef Medline
- van Noort-van der Spek IL, Franken MC, Weisglas-Kuperus N (2012) Language functions in preterm-born children: a systematic review and meta-analysis. Pediatrics 129:745–754. CrossRef Medline
- Vohr B (2014) Speech and language outcomes of very preterm infants. Semin Fetal Neonatal Med 19:78–83. CrossRef
- Weitzman ED, Graziani LJ (1968) Maturation and topography of the auditory evoked response of the prematurely born infant. Dev Psychobiol 1:79–89. CrossRef