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The Fos-Related Antigen 1–JUNB/Activator Protein 1 Transcription Complex, a Downstream Target of Signal Transducer and Activator of Transcription 3, Induces T Helper 17 Differentiation and Promotes Experimental Autoimmune Arthritis

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Dysfunction of T helper 17 (Th17) cells leads to chronic inflammatory disorders. Signal transducer and activator of transcription 3 (STAT3) orchestrates the expression of proinflammatory cytokines and pathogenic cell differentiation from interleukin (IL)-17-producing Th17 cells. However, the pathways mediated by STAT3 signaling are not fully understood. Here, we observed that Fos-related antigen 1 (FRA1) and JUNB are directly involved in STAT3 binding to sites in the promoters of *Fos11* and *Junb*. Promoter binding increased expression of IL-17 and the development of Th17 cells. Overexpression of *Fra1* and *Junb* in mice resulted in susceptibility to collagen-induced arthritis and an increase in Th17 cell numbers and inflammatory cytokine production. In patients with rheumatoid arthritis, FRA1 and JUNB were colocalized with STAT3 in the inflamed synovium. These observations suggest that FRA1 and JUNB are associated closely with STAT3 activation, and that this activation leads to Th17 cell differentiation in autoimmune diseases and inflammation.

Keywords: Fos-related antigen 1–JUNB, signal transducer and activator of transcription 3, T helper 17, autoimmune arthritis, inflammation

INTRODUCTION

T helper 17 (Th17) cells are a pathogenic subset of T helper lymphocytes that play a key role in inflammatory disorders leading to a severe chronic immune inflammatory response. Th17 cells release several proinflammatory cytokines including interleukin (IL)-17A, IL-21, and IL-22 (1). IL-17A is an inflammatory cytokine produced predominantly by Th17 cells with strong effects on stromal cells. IL-17A induces inflammatory cytokine production and leukocyte recruitment, which act to link the innate and adaptive branches of immunity (2). Both IL-17A and Th17 cells are highly involved in the pathogenesis of several autoimmune diseases, including rheumatoid arthritis (RA) (3, 4), while playing a significant role in autoimmune disorders mediated by excessive inflammation.

Signal transducer and activator of transcription 3 (STAT3) is an important transcription factor with DNA-binding properties whose activity is paramount to the inflammatory response. It has been suggested that STAT3 activation upregulates IL-17 production *via* Th17 cell proliferation (5, 6). Several transcription factors including STAT3 regulate Th17 cell differentiation (7–12); however, STAT3 also plays a key role in the immune inflammatory response. There is a general consensus that STAT3 is essential for Th17 cell differentiation (13). Moreover, STAT3 modulates the production of several cytokines including IL-17A and activates downstream transcription factors, such as RAR-related orphan receptor gamma isoform 2 (ROR γ t), which is responsible for the Th17 phenotype (14, 15).

The activator protein 1 (AP-1) family is a group of structurally and functionally related JUN (c-JUN, JUNB, and JUND) and FOS [c-FOS, FOSB, Fos-related antigen 1 (FRA1), and FRA2] transcription factors. AP-1 heterodimers are involved in a variety of biological processes including cell proliferation, differentiation, apoptosis, and inflammation (16, 17). It has been suggested that AP-1 proteins are involved in several pathological conditions (18–21), while JUN and FOS proteins are also associated with the immune inflammatory response. Modulation of c-FOS and c-JUN expression is critical for inhibition of IL-17 production (22) and the maintenance of suppressive regulatory T-cell function (23). Additionally, production of FRA1, a member of the FOS protein family, is increased by B cell stimulation (24). Furthermore, JUNB modulates the proliferation of B cells (25). This evidence suggests that FRA1 and JUNB may be involved in regulating the inflammatory immune response.

We hypothesized that FRA1 and JUNB modulate the Th17 cell-mediated inflammatory response. The aim of this study was to elucidate whether FRA1 and JUNB regulate autoimmune arthritis *via* Th17 cell differentiation and factors downstream of STAT3. We used *in vitro* models, *in vivo* animal models, and clinical specimens from patients with RA to investigate the biological importance of this pathway.

MATERIALS AND METHODS

Mice

Collagen-induced arthritis (CIA) was induced in 6–8-week-old male DBA/1J, BALB/c, and C57BL/6 mice (Orient, Korea).

To generate *Fra1/Junb* Tg mice, a pcDNA3.1+HA (Invitrogen, CA, USA) vector containing the FRA1 and JUNB proteins coupled to a linker peptide (3 \times GGGGS) was constructed. The *Fra1/Junb* fragment was synthesized by GenScript Corporation (NJ, USA), with codon optimization for expression in mammalian cells. *Fra1/Junb* Tg mice were bred from the C57BL/6 line and maintained in facilities at Macrogen Laboratories (Seoul, Korea). All mice were maintained under specific-pathogen-free conditions at the Institute of Medical Science, The Catholic University of Korea. The presence of the transgene in the founders was confirmed by PCR of genomic DNA extracted from the tail samples. Genotyping was performed by PCR analysis of genomic DNA obtained from mice at 3 weeks of age. All experimental procedures were examined and approved by the Animal Research Ethics Committee at the Catholic University of Korea.

Accession Codes

Raw RNA-seq data have been deposited in the NCBI Sequence Read Archive (SRR6320798 and SRR6320799).

A detailed description of all other experimental procedures and the statistical analysis is provided in the Section “Supplementary Materials and Methods” in Data Sheet S1 in Supplementary Material.

RESULTS

STAT3 Target Genes Are Differentially Expressed in Mouse Th17 Cells

Potential STAT3-binding sites were identified using publicly available chromatin immunoprecipitation sequencing (ChIP-Seq) data cross-referenced with differentially expressed genes in Th17 cells (14). We sequenced mRNA obtained from Th17 cells and naïve T cells and compared the results with potential STAT3-binding targets identified by ChIP-Seq to identify STAT3-regulated genes involved in Th17 cell differentiation. The literature was systematically reviewed for downstream STAT3 targets in humans and mice in multiple biological contexts. By combining these three datasets, we searched for genes with STAT3-binding sites that are upregulated during Th17 cell differentiation (**Figure 1A**; Table S1 in Supplementary Material). RNA-Seq was performed to identify genes differentially expressed between Th17 cells and naïve T cells. This integrated approach suggested that *Fosl1* (the gene locus of *Fra1*) and *Junb* are Th17 cell differentiation factors downstream of STAT3. As shown in **Figures 1B–F** and Table S2 in Supplementary Material, *Fos* and *Jun* were the predominant AP-1 subtypes expressed in naïve (CD4⁺CD62L⁺) T cells, whereas *Fosl1* and *Junb* were the predominant subtypes in Th17 cells. Other AP-1 subtypes were downregulated, suggesting that *Fosl1* and *Junb* AP-1 subtypes play an important role in Th17 cell differentiation.

Fra1 and *Junb* Are Highly Expressed in Th17 Cells, and Their Expression Is Regulated Directly by STAT3 in Mice

Quantitative real-time reverse transcription PCR (qRT-PCR) and immunoblot analyzes were used to confirm *Fra1* and *Junb*

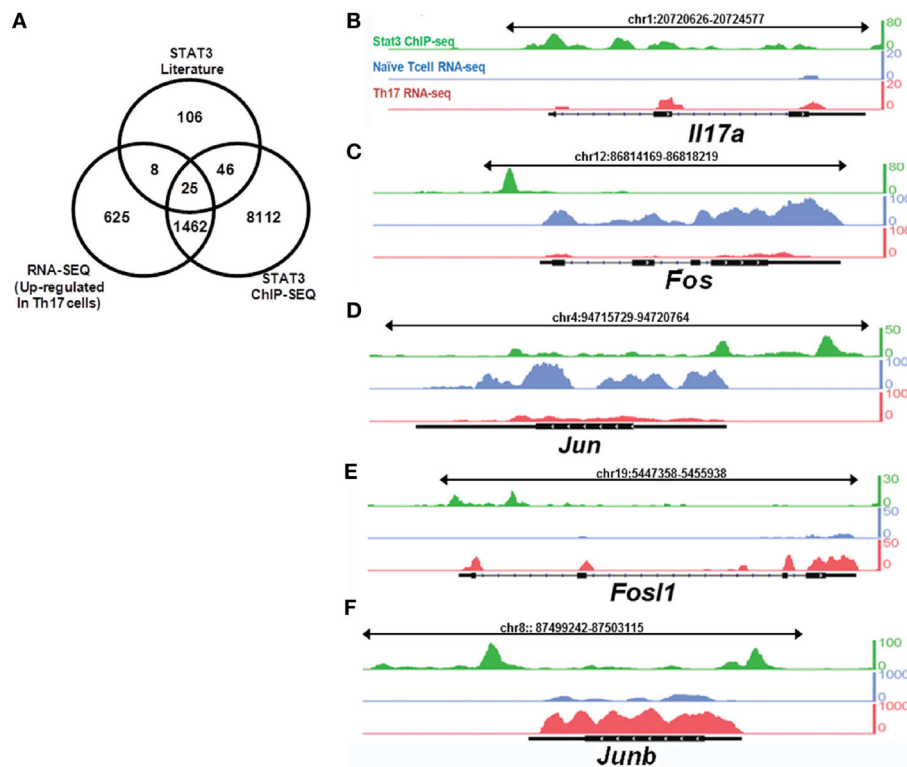


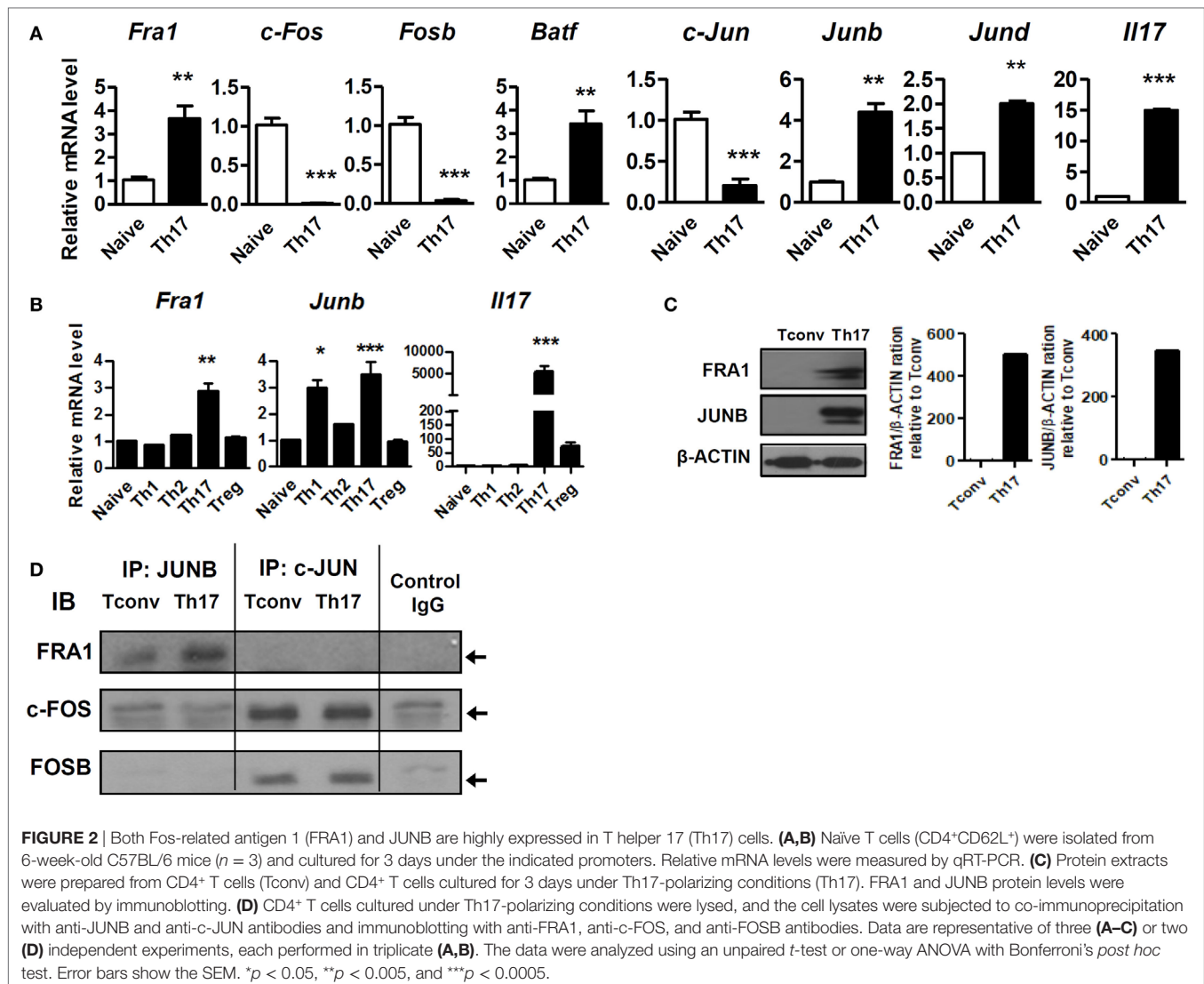
FIGURE 1 | Signal transducer and activator of transcription 3 (STAT3) target genes are differentially expressed in naïve and T helper 17 (Th17) cells. Naïve T cells (CD4⁺CD62L⁺) were isolated from 6-week-old mice ($n = 3$), stimulated under Th17-polarizing conditions for 3 days, and then sorted for CD4⁺CD62L⁻ expression. RNA was sequenced using the Illumina GA IIx. STAT3 chromatin immunoprecipitation sequencing (ChIP-Seq) data for CD4⁺ T cells were downloaded from the NCBI Sequence Read Archive (SRP002451). **(A)** A comparison of differentially expressed genes identified by RNA-Seq, STAT3-binding target genes identified by ChIP-Seq, and the downstream targets of STAT3. The expression of **(B)** *IL17a* and **(C-F)** activator protein 1 transcription factor target genes [**(C)** *Fos*, **(D)** *Jun*, **(E)** *Fos1*, and **(F)** *Junb*] was compared between naïve T cells and Th17 cells, with reference to STAT3-binding target genes (obtained from STAT3 ChIP-Seq data in Th17 cells). The peak profiles are color-coded as follows: naïve T cells (RNA-Seq: blue), Th17 cells (RNA-Seq: red), and STAT3-binding genes (ChIP-Seq: green).

expression. Among the various AP-1 subtypes, only *Fra1* and *Junb* mRNA levels were significantly increased in Th17 cells compared with naïve T cells (**Figures 2A,B**). These findings supported the RNA-Seq results. Immunoblot analysis also showed high levels of FRA1 and JUNB expression in Th17 cells (**Figure 2C**). Co-immunoprecipitation studies were performed using antibodies against Jun family proteins (i.e., JUNB and c-JUN), followed by immunoblotting using antibodies against FOS family proteins (i.e., FRA1, c-FOS, and FOSB) to determine whether FRA1 and JUNB form a heterodimeric AP-1 complex. It was found that FRA1 forms a complex with JUNB, but not with c-JUN (**Figure 2D**).

Next, we investigated the functional association between the FRA1–JUNB complex and STAT3. The STAT3 signaling pathway was inhibited using genetically engineered STAT3-deficient CD4⁺ T cells and STA-21, which blocks the DNA-binding activity of STAT3 (26). As shown in **Figure 3A**, CD4⁺ T cells harboring *Stat3^{fl/fl}Cd4-Cre* had markedly decreased levels of *Fra1* and *Junb* mRNA under both neutral and Th17-polarizing conditions. Treatment with STA-21 profoundly decreased the expression of FRA1, JUNB, and STAT3 (**Figure 3B**). Moreover, STA-21 decreased the levels of STAT3 phosphorylation under

Th17-polarizing conditions (**Figure 3C**). STAT3 overexpression increased the relative levels of *Fra1* and *Junb* mRNA (**Figure 3D**), while *in vitro* treatment with IL-6 upregulated *Fra1* and *Junb* mRNA (**Figure 3E**).

Co-immunoprecipitation studies were performed using anti-FRA1 and anti-JUNB antibodies followed by immunoblotting with antibodies against STAT3 to determine whether FRA1 and JUNB physically interact with STAT3. Both FRA1 and JUNB were found to form a complex with STAT3 (**Figure 3F**). We performed ChIP and qPCR to examine whether STAT3 binds directly to the promoter regions of *Fos1* and *Junb*. The ChIP-qPCR results were compared with publicly available ChIP-Seq data from Th17 cells. We found that STAT3 binds to the predicted recognition sites upstream of *Fos1* and *Junb*, suggesting that it regulates their expression by binding directly to their promoters (**Figure 3G**). Using confocal microscopy, we found that FRA1 and JUNB were colocalized with phosphorylated STAT3 in CD4⁺ splenic T cells and Th17 cells from arthritic mice (**Figures 3H,I**). These findings suggest that STAT3 signaling is critical for the expression and activation of FRA1 and JUNB during Th17 cell differentiation.

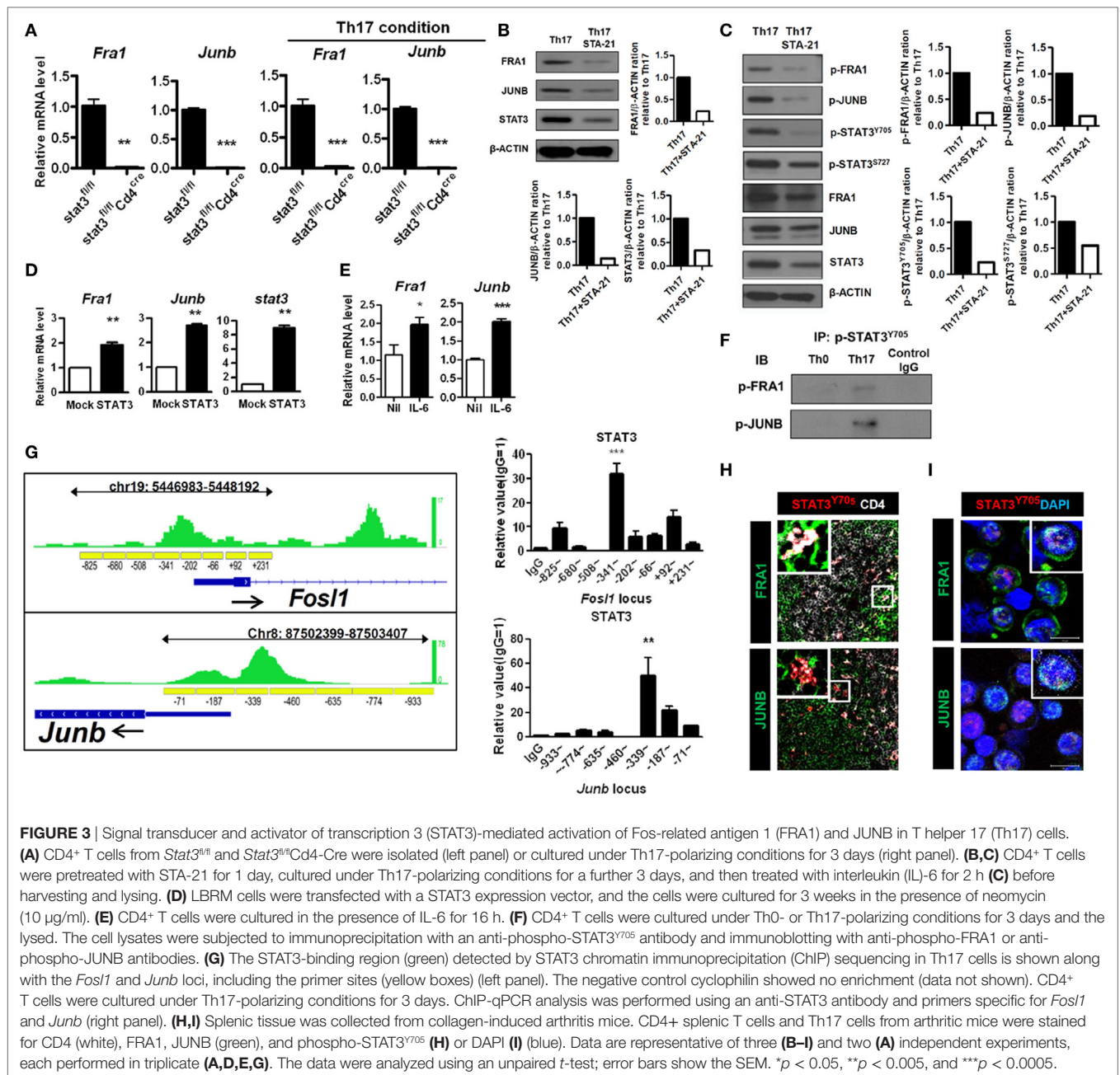


Expression of IL-17 Is Directly Regulated by the FRA1–JUNB Complex

The AP-1 family plays a versatile role in T-cell development (27). Levels of IL-17 were measured in *Fra1*- and *Junb*-overexpressing and silenced cells. The relative levels of *IL17* mRNA were significantly increased in cells overexpressing *Fra1* and *Junb*. The relative mRNA levels of other genes associated with Th17 cells, such as *IL21*, *IL22*, and *Rorc* (the ROR γ t gene locus), were also increased (Figures 4A,B). CD4⁺IL-17⁺ cells and *IL17* mRNA levels were significantly decreased when *Fra1* and *Junb* were silenced under Th17-polarizing conditions (Figures 4C,D). By contrast, when *c-Fos* and *c-Jun*, the dominant AP-1 subtypes in naïve T cells, were overexpressed, *IL17* mRNA levels were decreased significantly (Figures 4E,F). These findings support the specificity of the role of the FRA1–JUNB complex in Th17 cell differentiation.

We next investigated whether the FRA1–JUNB complex directly regulates IL-17 expression by binding to the *IL17a*

promoter. The interaction between the FRA1–JUNB complex and the AP-1-binding motif within the *IL17a* promoter region was examined by electrophoretic mobility shift assay. The anti-FRA1 and anti-JUNB antibodies caused a super shift of the labeled DNA probe, indicating that the AP-1-binding motif interacts specifically with FRA1 and JUNB (Figure 4G). This finding was confirmed by an analysis of the *IL17a* promoter using a luciferase reporter system. Overexpression of both FRA1 and JUNB increased luciferase activity in EL4 cells. Simultaneous overexpression of FRA1 and JUNB caused luciferase activity to increase twofold (Figure 4H). Silencing of *Fra1* and *Junb* caused a greater than twofold reduction in luciferase activity. Simultaneous overexpression of c-FOS and c-JUN caused a significant decrease in the luciferase activity induced by the *IL17a* promoter (Figure 4I). ChIP-qPCR analyses showed that FRA1 and JUNB bind directly to the *IL17a* gene loci and to gene loci encoding other Th17 cell differentiation activators, such as *IL21*, *IL22*, and *Rorc* (Figure 4J). These findings indicate that the FRA1–JUNB complex binds to



the *IL17a* promoter region, and that this binding activates *IL17* gene transcription.

FRA1 and JUNB Are Critical for the Pathogenesis of CIA

T helper 17 cells and IL-17 contribute significantly to the development of RA (28, 29). To examine the *in vivo* role of the FRA1–JUNB complex in rheumatoid inflammation, *Fra1* and *Junb* were silenced in CIA mice by injecting *Fra1* and *Junb* short hairpin RNA vectors. This markedly reduced the incidence of arthritis and decreased the arthritis score (Figure 5A). The numbers of FRA1⁺, JUNB⁺, and IL-17⁺ cells were significantly lower within

the joint tissues (Figure 5B). Histological examination showed that the ankle joints of *Fra1/Junb*-silenced mice had less inflammation and cartilage damage (Figure 5C). Inhibition of *Fra1/Junb* also reduced the expression of Th17 cytokines such as IL-6, IL-17, IL-1β, and tumor necrosis factor-α in joint tissue. Other mediators of joint destruction such as vascular endothelial growth factor and receptor activator of nuclear factor κB ligand (RANKL) were also downregulated (Figure S1A in Supplementary Material). In CIA mice, disease severity was correlated with the level of type II collagen (CII)-specific IgG antibodies (30). Silencing of *Fra1* and *Junb* reduced T-cell proliferation and CII-specific antibody production (Figures S1B,C in Supplementary Material). It also

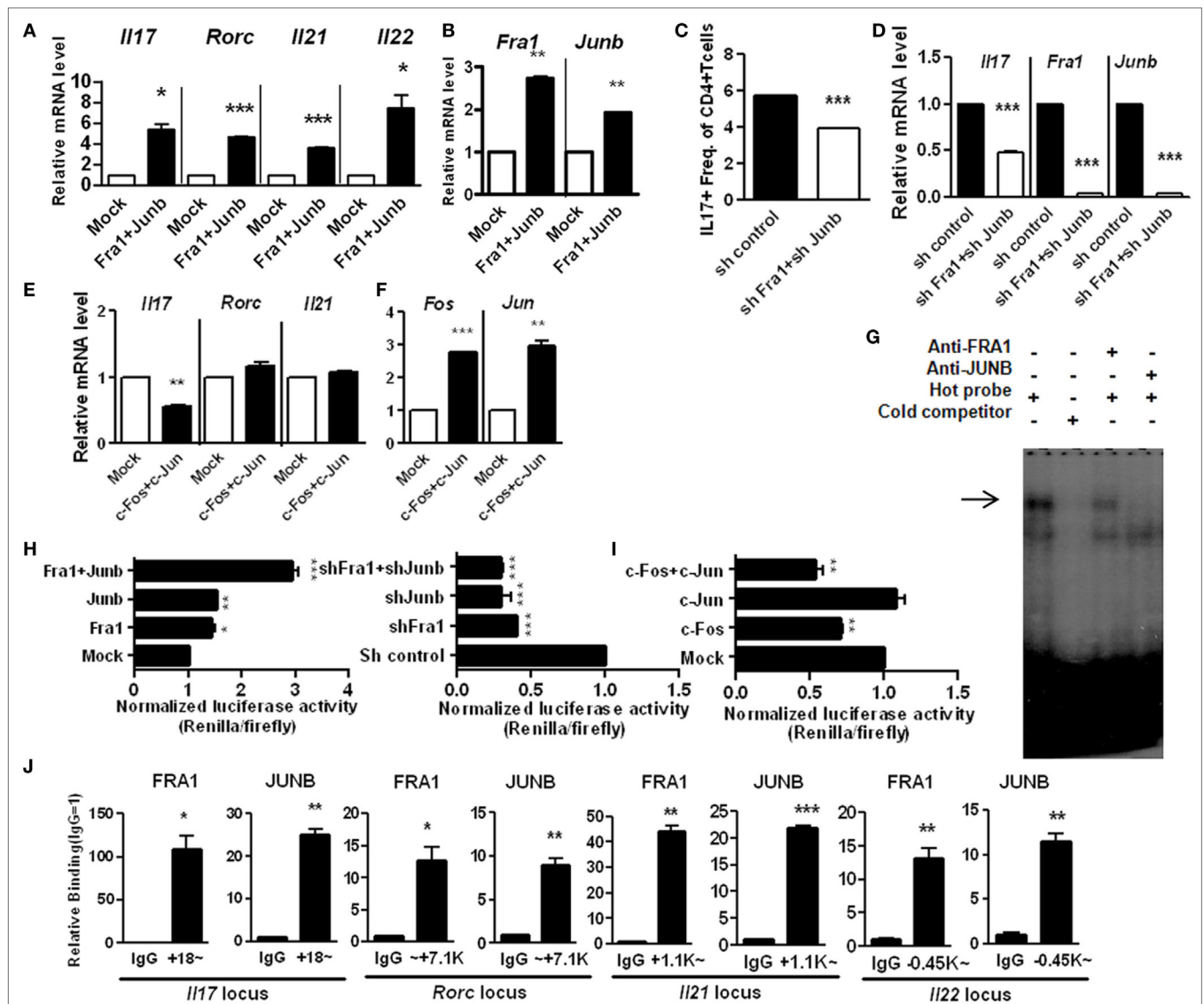
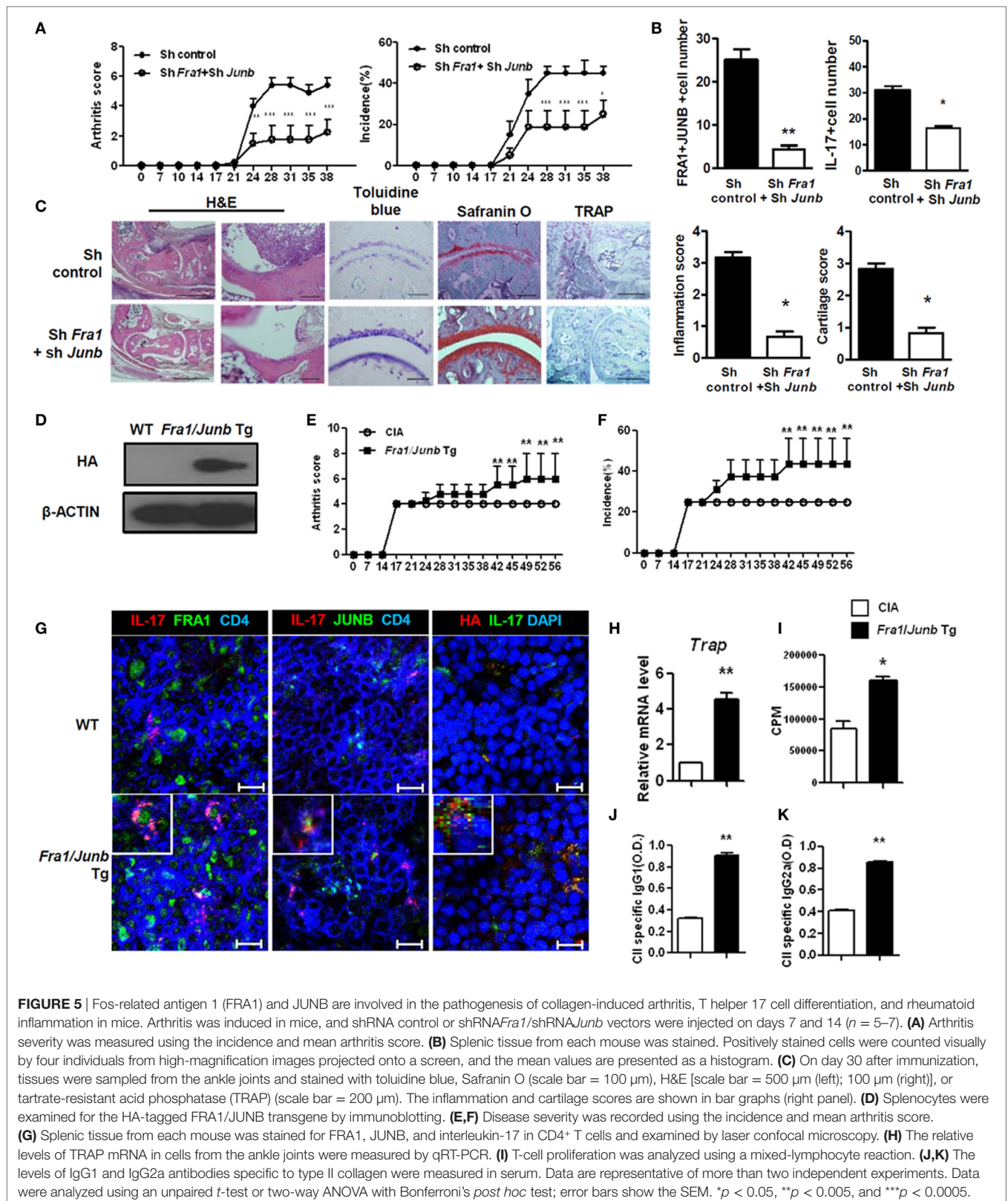


FIGURE 4 | Expression of interleukin (IL)-17 is directly mediated by the Fos-related antigen 1 (FRA1)-JUNB complex in T helper 17 (Th17) cells. LBRM cells were transfected with vectors and cultured for 3 weeks with neomycin (10 μ g/ml) (A,B,E,F). (A,B) The relative mRNA levels of each gene were measured by qRT-PCR. (C,D) CD4⁺ T cells were infected with lentiviruses harboring constructs designed to knockdown *Fra1* and *Junb*. Cells were then cultured under Th17-polarizing conditions for 3 days. CD4⁺IL-17⁺ T cells were identified by flow cytometry, and relative mRNA levels were measured by qRT-PCR. (E,F) The relative mRNA level of each gene was measured by qRT-PCR. (G) Nuclear extracts from Th17 cells were analyzed by electrophoretic mobility shift assay incorporating a radiolabeled probe derived from the activator protein 1-binding site of the *IL17a* locus and antibodies specific for FRA1 or JUNB. (H,I) EL4 cells were transiently transfected with expression vectors and *IL17a* promoter constructs with a CNS2 enhancer region. Luciferase activity was measured using a dual-luciferase reporter assay system. (J) CD4⁺ T cells were cultured under Th17-polarizing conditions. Chromatin immunoprecipitation-qPCR was performed using anti-FRA1 or anti-JUNB antibodies with primers specific for each gene locus. Data are representative of at least three independent experiments, performed in triplicate. Data were analyzed using an unpaired *t*-test; error bars show the SEM. **p* < 0.05, ***p* < 0.005, and ****p* < 0.0005.

substantially reduced the population of IL-17-producing CD4⁺ T cells, considered to be Th17 cells, in the spleen and draining lymph nodes (Figure S1D in Supplementary Material).

CD4⁺ T cells in *Fra1/Junb* Tg mice had different proportions of Th1 and Th17 cells (Figure S2 in Supplementary Material). CIA was induced in *Fra1/Junb* transgenic mice to investigate the *in vivo* effects of *Fra1* and *Junb* overexpression. Splenocytes were examined for HA-tagged FRA1/JUNB transgene by

immunoblotting (Figure 5D). The incidence of arthritis and the arthritis score were significantly increased (Figures 5E,F). There was a marked increase in the levels of IL-17, FRA1, and JUNB in the spleen (Figure 5G). The relative mRNA level of the osteoclastogenic marker tartrate-resistant acid phosphatase was significantly higher (Figure 5H), while CII-specific T-cell proliferation and IgG production were also increased (Figures 5I-K). Overexpression of FRA1/JUNB profoundly increased CIA



joint pathology (Figure S3A in Supplementary Material). Immunostaining showed that the expression of various proinflammatory cytokines and RANKL, a mediator of osteoclastogenesis,

was significantly greater in the joint tissues of transgenic mice with CIA (Figure S3B in Supplementary Material). IL-17 mRNA levels and Th17 cell numbers were increased markedly (Figures

S3C,D in Supplementary Material). These findings suggest that the FRA1–JUNB complex plays a major role in the development of CIA, likely through the induction of Th17 cell differentiation.

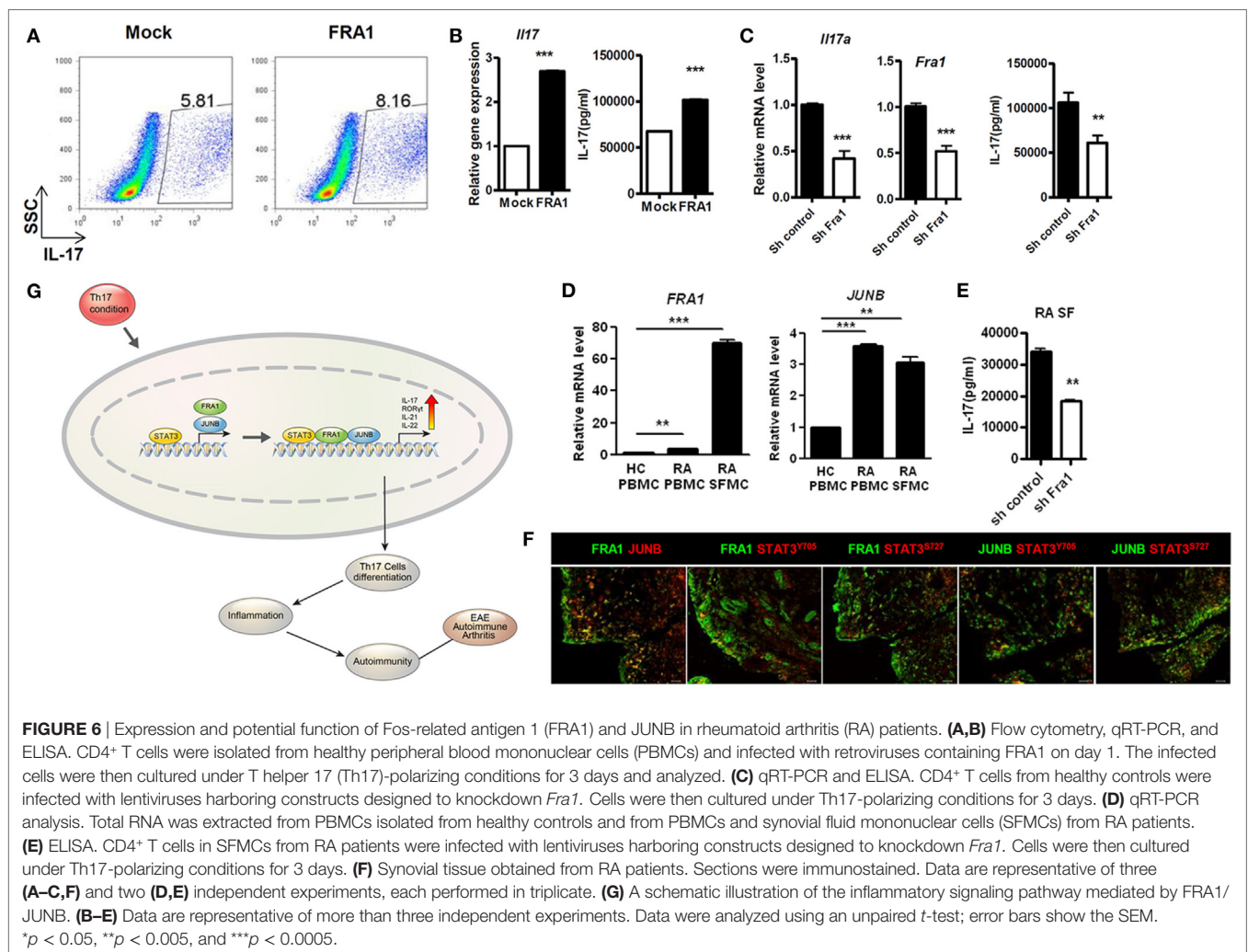
Expression and Potential Functions of FRA1 and JUNB in RA Patients

We demonstrated *in vivo* roles of FRA1 and JUNB in mice with CIA, an animal disease model for RA. To confirm the roles of FRA1 and JUNB in patients with RA, FRA1 was overexpressed in CD4⁺ T cells isolated from healthy controls. FRA1 overexpression increased the proportion of IL-17-producing CD4⁺ T cells during Th17 polarization (Figure 6A). These cells produced significantly higher levels of *IL17a* mRNA and cytokines (Figure 6B). *Fra1* silencing decreased *IL17a* mRNA levels and IL-17 secretion (Figure 6C). There were significantly increased the transcription levels of *Fra1* and *Junb* among peripheral blood mononuclear cells (PBMCs) and synovial fluid mononuclear cells (SFMCs) from RA patients compared with PBMCs from healthy controls (Figure 6D). *Junb* was also upregulated in RA patients. When

Fra1 was silenced in CD4⁺ T cells isolated from SFMCs of RA patients, IL-17 secretion was decreased during Th17 polarization (Figure 6E). The expression of both FRA1 and JUNB was particularly high in the inflamed synovium of RA patients, a region enriched with inflammatory immune cells (Figure 6F). Of note, FRA1, JUNB, and phosphorylated STAT3 were colocalized in the inflamed synovium, which further supported our *in vitro* experimental results. A schematic illustration of the inflammatory signaling pathway induced by FRA1/JUNB is shown in Figure 6G. Th17 conditioning induces an inflammatory signal by promoting FRA1/JUNB. The inflammatory response is mediated by activation of STAT3, which regulates genes involved in Th17 cell differentiation. This cascade induces excessive inflammation and autoimmunity resulting in RA.

DISCUSSION

Although FRA1 and JUNB may have modulatory activity in the immune inflammatory response (18, 19), there is currently no



evidence in the literature to support this notion in autoimmune disorders mediated by STAT3 and Th17 cells. In this study, we found evidence of previously unidentified functions of FRA1 and JUNB in Th17 cell differentiation and autoimmune disease. We investigated the modulatory function and underlying mechanisms of FRA1 and JUNB in a murine model of inflammatory arthritis. The predominant conclusion from this study was that FRA1 and JUNB exacerbate inflammation *via* the induction of Th17 cells. To our knowledge, this is the first study to implicate FRA1 and JUNB directly in STAT3 activation and regulation of Th17 cell proliferation. These observations suggest a previously unidentified function of these factors in autoimmune disease.

Activator protein 1 is comprised of dimers of FOS and JUN protein family members (31, 32). The diverse functions of AP-1 family members result from heterodimer formation between the FOS and JUN proteins (33). AP-1 is also involved in cell functions including proliferation, differentiation, and apoptosis. For example, the FRA2–JUNB complex is involved in the terminal differentiation of granulosa cells into luteal cells (34) and T-cell activation (35). Additionally, AP-1 factors may play important roles in autoimmune disease and Th17 cell differentiation. It was suggested that basic leucine zipper transcription factor, ATF-like (BATF), an AP-1 protein, leads to increased Th17 cell differentiation, whereas BATF deficiency results in IL-17 downregulation (12). Loss of the AP-1 family member *Fosl2* in T cells improved EAE severity *via* suppression of Th17 cell plasticity (36). We performed an integrative analysis using RNA-Seq and ChIP-Seq data to identify key factors downstream of STAT3 involved in Th17 cell differentiation. This integrative analysis suggested that upregulation of *Fra1* and *Junb* drives and promotes Th17 cell differentiation.

Signal transducer and activator of transcription 3-mediated IL-17 expression and Th17 cell differentiation involve AP-1 family proteins. Previous studies have shown a biological link between the STAT3 signaling pathway and the AP-1 family of transcription factors (12, 37, 38). In particular, STAT3-mediated expression of BATF, an AP-1 subfamily member, is associated with Th17 cell differentiation (12). In this study, RNA-Seq analysis showed that the expression of *cFos* and *cJun* was higher in naïve T cells. This suggests that AP-1 subtype switching from c-FOS–c-JUN to FRA1/JUNB is essential for Th17 cell differentiation. It can also be postulated that activation of c-FOS–c-JUN must be suppressed in parallel with activation of FRA1/JUNB by STAT3 during polarization toward the Th17 subtype. The present study identified an AP-1 complex comprising FRA1 and JUNB, which appears to function as a key factor downstream of STAT3 in Th17 cell differentiation. It appears that AP-1 subtype switching, particularly from c-FOS, FOSB, and c-JUN to FRA1 and JUNB, is required for STAT3-mediated Th17 cell differentiation.

The AP-1 family includes the c-JUN and c-FOS proteins, which play important roles in IL-17 expression and Th17 cell functions. It has been demonstrated that BATF binds to intergenic components in the *IL17a* locus and to *IL17* promoters, both of which are essential for Th17 differentiation (12). Additionally, the BATF–JUNB and BATF–JUND complexes have been shown to cooperate with IRF4 in Th17 cells to promote transcription of *IL17* (39). The c-JUN transcription factor that heterodimerizes with c-FOS

to generate the AP-1 transcription factor complex is involved in suppression of IL-17 production in developing Th17 cells (16, 22). FRA2 is also a negative regulator of IL-17 (36). We observed that overexpression of FRA2 decreased the expression of *IL17a* in a promoter-fused luciferase reporter assay (data not shown). We also found that expression levels of *c-Fos* and *c-Jun* were significantly reduced, whereas *Batf* expression was increased in Th17 cells compared with naïve T cells. Overexpression of *c-Fos* and *c-Jun* significantly decreased *IL17* mRNA levels. The FRA1–JUNB complex directly activates expression of IL-17, a cytokine produced by Th17 cells. This complex plays a role in promoting the expression of other cytokines associated with Th17 cells, such as IL-21 and IL-22, by directly binding to their promoter regions. These results suggest that FRA1 and JUNB are involved in IL-17 production and Th17 cell differentiation.

Although FRA1/JUNB and c-FOS/c-JUN compete for the same AP-1-binding sites within the *IL17a* promoter, we found in this study that *IL17a* promoter activity was increased by FRA1 and JUNB overexpression but decreased by c-FOS and c-JUN overexpression. We observed an interaction between p-STAT3 and p-FRA1 or p-JUNB in Th17 cells; therefore, FRA1/JUNB and c-FOS/c-JUN may be involved in AP-1 subtype switching. Additionally, p-FRA1 binds to p-JUNB in Th17 cells. Further study is needed to confirm the AP-1 subtype switching factors involved and the mechanism of interaction between p-FRA1 and p-JUNB in Th17 cells.

This study has certain limitations. One is that the animal studies were conducted using FRA1/JUNB-1 overexpression and knockdown vectors. Adoptive transfer studies in conditional overexpression and knockout mice are required to confirm that FRA1/JUNB-1 regulates CD4⁺ T cells, leading to autoimmune arthritis. Nonetheless, this study is the first to demonstrate functions of FRA1/JUNB-1 in Th17 cell differentiation and autoimmune arthritis. Future studies using adoptive transfer models with CD4⁺ T cells differentially expressing FRA1/JUNB-1 are required to validate our data more precisely.

Uncontrolled Th17 cell activation is responsible for the onset of several autoimmune diseases. Th17 cells cause tissue injury *via* production of IL-17, as observed in CIA mouse models of disease. The present study showed that the FRA1–JUNB complex plays a central role in the development of RA, which is a Th17-mediated autoimmune disease. The FRA1–JUNB complex may be clinically relevant as it functions as a Th17 cell-specific regulator of proinflammatory cytokine production in CIA mouse models and hence RA. In the RA mouse model used in this study, FRA1/JUNB overexpression resulted in the development of severe inflammatory symptoms and augmented expression of IL-17 in mice. Notably, this has fueled interest in the genetic importance of FRA1/JUNB in human RA in the context of promoted STAT3 activity. FRA1 overexpression activated Th17 cell differentiation in PBMCs from healthy controls; however, FRA1 silencing downregulated IL-17 production in SFMCs from RA patients. This may indicate the potential of the FRA1–JUNB complex as a therapeutic target in human autoimmune disease. In conclusion, our results suggest a previously unidentified function of FRA1/JUNB in T-cell-mediated autoimmune diseases. FRA1/JUNB directly modulates STAT3 activation and Th17 cell differentiation. Our

identification of the involvement of FRA1/JUNB in T-cell development provides a potential therapeutic target for the treatment of autoimmune diseases.

ETHICS STATEMENT

The Animal Care Committee of The Catholic University of Korea approved the experimental protocol. All experimental procedures were evaluated and carried out in accordance with the protocols approved by the Animal Research Ethics Committee at the Catholic University of Korea (CMCU-2012-0156-01). All procedures performed followed the ethical guidelines on animal use. Approval by the ethics committee of Seoul St. Mary's Hospital (Seoul, Republic of Korea) was obtained for all procedures. All human experimental procedures were approved by the Ethics Committee of Seoul St. Mary's Hospital (Seoul, Republic of Korea, KC13TISE0032).

AUTHOR CONTRIBUTIONS

Y-MM, S-YL, S-KK, SL, S-MA, and M-LC designed the experiments and analyzed the data. Y-MM, S-YL, S-KK, and SL wrote the manuscript along with input from DK, WK, Y-MH, H-JS, E-KK, J-GR, H-BS, J-EK, S-YH, and JY. Y-MM and S-YL performed all *in vitro* assays with help from S-KK, SL, DK, WK, and H-BS. S-YL, H-JS, and J-GR performed the animal experiments. E-KK conducted all immunohistochemistry experiments. Y-MM, S-YL, S-KK, SL, RS, D-MJ, H-YK, S-HP, S-MA, and M-LC discussed

and developed the study concept. All authors critically reviewed and approved the final form of the manuscript.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at <http://www.frontiersin.org/articles/10.3389/fimmu.2017.01793/full#supplementary-material>.

FIGURE S1 | Knockdown of FRA1/JUNB Decreases Th17 Cell Differentiation and Attenuates Rheumatoid Inflammation in Mice.

FIGURE S2 | T cell proportion of CD4⁺Tcells in Fra1/Junb Tg mice.

FIGURE S3 | Overexpression of FRA1/JUNB Causes Th17 Cell Differentiation and Increases CIA Disease Severity.

REFERENCES

- Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, et al. Interleukin 17-producing CD4⁺ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol* (2005) 6:1123–32. doi:10.1038/ni1254
- Iwakura Y, Nakae S, Saijo S, Ishigame H. The roles of IL-17A in inflammatory immune responses and host defense against pathogens. *Immunol Rev* (2008) 226:57–79. doi:10.1111/j.1600-065X.2008.00699.x
- Komiyama Y, Nakae S, Matsuki T, Nambu A, Ishigame H, Kakuta S, et al. IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. *J Immunol* (2006) 177:566–73. doi:10.4049/jimmunol.177.1.566
- Kotake S, Udagawa N, Takahashi N, Matsuzaki K, Itoh K, Ishiyama S, et al. IL-17 in synovial fluids from patients with rheumatoid arthritis is a potent stimulator of osteoclastogenesis. *J Clin Invest* (1999) 103:1345–52. doi:10.1172/JCI5703
- Mathur AN, Chang HC, Zisoulis DG, Stritesky GL, Yu Q, O'Malley JT, et al. STAT3 and STAT4 direct development of IL-17-secreting Th cells. *J Immunol* (2007) 178:4901–7. doi:10.4049/jimmunol.178.8.4901
- O'Shea JJ, Paul WE. Mechanisms underlying lineage commitment and plasticity of helper CD4⁺ T cells. *Science* (2010) 327:1098–102. doi:10.1126/science.1178334
- Ivanov II, McKenzie BS, Zhou L, Tadokoro CE, Lepelley A, Lafaille JJ, et al. The orphan nuclear receptor ROR γ directs the differentiation program of proinflammatory IL-17+ T helper cells. *Cell* (2006) 126:1121–33. doi:10.1016/j.cell.2006.07.035
- Chaudhry A, Rudra D, Treuting P, Samstein RM, Liang Y, Kas A, et al. CD4⁺ regulatory T cells control Th17 responses in a STAT3-dependent manner. *Science* (2009) 326:986–91. doi:10.1126/science.1172702
- Yang XO, Panopoulos AD, Nurieva R, Chang SH, Wang D, Watowich SS, et al. STAT3 regulates cytokine-mediated generation of inflammatory helper T cells. *J Biol Chem* (2007) 282:9358–63. doi:10.1074/jbc.C600321200
- Pan F, Ye Z, Cheng L, Liu JO. Myocyte enhancer factor 2 mediates calcium-dependent transcription of the interleukin-2 gene in T lymphocytes: a calcium signaling module that is distinct from but collaborates with the nuclear factor of activated T cells (NFAT). *J Biol Chem* (2004) 279:14477–80. doi:10.1074/jbc.C300487200
- Zhang F, Meng G, Strober W. Interactions among the transcription factors Runx1, ROR γ and Foxp3 regulate the differentiation of interleukin 17-producing T cells. *Nat Immunol* (2008) 9:1297–306. doi:10.1038/ni.1663
- Schraml BU, Hildner K, Ise W, Lee WL, Smith WA, Solomon B, et al. The AP-1 transcription factor BATF controls T(H)17 differentiation. *Nature* (2009) 460:405–9. doi:10.1038/nature08114
- Chen Z, Laurence A, Kanno Y, Pacher-Zavisin M, Zhu BM, Tato C, et al. Selective regulatory function of Socs3 in the formation of IL-17-secreting T cells. *Proc Natl Acad Sci U S A* (2006) 103:8137–42. doi:10.1073/pnas.0600666103
- Durant L, Watford WT, Ramos HL, Laurence A, Vahedi G, Wei L, et al. Diverse targets of the transcription factor STAT3 contribute to T cell pathogenicity and homeostasis. *Immunity* (2010) 32:605–15. doi:10.1016/j.immuni.2010.05.003
- Bi Y, Liu G, Yang R. Th17 cell induction and immune regulatory effects. *J Cell Physiol* (2007) 211:273–8. doi:10.1002/jcp.20973
- Hess J, Angel P, Schorpp-Kistner M. AP-1 subunits: quarrel and harmony among siblings. *J Cell Sci* (2004) 117:5965–73. doi:10.1242/jcs.01589
- Thomsen MK, Bakiri L, Hasenfuss SC, Hamacher R, Martinez L, Wagner EF. JUNB/AP-1 controls IFN- γ during inflammatory liver disease. *J Clin Invest* (2013) 123:5258–68. doi:10.1172/JCI70405
- Pirusian ES, Nikol'skaia TA, Abdeev RM, Bruskin SA. Transcription factor AP-1 components as a psoriasis candidate genes. *Mol Biol (Mosk)* (2007) 41:1069–80.
- Sabatokos G, Rowe GC, Kveiborg M, Wu M, Neff L, Chiusaroli R, et al. Doubly truncated FosB isoform (Delta2DeltaFosB) induces osteosclerosis in transgenic mice and modulates expression and phosphorylation of Smads in osteoblasts independent of intrinsic AP-1 activity. *J Bone Miner Res* (2008) 23:584–95. doi:10.1359/jbmr.080110

20. Eferl R, Hasselblatt P, Rath M, Popper H, Zenz R, Komnenovic V, et al. Development of pulmonary fibrosis through a pathway involving the transcription factor FRA-2/AP-1. *Proc Natl Acad Sci U S A* (2008) 105:10525–30. doi:10.1073/pnas.0801414105
21. Min L, Ji Y, Bakiri L, Qiu Z, Cen J, Chen X, et al. Liver cancer initiation is controlled by AP-1 through SIRT6-dependent inhibition of survivin. *Nat Cell Biol* (2012) 14:1203–11. doi:10.1038/ncb2590
22. Coquet JM, Middendorp S, van der Horst G, Kind J, Veraar EA, Xiao Y, et al. The CD27 and CD70 costimulatory pathway inhibits effector function of T helper 17 cells and attenuates associated autoimmunity. *Immunity* (2013) 38:53–65. doi:10.1016/j.immuni.2012.09.009
23. Lee SM, Gao B, Fang D. FoxP3 maintains Treg unresponsiveness by selectively inhibiting the promoter DNA-binding activity of AP-1. *Blood* (2008) 111:3599–606. doi:10.1182/blood-2007-09-115014
24. Huo L, Rothstein TL. Isolation and characterization of murine FRA-1: induction mediated by CD40 and surface Ig is protein kinase C dependent. *J Immunol* (1996) 157:3812–8.
25. Szremska AP, Kenner L, Weisz E, Ott RG, Passegue E, Artwohl M, et al. JUNB inhibits proliferation and transformation in B-lymphoid cells. *Blood* (2003) 102:4159–65. doi:10.1182/blood-2003-03-0915
26. Song H, Wang R, Wang S, Lin J. A low-molecular-weight compound discovered through virtual database screening inhibits STAT3 function in breast cancer cells. *Proc Natl Acad Sci U S A* (2005) 102:4700–5. doi:10.1073/pnas.0409894102
27. Jochum W, Passegue E, Wagner EF. AP-1 in mouse development and tumorigenesis. *Oncogene* (2001) 20:2401–12. doi:10.1038/sj.onc.1204389
28. Lundy SK, Sarkar S, Tesmer LA, Fox DA. Cells of the synovium in rheumatoid arthritis. T lymphocytes. *Arthritis Res Ther* (2007) 9:202. doi:10.1186/ar2107
29. Tesmer LA, Lundy SK, Sarkar S, Fox DA. Th17 cells in human disease. *Immunol Rev* (2008) 223:87–113. doi:10.1111/j.1600-065X.2008.00628.x
30. Hietala MA, Jonsson IM, Tarkowski A, Kleinau S, Pekna M. Complement deficiency ameliorates collagen-induced arthritis in mice. *J Immunol* (2002) 169:454–9. doi:10.4049/jimmunol.169.1.454
31. Jariel-Encontre I, Salvat C, Steff AM, Pariat M, Acquaviva C, Furstoss O, et al. Complex mechanisms for c-FOS and c-JUN degradation. *Mol Biol Rep* (1997) 24:51–6. doi:10.1023/A:1006804723722
32. Stancovski I, Gonen H, Orian A, Schwartz AL, Ciechanover A. Degradation of the proto-oncogene product c-FOS by the ubiquitin proteolytic system in vivo and in vitro: identification and characterization of the conjugating enzymes. *Mol Cell Biol* (1995) 15:7106–16. doi:10.1128/MCB.15.12.7106
33. Bakiri L, Matsuo K, Wisniewska M, Wagner EF, Yaniv M. Promoter specificity and biological activity of tethered AP-1 dimers. *Mol Cell Biol* (2002) 22:4952–64. doi:10.1128/MCB.22.13.4952-4964.2002
34. Sharma SC, Richards JS. Regulation of AP1 (JUN/FOS) factor expression and activation in ovarian granulosa cells. Relation of JUNB and FRA2 to terminal differentiation. *J Biol Chem* (2000) 275:33718–28. doi:10.1074/jbc.M003555200
35. Boise LH, Petryniak B, Mao X, June CH, Wang CY, Lindsten T, et al. The NFAT-1 DNA binding complex in activated T cells contains FRA-1 and JUNB. *Mol Cell Biol* (1993) 13:1911–9. doi:10.1128/MCB.13.3.1911
36. Ciofani M, Madar A, Galan C, Sellars M, Mace K, Pauli F, et al. A validated regulatory network for Th17 cell specification. *Cell* (2012) 151:289–303. doi:10.1016/j.cell.2012.09.016
37. Leu JI, Crissey MA, Leu JP, Ciliberto G, Taub R. Interleukin-6-induced STAT3 and AP-1 amplify hepatocyte nuclear factor 1-mediated transactivation of hepatic genes, an adaptive response to liver injury. *Mol Cell Biol* (2001) 21:414–24. doi:10.1128/MCB.21.2.414-424.2001
38. Song Y, Qian L, Song S, Chen L, Zhang Y, Yuan G, et al. FRA-1 and STAT3 synergistically regulate activation of human MMP-9 gene. *Mol Immunol* (2008) 45:137–43. doi:10.1016/j.molimm.2007.04.031
39. Li P, Spolski R, Liao W, Wang L, Murphy TL, Murphy KM, et al. BATF-JUN is critical for IRF4-mediated transcription in T cells. *Nature* (2012) 490:543–6. doi:10.1038/nature11530

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