





Genome Sequences of Two Single-Stranded DNA Viruses Identified in Varroa destructor

Simona Kraberger, a Gabriel A. Visnovsky, Pon F. van Toor, Maketalena F. Male, Kara Waits, Rafaela S. Fontenele, a Arvind Varsania,d,f

- ^aThe Biodesign Center for Fundamental and Applied Microbiomics, Center for Evolution and Medicine, School of Life Sciences, Arizona State University, Tempe, Arizona, USA
- ^bDepartment of Chemical and Process Engineering, Biomolecular Interaction Centre, University of Canterbury, Christchurch, New Zealand
- ^cThe New Zealand Institute for Plant and Food Research Limited, Christchurch, New Zealand
- ^dSchool of Biological Sciences, Biomolecular Interaction Centre, University of Canterbury, Christchurch, New Zealand
- eSchool of Environmental and Life Sciences, The University of Newcastle, Callaghan, New South Wales,
- fStructural Biology Research Unit, Department of Clinical Laboratory Sciences, University of Cape Town, Cape Town, South Africa

ABSTRACT Varroa destructor is a ubiquitous and parasitic mite of honey bees, infecting them with pathogenic viruses having a major impact on apiculture. We identified two novel circular replication-associated protein (Rep)-encoding singlestranded (CRESS) DNA viruses from V. destructor sampled from a honey bee hive near Christchurch in New Zealand.

arroa destructor is a parasitic mite that feeds on the hemolymph of honey bee (Apis mellifera). This mite is implicated in colony collapse and transmits several viruses (1). So far, only positive-sense RNA viruses belonging to the Dicistroviridae and Iflaviridae genera (order Picornavirales) have been associated with V. destructor, and these viruses also infect honey bees (2, 3). However, DNA viruses associated with V. destructor have yet been identified. To determine if DNA viruses are present, 10 V. destructor individuals were collected from a honey bee (A. mellifera liqustica) hive near Christchurch (New Zealand; 43.6401°S, 172.4842°E) on 1 July 2015, 15 years after Varroa was first identified in New Zealand (4).

The Varroa specimens were pooled and homogenized with 1 ml of SM buffer (0.1 M NaCl, 50 mM Tris-HCl [pH 7.4]). The homogenate was centrifuged at $10,000 \times g$ for 5 min and the supernatant filtered through a 0.2- μ m-pore-size syringe filter. Viral DNA was extracted from the filtrate using a High Pure viral nucleic acid kit (Roche Diagnostics, USA), followed by amplification of circular DNA molecules using the illustra TempliPhi kit (GE Healthcare, USA) and sequencing in a Illumina HiSeq 2500 platform at Novogene, Hong Kong. The paired-end reads were de novo assembled using ABySS 1.9 (5), and contigs >500 nucleotides (nt) were analyzed against a viral sequence database. Two contigs sharing similarities to circular replication-associated protein (Rep)-encoding single-stranded (CRESS) DNA viruses were identified. Abutting primers were designed and used together with a HiFi HotStart DNA polymerase (Kapa Biosystems, USA) to amplify full viral genomes. These were cloned and Sanger sequenced at Macrogen, South Korea.

CRESS DNA viruses are classified into eight recognized families, and a large number are currently unclassified. Various CRESS DNA viruses have been identified in various insect samples, including whiteflies (6, 7), mosquitoes (8), dragonflies and damselflies

Received 1 February 2018 Accepted 8 February 2018 **Published** 1 March 2018

Citation Kraberger S, Visnovsky GA, van Toor RF, Male MF, Waits K, Fontenele RS, Varsani A. 2018. Genome sequences of two singlestranded DNA viruses identified in Varroa destructor. Genome Announc 6:e00107-18. https://doi.org/10.1128/genomeA.00107-18.

Copyright © 2018 Kraberger et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Gabriel A. Visnovsky, gabriel.visnovsky@canterbury.ac.nz. or Arvind Varsani, arvind.varsani@asu.edu.

Kraberger et al.

(9-11), ticks (12), and thrips (13). Of the two CRESS DNA viruses identified, one is a member of a recently established virus family, Genomoviridae (14, 15), and the other is a novel virus. Varroa mite associated genomovirus 1 (VmaGV-1; 2,194 nt), with a TAATATTAT nonanucleotide motif, shares 77.3% genome-wide pairwise identity with Pacific flying fox feces-associated gemycircularvirus-9 (PfffaGmV-9; GenBank accession no. KT732803) and thus is a member of the genus Gemycircularvirus of the family Genomoviridae. VmaGV-1 encodes three proteins, a capsid protein (CP) in the virion sense and a Rep and RepA in the complementary sense that share 46%, 84%, and 94% amino acid identity with the CP, Rep, and RepA of PfffaGmV-9, respectively. The second genome, Varroa mite associated virus 1 (VmaV-1; 1,811 nt), with a CAGTATTAC nonanucleotide motif, is a novel CRESS DNA virus that encodes a Rep and CP in the virion sense. This genome shares low similarity to other known viruses and is most closely related to Giant panda circovirus 4 (GenBank accession no. MF327576), sharing 60% genome-wide identity, 36% amino acid identity to the Rep, and no similarities to any known CP. The two genome sequences described here represent the first documented DNA viruses to be associated with V. destructor, and the pathogenicity of these viruses in Varroa or their parasitized host, honey bees, is currently unknown.

Accession number(s). The complete genome sequences of *Varroa* mite-associated genomovirus 1 isolate VPVL_36 and *Varroa* mite-associated virus 1 isolate VPVL_46 were deposited in GenBank with accession numbers MG571087 and MG571088, respectively.

ACKNOWLEDGMENT

This study was supported by a seed funding from the Biomolecular Interaction Centre, University of Canterbury, New Zealand, awarded to Gabriel A. Visnovsky and Arvind Varsani.

REFERENCES

- Le Conte Y, Ellis M, Ritter W. 2010. Varroa mites and honey bee health: can Varroa explain part of the colony losses? Apidologie 41:353–363. https://doi.org/10.1051/apido/2010017.
- Chen Y, Evans J, Feldlaufer M. 2006. Horizontal and vertical transmission of viruses in the honey bee, *Apis mellifera*. J Invertebr Pathol 92:152–159. https://doi.org/10.1016/j.jip.2006.03.010.
- Ongus JR, Peters D, Bonmatin JM, Bengsch E, Vlak JM, van Oers MM. 2004. Complete sequence of a picorna-like virus of the genus *Iflavirus* replicating in the mite *Varroa destructor*. J Gen Virol 85:3747–3755. https://doi.org/10.1099/vir.0.80470-0.
- Ministry of Agriculture and Forestry. 2000. Varroa jacobsoni: economic impact assessment. Ministry of Agriculture and Forestry, Wellington, New Zealand.
- Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJ, Birol I. 2009. ABySS: a parallel assembler for short read sequence data. Genome Res 19:1117–1123. https://doi.org/10.1101/gr.089532.108.
- Nakasu EY, Melo FL, Michereff-Filho M, Nagata T, Ribeiro BM, Ribeiro SG, Lacorte C, Inoue-Nagata AK. 2017. Discovery of two small circular ssDNA viruses associated with the whitefly *Bemisia tabaci*. Arch Virol 162: 2835–2838. https://doi.org/10.1007/s00705-017-3425-y.
- Ng TF, Duffy S, Polston JE, Bixby E, Vallad GE, Breitbart M. 2011. Exploring the diversity of plant DNA viruses and their satellites using vectorenabled metagenomics on whiteflies. PLoS One 6:e19050. https://doi .org/10.1371/journal.pone.0019050.
- Ng TFF, Willner DL, Lim YW, Schmieder R, Chau B, Nilsson C, Anthony S, Ruan YJ, Rohwer F, Breitbart M. 2011. Broad surveys of DNA viral diversity obtained through viral metagenomics of mosquitoes. PLoS One 6:e20579. https://doi.org/10.1371/journal.pone.0020579.

- Dayaram A, Potter KA, Pailes R, Marinov M, Rosenstein DD, Varsani A. 2015. Identification of diverse circular single-stranded DNA viruses in adult dragonflies and damselflies (Insecta: Odonata) of Arizona and Oklahoma, USA. Infect Genet Evol 30:278–287. https://doi.org/10.1016/ j.meegid.2014.12.037.
- Rosario K, Dayaram A, Marinov M, Ware J, Kraberger S, Stainton D, Breitbart M, Varsani A. 2012. Diverse circular ssDNA viruses discovered in dragonflies (Odonata: Epiprocta). J Gen Virol 93:2668–2681. https://doi.org/10.1099/vir.0.045948-0.
- Dayaram A, Galatowitsch ML, Argüello-Astorga GR, van Bysterveldt K, Kraberger S, Stainton D, Harding JS, Roumagnac P, Martin DP, Lefeuvre P, Varsani A. 2016. Diverse circular replication-associated protein encoding viruses circulating in invertebrates within a lake ecosystem. Infect Genet Evol 39:304–316. https://doi.org/10.1016/j.meegid.2016.02.011.
- Waits K, Edwards MJ, Cobb IN, Fontenele RS, Varsani A. 2017. Identification of an anellovirus and genomoviruses in ixodid ticks. Virus Genes. https://doi.org/10.1007/s11262-017-1520-5.
- Kraberger S, Polston JE, Capobianco HM, Alcala-Briseno RI, Fontenele RS, Varsani A. 2017. Genomovirus genomes recovered from *Echinothrips americanus* sampled in Florida, USA. Genome Announc 5:e00445-17. https://doi.org/10.1128/genomeA.00445-17.
- Varsani A, Krupovic M. 2017. Sequence-based taxonomic framework for the classification of uncultured single-stranded DNA viruses of the family Genomoviridae. Virus Evol 3:vew037. https://doi.org/10.1093/ve/vew037.
- Krupovic M, Ghabrial SA, Jiang D, Varsani A. 2016. Genomoviridae: a new family of widespread single-stranded DNA viruses. Arch Virol 161:2633–2643. https://doi.org/10.1007/s00705-016-2943-3.

Volume 6 Issue 9 e00107-18 genomea.asm.org 2