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## Complete Genome Sequence of *Corynebacterium ureicelerivorans* DSM 45051, a Lipophilic and Urea-Splitting Isolate from the Blood Culture of a Septicemia Patient

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*Corynebacterium ureicelerivorans* is an opportunistic pathogen with a lipophilic lifestyle and an exceptionally high urease activity. The genome sequence of the type strain revealed that lipophilism is caused by the lack of a fatty acid synthase gene. The *ureABCEFGD* genes are similar to the urease gene region of *Corynebacterium glucuronolyticum*.

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orynebacterium ureicelerivorans is a lipophilic coryneform bacterium that is biochemically characterized by the very rapid enzymatic conversion of urea in a standard urease test (1). The taxonomic description of this species is based on a single isolate from the blood culture of a patient suffering from fever and exhibiting symptoms of septicemia (1). C. ureicelerivorans is related to Corynebacterium mucifaciens and Corynebacterium pilbarense by 16S rRNA gene sequence comparison but can be differentiated by DNA-DNA hybridization and biochemical phenotypic methods (1, 2). The rpoB gene sequence was determined in six strains of C. ureicelerivorans, finding a similarity of 91 to 97% with the type strain of C. mucifaciens (3). C. ureicelerivorans was recovered from two unspecified clinical samples (4, 5)and from blood cultures and ascitic fluid of patients with digestive disorders (3). Antimicrobial susceptibility assays revealed a common resistance of the latter C. ureicelerivorans isolates to macrolide and lincosamide antibiotics (3). To attain genetic knowledge of this disease-associated pathogen, we sequenced the complete genome of the type strain C. ureicelerivorans DSM 45051, initially named IMMIB RIV-2301 (1).

*C. ureicelerivorans* DSM 45051 was obtained from the Leibniz Institute DSMZ and was grown at 37°C in brain heart infusion broth-yeast extract medium (6) supplemented with 1% (vol/vol) Tween 80. Genomic DNA was purified with a Genomic-tip 500/G and the Genomic DNA buffer set (Qiagen). A sequencing-ready DNA library was generated with the Nextera DNA sample preparation kit (Illumina) and was sequenced in a 2  $\times$  300 nucleotide paired-end run using the MiSeq reagent kit version 3 (600 cycles) and the MiSeq desktop sequencer (Illumina). This sequencing approach resulted in 3,404,646 paired reads and 818,435,952 detected bases. The paired reads were assembled with the Roche GS De Novo Assembler software (release 2.8), yielding 43 contigs in 36 scaffolds. The scaffolds were ordered by synteny analysis with the r2cat tool (7). The remaining gaps in the genome sequence were closed by PCR assays with the BIOTAQ DNA polymerase (Bioline) and subsequent sequencing of the PCR products was performed on an ABI 3730xl DNA analyzer (Applied Biosystems). The Consed tool (version 26) was used for editing and finishing of the sequence assemblies (8).

The genome of *C. ureicelerivorans* DSM 45051 consists of a circular chromosome with a size of 2,279,990 bp (65.01% mean G+C content) and an extrachromosomal copy of the corynephage  $\Phi$ CUREI2301I, specified by a size of 48,288 bp and a mean G+C content of 64.56%. The *C. ureicelerivorans* genome lacks a gene encoding a fatty acid synthase, which defines the lipophilism of this species as lipid auxotrophy, just as previously demonstrated for other lipophilic corynebacteria (9–11). A potential macrolide and lincosamide resistance of *C. ureicelerivorans* is encoded by an *erm*(X) gene region showing the best BLASTN hits to a corresponding gene region in *Actinobaculum schaalii* strain 16 from a human urine specimen (12). The urease locus of *C. ureicelerivorans* comprises the *ureABCEFGD* genes revealing the best BLASTX hits to the urease subunits of *Corynebacterium glucuronolyticum*, isolated from the genital tract of humans (13, 14).

Nucleotide sequence accession numbers. This genome project has been deposited in the GenBank database under the accession numbers CP009215 (chromosome) and CP009216 ( $\Phi$ CUREI2301I).

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## REFERENCES

- Yassin AF. 2007. Corynebacterium ureicelerivorans sp. nov., a lipophilic bacterium isolated from blood culture. Int. J. Syst. Evol. Microbiol. 57: 1200–1203. http://dx.doi.org/10.1099/ijs.0.64832-0.
- 2. Aravena-Roman M, Spröer C, Sträubler B, Inglis T, Yassin AF. 2010. Corynebacterium pilbarense sp. nov., a non-lipophilic corynebacterium

isolated from a human ankle aspirate. Int. J. Syst. Evol. Microbiol. **60**: 1484–1487.

- Fernández-Natal MI, Sáez-Nieto JA, Valdezate S, Rodríguez-Pollán RH, Lapeña S, Cachón F, Soriano F. 2009. Isolation of *Corynebacterium ureicelerivorans* from normally sterile sites in humans. Eur. J. Clin. Microbiol. Infect. Dis. 28:677–681. http://dx.doi.org/10.1007/s10096-008-0677-1.
- Keller PM, Rampini SK, Büchler AC, Eich G, Wanner RM, Speck RF, Böttger EC, Bloemberg GV. 2010. Recognition of potentially novel human disease-associated pathogens by implementation of systematic 16S rRNA gene sequencing in the diagnostic laboratory. J. Clin. Microbiol. 48:3397–3402. http://dx.doi.org/10.1128/JCM.01098-10.
- Seng P, Abat C, Rolain JM, Colson P, Lagier JC, Gouriet F, Fournier PE, Drancourt M, La Scola B, Raoult D. 2013. Identification of rare pathogenic bacteria in a clinical microbiology laboratory: impact of matrixassisted laser desorption ionization-time of flight mass spectrometry. J. Clin. Microbiol. 51:2182–2194. http://dx.doi.org/10.1128/JCM.00492-13.
- Bomholt C, Glaub A, Gravermann K, Albersmeier A, Brinkrolf K, Rückert C, Tauch A. 2013. Whole-genome sequence of the clinical strain *Corynebacterium argentoratense* DSM 44202, isolated from a human throat specimen. Genome Announc. 1(5):e00793-13. http://dx.doi.org/ 10.1128/genomeA.00793-13.
- 7. Husemann P, Stoye J. 2010. r2cat: synteny plots and comparative assembly. Bioinformatics 26:570-571. http://dx.doi.org/10.1093/bioinformatics/btp690.
- Gordon D, Green P. 2013. Consed: a graphical Editor for next-generation sequencing. Bioinformatics 29:2936–2937. http://dx.doi.org/10.1093/ bioinformatics/btt515.
- 9. Tauch A, Kaiser O, Hain T, Goesmann A, Weisshaar B, Albersmeier A, Bekel T, Bischoff N, Brune I, Chakraborty T, Kalinowski J, Meyer F, Rupp

O, Schneiker S, Viehoever P, Pühler A. 2005. Complete genome sequence and analysis of the multiresistant nosocomial pathogen *Corynebacterium jeikeium* K411, a lipid-requiring bacterium of the human skin flora. J. Bacteriol. **187**:4671–4682. http://dx.doi.org/10.1128/JB.187.13.4671-4682.2005.

- Tauch A, Trost E, Tilker A, Ludewig U, Schneiker S, Goesmann A, Arnold W, Bekel T, Brinkrolf K, Brune I, Götker S, Kalinowski J, Kamp PB, Lobo FP, Viehoever P, Weisshaar B, Soriano F, Dröge M, Pühler A. 2008. The lifestyle of *Corynebacterium urealyticum* derived from its complete genome sequence established by pyrosequencing. J. Biotechnol. 136: 11–21. http://dx.doi.org/10.1016/j.jbiotec.2008.02.009.
- 11. Tauch A, Schneider J, Szczepanowski R, Tilker A, Viehoever P, Gartemann KH, Arnold W, Blom J, Brinkrolf K, Brune I, Götker S, Weisshaar B, Goesmann A, Dröge M, Pühler A. 2008. Ultrafast pyrosequencing of *Corynebacterium kroppenstedtii* DSM44385 revealed insights into the physiology of a lipophilic corynebacterium that lacks mycolic acids. J. Biotechnol. 136:22–30. http://dx.doi.org/10.1016/j.jbiotec.2008.03.004.
- Hays C, Lienhard R, Auzou M, Barraud O, Guérin F, Ploy MC, Cattoir V. 2014. Erm(X)-mediated resistance to macrolides, lincosamides and streptogramins in *Actinobaculum schaalii*. J. Antimicrob. Chemother. 69: 2056–2060. http://dx.doi.org/10.1093/jac/dku099.
- 13. Riegel P, Ruimy R, de Briel D, Prévost G, Jehl F, Bimet F, Christen R, Monteil H. 1995. *Corynebacterium seminale* sp. nov., a new species associated with genital infections in male patients. J. Clin. Microbiol. 33: 2244–2249.
- Devriese LA, Riegel P, Hommez J, Vaneechoutte M, de Baere T, Haesebrouck F. 2000. Identification of *Corynebacterium glucuronolyticum* strains from the urogenital tract of humans and pigs. J. Clin. Microbiol. 38:4657–4659.