Journal of the Marine Biological Association of the United Kingdom, 2016, 96(2), 291–295.
doi:10.1017/S0025315414001805

Symplectella rowi (Porifera: Hexactinellida: Lyssacinosida) is a rossellid, not a euplectellid

MARTIN DOHRMANN

Department of Earth and Environmental Sciences, Ludwig-Maximilians-University Munich, Palaeontology and Geobiology, Molecular Geo- and Palaeobiology Lab, Richard-Wagner-Str. 10, 80333 Munich, Germany

The monospecific hexactinellid sponge genus Symplectella endemic to New Zealand waters was originally assigned to family Rossellidae within the order Lyssacinosida (subclass Hexasterophora), although affinities to family Euplectellidae were also noted. Seventy-eight years later, the genus was transferred to Euplectellidae (subfamily Corbitellinae) on rather subjective grounds. Here, I test these two competing taxonomic hypotheses with molecular phylogenetic methods and demonstrate that Symplectella rowi *is indeed a rossellid, as was originally suggested. The genus is officially transferred back to Rossellidae (subfamily Rossellinae), which represents another small step towards a more natural classification system of glass sponges.*

Keywords: Classification, Euplectellidae, Hexactinellida, integrative taxonomy, molecular phylogenetics, Porifera, Rossellidae, *Symplectella*

Submitted 5 September 2014; accepted 2 November 2014; first published online 26 November 2014

INTRODUCTION

Symplectella rowi Dendy, 1924 is a lyssacine hexasterophoran glass sponge species (Porifera: Hexactinellida: Hexasterophora: Lyssacinosida) endemic to New Zealand waters (Dendy, 1924; Tabachnick, 2002a; Van Soest *et al.*, 2014). Although Dendy (1924, p. 287) noted that his new genus 'appears to be not without affinities with the Euplectellidae', he assigned it to family Rossellidae. Seventy-eight years later, in the seminal revision of all sponge genera compiled by Hooper & Van Soest (2002), Tabachnick (2002a) transferred the genus to Euplectellidae (Corbitellinae). The reasons for this move were rather vague, however: although Tabachnick (2002a, p. 1414) admitted that the absence of hypodermal pentactins also characterizes some rossellid species, he asserted that this feature supports assignment of *Symplectella* to Euplectellidae. Likewise, the absence of atrialia was considered '*rather* a feature of *some* Corbitellinae and *some* Euplectellinae than of Rossellidae' (my italics). Finally, the absence of the euplectellid-specific large dermalia with long rays – *Symplectella* has small dermalia with short rays as is characteristic for Rossellidae – did not prevent him from advocating a euplectellid affinity. Instead, he argued that the thick-rayed nature of *Symplectella*'s dermal hexactins and pentactins closely resembles spicules of the osculum-covering sieve plate found in many Corbitellinae. However, the sieve plate of *Symplectella* is constructed from regular diactins (Dendy, 1924, p. 288), and normal dermalia can hardly be homologized with specialized spicules restricted to the oscular region. The presence of a sieve plate *per se* also does not provide an unambiguous argument for a euplectellid affinity of *Symplectella*. Although sieve plates are widespread among euplectellids and unknown from any other rossellids, they are also found in distantly related families, namely Aphrocallistidae, Aulocalycidae and Hyalonematidae (Reiswig 2002a, b; Tabachnick & Menshenina, 2002), and therefore seem to be prone to convergent evolution. In my view, none of the characters cited by Tabachnick (2002a) provide compelling arguments for a placement of *Symplectella* in Euplectellidae. On the other hand, Dendy (1924) did not provide any justification for the placement of his new genus in Rossellidae, either (besides, perhaps, the absence of floricomes, a spicule type that is however not present in all genera of Euplectellidae). The contrasting decisions of Dendy and Tabachnick seem to be largely subjective, necessitating independent evidence for a firm placement of *Symplectella* among the families of Lyssacinosida. Fortunately, I was able to obtain molecular sequence data from a specimen of *S. rowi* included in the National Cancer Institute (NCI) collection housed at the Smithsonian Institution's National Museum of Natural History (NMNH), which allowed me to test the competing hypotheses by means of molecular phylogenetic analysis.

Corresponding author:
M. Dohrmann
Email: m.dohrmann@lrz.uni-muenchen.de

MATERIALS AND METHODS

A specimen of *S. rowi* (NCI OCDN 6625-L), collected on 16 April 1999 off New Zealand at 200 m depth, was subsampled for molecular work at the NMNH, Washington, DC, and further processed at the Molecular Geo- & Palaeobiology

Lab at LMU Munich. Identification as *Symplectella rowi* was confirmed by light microscopy investigation of temporary spicule preparations made by digesting small pieces of tissue in commercial bleach. DNA extraction, polymerase chain reaction (PCR) and Sanger sequencing to obtain phylogenetic markers established for Hexactinellida (Dohrmann *et al.*, 2012b) were carried out using previously developed protocols (Dohrmann *et al.*, 2008, 2009, 2012b). Despite substantial efforts, the cytochrome oxidase subunit I gene (COI), the 18S ribosomal RNA gene (18S rDNA), and the 5′-half of the 28S rDNA fragment could not be amplified (presumably because of DNA degradation during preservation of the specimen), leaving only the 16S rDNA fragment and the 3′-half of the 28S rDNA fragment for phylogenetic inference. The new sequences (GenBank accession numbers LN624751 and LN624750) were manually added to established alignments (Dohrmann *et al.*, 2012a, b). Initial phylogenetic analyses involving all available hexactinellid orthologues confirmed placement of *S. rowi* in Lyssacinosida (results not shown). For the final analyses, alignments were thus restricted to Lyssacinosida (plus *Iphiteon panicea* [Hexactinosida: Dactylocalycidae] as an outgroup) to allow inclusion of additional informative nucleotide positions that were unalignable across the whole taxon set. Phylogenetic analyses were conducted using maximum likelihood (ML) as implemented in RAxML v. 8.0.26 (Stamatakis, 2014) and were carried out on the 16S rDNA alignment (412 bp after removal of unalignable positions), the 28S rDNA alignment (1199 bp), a combination of the two (1610 bp) and a combination of all four markers (4805 bp). For COI and 16S rDNA, GTR + G models (Lanave *et al.*, 1984; Yang, 1994) were employed, and for 18S and 28S rDNA the S16 + G paired-sites model (see Savill *et al.*, 2001) was assigned to stem-encoding regions in addition to GTR + G for loop-encoding regions. In the combined analyses, all model parameters except topology and branch lengths were unlinked across partitions (except the stem-encoding regions of 18S and 28S rDNA, which were treated as a single partition due to software limitations). Clade support was estimated using rapid nonparametric bootstrapping (Felsenstein, 1985; Stamatakis *et al.*, 2008) under the 'autoMRE' option to automatically determine the sufficient number of pseudoreplicates (Pattengale *et al.*, 2010). Alignments and trees are available at OpenData LMU (doi: 10.5282/ubm/data.62).

RESULTS

In the phylogeny reconstructed from the 16S rDNA alignment (Figure 1), neither Euplectellidae nor Rossellidae is monophyletic. However, *Symplectella* nests within a well-supported clade (89% bootstrap support [BS]) that includes all rossellids as well as *Leucopsacus* + *Oopsacas* (Leucopsacidae) and *Clathrochone* (*incertae sedis*) to the exclusion of the euplectellids. In the 28S rDNA phylogeny (Figure 2), Euplectellidae is likewise paraphyletic but Rossellidae including *Symplectella* is reconstructed as a clade (72% BS). In the combined 16S + 28S rDNA phylogeny (Figure 3), this clade has 84% BS and the Rossellidae (including *Symplectella*) + Leucopsacidae + *Clathrochone* clade has maximal support (100% BS). Finally, in the phylogeny based on all four markers (Figure 4), *Symplectella* firmly nests within Rossellidae (100% BS), in a well-supported (94% BS) subclade containing most members of subfamily Rossellinae (except *Caulophacus* and

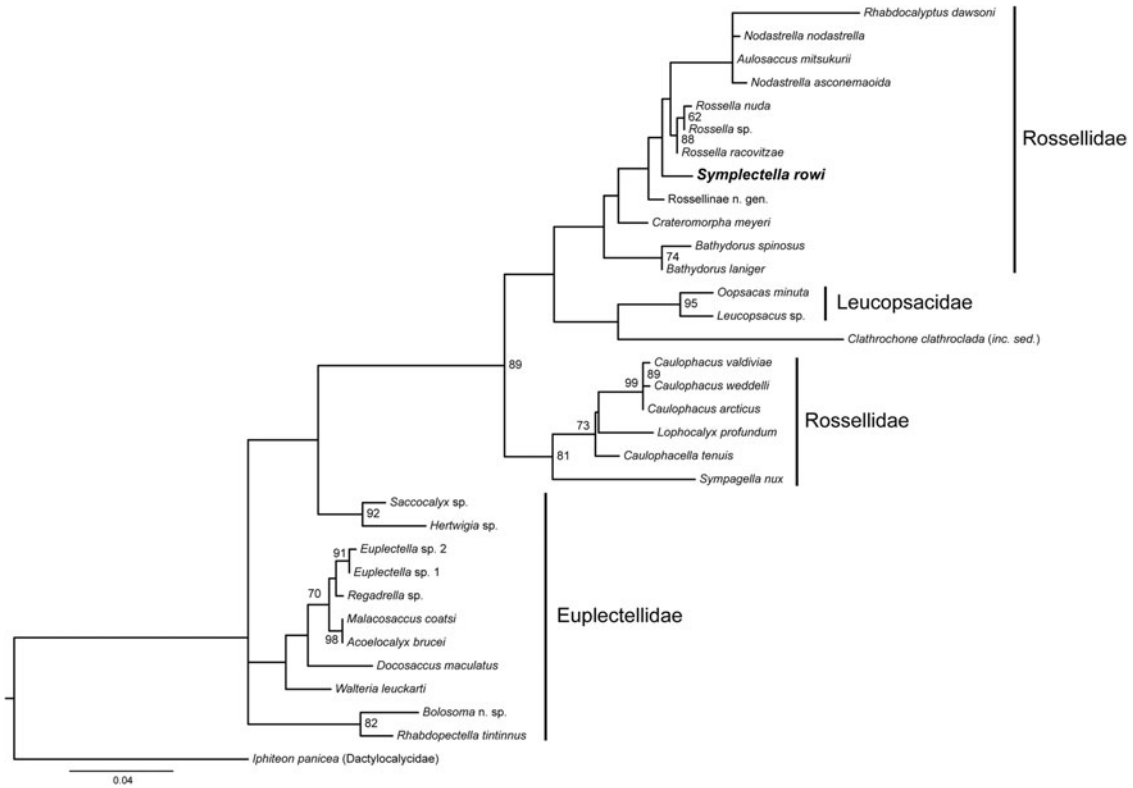

Fig. 1. Maximum-likelihood phylogeny of Lyssacinosida inferred from 16S rDNA alignment. Numbers at nodes are rapid-bootstrap support values obtained from 900 pseudoreplicates. Only values ≥70% are shown (cf. Hillis & Bull 1993). Scale bar indicates expected number of substitutions per site.

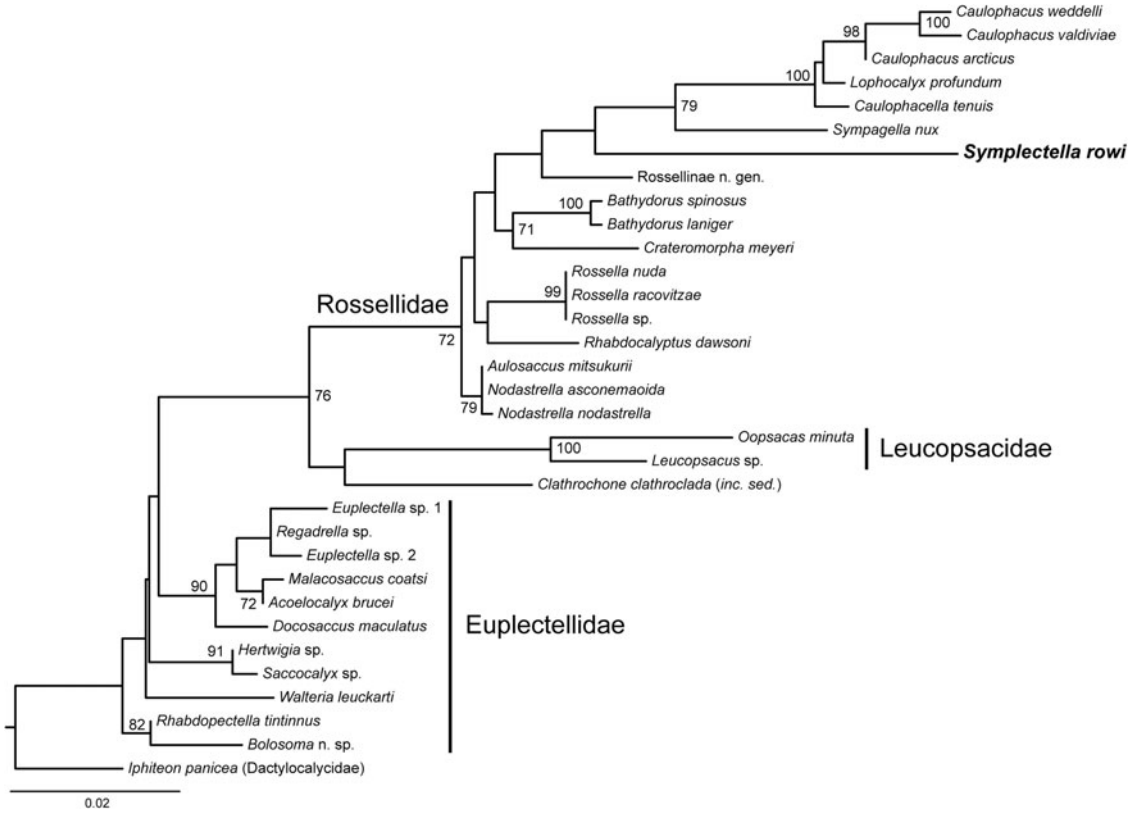

Fig. 2. Maximum-likelihood phylogeny of Lyssacinosida inferred from 28S rDNA alignment. Numbers at nodes are rapid-bootstrap support values obtained from 1000 pseudoreplicates. Only values ≥70% are shown (cf. Hillis & Bull, 1993). Scale bar indicates expected number of substitutions per site.

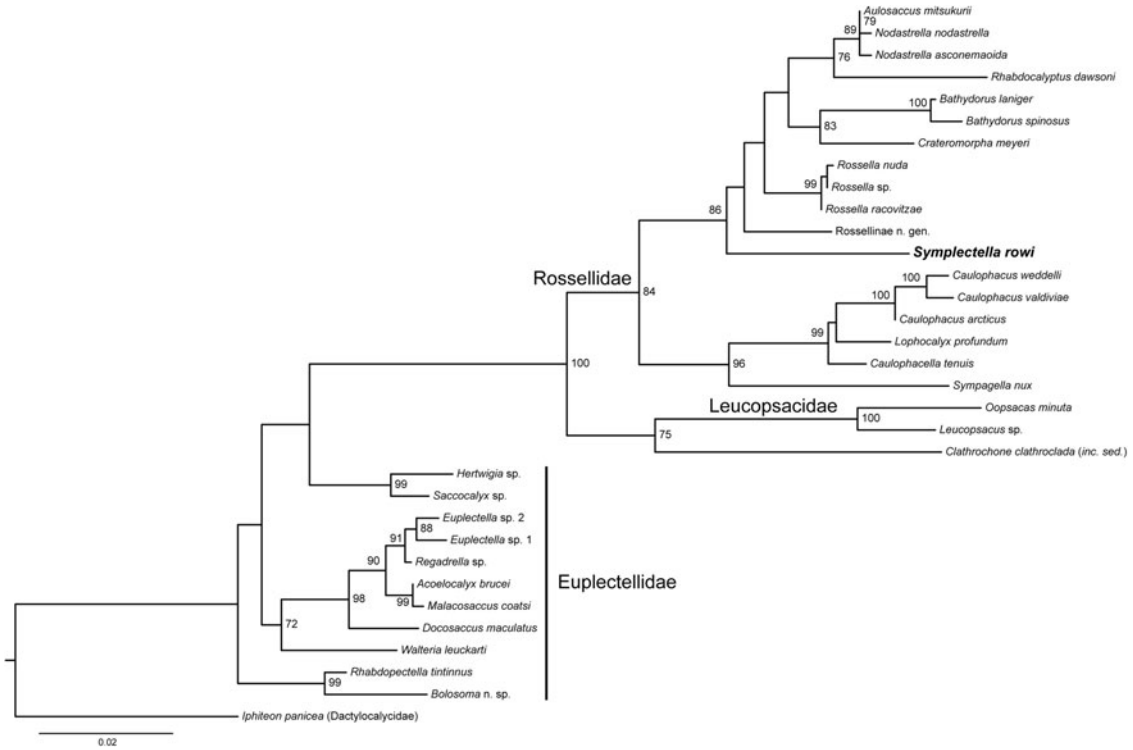

Fig. 3. Maximum-likelihood phylogeny of Lyssacinosida inferred from combined 16S + 28S rDNA alignments. Numbers at nodes are rapid-bootstrap support values obtained from 750 pseudoreplicates. Only values ≥70% are shown (cf. Hillis & Bull, 1993). Scale bar indicates expected number of substitutions per site.

https://doi.org/10.1017/S0025315414001805

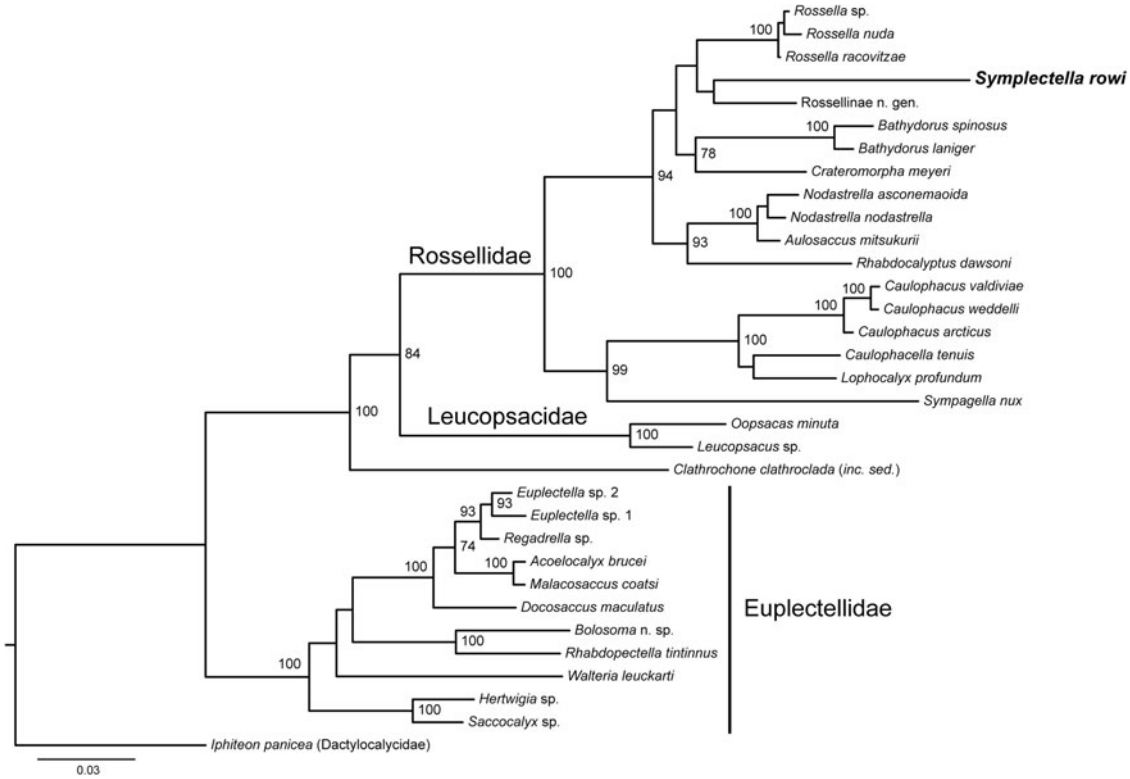

Fig. 4. Maximum-likelihood phylogeny of Lyssacinosida inferred from combined 18S + 28S + 16S rDNA + COI alignments. Numbers at nodes are rapid-bootstrap support values obtained from 250 pseudoreplicates. Only values ≥70% are shown (cf. Hillis & Bull, 1993). Scale bar indicates expected number of substitutions per site.

Caulophacella, which group with the two sampled representatives of Lanuginellinae) and *Rhabdocalyptus* (Acanthascinae); Euplectellidae excluding *Symplectella* is supported as a clade by 100% BS.

DISCUSSION

Two different hypotheses for the family assignment of the monospecific glass sponge genus *Symplectella* have been proposed in the literature: in Rossellidae (Dendy, 1924) and in Euplectellidae (Tabachnick, 2002a). The molecular phylogenetic analyses presented here clearly support the original hypothesis of Dendy (1924). Morphologically, there are no strong characters in support of Tabachnick's hypothesis (see Introduction). Thus, I here move *Symplectella* back to its original position in Rossellidae. This placement implies secondary loss of hypodermal pentactins and atrialia, as well as convergent evolution of a sieve plate in *Symplectella*, but the family diagnoses of Rossellidae and Euplectellidae (Tabachnick, 2002a, b) are sufficiently vague such that no emendations are required. Because neither strobiloplumicomes (cf. Tabachnick, 2002b) nor discoctasters (cf. Reiswig & Stone, 2013) are known from *Symplectella*, it has to be assigned to subfamily Rossellinae (which is unfortunately an artificial taxon; see Dohrmann *et al.*, 2012b). Although the sister genus of *Symplectella* could not be determined here with confidence, likely candidates are *Rossella*, *Nodastrella* or *Caulophacus*, as in these taxa calycocomes also occur (see Dohrmann *et al.*, 2012a). However, *Nodastrella* and *Caulophacus* appear to be firmly nested in well-supported clades with other genera (Figure 4), leaving only *Rossella* as a good candidate for the closest relative of *Symplectella*, a hypothesis that awaits to be tested with additional data.

ACKNOWLEDGEMENTS

I thank Allen G. Collins for hosting me at NMNH, the NMNH collection staff for providing access to the NCI collection, and the National Cancer Institute and David J. Newman for allowing the NCI natural product voucher to be used for taxonomic purposes, which was arranged within the framework of the National Science Foundation (NSF)-funded Porifera Tree of Life project. I further thank Gert Wörheide for access to computational resources at LMU and two anonymous reviewers for comments on the manuscript.

FINANCIAL SUPPORT

This research was supported by a Smithsonian Institution Postdoctoral Fellowship and grant DO 1742/1-1 of the Deutsche Forschungsgemeinschaft (DFG).

REFERENCES

Dendy A. (1924) Porifera. Part I. Non-Antarctic sponges. *Natural History Report. British Antarctic ('Terra Nova') Expedition, 1910 (Zoology)* 6, 269–392.

Dohrmann M., Collins A.G. and Wörheide G. (2009) New insights into the phylogeny of glass sponges (Porifera, Hexactinellida): monophyly of Lyssacinosida and Euplectellinae, and the phylogenetic position of Euretidae. *Molecular Phylogenetics and Evolution* 52, 257–262.

Dohrmann M., Göcke C., Reed J. and Janussen D. (2012a) Integrative taxonomy justifies a new genus, *Nodastrella* gen. nov., for North Atlantic "*Rossella*" species (Porifera: Hexactinellida: Rossellidae). *Zootaxa* 3383, 1–13.

Dohrmann M., Haen K.M., Lavrov D.V. and Wörheide G. (2012b) Molecular phylogeny of glass sponges (Porifera, Hexactinellida): increased taxon sampling and inclusion of the mitochondrial protein-coding gene, cytochrome oxidase subunit I. *Hydrobiologia* 687, 11–20.

Dohrmann M., Janussen D., Reitner J., Collins A.G. and Wörheide G. (2008) Phylogeny and evolution of glass sponges (Porifera, Hexactinellida). *Systematic Biology* 57, 388–405.

Felsenstein J. (1985) Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 39, 783–791.

Hillis D.M. and Bull J.J. (1993) An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. *Systematic Biology* 42, 182–192.

Hooper J.N.A. and Van Soest R.W.M. (2002) *Systema Porifera. A guide to the classification of sponges*. New York, NY: Plenum.

Lanave C., Preparata G., Saccone C. and Serio G. (1984) A new method for calculating evolutionary substitution rates. *Journal of Molecular Evolution* 20, 86–93.

Pattengale N.D., Alipour M., Bininda-Emonds O.R.P., Moret B.M.E. and Stamatakis A. (2010) How many bootstrap replicates are necessary? *Journal of Computational Biology* 17, 337–354.

Reiswig H.M. (2002a) Family Aulocalycidae Ijima, 1927. In Hooper J.N.A. and Van Soest R.W.M. (eds) *Systema Porifera. A guide to the classification of sponges*. New York, NY: Plenum, pp. 1362–1371.

Reiswig H.M. (2002b) Family Aphrocallistidae Gray, 1867. In Hooper J.N.A. and Van Soest R.W.M. (eds) *Systema Porifera. A guide to the classification of sponges*. New York, NY: Plenum, pp. 1282–1286.

Reiswig H.M. and Stone R.P. (2013) New glass sponges (Porifera: Hexactinellida) from deep waters of the central Aleutian Islands, Alaska. *Zootaxa* 3628, 1–64.

Savill N.J., Hoyle D.C. and Higgs P.G. (2001) RNA sequence evolution with secondary structure constraints: comparison of substitution rate models using maximum-likelihood methods. *Genetics* 157, 399–411.

Stamatakis A. (2014) RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30, 1312–1313.

Stamatakis A., Hoover P. and Rougemont J. (2008) A rapid bootstrap algorithm for the RAxML web servers. *Systematic Biology* 57, 758–771.

Tabachnick K.R. (2002a) Family Euplectellidae Gray, 1867. In Hooper J.N.A. and Van Soest R.W.M. (eds) *Systema Porifera. A guide to the classification of sponges*. New York, NY: Plenum, pp. 1388–1434.

Tabachnick K.R. (2002b) Family Rossellidae Schulze, 1885. In Hooper J.N.A. and Van Soest R.W.M. (eds) *Systema Porifera. A guide to the classification of sponges*. New York, NY: Plenum, pp. 1441–1505.

Tabachnick K.R. and Menshenina L.L. (2002) Family Hyalonematidae Gray, 1857. In Hooper J.N.A. and Van Soest R.W.M. (eds) *Systema porifera. A guide to the classification of sponges*. New York, NY: Plenum, pp. 1232–1263.

Van Soest R.W.M., Boury-Esnault N., Hooper J.N.A., Rützler K., de Voogd N.J., Alvarez de Glasby B., Hajdu E., Pisera A.B., Manconi R., Schönberg C., Janussen D., Tabachnick K.R., Klautau M., Picton B., Kelly M., Vacelet J., Dohrmann M., Díaz C. and Cárdenas P. (2014) *World Porifera Database*. http://www.marinespecies.org/porifera.

and

Yang Z. (1994) Maximum likelihood phylogenetic estimation from DNA sequences with variable rates over sites: approximate methods. *Journal of Molecular Evolution* 39, 306–314.

Correspondence should be addressed to:
M. Dohrmann
Department of Earth and Environmental Sciences, Ludwig-Maximilians-University Munich, Palaeontology and Geobiology, Molecular Geo- and Palaeobiology Lab, Richard-Wagner-Str. 10, 80333 Munich, Germany
email: m.dohrmann@lrz.uni-muenchen.de