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Large fine-scale spatiotemporal variations of CH₄ diffusive fluxes from shrimp aquaculture ponds affected by organic matter supply and aeration in Southeast China

Ping Yang^{1,2,3‡}, Yan Zhang^{1,2,3‡}, Hong Yang^{4,5,6*}, Yifei Zhang^{1,2}, Jin Xu², Lishan Tan^{1,2}, Chuan Tong^{1,2,3*}, Derrick Y.F. Lai⁷

¹Key Laboratory of Humid Subtropical Eco-geographical Process of Ministry of Education, Fujian Normal University, Fuzhou 350007, P.R. China,

²School of Geographical Sciences, Fujian Normal University, Fuzhou 350007, P.R. China,

³Research Centre of Wetlands in Subtropical Region, Fujian Normal University, Fuzhou 350007, P.R. China,

⁴College of Environmental Science and Engineering, Fujian Normal University, Fuzhou, 350007, China,

⁵Collaborative Innovation Center of Atmospheric Environment and Equipment Technology, Jiangsu Key Laboratory of Atmospheric Environment Monitoring and Pollution Control, School of Environmental Science and Engineering, Nanjing University of Information Science & Technology, 219 Ningliu Road, Nanjing 210044, China,

⁶Department of Geography and Environmental Science, University of Reading, Reading, RG6 6AB, UK,

⁷Department of Geography and Resource Management, The Chinese University of Hong Kong, Shatin, New Territories, Hong Kong SAR, China

*Correspondence to:

H. Yang (hongyanghy@gmail.com, Telephone 0044(0)1183787750) or C. Tong (tongch@fjnu.edu.cn, Telephone 086-0591-87445659)

‡ Ping Yang and Yan Zhang contributed equally to this work.

Abstract Mariculture shrimp ponds are important CH₄ sources to the atmosphere. However, the spatiotemporal variations of CH₄ concentration and flux at fine spatial scales in mariculture ponds are poorly known, particularly in China, world largest aquaculture producer. In this study, the plot-scale spatiotemporal variations of water CH₄ concentration and flux, both within and among ponds, were researched in shrimp ponds in Shanyutan wetland, Min River Estuary, Southeast China. The average water CH₄ concentration and diffusion flux across the water-air interface in the shrimp ponds over the shrimp aquaculture period varied from 2.29 ± 0.29 to 50.48 ± 20.91 μM and from 0.09 ± 0.01 to 2.32 ± 0.95 $\text{mmol m}^{-2} \text{h}^{-1}$, respectively. The CH₄ emissions from the estuarine ponds varied greatly in seasonal dynamics, with peaks in August and September, which was similar to the trend of water temperature and dissolved oxygen (DO) concentrations. There was no remarkable difference in CH₄ concentration and flux between shrimp ponds, but significantly spatiotemporal differences in CH₄ concentration and flux within the ponds. Significantly higher emissions occurred in the feeding zone, accounting for approximately 60% of total CH₄ emission flux, while much lower CH₄ emissions appeared in aeration zone, contributing 14% to total flux. This study suggests the importance of considering spatiotemporal variation in the whole-pond estimates of CH₄ concentration and flux. In light of such high spatial variation within ponds, improving aeration and feed utilization efficiency would help to mitigate CH₄ emissions from mariculture ponds.

1. Introduction

Since the beginning of the industrial revolution, greenhouse gases (GHGs) emissions produced by human activities have increased markedly (IPCC, 2013). Methane (CH₄) is an important greenhouse gas that has a much larger global warming potential than CO₂ and contributes to approximately 20% of global radiative forcing (IPCC, 2013). Global atmospheric CH₄ levels have increased from 0.7 μ atm in 1750 to 1.8 μ atm in 2015, exceeding the pre-industrial levels by about 150% (World Meteorological Organization, 2016). Worse, some projections indicate a further doubling by 2100 (Cotovicz Jr., et al., 2016; IPCC, 2013). Accurately quantifying CH₄ emission and concentration in various ecosystems provides an indispensable basis for predicting future CH₄ emissions and climate change.

Aquatic ecosystems (e.g., lakes, rivers, and reservoirs) actively process terrestrial carbon, and frequently supersaturated with CH₄ in most time (Blees et al., 2015; Diem et al., 2012; Wen et al., 2016; Yang & Flower, 2012). They are important sources to the global CH₄ budget (Bastviken et al., 2011; Tangen et al., 2016; Yang et al., 2011), and it was estimated that global inland waters emit 0.65 Pg of C (CO₂-eq) year⁻¹ in the form of CH₄ (Bastviken et al., 2011). Due to the limitation of data (Bastviken et al., 2011), the CH₄ emissions from tropical rivers have been markedly underestimated (Borges et al., 2015). Furthermore, the accurate estimate of the regional and global CH₄ budgets remains challenging also because of overlooked the role of small ponds (Holgerson, 2015; Holgerson & Raymond, 2016; Long et al., 2016). As an indispensable part of the global small ponds, some recent studies have suggested that aquaculture ponds can be indispensable CH₄ emissions (Chen et al., 2016; Hu et al., 2012; Wu et al., 2018; Yang et al., 2015b; Yuan et al., 2019). As an important part of the global small ponds, some recent studies have suggested that aquaculture ponds are indispensable CH₄ emission sources (Chen et al., 2016; Hu et al., 2012; Wu et al., 2018; Yang et al., 2015b; Yuan et al., 2019). Although some efforts have been made on characterizing CH₄ fluxes in aquaculture ponds, the number of field records of CH₄ emissions from aquaculture ponds remains very scarce as compared to those from other aquatic systems (e.g., lakes and reservoirs) (Yang et al., 2018a). More importantly, the magnitude of spatial variation in CH₄ fluxes, both within pond and between nearby ponds, is poorly understood so far, and there is a lack of integrated analysis of both spatial and temporal variations. Furthermore, the dominant pathway of CH₄ release from aquaculture ponds into atmospheric environment remains poorly documented. Detailed field studies including both the spatial and temporal dimensions are critical to better understand the variation, and to develop more accurate approaches for upscaling to whole-pond CH₄ emissions and further large-scale assessments of pond CH₄ fluxes.

China has world's largest mariculture industry, contributing more than 17% of world's mariculture volume and approximately a third of global value in 2014 (FAO, 2017). Shrimp aquaculture is one of the most important mariculture productions in China and it is widely distributed in the subtropical estuaries along the coastal regions (Yang et al., 2017a). These mariculture ponds are highly heterogeneous over time and space

owing to variations in topography, environmental factors (e.g., temperature, nutrient levels, dissolved oxygen, and others), astronomical tidal levels and other factors (Yang et al., 2018a), which may in turn lead to large uncertainties in the emission of CH₄. To improve the understanding of fine-scale spatiotemporal variation in CH₄ dynamics, and their implications for effectively upscaling pond fluxes to regional scales, this study researched fine-scale CH₄ flux dynamics across the water-atmosphere interface of shrimp ponds in Southeast China. The research aims are 1) to determine the spatial variations in CH₄ fluxes both within pond and among ponds; 2) to assess the seasonal dynamics of CH₄ flux in the shrimp ponds and main influencing factors; and 3) to determine the dominant pathway of CH₄ release from the shrimp ponds into atmospheric environment.

2. Materials and Methods

2.1. Study Area Description

This study was conducted within the central-western part of the Shanyutan wetland (26°00'36"–26°03'42"N, 119°34'12"–119°40'40"E) located in the Min River estuary (MRE) in Southeast China (Figure 1). Climate in the region is characterized by warm and wet, with a mean annual temperature of 19.6°C and a mean annual precipitation of 1350 mm (Tong et al., 2010). The tides at the wetland are typically semidiurnal, with an average range of approximately 4.5 m. The average salinity of tidal water in the Min River estuary is 4.2±2.5‰. The dominant vegetation species of the Shanyutan wetland include the native *Cyperus malaccensis* and *Phragmites australis* and the invasive *Spartina alterniflora* (smooth cordgrass). Conversion of the tidal marsh ecosystem was performed in the Shanyutan wetland of the Min River estuary in recent years, and almost all of the converted lands were used for aquaculture (Yang et al., 2017a).

2.2. Shrimp Pond System and Management

Shrimp pond is one of the dominant landscapes in the Min River estuary. Most of the ponds were converted by the complete removal of original marsh vegetation. The aquaculture period usually starts in June and ends in November, with only one single crop of shrimps being produced each year (Yang et al., 2017b). Prior to shrimp production, these ponds were filled with salt water from an adjacent estuary using a submerged pump. The water depth in these shrimp ponds ranged between 1.1 and 1.8 m during the culture period. There was no water exchange during the farming period. The shrimps were fed with commercial aquatic feed pellets containing 42% protein (Yuehai™, Guangzhou, China) twice per day at 07:00 AM and 16:00 PM (local standard time), respectively, by direct application from a small boat. In each pond, three to five 1500 W paddlewheel aerators were operated four times in 07:00–09:00, 12:00–14:00, 18:00–20:00, and 00:00–03:00 (local standard time) to improve oxygen supply. Further details about the shrimp pond system and the associated management practices can be found in Yang et al. (2017b) and Yang et al. (2018a).

The pond is divided into three zones according to microtopography feature, water depth, and management practices (Figure 1c). Zone N is a nearshore area and inhabited by the tiny minority of submerged vegetation. Zone F is a deepwater area

(ditch) used for bait feeding and it is the major area for foraging, habitating and metabolic activity of shrimps. Zone A is a shallow area (platform) used for aeration activities, and to improve ponds oxygen supply. Water depth in Zone N typically 0.3–0.5 m, for Zone F (ditch) 1.5–1.8 m, and for Zone A (platform) 0.8–1.2 m. More details about the three zones of shrimp pond can be found in [Zhang et al. \(2019\)](#). To assess the plot-scale spatiotemporal variation of CH₄ emission from shrimp aquaculture ponds, water, sediment, and gas samples were collected from three commercial shrimp ponds in Shanyutan Wetland of the MRE ([Figure 1](#)), respectively. Basic characteristics about the selected shrimp ponds in the estuary are given in [Zhang et al. \(2019\)](#).

2.3. Measurement of the CH₄ Concentration and Flux

2.3.1. CH₄ Concentration

Three transects were chosen in each pond for the measurement of water dissolved CH₄ concentration. Taking into account the shrimp grow-out cycle as well as the logistical feasibility of sampling in the shrimp farms, water samples from the shrimp ponds were collected in June and November 2017. At each pond, three sampling sites were deployed on a transect from the nearshore zone to the aeration zone in each pond (zones N, F, and A) ([Figure 1c](#)). Each whole-pond survey was completed between 10:00 and 16:00 (local standard time). Water samples for the determination of dissolved CH₄ concentration were collected in 55 mL pre-weighted serum glass bottles with silicone tubing, left to overflow, poisoned with a saturated solution of HgCl₂ (0.2 mL⁻¹), sealed with a butyl stopper, and crimped with an aluminum cap ([Abril et al., 2007](#); [Borges et al., 2017](#); [Cotovicz Jr., et al., 2016](#)). CH₄ concentration was determined using the headspace technique and a gas chromatograph. Ultrahigh purity N₂ gas (99.999%) was injected into the glass bottle to create a 25 mL headspace. The N₂ gas entered the bottle via a syringe inserted in the rubber stopper at a slight positive pressure of 50 hPa, and 25 mL of water was pushed out of the bottle via a second syringe inserted in the stopper ([Xiao et al., 2017](#)). The samples were vigorously shaken to obtain complete equilibration between air and water phases ([Cotovicz Jr., et al., 2016](#)). After waiting for 0.5 h, the headspace CH₄ concentrations were determined using gas chromatography (GC-2010, Shimadzu, Kyoto, Japan) with flame ionization detection (FID). Five different concentrations of standard CH₄ gas, namely 2, 8, 500, 1000 and 10000 ppm, were used to calibrate the FID of gas chromatograph. The detection limits for CH₄ were 0.3 ppm, and the relative standard deviations of CH₄ analyses were $\leq 2.0\%$ in 24 h. The dissolved CH₄ concentration *in situ* surface water was calculated according to a temperature and salinity-dependent Henry's law constant and accounted for CH₄ in the headspace and in the water ([Farías et al., 2017](#); [Wanninkhof, 1992](#); [Xiao et al., 2017](#)).

2.3.2. CH₄ Flux from the Transfer Coefficient Method

Transfer coefficient method (Eq.1) was used to quantify the diffusive CH₄ flux ($F_{m,d}$, mmol m⁻² h⁻¹) across the water-atmosphere interface at three transects across the ponds and in different months of the aquaculture period.

$$F_{m,d} = k(C_w - C_{eq}) \quad (1)$$

where k is the gas exchange velocity (cm h^{-1}), C_w is the measured surface water (here at the depth 20 cm) dissolved CH_4 concentration (mmol L^{-1}), and C_{eq} is the equilibrium dissolved CH_4 concentration relative to the atmospheric concentration at the prevailing *in situ* conditions (mmol L^{-1}). The gas transfer coefficient k is dependent on wind speed and is normalized to a Schmidt number of 600 (Jahne et al., 1987; MacIntyre et al., 1995; Xiao et al., 2017). The wind speed was collected from the automatic weather station of the Min River Estuary Ecological Station in the Shanyutan wetland. k values were obtained from the model described by Cole & Caraco (1998) due to that their experiment environment (considering the influence of varying wind speeds on the estimate of k value) were closest to the studied shrimp ponds.

2.3.3. CH_4 Flux from Direct Measurement Using Chamber

In order to evaluate the potential role of CH_4 ebullition flux from the shrimp ponds, total CH_4 fluxes were determined by floating chamber. On each sampling date, three plastic floating chambers were deployed on transects L1 from the nearshore zone to the aeration zone of each pond. Chambers were with an area of 0.1 m^2 and a volume of 5.2 L, and they were fitted with Styrofoam floats on their sides. They were covered with aluminum tape to minimize internal heating by sunlight. More details about the floating chambers can be found in Natchimuthu et al. (2016, 2017). Two air samples inside the chamber headspace were collected began at 9:00–11:00 AM on the 1st day and ended at the same time on the 2nd day over a 24 h period from chamber enclosure by using 60 mL plastic syringes equipped with three-way stopcocks. The samples were then immediately transferred to pre-evacuated airtight gas sampling bags (Dalian Delin Gas Packing Co., Ltd., China), transported to the laboratory, and analyzed using a gas chromatograph (GC-2010, Shimadzu, Kyoto, Japan) equipped with a FID within 24 h after sampling. The gas flux ($F_{m, t}$, $\text{mmol m}^{-2} \text{ h}^{-1}$) was calculated with the following Equation (2):

$$F_{m, t} = \frac{dn}{dt} \times \frac{1}{A} \quad (2)$$

where dn/dt is the slope of the amount of substance for CH_4 over the sampling period (mol h^{-1}) and A is the chamber area (m^2). The amounts of CH_4 in the chamber at different times were calculated using Equation (3):

$$n = (C_{\text{end}} - C_{\text{init}}) \times 10^{-6} \times \frac{P_{\text{tot}} \times V}{R \times T} \quad (3)$$

where C_{init} and C_{end} are respectively the initial and end concentration of CH_4 across the water-air interface (ppm) (usually comes from GC measurement); P_{tot} is the total air pressure (usually $\sim 1 \text{ atm} = 1013.15 \text{ hPa}$); V is the chamber volume (L); R is the common gas constant ($0.082056 \text{ L atm K}^{-1} \text{ mol}^{-1}$); and T is the absolute temperature during the gas sampling (K). Since chambers showed distinct nonlinear increases in methane concentration, this research considered the chambers captured the flux to the atmosphere including both flux by diffusion and by ebullition (bubble flux) from the shrimp ponds. Therefore, same as previous studies (Bastviken et al., 2004; Chuang et al., 2017; Keller & Stallard, 1994; Miller & Oremland, 1988; Natchimuthu et al., 2014), the contribution of ebullition was determined by comparing the flux measured

with the transfer coefficient method against the total flux measured with the floating chambers flux.

2.4. Measurement of Ancillary Variables

Meteorological data (air temperature, air pressure, wind speed, and precipitation) were obtained from the local weather stations, which provided meteorological information at a 30 min interval. During each sampling campaign, surface water temperature, pH, salinity, and dissolved oxygen (DO) concentration at 20 cm water depth were measured *in situ* in each sampling site. Water temperature and pH were measured using a handheld pH/mV/Temperature meter system (IQ150, IQ Scientific Instruments, USA) and the salinity was measured using a salinity meter (Eutech Instruments-Salt6, USA). The dissolved oxygen concentration was determined *in situ* with a multiparameter probe (550A YSI, USA) at 20 cm depth.

During each sampling campaign, surface water samples (~ 20 cm depth) were collected from the above mentioned positions from different zones by using organic glass hydrophores, and then transferred into 150 mL polyethylene bottles. Approximately 0.2 mL of saturated HgCl₂ solution was injected into each bottle to inhibit microbial activity (Zhang et al., 2013; Yang et al., 2017b). The water samples were subsequently transported to the laboratory within 4–6 h, stored in a 4 °C cooler, and analyzed within one week.

Approximately 100 mL of the water sample was filtered through a 0.45 µm filter (Biotrans™ nylon membranes) and subsequently analyzed for the concentrations of N-NO₃⁻ and total organic carbon (TOC). N-NO₃⁻ and TOC concentrations in the surface water samples were analyzed using flow injection analysis (Skalar Analytical SAN⁺⁺, Netherlands) and a total organic carbon analyzer (Schimadzu TOC-V_{CPH/CPN}, Kyoto, Japan), respectively. The detection limits for N-NO₃⁻ and TOC were 6 µg L⁻¹ and 4 µg L⁻¹, respectively. The relative standard deviations of N-NO₃⁻ and TOC analyses were ≤3.0% and ≤1.0%, respectively.

2.5. Data Analysis

The calculations of basic statistical parameters (e.g. mean, standard error (SE), and others) were carried out using SPSS 17.0 (SPSS, Inc., USA). Data transformations were performed using both the Box-Cox procedure (including CH₄ fluxes |λ = -0.12, CH₄ concentrations |λ = -0.13, wind speed |λ = 0.23, salinity |λ = -0.22 and NO₃⁻-N |λ = -0.19, and λ herein is the Box-Cox exponent) and log transformation (TOC) to ensure *a priori* that the assumptions for the analyses of variance and the linear model analysis were not violated. Significance tests were calculated based on the transformed data, while untransformed data are used to plot the figures. Two-way analysis of variance (ANOVA) was conducted to analyze the effects of sampling zones (nearshore zone, feeding zone, and aeration zone), culturing time, and their interactions on the CH₄ diffusion fluxes (or CH₄ concentration) in shrimp ponds with ponds specified as the random term. Linear mixed effects models accounting the pond random effect were also fitted to explore the relationships between environmental variables and the CH₄ diffusion fluxes (or CH₄ concentration) using the nlme package of R (Bates et al., 2014; Holgerson, 2015). The stepAIC() function in the R package “MASS” was used for the model selection (Ripley et al., 2016). The model with the

lowest AIC value was chosen, and the relationship between the dependent variables and chosen predictors was further tested by Type II Wald's test implemented in the R package "car" (Fox et al., 2018). To test whether there is a significant random effect, this study used the rand function in the R package "lmerTest" (Kuznetsova et al., 2017) to perform a likelihood ratio test. The Chi square statistics and the corresponding p-values of this test were implemented in the 2-way ANOVA table, and this kind of statistics thus indicate whether the variation in the CH₄ diffusion fluxes (or CH₄ concentration) were dependent on the random pond selection. Other analyses and graphics were conducted with SPSS 17.0 (SPSS, Inc., USA) and OriginPro 7.5 (OriginLab Corporation, USA), respectively. The results were considered significant at the 0.05 significance level and summarized as "mean ± standard error". The concentration and diffusion fluxes of CH₄ data from the three shrimp ponds were interpolated for mapping by using Inverse Distance Weighted method (IDW) in ArcGIS 10.2 (ESRI Inc., Redlands, CA, USA).

3. Results

3.1. Meteorological Parameters and Surface Water Quality

The variations in meteorological parameters from the shrimp ponds during the aquaculture period are shown in Figure 2. Atmospheric pressure in the estuarine ponds showed an increasing trend with time (Figure 2a), and the differences were more than 35 hPa ($p < 0.05$, ranged from 985 to 1020 hPa). Wind speed (Figure 2a) and air temperature (Figure 2c) from the estuarine ponds varied greatly between months, with considerably higher values from August to October. Meteorological parameters showed insignificantly spatial changes inside and between ponds ($p > 0.05$).

The temporal variations in water quality parameters from the shrimp ponds during the aquaculture period are shown in Figure 3. Water temperature displayed obvious temporal changes, and the mean values changed from 18.11°C (in November) to 34.35°C (in August) (Figure 3a). The other five water quality parameters, namely pH, DO, salinity, N-NO₃⁻, and TOC, also showed prominently temporal changes (Figure 3b-f). Water pH and DO in the shrimp ponds over the study period ranged from 9.11 to 10.01, and from 12.17 to 18.84 mg L⁻¹, respectively, with lower pH in July ($p < 0.05$; Figure 3b), and lower DO in September and October ($p < 0.05$; Figure 3c). The mean salinity and N-NO₃⁻ concentrations were lower in July and August compared with other months ($p < 0.05$; Figure 3d and 3e). The TOC concentrations of the shrimp ponds was generally between 11.40 and 41.59 mg L⁻¹ with lower TOC in July and higher value in October ($p < 0.05$; Figure 3f).

The spatial variations in water quality parameters within- and between-ponds during the aquaculture period are shown in Figure S1. Most of the measured water quality variables, namely temperature, pH, salinity, N-NO₃⁻ and TOC, did not differ significantly between the sampling sites within-ponds ($p > 0.05$; Figure S1). The differences of mean values of the water quality parameters between three ponds were insignificantly ($p > 0.05$; Figure S1). However, significant differences in DO concentrations were observed between the sampling sites within ponds ($p < 0.05$; Figure S1E), and the average DO concentrations in the Zone F were generally higher

than those in the Zones N and A.

3.2. Spatial Variation in CH₄ Concentration and Diffusion Flux

3.2.1. Within-Pond Variation

Surface water CH₄ concentration from the estuarine shrimp ponds changed considerably between different zones within ponds (Figure 4 and Figure S2). Across all sampling ponds, mean CH₄ concentrations ranged from 2.19 ± 0.28 to 18.69 ± 4.17 μM , 4.28 ± 0.67 to 88.82 ± 17.69 μM , and 1.64 ± 0.15 to 9.12 ± 2.96 μM , in the Zones N, F and A, respectively, with average values of 7.84 ± 1.11 , 33.09 ± 6.07 and 4.01 ± 0.67 μM . The CH₄ diffusion flux also showed very large spatial variations across the different zones within ponds (Figure 5 and Figure S2A), ranging from 0.09 ± 0.01 to 0.86 ± 0.20 $\text{mmol m}^{-2} \text{h}^{-1}$, 0.17 ± 0.02 to 4.04 ± 1.19 $\text{mmol m}^{-2} \text{h}^{-1}$, and 0.06 ± 0.01 to 0.44 ± 0.14 $\text{mmol m}^{-2} \text{h}^{-1}$ in the Zones N, F and A, respectively. Over the study period, the Zone F was hot spot of CH₄ emission (1.69 ± 0.33 $\text{mmol m}^{-2} \text{h}^{-1}$), followed by the Zone N (0.34 ± 0.05 $\text{mmol m}^{-2} \text{h}^{-1}$) and Zone A (0.19 ± 0.03 $\text{mmol m}^{-2} \text{h}^{-1}$) (Figure S2A). There were significant differences in pond water CH₄ concentration and fluxes between different zones within ponds ($p < 0.01$; Figure S2A and Table 1).

3.2.2. Between-Pond Variation

Across all sampling months and sites, the mean CH₄ concentrations were 18.81 ± 4.79 μM , 11.65 ± 3.67 μM , and 14.48 ± 3.33 μM in Ponds I, II and III, respectively (Figure 4). The overall median and mean from all ponds were 4.57 and 14.98 μM . CH₄ concentrations were supersaturated across all ponds during the aquaculture period, indicating that aquaculture ponds are CH₄ emission source (Figure 5). The mean CH₄ emission fluxes were 0.95 ± 0.25 $\text{mmol m}^{-2} \text{h}^{-1}$, 0.58 ± 0.21 $\text{mmol m}^{-2} \text{h}^{-1}$, and 0.70 ± 0.17 $\text{mmol m}^{-2} \text{h}^{-1}$ in Ponds I, II and III, respectively (Figure 5). The overall median and mean of CH₄ fluxes from all three ponds were 0.19 and 0.74 $\text{mmol m}^{-2} \text{h}^{-1}$, respectively. The CH₄ concentrations and flux in Pond I were largest, followed by Pond III and Pond II (Figures 4 and 5), and there was significant difference in CH₄ concentrations and flux between ponds ($p < 0.05$; Table 1).

3.3. Temporal Variation in CH₄ Concentration and Diffusion Flux

CH₄ concentration and diffusion flux in three shrimp ponds showed similar temporal patterns, with the highest CH₄ concentration and flux generally in August and September, and the lowest flux always in June and November (Figures 5 and 6). When averaging the monthly concentrations (or fluxes) over three ponds, a strong temporal pattern in CH₄ concentrations (or fluxes) emerged, with the minimum in June (2.71 ± 0.33 μM and 0.11 ± 0.01 $\text{mmol m}^{-2} \text{h}^{-1}$), the maximum in September (38.88 ± 9.13 μM and 1.76 ± 0.41 $\text{mmol m}^{-2} \text{h}^{-1}$), and generally low values in November (4.89 ± 0.64 μM and 0.17 ± 0.02 $\text{mmol m}^{-2} \text{h}^{-1}$). According to the AIC-based model selection, monthly CH₄ concentration / flux (temporal variations) in the estuarine ponds were best predicted by dissolved oxygen (DO), atmospheric pressure and salinity / pH (Table 2).

4. Discussions

4.1. Role of Dissolved Oxygen and Organic Matter

Large spatial variation in CH₄ concentration and flux at small spatial scales (e.g., within system, and between systems) has been reported in rivers (Crawford et al., 2017; Zhao et al., 2013), reservoirs (Musenze et al., 2014; Zhao et al., 2013), and lakes (Borges et al., 2011; Chuang et al., 2017; Natchimuthu et al., 2016; Schrier-Uijl et al., 2011; Xiao et al., 2017; Xing et al., 2005; Yang et al., 2008). Many of these studies have attributed the spatial variation of CH₄ concentration and flux to direct or indirect effects of primary productivity, nutrient status (e.g., organic carbon), meteorology, and morphometry (e.g., area and depth) (e.g., Chuang et al., 2017; Holgerson et al., 2015; Natchimuthu et al., 2016; Schrier-Uijl et al., 2011; Xiao et al., 2017; Xing et al., 2006). To the best of our knowledge, such information is limited for aquaculture ponds. In the current research, the small spatial scales variations in CH₄ concentration and flux across the shrimp ponds were analyzed. An interesting finding of this study was CH₄ concentration (or fluxes) differed significantly both within pond and among ponds (Table 1).

The average CH₄ concentration and emission flux in Pond-I was significantly higher than those in Pond-II and Pond-III (Figures 4 and 5; $p < 0.01$). The spatial variability of CH₄ dynamics might be related to the physical and chemical parameters of sediment / water differed in their magnitude among the three ponds, which influence sediment CH₄ production. Among the several environmental variables of the study (Figure S1), only water N-NO₃⁻ concentration differed significantly among ponds ($p < 0.01$), and the average concentration followed the orders: Pond-II ($99.7 \pm 15.7 \mu\text{g L}^{-1}$) > Pond-III ($50.3 \pm 5.1 \mu\text{g L}^{-1}$) > Pond-I ($22.9 \pm 3.1 \mu\text{g L}^{-1}$). The spatial patterns of N-NO₃⁻ concentration and CH₄ dynamics in the MRE ponds were largely similar. Previous research has shown that some microorganisms preferentially use N-NO₃⁻ as an alternative electron acceptor to oxidize organic substrates (such as acetate) in anaerobic environments (Hu et al., 2017; Nykänen et al., 2002), thereby outcompeting methanogens and inhibiting methanogenesis. Therefore, it is considered that high CH₄ emission flux occurred in Pond-I and low flux occurred in Pond-II, to some extent, were dependent on the difference in N-NO₃⁻ concentration between ponds.

Net CH₄ release rate in aquatic ecosystems is determined by the production of methanogens, consumption by methanotrophs, and transport processes, which are essentially affected by a series of biotic and abiotic parameters. The role of DO in methane dynamic has been evaluated in various aquatic ecosystems (e.g., Liu et al., 2015; Schrier-Uijl et al., 2011; Xiao et al., 2017; Yang et al., 2015a). High water DO concentration would promote CH₄ oxidation at the sediment-water interface or during the passway in transportation but inhibit methanogens (Liu et al., 2015; Schrier-Uijl et al., 2011), ultimately resulting in a lower water CH₄ concentration and the subsequent emission (Xiao et al., 2017). This study found the Zone F with the smallest surface water DO (Figure S1E) and largest CH₄ concentration (Figure S2A) and diffusion flux (Figure S2B). Also, CH₄ concentration and flux significantly and negatively correlated with DO concentrations ($p < 0.05$; Figures 6 and 7). These results suggested that the DO variations could be one of possible reasons for the difference in CH₄ concentration and flux among the three zones within ponds in our study site.

In addition to DO, sediment total carbon (TC) content (P. Yang, unpublished data) and

water TOC concentration differ markedly between the three zones within ponds. Sediment TC and water TOC (Figure S1G) in the Zone F were largest, followed by the Zones N and A, which was similar to the spatial distribution of water CH₄ concentration and release flux (Figure S2). This indicates that organic matters (e.g., bait) was also a variable causing spatial variations in CH₄ flux inside the pond. It is well known that CH₄ in aquatic ecosystem is mainly generated from sediments containing organic matters (e.g., Bastviken et al., 2008; Grinham et al., 2018; Xiao et al., 2017). Large organic matter loading in sediment not only fuels CH₄ production, but also increases oxygen consumption, which suppresses CH₄ oxidation (Huttunen et al., 2003; Xiao et al., 2017). Consequently, large amounts of CH₄ was produced in the feeding zone and emitted into water and atmosphere. These finding highlights pond aeration (DO) and organic matter supply play an important role in the large spatial variation in CH₄ concentration and flux within pond.

4.2. Factors Influencing the Temporal Variations of CH₄ Flux

At the month scale, the mean CH₄ concentration and diffusion flux in the three ponds showed considerable variation (Figures 4 and 5). Overall, higher CH₄ concentration and flux occurred in August and September and lower value appeared in June and November. Markedly temporal variations in CH₄ concentration / flux have been reported in lakes (Natchimuthu et al., 2016; Xiao et al., 2017), rivers (Borges et al., 2018; Zhao, et al., 2013), shallow ponds (Holgerson, 2015; Yang et al., 2018a), coastal and continental shelf zones (e.g., Borges et al., 2018; Cunada et al., 2018; Gü l zow et al., 2014; Jakobs et al., 2014; Sierra et al., 2017). Most of these studies have related the seasonal patterns of CH₄ with variation in temperature (e.g., Borges, et al., 2017; Natchimuthu et al., 2016; Sierra et al., 2017; Xiao et al., 2017; Yang et al., 2018a; Zhao, et al., 2013), particularly the increase in sediment CH₄ production rates in response to the increasing temperature (Vizza et al., 2017; Yang et al., 2018a; Yvon-Durocher et al., 2014). In addition, some studies found that the seasonal variation of CH₄ could be governed by the changes in DO concentrations in aquatic systems (Holgerson, 2015; Hu et al., 2018; Zhao, et al., 2013). Generally, when DO concentration is low in water, the methanogenic (anaerobic bacteria) activity increases and the CH₄ oxidation capacity declines, which leads to the increase in sediment CH₄ production and subsequent emission (Hu et al., 2018; Ivanov et al., 2002). According to the AIC-based model selection, CH₄ emission fluxes were best predicted by a negative relationship with DO (Table 2), indicating that DO level was also play a major role in influencing the temporal variation in CH₄ emissions from the aquaculture ponds in subtropical estuaries.

Salinity is an important environmental factor governing CH₄ dynamics in coastal areas (Tong et al., 2010; Vizza et al., 2017). On one hand, salinity allows the occurrence of sulfate-reduction that leads to enhanced anaerobic oxidation of CH₄ in sediments, and strong competition of sulfate-reducers with methanogens (Vizza et al., 2017). On the other hand, high salinity induces ion (e.g., Cl⁻ and Fe³⁺) stress (Chambers et al., 2013; Neubauer et al., 2013) or harm to methanogens (Sun et al., 2013), with the consequence of reducing sediment CH₄ production. Many studies found CH₄ emission fluxes from coastal wetlands and aquatic ecosystems decreases

with the increase in salinity (e.g., Poffenbarger et al., 2011; Vizza et al., 2017; Welti et al., 2017; Wilson et al., 2015; Yang et al., 2018a). Similarly, the significantly lower water salinity between August and September in this study (Figure 3d) could significantly increase sediment CH₄ production owing to the enhanced methanogenic activities. Thus, the higher concentration and flux of CH₄ in the August and September was likely due to the higher CH₄ production rates supported by lower water salinity. Although CH₄ production data are unavailable in the present study, the CH₄ concentration in the ponds showed a significantly negative relationship with water salinity ($p < 0.05$; Table 2), which supports the above hypothesis. Further studies merit to explore the exact impacts of salinity on CH₄ production and emission.

Previous studies show that low pressures may facilitate the transport of CH₄ from sediments to the atmosphere and reduce the amount of time available for methane oxidation (Chen et al., 2014; Natchimuthu et al., 2014, 2016; Zhu et al., 2016). This study observed high CH₄ concentration and flux in the August and September and low values in the June and November, which were opposite to the trend of atmospheric pressure ($p < 0.05$; Table 2). The results showed that the seasonal variation of ponds CH₄ dynamics in the aquaculture ponds also could be governed by the changes in atmospheric pressure.

In addition, this study also found a significantly positive relationship between monthly CH₄ flux and pH ($p < 0.01$; Table 2). Methanogens are pH sensitive and grow best around pH 6–8 in wetland and aquatic systems (Chang and Yang, 2003; Hu et al., 2017; Yang et al., 2017b). Hence CH₄ flux is expected to decrease as pH values move away from the optimal range of 6–8 (e.g., Hu et al., 2017; Le Mer and Roger, 2001; Schrier-Uijl et al., 2011). The positive relationship observed between CH₄ flux and pH in this study could be related, at least in part, to the influence of primary production. A higher primary production will enhance the uptake of CO₂ in the water column, which in turn increase water pH and alkalinity (Gruca-Rokosz et al., 2017; Portielje and Lijklema, 1995). At the same time, a higher primary production can increase the supply of organic matter to pond sediments, thereby reducing soil redox potential and stimulating methanogenic activities. Further studies should be done using controlled experiments to examine whether pH exerts a direct influence on CH₄ emissions.

4.3. Diffusion isn't a Major Pathway of CH₄ Emission in Mariculture Pond

Anoxic sediment is “hot spot” of methane production in aquatic ecosystems. Methane can be exported from the sediment through molecule diffusion, ebullition (bubbles), or combination of them (Bastviken et al., 2004; Hu et al., 2016). Ebullition is often considered as the main CH₄ emission pathway in reservoirs, rivers and lakes (e.g., Bastviken et al., 2004; Chuang et al., 2017; Deshmukh et al., 2014; Natchimuthu et al., 2016; Rodriguez and Casper, 2018; Xiao et al., 2017). However, such information from aquaculture ponds is still very limited (Yang et al., 2017a). In the current research, the contribution of ebullition was estimated by comparing the diffusion flux measured with the transfer coefficient method against the total flux measured with the floating chambers (Bastviken et al., 2004; Chuang et al., 2017; Keller and Stallard, 1994; Miller and Oremland, 1988; Natchimuthu et al., 2014). The average CH₄

diffusion flux and total flux ranged from 0.11 to 1.76 mmol m⁻² h⁻¹ and 0.18 to 8.52 mmol m⁻² h⁻¹ (Figure S3), respectively, with mean values of 0.74 ± 0.30 and 3.86 ± 1.38 mmol m⁻² h⁻¹, respectively. Consequently, the average CH₄ ebullition flux ranged from 0.01 to 6.98 mmol m⁻² h⁻¹, with mean values of 3.12 ± 1.21 mmol m⁻² h⁻¹ (Figure S3). Ebullition emission comprises over 70% (ranged 5.0 to 96.3%) of the total CH₄ flux. In spite of limited number of floating chambers, our results clearly show that CH₄ emission was dominated by ebullition. In the meantime, our results highlight that diffusion isn't the main CH₄ emission pathway in aquaculture ponds. Given the episodic nature of ebullition (Xiao et al., 2017), obviously more fine-scale temporal and spatial measurements data are needed to increase the accuracy in the flux estimate.

4.4. Implications of CH₄ Emission Flux from Aquaculture Ponds

4.4.1. Implications of CH₄ Flux Spatiotemporal Variations

Our results highlight that subtropical aquaculture ponds are large atmospheric CH₄ sources with strong spatial variability. The large spatial variation of CH₄ flux within ponds (Figure S2) implies a large uncertainty of whole-pond CH₄ fluxes budgets estimated by earlier studies that based on single or limited number of site measurements (e.g., Hu et al., 2016; Ma et al., 2018; Yang et al., 2017a; Yang et al., 2018a). For a more accurate estimate of whole pond CH₄ emissions, it is of paramount importance to take into account of measurements from a number of strategically located sites that would adequately capture a representative areal extent of the emitting surface. The markedly spatial variation in CH₄ fluxes means that extrapolation of a few ponds' measurements during regional CH₄ budgeting should be done cautiously. Similarly, the significant temporal variation of pond CH₄ fluxes (Figure 5) implies the large uncertainty during extrapolating a single month CH₄ emission measurement to annual emissions. Therefore, it is very important to measure from as many sites as practicable over a number of months in order to reduce the uncertainty of CH₄ flux estimations and improve our understanding of CH₄ dynamics in aquaculture ponds.

4.4.2. Implications of Large CH₄ Emission Flux

An earlier study estimated that GHGs (CO₂ and CH₄) efflux from mariculture ponds across the subtropical estuaries of China would be equivalent to ~15% of the net carbon emissions from the terrestrial natural ecosystems in China (Yang et al., 2018a). It is worth noting that the CH₄ emissions fluxes in subtropical estuarine aquaculture ponds were substantially higher than those from the freshwater aquaculture systems (e.g., Da Silva et al., 2018; Hu et al., 2014; Hu et al., 2016; Liu et al., 2015; Wu et al., 2018) and were one to three orders of magnitude larger than those observed in most reservoirs and lakes (e.g., Gerardo-Nieto et al., 2017; Huttunen et al., 2003; Musenze et al., 2014; Natchimuthu et al., 2016; Wen et al., 2016; Xiao et al., 2017; Zhao et al., 2013; Zhu et al., 2010). CH₄ diffusion fluxes in our ponds were also substantially higher than those in coastal aquatic ecosystems (Sierra et al., 2017), and was approximately 8 times higher than the average of 0.09 mmol m⁻² h⁻¹ found in China's natural wetlands (Wei and Wang, 2017). Moreover, the magnitude of CH₄ emissions observed in our ponds were much larger than those from the estuarine brackish

Cyperus malaccensis marsh (ranged from 0.04 to 0.32 mmol m⁻² h⁻¹) (Yang et al., 2019). These results suggest that subtropical estuarine aquaculture ponds could be important sources of atmospheric CH₄, which could play an important yet overlooked role in regional and global CH₄ budgets.

It is a big challenge to balance the economic development and environmental protection (Yang, 2014), for example seafood production and CH₄ mitigation from subtropical estuarine aquaculture ponds. This study found that the Zone F was hot spot of CH₄ emission, followed by the Zone N and A (Figure 5 and Figure S2), which accounting for approximately 60%, 26% and 14% of total pond CH₄ emission fluxes, respectively. Aquaculture ponds are generally maintained through daily feed supply to produce aquatic animals (Chen et al., 2015, 2016). However, only a small portion of the feed input is actually converted into shrimp biomass, with the feed utilization efficiency of ~4.0%–27.4% (Chen et al., 2015; Molnar et al., 2013). Most of the feed input remains in aquaculture systems. Thus large CH₄ emission fluxes occurred the feeding zone, to a large extent, were dependent on the plentiful supply of organic matter from residual feed and faeces, which are more favorable for the majority of CH₄ production. These findings indicate that improving feed utilization efficiency, reducing organic matter (e.g., residual feed and faeces) accumulation on the bottom of ponds feeding zone, and increasing the area of aeration activities might be important strategies to mitigate CH₄ emissions from aquaculture ponds.

4.5. Limitation and Future Research

Similar as many studies, there are some limitations in the current study. CH₄ measurement and estimation were conducted in one estuary during the aquaculture period (from June to November) in the present study. Significantly spatiotemporal variations in CH₄ fluxes at various sites in different shrimp ponds have been found in our study. Obviously, future research should increase the frequency of *in situ* sampling and include more innovative techniques to measure CH₄ flux in aquaculture ponds at multiple estuaries. Moreover, our study did not thoroughly quantify event-driven CH₄ exchange, such as the effect of weather conditions, particularly the extreme weather (e.g., typhoon), on water-atmosphere CH₄ fluxes. Many previous studies have found a large amount of ebullition coinciding with a low atmosphere pressure (Casper et al., 2000; Chen et al., 2014; Mattson & Likens, 1990; Natchimuthu et al., 2015, 2016). Thereby many low pressure induced CH₄ flux events were likely missed, in turn generating underestimated CH₄ fluxes (Natchimuthu et al., 2016). Event-driven CH₄ fluxes during the sampling period should be further investigated. Furthermore, CH₄ fluxes in the aquatic systems varied greatly in diurnal cycle (e.g., Erkkilä et al., 2018; Hirota et al., 2007; Natchimuthu et al., 2014; Xing et al., 2004). Most of the research found that high CH₄ flux generally occurred during the daytime and low flux occurred during the nighttime (e.g., Bastviken et al., 2004; Hirota et al., 2007; Natchimuthu et al., 2014; Xing et al., 2004). In spite of no direct measurement of day-night pattern of CH₄ fluxes in the current study, a similar diurnal pattern can happen in aquaculture ponds. Therefore, increase in sampling frequency in the further studies can improve the estimate accuracy of CH₄ fluxes at diurnal temporal scales before upscaling them to calculate seasonal and annual flux.

Significant amount of CH₄ ebullition fluxes in shrimp ponds have been found in our study, which are consistent with those of previous studies in shallow and nutrient-rich ponds (e.g., [Holgerson, 2015](#); [Natchimuthu et al., 2014](#)). However, the accuracy of our estimates was eroded due to the CH₄ ebullition fluxes was estimated by comparing the diffusion flux measured with the transfer coefficient method against the total flux measured with the floating chambers. In the future work, therefore, there is an urgent need for utilizing advancing technologies to directly measure the CH₄ ebullition in aquaculture ponds.

5. Conclusions

CH₄ concentrations were supersaturated across all ponds and all sampling dates, indicating that aquaculture shrimp ponds were important CH₄ emitters to the atmosphere. CH₄ emissions from the estuarine ponds varied greatly between months, reaching a peak in August and September, which was similar to the trend of temperature and water DO concentrations. Duplicate CH₄ measurements at various sites within ponds yielded new insights into the spatial variations of CH₄ concentration and emission flux. The patterns clearly show that the common single-point is not representative for estimating whole-pond CH₄ emissions. The integrated assessment of both spatial (at various sites within pond) and temporal variations in this study showed that it is important to measure from as many sites as practicable over a number of months to improve the accuracy of whole-pond CH₄ flux estimates. Mariculture shrimp ponds in the subtropical estuaries are large sources of atmospheric CH₄. The high spatial CH₄ flux variation within ponds implies better aeration and higher feed utilization efficiency would help to mitigate CH₄ emissions from mariculture ponds.

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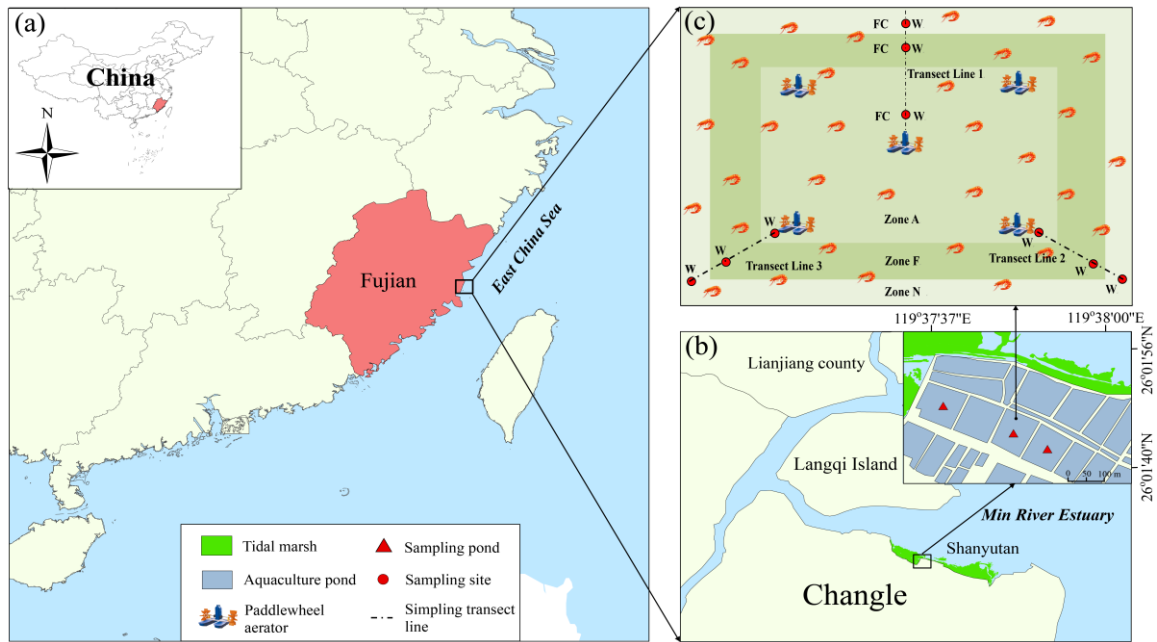


Figure 1. Location of the study area (a) and sampling sites (b) in Shanyutan wetland of Min River estuary. Design of aquaculture shrimp pond and the location of spatial sampling sites (red dots) (c). Zone N, F and A were nearshore area, feeding area and aeration area, respectively. W, water concentration samples; and FC, air samples collection by floating chambers.

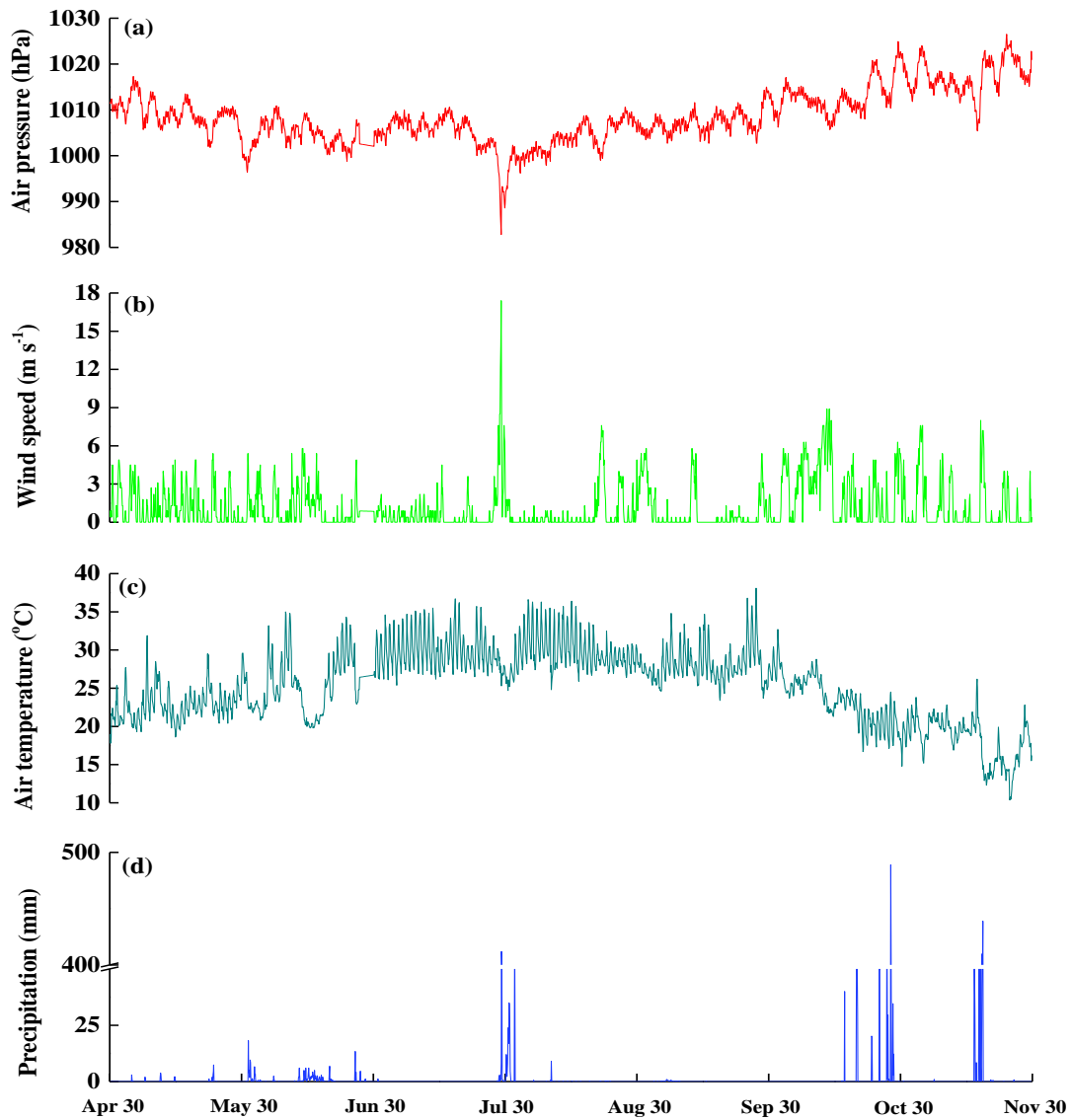


Figure 2. Temporal variation of the (a) air pressure, (b) wind speed, (c) air temperature, and (d) precipitation in the shrimp ponds at the Min River estuary during the aquaculture period (from June to November). Bars represent mean \pm SE ($n = 3$ ponds).

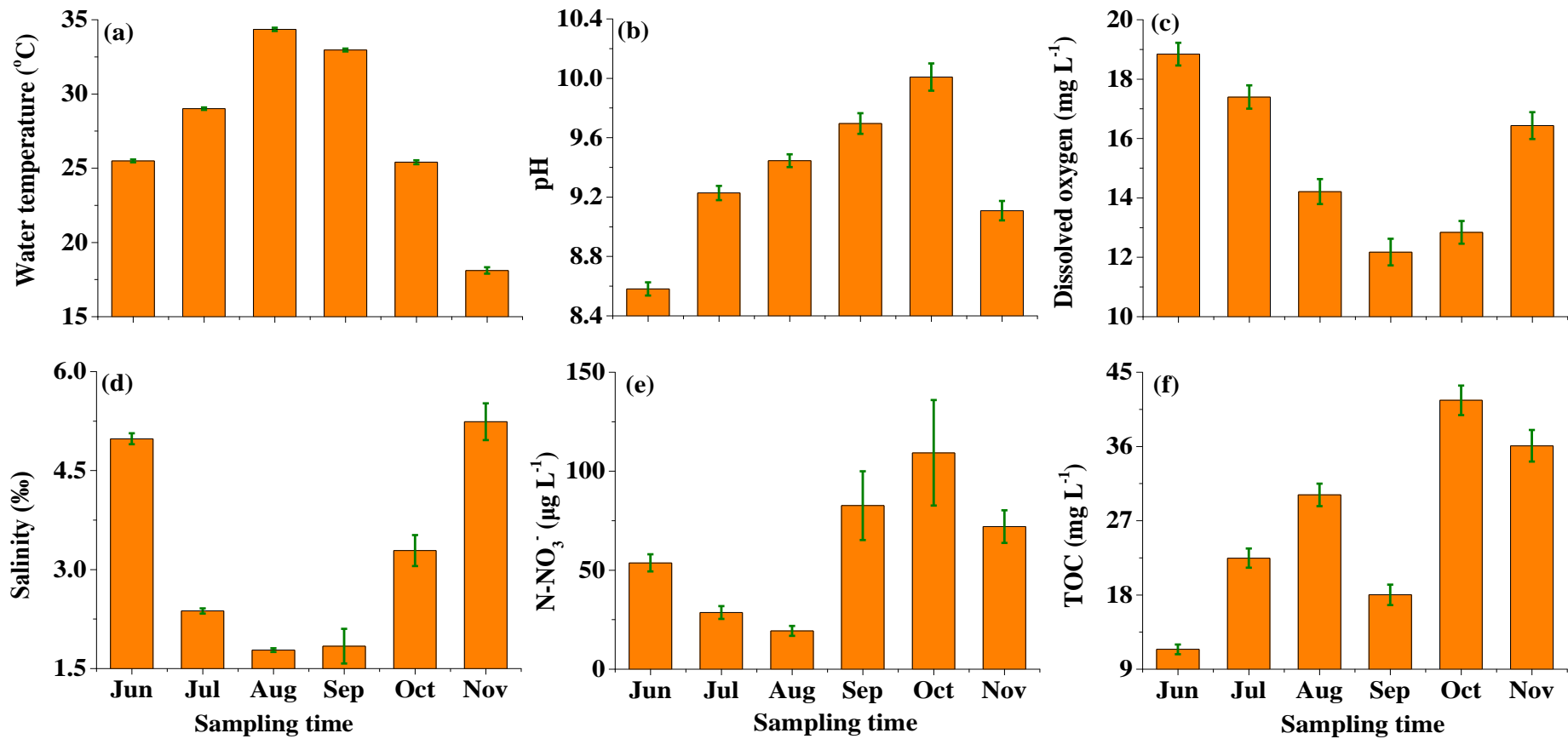
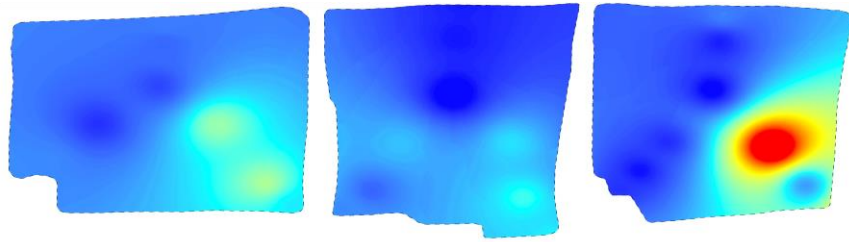
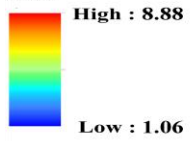


Figure 3. Temporal variation of (a) temperature, (b) pH, (c) dissolved oxygen (DO), (d) salinity, (e) N-NO₃⁻, and (f) total organic carbon (TOC) in the surface water (20 cm depth) of shrimp ponds at the Min River estuary during the aquaculture period (from June to November). Bars represent mean±SE (*n* = 3 ponds).

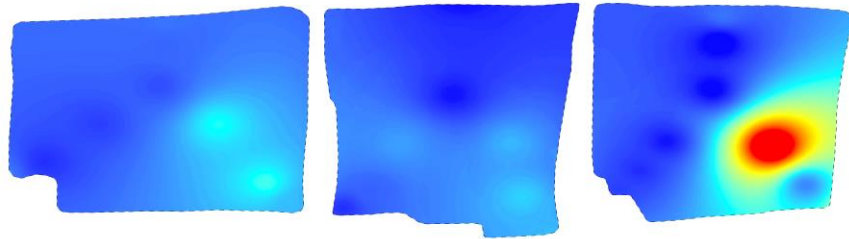
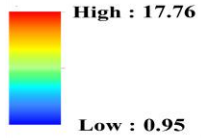
a June

CH₄ Concentrations (μM)



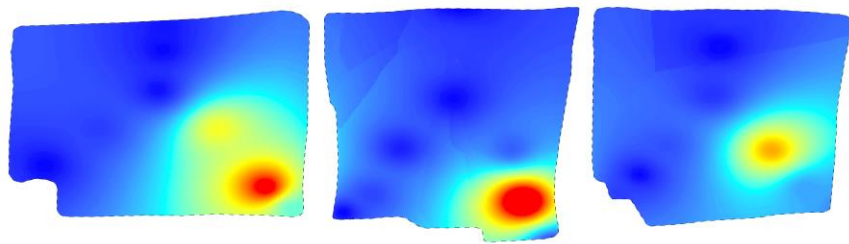
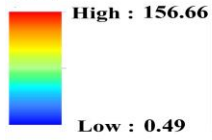
b July

CH₄ Concentrations (μM)



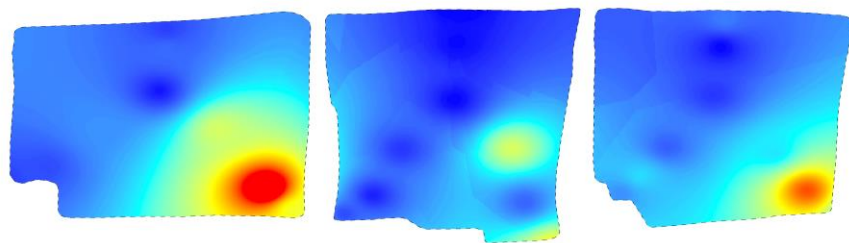
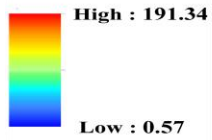
c August

CH₄ Concentrations (μM)



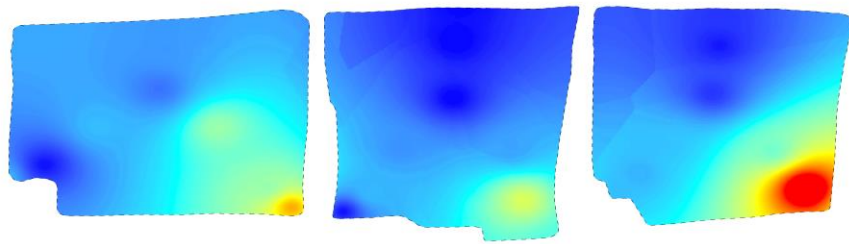
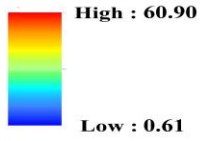
d September

CH₄ Concentrations (μM)



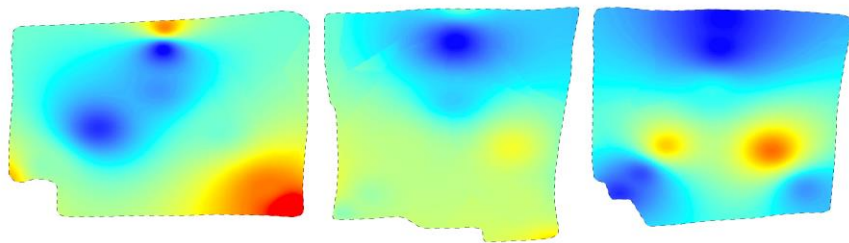
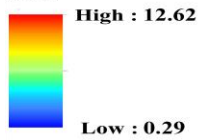
e October

CH₄ Concentrations (μM)



f November

CH₄ Concentrations (μM)



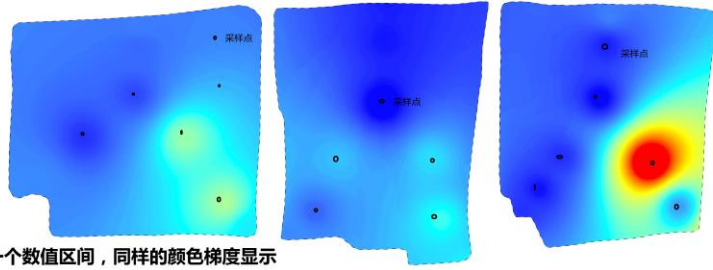
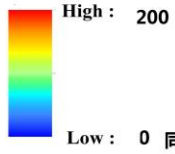
Pond I

Pond II

Pond III

a June

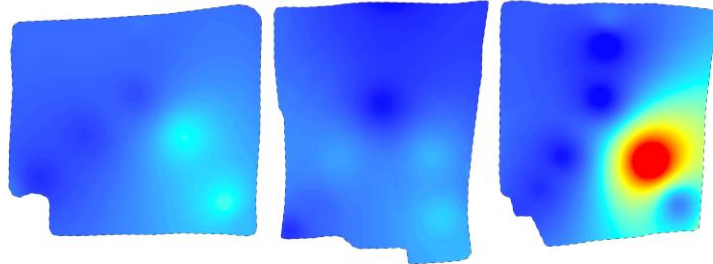
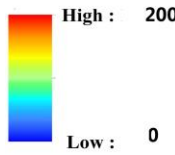
CH₄ Concentrations (μM)



同一个数值区间, 同样的颜色梯度显示

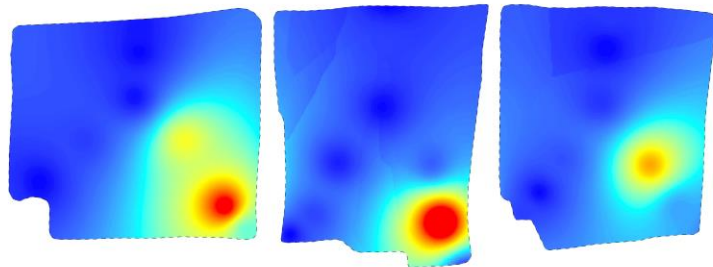
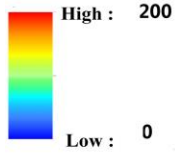
b July

CH₄ Concentrations (μM)



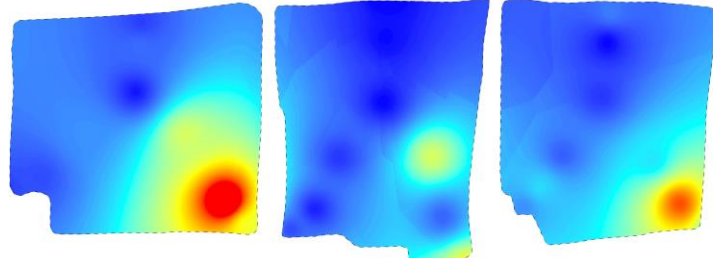
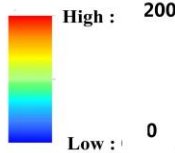
c August

CH₄ Concentrations (μM)



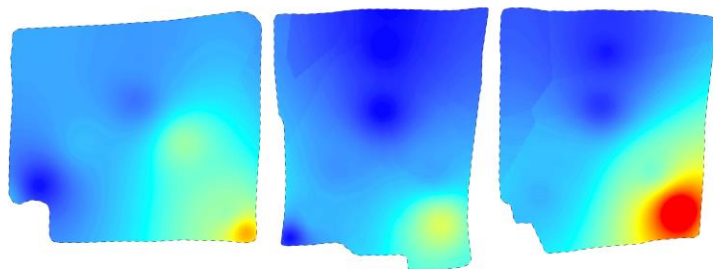
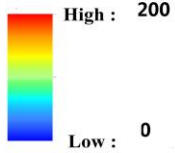
d September

CH₄ Concentrations (μM)



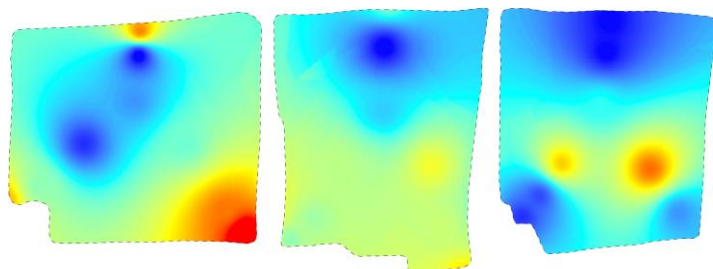
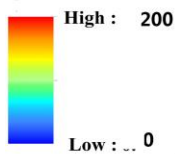
e October

CH₄ Concentrations (μM)



f November

CH₄ Concentrations (μM)



Pond I

Pond II

Pond III

Figure 4. Spatial variation of the CH₄ concentration in the surface water (20 cm depth) of shrimp ponds at the Min River estuary during the aquaculture period (from June to November).

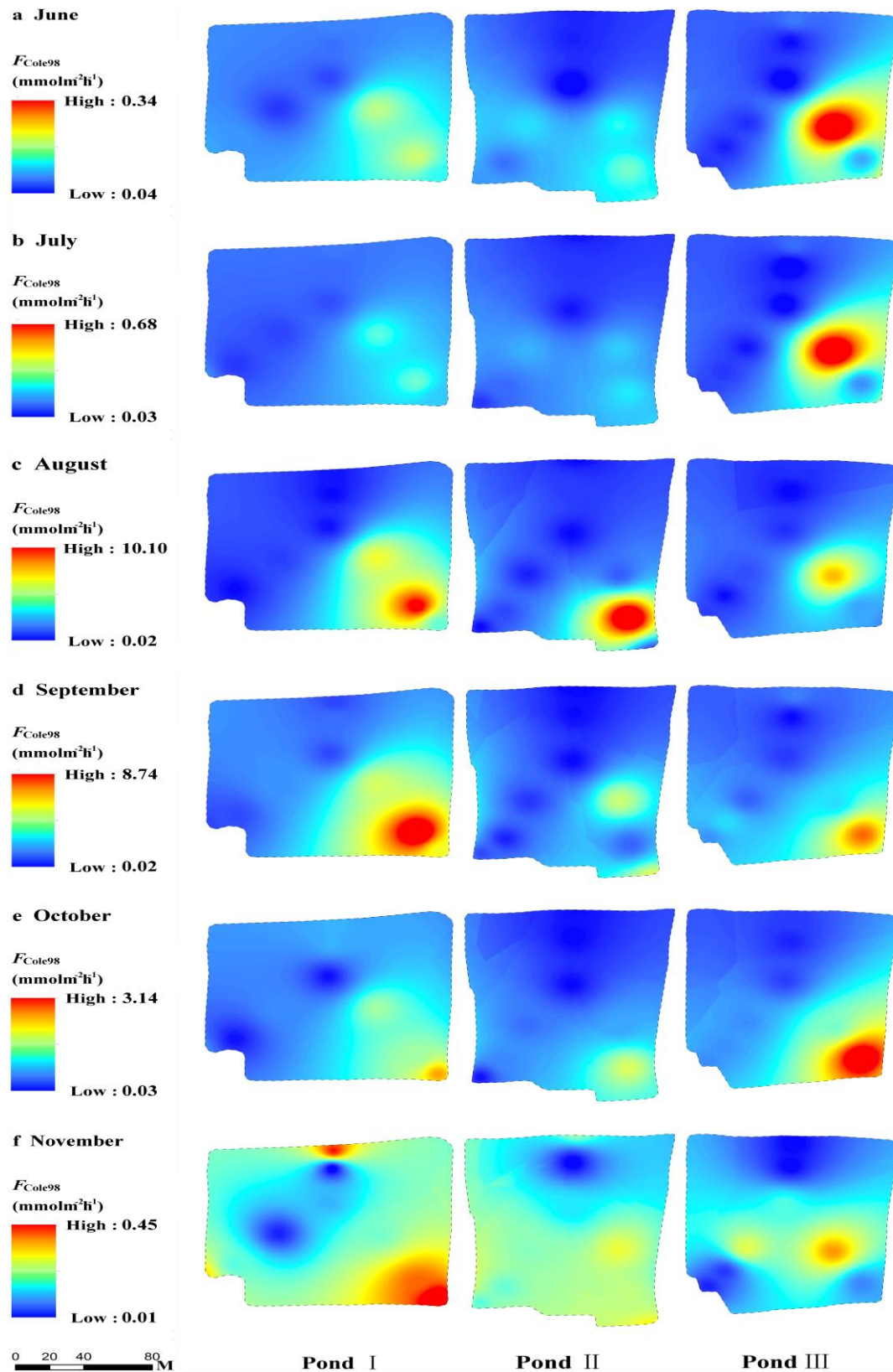


Figure 5. Spatial variation of the CH₄ diffusion flux in the surface water (20 cm depth) of shrimp ponds at the Min River estuary during the aquaculture period (from June to

November)

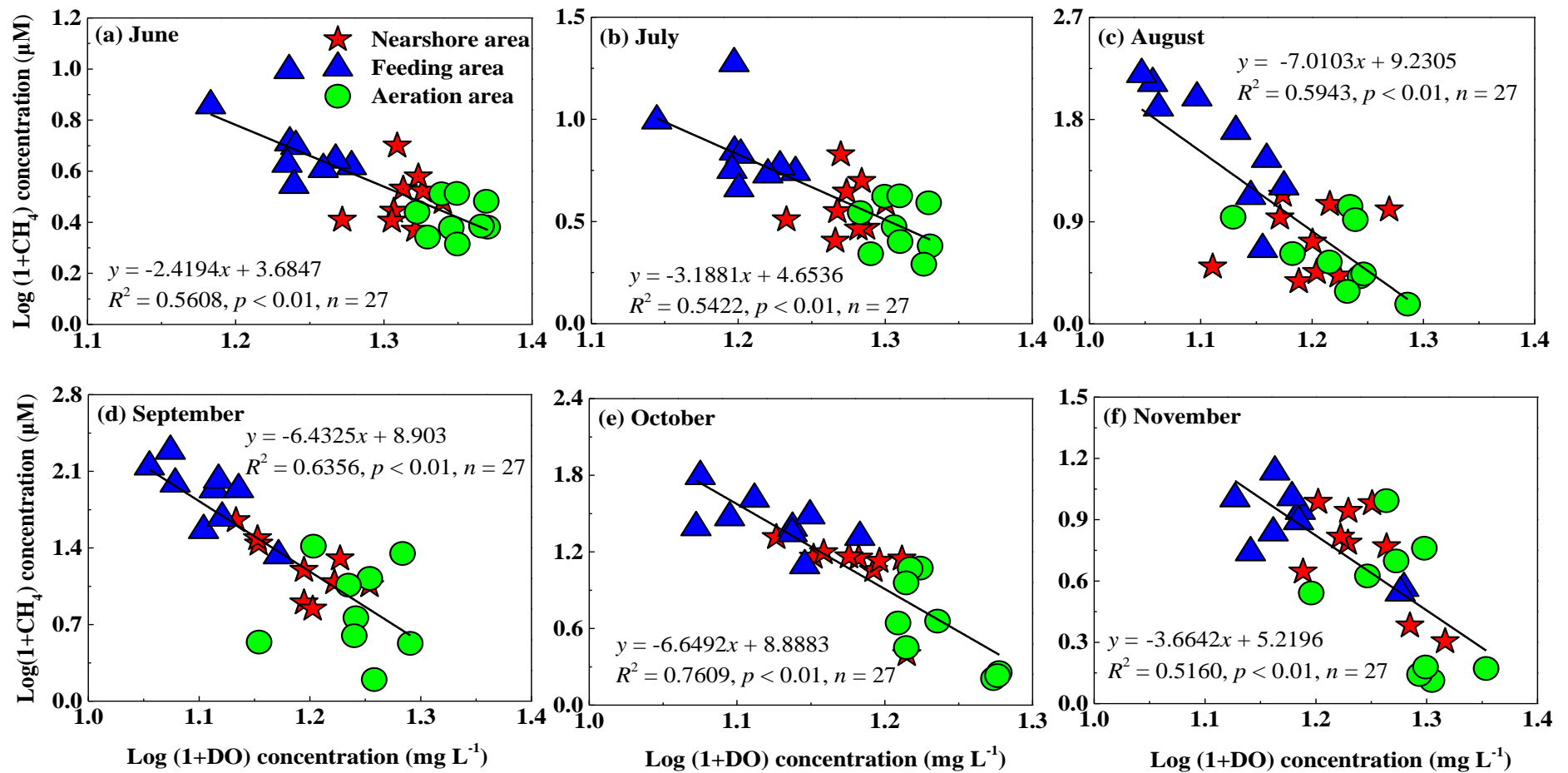


Figure 6. Relationship between the CH_4 concentrations and dissolved oxygen (DO) concentration in the surface water (20 cm depth) at shrimp ponds in the Min River estuary during each sampling campaign. Parameter bounds on the regression coefficients are 95% confidence limits.

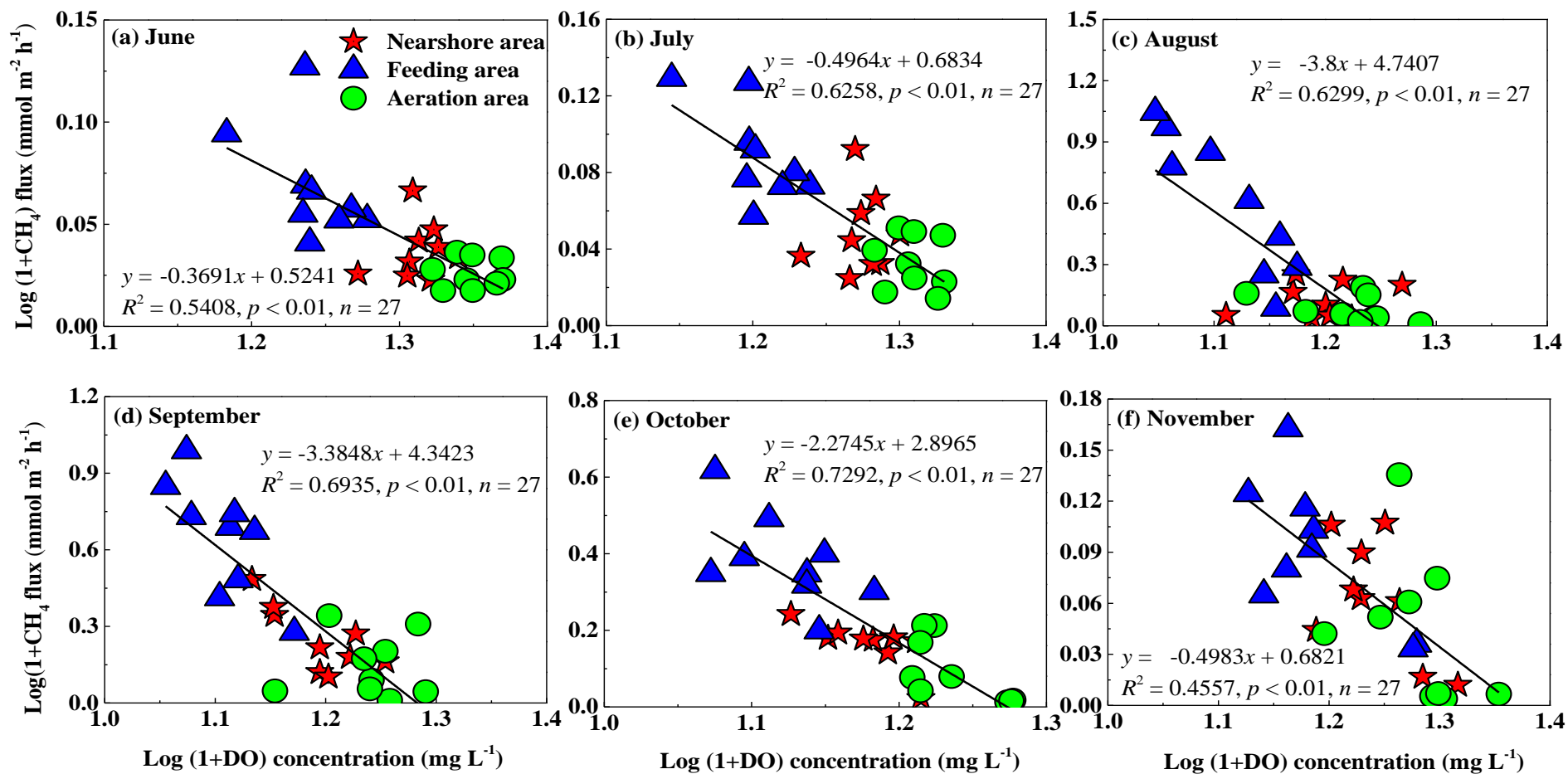


Figure 7. Linear relationship between CH_4 diffusion flux and dissolved oxygen (DO) concentration in the surface water (20 cm depth) at shrimp ponds in the Min River estuary during each campaign. Parameter bounds on the regression coefficients are 95% confidence limits.

Table 1. Summary of two-way ANOVAs (with ponds ID specified as the random term) that examining the effect of sampling zones, sampling time (months) and their interactions on water CH₄ concentration and on CH₄ fluxes at the shrimp ponds in the Min River Estuary.

	Water CH ₄ concentration					CH ₄ fluxes across the water-air				
<i>Fixed effect</i>	<i>Sum of squares</i>	<i>Mean square</i>	<i>df</i>	<i>F values</i>	<i>P values</i>	<i>Sum of squares</i>	<i>Mean square</i>	<i>df</i>	<i>F values</i>	<i>P values</i>
Zones	55.89	27.946	2	69.167	<0.001	126.59	63.29	2	65.281	<0.001
Months	45.66	9.132	5	22.603	<0.001	146.67	29.33	5	30.255	<0.001
Zones × Months	8.72	0.872	10	2.159	0.0236	14.61	1.46	10	1.507	0.142
Residuals	57.37	0.404	142			137.68	0.97	142		
<i>Random effect</i>			<i>df</i>	<i>Chi-square</i>	<i>P values</i>			<i>df</i>	<i>Chi-square</i>	<i>P values</i>
Ponds			1	7.680	0.0055			1	7.828	0.0051

Table 2. Summary of linear mixed-effect models fitted for water CH₄ concentration and CH₄ fluxes across the water-air interface. Models are ranked in order of the lowest Akaike information criterion corrected for low samples sizes (AICc) along with delta AICc. The predictors of the best model with lowest AICc were tested by Type II Wald test and the significant positive (↑) or negative effects (↓) of chosen continuous predictors are indicated.

	AICc	Variab
Water CH₄ concentration		
Air temperature+Atmospheric pressure+Water temperature+Wind speed+pH+DO+TOC+Salinity+NO ₃ ⁻ -N	388.94	-Air te
Atmospheric pressure+Water temperature+Wind speed+pH+DO+TOC+Salinity+NO ₃ ⁻ -N	382.31	-Water
Atmospheric pressure+Wind speed+pH+DO+TOC+Salinity+NO ₃ ⁻ -N	375.89	-Wind
Atmospheric pressure+pH+DO+TOC+Salinity+NO ₃ ⁻ -N	369.75	-TOC
Atmospheric pressure+pH+DO+Salinity+NO ₃ ⁻ -N	364.51	-NO ₃ ⁻ -
Atmospheric pressure+pH+DO+Salinity	359.45	-pH
Atmospheric pressure+DO+Salinity	358.91	
<i>Predictors from best model tested</i>		
DO (↓)		
Atmospheric pressure (↓)		
Salinity (↓)		
CH₄ fluxes across the water-air		
Air temperature+Atmospheric pressure+Water temperature+Wind speed+pH+DO+TOC+Salinity+NO ₃ ⁻ -N	519.57	-Air te
Atmospheric pressure+Water temperature+Wind speed+pH+DO+TOC+Salinity+NO ₃ ⁻ -N	513.06	-Wind
Atmospheric pressure+Water temperature+pH+DO+TOC+Salinity+NO ₃ ⁻ -N	506.78	-Water
Atmospheric pressure+pH+DO+TOC+Salinity+NO ₃ ⁻ -N	501.65	-NO ₃ ⁻ -
Atmospheric pressure+pH+DO+TOC+Salinity	495.90	-TOC
Atmospheric pressure+pH+DO+Salinity	490.45	-Salini
Atmospheric pressure+pH+DO	490.18	
<i>Predictors from best model tested</i>		
DO (↓)		
Atmospheric pressure (↓)		
pH (↑)		