

Update on Sequencing the Sorghum Genome

Toward Sequencing the Sorghum Genome. A U.S. National Science Foundation-Sponsored Workshop Report^[w]

Sorghum Genomics Planning Workshop Participants¹

Members of the worldwide sorghum (*Sorghum* spp.) community, including private sector and international

¹ Stephen Kresovich, Brad Barbazuk, Joseph A. Bedell, Andrew Borrell, C. Robin Buell, John Burke, Sandra Clifton, Marie-Michèle Cordonnier-Pratt, Stan Cox, Jeff Dahlberg, John Erpelding, Theresa M. Fulton, Bob Fulton, Lucinda Fulton, Alan R. Gingle, C. Tom Hash, Yinghua Huang, David Jordan, Patricia E. Klein, Robert R. Klein, Jurandir Magalhaes, Richard McCombie, Paul Moore, John E. Mullet, Peggy Ozias-Akins, Andrew H. Paterson*, Kay Porter, Lee Pratt, Bruce Roe, William Rooney, Patrick S. Schnable, David M. Stelly, Mitchell Tuinstra, Doreen Ware, and Ujwala Warek.

Donald Danforth Plant Science Center, St. Louis, Missouri 63132 (B.B.); Orion Genomics, St. Louis, Missouri 63108 (J.A.B., U.W.); Department of Primary Industries and Fisheries, Brisbane, Queensland 4350, Australia (A.B., D.J.); The Institute for Genomic Research, Rockville, Maryland 20850 (C.R.B.); Plant Stress and Germplasm Development Research Unit, United States Department of Agriculture Agricultural Research Service, Lubbock, Texas 79415 (J.B.); Genome Sequencing Center, Washington University, St. Louis, Missouri 63110 (S. Clifton, B.F., L.F.); Laboratory for Genomics and Bioinformatics, Department of Plant Biology, University of Georgia, Athens, Georgia 30602-7271 (M.-M.C.-P., L.P.); The Land Institute, Salina, Kansas 67401 (S. Cox); National Grain Sorghum Producers, Lubbock, Texas 79403 (J.D.); Tropical Agriculture Research Station, United States Department of Agriculture Agricultural Research Service, Mayaguez, Puerto Rico 00680-5470 (J.E.); Institute for Genomic Diversity, Cornell University, Ithaca, New York 14853 (S.K., T.M.F.); Plant Genome Mapping Laboratory, University of Georgia, Athens, Georgia 30602-6810 (A.R.G., A.H.P.); ICRISAT, Patancheru, AP 502 324, India (C.T.H.); United States Department of Agriculture Agricultural Research Service, Oklahoma State University, Stillwater, Oklahoma 74075-2714 (Y.H.); Institute for Plant Genomics and Biotechnology, Texas A&M University, College Station, Texas 77843 (J.E.M., P.E.K.); United States Department of Agriculture Agricultural Research Service, Texas A&M University, College Station, Texas 77843 (R.R.K.); EMBRAPA Maize and Sorghum, 35701-970, Sete Lagoas-MG, Brazil (J.M.); Cold Spring Harbor Laboratory, Cold Spring Harbor, New York 11724 (R.M., D.W.); United States Department of Agriculture Agricultural Research Service, Pacific Basin Agricultural Research Center, Aiea, Hawaii 96701 (P.M.); Department of Horticulture, University of Georgia, Coastal Plain Experiment Station, Tifton, Georgia 31793 (P.O.-A.); Pioneer HiBred International, Plainview, Texas 79072 (K.P.); Department of Chemistry and Biochemistry, University of Oklahoma, Norman, Oklahoma 73072 (B.R.); Department of Soil and Crop Sciences, Texas A&M University, College Station, Texas 77843 (W.R., D.M.S.); Center for Plant Genomics, Iowa State University, Ames, Iowa 50011 (P.S.S.); and Department of Agronomy, Kansas State University, Manhattan, Kansas 66506 (M.T.)

* Corresponding author; e-mail paterson@uga.edu; fax 706-583-0160.

^[w] The online version of this article contains Web-only data.
www.plantphysiol.org/cgi/doi/10.1104/pp.105.065136.

scientists as well as community representatives from closely related crops such as sugarcane (*Saccharum* spp.) and maize (*Zea mays*), met in St. Louis, Missouri, on November 9, 2004, to lay the groundwork for future advances in sorghum genomics and, in particular, to coordinate plans for sequencing of the sorghum genome. Key developments that made this workshop timely included advances in knowledge of the sorghum genome that provide for the development of a genetically anchored physical map to guide sequence assembly and annotation, the growing role of the sorghum genome as a nucleation point for comparative genomics of diverse tropical grasses including many leading crops, and the need for dramatically increased sorghum production to sustain human populations in many regions where its inherent abiotic stress tolerance makes it an essential staple. This report reviews current knowledge of the sorghum genome, a community-endorsed schema for integrating this knowledge into a finished sequence, and early plans for translating the sequence into sustained advances to benefit a worldwide group of stakeholders.

WHAT ARE SOME OF THE UNIQUE CONTRIBUTIONS TO BIOLOGY AND AGRICULTURE THAT WILL RESULT FROM SEQUENCING OF THE SORGHUM GENOME?

Sorghum (*Sorghum bicolor* L. Moench) is one of the world's leading cereal crops, providing food, feed, fiber, fuel, and chemical/biofuels feedstocks across a range of environments and production systems. Worldwide, sorghum is the fifth most important cereal crop (<http://apps.fao.org/default.jsp>). Its remarkable ability to produce a crop under adverse conditions, in particular with much less water than most other grain crops, makes sorghum an important "failsafe" source of food, feed, fiber, and fuel in the global agroecosystem. For example, in arid countries of northeast Africa such as Sudan, sorghum contributes about 39% of the calories in the human diet (<http://www.fao.org>, 1999 statistics). Increased demand for limited fresh water supplies, coupled with global climatic trends and expanding populations, suggests that "dryland" crops such as sorghum will be of increasing importance.

As a model organism for tropical grasses that carry out "C₄" photosynthesis, sorghum is a logical complement to the C₃ grass *Oryza* (rice), the first monocot

plant with a fully sequenced genome. Sorghum is more closely related than rice to major crops of tropical origin such as maize, sugarcane, and pearl millet (*Pennisetum typhoides*), and thus provides a better roadmap for study of these crops at the DNA level. C_4 plants contribute disproportionately to global primary productivity, in part because of biochemical and morphological specializations that result in more efficient carbon assimilation and water use at high temperatures and in water-limited environments. Anchoring of the sorghum maps to those of rice (Paterson et al., 1995, 2004), maize (Whitkus et al., 1992; Bowers et al., 2003), sugarcane (Dufour et al., 1997; Ming et al., 1998), millet (Jessup et al., 2003), switchgrass (*Panicum virgatum*; Missaoui et al., 2005), Bermuda grass (*Cynodon dactylon*; C. Bethel and A.H. Paterson, unpublished data), and others provides for the cross-utilization of results to simultaneously advance knowledge of many important crops.

The most recent whole-genome duplication in sorghum was about 70 million years ago, prior to the divergence of the major cereals from a common ancestor (Paterson et al., 2004). As such, sorghum functional genomics enjoys important advantages relative to organisms that retain duplicated copies of larger numbers of genes, such as maize, which reduplicated about 12 million years ago (Swigonova et al., 2004), and sugarcane, which has independently reduplicated at least twice in the 5 to 10 million years since its divergence from sorghum (Ming et al., 1998).

The Sorghum genus is also noteworthy because it includes one of the world's most noxious weeds, Johnson grass (*Sorghum halepense*). The rapid dispersal, high growth rate, and durability that make Johnson grass such a troublesome weed are actually desirable in many forage, turf, and biomass crops that are genetically complex. Therefore, sorghum offers novel learning opportunities relevant to weed biology as well as to improvement of a wide range of other forage, turf, and biomass crops.

Finally, sorghum fills a key gap in biogeography, with the African origin of *S. bicolor* complementing the Asian origin of rice, American origin of maize, and Middle Eastern origin of the Triticeae (wheat, barley, and others). Sorghum has made unique contributions to understanding the genetic basis of cereal domestication, which appears to have occurred independently on these different continents by the imposition of many parallel selective pressures to divergent taxa (Paterson et al., 1995). In addition, a finished sorghum sequence will be valuable for determination of the provenance of differences between rice and maize in sequence repertoire and organization. Specifically, phylogenetic "triangulation" using parsimony-based approaches to compare sorghum, maize, and rice will permit one to infer whether polymorphisms among these species are recent (for example, specific to maize), or ancient (for example, shared by maize and sorghum but differing from rice).

WHAT IS THE NATURE AND ORGANIZATION OF THE SORGHUM GENOME?

Estimates of the physical size of the sorghum genome range from 700 Mb based on Cot analysis (Peterson et al., 2002) to 772 Mb based on flow cytometry (Arumuganathan and Earle, 1991). This makes the sorghum genome about 60% larger than that of rice, but only about one-fourth the size of the genomes of maize or human. GC content is estimated at 37.7% (Peterson et al., 2002). Because sorghum is a predominantly self-pollinated plant, most genotypes are homozygous, including each of the three genotypes for which extensive genetic maps, bacterial artificial chromosome (BAC) resources, and physical maps have been constructed.

DNA renaturation kinetic analysis (Peterson et al., 2002) shows the sorghum genome to be comprised of about 16% foldback DNA, 15% highly repetitive DNA (with individual families occurring at an average of 5,200 copies per genome), 41% middle-repetitive DNA (average 72 copies), and 24% low-copy DNA. About 4% of the DNA remained single stranded at very high Cot values and is assumed to have been damaged (thus, the other percentages are slight underestimates).

Building on a rich history of genetics research supported by a wide range of sources, recent National Science Foundation (NSF)-funded activities have significantly advanced current knowledge of the sorghum genome. High-density maps of one intraspecific *S. bicolor* (Klein et al., 2000; Menz et al., 2002) and one interspecific *S. bicolor* × *Sorghum propinquum* (Chittenden et al., 1994; Bowers et al., 2003) cross provide about 2,600 sequence-tagged sites (based on low-copy probes that have been sequenced), 2,454 amplified fragment length polymorphisms, and approximately 1,375 sequence-scanned (based on sequences of genetically anchored BAC clones) loci. More than 800 markers mapped in sorghum are derived from other taxa (hence serve as comparative anchors), and additional sorghum markers have been mapped directly in other taxa or can be plotted based on sequence similarity. The two maps share one common parent (*S. bicolor* BTx623) and are essentially colinear (F. Feltus, G. Hart, K. Schertz, A. Casa, S. Kresovich, P. Klein, P. Brown, and A.H. Paterson, unpublished data). Recent cytological characterization of the individual sorghum chromosomes has provided a generally accepted numbering system (Kim et al., 2005).

The small size of the sorghum genome facilitates its use as a tropical grass model. While the maize and sugarcane genomes are similar in size to the human genome, the sorghum genome is approximately 75% smaller, variously estimated at 690 to 760 Mb. BAC libraries are available for BTx623 (about 12× coverage from *Hind*III and 8× from *Bam*HI), *S. propinquum* (13–14× coverage from *Eco*RI [approximately 7×] and *Hind*III [approximately 7×]) and IS3620C (approximately 9× coverage from *Hind*III). A total of 69,545 agarose-based fingerprints from BTx623 BACs are also

anchored with 139,434 hybridization loci from 5,147 probes (about 2,000 of which are genetically mapped). In parallel, 40,957 agarose-based fingerprints from *S. propinquum* are anchored with 148,758 hybridization loci from 5,683 probes (2,000 genetically mapped). Each of these has been assembled into WebFPC-accessible physical maps (<http://www.stardaddy.uga.edu/fpc/bicolor/WebAGCoL/WebFPC/> and <http://www.stardaddy.uga.edu/fpc/propinquum/WebAGCoL/WebFPC/>). Additional resources include 20,000 high information content fingerprint (HICF) fingerprints (from genetically mapped contigs) and six-dimensional BAC pools (5× deep) from BTx623, and 10,000 HICF fingerprints and six-dimensional BAC pools (5× deep) from IS3620C. Targeted HICF of additional contig-terminal BACs is in progress to fill gaps. About 456 *S. propinquum* and 303 *S. bicolor* BAC contigs (41% of BACs, 80% of single-copy loci) appear to be well anchored to euchromatic regions, with the percentage of the genome attributable to euchromatin likely to rise appreciably with additional anchoring. The finding that 41% of BACs are already anchored to euchromatin while only 24% of the sorghum genomic DNA is single or low copy (with an overall kinetic complexity of 1.64×10^8 ; Peterson et al., 2002) suggests that euchromatin includes a mixture of low-copy and repetitive DNA.

A detailed report of the Sorghum Genomics Planning Workshop, sponsored by NSF, is available as supplemental data. The goals of the workshop were to (1) obtain a status report on the development and accessibility of sorghum genome research information, technologies, and infrastructure; (2) identify future priorities and needs for sorghum genomics research; (3) better organize the sorghum community; and (4) foster sorghum improvement.

Prior to the meeting, a survey was conducted to establish priority needs of the broader user community. The findings of the survey, obtained from the input of approximately 60 respondents among 140 members of the international sorghum (and closely related sugarcane) communities polled, are attached to the report. Topics included in the workshop were those identified as key issues by the user community. In summary, these can be classified into four focus areas, mapping, sequencing, germplasm, and database/bioinformatics, each of which is addressed below.

An interim Sorghum Genomics Steering Committee was charged with the development of the "white paper" outlining key priorities for sorghum genomics for the next 5 to 10 years, as well as devising a mechanism for, and carrying out, an election process to maintain and enhance community activities and impact.

While a natural long-term goal is a high-quality assembled sequence that is finished to Bermuda standards, this is likely to be accomplished in stages that build on one another.

The white paper (see supplemental data) details a three-stage strategy, also noting that aspects of the three stages are proceeding to some degree in parallel.

Stage 1: Gene Space Characterization

The sorghum gene space is presently represented by approximately 200,000 expressed sequence tags (ESTs) that have been clustered into approximately 22,000 uniscripts, representing more than 20 diverse libraries from several genotypes. Genome annotation will benefit from additional EST sequencing, emphasizing full-length clones. This also represents an opportunity to sample both physiological and genetic (single-nucleotide polymorphism) diversity by drawing these ESTs from diverse genotypes.

About 500,000 methyl-filtered (MF) reads that provide estimated 1× coverage of the MF-estimated gene space (Bedell et al., 2005) have been assembled into contigs (SAMIs; <http://magi.plantgenomics.iastate.edu/>). Another reduced-representation strategy, Cot-based cloning and sequencing (CBCS), was first demonstrated in sorghum in 2001 (as noted in GenBank accessions AZ921847–AZ923007) and further detailed subsequently (Peterson et al., 2002). This method offers the potential to further enhance gene space coverage beyond that offered by ESTs and MF, in a complementary manner as demonstrated for maize. Sequencing of the low-copy DNA to similar levels of coverage, by MF- and CBCS-based methods, is viewed as a logical intermediate step toward efficiently capturing the sequence complexity of sorghum. Primary sequencing of genomic DNA should be focused on the inbred genotype BTx623 (see below), to foster sequence assembly.

Stage 2: Gold-Standard Physical Map

Most genomic resources for sorghum have been developed using a U.S. inbred, BTx623, which was selected as a focal point for genomic sequencing and which enjoys about 20× genome coverage by two sets of BACs cloned using two different restriction enzymes. However, a gold-standard physical map will necessarily integrate data from multiple genotypes to help resolve genomic instabilities or other genotype- or species-specific features that interfere with cloning and/or sequencing. As such, our integrated physical map will comprise detailed alignment among BTx623, *S. propinquum*, and *S. bicolor* accession IS3620C. This will not only provide for filling gaps, but also advance application to studies of unique aspects of plant biology for which these diverse genotypes represent botanical models.

Stage 3: Finished Sequencing

Sorghum is a relatively complex genome, but with a smaller overall genome size and less repetitive DNA than many of its relatives, such as maize and sugarcane. While the physical map will provide the means to conduct BAC-by-BAC finished sequencing of a minimum tiling path, ongoing technological and computational improvements may offer compelling efficiencies to whole-genome shotgun (WGS)-based approaches, or more probably to hybrid approaches

that integrate aspects of BAC-based and WGS approaches. A high-quality genetically oriented physical map provides a robust guide for assembly by either BAC-based or WGS approaches, and the community remains open to considering a range of options for completion of the sequence based on economics and the state of the art as funding becomes available. In addition, it is noted that the relationship between the physical map and the sequence may be iterative; for example, targeted physical anchoring of WGS contigs may expedite assembly and closure.

Database Resources/Bioinformatics

While existing Web-based resources focus on comparative structural and evolutionary genomics (<http://cggc.agtec.uga.edu/>), functional genomics of the transcriptome (<http://funken.botany.uga.edu/>), and genomics of the unique abiotic stress responses of sorghum (<http://sorgblast2.tamu.edu/>), the community recognizes a growing need to develop a unified sorghum database much like Maize GDB or rice-centric Gramene. This database may be centralized or federated but should maintain critical links to the individual groups' databases, thus taking advantage of the respective strengths of individual groups in annotation and curation of data that they have first-hand knowledge of and that is of primary importance to them. In addition, the establishment of a "Sorghum Portal," with links to relevant Web resources, is recommended. Finally, data also need to be accessible in formats compatible with usage by scientists in regions where Internet access remains unavailable or too slow to efficiently download genomic data sets.

Applications to Benefit Worldwide Stakeholders

Much of the value of a sorghum sequence would be realized through better understanding of the levels and patterns of diversity in extant germplasm, which can contribute both to functional analysis of specific sorghum genes and to deterministic improvement of sorghum for specific needs and environments. Extensive *ex situ* sorghum germplasm collections exist within the U.S. National Plant Germplasm System and the International Crops Research Institute for the Semi-Arid Tropics. An informal meeting of breeders and geneticists at Cornell in September 2004 laid the groundwork for development of "core" sorghum panels (including wild species, landraces, and elite genotypes), which will capture as much genetic diversity as possible while minimizing redundancies. These panels are expected to be suitable for association genetics-based approaches that explore relationships between phenotypes and haplotype variation at candidate genes directly, or for markers distributed either throughout the genome (genome-wide association tests) or closely linked to a target locus. Plans were also outlined for development of about 25 recombinant inbred line populations initially needed to foster joint

quantitative trait loci-association genetics studies. The germplasm planning group met again on February 21, 2005, to further advance a recommendation to be circulated to the broader sorghum community for feedback.

Africa is recognized as the center of origin and diversity for sorghum, the source of about 50% of the accessions in the world collections, and the location of much *in situ* diversity lacking from existing collections. Sorghum is thus an attractive vehicle for engagement of the African scientific community in genomics and its applications, in particular regarding documentation and analysis of *in situ* diversity that is presently inaccessible to Western scientists. However, the problems of limited communications infrastructure, slow Internet connectivity, and lack of information technology support will present special challenges. Periodic data dissemination in CD format, together with the development of simplified database structures that use off-the-shelf software but are cross-compatible with more sophisticated structures such as the planned centralized database (above), will be essential. Implementation of these new capabilities will benefit from the engagement of institutions such as the Consultative Group on International Agricultural Research Centers, the International Sorghum and Millet Collaborative Research Support Program, the Rockefeller and Syngenta foundations, and others.

To sustain coordination and communication, the initial Sorghum Genomics Executive Committee will coordinate the identification and election of its successors, to comprise a 15-member committee with at least five and two seats filled by non-U.S. and private sector members, respectively, to serve 3-year terms on a staggered basis (five new members per year). The elected committee will establish guidelines for future committee actions; collect, collate, and disseminate information; organize consortium meetings; and serve as an advocacy group for each country and area of research, with emphasis on cohesiveness of the community and importance of the crop.

SYNTHESIS

As a model organism for tropical grasses that carry out C_4 photosynthesis, sorghum is a logical complement to the C_3 grass *Oryza*, the first monocot plant with a near-completely sequenced genome. The relatively small genome of sorghum is likely to be appreciably less complex to assemble than the larger and more repetitive genomes of other major C_4 crops, such as maize and sugarcane. Detailed physical maps provide a foundation upon which to overlay sequence assemblies, linking them to a rich history of genetics and genomics research based on inclusion of genetically mapped sequence-tagged sites in the physical map. Sequencing of sorghum will fill a key gap in plant biogeography in view of its African origin, permitting phylogenetic triangulation of key events in cereal evolution and in particular leading to new

insights into parallel but independent domestication of the cereals. Its remarkable ability to produce a crop under adverse conditions, in particular with much less water than most other grain crops, makes sorghum an important failsafe source of food, feed, fiber, and fuel in the global agroecosystem with special relevance to Africa. Moreover, sorghum genome analysis offers novel learning opportunities relevant to weed biology as well as to improvement of a wide range of other forage, turf, and biomass crops. The infrastructure to link a finished sequence to a rich history of genetics research, toward resolution of a wide range of challenges facing a worldwide set of stakeholders, is largely in place.

ACKNOWLEDGMENT

The U.S. National Science Foundation supported the Sorghum Genomics Workshop (under award no. DBI-0456171).

Received May 3, 2005; revised May 3, 2005; accepted June 5, 2005; published August 11, 2005.

LITERATURE CITED

- Arumuganathan K, Earle E** (1991) Estimation of nuclear DNA content of plants by flow cytometry. *Plant Mol Biol Rep* **9**: 208–218
- Bedell JA, Budiman MA, Nunberg A, Citek RW, Robbins D, Jones J, Flick E, Rohlfing T, Fries J, Bradford K, et al** (2005) Sorghum genome sequencing by methylation filtration. *PLoS Biol* **3**: 103–115
- Bowers JE, Abbey C, Anderson S, Chang C, Draye X, Hoppe AH, Jessup R, Lemke C, Lenington J, Li Z, et al** (2003) A high-density genetic recombination map of sequence-tagged sites for sorghum, as a framework for comparative structural and evolutionary genomics of tropical grains and grasses. *Genetics* **165**: 367–386
- Chittenden LM, Schertz KE, Lin YR, Wing RA, Paterson AH** (1994) A detailed RFLP map of *Sorghum bicolor* X *S. propinquum*, suitable for high-density mapping, suggests ancestral duplication of sorghum chromosomes or chromosomal segments. *Theor Appl Genet* **87**: 925–933
- Dufour P, Deu M, Grivet L, Dhont A, Paulet F, Bouet A, Lanaud C, Glaszmann JC, Hamon P** (1997) Construction of a composite sorghum genome map and comparison with sugarcane, a related complex polyploid. *Theor Appl Genet* **94**: 409–418
- Jessup RW, Burson BL, Burow G, Wang YW, Chang C, Li Z, Paterson AH, Hussey MA** (2003) Segmental allotetraploidy and allelic interactions in buffelgrass (*Pennisetum ciliare* (L.) Link syn. *Cenchrus ciliaris* L.) as revealed by genome mapping. *Genome* **46**: 304–313
- Kim JS, Klein PE, Klein RR, Price HJ, Mullet JE, Stelly DM** (2005) Chromosome identification and nomenclature of *Sorghum bicolor*. *Genetics* **169**: 1169–1173
- Klein PE, Klein RR, Cartinhour SW, Ulanich PE, Dong JM, Obert JA, Morishige DT, Schlueter SD, Childs KL, Ale M, et al** (2000) A high-throughput AFLP-based method for constructing integrated genetic and physical maps: progress toward a sorghum genome map. *Genome Res* **10**: 789–807
- Menz MA, Klein RR, Mullet JE, Obert JA, Unruh NC, Klein PE** (2002) A high-density genetic map of *Sorghum bicolor* (L.) Moench based on 2926 AFLP, RFLP and SSR markers. *Plant Mol Biol* **48**: 483–499
- Ming R, Liu SC, Lin YR, da Silva J, Wilson W, Braga D, van Deynze A, Wenslaff TF, Wu KK, Moore PH, et al** (1998) Detailed alignment of Saccharum and Sorghum chromosomes: comparative organization of closely related diploid and polyploid genomes. *Genetics* **150**: 1663–1682
- Missaoui A, Paterson AH, Bouton JH** (2005) Investigation of genome organization in switchgrass (*Panicum virgatum* L.) using DNA markers. *Theor Appl Genet* **110**: 1372–1383
- Paterson A, Lin Y, Li Z, Schertz K, Doebley J, Pinson S, Liu S, Stansel J, Irvine J** (1995) Convergent domestication of cereal crops by independent mutations at corresponding genetic loci. *Science* **269**: 1714–1718
- Paterson AH, Bowers JE, Chapman BA** (2004) Ancient polyploidization predating divergence of the cereals, and its consequences for comparative genomics. *Proc Natl Acad Sci USA* **101**: 9903–9908
- Peterson DG, Schulze SR, Sciara EB, Lee SA, Bowers JE, Nagel ANJ, Tibbitts DC, Wessler SR, Paterson AH** (2002) Integration of Cot analysis, DNA cloning, and high-throughput sequencing facilitates genome characterization and gene discovery. *Genome Res* **12**: 795–807
- Swigonova Z, Lai J, Ma J, Ramakrishna W, Llaca V, Bennetzen JL, Messing J** (2004) Close split of sorghum and maize genome progenitors. *Genome Res* **14**: 1916–1923
- Whitkus R, Doebley J, Lee M** (1992) Comparative genetic mapping of sorghum and maize. *Genetics* **132**: 1119–1130