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The timing of herbivore-induced volatile emission in black poplar (*Populus nigra*) and the influence of herbivore age and identity affect the value of individual volatiles as cues for herbivore enemies

Clavijo McCormick et al.





## **RESEARCH ARTICLE**

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- The timing of herbivore-induced volatile emission in black poplar (*Populus nigra*) and the influence of
- herbivore age and identity affect the value of
- individual volatiles as cues for herbivore enemies
- Andrea Clavijo McCormick<sup>†</sup>, G Andreas Boeckler<sup>†</sup>, Tobias G Köllner, Jonathan Gershenzon and Sybille B Unsicker<sup>\*</sup>

### **Abstract**

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**Background:** The role of herbivore-induced plant volatiles as signals mediating the attraction of herbivore enemies is a well-known phenomenon. Studies with short-lived herbaceous plant species have shown that various biotic and abiotic factors can strongly affect the quantity, composition and timing of volatile emission dynamics. However, there is little knowledge on how these factors influence the volatile emission of long-lived woody perennials. The aim of this study was to investigate the temporal dynamics of herbivore-induced volatile emission of black poplar (*Populus nigra*) through several day-night cycles following the onset of herbivory. We also determined the influence of different herbivore species, caterpillars of the gypsy moth (*Lymantria dispar*) and poplar hawkmoth (*Laothoe populi*), and different herbivore developmental stages on emission.

**Results:** The emission dynamics of major groups of volatile compounds differed strikingly in response to the timing of herbivory and the day-night cycle. The emission of aldoximes, salicyl aldehyde, and to a lesser extent, green leaf volatiles began shortly after herbivore attack and ceased quickly after herbivore removal, irrespective of the day-night cycle. However, the emission of most terpenes showed a more delayed reaction to the start and end of herbivory, and emission was significantly greater during the day compared to the night. The identity of the caterpillar species caused only slight changes in emission, but variation in developmental stage had a strong impact on volatile emission with early instar *L. dispar* inducing more nitrogenous volatiles and terpenoids than late instar caterpillars of the same species.

**Conclusions:** The results indicate that only a few of the many herbivore-induced black poplar volatiles are released in tight correlation with the timing of herbivory. These may represent the most reliable cues for herbivore enemies and, interestingly, have been shown in a recent study to be the best attractants for an herbivore enemy that parasitizes gypsy moth larvae feeding on black poplar.

**Keywords:** Diurnal rhythm, Herbivore-induced plant volatiles (HIPV), Herbivore feeding pattern, Lepidoptera, Salicaceae, Signaling molecules in indirect defense, Tree defense

Department of Biochemistry, Max Planck Institute for Chemical Ecology, Hans-Knöll-Straβe 8, 07745 Jena, Germany



<sup>\*</sup> Correspondence: sunsicker@ice.mpg.de

<sup>&</sup>lt;sup>†</sup>Equal contributors

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### **Background**

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Herbivory induces dramatic changes in the volatile emission of plants. This phenomenon has been reported for many plant species from different orders, and possibly originated in photosynthetic bacteria long before the appearance of eukaryotic cells, leading to the belief that this is an ancestral feature of plants [1,2]. Herbivore-induced plant volatiles are well known to attract predators and parasitoids of herbivores and so have been frequently termed a "cry for help" from the plant to reduce herbivore pressure [2-4]. However, it is still unclear if herbivore enemy recruitment has a real fitness benefit for the plant or if plant volatiles are reliable cues for natural enemies of herbivores [5-7]. Major limitations in understanding the ecological roles of plant volatiles are the complexity of the emitted blends and our lack of knowledge on how insects perceive and process olfactory information [4].

One interesting aspect about volatile emission upon herbivory is its dynamic nature. Volatile emission patterns change during the course of herbivory with variation in how soon compounds are emitted after the start of herbivory [8-12], how soon emission decreases after herbivory stops [9,10,13] and changes in day and night cycles [14]. The emission patterns of abundant herbivore-induced volatiles, such as green leaf volatiles (GLVs) and terpenoids, are well described in the literature. However, much less is known about compounds emitted in lower amounts, such as aromatic compounds and amino acid derivatives (nitrogen and sulfur containing compounds) [2,4,15,16], although there is evidence that such minor compounds could have a high ecological value for both herbivores and their natural enemies [17-20].

Herbivore enemies have been shown to use differences in plant volatile emission to successfully discriminate between host plant species or cultivars [21-26] and between plants under different physiological stress conditions [27]. Herbivore parasitoids and predators can also obtain detailed information from volatile cues about the nature of the attacking herbivore species, and its developmental stage or parasitization status [14,28-30]. The presence of multiple herbivores adds another level of complexity to volatile emission causing increased attraction of herbivore enemies in some cases [31-35].

Understanding how herbivore enemies respond to volatiles emitted by different plant-herbivore combinations will increase our understanding about the ecological roles of specific compounds, but there are many gaps in our knowledge of what affects volatile emission in such circumstances. For example, how the spectrum of volatiles is altered by different herbivore species or different feeding stages is seldom taken into account e.g. [36]. Additionally, most studies on herbivore enemy recruitment focus on volatiles present at just one time point after herbivory starts (but see [37]).

Despite the long history of research on plant volatiles, 85 most research has concentrated on herbaceous species and relatively few studies have explored the emission of 87 herbivore-induced volatiles from woody perennial species and their ecological roles e.g. [18,38-43].

Among woody plants, poplar has become a model organism because of its ecological and economic importance. In addition, since the completion of the genome of 92 Populus trichocarpa [44], many genetic, genomic, biochemical and molecular tools are now available and a growing amount of information is accumulating that has opened the doors to studying many aspects of poplar biology, including direct and indirect defense [18,45].

In a previous study, we documented the enormous diversity of volatile compounds emitted by black poplar (Populus nigra) upon herbivore attack and established that the parasitoid Glyptapanteles liparidis, which preferentially parasitizes second instar gypsy moth (L. dispar) caterpillars on black poplar, is attracted to minor nitrogencontaining volatiles emitted by poplar locally at the sites of herbivory. Parasitoid wasps were also attracted to these 105 minor volatiles and green leaf volatiles when compounds were presented individually under field conditions, indicating that these substances might be important cues for a broad range of natural enemies of herbivores feeding on poplar trees [18]. However, in this earlier study, we did not explore the reasons why these compounds might be preferred by parasitoids over other more abundant poplar volatiles such as terpenoids.

We hypothesize that compounds which are important 114 cues for herbivore enemies should possess certain traits. They should A) indicate the actual presence of the herbivore (being rapidly emitted after the onset of herbivory 117 with emission ceasing quickly after herbivore departure), B) be emitted independently of light and dark conditions at times when herbivore enemies are foraging, and C) provide information about the identity, age and abundance of the herbivore. The aim of this study was to investigate the temporal dynamics of herbivore-induced volatile emission of black poplar (Populus nigra) during and after herbivory, and to investigate the differences in volatile emission in response to different herbivore species, developmental stages of a herbivore and amount of feeding. These data should help establish which compounds could be most useful sources of information for herbivore enemies.

### Results

### Temporal dynamics of volatile emission in black poplar after gypsy moth herbivory

To investigate the diurnal patterns of black poplar (Populus nigra) volatile emission, we selected 20 compounds as representatives of each of the major classes of volatiles found in this species: green leaf volatiles (GLVs), monoterpenes (cyclic and acyclic), homoterpenes, sesquiterpenes,

nitrogen-containing compounds and aromatic compounds. 138 The volatile blend from undamaged trees was dominated 139 by GLVs and cyclic monoterpenes, and these volatiles were 140 almost exclusively emitted during light periods (Figure 1, 141 Additional file 1: Figure S1). Feeding by 4<sup>th</sup> instar larvae of the generalist herbivore Lymantria dispar caused an in-143 creased emission of all volatiles measured, although the

extent of increase varied with the compound class, diurnal 145 cycle, and the timing of herbivory.

GLVs such as (Z)-3-hexenyl acetate were emitted rapidly upon the onset of herbivory, and emission declined 148 after herbivore removal. They were released both day 149 and night, with a greater emission during the day. The 150 emission of terpenoids was also greater during the day 151

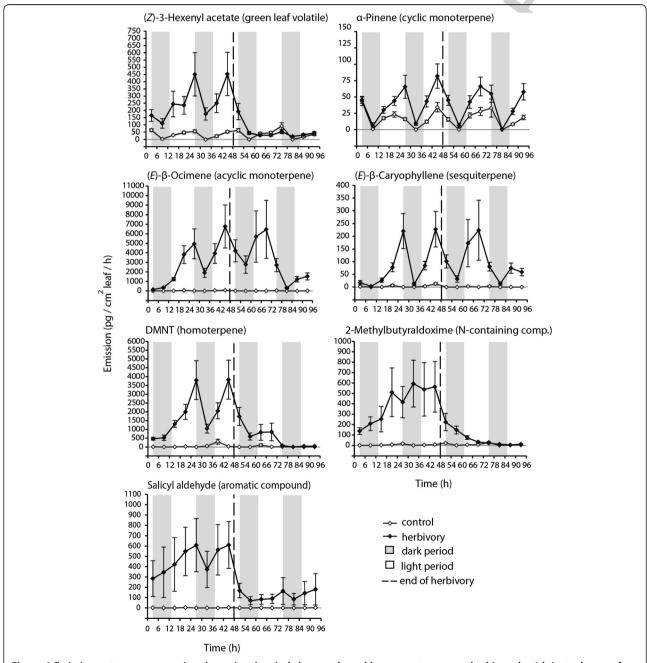


Figure 1 Emission patterns representing the major chemical classes released by young trees upon herbivory by 4th instar larvae of Lymantria dispar (gypsy moth) or from undamaged controls over a 4-day experiment. The graphs depict the rates of emission for individual compounds over the course of herbivory (initiated at the beginning of the experiment for herbivory treatment) as well as after herbivore removal. Volatiles were continuously sampled day and night in 6 h intervals. Means and ± SEM are given.

than at night, but the increase in emission after herbivory did not coincide with the onset of herbivory, but occurred 153 only several hours after caterpillar damage had begun. Fur-154 thermore, terpenoids continued to be emitted several hours 155 after herbivore removal and in some cases even until the 156 end of the experiment 48 hours later (Figure 1, Additional 157 file 1: Figure S1). Among the terpenes, cyclic monoterpenes 158 159 showed only a modest increase in emission after induction (roughly two-fold). By contrast, acyclic monoterpenes, ses-160 quiterpenes, and the homoterpene DMNT, which were 161 only present in minute amounts in the headspace of uninfested plants, showed a many-fold increase (e.g. 7000-fold 163 for (E)-β-ocimene, 4000-fold for DMNT and 250-fold for 164 (E)-β-caryophyllene) after herbivory (Figure 1, Additional 165 file 1: Figure S1). 166

Of the nitrogen-containing compounds, the emission of 2-methylbutyraldoxime was induced immediately by herbivory and increased to its highest levels during the first full light period. The rate of emission was not influenced by the light or dark period, and it declined to baseline levels after herbivory ended (Figure 1). The emission patterns of two other nitrogen-containing compounds, benzyl cyanide and indole, were different in displaying significant diurnal rhythms (emission 2-3-fold greater during the day as during the night) and a less rapid decline after caterpillars were removed (Additional file 1: Figure S1).

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Among the aromatic compounds, salicyl aldehyde was emitted almost from the onset of herbivory in substantial rates, both day and night, ceasing abruptly after herbivore removal (Figure 1). Two other aromatic compounds, benzyl alcohol and benzene ethanol, showed much more of a biphasic emission pattern, elevated during the day and reduced at night (Additional file 1: Figure S1). Emission was induced by herbivory more slowly than for salicyl aldehyde and stayed at significantly higher emission levels than in controls until almost the end of the experiment rather than declining rapidly after herbivory stopped as for salicyl aldehyde.

#### Effect of herbivore species, its developmental stage, and feeding intensity on volatile emission 190

In comparing the herbivory of 5<sup>th</sup> instar *L. dispar* larvae to that of 2<sup>nd</sup> instar L. dispar and larvae of another lepidop-192 teran, the specialist Laothoe populi, we observed that the three treatments had very characteristic damage patterns 194 (Figure 2A). For example, 5<sup>th</sup> instar *L. populi* larvae caused F2 few, but very extensive lesions in a few leaves, often com-196 197 pletely consuming the whole leaf blade. Fifth instar L. dispar caterpillars also caused extensive lesions on a few 198 leaves, but mostly avoided the leaf venation and only rarely consumed whole leaf blades (Figure 2A). Further-200 more 5<sup>th</sup> instar *L. dispar* caterpillars moved more often 201 from one leaf to another thus damaging more leaves over-202 all than L. populi. In contrast, second instar L. dispar caterpillars caused numerous small lesions and frequently changed feeding position causing minor to moderate damage on a larger number of leaves. When fifth instar L. dispar and L. populi were combined, there was an intermediate damage pattern between that of both herbivores measured separately (Figure 2B).

To quantify the feeding differences among herbivore 210 treatments in relation to volatile emission, we calculated 211 the number of damaged leaves and number of lesions as observed in each treatment. We found a significant positive correlation between total volatile emission and the two parameters: number of damaged leaves and number of lesions 215  $(R^2 = 0.603, p = 0.005 \text{ and } R^2 = 0.735, p = 0.0002, respect$ ively), as well as a significant correlation between these two damage parameters ( $R^2 = 0.739$ , p = 0.0002) (Figure 2B). Therefore we used principal component analysis as a factor reduction technique to combine these two parameters into a single component which we termed feeding intensity. Then we applied a generalized least square model (GLS) to 222 calculate the effect of the herbivory treatment (larval species and instar), the feeding intensity (regardless of treatment), and their interaction on the rate of emission of each of the 20 studied compounds (Table 1). Emission was calculated relative to total leaf area consumed in each treatment to control for variation in the extent of herbivory among treatments.

In comparing *P. nigra* volatiles among treatments, only 230 four compounds differed significantly in emission upon feeding by the two caterpillar species tested (the specialist *L. populi* and the generalist *L. dispar* both 5<sup>th</sup> instar): (*E*)-β-caryophyllene, 3-methylbutyraldoxime, myrcene and nerolidol (Figure 3), all emitted in greater abundance after damage by L. dispar. Four compounds were also different between combined damage by the two herbivore species vs. damage by the generalist herbivore alone: (E)- $\beta$ -caryophyllene, 3-methylbutyraldoxime, (Z)-3-hexenol and nerolidol (Figure 3). These compounds were emitted in higher amounts by *L. dispar* than by the 241 two species combined. The emission in the combined damage treatment did not differ significantly from that induced by the specialist herbivore (L. populi) alone (Figure 3, Additional file 2: Figure S2). Herbivore instar had very strong effect on volatile emission caused by L. dispar: early instar L. dispar induced significantly more emission of nitrogen-containing volatiles and most terpenoids than late instar *L. dispar* and *L. populi* (Figure 3, Additional file 2: Figure S2).

The feeding intensity had also a significant direct effect on the emission of the majority of black poplar volatiles tested: all monoterpenes, the sesquiterpenes nerolidol and (E)-β-caryophyllene, all nitrogen containing volatiles excluding indole and the GLV (Z)-3-hexenyl acetate (Figure 4, Additional file 3: Figure S3, Table 1). Interestingly the emission of DMNT, which is one of the most abundant herbivore induced volatiles, was shown not to be influenced by

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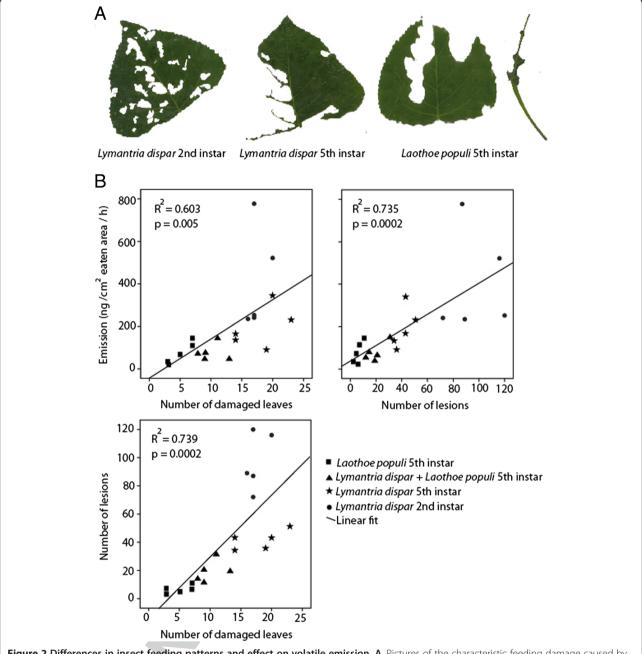
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**Figure 2 Differences in insect feeding patterns and effect on volatile emission. A.** Pictures of the characteristic feeding damage caused by second instar *Lymantria dispar*, fifth instar *L. dispar* and fifth instar *Laothoe populi* on *Populus nigra* leaves. **B.** Correlation between two insect feeding parameters (number of leaves damaged and number of lesions) and total volatile emission of *P. nigra* leaves (combined emission of the 20 compounds investigated) in relation to the amount of leaf area eaten. Correlation between the two feeding parameters is also shown. Herbivory treatments are depicted by different symbols.

feeding intensity or the identity and developmental stage of the herbivore, which is also the case for the aromatic compounds benzyl alcohol and benzene ethanol (Table 1). For the interaction between herbivory treatment (herbivore identity and developmental stage) and feeding intensity, we only observed a significant effect for two aromatic compounds, salicyl aldehyde and benzene ethanol.

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### **Discussion and conclusions**

# Major groups of herbivore-induced volatiles in poplar show different temporal emission patterns

The value of herbivore-induced plant volatiles as cues for 269 herbivore enemies depends on how closely their emission 270 correlates with the presence of herbivores. While some 271 compounds were emitted almost immediately after the 272

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Table 1 Effect of herbivore identity, feeding intensity and their interaction on black poplar volatile emission

t1.2	Compound		Interaction (feeding intensity x Herbivore treatment)		Herbivory treatment		Feeding intensity	
t1.3		Variance structure	Likelihood ratio	p. value	Likelihood ratio	p. value	Likelihood ratio	p. value
t1.4	Monoterpenes							
t1.5	α-Pinene (cyclic)	7	0.906	0.824	15.227	0.002**	15.393	<0.001***
t1.6	Camphene (cyclic)	2	0.999	0.802	8.246	0.038*	8.883	0.003**
t1.7	Myrcene (cyclic)	4	0.642	0.887	8.364	0.04*	7.533	0.006**
t1.8	Borneol (cyclic)	2	0.779	0.855	6.925	0.074	9.052	0.003**
t1.9	(Z)-Ocimene (acyclic)	8	3.951	0.267	7.286	0.063	8.424	0.004**
t1.10	(E)-β-Ocimene (acyclic)	4	1.384	0.709	9.797	0.020*	14.036	<0.001***
t1.11	Linalool (acyclic)	2	1.441	0.696	2.456	0.483	11.012	<0.001***
t1.12	Homoterpene							
t1.13	DMNT	8	7.797	0.051	3.786	0.286	0.444	0.505
t1.14	Sesquiterpenes					7		
t1.15	( <i>E</i> )-β-Caryophyllene	2	0.667	0.881	11.371	0.01*	4.334	0.037*
t1.16	α-Humulene	2	1.526	0.676	9.014	0.029*	1.676	0.196
t1.17	Nerolidol	4	6.410	0.093	10.387	0.016*	12.891	<0.001***
t1.18	Green leaf volatiles							
t1.19	(Z)-3-Hexenyl acetate	2	0.656	0.884	6.067	0.108	6.454	0.011*
t1.20	(Z)-3-Hexenol	1	2.284	0.516	16.015	0.001**	0.138	0.71
t1.21	N-containing comp.			X				
t1.22	2-Methylbutyraldoxime	7	0.522	0.914	10.821	0.013*	4.454	0.035*
t1.23	3-Methylbutyraldoxime	4	0.536	0.911	14.950	0.002**	10.335	0.001**
t1.24	Benzyl cyanide	3	2.723	0.466	10.852	0.013*	9.464	0.002**
t1.25	Indol	3	2.136	0.545	9.688	0.021*	1.537	0.215
t1.26	Aromatic compounds							
t1.27	Salicyl aldehyde	8	8.734	0.033*	13.706	0.003**	0.535	0.464
t1.28	Benzyl alcohol	4	4.867	0.182	4.624	0.202	6.770	0.386
t1.29	Benzene ethanol	4	8.003	0.046*	7.629	0.054	0.703	0.402

For each parameter the F and p values are given. Asterisks indicate significant differences, p < 0.001 = \*\*\*, p < 0.01 = \*\*\*, p < 0.05 = \*, no asterisk = not significant.

t1.35 Combined variance (varident for herbivory treatment, varExp for feeding intensity). A detailed description of the variance structures is given by [80].

onset of herbivory and ceased emission soon after herbivory had stopped, others, especially terpenes, were first emitted only 12 hours after the beginning of herbivory and contin-275 276 ued being emitted for a day or more after herbivory had stopped. These differences suggest very divergent mecha-277 nisms triggering and controlling the biosynthesis of these 278 279 compounds [2,15,16]. There are also differences for the same compound class among different plant species. For 280 instance, GLV emission is often considered to be restricted to the time when actual leaf damage occurs [46], but here 282 (Z)-3-hexenol emission continued for 24 hours after herbiv-283 284 ory had stopped (Additional file 1: Figure S1). The volatiles that are the most diagnostic cues for herbivore enemies, 285 should be emitted as long as herbivores are present.

Variation of emission with day-night rhythm may 287 also affect the value of volatiles as herbivore enemy attractants. The emission of most herbivore-induced and constitutive volatiles was found to vary strongly in 290 a diurnal fashion. The terpenoids followed this trend 291 especially well with emission being much higher in 292 light vs. dark periods for all compounds measured. 293 Previous work with herbaceous plants also found the 294 emission of monoterpenes ( $C_{10}$ ), sesquiterpenes ( $C_{15}$ ) and homoterpenes (the C<sub>15</sub>-derived homoterpene 296 DMNT) to be much higher in the day than the night 297 [9,47]. A correlation with light may arise because 298 much of the substrate for the biosynthesis of volatile 299 terpenes arises from the methylerythritol phosphate 300

t1.31 Compounds are grouped according to their chemical classes, the second column shows the variance structure with the lowest Akaike Information Criterion (AIC),

which was used in the Generalized Least Square model (GLS). Variance structures tested were as follows: 1. varFixed variance for feeding intensity, 2. varIdent

t1.33 variance for herbivory treatment, 3. varPower variance for herbivory treatment, 4. varExp variance for feeding intensity, 5. varConstPower for feeding intensity,

<sup>6.</sup> varConstPower for feeding intensity and herbivory treatment, 7. Combined variance (varIdent for herbivory treatment, varFixed for feeding intensity) and 8.

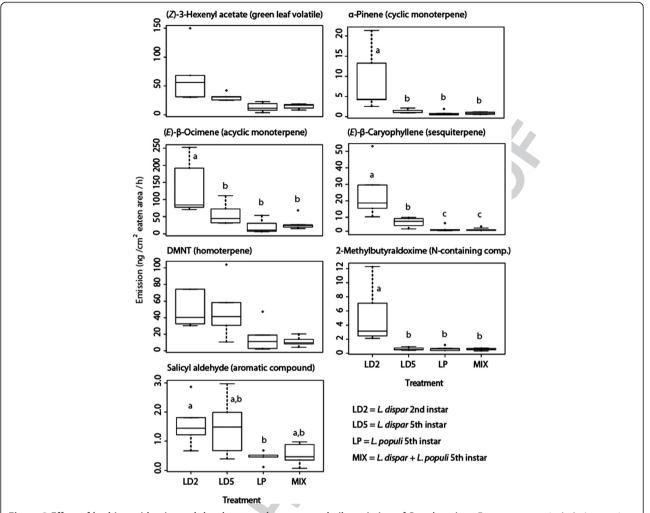


Figure 3 Effect of herbivore identity and developmental stage on volatile emission of Populua nigra. Four treatments include Lymantria dispar (2nd instar), L. dispar (5th instar), Laothoe populi (5th instar), and a mixture of L. dispar (5th instar) and L. populi (5th instar). Box-plots showing the same letter are not statistically significant from one another after a Tukey test performed on the fitted values after applying a GLS model, excluding the effect of the feeding intensity. P values are given in Table 1. Plots showing no letters indicate that there was no effect of the treatment on volatile emission.

pathway [48-50], which is closely connected with photosynthesis [51]. 302

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Other groups of compounds showed less clear trends in day-night emission patterns. Certain green leaf vola-304 tiles (GLVs) [(Z)-3-hexenyl acetate], nitrogen-containing compounds (benzyl cyanide, indole) and aromatic com-306 pounds (benzene ethanol) displayed a strong diurnal 308 rhythm with more emission in the light, but other mem-309 bers of these groups showed weaker rhythms or none at all. GLVs are sometimes reported to be emitted inde-310 pendently of any diurnal rhythm [38,52] or only at night [47]. Diurnal variation in volatile emission has been re-312 ported for many compounds in a range of plant species, 313 both herbaceous and woody, induced by herbivores or pathogens [9,47,53-55], including poplar [38,42,56], but the regulatory mechanisms are not known.

The significance of day-night variation for herbivore 317 enemy attraction depends on the activity rhythms of en- 318 emies. If enemies are active throughout the 24 hr cycle, 319 an emission pattern independent of light and dark, such 320 as that of 2-methylbutyraldoxime, salicyl aldehyde or 321 some GLVs, may be most advantageous. For enemies 322 that are only active at specific periods, emission during those times is most critical.

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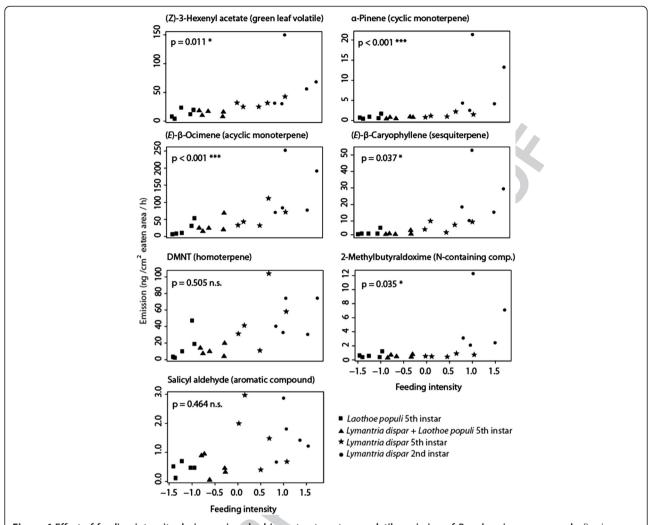
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### Emission varies in response to herbivore developmental stage, but not to herbivore species

In our study we found very few differences in volatile 327 emission among black poplar fed upon by two different 328 herbivore species, Lymantria dispar and Laothoe populi. 329 Possible explanations for this lack of species-specificity are that the two lepidopteran species tested feed in the 331



**Figure 4** Effect of feeding intensity during various herbivory treatments on volatile emission of *Populus nigra* compounds. P values indicate significant differences after applying a GLS model (excluding the effect of herbivore identity). p < 0.001 = \*\*\*\*, p < 0.05 = \*\*, n.s. = not significant. Herbivory treatments are depicted by different symbols.

same manner and share similar elicitors in their saliva. In previous studies, feeding by leaf-chewing lepidopteran larvae and grasshoppers has induced similar blends of volatiles [12,57,58] suggesting that these volatiles are a general response to attack by chewing insects. In support of this suggestion is the fact that, despite the great diversity of herbivores, only a few elicitors triggering defense responses in plants have been identified in herbivore oral secretions so far [59-62].

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In contrast to arthropod herbivores from a single feeding guild, such as leaf chewers or phloem feeders, it is likely that arthropods from different feeding guilds induce different patterns of plant volatile emission [63,64], although there are exceptions in the literature that challenge this idea [65,66]. If there are differences in feeding mode between younger and older larvae of a single species, these might also lead to differences in emission. In our study, we found that "feeding intensity" (a factor

combining number of damaged leaves and number of lesions) differed between early and late instar Lymantria dispar. In fact, there was more similarity in feeding intensity between late instar L. dispar and late instar 353 Laothoe populi than between early and late instar L. dis- 354 par which led to corresponding differences in volatile 355 emission. Nitrogen-containing volatiles and most ter- 356 penes were emitted at greater rates from early vs. late instar *L. dispar*. Thus volatile emission profiles were more influenced by instar and damage intensity than the identity of the herbivore species. Alterations in emission profiles induced by feeding of different instars of a single 361 herbivore have also been reported in previous studies [30,67,68], and may aid herbivore enemies in finding their favored prey or host stage. The differences between 364 instars in our study may also be due to the fact that, although we controlled for herbivore weight, the number 366 of feeding caterpillars was much higher in the second 367

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instar herbivory treatment than in the 5th instar treatments. However, under natural conditions, many moth 369 and butterfly caterpillars are gregarious early in develop-370 ment, and become solitary in late instars [69]. Thus the 371 differences in our treatments reflect natural conditions. 372

When young black poplar trees were simultaneously attacked by late instar *L. dispar* and *L. populi* caterpillars the emission of a few compounds decreased in comparison to trees infested by late instar *L. dispar* alone. Previous studies have already documented attenuation in volatile emission upon multiple herbivore species attack, however, examples for enhanced volatile emission in response to simultaneous feeding by different species also exist [70].

Further studies on the effects of larval stage and simultaneous attack by different herbivore species on volatile emission are necessary to better define these differences and survey their impact on herbivore enemies.

#### The potential role of black poplar volatiles in attraction of 385 herbivore enemies 386

Our initial hypothesis was that plant volatile compounds 387 employed as cues by herbivore enemies should: a) reli-388 ably indicate the actual presence of herbivores, b) be 389 390 emitted independently of light or dark cycles as long as herbivore enemies are active, and c) provide specific in-391 formation about the identity, developmental stage and 392 abundance of the herbivore. Although no individual 393 compound released from P. nigra meets all the require-394 ments, 2-methylbutyraldoxime and salicyl aldehyde ful-395 fill the first two requirements best, whereas a number of 396 compounds are informative regarding herbivore identity 397 (3-methylbutyraldoxime, myrcene, (E)- $\beta$ -caryophyllene 398 and nerolidol), herbivore instar (aldoximes, most ter-399 400 penes) and herbivore abundance (most volatiles).

The list of volatiles that best meet the criteria to serve as good signals for enemies of P. nigra herbivores shows a remarkable correspondence with those compounds found previously to be attractive to the braconid koinobiont parasitoid Glyptapanteles liparidis, which is a specialist on early instar L. dispar caterpillars. The aldoximes, 2and 3-methylbutyraldoxime, were the only compounds showing attraction in laboratory bioassays, while 2methylbutyraldoxime (3-methylbutyraldoxime was not tested), benzyl cyanide, (Z)-3-hexenol, (Z)-3-hexenyl acetate and linalool were attractive to a community of different parasitoid species in a natural P. nigra stand [18]. It would be interesting to know the major attractive cues for other enemies of L. dispar. The importance of individual herbivore-induced volatiles to herbivore enemies may also depend on their degree of host or prey specificity [71]. Generalist parasitoids and predators might orient towards abundant widespread compounds which generally signal herbivory (such as GLVs), whereas specialists may benefit from responding only to more specific compounds (such as aldoximes). Even though the differences in black poplar 421 volatile emission upon damage by late instar L. populi and L. dispar are minor, parasitoids may still be able to locate 423 their prey under natural conditions, as they possess very sensitive olfactory systems to detect slight changes in volatile cues that we cannot detect with our analytical devices.

In the case of koinobiont parasitoids which develop inside a living host, there is a preference to oviposit in early instar larvae to prevent the risk of encapsulation as well as to allow the completion of the endoparasitic larval stage which would not be possible if the host entered pupation [72,73]. In this sense, compounds signaling early instar damage should be of great importance for koinobiont parasitoids.

The emission patterns of herbivore-induced volatiles may also reflect other roles of these substances in the plant. Plant volatiles have been implicated in direct defense against herbivores [74], communication within and among 438 plants [75], and resistance to abiotic stresses, such as high light and temperature [76]. The importance of some of these roles could vary during the diurnal cycle. For example, since light and high temperature stresses would occur during the day, volatiles such as isoprene and monoterpenes involved in resistance to these stresses might be emitted in greater amounts during the day.

### Critical conclusion

Upon herbivore damage, plants typically emit a large, diverse blend of volatile compounds that have been shown to have importance in direct defense against herbivores and the attraction of herbivore enemies. In black poplar, a few individual compounds of the blend have been shown to be active in enemy attraction [18]. Here we show that 452 these active compounds may have been selected as cues by herbivore enemies because they are more reliable indicators of herbivore presence and provide information about the age and identity of the damaging species.

### Methods

## Plants & insects

### Populus nigra

Black poplar trees were grown from stem cuttings obtained from old-growth trees and raised under summer conditions in a climate chamber (~14:10 h day:night photoperiod, 22°C day - 19°C night, 60% humidity). The light period started at 6:00 a.m. and ended at 8:00 p.m. Cuttings were planted in 2 L pots containing a 2:2:1 mixture of clay, humus and sand. Fertilizer and water were applied regularly until the experiment started.

### Lymantria dispar

L. dispar caterpillars were hatched from egg clutches 469 (kindly provided by Melody Keena and Hanna Nadel 470 from the, US Department of Agriculture - Mill Pond 471

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Road Hamden, CT and Buzzard's Bay, MA) and reared on artificial gypsy moth diet (MP Biomedicals LLC, Illkirch, France) until two days before the experiments started, where they were then fed with P. nigra leaves to get adapted to this food source. All caterpillars were maintained in a climate chamber with the same photoperiod, 477 temperature and relative humidity conditions as described 479 above.

#### Laothoe populi 480

L. populi caterpillars were hatched from eggs (purchased from the Lepidoptera Breeders Association, Seaford, UK) and reared on fresh poplar leaves at ambient temperatures in the laboratory. 484

#### Volatile collection and analysis 485

#### Temporal dynamics experiment 486

To investigate the diurnal variation of volatile emission 487 in poplar, volatiles were collected in a climate chamber 488 using a push-pull system that consisted of a circular 489 Plexiglas top (Ø 26 cm) attached to a cylindrical PET 490 bag (Ø 26 cm, 50 cm height). Two holes were drilled 491 through the top to hold the valves for incoming and out-492 493 going air. A young tree (~40 cm tall, 2 months old) was introduced into the system through the bottom opening 494 and the PET-bag was fastened to the pot with a cable binder. During the volatile collection, charcoal purified 496 air was pumped through Teflon tubing into the system 497 at a flow rate of 2.5 L min<sup>-1</sup>. At the same time, 498 1.5 L min<sup>-1</sup> of air from the plant headspace was pumped out of the system through a Teflon tube passing through 500 a 20 mg Super-Q (Alltech, FL, USA) filter to absorb vol-501 atiles. The abiotic conditions in the climate chamber 502 were kept the same as described above. Ten trees were 503 assigned to each of two treatments (herbivory, control) 504 and placed inside the collection system. Fifteen 4<sup>th</sup> instar 505 gypsy moth caterpillars were released on the trees in the 506 herbivore treatment shortly before the first volatile col-507 508 lection. The first volatile collection started at 5 pm with 3 h light period remaining. Volatile emission was con-509 tinuously sampled in 6 h intervals for a total of 96 h, both during day and night. Gypsy moth caterpillars were removed from herbivore-treated trees after 48 h. By the end of the experiment, all leaves were excised and 513 photographed to determine the leaf area as described in [77]. Volatiles were eluted from Super-Q Traps with 200 μL dichloromethane containing 10 ng/μL of nonyl acetate as an internal standard. A portion (2 µL) of the eluate was splitlessly injected in a GC/MS equipped with a 30 m  $\times$  250  $\mu$ m  $\times$  0.25  $\mu$ m DB5-MS column (Wicom GmbH, Heppenheim, Germany). The injector was held at 230°C and helium was used as a carrier gas at 1 mL/ min. The oven temperature of the GC/MS was held at 50°C for 3 minutes after injection and then increased to

95°C at a rate of 4°C/min. Afterwards, the oven was 524 heated to 145°C with a 15°C/min gradient and then to 180°C with a 10°C/min gradient. Finally, the oven 526 temperature was held for 3 min at 300°C. Mass spectra were recorded with a 3 min solvent delay using a Hewlett-Packard MSD 5973 mass spectrometer (transfer 529 line temp: 230°C, source temp: 230°C, quadrupole temp: 530 150°C, ionization energy: 70 eV, mass range: 40–500 m/z). Compounds were identified by comparing their retention time to those of authentic standards. Quantifica- 533 tion was carried out by mass spectrometry since the 534 emission of some volatiles during dark periods turned 535 out to be too low for flame ionization detection. This however, limited the quantification to compounds that 537 could be acquired commercially in acceptable purity (>90%). Selected ion monitoring was used for quantification in a way that a specific m/z of each compound was referenced to the m/z = 69 of the internal standard. The 541 compound and m/z specific response factors required 542 for absolute quantification were calculated from dilutions 543 of the authentic standards in dichloromethane with a constant internal standard concentration of 8.64 ng/µL. 545 For each compound, two response factors were averaged from two six point calibration curves, one for a lower concentration range (0.2-1 ng/µL) and one for a higher concentration range (1–10 ng/ $\mu$ L). The amount of volatiles emitted was normalized to the leaf area.

### Effect of herbivore species and developmental stage experiment

To investigate the differences in volatile emission of black polar trees infested with different species of caterpillars and different instars of the same species, five trees were 555 assigned to each of the following treatments: control (undamaged trees), L. dispar second instar herbivory (3000 mg of larval weight -approximately 60 caterpillars, LD2), L. dispar fifth instar (3000 mg of larval weight – 3 to 4 caterpillars, LD5), L. populi fifth instar (3000 mg of 560 larval weight – 3 to 4 caterpillars, LP), mixed herbivory (3000 mg of larval weight 1500 mg for L. populi and 1500 562 for L. dispar – 2 caterpillars of each species, MIX). Caterpillars were weighed, separated by groups and starved the day before the experiment. The experiment was conducted in a climate chamber under the same conditions as 566 described above. At the beginning of the experiment, at 9:00 am the caterpillars were placed on the trees according to treatment. Volatiles were collected during four hours between 48 and 52 h after the herbivores were added. The 570 caterpillars remained on the trees during volatile collection. The experimental setup for volatile collection and filter elution are described above. Qualitative and quanti- 573 tative volatile analysis was conducted using an Agilent 574 6890 Series gas chromatograph coupled to an Agilent 575 5973 quadrupole mass selective detector (interface temp, 576

270°C; quadrupole temp, 150°C; source temp, 230°C; electron energy, 70 eV) and a flame ionization detector (FID) 578 operated at 300°C, respectively. The constituents of the 579 volatile bouquet were separated using a ZB-WAX column 580 (Phenomenex, Aschaffenburg, Germany, 60 m × 0.25 mm 581  $\times$  0.15 µm) and He (MS) or H<sub>2</sub> (FID) as carrier gas. A por-582 tion (1 µl) of the sample was injected without split at an 583 initial oven temperature of 40°C. The temperature was 584 585 held for 2 minutes and then increased to 225°C with a gradient of 5°C/min, held for another 2 minutes and then fur-586 ther increased to 250°C with 100°C/min and a hold for 1 min. Compounds were identified by comparison of 588 retention times and mass spectra to those of authentic 589 standards. The absolute amount of all compounds was de-590 termined based on their FID peak area in relation to the 591 area of the internal standard using the effective carbon 592 number (ECN) concept as described by Scanion and 593 Willis [78]. We restricted our analyses to 20 compounds 594 for which standards were available in high purity (>90%) 595 (Table 1). After termination of the experiment, volatile 596 collections of the caterpillars removed from the leaves 597 along with the frass produced throughout the experiment 598 were performed as described above (Additional file 4: 599 600 Table S1). Leaves from individual trees were harvested separately, and photographed to determine the area of leaf 601 damage as described in [77]. In addition we recorded the number of lesions and damaged leaves per tree. 603

### Statistical analyses

604 All statistical assumptions such as normal distribution 605 and heteroscedasticity were checked. Throughout the 606 manuscript means are always given with standard errors 607 (SE). To determine the importance of volatiles emitted 608 609 from P. nigra in characterizing the different herbivory treatments (L. dispar second instar, L. dispar fifth instar, L. populi fifth instar, and mixed herbivory), we combined 611 the effect of the covariates "number of damaged leaves" and "number of lesions" by performing a principal component analysis for factor reduction as described in [79]. We termed the new variable feeding intensity. Due to 615 the high variability among treatments and the fact that compounds showed different emission patterns, we tested eight models with different variance structures for each compound according to [80]. Model comparison 619 was performed by a maximum likelihood ratio test using the Akaike Information Criterion (AIC) as a measure for the predictive power of the respective statistical model. 622 The model with the lowest AIC value was then selected 623 for the analysis. Table 1 gives an overview of the statis-624 tical models applied. For the selected model we applied 625 a generalized least square model (GLS) to calculate 626 effect of the herbivory treatment (different species, instars and combined damage), the feeding amount and their interaction on the emission of a given compound.

Whenever the herbivory treatment was significantly dif- 630 ferent, we performed a Tukey test for comparison of 631 means on the fitted values. Statistical analyses were 632 performed using R 2.15.2 (R Development Core Team; http://www.R-project.org).

### **Additional files**

Additional file 1: Figure S1. Volatile emission pattern of thirteen further volatiles of Populus nigra foliage representing the major chemical classes released by young trees upon herbivory by fifth instar larvae of Lymantria dispar (gypsy moth) or from undamaged controls over a 4-day experiment. The graphs depict the rates of emission for individual compounds over the course of herbivory (initiated at the beginning of the experiment for herbivory treatment as well as after herbivore removal) during day and night in 6 h intervals. Means + SEM are given at the end of each measuring period.

Additional file 2: Figure S2. Effect of herbivore identity and developmental stage on volatile emission of *Populus nigra* (for thirteen further volatile compounds), Four treatments include Lymantria dispar (2nd instar), L, dispar (5th instar), Laothoe populi (5th instar), and a mixture of L, dispar (5th instar) and L, populi (5th instar), Box-plots showing the same letter are not statistically significant from one another after a Tukey test performed on the fitted values after applying a GLS model, excluding the effect of the feeding intensity, P values are given in Table 1, Plots showing no letters indicate that there was no effect of the treatment on volatile emission.

Additional file 3: Figure S3. Effect of feeding intensity during various herbivory treatments on volatile emission of Populus nigra compounds (for thirteen further volatile compounds), P values indicate significant differences after applying a GLS model (excluding the effect of herbivore identity), p < 0.001 = \*\*\*, p < 0.01 = \*\*, p < 0.05 = \*, ns, = not significant,Herbivory treatments are depicted by different symbols.

**Additional file 4: Table S1.** Mean and  $\pm$  SEM of volatile emission of frass and larvae, after removing them from the respective treatments. Values are expressed as nanograms emitted per gram of fresh weight per hour (ng/mg FW/h), GC-FID retention times for each compound are shown; unidentified compounds are labeled UN ID

### Competing interests

The authors declare that they have no competing interests.

### Authors' contributions

SBU, GAB, ACM and TGK conceived the study and GAB, ACM and SBU designed the experiments; GAB and ACM carried out the experiments, analyzed the data and drafted the manuscript together with SBU. JG substantially revised the manuscript. All authors read and approved the final manuscript.

### Acknowledgments

We thank Beate Rothe, Simone Frommeyer, Elisabeth Fial, Isabell Georgy, and the MPI-ICE greenhouse team for their help in the lab. We also thank Drs. Hannah Nadel and Melody Kenah from the US Department of Agriculture for kindly providing the L. dispar egg clutches and Stefan Bartram from the Department of Bio-organic Chemistry at the MPI-CE for the synthesis of DMNT. We specially thank Daniel Veit, from the Department of Scientific Instrumentation and Utilities Management at the MPI-CE for the design and technical support of the volatile collection systems and Grit Kunert for advice on statistical procedures. This project was funded by the Max Planck Society. Andrea Clavijo McCormick was the recipient of a stipend from the Inter national Max Planck Research School, Jena.

# Received: 16 September 2014 Accepted: 23 October 2014

### References

Baldwin IT: Plant volatiles. Curr Biol 2010, 20(9):392-397.

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- 692 2. Mumm R, Dicke M: Variation in natural plant products and the attraction
   693 of bodyguards involved in indirect plant defense. Can J Zool 2010,
   694 88(7):628-667.
- Arimura G, Matsui K, Takabayashi J: Chemical and molecular ecology of herbivore-induced plant volatiles: proximate factors and their ultimate functions. Plant Cell Physiol 2009, 50(5):911–923.
- 698 4. McCormick AC, Unsicker SB, Gershenzon J: The specificity of herbivore-induced plant volatiles in attracting herbivore enemies. *Trends Plant Sci* 700 2012, 17(5):303–310.
- 701 5. Dicke M, Baldwin IT: The evolutionary context for herbivore-induced plant volatiles: beyond the 'cry for help'. *Trends Plant Sci* 2010, **15**(3):167–175.
- 703 6. Hare JD, Sun JJ: Production of induced volatiles by *Datura wrightii* in
   704 response to damage by insects: effect of herbivore species and time.
   705 J Chem Ecol 2011. 37(7):751–764.
- 706 7. Kessler A, Heil M: The multiple faces of indirect defences and their agents of natural selection. Funct Ecol 2011, 25(2):348–357.
- 708 8. Hoballah ME, Turlings TCJ: The role of fresh versus old leaf damage in the
   709 attraction of parasitic wasps to herbivore-induced maize volatiles.
   710 J Chem Ecol 2005, 31(9):2003–2018.
- 711 9. Loughrin JH, Manukian A, Heath RR, Turlings TCJ, Tumlinson JH: Diurnal
   712 cycle of emission of induced volatile terpenoids herbivore-injured cotton
   713 plants. Proc Natl Acad Sci U S A 1994, 91(25):11836–11840.
- Mattiacci L, Rocca BA, Scascighini N, D'Alessandro M, Hern A, Dorn S:
   Systemically induced plant volatiles emitted at the time of "danger".
   J Chem Ecol 2001, 27(11):2233–2252.
- 717 11. Scascighini N, Mattiacci L, D'Alessandro M, Hern A, Rott AS, Dorn S: New
   718 insights in analysing parasitoid attracting synomones: early volatile
   719 emission and use of stir bar sorptive extraction. Chemoecology 2005,
   720 15(2):97–104.
- Turlings TCJ, Bernasconi M, Bertossa R, Bigler F, Caloz G, Dorn S: The induction of volatile emissions in maize by three herbivore species with different feeding habits: Possible consequences for their natural enemies. *Biol Control* 1998, 11(2):122–129.
- 725 13. Kugimiya S, Shimoda T, Tabata J, Takabayashi J: Present or Past Herbivory:
   726 A screening of volatiles released from *Brassica rapa* under caterpillar
   727 attacks as attractants for the solitary parasitoid, *Cotesia vestalis*. *J Chem Ecol* 2010, 36(6):620–628.
- 729 14. De Moraes CM, Lewis WJ, Pare PW, Alborn HT, Tumlinson JH: Herbivore infested plants selectively attract parasitoids. *Nature* 1998,
   731 393(6685):570–573.
- 732 15. Arimura G, Kost C, Boland W: Herbivore-induced, indirect plant defences.
   733 BBA-Mol Cell Biol L 2005, 1734(2):91–111.
- 734 16. Dudareva N, Picherski E, Gershenzon J: **Biochemistry of plant volatiles.** *Plant Physiol* 2004, **135**(4):1893–1902.
- 736 17. Clavijo McCormick A, Gershenzon J, Unsicker SB: Little peaks with big 737 effects: establishing the role of minor plant volatiles in plant-insect
- interactions. Plant Cell Environ 2014, 37(8):1836–1844.
   Clavijo McCormick A, Irmisch S, Reinecke A, Boeckler A, Veit D, Reichelt M,
   Hansson B, Gershenzon J, Köllner TG, Unsicker SB: Herbivore-induced
- volatile emission in black poplar regulation and role in attracting
   herbivore enemies. Plant Cell Environ 2014, 37(8):1909–1923.
   D'Alessandro M, Brunner V, von Merey G, Turlings TCJ: Strong attraction of
- the parasitoid *Cotesia marginiventris* towards minor volatile compounds of maize. *J Chem Ecol* 2009, **35**(9):999–1008.
- 746 20. Evans KA, Allenwilliams LJ: Electroantennogram responses of the cabbage
   747 seed weevil, Ceutorynchus assimilis, to oilseed rape, Brassica napus spp.
   748 oleifera, volatiles. J Chem Ecol 1992, 18(9):1641–1659.
- Geervliet JBF, Posthumus MA, Vet LEM, Dicke M: Comparative analysis of headspace volatiles from different caterpillar-infested or uninfested food plants of *Pieris* species. J Chem Ecol 1997, 23(12):2935–2954.
- 752 22. Gouinguene S, Degen T, Turlings TCJ: Variability in herbivore-induced odour emissions among maize cultivars and their wild ancestors
   754 (teosinte). Chemoecology 2001, 11(1):9–16.
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- 759 24. Kappers IF, Hoogerbrugge H, Bouwmeester HJ, Dicke M: Variation in herbivory-induced volatiles among cucumber (*Cucumis sativus* L.)
   761 varieties has consequences for the attraction of carnivorous natural enemies. *J Chem Ecol* 2011, 37(2):150–160.

- Krips OE, Willems PEL, Gols R, Posthumus MA, Gort G, Dicke M: Comparison of cultivars of ornamental crop Gerbera jamesonii on production of spider mite-induced volatiles, and their attractiveness to the predator Phytoseiulus persimilis. J Chem Ecol 2001, 27(7):1355–1372.
- Mumm R, Tiemann T, Varama M, Hilker M: Choosy egg parasitoids: Specificity of oviposition-induced pine volatiles exploited by an egg parasitoid of pine sawflies. Entomol Exp et Appl 2005, 115(1):217–225.
- Olson DM, Cortesero AM, Rains GC, Potter T, Lewis WJ: Nitrogen and water affect direct and indirect plant systemic induced defense in cotton. Biol Control 2009, 49(3):239–244.
- De Boer JG, Posthumus MA, Dicke M: Identification of volatiles that are used in discrimination between plants infested with prey or nonprey herbivores by a predatory mite. J Chem Ecol 2004, 30(11):2215–2230.
- Fatouros NE, Huigens ME, van Loon JJA, Dicke M, Hilker M: Chemical communication - Butterfly anti-aphrodisiac lures parasitic wasps. Nature 2005, 433(7027):704.
- Yoneya K, Kugimiya S, Takabayashi J: Can herbivore-induced plant volatiles inform predatory insect about the most suitable stage of its prey? Physiol Entomol 2009, 34(4):379–386.
- de Boer JG, Hordijk CA, Posthumus MA, Dicke M: Prey and non-prey arthropods sharing a host plant: Effects on induced volatile emission and predator attraction. J Chem Ecol 2008, 34(3):281–290.
- 32. Erb M, Foresti N, Turlings TCJ: A tritrophic signal that attracts parasitoids to host-damaged plants withstands disruption by non-host herbivores. BMC Plant Biol 2010, 1(247):1–11.
- Rodriguez-Saona C, Chalmers JA, Raj S, Thaler JS: Induced plant responses to multiple damagers: differential effects on an herbivore and its parasitoid. *Oecologia* 2005, 143(4):566–577.
- Shiojiri K, Takabayashi J, Yano S, Takufuji A: Infochemically mediated tritrophic interaction webs on cabbage plants. Popul Ecol 2001, 34:23–29.
- 35. Zhang PJ, Zheng SJ, van Loon JJA, Boland W, David A, Mumm R, Dicke M: Whiteflies interfere with indirect plant defense against spider mites in Lima bean. *Proc Natl Acad Sci U S A* 2009, **106**(50):21202–21207.
- Gouinguene S, Alborn H, Turlings TCJ: Induction of volatile emissions in maize by different larval instars of Spodoptera littoralis. J Chem Ecol 2003, 29(1):145–162.
- Köpke D, Schroder R, Fischer HM, Gershenzon J, Hilker M, Schmidt A: Does egg deposition by herbivorous pine sawflies affect transcription of sesquiterpene synthases in pine? *Planta* 2008, 228(3):427–438.
- Arimura G, Huber DPW, Bohlmann J: Forest tent caterpillars (Malacosoma disstria) induce local and systemic diurnal emissions of terpenoid volatiles in hybrid poplar (Populus trichocarpa x deltoides): cDNA cloning, functional characterization, and patterns of gene expression of (–)-germacrene D synthase, PtdTPS1. Plant J 2004, 37(4):603–616.
- Blande JD, Tiiva P, Oksanen E, Holopainen JK: Emission of herbivoreinduced volatile terpenoids from two hybrid aspen (*Populus tremula x tremuloides*) clones under ambient and elevated ozone concentrations in the field. *Glob Change Biol* 2007, 13(12):2538–2550.
- Frost CJ, Appel M, Carlson JE, De Moraes CM, Mescher MC, Schultz JC: Within-plant signalling via volatiles overcomes vascular constraints on systemic signalling and primes responses against herbivores. *Ecol Lett* 2007, 10(6):490–498.
- Gossner MM, Weisser WW, Gershenzon J, Unsicker SB: Insect attraction to herbivore-induced beech volatiles under different forest management regimes. *Oecologia* 2014, 176(2):569–580.
- 42. Schaub A, Blande JD, Graus M, Oksanen E, Holopainen JK, Hansel A: Real-time monitoring of herbivore induced volatile emissions in the field. *Physiol Plant* 2010, **138**(2):123–133.
- Staudt M, Lhoutellier L: Volatile organic compound emission from hohn oak infested by gypsy moth larvae: evidence for distinct responses in damaged and undamaged leaves. Tree Physiol 2007, 27(10):1433–1440.
- 44. Tuskan GA, Di Fazio S, Jansson S, Bohlmann J, Grigoriev I, Hellsten U, Putnam N, Ralph S, Rombauts S, Salamov A, Schein J, Sterck L, Aerts A, Bhalerao RR, Bhalerao RP, Blaudez D, Boerjan W, Brun A, Brunner A, Busov V, Campbell M, Carlson J, Chalot M, Chapman J, Chen GL, Cooper D, Coutinho PM, Couturier J, Covert S, Cronk Q, et al: The genome of black cottonwood, Populus trichocarpa (Torr, & Gray). Science 2006, 313 (5793):1596–1604.
- 45. Irmisch S, McCormick AC, Boeckler GA, Schmidt A, Reichelt M, Schneider B, Block K, Schnitzler JP, Gershenzon J, Unsicker SB, Köllner TG: Two herbivore-induced cytochrome P450 enzymes CYP79D6 and CYP79D7 catalyze the

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- formation of volatile aldoximes involved in poplar defense. *Plant Cell* 2013, **25**(11):4737–4754.
- 46. D'Auria JC, Pichersky E, Schaub A, Hansel A, Gershenzon J: Characterization of a BAHD acyltransferase responsible for producing the green leaf volatile (Z)-3-hexen-1-yl acetate in *Arabidopsis thaliana*. *Plant J* 2007, 49(2):194–207.
- De Moraes CM, Mescher MC, Tumlinson JH: Caterpillar-induced nocturnal plant volatiles repel conspecific females. *Nature* 2001, 410(6828):577–580.
- 842 48. Dudareva N, Andersson S, Orlova I, Gatto N, Reichelt M, Rhodes D, Boland W,
   843 Gershenzon J: The nonmevalonate pathway supports both monoterpene
   844 and sesquiterpene formation in snapdragon flowers. Proc Natl Acad Sci U S
   845 A 2005. 102(3):933–938.
- 846 49. Hemmerlin A, Harwood JL, Bach TJ: A raison d'etre for two distinct
   847 pathways in the early steps of plant isoprenoid biosynthesis? *Prog Lipid* 848 *Res* 2012, 51(2):95–148.
- 849 50. Opitz S, Nes WD, Gershenzon J: Both methylerythritol phosphate and mevalonate pathways contribute to biosynthesis of each of the major isoprenoid classes in young cotton seedlings. *Phytochemistry* 2014, 98:110–119.
- 853 51. Li ZR, Sharkey TD: Metabolic profiling of the methylerythritol phosphate
   854 pathway reveals the source of post-illumination isoprene burst from
   855 leaves. Plant Cell Environ 2013, 36(2):429–437.
- 856
   852. Arimura GI, Kopke S, Kunert M, Volpe V, David A, Brand P, Dabrowska P,
   857 Maffei ME, Boland W: Effects of feeding Spodoptera littoralis on lima
   858 bean leaves: IV. Diurnal and nocturnal damage differentially initiate
   859 plant volatile emission. Plant Physiol 2008, 146(3):965–973.
- 860 53. Cai XM, Sun XL, Dong WX, Wang GC, Chen ZM: Variability and stability of tea weevil-induced volatile emissions from tea plants with different weevil densities, photoperiod and infestation duration. *Insect Sci* 2012, 19(4):507–517.
- 864 54. Huang J, Cardoza YJ, Schmelz EA, Raina R, Engelberth J, Tumlinson JH:
   Differential volatile emissions and salicylic acid levels from tobacco
   plants in response to different strains of Pseudomonas syringae.
   Planta 2003, 217(5):767-775.
- 868 55. Martin DM, Gershenzon J, Bohlmann J: Induction of volatile terpene
   869 biosynthesis and diurnal emission by methyl jasmonate in foliage of
   870 Norway spruce. Plant Physiol 2003, 132(3):1586–1599.
- 871 56. Pio CA, Silva PA, Cerqueira MA, Nunes TV: Diurnal and seasonal emissions
   872 of volatile organic compounds from cork oak (*Quercus suber*) trees.
   873 Atmosph Environ 2005, 39(10):1817–1827.
- 874 57. Rose USR, Lewis WJ, Tumlinson JH: Specificity of systemically released
   875 cotton volatiles as attractants for specialist and generalist parasitic
   876 wasps. J Chem Ecol 1998, 24(2):303–319.
- 877 58. Turlings TCJ, McCall PJ, Alborn HT, Tumlinson JH: An elicitor in caterpillar
   878 oral secretion that induces corn seedlings to emmit chemical signals
   879 attractive to parasitic wasps. J Chem Ecol 1993, 19(3):411–425.
- 880 59. Alborn HT, Hansen TV, Jones TH, Bennett DC, Tumlinson JH, Schmelz EA,
   881 Teal PEA: Disulfooxy fatty acids from the American bird grasshopper
   882 Schistocerca americana, elicitors of plant volatiles. Proc Natl Acad Sci U S A
   883 2007, 104(32):12976–12981.
- 884 60. Felton GW, Eichenseer H: Herbivore saliva and induction of resistance to herbivores and pathogens. In Induced Plant Defenses against Pathogens and Herbivores: Biochemistry, Ecology and Agriculture. Edited by Agurwal A, Tuzun S, Bent E. St. Paul, MN, USA: Phytopathological Society; 1999:19–36.
- 888 61. Funk CJ: Alkaline phosphatase activity in whitefly salivary glands and saliva. Arch Insect Biochem Physiol 2001, 46(4):165–174.
- 890 62. Mattiacci L, Dicke M, Posthumus MA: Beta-glucosidase: an elicitor of herbivore-induced plant odor that attracts host-searching parasitic wasps. *Proc Natl Acad Sci U S A* 1995, 92(6):2036–2040.
- 893 63. De Boer JG, Snoeren TAL, Dicke M: Predatory mites learn to discriminate
   894 between plant volatiles induced by prey and nonprey herbivores.
   895 Anim Behav 2005, 69:869–879.
- 896 64. Dicke M, Poecke R: Signalling in plant-insect interactions: signal transduction in direct and indirect plant defence. In Plant Signal Transduction. Edited by D
   898 Scheel CW. Oxford, UK: Oxford University Press; 2002:289–316.
- 899 65. Walling LL: The myriad plant responses to herbivores. J Pl Growth Regul 2000, 19(2):195–216.
- 901 66. Bostock RM: Signal crosstalk and induced resistance: Straddling the line
   902 between cost and benefit. In Ann Rev Phytopath, Volume 43. Palo Alto:
   903 Annual Reviews; 2005:545–580.

- Fatouros NE, Van Loon JJA, Hordijk KA, Smid HM, Dicke M: Herbivoreinduced plant volatiles mediate in-flight host discrimination by parasitoids. J Chem Ecol 2005, 31(9):2033–2047.
- Takabayashi J, Takahashi S, Dicke M, Posthumus MA: Developmental stage of herbivore *Pseudaletia separata* affects production of herbivoreinduced synomone by corn plants. *J Chem Ecol* 1995, 21(3):273–287.
- Reader T, Hochuli DF: Understanding gregariousness in a larval Lepidopteran: the roles of host plant, predation, and microclimate. Ecol Entomol 2003, 28(6):729–737.
- Dicke M, van Loon JJA, Soler R: Chemical complexity of volatiles from plants induced by multiple attack. Nat Chem Biol 2009, 5(5):317–324.
- 71. Vet LEM, Dicke M: Ecology of infochemical use by natural enemies in a tritrophic context. *Annu Rev Entomol* 1992, **37**:141–172.
- Poirie M, Carton Y, Dubuffet A: Virulence strategies in parasitoid Hymenoptera as an example of adaptive diversity. C R Biol 2009, 332(2–3):311–320.
- Vinson SB: Host selection by insect parasitoids. Annu Rev Entomol 1976, 21:109–133.
- Unsicker SB, Kunert G, Gershenzon J: Protective perfumes: the role of vegetative volatiles in plant defense against herbivores. Curr Opin Pl Biol 2009, 12(4):479–485.
- 75. Heil M, Silva Bueno JC: Within-plant signaling by volatiles leads to induction and priming of an indirect plant defense in nature. *Proc Natl Acad Sci U S A* 2007, **104**(13):5467–5472.
- Vickers CE, Gershenzon J, Lerdau MT, Loreto F: A unified mechanism of action for volatile isoprenoids in plant abiotic stress. Nat Chem Biol 2009, 5(5):283–291.
- Boeckler GA, Towns M, Unsicker SB, Mellway RD, Yip L, Hilke I, Gershenzon J, Constabel CP: Transgenic upregulation of the condensed tannin pathway in poplar leads to a dramatic shift in leaf palatability for two tree-feeding Lepidoptera. J Chem Ecol 2014, 40(2):150–158.
- 78. Scanion JT, Willis DE: Calculation of flame ionization detector relative response factors using the effective carbon number concept.

  J Chromatogr Sci 1985, 23:333–340.
- 79. Fodor Kl: A Survey of Dimension Reduction Techniques. Livermore, CA, USA:
  Center for Applied Scientific Computing, Lawrence Livermore National
  Laboratory; 2002.
- 80. Zuur AF, İeno EN, Walker N, Saveliev AA, Smith GM: Mixed Effect Models and Extensions in Ecolgy with R. Springer Science + Business Media, LLC; 2009.

### doi:10.1186/s12870-014-0304-5

Cite this article as: Clavijo McCormick *et al.*: The timing of herbivore-induced volatile emission in black poplar (*Populus nigra*) and the influence of herbivore age and identity affect the value of individual volatiles as cues for herbivore enemies. *BMC Plant Biology* 2014 14:304.

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