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Trends

Progress in correlation spectroscopy at ultra-fast magic-angle spinning: Basic building blocks and complex experiments for the study of protein structure and dynamics

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ABSTRACT

Recent progress in multi-dimensional solid-state NMR correlation spectroscopy at high static magnetic fields and ultra-fast magic-angle spinning is discussed. A focus of the review is on applications to protein resonance assignment and structure determination as well as on the characterization of protein dynamics in the solid state. First, the consequences of ultra-fast spinning on sensitivity and sample heating are considered. Recoupling and decoupling techniques at ultra-fast MAS are then presented, as well as more complex experiments assembled from these basic building blocks. Furthermore, we discuss new avenues in biomolecular solid-state NMR spectroscopy that become feasible in the ultra-fast spinning regime, such as sensitivity enhancement based on paramagnetic doping, and the prospect of direct proton detection.

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1. Introduction

The last decade has seen tremendous technological and methodological developments in the field of magic-angle spinning (MAS) solid-state NMR (ssNMR) [1,2]. These developments have allowed for the structural characterization of many samples of high biological interest: microcrystalline proteins, amyloid fibrils, oligomeric assemblies, and membrane proteins in a native-like

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environment. Part of those developments can be attributed to the widespread adoption of high static magnetic fields.

At the same time, fast spinning MAS probe-heads of a new generation have been developed and are now commercially available. This promises to further improve the resolution and sensitivity of biological solid-state NMR. Additionally in the ultrafast spinning regime, fundamentally new applications have become feasible. In this review, we will discuss these recent improvements: the study of paramagnetic metal binding centers, new methods for resonance assignment and collection of longrange distance restraints, proton detection at ultra-fast MAS, and the study of protein dynamics.

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It should be noted that complementary information to the current article is available. In a review by Anne Lesage [1], the advantages of fast MAS are presented in the context of organic solid materials along with recent advances in structural investigations. The mechanical aspect of fast rotation and the impact of coil dimension on radio-frequencies (RF) are discussed in a recent chapter of the Encyclopedia of Magnetic Resonance by Ago Samoson and coworkers [3]. Another chapter of the Encyclopedia of Magnetic Resonance by Donghua H. Zhou [4] introduces fast MAS in combination with proton detection and paramagnetic doping.

In this review, we will distinguish between three regimes of spinning frequencies: low MAS below 20 kHz, fast MAS between 20 and 40 kHz, and ultra-fast MAS above 40 kHz. So far, ultra-fast MAS can be attained with rotors of reduced size: 40–50 kHz MAS for rotors of outer diameter (o.d.) 1.8 mm and 1.6 mm, and 60–70 kHz MAS for rotors of o.d. 1.3 mm and 1.2 mm. More recently, MAS frequencies of up to 80 kHz have been reached by rotors with an o.d. of 1.0 mm (JEOL, Japan) [5]. For comparison purposes, the characteristics of some commercially available rotors (Fig. 1a) are summarized in Table 1. The reduction in rotor dimensions has important repercussions with respect to sensitivity, RF performance, and sample heating.

In NMR, sensitivity per unit square root time $(S/N)_t$, or more simply sensitivity, is generally defined as the signal to noise ratio (S/N)in the frequency domain per unit square root time (Eq. (1)) [7]. It is closely related to the time-domain S/N directly following a 90° pulse, which has been described in a closed-form expression by Hoult and Richards [8]. The signal part (Eq. (2)) depends notably on the sample volume V_{sample} and on the coil sensitivity $(B_1)_{xy}$, defined as the magnitude of the transverse magnetic field induced in the sample by a current of 1 A flowing in the RF coil [9]. In ultra-fast MAS rotors. the large reduction in active volume expectedly results in a reduction of the absolute signal. Contrarily, for a solenoid coil, the coil sensitivity $(B_1)_{xy}$ is improved with smaller diameters (Eq. (3)), reflecting an improvement of the filling factor. The noise amplitude σ_{noise} (Eq. (4)) depends on the spectral bandwidth Δf , and resistance losses R_{noise} (conductive, magnetic, and dielectric losses) which are mostly dictated by the wire geometry of the receiver coil. If we make the fair assumption that the height to radius ratio is conserved among the different coils, then noise amplitude is unaffected by the rotor diameter. The outcome is that the absolute S/N is roughly proportional to the rotor diameter, as described in Refs. [9,10]. The S/N relative to the amount of material is improved in rotors of smaller diameter, which is advantageous for mass-limited samples (Fig. 1b).

$$(S/N)_t \equiv \frac{S}{\sigma_{noise}\sqrt{T_{tot}}} \tag{1}$$

S is the frequency domain signal peak amplitude, σ_{noise} is the frequency domain R.M.S. noise amplitude, and T_{tot} is the total experimental time.

$$(S/N)_t \propto V_{\text{sample}}(B_1)_{xy} M_0 \omega_0 K$$
 (2)

 $(B_1)_{xy}$ is the sensitivity of the RF coil, M_0 is the initial magnetization, V_{sample} is the sample volume, ω_0 is the Larmor frequency, and K is the RF inhomogeneity factor.

$$(B_1)_{xy} = \frac{\mu_0 n}{2} \frac{1}{\sqrt{r_{coil}^2 + ((1/2)h_{coil})^2}}$$
(3)

 μ_0 is the vacuum permeability $(4\pi \times 10^{-7} \text{NA}^{-2})$, n is the number of wire turns in the solenoid, and r_{coil} and h_{coil} are the radius and height of the solenoid coil, respectively.

$$\sigma_{\text{noise}} = \sqrt{4k_B T_c R_{\text{noise}} \Delta f} \tag{4}$$

 k_B is the Boltzmann constant, T_c is the coil temperature, R_{noise} is the resistance losses, and Δf is the bandwidth of the receiver.

The high coil sensitivity $(B_1)_{xy}$ also entails high RF performance for ultra-fast MAS probe-heads, meaning that large RF fields can be generated by the coil without requiring a large input power from the amplifiers [4]. However, such strong RF irradiation has the drawback of depositing a high amount of energy in the sample. For instance, rapidly oscillating electric fields will bring free charge carriers in motion and will induce the realignment of molecular dipoles [11]. The resistance of the environment to such motion draws energy from the electric field; a sample with high resistance is termed "lossy". The situation is particularly critical in the study of biological samples [12,13], which are fragile and often preserved in ionic buffers or in a lipid environment. Eventually, the heat contributed can lead to irremediable alteration of the sample, for instance degradation, denaturation, or aggregation. Another cause of concern is the effect of heating on experimental results. RF absorption and heat dissipation depend on numerous factors, crippling reproducibility of results if different pulse sequences, sample conditions, or instruments are

Table 1Comparison of MAS rotor sizes.
Data according to Ref. [6].

Outer diameter (mm)	7	4	3.2	2.5	2.5	1.3	1.6	1.2
Inner diameter (mm)	5.6	3.0	2.2	1.7	1.3	0.9		
Sample volume ^a (μL)	246	70	30	14	8	1.7	8.1	1.2
Max. spinning frequency (kHz)	7	15	24	35	35	67	45	65
Manufacturer	Bruker Biospin						Agilent	

^a For Bruker Biospin rotors, the sample volume refers to the active sample volume inside the NMR RF coil (data provided by Dr. Frank Engelke, Bruker Biospin, Germany); for Agilent Technologies, it refers to the total sample volume.

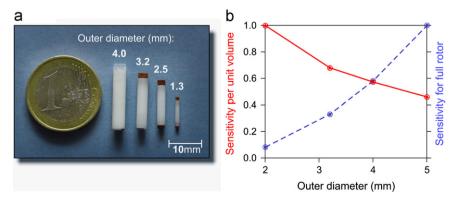


Fig. 1. Comparison of MAS rotors. (a) MAS rotors of 1.3 mm o.d. have an active volume of 1.7 μL and can spin up to 67 kHz. (b) While larger rotors are more sensitive, (total sensitivity $(S/N)_t$, blue) small rotor sizes exhibit a higher sensitivity per unit volume $((S/N)_t/V_{sample}, red)$. Figure adapted from Ref. [15].

used. The heterogeneity of spectroscopic observables due to spatial temperature gradients is exacerbated at strong RF fields and can persist for several hundreds of milliseconds after the heating pulse [13]. Small rotors are beneficial in this respect, as temperature gradients due to irradiation increase as the fourth power of the coil diameter [14].

To overcome the transient heat deposition by RF, one strategy is the development of pulse sequences requiring minimal amounts of irradiation power. Another strategy is to change the design of the RF coil in order to reduce the generation of timevarying electric fields during irradiation. Multiple designs have been introduced to replace or improve the typically employed solenoidal coils and are discussed in a review by Christopher V. Grant and coworkers [16]. Designs which have been combined with MAS include the scroll coil [17], the cross coil/solenoid [14], the Z coil [18], the loop-gap loaded coil [13], and "low-E" coils [19]. Both of those strategies directly improve spectral resolution since they allow longer acquisition times to be tolerated.

An additional cause of heating in MAS solid-state NMR is the power loss due to friction between the pressured gas and the bearings. The power loss is proportional to the square of the spinning frequency [20] and can lead to temperature increases of 40–60 °C at 60 kHz MAS in comparison with the static case [4]. In contrast to RF heating, frictional heating remains uniform over time, although non-uniform spatially. It is likely that cooling gas remedies frictional heating more effectively than it does RF heating [14]. Because of the large temperature difference, the flow and temperature of the cooling gas have to be gradually adapted during spinning-up and spinning-down of the rotor. For biological

samples, a facile temperature calibration can be done on the water ¹H signal in combination with internal DSS chemical shift referencing [21].

The effects of high spinning rates are decisive in preserving a competitive sensitivity compared to larger rotors. For resonances with large chemical shift anisotropies, the very fast rotation has the consequence of moving spinning sidebands out of the spectral window, increasing the intensity of the center band [10,22]. Homonuclear dipolar couplings are as well greatly reduced (see Fig. 8 and Section 5 on Proton NMR at ultra-fast MAS), resulting in narrower and more intense lines [23]. Pulse sequences developed for ultra-fast MAS promise further improvements in sensitivity and resolution by employing four complementing strategies: (1) increased transfer to cross-peaks during magnetization mixing steps, (2) reduced decay of spin-locked and transverse magnetization, (3) low-power sequences which enable fast recycling and long acquisition times, (4) proton detection. The application of ultra-fast MAS techniques to the study of proteins and other biomolecules is discussed in the following sections.

2. Protein resonance assignment and structural characterization

Achieving a complete assignment of the NMR signals constitutes the first step in structural studies using solid-state NMR and still stands as a major obstacle in obtaining site-specific structural information. Usually, a combination of two-dimensional correlation spectra is required for the sequential resonance assignment

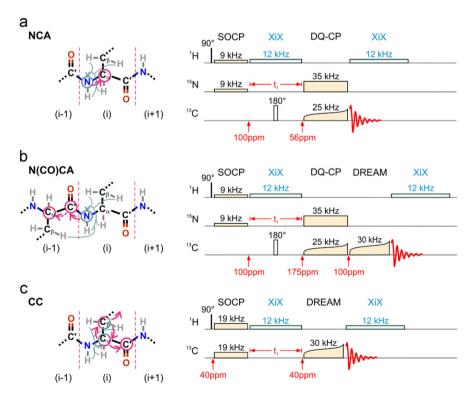


Fig. 2. Complete set of low-power experiments for assignment of uniformly [13 C, 15 N]-labeled proteins. (*Left panels*) Assignment of backbone N–C correlation proceeds through (a) NCA for intra-residue correlations and (b) N(CO)CA for correlation to carbons of the preceding residue. (c) Intra-residue side-chain 13 C correlations are obtained from the CC spectrum. (*Right panels*) Corresponding pulse schemes. Initial 1 H to 15 N transfer is carried out through low-power SOCP at 9 kHz. The RF strength of SOCP is increased to 19 kHz in the CC experiment as in Ref. [44] in order to excite the full 13 C aliphatic region. For N–C experiments, J_{NC} —couplings are removed in the indirect dimension by application of a 180 ° pulse midway through t_1 evolution. Adiabatic shaped pulses are employed on the 13 C channel for SPECIFIC-CP and DREAM recoupling elements; the indicated frequency corresponds to the average RF strength. Low-power XiX decoupling is applied during t_1 evolution and acquisition. The 13 C carrier position is changed during experiments, indicated by red arrows. Figure adapted from Ref. [41].

of uniformly [13 C, 15 N]-labeled proteins. For larger systems, often assignment ambiguities remain and 3D or even 4D solid-state NMR experiments need to be performed [24 - 27].

At low-MAS frequencies (\sim 7–13 kHz), the standard repertoire of experiments for protein resonance assignment consists of homonuclear ¹³C-¹³C correlation experiments such as protondriven spin-diffusion (PDSD) [28] or alternatively RAD/DARR [29,30] as well as heteronuclear ¹⁵N-¹³C correlation experiments such as NCACX or NCOCX. In the latter case, the transfer from C_{α} or C' to CX often relies on PDSD or DARR. However, PDSD and DARR become ineffective at high spinning frequencies and at high static magnetic fields. In the case of ultra-fast MAS, the MIRROR [31], RESORT [32], and PARIS [33,34] experiments can be used analogously to PDSD and DARR (see Section 4 on Dipolar recoupling at ultra-fast MAS). As well, Hou and coworkers reported homonuclear 13C-13C spin-diffusion experiments based on R-symmetry sequences [35]. They found that such experiments work well at fast MAS frequencies of 40 kHz where conventional PDSD or DARR experiments already fail.

Several first-order recoupling sequences for $^{13}C^{-13}C$ transfer have been introduced which work well at ultra-fast spinning, for example DREAM [36] providing double-quantum transfer, or RFDR [37,38] providing zero-quantum transfer. A general difference between PDSD which is a second-order process and first-order dipolar recoupling experiments such as DREAM and RFDR is that the latter ones are strongly influenced by dipolar truncation [39]. Here, transfer is restricted to the strongest dipolar couplings present, usually between directly bonded nuclei, e.g. C_{α} and C_{β} . Relying only on first-order sequences therefore restricts the number of observed correlations. The detection of sequential transfer, for instance between C_{α} nuclei of adjacent residues, feasible with PDSD [40], is therefore difficult to achieve with first-order recoupling sequences.

Nevertheless, it is in principle possible to compile a set of experiments suitable for the complete resonance assignment of uniformly labeled proteins at ultra-fast MAS. For instance, such a complete set of experiments was introduced by Vijayan et al. and is depicted in Fig. 2 [41]. The set comprises CC, NCA, and N(CO)CA two-dimensional correlation experiments. Applications to ubiquitin and tau paired helical filaments are described in Ref. [41]. After resonance assignments have been obtained, ¹³C and ¹⁵N chemical shifts can be readily analyzed in terms of secondary structure using secondary chemical shifts [42] or statistical database approaches such as TALOS [43].

Experiments for gathering distance restraints at ultra-fast MAS are still scarce. Since approaches based on PDSD will fail under ultra-fast MAS conditions, other strategies have to be pursued. One successful approach has been presented by Lewandowski et al. [45]. In this study, the authors demonstrated that the PAR experiment works even at an MAS frequency of 65 kHz. Application to GB1 yielded highly resolved 2D spectra which allow for the extraction of long-range ¹³C-¹³C proximities.

Another promising approach for the detection of long-range restraints has recently been reported by Huber et al. [46]. This approach relies on the measurement of long-range proton-proton distances in extensively deuterated samples with proton-detected 3D and 4D correlation experiments. The method utilizes DREAM mixing for efficient proton-proton transfer and was applied to a sparsely methyl- and amide-labeled ubiquitin sample. Simultaneously, a very similar approach has also been presented by Linser et al. [47].

Meier and coworkers also demonstrated that $^{3h}J_{NC'}$ hydrogenbond scalar couplings can be directly detected at ultra-fast MAS (55–57 kHz) using extensively deuterated samples [48]. The measurement of these sub-Hertz scalar couplings currently remains challenging but the approach is potentially very useful

for structural biology applications and could further research on $\beta\mbox{-sheet-rich}$ amyloid fibrils.

3. Paramagnetic doping and paramagnetic proteins

As previously mentioned, the ultra-fast spinning regime has the advantage that entirely low-power sequences can be applied [41,49]. While low-MAS heteronuclear decoupling sequences are a priori viable at faster MAS, an important realization from the beginning of the last decade was that commensurate or improved decoupling performances are realized at low RF powers in the ultra-fast MAS regime [50]. The sequences which are currently considered to offer the best decoupling are low-power XiX [51], low-power TPPM [52], swept low-power TPPM [53], and low-amplitude PISSARRO [54]. In practice, more than one decoupling sequence should be tested, as the relative performance can fluctuate depending on the MAS frequency, static magnetic field B_0 , and on the molecular system, similar to what was investigated at low MAS [55,56].

Since only weak RF fields in the order of 10–20 kHz are used for heteronuclear 1 H decoupling, the strain on the instrumentation and the heat deposition are mitigated. Multi-dimensional spectra can be recorded with longer acquisition times and faster repetition rates compared with experiments at low spinning frequencies and high-power decoupling. For instance, an extremely well-resolved band-selective INADEQUATE- 3 E spectrum of microcrystalline GB1 is reported in Ref. [57], recorded with maximum acquisition times of 37 ms in t_{1} and 50 ms in t_{2} at 60 kHz MAS on a 1 GHz (23.5 T) spectrometer.

Such low-power approaches for resonance assignment and detection of structural restraints can be combined with paramagnetic optimized relaxation times [58,59]. The recycle delay can then be reduced and the resulting sensitivity enhancement compensates for the loss of sensitivity due the small sample volume of fast-spinning MAS rotors (Fig. 3). For this purpose, paramagnetic dopants such as Cu–EDTA are added in

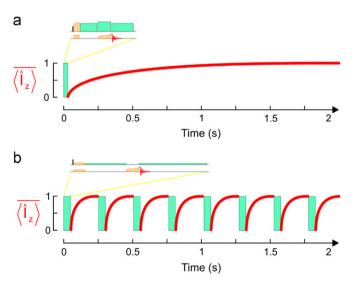


Fig. 3. Fast repetition rates and paramagnetic doping. (a) In traditional acquisition, the duration of the inter-scan delay is dictated by the delay necessary for heat dissipation, and by the recovery rate of proton longitudinal magnetization, as depicted by the red curve. (b) In condensed acquisition, the experiment can be repeated at a faster rate, approximately an order of magnitude faster, owing to the acceleration of ¹H longitudinal relaxation rate by paramagnetic dopants. The use of entirely low-power sequences prevents excessive deposition of heat and allows for longer acquisition times, therefore increasing resolution. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

concentrations of 10–250 mM. This shortens the 1 H T_1 relaxation times down to 50–100 ms [60]. Reif and coworkers have proposed to combine ultra-fast MAS, paramagnetic doping, and deuteration [12]. Nadaud and coworkers have demonstrated that a similar approach comprising 2D and 3D correlation spectra can also be applied to proteins with covalently bound paramagnetic tags [61].

Furthermore, fast recycling experiments can be readily applied to paramagnetic metalloproteins. For example, a 2D ¹³C-¹³C correlation spectrum of the oxidized, paramagnetic form of human copper zinc superoxide dismutase (SOD) could be acquired with a shortened recycle delay of 500 ms [62]. Bertini et al. reported the application of ultra-fast MAS and fast recycling protocols to detect residues as close as 5.6 Å from the Co^{II} in MMP-12 [63]. This opens the way to characterize the metal coordination environment of metalloproteins by ssNMR which was previously unobservable at lower spinning frequencies due to large and highly anisotropic paramagnetic chemical shifts.

4. Dipolar recoupling at ultra-fast MAS

The standard building units for multi-dimensional ssNMR experiments are homonuclear and heteronuclear recoupling sequences, along with periods of evolution where decoupling is often required. Magnetization transfer can be mediated by a choice of mechanisms: through space via dipolar couplings, or through chemical bonds via J-couplings. The progress made in J-based sequences, see Refs. [57,64] for instance, has been recently discussed [1,65,66] and will not be considered here in detail. As previously mentioned, in many cases the homonuclear dipolar recoupling schemes must be adapted, as the recoupling sequences that are employed at low MAS are either not efficient anymore at ultra-fast MAS or their RF requirements are too elevated for the current instruments and samples. For example, symmetry-based pulse sequences such as SPC5 [67] or C7 and POST-C7 [68,69] require an irradiation of, respectively, 5 and 7 times the MAS frequency on the low-frequency channel. An important advantage of ultra-fast MAS is that ¹H decoupling can be omitted in many recoupling sequences such as DREAM [49], RFDR [70], CMpRR [71], and ¹⁵N-¹³C SPECIFIC-CP [41,60]. Another important advance in recent years is the emergence and better understanding of second-order recoupling sequences. The present section will highlight the particularities of dipolar recoupling techniques at ultra-fast MAS. We will discuss separately firstorder and second-order recoupling techniques. Here, first-order indicates that the effect occurs due to the first term of the Magnus expansion in the average Hamiltonian treatment of the pulse sequence [72], while second-order effects arise due to the second term (commutator terms) of the Magnus expansion.

4.1. First-order dipolar recoupling: homonuclear

First-order dipolar recoupling experiments fall into two categories depending on whether their effective Hamiltonian is of double-quantum (DQ) or zero-quantum (ZQ) nature [73]. The important consequence is that the transferred magnetization will change its sign for DQ transfer but not for ZQ. For DQ mixing, this makes it possible to distinguish between direct transfers, which have negative cross-peaks relative to the diagonal, and relay transfers (i.e. two subsequent transfers during the same mixing period) which have positive cross-peaks.

Because of its low RF requirements, the double-quantum homonuclear rotary-resonance condition, DQ-HORROR [74], is frequently employed (Eq. (5)). In this sequence, cw irradiation is applied on the X channel at approximately $\frac{1}{2}v_r$, such that the sum of effective fields on A and B equals the MAS frequency. Here, X represents the

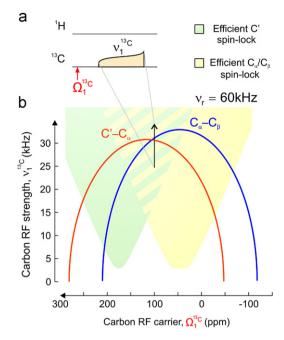


Fig. 4. Optimization of 13 C $^{-13}$ C DREAM transfer. (a) The DREAM recoupling element is characterized by the carrier position $\Omega_1^{^{13}}$ C and the average RF strength $\nu_1^{\rm X}$. At ultra-fast MAS, no $^{^{1}}$ H decoupling is applied during recoupling. (b) Calculated transfer conditions for C'- C_{α} transfer (orange) and C_{α} - C_{β} transfer (blue). Transfer occurs at different RF strengths because of their different chemical shift separation, as calculated from Eq. (5). Typical $^{^{13}}$ C isotropic chemical shifts are considered: C' (175.8 ppm), C_{α} (57.5 ppm) and C_{β} (34.6 ppm). The black arrow represents the amplitude of the RF field during a DREAM adiabatic sweep with carrier position at 100 ppm. Recoupling can be achieved off-resonance but the spin-lock is inefficient, leading to magnetization loss. Approximate regions of efficient spin-lock, as observed in ubiquitin at 18.8 T, are highlighted in green for C' and in yellow for C_{α}/C_{β} . The mixing duration for DREAM is optimized between 2 and 10 ms.

nucleus for which recoupling occurs (e.g. ¹³C and ¹⁵N). The recoupling is commonly realized with the adiabatic version of DQ-HORROR, namely the DREAM scheme [36], which was first presented as a DQ filter scheme [75]. The initial magnetization is spin-locked with an applied RF of amplitude far inferior or far superior to the HORROR condition. The RF strength is then gradually changed, effectively 'dragging' the spin-system adiabatically through the recoupling condition (Fig. 4a). This makes the sequence robust against RF mismatch and distribution of dipolar coupling frequencies due to crystallite orientation and isotropic chemical shift distribution.

$$v_r = \underbrace{\sqrt{v_1^2 + (\Omega_{iso}^A)^2}}_{\text{effective field on spin A}} + \underbrace{\sqrt{v_1^2 + (\Omega_{iso}^B)^2}}_{\text{effective field on spin B}}$$
 (5)

 v_r is the MAS frequency, v_1 is the RF nutation frequency, and Ω_{iso} is the chemical shift offset relative to carrier.

DREAM is narrow-banded at low MAS and broadband at ultrafast MAS, as it can recouple two spins with an isotropic chemical shift separation of up to $\frac{1}{2}\nu_r$. This feature is nicely illustrated in the work of Ernst et al. [49], where $^{13}\text{C}-^{13}\text{C}$ cross-peaks with increasing chemical shift separation appear as the spinning rate is increased from 30 to 60 kHz. This work also demonstrates that above a given spinning rate (\geq 50 kHz MAS), cross-peak intensities are unaffected by the presence or absence of ^1H decoupling. The RF field strength employed for the DREAM shaped pulse depends on both the carrier position Ω_1^{Υ} and the chemical shift separation of the recoupled spin pair. Fig. 4b illustrates the different RF fields required for C_{α} –C' and C_{α} – C_{β} , transfers. Practically, the best transfer conditions are obtained with the carrier

frequency set to the middle of the range of resonances to be recoupled and the average irradiation frequency slightly below $\frac{1}{2}\nu_r$ [36]. Due to the distribution of chemical shifts in proteins, different spin pairs cross the resonance condition at different times during the adiabatic sweep [76]. For example, as seen from the arrow depicted in Fig. 4b, the $C'-C_\alpha$ condition is traversed before the $C_\alpha-C_\beta$ condition. This leads to an intricate pattern of magnetization transfer and asymmetric cross-peak intensities in 2D spectra.

New DQ recoupling schemes have also been recently introduced for the study of $^{19}F^{-19}F$ proximities [77,78] or for ^{31}P recoupling [79]. Symmetry-based techniques are utilized in these schemes in order to be more robust against the broad range of ^{19}F chemical shifts, or against the large ^{31}P CSA.

The sign alternation is not encountered when using ZQ recoupling, of which the chief sequence at ultra-fast MAS is RFDR. In this sequence, a train of rotor-synchronized π (180°) pulses is applied on the X channel in the middle of every rotor period (Fig. 5b). RFDR has been firstly analyzed considering π pulses as delta functions [37,38], where dipolar couplings are restored only in an interaction frame which involves both RF and isotropic chemical shifts. However, it was then found that when finite pulse-width effects are considered, an interaction frame involving the RF is sufficient to re-introduce the dipolar coupling, making RFDR largely insensitive to chemical shift offsets [81,82]. RFDR is versatile in the slow and fast MAS regimes (<40 kHz), as it can be used in broadband or band-selective experiments. The band-specific implementation requires weaker-RF π pulses which occupy a larger fraction of the rotor period. In a study by Bayro and coworkers [83], this specificity has been combined with sparse labeling in order to reduce the number of actively recoupled nuclei and circumvent dipolar truncation. A large amount of aliphatic correlations, including long-range C_{α} - C_{α} contacts, could be obtained at 12.5 kHz MAS in [2-13C]glycerol-labeled PI3-SH3 amyloid fibrils. At ultra-fast MAS, RFDR is used as a broadband sequence

Homonuclear recoupling

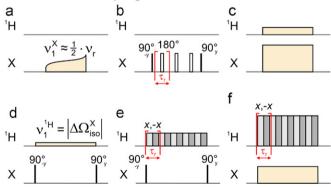


Fig. 5. Building blocks for homonuclear recoupling at ultra-fast MAS. Representation of pulse sequence elements for homonuclear recoupling and spin diffusion at ultra-fast MAS: (a)-(b) first-order sequences and (c)-(f) second-order sequences. Initial magnetization on the x-axis of a first X-nuclei spin is transferred to x-axis magnetization of a second spin of the same isotope. Continuous-wave irradiation is denoted by yellow pulses and phase-alternated irradiation (x, -x) by light and dark gray blocks. For both channels, the carrier position is set in the mid-range $\Omega_1^X = \frac{1}{2}(\Omega_{min}^X + \Omega_{max}^X)$ of frequencies to be recoupled. (a) In DREAM, v_1^X is the average RF strength at midpoint during the sweep, as selected from Fig. 4b. Rotorsynchronized pulses are employed for (b) RFDR, (e) PARIS, and (f) RESORT, with the period indicated in red (n.b. $\tau_r = v_r^{-1}$). For RFDR, the pulse is centered during one rotor period and $au_{pulse} \ll au_r$. For PARIS and RESORT, the duration for each pulse is half the rotor period. (c) For ¹³C-¹³C PAR at 65 kHz MAS, a low-power condition of $(v_1^{12}C \approx 1.12v_r, v_1^{14} \approx 0.3v_r)$ can be exploited for biological samples. (d) In MIRROR-SD, $|\Delta\Omega_{iso}^X|$ refers to the isotropic chemical shift separation between the two low-y nuclei of interest. More information on pulse settings for PAR and RESORT is available in Ref. [45,80,32], respectively. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article.)

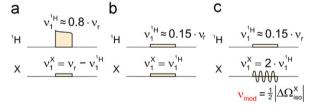
and 1H decoupling is generally omitted [62]. Proton irradiation can reintroduce depolarization conditions which are only avoided through a careful selection of 1H RF strength, with independent levels chosen for the duration of the π pulse and for the window between pulses [70]. Proper phase cycling of the π pulses is critical to eliminate residual DQ dipolar terms or chemical shift terms. Usually, supercycles of the XY – 4 scheme (XY – 8, XY – 16, XY – 32) are employed [81–83]. These phase cycling schemes are beneficial in attenuating the imperfections introduced by the large number of consecutive π pulses.

4.2. First-order dipolar recoupling: heteronuclear

For sensitivity reasons, the initial polarization in ssNMR experiments originates from protons. It is then transferred to low-γ nuclei (e.g. ¹³C or ¹⁵N). Hartmann–Hahn cross-polarization (CP) is the most common technique for this transfer [84,85]. The ZO and DO recoupling conditions used in Hartmann-Hahn CP are described in Fig. 7a. The use of an adiabatic ramp is recommended to increase transfer efficiency [86–88], for reasons similar to those previously exposed for the DREAM scheme. While the RF strengths employed must be sufficiently strong to spin-lock the desired chemical shift ranges, a special attention must be paid to minimize the power deposition. The DO n=1 condition constitutes the first-order recoupling condition with the lowest RF requirements. In a work by Laage et al. [89], this condition is used in a band-selective fashion in order to excite either the carbonyl or the aliphatic 13 C region of human SOD. RF field strengths of v_1^{13} C = 14 kHz, v_1^{1} H = 46 kHz are employed at 60 kHz MAS, as illustrated in Fig. 6a.

Hartmann–Hahn transfers between ^{15}N and ^{13}C are an important component of N–C, N–C–C, and C–N–C correlation experiments, sometimes referred to as double cross-polarization (DCP) experiments. Band-specific transfers are employed similarly to the SPECIFIC-CP technique introduced by Baldus and coworkers [91,92]. As any ZQ condition would require strong irradiation on at least one of the low-frequency channels, the DQ n=1 condition

¹H-X recoupling



15N-13C recoupling

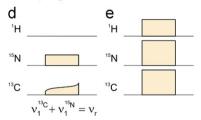


Fig. 6. Building blocks for heteronuclear recoupling at ultra-fast MAS. Pulse sequence elements for ${}^{1}\text{H}-\text{X}$ and ${}^{15}\text{N}-{}^{13}\text{C}$ transfers. The initial state of the spin-system are (a)–(c) \hat{i}_{x}^{1} , and (d)–(e) \hat{S}_{x}^{1} ; the desired final states are (a)–(c) \hat{S}_{x}^{2} , and (d)–(e) \hat{S}_{x}^{2} . For all channels, the carrier position is set to the mid-range of frequencies to be recoupled. RF field strength recommendations are indicated for (a)–(d), see also Fig. 7b and c. For MOD-CP, a cosine amplitude modulation of frequency v_{mod} is applied on the X channel. Adiabatic amplitude sweeps are applied for ${}^{1}\text{H}-\text{X}$ band-specific CP and ${}^{15}\text{N}-{}^{13}\text{C}$ SPECIFIC-CP. (e) Pulse settings and durations for PAIN-CP are discussed in Ref. [90].

(Eq. (6)) is the condition of choice at ultra-fast MAS (Fig. 6d).

$$v_1^{13}C + v_1^{15}N = v_r \tag{6}$$

An adiabatic amplitude sweep is commonly employed on ^{13}C to increase the long-term stability of the experiment [24]. The RF fields on both channels must be chosen so as to prevent homonuclear recoupling conditions such as HORROR $(v_{RF} = \frac{1}{2}v_r)$, or rotary resonance $(v_{RF} = v_r)$. As ^{13}C has a large chemical shift range, RF frequencies between $\frac{1}{3}v_r$ and $\frac{2}{3}v_r$ should be avoided, especially at high fields or with a large adiabatic sweep. Alternatively, the adiabatic amplitude sweep can be done on the ^{15}N channel. Efficient transfers have been accomplished without proton decoupling at 40 kHz MAS and above [41,60]. Strong ^{1}H decoupling $(v_1^{1\text{H}} \approx 2.5v_r)$ is required in the fast MAS regime (20 kHz) [90,93], although efficient transfers might be possible in some conditions in the absence of ^{1}H decoupling [94].

4.3. Second-order recoupling

The previously mentioned first-order recoupling sequences are necessarily susceptible to the phenomenon of dipolar truncation [40,95]: in a multi-spin system, magnetization is preferentially transferred to a strongly coupled partner, extinguishing transfer to a weakly coupled partner. Sequences operating through a second-order mechanism, less prone to dipolar truncation, are appropriate for transfer to remote nuclei and collection of longrange distance restraints. Second-order recoupling sequences that work well in the ultra-fast MAS regime include PAR [45,80,96], MIRROR-SD [31], RESORT [32], and PARIS [33,34,97] in the homonuclear case; PAIN-CP [90,93], SOCP [98], MOD-CP [44], and MIRROR-CP [99] in the heteronuclear case.

The general designation of third spin assisted recoupling (TSAR) describes a mechanism B-[A]-C where two nuclei B and C are recoupled through their respective dipolar coupling with a third spin A (i.e. B-A and C-A couplings) rather than directly [93,96,100]. The first use of the TSAR mechanism for magnetization transfer has been introduced in a work by Lewandowski and coworkers with Proton assisted insensitive nuclei cross-polarization (PAIN-CP) where second-order N-C transfer was demonstrated at 20 kHz MAS [93]. The ¹⁵N-[¹H]-¹³C TSAR mechanism is obtained by applying cw irradiation simultaneously on three channels (Fig. 6e): ¹H, ¹³C, and ¹⁵N. A thorough theoretical description of PAIN was recently published [90]. The homonuclear analog of PAIN, Proton assisted recoupling (PAR), generates X-[¹H]-X transfer by concurrent cw irradiation on the proton and X channel (Fig. 5c), For $^{15}\text{N}-^{15}\text{N}$ PAR at 20 kHz MAS $(v_1^{^1}=v_1^{^{15}\text{N}}>2v_r)$ and $(v_1^{^1}\approx 0.2v_r,\ v_1^{^1}\approx 3v_r)$ are suggested. Inter-nuclear proximities can be probed from the build-up of transferred intensity during the PAR element, as demonstrated with ¹³C-¹³C cross-peak build-ups in microcrystalline Crh [96]. The use of PAR at ultra-fast MAS (65 kHz MAS) was demonstrated on microcrystalline GB1 [45] where ¹³C-¹³C correlations could be obtained with a low-power condition of $(v_1^{^{13}C} \approx 1.12v_r, v_1^{^{1}H} \approx 0.3v_r)$. A condition at $(v_1^{^{13}C} \approx 1.75v_r, v_1^{^{1}H} \approx 1.3v_r)$ has also been used. Possible optimization protocols for PAR are discussed in Refs. [45,80]. When setting up a PAR or a PAIN experiment, an important aspect is to avoid RF frequencies which lead to firstorder recoupling conditions (e.g. HORROR, rotary resonance, and Hartmann-Hahn matching). First, the regions to be excluded from the optimization space are identified by simulating an interference map of the spin system. RF power levels are then optimized around regions of known high transfer, identified from a polarization transfer map. Scripts for generating such maps, including C_{α} – C_{β} , C_{α} –C', and C_{α} – C_{γ} transfers, are available [45].

Analogous to PAR, the RESORT experiment has been presented for homonuclear transfer [32]. The low- γ nuclei are spin-locked with *cw* irradiation; however in this case, phase-alternated irradiation is applied on 1H (Fig. 5f). This generates an effective Hamiltonian with the same form as for PAR but with different transfer conditions. RESORT has been demonstrated at 40 kHz MAS on microcrystalline ubiquitin.

In contrast to previous sequences, second-order recoupling can also be accomplished with sequences where no irradiation is applied on the X channel. For example, proton spin diffusion $(^{1}H-^{1}H)$ and proton-driven spin diffusion $(^{13}C-^{13}C$ or $^{15}N-^{15}N)$ do not require any irradiation during mixing. However, as PDSD relies on energy-level broadening due to homonuclear ¹H-¹H and heteronuclear ¹H-X dipolar interactions to increase the spectral overlap between low- γ spin pairs, it only functions at low MAS. In the fast MAS range, the dipolar-assisted rotational resonance (DARR) method is used to accelerate the transfer [29,101]. According to Takegoshi and coworkers [101], the effect of irradiating the protons at the rotary-resonance condition (Eq. (7)) is to restore the spectral overlap between a sideband of an X spin and the ¹H–X dipolar pattern of another X spin. Spin-diffusion mediated by DARR is abolished at ultra-fast MAS. Indeed, in a study by Scholz and coworkers, it was found that irradiating at the n=1 or n=2 DARR condition could not induce C' to C_{α} transfer at 45 kHz MAS [31]. Instead, proton irradiation at the mixed rotational and rotary-resonance (MIRROR) condition was found to mediate spin-diffusion (Fig. 5d). The strongest transfer occurs when the applied field on ¹H matches the ¹³C chemical shift difference (Eq. (8), n=0). The recoupling can be carried out using phase-alternated irradiation instead of cw, providing increased robustness against $v_1^{\rm H}$ mismatch. Because of the dependence on ¹³C chemical shift, MIRROR is an intrinsically band-selective sequence. This building block is particular in that it has a dual function and can promote both $^{13}C^{-13}C$ recoupling (MIRROR-SD) and ¹H-¹³C transfer (MIRROR-CP) [99]. The PARIS scheme has also been proposed to promote ¹³C-¹³C recoupling [33] and it employs phase-alternated ${}^{1}H$ pulses which have a duration of either $\frac{1}{2}$ or 2 times the rotor period (Fig. 5e). PARIS-xy, a variant of PARIS with the x, -x, -y, y phase cycle [34], was recently used in the 39–52 kHz MAS range to record ¹³C–¹³C correlations in selectively labeled amyloid-β peptides [97].

$$v_1^{^{1}H} = nv_r, \quad n = \{1, 2\}$$
 (7)

$$v_1^{^{1}H} = nv_r \pm \Delta \Omega_{iso}^{^{13}C}, \quad n = \{0, \pm 1, \pm 2, \pm 3, \pm 4\}$$
 (8)

Second-order processes can also accomplish the initial crosspolarization from ¹H to low-γ nuclei. Indeed, second-order cross-polarization (SOCP) was introduced in a study by Lange et al. [98]. An important observation is that decay during ¹H spinlock is highly dependent on the applied RF strength. Efficient spin-locking of ¹H magnetization does not require strong RF fields, but can be accomplished with low-power irradiation, e.g. 9.4 kHz at 65 kHz MAS. The most efficient ¹H RF frequencies only appear at certain ratios of the MAS frequency, usually with vi equal to 0.15, 1.67, and 2.55 times v_r (Fig. 7c). Those ratios are identified by monitoring the signal intensity as a function of ¹H spin-lock field. SOCP is achieved with the n=0 Hartman-Hahn condition: $v_1^{'H} = v_1^X$ (Fig. 6b). The predominant transfer mechanism is due to second-order cross-terms between homonuclear and heteronuclear dipolar couplings, although transfer can in principle also occur through J-couplings. Notable advantages of SOCP are the higher sensitivity compared to first-order highpower CP, as initial magnetization can originate from protons not directly coupled to the S spin, and the robustness against RF field inhomogeneities, due to matching at the n=0 condition. SOCP is

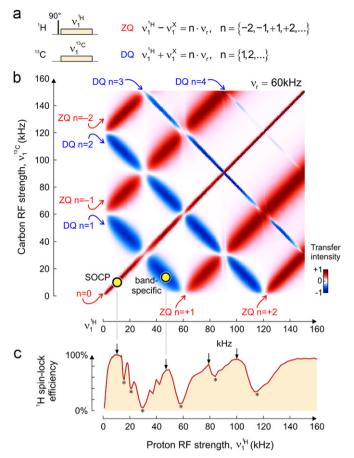


Fig. 7. Selecting optimal $^1\text{H}-\text{X}$ cross-polarization conditions at ultra-fast MAS. (a) Pulse scheme for $^1\text{H}-^{13}\text{C}$ CP. The ZQ (red) and DQ (blue) Hartmann-Hahn conditions lead, respectively, to positive and negative transfers. (b) Map of magnetization transfer at 60 kHz MAS simulated for a CH₂ group, considering the $^1\text{H}-^1\text{H}$ and $^1\text{H}-^{13}\text{C}$ dipolar couplings. In addition to $n=\pm 1, \pm 2$ ZQ and n=1,2 DQ conditions, transfer conditions due to higher order terms appear: the n=0 condition used in SOCP, as well as the n=3 and n=4 DQ conditions. The RF settings for SOCP and band-specific CP are indicated. Intensities correspond to the average ^{13}C magnetization for time points between 6 and 8 ms. (c) Efficiency of ^1H spin-lock as a function of RF strength, as measured after 2.5 ms of spin-lock in $[^1\text{SN},^{13}\text{C}]$ -labeled glutamine. Conditions of least decay, indicated by arrows, should be employed for CP. Deleterious recoupling conditions at ratio of $\frac{1}{4}$, $\frac{1}{3}$, $\frac{1}{2}$, $\frac{3}{2}$, and 2 times v_r are indicated by asterisks. Part (c) is adapted from Ref. [44].

employed in a band-selective fashion in the construction of a complete set of experiments for protein assignment at ultra-fast MAS (Fig. 2). For the initial H-X transfer, a limited bandwidth is adequate for N-C experiments; however, broadband excitation may be desired in the case of C-C spectra. The RF strength on both channels can then be slightly increased to 19 kHz at 60 kHz MAS, therefore avoiding the higher order $v_1 = \frac{1}{4}v_r$ and $v_1 = \frac{1}{3}v_r$ recoupling conditions. When two regions present a large chemical shift separation $|\Delta\Omega_{iso}^X|$, for instance C' and C_{α} in proteins, the simultaneous excitation of both regions is achievable at low-power through the amplitude-modulated second-order cross-polarization scheme (MOD-CP) [44]. In the MOD-CP scheme, a cosine amplitude modulation with frequency $v_{mod} = \frac{1}{2} |\Delta \Omega_{iso}^X|$ is applied on the X channel (Fig. 6c). The ¹³C RF strength is doubled, $v_1^X = 2v_1^{1H}$, creating the two distinct excitation regions. The simultaneous excitation of C' and C_{α} , or C' and C_{β} regions is demonstrated in microcrystalline ubiquitin at 60 kHz. MOD-CP restores the high information content per spectrum as found in highpower broadband excitation, while limiting the RF expenditure of the pulse sequence. MOD-CP can potentially be exploited for the

study of organic materials, since several other spin-1/2 nuclei such as $^{31}\rm{P},~^{15}\rm{N},~^{29}\rm{Si},~$ and $^{19}\rm{F}$ cover a large range of isotropic chemical shifts.

5. Proton NMR at ultra-fast MAS

So far in ssNMR, mostly rare low- γ nuclei such as 13 C and 15 N have been employed for detection, for observation in indirect dimensions, and for obtaining distance restraints. On the other hand, protons have the advantage of a high gyro-magnetic ratio γ , an almost 100% natural abundance, and a molar content of more than 50% in biomolecules and in numerous other systems of interest. As sensitivity at detection is proportional to $\gamma^{3/2}$, at the same line-width, detection via 1 H would be, respectively, 8 and 31 times more sensitive than detection via 13 C and 15 N. In general, sensitivity gains via proton detection in 2D HETCOR experiments depend as well on line-width and quality factor of the probe coils, as it has been described by Ishii and Tycko [102].

Inter-nuclear proximities in solid-state NMR are estimated from dipole–dipole couplings, which are proportional to the product of gyro-magnetic ratios γ_1 and γ_2 of both nuclei involved in the interaction. As a consequence of ultra-fast MAS, $^{13}\text{C}-^{13}\text{C}$ and $^{15}\text{N}-^{15}\text{N}$ spin diffusion is significantly reduced [53,103,104] and cannot be efficiently used for the collection of distance restraints. For the same inter-nuclear distance, $^{1}\text{H}-^{1}\text{H}$ dipolar couplings are 16 and 97 times stronger than $^{13}\text{C}-^{13}\text{C}$ and $^{15}\text{N}-^{15}\text{N}$ couplings, respectively. This would potentially allow to obtain long-range distance restraints based on $^{1}\text{H}-^{1}\text{H}$ magnetization transfer [46,47,105]. Another benefit of proton observation is resolution enhancement by the introduction of additional proton dimension(s), which can simplify the process of resonance assignment.

However, the major limitation for ^1H observation in ssNMR relates directly to its high natural abundance, high density, and high gyro-magnetic ratio: networks of strong proton–proton dipolar couplings result in very broad lines of ca. 50 kHz for static samples. In the high-field approximation, the spin Hamiltonian of the homonuclear dipolar coupling of spins S and K is given by Eq. (9). The instantaneous dipolar coupling $d(\theta(t))$ depends on the orientation of the inter–nuclear vector with respect to the external magnetic field, given by the Euler angle θ (Eq. (10)). The angle θ is time-modulated due to MAS. The maximum value of $d(\theta(t))$ for two protons at a distance of 2.5 Å is 7.8 kHz.

$$H_D = d(\theta(t))[2S_z K_z - S_x K_x - S_y K_y]$$
(9)

 $S_{x,y,z}$ are the Cartesian components of spin operators S and K.

$$d(\theta(t)) = \frac{\mu_0}{4\pi} \frac{\gamma^2 h}{r^3} \frac{1 - 3\cos^2(\theta(t))}{2}$$
 (10)

r is the inter-nuclear distance, γ is the gyro-magnetic ratio, \hbar is the reduced Planck constant.

There are now three major approaches to increase proton resolution [6]: (1) proton dilution with deuterium [106], (2) proton–proton radio-frequency homonuclear decoupling [107], and (3) magic-angle spinning at rates (> 30 kHz MAS) that are comparable with homonuclear dipolar interactions [108]. Additionally, a combination of these techniques is commonly exploited. The first approach provides the best ¹H line narrowing so far, with line-widths on the order of 20 Hz in microcrystalline SH3 [109] at MAS rates below 20 kHz. However, the use deuteration can require adopting or establishing new sample preparation protocols. Radio-frequency ¹H–¹H decoupling is efficient in combination with MAS, which removes other anisotropic interactions such as CSA and heteronuclear dipolar couplings as well. Numerous experimental techniques were developed in this field since the

early days of NMR and resulted in achievable line-widths on the order of 150-200 Hz in dry model compounds [110]. A major advantage of this concept is the possibility of routine experimental implementation without the need for special ssNMR instrumentation (i.e. ultra-fast MAS probe-head) and on samples with natural hydrogen abundance. In the third approach of using only mechanical spinning of the sample for suppression of strong homonuclear couplings, it was possible to observe resolved proton resonances in model compounds at the relatively slow spinning rate of 15 kHz [111]. Recently, ultra-fast commercial MAS probe-heads have become available with reachable spinning rates up to 80 kHz (IEOL, Japan) [5]. Certainly, these rates are not sufficient to suppress proton-proton dipolar interactions to an insignificant level, but they still provide remarkably good averaging of protonproton homonuclear dipolar couplings and yield high-resolution proton spectra (Fig. 8). Remarkably, the spinning frequencies in ultra-fast MAS are getting very close to effective RF field frequencies which are commonly applied for homonuclear proton decoupling in experiments on proteins [112,113]. Therefore, one may expect improved decoupling efficiencies of the MAS-alone approach in the near future. Advantages of using ultra-fast MAS alone are simplicity of pulse technique implementation, artifactfree spectra, absence of an isotropic chemical shift scaling factor, and higher sensitivity because of the absence of RF irradiation during acquisition. As well, prevention of sample heating due to RF irradiation is a necessity for short repetition delay experiments and is one of the major advantages of the ultra-fast MAS approach [60] (see also Sections 1 and 3 on Paramagnetic doping and paramagnetic proteins).

For three and more like spins which are interacting through dipolar couplings and are arbitrarily positioned in space, the spin Hamiltonian derived with the help of the Eq. (9) is, in general, not self-commuting at different time points $[H(t'), H(t'')] \neq 0$ and has therefore a "homogeneous" nature according to the classification introduced by Maricq and Waugh [114]. As a consequence, proton line-widths are broad even at spinning frequencies exceeding the sum over all proton-proton interactions, and decrease slowly at increasing spinning rates. Maricq and Waugh derived in their pioneering work that the line-width of homonuclear spin systems is inversely proportional to the spinning rate v_r . Later, the proton line-shape dependence on sample spinning has been theoretically discussed by a number of other authors [73,108,115,116]. Levitt et al. [73] found that at some geometrical configurations of proton systems the line-widths can be proportional to $(\Delta \delta v_r^2)^{-1}$, where $\Delta\delta$ is the chemical shift difference of two spins. Using Floquet theory together with standard perturbation theory and numerical

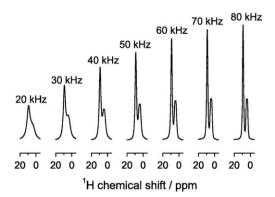


Fig. 8. Single pulse ¹H spectra of glycine at various spinning frequencies. This figure illustrates the impact of ultra-fast spinning on proton resolution and sensitivity. The undecoupled, proton-detected glycine spectra were obtained in the range of spinning rates between 20 kHz and 80 kHz in studies of Nishiyama and coworkers [5].

simulations, Ray and coworkers [115] found different power dependences of the proton line-width on v_r , falling in a range between -1.1 and -2 depending on local spin density and mutual proton orientation. Still, the comprehensive theoretical and even numerical treatment of homonuclear multi-spin systems under MAS remains a formidable challenge.

Previously, extensive experimental studies of proton linewidth in different systems had been performed by a number of research groups in the fast spinning regime (20-40 kHz), limited by the technologically allowed spinning rate at that time [102.116.119–121]. They could confirm the prediction that the line-width is approximately inversely proportional to the spinning rate and strongly dependent on the local proton density, as well as on other sources of broadening like anisotropic magnetic susceptibility [119,121] and crystallite orientation [118,122]. These studies demonstrate the possibility of sufficient ¹H resolution in ssNMR, and are one of the driving forces to increase spinning rates up to 80 kHz. Nowadays, many proton-detected experiments are performed in the fast and ultra-fast MAS regime: the rapid characterization of small compounds [123,124], linewidth measurements at different external magnetic fields and MAS rates, and studies on the influence of anisotropic magnetic susceptibility [119] and crystallite orientation [118] on line broadening.

A number of pulse schemes for proton-detected heteronuclear 2D correlation experiments have been designed and can be easily extended to 3D or 4D [46,47,105]. Magnetization transfer in HSQC experiments can be performed through CP (Fig. 9a) [125], INEPT (Fig. 9b) [117], or refocused INEPT (Fig. 9c) [46,118]. An HMOCtype experiment [5,117] is shown in Fig. 9d. Zhou and coworkers demonstrated the efficiency of indirect proton detection in 2D HETCOR experiments recorded on fully protonated, uniformly [13C, 15N]-labeled GB1 at an MAS rate of 40 kHz [126], On a spectrometer operating at 500 MHz proton Larmor frequency (11.75 T), ¹H detection was more sensitive by a factor of 3 and 4 compared with ¹³C and ¹⁵N detection, respectively. With an instrument operating at 750 MHz ¹H Larmor frequency (17.62 T), the sensitivity of proton-detected experiments was 14 times higher compared with 15N-detected heteronuclear correlation experiments, which can be attributed to narrower proton linewidths and improved probe design. Indeed, the average proton line-width of 500 + 150 Hz observed on the 500 MHz spectrometer decreased down to 360 ± 115 Hz in experiments performed on the 750 MHz spectrometer. The last effect has been attributed to truncation of homonuclear coupling networks due to the larger isotropic chemical shift dispersion. So far, significant truncation effects have not been observed by other studies on numerous model compounds [119,121] but instead were found for systems which have atypically large chemical shift dispersion and/or very weak dipolar couplings [127], such as systems containing unpaired electrons [63,128,129]. Proton line-widths were further reduced by using deuteration. Combined with the advantages of using proton detection and collecting proton-proton distance restraints, this allowed the determination of the 3D structure of microcrystalline GB1 [105]. The protein was uniformly ¹³C and ¹⁵N labeled, and protonated only on labile sites, resulting in proton line-widths of 140 ± 30 Hz on average. Spectra were measured at 39 kHz MAS on a spectrometer operating at 750 MHz proton

As well, proton line-widths can be modulated by internal motions via two concurrent effects: (1) line narrowing, due to the partial averaging of anisotropic interactions (e.g. dipolar couplings and CSA) [130], and (2) line broadening, due to chemical exchange and transverse relaxation [131,132]. In both cases, the magnitude of the effect depends on the amplitude of motion and will occur only if the motion is not completely

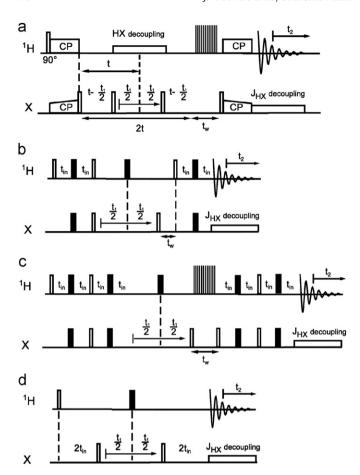


Fig. 9. Indirect proton-detected 2D HETCOR pulse schemes. HSQC experiment based on (a) CP transfer, (b) INEPT, and (c) refocused INEPT. (d) Pulse scheme to obtain HMQC correlations. Conventional notations are used. X refers to 13 C or 15 N nuclei. Evolution periods are represented by t_1 and t_2 . Open and black bars denote 90° and 180° pulses, respectively. Duration for water signal suppression is given by t_w , where blocks of lines represent saturation pulses. INEPT transfer times t_{in} in relaxation-free systems are defined by the HX J-coupling as $t_{in} = (4J_{HX})^{-1}$. Pulse schemes adapted from Refs. [117,118].

restricted (i.e. with an order parameter S < 1). To observe line narrowing, the frequency of motion τ_c^{-1} must be larger than the anisotropy of the NMR interaction. On the other hand, motions which have the same or slower frequency as the ¹H Larmor frequency will lead to relaxation line broadening. In the case of chemical exchange, motions which have a frequency on the same order of magnitude as the difference in proton chemical shift between sites will lead to exchange broadening. Thus, very fast motions $(\tau_c^{-1} \gg v_0^{1_H})$ can facilitate the suppression of ${}^{1}H - {}^{1}H$ homonuclear couplings by ultra-fast MAS, as they induce line narrowing but not line broadening. Studies on the local dynamics of SH3 domain and ubiquitin [133,134], as well as a number of highly resolved proton spectra for those proteins [46,109], indicated that most residues do not present slow motions of large amplitude, which would lead to significant proton line broadening.

The high sensitivity and resolution approach of using MAS alone was used in the study of model compounds. Zhou and coworkers introduced protocols for rapid analysis of small unlabeled organic compounds [123]. Salager and coworkers [135] reported 1H line-widths between 205 Hz and 560 Hz for the model compound $\beta\text{-L-Asp-L-Ala}$ studied at spinning rates of 65 kHz on a 500 MHz spectrometer. Studies from Holland et al. [118] corroborate the utility of proton detection for the

characterization of small molecules like L-alanine, L-isoleucine, and black widow dragline silk.

Additionally, indirect 1 H detection offers good opportunities for NMR of 14 N, which has high natural abundance and importance in biomolecules and a number of materials, while very poorly suitable for direct detection due to its spin number I=1 and related to its quadrupolar interaction. Spinning frequencies up to 80 kHz were used in experiments of Nishiyama et al., resulting in proton-detected 14 N- 1 H 2D spectra of glycine and glycyl-L-alanine with high sensitivity and resolution [5].

Recent progress has been made in studies of ¹H homonuclear decoupling through RF irradiation at ultra-fast MAS. It has been shown that a number of proton-proton decoupling pulse sequences can be successfully applied at MAS rates of 60-65 kHz [136-140] by using RF strengths on the order of 200 kHz. The widely used family of PMLG [141,142] and DUMBO [143] homonuclear decoupling sequences have shown a good performance for use in both indirect and direct dimensions [135,136]. That is despite the fact, that they have been developed in the quasi-static approximation, where rotor spinning is much slower than the frequency of the applied RF decoupling cycle. Two recently introduced rotor-synchronized sequences, smooth amplitude modulation (SAM) [137] and the RN_n^{ν} family [139], can yield good proton resolution as well. The TIMES sequence [140] shows good performance at a relatively low RF field of 130 kHz. The RF irradiation alone is not designed to average out the proton-proton couplings completely for static samples in first-order approximation, but to reduce them significantly, while any remaining homonuclear interactions are further suppressed by ultra-fast MAS.

6. Protein dynamics

Compared to NMR in solution, solid-state NMR has the advantage that molecular internal motion can be studied in the absence of overall molecular tumbling. Indeed, the dynamics of proteins in the solid state can directly manifest itself unhindered by global molecular reorientation. Recent achievements in solidstate NMR lay the ground for studying protein dynamics with single residue resolution. The internal backbone motions of small proteins have been subsequently quantified within the frame of an extended model-free approach [133,134]. In general, there are two types of experiments which provide information on motion in solid-state NMR. The first approach observes spin evolution caused by relaxation and extracts the amplitude and frequency of incoherent motions. A very common example of this approach is the measurement of longitudinal relaxation [144]. In a second type of experiment, partially motionally averaged anisotropic interactions are measured and compared to their static limits which are known *a priori*. This approach can give only information about the amplitude of incoherent motion. One prominent example is the measurement of ${}^{1}H^{-15}N$ or ${}^{1}H^{-13}C$ dipolar couplings and consequently dipolar order parameters [112,134,145]. Usually, the influence of incoherent motion on the spin evolution is difficult to quantify due to the presence of strong anisotropic interactions. Ultra-fast spinning can increase the accuracy of relaxation experiments and even give access to information previously not available, owing to the near-complete averaging of anisotropic interactions. At the same time, incoherent motions on timescales much faster than the MAS stay unaffected by sample rotation. So far, only few studies of protein dynamics were performed in the ultra-fast MAS regime. Employing ultrafast MAS of 60 kHz, Lewandowski and coworkers [53] could almost truncate ¹³C-¹³C PDSD during T₁ measurements, performed on protonated, uniformly [13C, 15N]-labeled GB1. The experiments provide high-accuracy information about backbone and side chain dynamics and its variation. Schanda et al. [134] obtained a vast set of relaxation parameters including 15 N T_1 , 1 H $^{-15}$ N order parameters, 1 H $^{-15}$ N dipole and 15 N CSA cross-correlated relaxation rates for the backbone of highly deuterated ubiquitin at a fast MAS rate of 40–45 kHz.

7. Conclusions

A new generation of fast spinning MAS probe-heads has opened ways to improved resolution and sensitivity of biological solid-state NMR. Additionally, fundamentally new applications such as the study of paramagnetic metalloproteins have become feasible in the ultra-fast spinning regime. In this review, we have discussed those recent developments: the study of paramagnetic metal binding centers, new methods for resonance assignment and detection of long-range distance restraints, proton NMR at ultra-fast MAS, and the study of protein dynamics. We expect that ongoing research from an increasing number of laboratories in this still largely unexplored area of solid-state NMR will lead to even more powerful methods and exciting new applications.

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- ⁵ Demonstrates ¹³C–¹³C magnetization transfer in peptides and proteins through the Proton Assisted Recoupling (PAR) mechanism at 65 kHz MAS.
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 $^{^{19}}$ Systematic study of proton line-width at 40 kHz MAS in a fully protonated protein and a model compound at different external magnetic fields.