

**Persistence and potential causes of reduced net CH₄ consumption under elevated CO₂
in a temperate forest**

Lindsay Laura Dubbs

A dissertation submitted to the faculty of the University of North Carolina at Chapel Hill
in partial fulfillment of the requirements for the degree of Doctor of Philosophy in the
Department of Environmental Sciences and Engineering.

Chapel Hill
2009

Approved by:
Stephen Whalen (Advisor)
Emily Bernhardt
Rob Jackson
Frederic Pfaender
Howard Weinberg

© 2009
Lindsay Laura Dubbs
ALL RIGHTS RESERVED

ABSTRACT

Lindsay Laura Dubbs: Persistence and potential causes of reduced net CH₄ consumption under elevated CO₂ in a temperate forest
(Under the direction of Stephen C. Whalen)

Impacts of the projected increase in atmospheric CO₂ on other biogeochemical cycles are uncertain. In a two-year study, Phillips et al. (2001) reported a 16 to 30% decrease in net consumption of atmospheric CH₄ by soils in CO₂-enriched plots in a temperate loblolly pine (*Pinus taeda*) forest. Consumption by upland soils accounts for ~30 Tg CH₄ y⁻¹ and is the only terrestrial sink for atmospheric CH₄, which is a greenhouse gas with radiative forcing second only to CO₂. However, it is uncertain whether decreased atmospheric CH₄ consumption represents a transient or sustained response of forest-soil systems to elevated CO₂.

This research focused on field observations aimed at investigating the strength and persistence of reduced atmospheric CH₄ consumption by temperate forest soils under elevated CO₂ at the same study site. It further investigates the causes of this response by CH₄ oxidizing and producing communities through field and laboratory experiments.

Rates of soil-atmosphere CH₄ exchange were repeatedly measured over 3 y from permanently established sampling sites at the Free Air Carbon Dioxide (FACE) site in the Duke Forest, where CO₂-enriched plots of a loblolly pine forest are maintained at approximately 200 mL L⁻¹ above ambient concentrations (380 mL L⁻¹), while control plots are exposed to ambient atmospheres. Reduced net atmospheric CH₄ consumption persisted in CO₂-enriched plots, showing annual declines of 19, 10 and 8% relative to control plots.

This study and previous work give a nearly continuous 8 y record of reduced net atmospheric CH₄ consumption in CO₂-enriched plots that suggests this is likely a sustained negative feedback to increasing atmospheric CO₂.

Causitive factors for the observed decrease in net CH₄ consumption under elevated CO₂ were difficult to identify because of high spatial and temporal variability in microbial activity and limited ability to collect soil samples. However, higher soil moisture and increased incidence and rates of CH₄ production in CO₂-enriched plots, along with transient inhibition by plant exudates and low overall soil diffusivity, begin to explain reduced rates of CH₄ consumption and increased rates of CH₄ production that result in long-term reduction in net CH₄ consumption in these soils.

ACKNOWLEDGEMENTS

Steve Whalen has been my mentor for seven years now. I am very grateful for all that I have learned from him including, but not limited to, how to be an honest and dedicated scientist, caring teacher, and quiet leader; how to build and fix instruments and field equipment myself; how to write more clearly and scientifically; and perhaps, most importantly, that I should lighten up a bit.

My dissertation committee has been incredibly helpful to the completion of my research and studies with their time and attention, pointed questioning, guidance, and kind words. Thank you to Drs. Emily Bernhardt, Rob Jackson, Frederic Pfaender, and Howard Weinberg.

Several other professors have also been great mentors to me during my graduate school years, especially Drs. Robert Wetzell, Donald Lauria, Joe von Fischer, and Ram Oren. I cannot thank them enough for their time, candid and interesting conversations, and inspiration.

Lance Leonhardt, an amazing biology and environmental science teacher, first lit the fire in me and gave me every reason to keep following my curiosity. I thank him for his excitement that was truly contagious.

I am grateful for soil moisture data provided by Hyun Seok-Kim and Ram Oren and root exudate data provided by Emily Bernhardt and Rich Phillips. I also thank many for field assistance over the years including Eric Fischer, Ryan Elting, Lauren Elich, Omar Monzon, Steve Artabane, William Dodge, and Robert Nettles. With their help and company, Robert

Nettles, David Cooley, and Jeff Phippen made visits to the FACE site something to look forward to. I thank Ramon Garcia who was very generous with statistics guidance. Whalen lab group members (Eric Fischer, Jeff DeBerardinis, Priscilla Benson, Brian Chalfant, Marsha Fisher, Gina Panasik, Joelene Diehl, Ken Fortino, and Dendy Lofton) have been great colleagues and I thank them for their company and conversation. David Singleton was an incredibly patient and knowledgeable teacher of molecular biology, even though our method never did work. I also am very thankful for the dedicated staff of ESE that I have interacted with on a regular basis. Jack Whaley, Donna Simmons, Deborah Williams, Robin Whitley, Jim Wallace, Melody Levy, Ann Goodwin, Elise Pohl, Rebecca Riggsbee Lloyd, and Linda Brezin have made navigating graduate school much more enjoyable and easy. I thank Glenn Walters, Randy Goodman, Cliff Burgess, and Fred Bevin of the ESE design studio, who were there to answer any question about instruments, find any part, or build anything I needed.

My family and friends have been resilient and determined in their encouragement, interest, and support of my research and studies. I would especially like to thank my parents, Barry and Dawn Dubbs, my brother, Nate, and William Dodge for their patience and constant flow of encouragement over the past few months.

This study was supported by EPA STAR Grant No. RD-83145101-0 to S.C. Whalen and R.G. Wetzel, and the Office of Science (BER), U.S. Department of Energy, Grant No. DE-FG02-95ER62083. The Edward R. Kuenzler award also provided financial assistance to complete my degree and for that, I am thankful.

TABLE OF CONTENTS

TABLE OF CONTENTS.....	vii
LIST OF TABLES	xi
LIST OF FIGURES	xii
LIST OF ABBREVIATIONS AND SYMBOLS	xiv
CHAPTER 1: INTRODUCTION.....	1
Elevated CO ₂ And Temperate Forests	1
Methane In The Pedosphere.....	3
Observed Effects Of Elevated CO ₂ On CH ₄ Dynamics In Soils	6
Ecosystem-Level Changes That May Influence CH ₄ Dynamics In Temperate Forests	7
Research Objectives.....	9
Dissertation Structure.....	10
References.....	12
CHAPTER 2: REDUCED NET CH ₄ CONSUMPTION IS A SUSTAINED RESPONSE TO ELEVATED CO ₂ IN A TEMPERATE FOREST	20
Abstract.....	20
Introduction.....	21
Methods.....	23

<i>Field Site</i>	23
<i>Gas Flux Measurements</i>	24
<i>Soil Physicochemical Measurements</i>	25
<i>Calculations And Statistics</i>	26
Results	27
<i>Environmental Variables</i>	27
<i>Patterns In Net CH₄ Flux</i>	27
Discussion	29
<i>Overall Patterns Of Net CH₄ Consumption And Environmental Correlates</i>	29
<i>Differences In Net CH₄ Consumption Between CO₂ Treatments</i>	31
<i>Potential Reasons For Reduced Net CH₄ Consumption Under Elevated CO₂</i>	33
References	37
CHAPTER 3: INHIBITION OF CH ₄ CONSUMPTION BY SECONDARY CARBON COMPOUNDS IN THE TISSUES AND EXUDATES OF TEMPERATE FOREST PLANTS EXPOSED TO ELEVATED CO ₂	48
Abstract	48
Introduction	49
Methods	51
<i>Field Site</i>	51
<i>Plant Exudate Collection</i>	52
<i>Soil Assays And Incubation</i>	54
<i>Statistical Analysis</i>	55
Results	55

<i>Throughfall And Leaf Leachates</i>	55
<i>Duff Leachates</i>	56
Organic Acids From Root Exudates	56
Discussion.....	57
References.....	62
CHAPTER 4: REDUCED NET CH ₄ CONSUMPTION CAUSED BY CHANGES IN THE SOURCES AND TRANSPORT OF SOIL GASES IN A TEMPERATE FOREST EXPOSED TO ELEVATED CO ₂	74
Abstract.....	74
Introduction.....	75
Methods.....	77
<i>Field Site</i>	77
<i>Soil Gas Sampling</i>	78
<i>Soil Cores</i>	79
<i>Methane Sample Measurements</i>	80
<i>Diffusivity</i>	80
<i>Environmental Measurements</i>	81
<i>Calculations And Statistical Analysis</i>	81
Results.....	83
<i>Depth Profiles Of CH₄ Concentrations</i>	83
<i>Depth Profiles Of Net CH₄ Consumption And CH₄ Production</i>	83
<i>Effective Diffusivity</i>	84
Discussion.....	85
References.....	90

CHAPTER 5: CONCLUSION	100
References.....	103

LIST OF TABLES

Table

2.1.	Annual time-integrated rates of net CH ₄ consumption and volumetric soil moisture.....	43
4.1.	Effective diffusivity (P _{CH₄}) and corresponding calculated net CH ₄ flux (J _{CH₄} ; μg m ⁻² h ⁻¹), measured net CH ₄ flux (μg m ⁻² h ⁻¹), and soil moisture in control and CO ₂ enriched plots	95

LIST OF FIGURES

Figure	
2.1.	Time series for rates of net atmospheric CH ₄ consumption by forest soils under CO ₂ -enriched and ambient atmospheres and time series for changes in mean soil and mean volumetric soil moisture.....44
2.2.	Relationship between net atmospheric CH ₄ consumption and volumetric soil moisture.....45
2.3.	Annual time-integrated net CH ₄ consumption by temperate forest soils under ambient and elevated concentrations of CO ₂ for 2004 through 2006.....46
2.4.	Conceptual model of the impact of forest ecosystem responses to elevated CO ₂ that influence soil CH ₄ cycling dynamics.....47
3.1.	Conceptual model of the impact of forest ecosystem responses to elevated CO ₂ that influence soil CH ₄ cycling dynamics.....67
3.2.	Mean first order rate constants (k; d ⁻¹) for CH ₄ consumption in temperate forest soils amended with deionized water or throughfall from CO ₂ -enriched or control plots.....68
3.3.	Mean first order rate constants (k; d ⁻¹) for CH ₄ consumption by temperate forest soils amended with leaf leachate from the four most dominant trees within the control and CO ₂ -enriched plots.....69
3.4.	Rates of CH ₄ consumption by forest soils amended with deionized water or duff leachates from CO ₂ -enriched or control plots.....70
3.5.	Rates of CH ₄ consumption by forest soils amended with duff leachates or deionized water.....71
3.6.	Rates of CH ₄ consumption by forest soils amended with deionized water or representative organic acids (100 μmol L ⁻¹) determined to be primary root exudates from loblolly pine (<i>Pinus taeda</i>) trees grown under elevated CO ₂ ...72

3.7.	Rates of CH ₄ consumption by forest soils amended with deionized water or levulinic acid, an organic acid determined to be a primary root exudate from loblolly pine (<i>Pinus taeda</i>) trees grown under elevated CO ₂	73
4.1.	Conceptual model of the impact of forest ecosystem responses to elevated CO ₂ that influence soil CH ₄ cycling dynamics.....	96
4.2.	Composite depth profiles of CH ₄ in soils in forest plots exposed to elevated CO ₂ or the ambient atmosphere (control)	97
4.3.	Mean rates of net CH ₄ consumption in forest soils from plots exposed to elevated CO ₂ or the ambient atmosphere (control)	98
4.4.	Mean rates of CH ₄ production in forest soils from plots exposed to elevated CO ₂ or the ambient atmosphere (control)	99

LIST OF ABBREVIATIONS AND SYMBOLS

FACE	free air CO ₂ enrichment
k	rate constant
α	p-value acceptance threshold
D_{0,CH_4}	diffusion coefficients of CH ₄
$D_{0,Rn}$	diffusion coefficients of Rn
P_{Rn}	permeability of Rn
J_{CH_4}	flux of CH ₄
ΔC_{CH_4}	linear change in CH ₄ concentration
Δz_{CH_4}	linear change in depth

CHAPTER 1: INTRODUCTION

Elevated CO₂ and temperate forests

The present-day atmospheric CO₂ concentration of approximately 380 mL L⁻¹ (NOAA 2008), exceeds the highest concentration measured in ice core samples from before the Industrial Revolution by almost 100 mL L⁻¹ (Barnola et al. 2003). The atmospheric concentration of CO₂ is expected to continue to increase, mainly as a result of fossil fuel emissions and destruction of vegetation (Forster et al. 2007). Models project that atmospheric CO₂ concentrations, by the end of the present century, will exceed the pre-industrial concentration by up to 270% (Friedlingstein et al. 2006). A rising atmospheric CO₂ concentration is of concern because it is a long-lived greenhouse gas with a radiative forcing of 1.66 W m⁻², exceeding the radiative forcing of all other trace atmospheric gases that control climate (Forster et al. 2007). Increasing atmospheric CO₂ is also of significance because it is continuously exchanged between the atmosphere, the ocean, and the terrestrial biosphere through biogenic processes such as photosynthesis and respiration (Schlesinger 1997). Rates of photosynthesis and respiration are further controlled by temperature and water availability, and changes in the concentrations of CO₂ and other greenhouse gases are expected to elicit changes in air temperature and the hydrologic cycle that may vary regionally (Denman, K. L. et al. 2007). While the atmospheric concentration and radiative forcing of CO₂ are well understood (Forster et al. 2007), the impacts of the CO₂-induced changes in atmospheric composition and climate on whole ecosystems and their components

are less clear. An understanding of ecosystem responses at all levels to elevated CO₂ is important to predicting future climates as they can, in turn, feed back to the biogeochemical cycling of CO₂ and other greenhouse gases.

Attempts at understanding terrestrial biological and biogeochemical responses to elevated CO₂ have ranged in size and complexity from individual potted plants, to open-top chambers containing a community of plants, to large scale manipulations of intact ecosystems designed to embrace the entire suite of interactions and feedbacks among plants, microbial communities and elemental cycles. Each of these approaches has associated strengths and weaknesses. Physiological studies conducted in small and simple modeled ecosystems have been ineffective at capturing the complexity of ecosystem component interactions and feedbacks. On the other end of the spectrum, free-air CO₂ exchange (FACE) technology has been employed to conduct ecosystem-level studies where tall vegetation and their surrounding ecosystems are exposed to elevated CO₂ with minimal alterations of surrounding microenvironments (Hendrey et al. 1999b). The primary criticism of FACE experiments is that they are initiated by exposing an ecosystem to an abrupt increase in atmospheric CO₂, which may not fully represent how ecosystem components will react to the contemporary monotonic increase in the concentration of CO₂ in the Earth's atmosphere (Klironomos et al. 2005).

Nonetheless, FACE studies have proven to be useful in predicting ecosystem level changes in a range of terrestrial environments, among them, temperate forests. Overall, temperate forests exposed to elevated CO₂ using FACE technology show increases in tree growth and net primary production (DeLucia et al. 1999, Finzi et al. 2002, Hamilton et al. 2002, DeLucia, E.H. et al. 2005, Norby et al. 2005, Finzi et al. 2006a), increased delivery of

C to roots, the forest floor and soils with a small increase in soil C storage (Allen et al. 2000, Matamala and Schlesinger 2000, Schlesinger and Lichter 2001, Jastrow et al. 2005, Lichter et al. 2005, Lichter et al. 2008, Pritchard et al. 2008, Hoosbeek and Scarascia-Mugnozza 2009), increased soil respiration (King et al. 2004, Bernhardt et al. 2006, Taneva et al. 2006), and variable changes in N cycling (Billings and Ziegler 2005, DeLucia 2005, Finzi et al. 2006a) and soil community composition (Larson et al. 2002, Billings and Ziegler 2005, Billings and Ziegler 2008). The initial increase in net primary production in response to elevated CO₂ is predicted to slow with time as ecosystems become more N-limited (Finzi et al. 2006b), although N-limitation has yet to appear in temperate forests after 6 y of CO₂-enrichment using FACE technology (Finzi et al. 2006a).

However, observations (McMurtrie and Comins 1996) and ecosystem models (Newton et al. 2001) indicate that biological responses to elevated CO₂ and biogeochemical feedbacks vary widely on different timescales. For instance, down-regulation of photosynthesis has been commonly reported for CO₂-fertilized model and intact forest ecosystems after as little as two years (reviewed by Amthor 1995, Leakey et al. 2009). Over longer time trajectories, initial response functions of all ecosystem components from trees to microbes can be expected to adjust physiologically and demographically on different time scales through modification of biogeochemical feedbacks (Korner 2000). Thus short-and long-term responses to elevated atmospheric CO₂ must be distinguished.

Methane in the pedosphere

Methane is another greenhouse gas that is cycled through temperate forests and thus may be affected by CO₂-induced changes to the ecosystem. Methane is the simplest, most

reduced hydrocarbon, and a long-lived (9 to 15 y) greenhouse gas directly and indirectly contributing more than half of the radiative forcing of CO₂ (0.9 and 1.6 W m⁻², respectively; Schindell et al. 2005), through warming of the troposphere and its participation in the stratospheric chemistry of ozone and water vapor formation (Wuebbles and Hayhoe 2002). The global atmospheric CH₄ concentration has more than doubled since the Industrial Revolution to reach a present-day average concentration of ~1780 μL L⁻¹ (NOAA 2008). Methane is spatially and temporally variable in the troposphere, with a higher concentration in the Northern Hemisphere where emissions are higher, and a minima corresponding to increased photochemical destruction during summer months.

While destruction by the hydroxyl radical in the atmosphere is the largest sink for CH₄, the only known biological sink for CH₄, and the largest natural source of CH₄ are sited in the pedosphere. The balance between rates of CH₄ production (methanogenesis) and CH₄ consumption (methanotrophy) determines whether a soil is a net source or sink for atmospheric CH₄, and the strength of that source/sink. Methane production usually exceeds consumption in wetland environments, accounting for about 69% of emissions to the atmosphere from natural sources (Wuebbles and Hayhoe 2002). Conversely, upland soils account for approximately 38 Tg of CH₄ removal from the atmosphere annually (Ridgwell et al. 1999). This net biological sink in upland soils includes atmospheric CH₄ consumption by methanotrophic bacteria in the largely oxic soil profile, and consumption of endogenously produced CH₄ by methanogenic bacteria in anoxic microsites (reviewed by Conrad 1996).

Methanotrophic bacteria oxidize CH₄ for energy and as their sole source of carbon (C) for biosynthesis (Hanson and Hanson 1996). Methanotrophs are responsible for both 'high affinity oxidation' of CH₄, which occurs at CH₄ concentrations close to atmospheric

concentrations ($< 12 \text{ mL L}^{-1}$), such as in upland soils, and 'low affinity oxidation', which occurs at CH_4 concentrations $> 40 \text{ mL L}^{-1}$, such as in the oxic zone of wetlands (Le Mer and Roger 2001). Known controls on CH_4 consumption by low affinity methanotrophs are water table position, which dictates the size of the oxic zone necessary for methanotrophy, pH, and temperature (reviewed by Whalen 2005). Demonstrated controls on atmospheric CH_4 consumption by high affinity methanotrophs in upland soils include temperature (Crill 1991, Castro et al. 1995, Phillips et al. 2001a, Steinkamp et al. 2001), soils nitrogen (Schnell and King 1994, 1995), and rate of supply of CH_4 to the subsurface aerobic zone of oxidation (King and Adamsen 1992, Dörr et al. 1993, King 1997).

Methane is produced by methanogenic Archaea through two different anaerobic metabolic processes, acetate splitting and CO_2 reduction. Of all metabolic pathways, methanogenesis yields the least free energy and methanogenic Archaea are typically out-competed by microbes with alternative metabolic pathways (Schlesinger 1997), except when the redox potential is very low, such as in persistently anoxic wetlands. The absence of oxygen, which is related to soil moisture, the availability of labile organic precursors, temperature, and pH are known controls on CH_4 production. Accordingly, wetlands and freshwater sediments, with low redox potentials and high levels of organic matter, provide natural environments favorable to methanogenesis. Low redox environments with high availability of labile organic matter have also been observed in aggregates of clay-rich forest soils (Sexstone, A.J. et al. 1985, Ramakrishnan et al. 2000). Independent reports of anoxic microzones (Sexstone, Alan J. et al. 1985, Zausig et al. 1993) and methanogenic activity in macroscopically oxygenated soils (Yavitt et al. 1995, Saari et al. 1997, von Fischer and Hedin 2002, Teh et al. 2005) indicate that simultaneous CH_4 production and consumption are

occurring in some well-drained upland soils. Waterlogged aggregates support localized zones of methanogenesis and oxic sites support methanotrophy.

Observed effects of elevated CO₂ on CH₄ dynamics in soils

Experiments examining the effect of elevated CO₂ on net CH₄ emissions from wetland soils unequivocally indicate that CH₄ emissions increase when wetland plants, plant communities, or ecosystems are grown under elevated CO₂. Increases in CH₄ emissions from wetland soils ranged from 10.9% in a pot study of a rice cultivars grown under CO₂ at 200 mL L⁻¹ above ambient concentrations (Lou et al. 2008) to 60% when rice fields were exposed to elevated CO₂ (300 mL L⁻¹ above ambient) in open-top chambers (Ziska et al. 1998). The increase in net CH₄ emissions was markedly similar to the range (38 to 58%) seen in a Japanese rice paddy exposed to elevated CO₂ via FACE technology (Inubushi et al. 2003).

Investigations of soil-atmosphere CH₄ exchange in CO₂-enriched ecosystems that normally function as atmospheric CH₄ sinks are few and show mixed results. Ambus and Robertson (1999) reported a 22% reduction in CH₄ consumption by soils in model *Populus tremuloides* (deciduous forest) ecosystem exposed to elevated CO₂, while Phillips et al. (2001a) showed annual reductions in CH₄ consumption of 16% and 30% in CO₂ fumigated plots (200 mL L⁻¹) relative to plots exposed ambient atmospheres in a 2 y study in a temperate forest. In grasslands, Ineson et al. (1998) observed that rates of atmospheric CH₄ uptake were three times greater in ambient CO₂ soils relative to CO₂-enriched plots in an N-fertilized sward of *Lolium perenne*, but a subsequent investigation (Baggs and Blum 2004) found a significant interaction between N fertilizer application rate and CO₂ on atmospheric

CH₄ consumption. Further, Mosier et al. (2002) saw no impact of CO₂ level on rates of CH₄ exchange between soils and the atmosphere in a semi-arid, mixed grassland community.

If the observed CO₂-induced increases in net CH₄ emissions are extrapolated to the global scale, the wetland (natural and agricultural environments) source strength in the atmospheric CH₄ budget will increase by 29 and 160 Tg annually with a 200 to 300 mL L⁻¹ increase in atmospheric CO₂ concentration (Chen and Prinn 2005). At the same time, models suggest that the annual forest sink of 24 Tg y⁻¹ for CH₄ (Ridgwell et al. 1999) can be expected to decline from between 3.8 to 7.2 Tg as atmospheric CO₂ concentrations increase by 200 mL L⁻¹. However, more empirical data are needed before we can rely on these predictions of changes in CH₄ source and sink terms with increasing atmospheric CO₂. The few extant observational records of < 2 y in forest ecosystems and < 3 y in wetlands are insufficient to distinguish between transient and equilibrium responses of forest and wetland ecosystems to elevated CO₂ and the impact of those responses on CH₄ cycling.

Ecosystem-level changes that may influence CH₄ dynamics in temperate forests

Several CO₂-induced changes in temperate forest ecosystems may help to explain the observed decline in net CH₄ consumption under elevated CO₂. Changes in plant productivity, chemistry, and allocation of C under elevated CO₂ impacts the quantity and quality of C in the ecosystem, and the supply and availability of C to soil organisms. Some C compounds, such as phenolics, tannins and terpenes inhibit metabolism and growth by some soil microorganisms. Examples of enhanced delivery of C to the soil under elevated CO₂ in FACE studies include increased labile dissolved organic C in throughfall, (Lichter et al. 2000b), a small increase in the storage of C in forest soils (Matamala and Schlesinger 2000,

Lichter et al. 2008) and increased root productivity and mortality (Pritchard et al. 2008). Enhanced root exudation of organic acids has been observed in a pot study of *Pinus echinata* seedlings (Norby et al. 1987), while greater litter fall in both FACE and microcosm studies (Allen et al. 2000, Lichter et al. 2005, Lichter et al. 2008, Liu et al. 2009) has been reported under elevated CO₂. Further, several researchers have seen changes in the abundance of secondary C compounds in tissues and root exudates between plants grown under elevated and ambient CO₂ (Peñuelas and Estiarte 1998, Verburg et al. 1999, Tuchman et al. 2002, Billings and Ziegler 2005, Wetzel and Tuchman 2005a). Secondary C compounds, such as phenolics and terpenes, inhibit metabolism and growth by broad groups of soil bacteria (Souto et al. 2000), and specifically, methanotrophs (Amaral and Knowles 1997, 1998).

Reduced net CH₄ consumption under elevated CO₂ in temperate forests may also be the result of higher soil moisture and the associated reduction in diffusion of atmospheric gases. Reduced gas diffusivity has been demonstrated (Dörr et al. 1993) to control rates of CH₄ supply to the usual subsurface locus of CH₄ oxidation (e.g. Whalen and Reeburgh 1992), which is itself substrate-limited in well-drained forest soils, based on kinetic considerations (Bradford et al. 2001). Thicker leaf litter in forests exposed to elevated CO₂ (Allen et al. 2000, Lichter et al. 2005, Lichter et al. 2008, Liu et al. 2009) can result in higher soil moisture because of reduced evaporation from the soil surface. Increased soil moisture in turn slows the transport of gases within the soil matrix (Suwa et al. 2004). In fact, a direct link between increased soil moisture and diffusion-limitation of substrate to CH₄ oxidizers is well established (Striegl 1993, Castro et al. 1995, Whalen and Reeburgh 1996). The excess of litterfall under elevated CO₂ additionally directly adds to diffusional resistance in soils, and experimental litter removal has been shown to increase rates of net atmospheric CH₄

consumption in forest soils by as much as 43% (Dong et al. 1998, Brumme and Borken 1999).

Finally, reduced diffusion of atmospheric O₂, because of thicker leaf litter and higher soil moisture, along with higher soil respiration (Bernhardt et al. 2006, Taneva et al. 2006), and increased soil aggregation (Hoosbeek and Scarascia-Mugnozza 2009) under elevated CO₂ may increase the incidence of anoxic microsites where anaerobic microbial metabolism, such as methanogenesis, is possible. Since net CH₄ consumption in upland soils is the net effect of CH₄ consumption in the oxic soil profiles and CH₄ production in anoxic microsites, increased incidence of anoxic loci can alter this balance, reducing rates of net CH₄ consumption or shifting localized areas to net CH₄ sources. Horn and Smucker (2005) found when soil aggregates were saturated with water, the redox potential decreased rapidly, making these soil aggregates transiently anoxic within an otherwise oxic profile.

Research objectives

This is a follow-up study to previous research reported by Phillips et al. (2001a), who saw 16 and 30% annual reductions in rates of net CH₄ consumption by soils in a temperate forest enriched with elevated CO₂. The cause(s) of the decline in rates of net CH₄ consumption were not identified and the persistence of such a reduction beyond 2 y was not determined. Therefore, the purpose of this dissertation is to a) determine if reduced net CH₄ consumption by the same temperate forest soils is a sustained response to elevated CO₂; and b) identify factor(s) contributing to the observed (Phillips et al. 2001a) decline in net CH₄ consumption under elevated CO₂ at the Duke Forest FACE site. Possible controls on CH₄ consumption resulting from elevated CO₂ concentrations include negative impacts of altered

organic compounds from the surrounding forest ecosystem on CH₄ oxidizing communities, higher soil moisture and an associated reduction in the supply of CH₄ to the zone of CH₄ oxidation, or a shift in the rates of consumption and production by CH₄ oxidizing and producing communities, respectively.

Model projections of future climates are strongly dependent on atmospheric concentrations of radiatively and chemically important trace gases, such as CH₄. Therefore, my intention is for this research to be used to improve model projections of future climates, with special attention to the feedbacks of elevated CO₂ on ecosystem components that control CH₄ dynamics within forest soils.

Dissertation structure

This dissertation has been written as 5 chapters. Chapters 2 through 4 were written with the intention of submitting each chapter as individual manuscripts. Chapters 1 and 5 introduce and conclude, respectively, the body of work. Chapter 2 shows an extension of the previous 2 y record of soil-atmosphere exchange of CH₄ in CO₂-enriched and free-air (control) plots to establish the long-term response of atmospheric CH₄ consumption under elevated CO₂. In Chapter 2, I also investigate if treatment-wise differences or interactions in environmental measures (soil moisture and temperature) account for reduced atmospheric CH₄ consumption in CO₂-enriched plots. Chapter 3 investigates the possibility of plant exudate control on CH₄ consumption in soils from the same study site. Chapter 4 evaluates the depth distribution of CH₄ in the soil profile, the effective diffusivity of CH₄ through the soil, as well as the extent and activity of CH₄ consuming and producing communities at the study site. This structure may result in some repetition in introductory material and

discussion of results.

References

- Allen, A. S., J. A. Andrews, A. C. Finzi, R. Matamala, D. D. Richter, and W. H. Schlesinger. 2000. Effects of free-air CO₂ enrichment (FACE) on belowground processes in a *Pinus taeda* forest. *Ecological Applications* **10**:437-448.
- Amaral, J. A. and R. Knowles. 1997. Inhibition of methane consumption in forest soils and pure cultures of methanotrophs by aqueous forest soil extracts. *Soil Biology and Biochemistry* **29**:1713-1720.
- Amaral, J. A. and R. Knowles. 1998. Inhibition of methane consumption in forest soils by monoterpines. *Journal of Chemical Ecology* **24**:723-734.
- Ambus, P. and G. P. Robertson. 1999. Fluxes of CH₄ and N₂O in aspen stands grown under ambient and twice-ambient CO₂. *Plant and Soil* **209**:1-8.
- Amthor, J. S. 1995. Terrestrial higher-plant response to increasing atmospheric CO₂ in relation to the global carbon cycle. *Global Change Biology* **1**:243-274.
- Baggs, E. M. and H. Blum. 2004. CH₄ oxidation and emissions of CH₄ and N₂O from *Lolium perenne* swards under elevated atmospheric CO₂. *Soil Biology and Biochemistry* **36**:713-723.
- Barnola, J.-M., D. Raynaud, C. Lorius, and N. I. Barkov. 2003. Historical Carbon Dioxide Record from the Vostok Ice Core. *in* O. R. N. L. Carbon Dioxide Information Analysis Center, Department of Energy, editor. *Trends: A Compendium of Data on Global Change*, Oak Ridge, Tenn., U.S.A.
- Bernhardt, E., J. Barber, J. Pippen, L. Taneva, J. Andrews, and W. Schlesinger. 2006. Long-term effects of free air CO₂ enrichment (FACE) on soil respiration. *Biogeochemistry* **77**:91-116.
- Billings, S. A. and S. E. Ziegler. 2005. Linking microbial activity and soil organic matter transformations in forest soils under elevated CO₂. *Global Change Biology* **11**:203-212.
- Billings, S. A. and S. E. Ziegler. 2008. Altered patterns of soil carbon substrate usage and heterotrophic respiration in a pine forest with elevated CO₂ and N fertilization. *Global Change Biology* **14**:1025-1036.
- Bradford, M. A., P. Ineson, P. A. Wookey, and H. M. Lappin-Scott. 2001. Role of CH₄ oxidation, production and transport in forest soil CH₄ flux. *Soil Biology and Biochemistry* **33**:1625-1631.

- Brumme, R. and W. Borken. 1999. Site variation in methane oxidation as affected by atmospheric deposition and type of temperate forest ecosystem. *Global Biogeochemical Cycles* **13**:493-501.
- Castro, M. S., P. A. Steudler, J. M. Melillo, J. D. Aber, and R. D. Bowden. 1995. Factors controlling atmospheric methane consumption by temperate forest soils. *Global Biogeochemical Cycles* **9**:1-10.
- Chen, Y.-H. and R. G. Prinn. 2005. Atmospheric modeling of high- and low-frequency methane observations: Importance of interannually varying transport. *Journal of Geophysical Research* **110**: D10303.
- Conrad, R. 1996. Soil microorganisms as controllers of atmospheric trace gases (H₂, CO, CH₄, OCS, N₂O, and NO). *Microbiological Reviews* **60**:609-640.
- Crill, P. M. 1991. Seasonal patterns of methane uptake and carbon dioxide release by a temperate woodland soil. *Global Biogeochemical Cycles* **5**:319-334.
- DeLucia, E. H., J. G. Hamilton, S. L. Naidu, R. B. Thomas, J. A. Andrews, A. Finzi, M. Lavine, R. Matamala, J. E. Mohan, G. R. Hendrey, and W. H. Schlesinger. 1999. Net primary production of a forest ecosystem with experimental CO₂ enrichment. *Science* **284**:1177-1179.
- DeLucia, E. H., D. J. Moore, and R. J. Norby. 2005. Contrasting responses of forest ecosystems to rising atmospheric CO₂: Implications for the global C cycle. *Global Biogeochemical Cycles* **19**: GB3006.
- Denman, K. L., G. Brasseur, A. Chidthaisong, P. Ciais, P. Cox, R. E. Dickinson, D. Hauglustaine, C. Heinze, E. Holland, D. Jacob, D. Lohmann, S. Ramachandran, P. L. da Silva Dias, S. C. Wofsy, and X. Zhang. 2007. Couplings Between Changes in the Climate System and Biogeochemistry. *in* S. Solomon, D. Qin, M. Manning, J. Chen, and M. Marquis, editors. *Climate Change 2007: The Physical Sciences Basis. Contributions of the Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Dong, Y., D. Scharffe, J. M. Lobert, P. J. Crutzen, and E. Sanhueza. 1998. Fluxes of CO₂, CH₄, and N₂O from a temperate forest soil: the effects of leaves and humus layers. *Tellus* **50B**:243-252.
- Dörr, H., L. Katruff, and I. Levin. 1993. Soil texture parameterization of the methane uptake in aerated soils. *Chemosphere* **26**:697-713.
- Finzi, A. C., E. H. DeLucia, J. G. Hamilton, D. D. Richter, and W. H. Schlesinger. 2002. The nitrogen budget of a pine forest under free air CO₂ enrichment. *Oecologia* **132**:567-578.

- Finzi, A. C., D. J. P. Moore, E. H. DeLucia, J. Lichter, K. S. Hofmockel, R. B. Jackson, H.-S. Kim, R. Matamala, H. R. McCarthy, R. Oren, J. S. Phippen, and W. H. Schlesinger. 2006a. Progressive nitrogen limitation of ecosystem processes under elevated CO₂ in a warm-temperate forest *Ecology* **87**:15-25.
- Finzi, A. C., R. L. Sinsabaugh, T. M. Long, and M. P. Osgood. 2006b. Microbial community responses to atmospheric carbon dioxide enrichment in a warm-temperate forest. *Ecosystems* **9**:215-226.
- Forster, P., V. Ramaswamy, P. Artaxo, T. Berntsen, R. Betts, D. W. Fahey, J. Haywood, J. Lean, D. C. Lowe, G. Myhre, J. R. Nganga, R. Prinn, G. Raga, M. Schulz, and R. Van Dorland. 2007. Changes in Atmospheric Constituents and in Radiative Forcing. *in* S. Solomon, D. Qin, M. Manning, J. Chen, M. Marquis, K. B. Averyt, M. Tignor, and A. J. Miller, editors. *Climate Change 2007: The Physical Sciences Basis. Contribution of the Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Friedlingstein, P., P. Cox, R. Betts, L. Bopp, W. von Bloh, V. Brovkin, P. Cadule, S. Doney, M. Eby, I. Fung, G. Bala, J. John, C. Jones, F. Joos, T. Kato, M. Kawamiya, W. Knorr, K. Lindsay, H. D. Matthews, and T. Raddatz. 2006. Climate- Carbon Cycle Feedback Analysis: Results from the C₄MIP Model Intercomparison. *Journal of Climate* **19**:3337-3353.
- Hamilton, J., E. DeLucia, K. George, S. Naidu, A. Finzi, and W. Schlesinger. 2002. Forest carbon balance under elevated CO₂. *Oecologia* **131**:250-260.
- Hanson, R. S. and T. E. Hanson. 1996. Methanotrophic bacteria. *Microbiological Reviews* **60**:439-471.
- Hendrey, G. R., D. S. Ellsworth, K. F. Lewin, and J. Nagy. 1999. A free-air enrichment system for exposing tall forest vegetation to elevated atmospheric CO₂. *Global Change Biology* **5**:293-309.
- Hoosbeek, M. and G. Scarascia-Mugnozza. 2009. Increased litter build up and soil organic matter stabilization in a poplar plantation after 6 years of atmospheric CO₂ enrichment (FACE): final results of POP-EuroFACE compared to other forest FACE experiments. *Ecosystems* **12**:220-239.
- Horn, R. and A. Smucker. 2005. Structure formation and its consequences for gas and water transport in unsaturated arable and forest soils. *Soil and Tillage Research* **82**:5-14.
- Ineson, P., P. A. Coward, and U. A. Hartwig. 1998. Soil gas fluxes of N₂O, CH₄ and CO₂ beneath *Lolium perenne* under elevated CO₂: The Swiss free air carbon dioxide enrichment experiment. *Plant and Soil* **198**:89-95.

- Inubushi, K., W. Cheng, S. Aonuma, M. M. Hoque, K. Kobayashi, S. Miura, H. Y. Kim, and M. Okada. 2003. Effects of free-air CO₂ enrichment (FACE) on CH₄ emission from a rice paddy field. *Global Change Biology* **9**:1458-1464.
- Jastrow, J. D., R. M. Miller, R. Matamala, R. J. Norby, T. W. Boutton, C. W. Rice, and C. E. Owensby. 2005. Elevated atmospheric carbon dioxide increase soil carbon. *Global Change Biology* **11**:2057-2064.
- King, G. M. 1997. Responses of atmospheric methane consumption by soils to global climate change. *Global Change Biology* **3**:351-362.
- King, G. M. and A. P. S. Adamsen. 1992. Effects of temperature on methane consumption in a forest soil and in pure cultures of the methanotroph *Methylobacterium rubra*. *Applied Environmental Microbiology* **58**:2758-2763.
- King, J. S., P. J. Hanson, E. Bernhardt, P. Deangelis, R. J. Norby, and K. Pregitzer. 2004. A multiyear synthesis of soil respiration responses to elevated atmospheric CO₂ from four forest FACE experiments. *Global Change Biology* **10**:1027-1042.
- Klironomos, J. N., M. F. Allen, M. C. Rillig, J. Piotrowski, S. Makvandi-Nejad, B. E. Wolfe, and J. R. Powell. 2005. Abrupt rise in atmospheric CO₂ overestimates community response in a model plant-soil system. *Nature* **433**:621-624.
- Korner, C. 2000. Biosphere responses to CO₂ enrichment. *Ecological Applications* **10**:1590-1619.
- Larson, J., D. R. Zak, and R. L. Sinsabaugh. 2002. Extracellular enzyme activity and metabolism of root-derived substrates beneath temperate trees growing under elevated CO₂ and O₃. *Soil Science Society of America Journal* **66**:1848-1856.
- Le Mer, J. and P. Roger. 2001. Production, oxidation, emission and consumption of methane by soils: A review. *European Journal of Soil Biology* **37**:25-50.
- Leakey, A. D. B., E. A. Ainsworth, C. J. Bernacchi, A. Rogers, S. P. Long, and D. R. Ort. 2009. Elevated CO₂ effects on plant carbon, nitrogen, and water relations: six important lessons from FACE. *Journal of Experimental Botany*:erp096.
- Lichter, J., S. H. Barron, C. E. Bevacqua, A. C. Finzi, K. F. Irving, E. M. Stemmler, and W. Schlesinger. 2005. Soil carbon sequestration and turnover in a pine forest after six years of atmospheric CO₂ enrichment. *Ecology* **86**:1835-1847.
- Lichter, J., S. A. Billings, S. E. Ziegler, D. Indh, R. Ryals, A. C. Finzi, R. B. Jackson, E. M. Stemmler, and W. H. Schlesinger. 2008. Soil carbon sequestration in a pine forest after 9 years of atmospheric CO₂ enrichment. *Global Change Biology* **14**:1-13.

- Lichter, J., M. Lavine, K. A. Mace, D. D. Richter, and W. H. Schlesinger. 2000. Throughfall chemistry in a loblolly pine plantation under elevated atmospheric CO₂ concentrations. *Biogeochemistry* **50**:73-93.
- Liu, L., J. S. King, F. L. Booker, C. P. Giardina, H. L. Allen, and S. Hu. 2009. Enhanced litter input rather than changes in litter chemistry drive soil carbon and nitrogen cycles under elevated CO₂: a microcosm study. *Global Change Biology* **15**:441-453.
- Lou, Y., K. Inubushi, T. Mizuno, T. Hasegawa, Y. Lin, H. Sakai, W. Cheng, and K. Kobayashi. 2008. CH₄ emission with differences in atmospheric CO₂ enrichment and rice cultivars in a Japanese paddy soil. *Global Change Biology* **14**:2678-2687.
- Matamala, R. and W. H. Schlesinger. 2000. Effects of elevated atmospheric CO₂ on fine root production and activity in an intact temperate forest ecosystem. *Global Change Biology* **6**:967-979.
- McMurtrie, R. E. and H. N. Comins. 1996. The temporal response of forest ecosystems to doubled atmospheric CO₂ concentration. *Global Change Biology* **2**:49-57.
- Mosier, A. R., J. A. Morgan, J. Y. King, D. LeCain, and D. G. Milchunas. 2002. Soil-atmosphere exchange of CH₄, CO₂, NO_x, and N₂O in the Colorado shortgrass steppe under elevated CO₂. *Plant and Soil* **240**:201-211.
- Newton, P. C. D., H. Clark, G. R. Edwards, and D. J. Ross. 2001. Experimental confirmation of ecosystem model predictions comparing transient and equilibrium plant responses to elevated atmospheric CO₂. *Ecology Letters* **4**:344-347.
- NOAA. 2008. Carbon dioxide, methane rise sharply. Page http://www.noaaneews.noaa.gov/stories2008/20080423_methane.html. NOAA News.
- Norby, R. J., E. H. DeLucia, B. Gielen, C. Calfapietra, C. P. Giardina, J. S. King, J. Ledford, H. R. McCarthy, D. J. P. Moore, R. Ceulemans, P. De Angelis, A. C. Finzi, D. F. Karnosky, M. E. Kubiske, M. Lukac, K. S. Pregitzer, G. E. Scarascia-Mugnozza, W. H. Schlesinger, and R. Oren. 2005. Forest response to elevated CO₂ is conserved across a broad range of productivity. *Proceedings of the National Academy of Sciences of the United States of America* **102**:18052-18056.
- Norby, R. J., E. G. O'Neill, W. G. Hood, and R. J. Luxmoore. 1987. Carbon allocation, root exudation, and mycorrhizal colonization of *Pinus echinata* seedlings grown under CO₂ enrichment. *Tree Physiology* **3**:203-210.
- Peñuelas, J. and M. Estiarte. 1998. Can elevated CO₂ affect secondary metabolism and ecosystem function? *Trees* **13**:20-24.

- Phillips, R. L., S. C. Whalen, and W. H. Schlesinger. 2001. Influence of atmospheric CO₂ enrichment on methane consumption in a temperate forest soil. *Global Change Biology* **7**:557-563.
- Pritchard, S. G., A. E. Strand, M. L. McCormack, M. A. Davis, A. C. Finzi, R. B. Jackson, R. Matamala, H. H. Rogers, and R. Oren. 2008. Fine root dynamics in a loblolly pine forest are influenced by free-air-CO₂-enrichment: a six-year-minirhizotron study. *Global Change Biology* **14**:588-602.
- Ramakrishnan, B., T. Lueders, R. Conrad, and M. Friedrich. 2000. Effect of soil aggregate size on methanogenesis and archaeal community structure in anoxic rice field soil. *Fems Microbiology Ecology* **32**:261-270.
- Ridgwell, A. J., S. J. Marshall, and K. Gregson. 1999. Consumption of atmospheric methane by soils: a process-based model. *Global Biogeochemical Cycles* **13**: 59-70.
- Saari, A., P. J. Martikainen, A. Ferm, J. Ruuskanen, W. De Boer, S. R. Troelstra, and H. J. Laanbroek. 1997. Methane oxidation in soil profiles of Dutch and Finnish coniferous forests with different soil texture and atmospheric nitrogen deposition. *Soil Biology and Biochemistry* **29**:1625-1632.
- Schindell, D. T., G. Faluvegi, N. Bell, and G. A. Schmidt. 2005. An emission-based view of climate forcing by methane and tropospheric ozone. *Geophysical Research Letters* **32**: LO4803.
- Schlesinger, W. H. 1997. *Biogeochemistry: An Analysis of Global Change*. 2nd edition. Academic Press, San Diego, CA.
- Schlesinger, W. H. and J. Lichter. 2001. Limited carbon storage in soil and litter of experimental forest plots under increased atmospheric CO₂. *Nature* **411**:466-469.
- Schnell, S. and G. M. King. 1994. Mechanistic analysis of ammonium inhibition of atmospheric methane consumption in forest soils. *Applied Environmental Microbiology* **60**:3514-3521.
- Schnell, S. and G. M. King. 1995. Stability of methane oxidation capacity to variations in methane and nutrient concentrations. *FEMS Microbiology Ecology* **18**:285-294.
- Sexstone, A. J., N. P. Revsbech, T. B. Parkin, and J. M. Tiedje. 1985. Direct measurement of oxygen profiles and denitrification rates in soil aggregates. *Soil Science Society of America Journal* **49**:645-651.
- Sexstone, A. J., N. P. Revsbech, T. B. Parkin, and J. M. Tiedje. 1985. Direct measurement of oxygen profiles and denitrification rates in soil aggregates. *Soil Science Society of America Journal* **49**:645-651.

- Souto, X. C., G. Chiapusio, and F. Pellissier. 2000. Relationships between phenolics and soil microorganisms in spruce forests: significance for natural regeneration. *Journal of Chemical Ecology* **26**:2025-2034.
- Steinkamp, R., K. Butterbach-Bahl, and H. Papen. 2001. Methane oxidation by soils of an N limited and N fertilized spruce forest in the Black Forest, Germany. *Soil Biology and Biochemistry* **33**:145-153.
- Striegl, R. G. 1993. Diffusional limits to the consumption of atmospheric methane by soils. *Chemosphere* **26**:715-720.
- Suwa, M., G. G. Katul, R. Oren, J. Andrews, J. Pippen, A. Mace, and W. H. Schlesinger. 2004. Impact of elevated atmospheric CO₂ on forest floor respiration in a temperate pine forest. *Global Biogeochemical Cycles* **18**: GB2013.
- Taneva, L., J. S. Pippen, W. H. Schlesinger, and M. A. Gonzalez-Meler. 2006. The turnover of carbon pools contributing to soil CO₂ and soil respiration in a temperate forest exposed to elevated CO₂ concentration. *Global Change Biology* **12**:983-994.
- Teh, Y. A., W. L. Silver, and R. Conrad. 2005. Oxygen effects on methane production and oxidation in humid tropical forest soils. *Global Change Biology* **11**:1283-1297.
- Tuchman, N. C., R. G. Wetzel, S. T. Rier, K. A. Wahtera, and J. A. Teeri. 2002. Elevated atmospheric CO₂ lowers leaf litter nutritional quality for stream ecosystem food webs. *Global Change Biology* **8**:163-170.
- Verburg, P. S. J., W. K. P. Van Loon, and A. L¹/₄kewille. 1999. The CLIMEX soil-heating experiment: soil response after 2 years of treatment. *Biology and Fertility of Soils* **28**:271-276.
- von Fischer, J. C. and L. O. Hedin. 2002. Separating methane production and consumption with a field-based isotope pool dilution technique. *Global Biogeochemical Cycles* **16**: 1034.
- Wetzel, R. G. and N. C. Tuchman. 2005. Effects of atmospheric CO₂ enrichment and sunlight on degradation of plant particulate and dissolved organic matter and microbial utilization. *Archiv für Hydrobiologie* **162**:287-308.
- Whalen, S. C. 2005. Biogeochemistry of Methane Exchange between Natural Wetlands and the Atmosphere. *Environmental Engineering Science* **22**:73-94.
- Whalen, S. C. and W. S. Reeburgh. 1992. Interannual variations in tundra methane emission: a 4-year time series at fixed sites. *Global Biogeochemical Cycles* **6**:139-159.

- Whalen, S. C. and W. S. Reeburgh. 1996. Moisture and temperature sensitivity of CH₄ oxidation in boreal soils. *Soil Biology and Biochemistry* **28**:1271-1281.
- Wuebbles, D. J. and K. Hayhoe. 2002. Atmospheric methane and global change. *Earth-Science Reviews* **57**:177-210.
- Yavitt, J. B., T. J. Fahey, and J. A. Simmons. 1995. Methane and carbon dioxide dynamics in a northern hardwood ecosystem. *Soil Science Society of America Journal* **59**:796-804.
- Zausig, J., W. Stepniewski, and R. Horn. 1993. Oxygen concentration and redox potential gradients in unsaturated model soil aggregates. *Soil Science Society of America Journal* **57**:908-916.
- Ziska, L. H., T. B. Moya, R. Wassmann, O. S. Namuco, R. S. Lantin, J. B. Aduna, A. Abao Jr., K. F. Bronson, H. U. Neue, and D. Olszyk. 1998. Long-term growth at elevated carbon dioxide stimulates methane emission in tropical paddy rice. *Global Change Biology* **4**:657-66.

CHAPTER 2: REDUCED NET CH₄ CONSUMPTION IS A SUSTAINED RESPONSE TO ELEVATED CO₂ IN A TEMPERATE FOREST

Abstract

We compared, from 2004 through 2006, rates of soil-atmosphere CH₄ exchange at permanently established sampling sites in a temperate forest exposed to ambient (control plots; ~380 mL L⁻¹) or elevated (ambient + 200 mL L⁻¹) CO₂ since August 1996. A total of 880 observations showed net atmospheric CH₄ consumption (flux from the atmosphere to the soil) from all static chambers most of the time at rates varying from 0.02 mg m⁻² d⁻¹ to 4.5 mg m⁻² d⁻¹. However, we infrequently found net CH₄ production (flux from the soil to the atmosphere) at lower rates, 0.01 mg m⁻² d⁻¹ to 0.08 mg m⁻² d⁻¹. For the entire study, the mean rate of net CH₄ consumption in control plots was higher than the mean for CO₂-enriched plots, 0.55 (± 0.03 SEM) versus 0.51 (± 0.03 SEM) mg m⁻² d⁻¹. Annual rates of 184, 196 and 197 mg m⁻² for net CH₄ consumption at control plots during the three calendar years of this study were 19, 10 and 8% higher than comparable values for CO₂ enriched plots. Differences between treatments were significant (p < 0.05) in 2004 and 2005 and nearly significant (p = 0.10) in 2006. Volumetric soil water content was consistently higher at CO₂-enriched sites and a mixed effects model identified a significant soil moisture x CO₂ interaction on net atmospheric CH₄ consumption. Increased soil moisture at CO₂-enriched sites likely increases diffusional resistance of surface soils and the frequency of anaerobic microsites supporting methanogenesis,

resulting in reduced rates of net atmospheric CH₄ consumption. Our study extends previous observations of reduced net atmospheric CH₄ consumption at CO₂-enriched plots at this site to nearly 8 continuous years, suggesting that this is likely a sustained negative feedback to increasing atmospheric CO₂.

Introduction

The atmospheric concentration of CH₄ has more than doubled since the Industrial Revolution to a present-day value of ~1782 μL L⁻¹ (Forster et al. 2007). This generalized increase is of concern because CH₄ is second only to CO₂ among trace atmospheric constituents with respect to radiative forcing and is also chemically active in the atmosphere, playing an important role in stratospheric and tropospheric ozone chemistry (Denman et al. 2007).

The atmospheric concentration of CO₂ has increased parallel to that of CH₄, and is projected to reach 730 mL L⁻¹ by 2100, a level that exceeds the preindustrial concentration by 260% (Forster et al. 2007). Although the unprecedented rate of change in the atmospheric concentrations of CO₂ and CH₄ over the last 250 y and the influences on climate are well documented, the reasons for changing abundances are not entirely clear. Model projections of future climate are strongly dependent on atmospheric concentrations of radiatively and chemically important trace gases. Cycling of long-lived greenhouse gases such as CO₂ and CH₄ are dominated or supported by a biospheric component responsible for the production and consumption of these gases, and for modulating or mediating gas exchange between the pedosphere or hydrosphere and atmosphere. However, improvements to current models require a comprehensive

understanding of the linkage between biogeochemical processes and the troposphere with respect to trace atmospheric constituents that influence climate, and further identification of the interactions between biogeochemical cycles that impact exchange of trace gases between soil or water and the atmosphere.

The balance between rates of CH₄ production (methanogenesis) and consumption (methanotrophy) determines whether a soil is a net source or sink for atmospheric CH₄. Methane production usually exceeds consumption in wetland environments, accounting for about 69% of emissions to the atmosphere from natural sources (Wuebbles and Hayhoe 2002). In contrast, well-drained soils generally display net consumption of CH₄, and constitute the only biological loss term in the atmospheric CH₄ budget. Little is understood about the effect of elevated CO₂ on biogeochemical processes affecting CH₄ cycling dynamics. However, Phillips et al. (2001a) previously showed, in a short term (2 y) study, annual reductions in CH₄ consumption of 16% and 30% in CO₂-fumigated plots relative to plots exposed to ambient atmospheres in an upland temperate forest. Similar investigations on shorter time scales (weeks to 2 mo) report reduced atmospheric CH₄ consumption under elevated CO₂ in a deciduous forest (Ambus and Robertson 1999), and give mixed results for a grassland (Ineson et al. 1998, Baggs and Blum 2004), although results in these studies include nitrogen x CO₂ fertilization interactions. In contrast, Mosier et al. (2002) reported no CO₂-induced response in atmospheric CH₄ consumption in a shortgrass steppe over 4 y.

A negative feedback on forest soil CH₄ consumption by rising CO₂ has important implications for the atmospheric CH₄ budget. Sink strength estimates for upland soils center around 30 Tg y⁻¹, or about 75% of the stratospheric sink of 40 Tg y⁻¹ (Denman et

al. 2007). The few extant observational records of < 2 y, however, are insufficient to distinguish between transient and equilibrium responses of forest ecosystems to elevated CO₂ or to determine whether the observed response will be sustained. Ecosystem models indicate that plant/community responses to elevated CO₂ and biogeochemical feedbacks can change over time (Newton et al. 2001). It is therefore critical to identify the long-term trajectory of the sign and magnitude of change.

Our study was conducted in an aggrading temperate forest where experimental plots had been continuously fumigated with CO₂. Our objectives were to: (a) extend a previous 2 y record of soil-atmosphere exchange of CH₄ in CO₂-enriched and free-air (control) plots (Phillips et al. 2001a) to establish the long term response of atmospheric CH₄ consumption under elevated CO₂; and (b) relate environmental measures (soil moisture and temperature) to rates of gas exchange to determine if treatment-wise differences or interactions in these well known controls on soil methanotrophy may account for reduced atmospheric CH₄ consumption in CO₂ enriched plots. A firmer understanding of the feedback between increasing atmospheric CO₂ and the rates and controls on CH₄ oxidation in forest soils will aid in the refinement of process-based models that contribute to larger efforts directed at predicting future climates.

Methods

Field site

Field measurements were conducted at the Duke Forest (North Carolina; USA) Free-Air CO₂ Enrichment (FACE) experiment sited in an even-aged stand of loblolly pine (*Pinus taeda* L.) planted in 1983. Soils are clay loam, Ultic Hapludalf's of the Enon

Series (Oh and Richter 2005). Average normal air temperature ranges from 3.6 ° C in January to 25.3 ° C in July and annual precipitation averages 1209 mm (State Climate Office of North State Climate Office of North Carolina 2003-2009). Soil physical characteristics are similar between CO₂ treatment plots, with the exception of soil organic matter, which averaged 4.6% in CO₂-enriched plots, and only 3.4% in control plots. Averages for all control and elevated CO₂ plots (0 to 20 cm depth zone) for soil particle density, bulk density, and pH were 2.5 g cm⁻³, 1.2 g cm⁻³, and 5.7 units, respectively. Soil texture was 9% clay, 42% silt, and 49% sand.

Site characteristics are fully documented in Hendrey et al. (1999a) and briefly described here. The experiment consists of eight circular 30-m diameter plots. Four treatment plots (referred to as “CO₂-enriched”) are fumigated with CO₂ to maintain atmospheric CO₂ concentrations 200 mL L⁻¹ above ambient levels, while three additional treatment plots are fumigated with ambient air to replicate micrometeorological effects associated with CO₂ addition. A fourth is subjected to ambient air without fumigation. The latter four plots are referred to as “controls”. Continuous (24 h d⁻¹) fumigation was initiated in August 1996, but was reduced to daylight hours only from 2003 to present.

Gas Flux Measurements

Each plot is partitioned into four quadrants for a total of 24 (2004 and 2005) and 32 (2006) individual sectors. Methane flux determinations within each sector were made ~biweekly in the 2004, 2005, and 2006 calendar years using the static chamber technique (Whalen et al. 1992), yielding 12 (2004 and 2005) and 16 (2006) measurements in both control and enriched plots on each sampling date. The polyvinyl chloride collars (20 cm

diameter x 11 cm height) permanently deployed in three plots of each treatment at the conception (1999) of our initial investigation remained intact and were revisited for this study. Collars of similar design were deployed in each quadrant of two additional plots (one plot, each treatment) in 2006. Polyvinyl chloride covers fitted with a sampling port and capillary bleed were emplaced on soil collars for CH₄ flux determinations.

Headspace samples were withdrawn into 10 mL gastight glass syringes at zero time and at 0.5 h intervals thereafter to 2 h. Collars were open to litterfall and rainfall between sampling sessions.

Gas samples were analyzed for CH₄ by flame ionization detection gas chromatography (Shimadzu model GC 8 A; precision expressed as the coefficient of variation for 10 replicate injections of a 0.94 mL CH₄ L⁻¹ standard was < 3%) within 10 h of collection, well within our predetermined holding time of 24 h. Sample separation was accomplished on a 1-m length x 0.32 cm diameter molecular sieve 5A column with an ultrahigh purity N₂ carrier gas (33 mL min⁻¹). Injector and detector temperatures were set at 90 °C and 140 °C.

Soil Physicochemical Measurements

Volumetric soil moisture (mL H₂O cm⁻³ soil) was continuously measured by time domain reflectometry using Campbell Scientific Model CS616 probes. Probes were located randomly in each quadrant of each plot in calendar years 2004 and 2005 (Hyun unpublished) and within 30 cm of each soil collar in 2006. The soil moisture probes integrate volumetric soil moisture from the soil surface to 30 cm depth at 30 s intervals, and average values over 24 h are recorded on Campbell Scientific Model CR200

dataloggers. In conjunction with CH₄ flux determinations, soil temperature was measured at 3 cm intervals from 1 cm to 19 cm depth with a multithermistor temperature probe.

Calculations and statistics

Area-based rates of net CH₄ consumption were calculated from chamber geometry and the time-linear change of CH₄ concentration in chamber headspaces. Annual rates of net CH₄ consumption were determined by integrating for the calendar year daily, area-based data from each sampling occasion. Average soil temperature was calculated as the mean of equally spaced observations taken to 19 cm.

We analyzed for differences in CH₄ flux between CO₂ treatments with the same statistical model used in a previous study (Phillips et al. 2001a). The mixed effects linear model considered CO₂ treatment as the main effect, with soil moisture, temperature and time (continuous) as covariates. The model was a nested, hierarchical design with plot nested inside CO₂ and quadrant nested within plot. Unequal sampling intervals required the use of a time-series covariance structure, where correlations decline as a function of time. Only significant interactions remained in the model. The same model with a different nesting structure was used to analyze overall net CH₄ flux and environmental variables, in which quadrant was nested within plot and CO₂ was simply an effect.

Student t-tests were used to analyze for statistical differences between CO₂ treatment averages for environmental variables. Treatment-wise differences between annual rates of net CH₄ consumption were determined by confidence interval overlap. All statistical analyses were performed at $\alpha=0.05$.

Results

Environmental variables

Air temperatures averaged 15 °C or 16 °C annually for each calendar year. Soil temperatures ranged from 4 °C to 25 °C (Fig. 2.1), closely tracking air temperatures (not shown) and showed an average of 16 °C for the entire study. Overall, soil moisture varied from 0.16 to 0.49 mL H₂O cm⁻³ soil, and averaged 0.27 mL H₂O cm⁻³ soil. Calendar year means for CO₂-enriched plots were consistently higher than means for control plots (Table 2.1). Differences between treatment means for soil moisture were significant in 2004 and 2005, but not 2006. Over the entire study (n = 68), the average soil moisture for the CO₂-enriched treatment (0.28 mL H₂O cm⁻³ soil) was significantly higher than for the control treatment (0.26 mL H₂O cm⁻³ soil).

Patterns in net CH₄ flux

Net CH₄ consumption (flux from the atmosphere to the soil) was generally found at all individual soil chambers and was always calculated for each plot if fluxes from all four quadrants were averaged. However, net CH₄ production (flux from the soil to the atmosphere) was also observed, from 17 individual quadrants on 16 separate dates, giving 22 observations of net CH₄ production in 880 total records. Net CH₄ production was found almost twice as often in CO₂-enriched chambers as in control chambers (14 versus 8 observations). Rates of net CH₄ production from individual chambers varied from 0.01 mg m⁻² d⁻¹ to 0.08 mg m⁻² d⁻¹ while rates of net CH₄ consumption from individual chambers were much higher, varying from 0.02 mg m⁻² d⁻¹ to 4.5 mg m⁻² d⁻¹. Chamber-wise analysis showed no pattern with respect to magnitude of flux, as no chamber

showed consistently high or low values. Although there was no clear seasonal pattern, rates of net CH₄ consumption were frequently higher in the summer than the winter months (Fig. 2.1).

There was a strong inverse relationship between soil moisture and plot-averaged rates of net CH₄ consumption (Fig. 2.2). At the chamber level, the mixed-effects model used to test the factors contributing to overall net CH₄ fluxes showed that soil moisture was significantly related to net CH₄ flux in 2006, when soil moisture probes were installed proximal to chamber collars, but not in 2004 and 2005 when probes were randomly located within quadrants. Overall, net CH₄ consumption decreased with increasing soil moisture. The model showed no relationship between soil temperature and net CH₄ flux.

Differences in net CH₄ consumption between CO₂ treatments

When the entire data were considered (880 observations; each treatment), the mean net rate of CH₄ consumption in control chambers was 7.5% higher than in CO₂-enriched chambers, 0.55 (\pm 0.03 SEM) versus 0.51 (\pm 0.03) mg m⁻² d⁻¹. The difference was significant. The disparity in net CH₄ consumption rates between treatments showed interannual variability. Mean rates for controls in 2004, 2005 and 2006 were 0.53 (\pm 0.06), 0.54 (\pm 0.06) and 0.56 (\pm 0.05) mg CH₄ m⁻² d⁻¹. These values were higher by 10, 4 and 9% than corresponding annual averages for CO₂-enriched chambers. There was no seasonal pattern in the relative difference in net CH₄ consumption rates between treatments.

The mixed-effects model used to test the factors contributing to the variability in net CH₄ consumption between treatments indicated that CO₂ significantly interacted with soil moisture. Soils from CO₂-enriched plots consumed less CH₄ than soils from control plots, and the difference between CO₂ treatments increased with increasing soil moisture. Soil temperature had no effect.

The time-integrated rates of net CH₄ consumption in control plots were 184, 196 and 197 mg m⁻² y⁻¹ in 2004, 2005, and 2006 (Fig. 2.3). Comparable values for CO₂-enriched plots were lower by 19, 10, and 8% at 150, 175 and 181 mg m⁻² y⁻¹. Differences between treatments were significant in 2004 and 2005 and nearly significant in 2006 (p=0.10).

Discussion

Overall patterns of net CH₄ consumption and environmental correlates

Consumption of atmospheric CH₄ by well-drained forest soils is a common observation in all climatic zones of the world. The mean net CH₄ consumption rate of 0.54 mg m⁻² d⁻¹ in the present study is consistent with the value of 0.6 mg m⁻² d⁻¹ reported by both our group (Phillips et al. 2001a) and others (McLain et al. 2002) for studies conducted roughly 6 y previously. Mean rates of net CH₄ consumption for this site falls toward the low end of worldwide reports for aerated temperate forest soils, which show averages ranging from 0.2 to 5.0 mg CH₄ m⁻² d⁻¹ and center around 1 mg CH₄ m⁻² d⁻¹ (summarized by Smith et al. 2000a, Butterbach-Bahl et al. 2002). The average annual rate of net CH₄ consumption in control plots (192 mg m⁻²) was markedly similar to the average (187 mg m⁻²) for a previous study (Phillips et al. 2001a). As with our average

day rate estimates of net CH₄ consumption, these annual values are at the low end of estimates for North American temperate forests, which largely fall between 320 and 2560 mg m⁻² y⁻¹, but have a strong New England bias (Smith et al. 2000b). Gas diffusivity has been demonstrated (Dörr et al. 1993) to control rates of CH₄ supply to the usual subsurface locus of CH₄ oxidation (e.g. Whalen et al. 1992), which is itself substrate-limited in well-drained forest soils, based on kinetic considerations (Bradford et al. 2001). Soil texture influences diffusivity (Ball et al. 1997), with clay soils showing net CH₄ consumption rates an order of magnitude lower than sandy soils (Dörr et al. 1993). It is likely that the fine texture of soil at our study site limits transport of atmospheric CH₄ down-profile, resulting in comparatively low area-based rates of net CH₄ consumption.

Net CH₄ consumption showed no relationship with soil temperature when the entire data over the temperature range 4 to 25 °C were considered, in agreement with the general lack of seasonality in flux (Fig. 2.1). However, this is at odds with the previous report of a significant, but weak temperature effect on net CH₄ consumption at this site (Phillips et al. 2001a). Other studies have frequently shown no or low influence of temperature on atmospheric CH₄ consumption in forest soils (e.g. Borken and Brumme 1997, Butterbach-Bahl and Papen 2002), an observation consistent with the dominance of diffusion limitation (substrate supply) over enzymatic limitation of methanotrophy that can be expected at typical atmospheric CH₄ concentrations (King and Adamsen 1992). However, some north temperate forest soils show an increased influence of temperature on atmospheric CH₄ consumption at values < 10 °C (Crill 1991, Castro et al. 1995, Steinkamp et al. 2001). Examination of our data with respect to this threshold extends

the observations of the influence of low temperatures on CH₄ consumption southward. The average net CH₄ consumption rate of 0.39 mg CH₄ m⁻² d⁻¹ for the 12 of 68 sampling dates at soil temperatures < 10 °C was lower by 32% than the mean of 0.58 mg CH₄ m⁻² d⁻¹ at higher temperatures. Differences in the strength of the temperature-CH₄ flux relationship at low temperatures between the past (Phillips et al. 2001a) and present investigations may have accounted for the disparity in the observed relationship between these variables when the entire data from each study were considered.

In contrast to temperature, we observed a strong (inverse) linear relationship between net CH₄ consumption and soil moisture (Fig. 2.2), which explained 34% of the variability in the entire data, and proved significant in the mixed-effects model for the 2006 data when moisture probes were sited in proximity to soil collars. This confirms previous observations of reduced net CH₄ consumption with increasing soil moisture at this site (Phillips et al. 2001a, McLain et al. 2002) and is consistent with other *in situ* seasonal studies in forest soils. However, our site is apparently less sensitive than many others to changes in soil moisture, as this factor explained 59 to 88% of the variability in net CH₄ consumption across a range of forest ecosystem types (Castro et al. 1994, Lessard et al. 1994, Steinkamp et al. 2001, Butterbach-Bahl and Papen 2002, Price et al. 2004).

Differences in net CH₄ consumption between CO₂ treatments

We found that CO₂ enrichment resulted in a per annum decline in net atmospheric CH₄ consumption of 8 to 19% relative to unamended controls, in accord with previous reports for this site (Phillips et al. 2001a, McLain et al. 2002). Moreover, there is no

compelling evidence for a temporal decline in the magnitude of the reduction in net CH₄ consumption in CO₂-enriched plots compared with controls when our entire data are considered (Table 1). Investigations of soil-atmosphere CH₄ exchange in CO₂-enriched ecosystems that normally function as an atmospheric CH₄ sink are few. In the most directly comparable study to our own, Ambus and Robertson (1999) reported a 22% reduction in CH₄ consumption by soils in model *Populus tremuloides* ecosystems exposed to elevated CO₂. Ineson et al. (1998) observed that rates of net atmospheric CH₄ uptake were three times greater in ambient CO₂ soils relative to CO₂-enriched plots in an N-fertilized sward of *Lolium perenne*. However, a subsequent investigation (Baggs and Blum 2004) found a significant interaction between N fertilizer application rate and CO₂ on net atmospheric CH₄ consumption. Mosier et al. (2002) saw no impact of CO₂ level on rates of CH₄ exchange between soils and the atmosphere in a semi-arid, mixed grassland community.

This study adds to previous efforts (Phillips et al. 2001a, Whalen unpublished) to uniquely provide a nearly continuous 8 y record of reduced atmospheric CH₄ consumption under elevated CO₂ at the same permanently installed soil collars in a representative southern forest. Short- and long-term responses to elevated atmospheric CO₂ must be distinguished. For instance, down-regulation of photosynthesis has been commonly reported for CO₂-fertilized model and intact forest ecosystems after as little as two years (reviewed by Amthor 1995). Over longer time trajectories, initial response functions of all ecosystem components from trees to microbes can be expected to adjust physiologically and demographically on different time scales through modification of biogeochemical feedbacks (Korner 2000). The lag of nearly 2 y between the initiation of

CO₂ fumigation and initial sampling (Phillips et al. 2001a), consistently lower annualized rates of net CH₄ consumption in soils from CO₂-enriched plots relative to controls, and lack of strong evidence that the magnitude of the CO₂ enrichment effect has declined with time all suggest that reduced net atmospheric CH₄ consumption is a sustained, equilibrium response of this forest soil to elevated CO₂.

Potential reasons for reduced net CH₄ consumption under elevated CO₂

Although reduced net atmospheric CH₄ consumption is likely a sustained negative feedback by soil to CO₂-enrichment at our study site, causative factors are difficult to identify, as the destructive sampling necessary for process-level investigations is limited to maintain ecosystem integrity. However, the significant moisture x treatment interaction in our mixed effects model indicates that site-wise differences in net CH₄ consumption are at least in part moisture-related. Several demonstrated effects of CO₂-enrichment on above- and below-ground processes within forest ecosystems feed back on soil moisture and by extension soil CH₄ cycling dynamics (Fig. 2.4).

The net soil-atmosphere CH₄ flux represents the balance between methanogenesis and methanotrophy, and changes in soil moisture elicit offsetting responses in these two microbial processes. Increased net primary production under elevated CO₂ (Fig. 2.4, pathway B) at our site (DeLucia et al. 1999, Hamilton et al. 2002, DeLucia, E.H. et al. 2005, Norby et al. 2005, Finzi et al. 2006a) is responsible for a continuous ~17% greater annual increment of litterfall since fumigation (Allen et al. 2000, Lichter et al. 2005, Lichter et al. 2008; with exception of the 2 y following a 2002 ice storm where litter fall increased regardless of CO₂ treatment). The associated insulating effect enhances

moisture conservation (Schäfer et al. 2003). The direct link between increased soil moisture and diffusion-limitation to CH₄ oxidizers is well established (Striegl 1993, Castro et al. 1995, Whalen and Reeburgh 1996). Persistently higher moisture content and reduced net atmospheric CH₄ consumption in CO₂-enriched plots relative to controls (Table 2) is consistent with a reduction in substrate supply to methanotrophs (Fig. 2.4; pathway B). The excess of litterfall under elevated CO₂ also directly adds to diffusional resistance in soils within these plots. Experimental litter removal has been shown to increase rates on net atmospheric consumption in forest soils by as much as 43% (Dong et al. 1998, Brumme and Borken 1999).

Independent reports of anoxic microzones (Sexstone et al. 1985, Zausig et al. 1993) and methanogenic activity in macroscopically oxygenated soils (Yavitt et al. 1995, Saari et al. 1997, von Fischer and Hedin 2002, Teh et al. 2005) indicate that simultaneous CH₄ production and consumption are occurring in some well-drained upland soils with anoxic aggregates supporting localized zones of methanogenesis and oxic sites supporting methanotrophy. Increased soil moisture under elevated CO₂ likely favored development of additional microsites supporting methanogenesis (Fig. 2.4; pathway B). Previously we found no evidence of methanogenic activity in sieved soils from this site (Phillips et al. 2001b), but a subsequent investigation (McLain and Ahmann 2008) reported CH₄ production in intact soil cores, with stronger activity in soils from CO₂-enriched plots. It was suggested (McLain and Ahmann 2008) that sieving in our earlier study destroyed anaerobic microsites. In the present study, our more frequent observations of net CH₄ emission in CO₂-enriched versus control chambers provide confirmatory evidence for at least episodic CH₄ production under both treatments and

higher rates in CO₂-enriched plots. Increased respiration in CO₂-enriched plots (Bernhardt et al. 2006) may have also directly provided additional substrate for methanogens, as the pathway in anoxic aggregates of forest soils appears to be CO₂ reduction rather than acetate cleavage (Teh et al. 2005).

Other feedbacks to CO₂-enrichment beyond plant-mediated changes in soil moisture may also have impacted CH₄ cycling dynamics in these soils. Hoosbeek et al. (2009) reported an increase in soil macro-aggregation (250–2000 μm) under elevated CO₂ in a temperate *Populus x euramericana* plantation. The soil aggregates contained higher concentrations of C and N, providing loci of microbial activity. Enhanced respiratory O₂ consumption by microbes may increase the incidence of anoxic microsites favorable for methanogenesis (Fig. 2.4, pathway C). Elevated CO₂ also induces increased concentrations of secondary compounds such as phenolic and tannins in plant tissues (Gebauer et al. 1997, Peñuelas and Estiarte 1998, Wetzal and Tuchman 2005b; Fig. 2.4; pathway A) and enhances root exudation of organic acids (Norby et al. 1987). Methanotrophs characteristically localized in upper mineral layers of forest soils (Whalen et al. 1992) show a high sensitivity to phenolics, monoterpenes and bulk organics from the overlying O horizon at environmentally relevant levels (Amaral and Knowles 1997, 1998). Enhanced production of inhibitory chemicals delivered in a larger mass of litterfall and subsequently leached to upper mineral layers could have reduced methanotrophic activity in enriched CO₂ plots.

It is unclear if this sustained reduction in net atmospheric CH₄ consumption can be broadly extrapolated to other forested ecosystems. Atmospheric CO₂ enrichment experiments have demonstrated significant increases in net primary production of forest

vegetation (DeLucia et al. 1999, Hamilton et al. 2002, DeLucia et al. 2005, Norby et al. 2005, Finzi et al. 2006a; Fig. 2.4; pathway B). Any attendant increase in soil moisture could effect a decrease in net atmospheric CH₄ consumption as observed here. A process-based model of atmospheric CH₄ consumption by soils indicates an aggregated forest sink of 24 Tg CH₄ y⁻¹ (Ridgwell et al. 1999). A decline in soil CH₄ consumption of the magnitude observed here (~15%; Table 2.1) across all forest biomes gives a decrease of 3.6 Tg CH₄ y⁻¹, a value that is not inconsequential as it represents 10% of the model estimate (Ridgwell et al. 1999) of 38 Tg CH₄ y⁻¹ for the total soil sink.

Our field study of the relationship between CH₄ flux, CO₂ enrichment and soil moisture suggests that moisture sensitivity of net atmospheric CH₄ results from diffusion limitation to methanotrophs and the availability of anaerobic microsites supporting methanogenic activity, although it yields no insights into the relative importance of these microbial processes or other potential controls on CH₄ exchange at the air-soil interface. Improvement of mechanistic models of global consumption of CH₄ by soils will require field and process-oriented laboratory studies across representative forest biomes and soil types to fully identify and quantify coupling mechanisms of net CH₄ oxidation to CO₂ enrichment and plant metabolism.

References

- Allen, A. S., J. A. Andrews, A. C. Finzi, R. Matamala, D. D. Richter, and W. H. Schlesinger. 2000. Effects of free-air CO₂ enrichment (FACE) on belowground processes in a *Pinus taeda* forest. *Ecological Applications* **10**:437-448.
- Amaral, J. A. and R. Knowles. 1997. Inhibition of methane consumption in forest soils and pure cultures of methanotrophs by aqueous forest soil extracts. *Soil Biology and Biochemistry* **29**:1713-1720.
- Amaral, J. A. and R. Knowles. 1998. Inhibition of methane consumption in forest soils by monoterpines. *Journal of Chemical Ecology* **24**:723-734.
- Ambus, P. and G. P. Robertson. 1999. Fluxes of CH₄ and N₂O in aspen stands grown under ambient and twice-ambient CO₂. *Plant and Soil* **209**:1-8.
- Amthor, J. S. 1995. Terrestrial higher-plant response to increasing atmospheric CO₂ in relation to the global carbon cycle. *Global Change Biology* **1**:243-274.
- Baggs, E. M. and H. Blum. 2004. CH₄ oxidation and emissions of CH₄ and N₂O from *Lolium perenne* swards under elevated atmospheric CO₂. *Soil Biology and Biochemistry* **36**:713-723.
- Ball, B. C., K. E. Dobbie, J. P. Parker, and K. A. Smith. 1997. The influence of gas transport and porosity on methane oxidation in soils. *Journal of Geophysical Research* **102**:23301-23308.
- Bernhardt, E., J. Barber, J. Phippen, L. Taneva, J. Andrews, and W. Schlesinger. 2006. Long-term effects of free air CO₂ enrichment (FACE) on soil respiration. *Biogeochemistry* **77**:91-116.
- Borken, W. and R. Brumme. 1997. Liming practice in temperate forest ecosystems and the effects on CO₂, N₂O and CH₄ fluxes. *Soil Use and Management* **13**:251-257.
- Bradford, M. A., P. Ineson, P. A. Wookey, and H. M. Lappin-Scott. 2001. Role of CH₄ oxidation, production and transport in forest soil CH₄ flux. *Soil Biology and Biochemistry* **33**:1625-1631.
- Brumme, R. and W. Borken. 1999. Site variation in methane oxidation as affected by atmospheric deposition and type of temperate forest ecosystem. *Global Biogeochemical Cycles* **13**:493-501.
- Butterbach-Bahl, K., L. Breuer, R. Gasche, G. Willibald, and H. Papen. 2002. Exchange of trace gases between soils and the atmosphere in Scots pine forest ecosystems of the northeastern German lowlands: 1. Fluxes of N₂O, NO/NO₂ and CH₄ at forest sites with different N-deposition. *Forest Ecology and Management* **167**:123-134.

- Butterbach-Bahl, K. and H. Papen. 2002. Four years continuous record of CH₄-exchange between the atmosphere and untreated and limed soil of a N-saturated spruce and beech forest ecosystem in Germany. *Plant and Soil* **240**:77-90.
- Castro, M. S., J. M. Melillo, P. A. Steudler, and J. W. Chapman. 1994. Soil moisture as a predictor of methane uptake by temperate forest soils. *Canadian Journal of Forest Resources* **24**:1805-1810.
- Castro, M. S., P. A. Steudler, J. M. Melillo, J. D. Aber, and R. D. Bowden. 1995. Factors controlling atmospheric methane consumption by temperate forest soils. *Global Biogeochemical Cycles* **9**:1-10.
- Crill, P. M. 1991. Seasonal patterns of methane uptake and carbon dioxide release by a temperate woodland soil. *Global Biogeochemical Cycles* **5**:319-334.
- DeLucia, E. H., J. G. Hamilton, S. L. Naidu, R. B. Thomas, J. A. Andrews, A. Finzi, M. Lavine, R. Matamala, J. E. Mohan, G. R. Hendrey, and W. H. Schlesinger. 1999. Net Primary Production of a Forest Ecosystem with Experimental CO₂ Enrichment. *Science* **284**:1177-1179.
- DeLucia, E. H., D. J. Moore, and R. J. Norby. 2005. Contrasting responses of forest ecosystems to rising atmospheric CO₂: Implications for the global C cycle. *Global Biogeochemical Cycles* **19**: GB3006.
- Denman, S. E., N. W. Tomkins, and C. S. McSweeney. 2007. Quantitation and diversity analysis of ruminal methanogenic populations in response to the antimethanogenic compound bromochloromethane. *FEMS Microbiology Ecology* **62**:313-322.
- Dong, Y., D. Scharffe, J. M. Lobert, P. J. Crutzen, and E. Sanhueza. 1998. Fluxes of CO₂, CH₄, and N₂O from a temperate forest soil: the effects of leaves and humus layers. *Tellus* **50B**:243-252.
- Dörr, H., L. Katruff, and I. Levin. 1993. Soil texture parameterization of the methane uptake in aerated soils. *Chemosphere* **26**:697-713.
- Finzi, A. C., D. J. P. Moore, E. H. DeLucia, J. Lichter, K. S. Hofmockel, R. B. Jackson, H.-S. Kim, R. Matamala, H. R. McCarthy, R. Oren, J. S. Phippen, and W. H. Schlesinger. 2006. Progressive nitrogen limitation of ecosystem processes under elevated CO₂ in a warm-temperate forest *Ecology* **87**:15-25.
- Forster, P., V. Ramaswamy, P. Artaxo, T. Berntsen, R. Betts, D. W. Fahey, J. Haywood, J. Lean, D. C. Lowe, G. Myhre, J. R. Nganga, R. Prinn, G. Raga, M. Schulz, and R. Van Dorland. 2007. Changes in Atmospheric Constituents and in Radiative Forcing. *in* S. Solomon, D. Qin, M. Manning, J. Chen, M. Marquis, K. B. Averyt, M. Tignor, and A. J. Miller, editors. *Climate Change 2007: The Physical Sciences*

- Basis. Contribution of the Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Gebauer, R. L. E., B. R. Strain, and J. F. Reynolds. 1997. The effect of elevated CO₂ and N availability on tissue concentrations and whole plant pools of carbon-based secondary compounds in loblolly pine (*Pinus taeda*). *Oecologia* **113**:29-36.
- Hamilton, J., E. DeLucia, K. George, S. Naidu, A. Finzi, and W. Schlesinger. 2002. Forest carbon balance under elevated CO₂. *Oecologia* **131**:250-260.
- Hendrey, G. R., D. S. Ellsworth, K. F. Lewin, and J. Nagy. 1999. A free-air enrichment system for exposing tall forest vegetation to elevated atmospheric CO₂. *Global Change Biology* **5**:293-309.
- Hoosbeek, M. and G. Scarascia-Mugnozza. 2009. Increased litter build up and soil organic matter stabilization in a poplar plantation after 6 years of atmospheric CO₂ enrichment (FACE): final results of POP-EuroFACE compared to other forest FACE experiments. *Ecosystems* **12**:220-239.
- Ineson, P., P. A. Coward, and U. A. Hartwig. 1998. Soil gas fluxes of N₂O, CH₄ and CO₂ beneath *Lolium perenne* under elevated CO₂: The Swiss free air carbon dioxide enrichment experiment. *Plant and Soil* **198**:89-95.
- King, G. M. and A. P. S. Adamsen. 1992. Effects of temperature on methane consumption in a forest soil and in pure cultures of the methanotroph *Methylomonas rubra*. *Applied Environmental Microbiology* **58**:2758-2763.
- Korner, C. 2000. Biosphere responses to CO₂ enrichment. *Ecological Applications* **10**:1590-1619.
- Lessard, R., P. Rochette, E. Topp, E. Pattey, R. L. Desjardins, and G. Beaumont. 1994. Methane and carbon dioxide fluxes from poorly drained adjacent cultivated and forest sites. *Canadian Journal of Forest Resources* **74**:139-146.
- Lichter, J., S. H. Barron, C. E. Bevacqua, A. C. Finzi, K. F. Irving, E. M. Stemmler, and W. Schlesinger. 2005. Soil carbon sequestration and turnover in a pine forest after six years of atmospheric CO₂ enrichment. *Ecology* **86**:1835-1847.
- Lichter, J., S. A. Billings, S. E. Ziegler, D. Indh, R. Ryals, A. C. Finzi, R. B. Jackson, E. M. Stemmler, and W. H. Schlesinger. 2008. Soil carbon sequestration in a pine forest after 9 years of atmospheric CO₂ enrichment. *Global Change Biology* **14**:1-13.

- McLain, J. and D. Ahmann. 2008. Increased moisture and methanogenesis contribute to reduced methane oxidation in elevated CO₂ soils. *Biology and Fertility of Soils* **44**:623-631.
- McLain, J. E. T., T. B. Keppeler, and D. M. Ahnman. 2002. Belowground factors mediating changes in methane consumption in a forest soil under elevated CO₂. *Global Biogeochemical Cycles* **16**:1050-1064.
- Mosier, A. R., J. A. Morgan, J. Y. King, D. LeCain, and D. G. Milchunas. 2002. Soil-atmosphere exchange of CH₄, CO₂, NO_x, and N₂O in the Colorado shortgrass steppe under elevated CO₂. *Plant and Soil* **240**:201-211.
- Newton, P. C. D., H. Clark, G. R. Edwards, and D. J. Ross. 2001. Experimental confirmation of ecosystem model predictions comparing transient and equilibrium plant responses to elevated atmospheric CO₂. *Ecology Letters* **4**:344-347.
- Norby, R. J., E. H. DeLucia, B. Gielen, C. Calfapietra, C. P. Giardina, J. S. King, J. Ledford, H. R. McCarthy, D. J. P. Moore, R. Ceulemans, P. De Angelis, A. C. Finzi, D. F. Karnosky, M. E. Kubiske, M. Lukac, K. S. Pregitzer, G. E. Scarascia-Mugnozza, W. H. Schlesinger, and R. Oren. 2005. Forest response to elevated CO₂ is conserved across a broad range of productivity. *Proceedings of the National Academy of Sciences of the United States of America* **102**:18052-18056.
- Norby, R. J., E. G. O'Neill, W. G. Hood, and R. J. Luxmoore. 1987. Carbon allocation, root exudation, and mycorrhizal colonization of *Pinus echinata* seedlings grown under CO₂ enrichment. *Tree Physiology* **3**:203-210.
- Oh, N. H. and D. D. Richter. 2005. Elemental translocation and loss from three highly weathered soil-bedrock profiles in the southeastern United States. *Geoderma* **126**:5-25.
- Peñuelas, J. and M. Estiarte. 1998. Can elevated CO₂ affect secondary metabolism and ecosystem function? *Trees* **13**:20-24.
- Phillips, R. L., S. C. Whalen, and W. H. Schlesinger. 2001a. Influence of atmospheric CO₂ enrichment on methane consumption in a temperate forest soil. *Global Change Biology* **7**:557-563.
- Phillips, R. L., S. C. Whalen, and W. H. Schlesinger. 2001b. Response of soil methanotrophic activity to carbon dioxide enrichment in a North Carolina coniferous forest. *Soil Biology and Biochemistry* **33**:793-800.
- Price, S. J., R. R. Sherlock, F. M. Kelliher, T. M. McSeveny, K. R. Tate, and L. M. Condron. 2004. Pristine New Zealand forest soil is a strong methane sink. *Global Change Biology* **10**:16-26.

- Ridgwell, A. J., S. J. Marshall, and K. Gregson. 1999. Consumption of atmospheric methane by soils: a process-based model. *Global Biogeochemical Cycles* **13**:59-70.
- Saari, A., P. J. Martikainen, A. Ferm, J. Ruuskanen, W. De Boer, S. R. Troelstra, and H. J. Laanbroek. 1997. Methane oxidation in soil profiles of Dutch and Finnish coniferous forests with different soil texture and atmospheric nitrogen deposition. *Soil Biology and Biochemistry* **29**:1625-1632.
- Schäfer, K. V. R., R. Oren, D. S. Ellsworth, C.-T. Lai, J. D. Herrick, A. C. Finzi, D. D. Richter, and G. G. Katul. 2003. Exposure to an enriched CO₂ atmosphere alters carbon assimilation and allocation in a pine forest ecosystem. *Global Change Biology* **9**:1378-1400.
- Schlesinger, W. H. and J. Lichter. 2001. Limited carbon storage in soil and litter of experimental forest plots under increased atmospheric CO₂. *Nature* **411**:466-469.
- Sexstone, A. J., N. P. Revsbech, T. B. Parkin, and J. M. Tiedje. 1985. Direct measurement of oxygen profiles and denitrification rates in soil aggregates. *Soil Science Society of America Journal* **49**:645-651.
- Smith, K. A., K. E. Dobbie, B. C. Ball, L. R. Bakken, B. K. Sitaula, S. Hansen, R. Brumme, W. Borken, S. Christensen, A. Priem, D. Fowler, J. A. Macdonald, U. Skiba, L. Klemetsson, A. Kasimir-Klemetsson, A. DegÅrska, and P. Orlanski. 2000b. Oxidation of atmospheric methane in Northern European soils, comparison with other ecosystems, and uncertainties in the global terrestrial sink. *Global Change Biology* **6**:791-803.
- Smith, P., D. S. Powlson, J. U. Smith, P. Falloon, and K. Coleman. 2000a. Meeting Europe's climate change commitments: quantitative estimates of the potential for carbon mitigation by agriculture. *Global Change Biology* **6**:525-539.
- State Climate Office of North Carolina. 2003-2009. NC Climate Retrieval and Observations Network of the Southeast Database (NCCRONOS).
- Steinkamp, R., K. Butterbach-Bahl, and H. Papen. 2001. Methane oxidation by soils of an N limited and N fertilized spruce forest in the Black Forest, Germany. *Soil Biology and Biochemistry* **33**:145-153.
- Striegl, R. G. 1993. Diffusional limits to the consumption of atmospheric methane by soils. *Chemosphere* **26**:715-720.
- Suwa, M., G. G. Katul, R. Oren, J. Andrews, J. Pippen, A. Mace, and W. H. Schlesinger. 2004. Impact of elevated atmospheric CO₂ on forest floor respiration in a temperate pine forest. *Global Biogeochemical Cycles* **18**: GB2013.

- Teh, Y. A., W. L. Silver, and R. Conrad. 2005. Oxygen effects on methane production and oxidation in humid tropical forest soils. *Global Change Biology* **11**:1283-1297.
- von Fischer, J. C. and L. O. Hedin. 2002. Separating methane production and consumption with a field-based isotope pool dilution technique. *Global Biogeochemical Cycles* **16**:1034.
- Wetzel, R. G. and N. C. Tuchman. 2005. Effects of elevated CO₂ on the lignin and total phenolic concentrations of cattail and trembling aspen leaves. *Archiv für Hydrobiologie* **162**:287-308.
- Whalen, S. C. and W. S. Reeburgh. 1996. Moisture and temperature sensitivity of CH₄ oxidation in boreal soils. *Soil Biology and Biochemistry* **28**:1271-1281.
- Whalen, S. C., W. S. Reeburgh, and V. A. Barber. 1992. Oxidation of methane in boreal forest soils: a comparison of seven measures. *Biogeochemistry* **16**:181-211.
- Wuebbles, D. J. and K. Hayhoe. 2002. Atmospheric methane and global change. *Earth-Science Reviews* **57**:177-210.
- Yavitt, J. B., T. J. Fahey, and J. A. Simmons. 1995. Methane and carbon dioxide dynamics in a northern hardwood ecosystem. *Soil Science Society of America Journal* **59**:796-804.
- Zausig, J., W. Stepniwski, and R. Horn. 1993. Oxygen concentration and redox potential gradients in unsaturated model soil aggregates. *Soil Science Society of America Journal* **57**:908-916.

Table 2.1. Annual time-integrated rates of net CH₄ consumption and volumetric soil moisture for six nearly consecutive years in forest plots exposed to ambient or elevated levels of CO₂. Annual time-integrated rates of net CH₄ consumption were determined by integrating for the calendar year daily, area-based data from each sampling occasion.

Year	Annual net CH ₄ consumption (mg m ⁻² y ⁻¹)		% Difference between treatments	Source	Volumetric soil moisture (mL H ₂ O g soil ⁻¹)		Source
	Control	CO ₂ -enriched			Control	CO ₂ -enriched	
1998	183	156	16 *	(Phillips et al. 2001a)	0.24	0.26 *	(Schäfer et al. 2003)
1999	191	136	30 *	(Phillips et al. 2001a)	0.27	0.34 *	(Schäfer et al. 2003)
2002	204	181	13 *	(Whalen unpubl.)	0.22	0.23 *	(Hyun unpubl.)
2004	184	150	19 *	Present study	0.29	0.31 *	(Hyun unpubl.)
2005	196	175	10 *	Present study	0.26	0.30 *	(Hyun unpubl.)
2006	197	181	8	Present study	0.23	0.24	Present study

*Differences between treatments significant at $\alpha=0.05$.

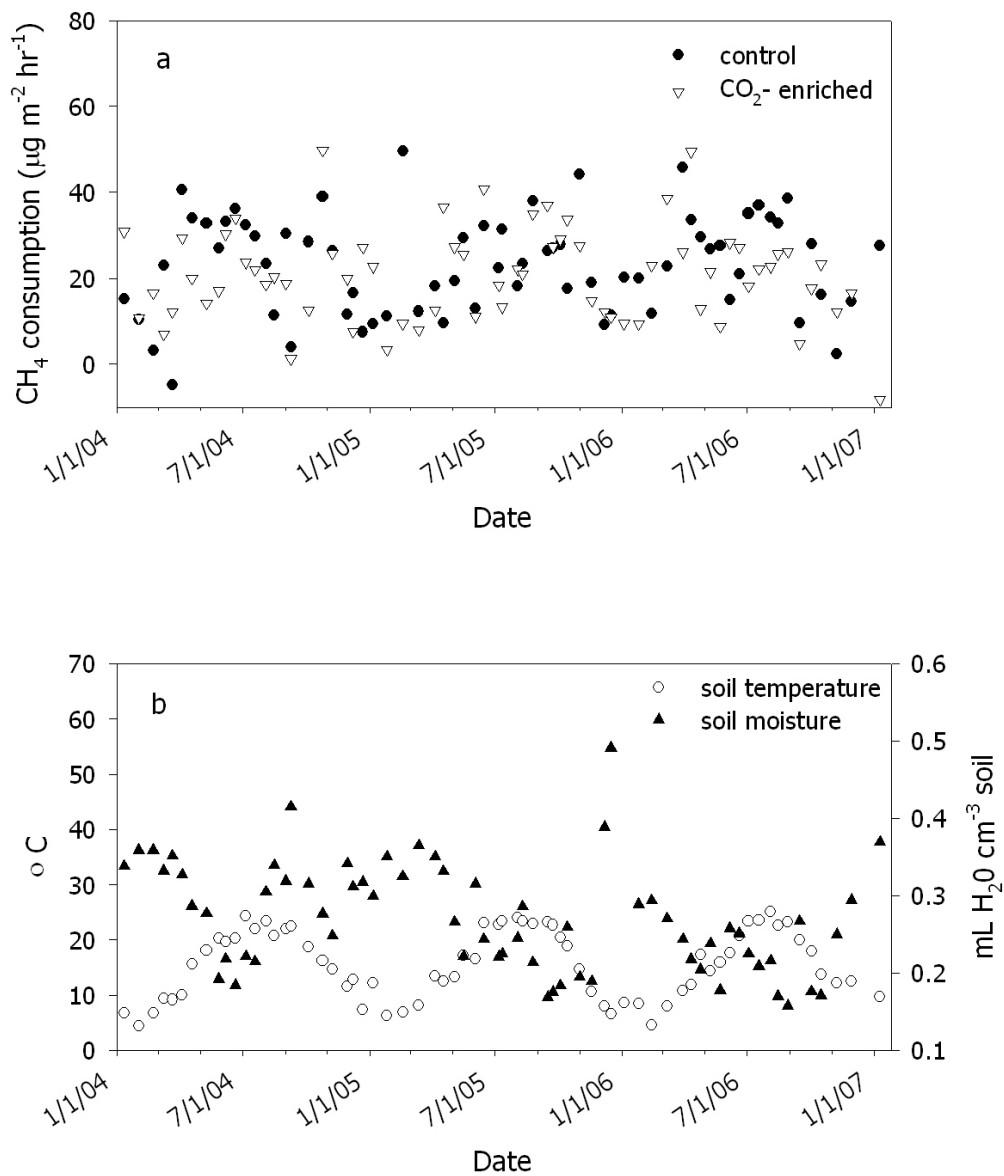


Figure 2.1. (a) Time series for rates of net atmospheric CH₄ consumption by forest soils under CO₂-enriched and ambient atmospheres (control). Each datum point represents the mean of 12 or 16 individual static chamber flux determinations for each treatment; (b) Time series for changes in mean soil temperature (1 to 19 cm depth interval) and mean volumetric soil moisture to 30 cm ($\text{mL H}_2\text{O cm}^{-3} \text{ soil}$). In all cases error bars are eliminated for clarity.

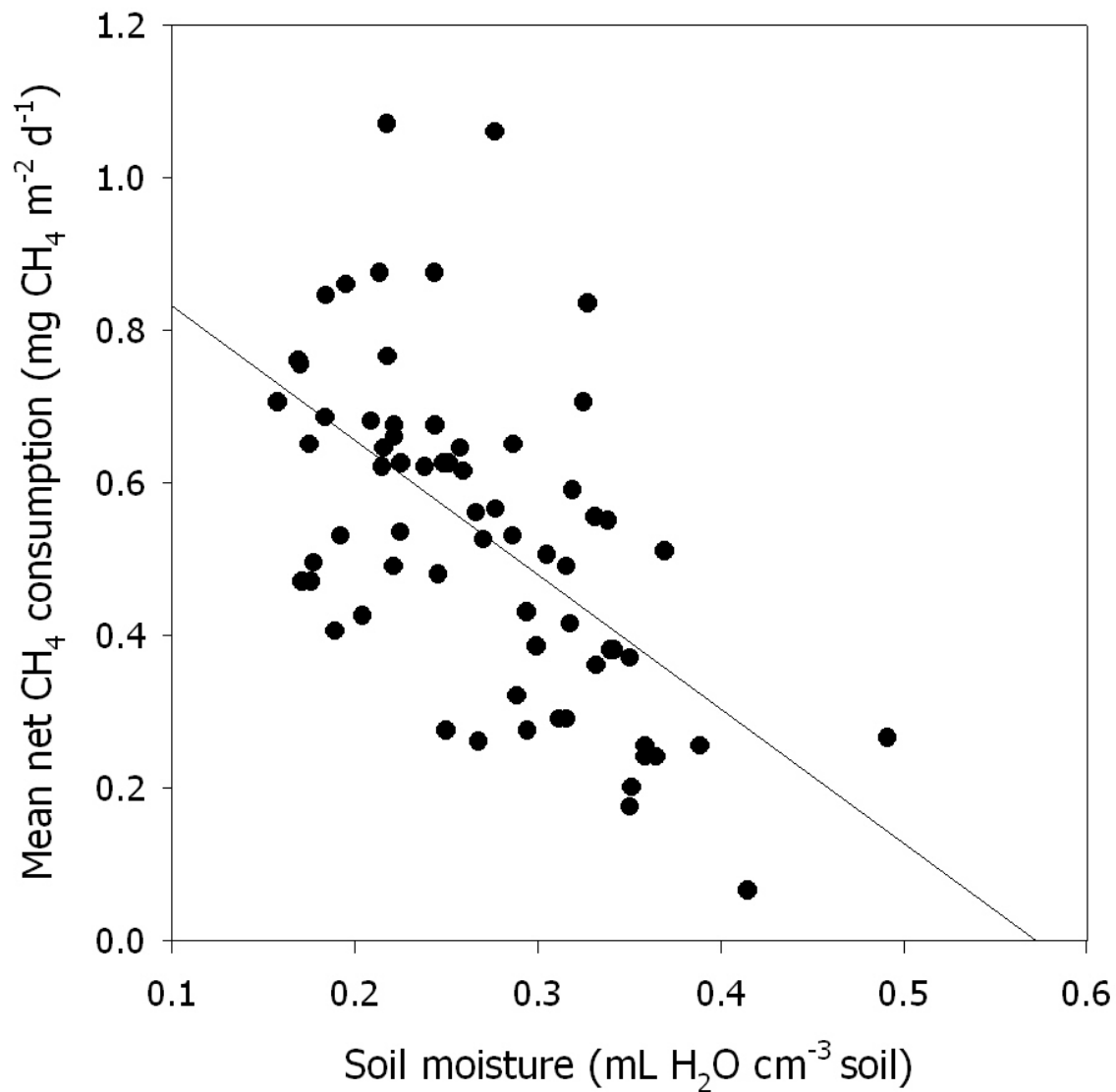


Figure 2.2. Relationship between net atmospheric CH₄ consumption and volumetric soil moisture to 30 cm depth for the entire study ($r^2 = 0.340$). Each datum point represents the mean of 24 (2004 and 2005) or 32 (2006) observations for a sampling date.

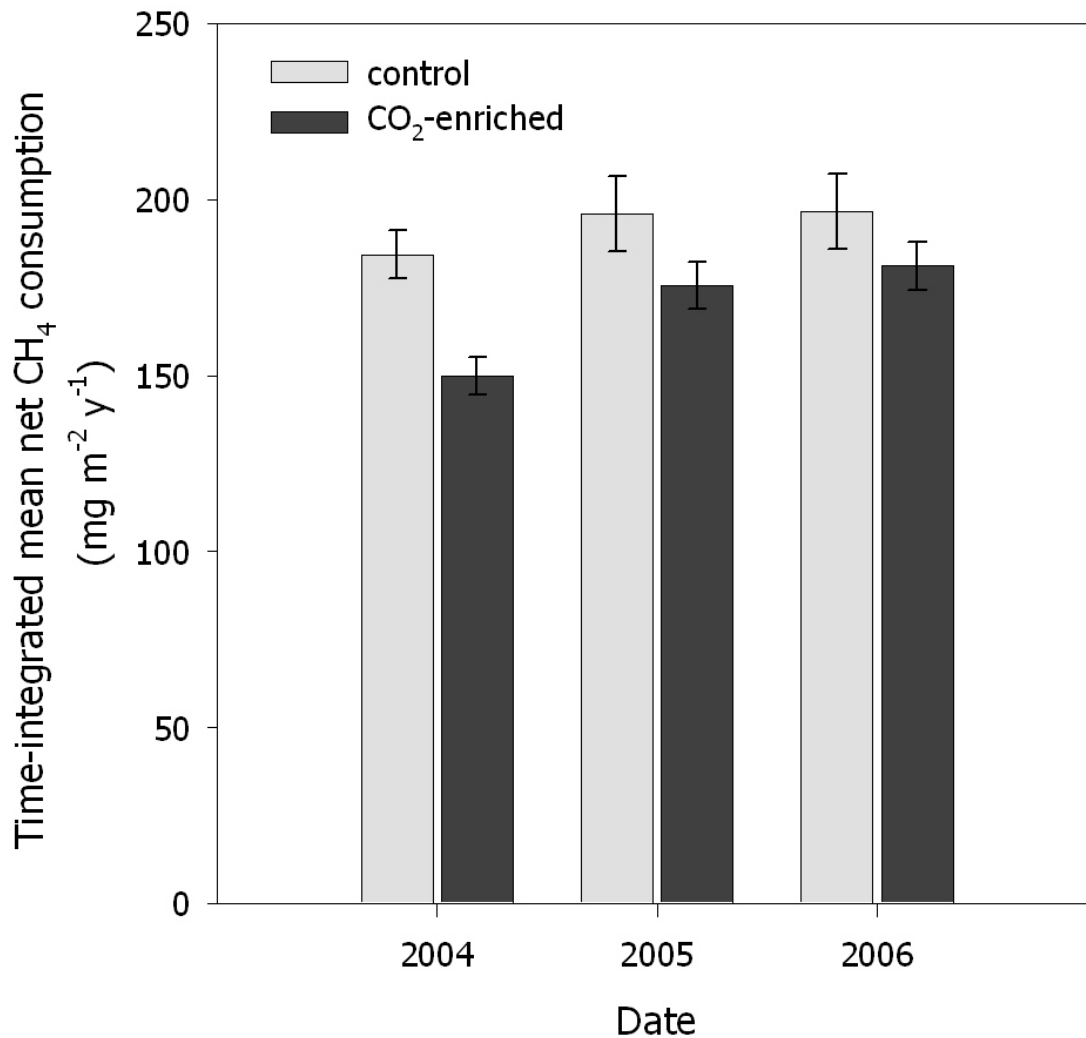
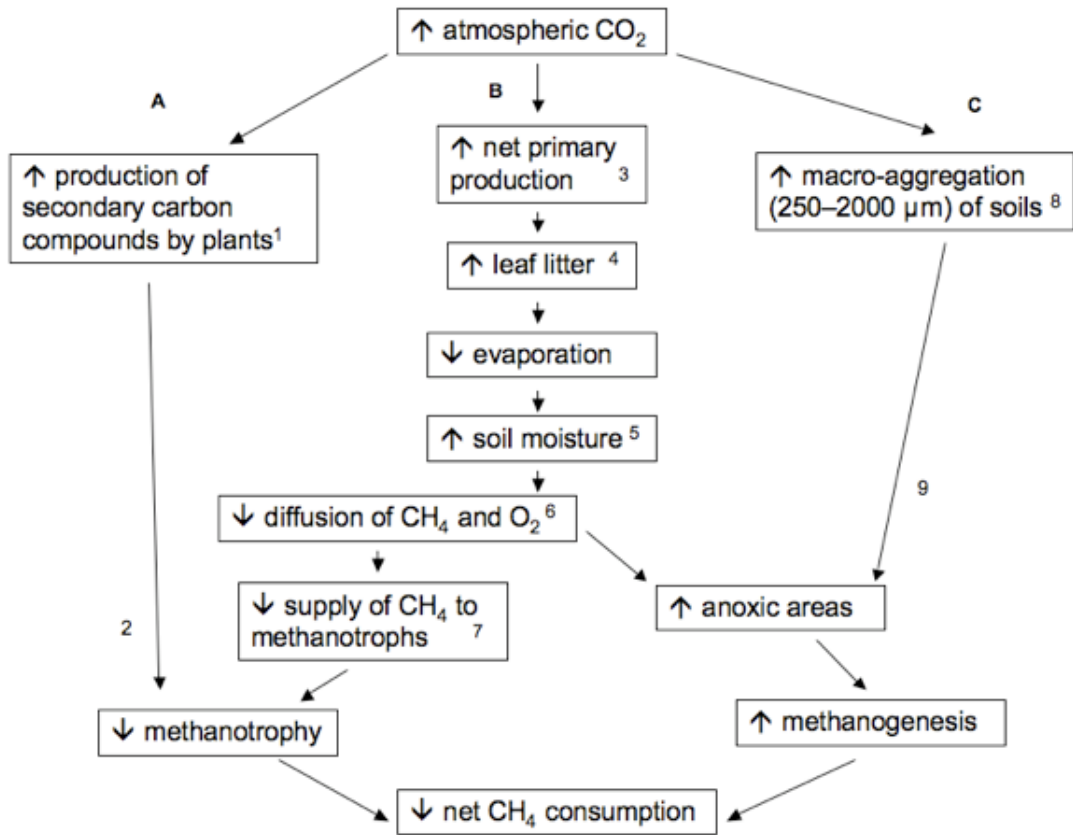


Figure 2.3. Annual time-integrated net CH₄ consumption by temperate forest soils at the Duke FACE site under ambient (control) and elevated (CO₂-enriched) concentrations of CO₂ for 2004 through 2006. Annual time-integrated rates of net CH₄ consumption were determined by integrating for the calendar year daily, area-based data from each sampling occasion (n = 23). The differences between CO₂ treatments were 19%, 10% and 8% for 2004, 2005, and 2006, respectively. Differences were significant for 2004 and 2005. Error bars represent one standard error of the mean.



¹ Gebauer et al. 1997; Peñuelas and Estiarte 1998; Wetzel and Tuchman 2005

² Amaral and Knowles 1997, 1998

³ DeLucia et al. 1999; Hamilton et al. 2002; DeLucia et al. 2005; Norby et al. 2005; Finzi et al. 2006

⁴ Allen et al. 2000; Schlesinger and Lichter 2001

⁵ Schäfer et al. 2002

⁶ Suwa et al. 2004

⁷ Striegl 1993; Castro et al. 1995; Whalen and Reeburgh 1996

⁸ Hoosbeek and Scarascia-Mugnozza 2009

⁹ Sextone et al. 1985; Zausig et al. 1993

Figure 2.4. Conceptual model of the impact of forest ecosystem responses to elevated CO₂ that influence soil CH₄ cycling dynamics. Up and down arrows within each response function indicate a positive or negative impact, respectively, of that factor on net atmospheric CH₄ consumption. Response functions are either documented or hypothesized by the associated references.

**CHAPTER 3: INHIBITION OF CH₄ CONSUMPTION BY SECONDARY
CARBON COMPOUNDS IN THE TISSUES AND EXUDATES OF TEMPERATE
FOREST PLANTS EXPOSED TO ELEVATED CO₂**

Abstract

We previously showed a sustained reduction in net atmospheric CH₄ consumption by temperate forest soils in response to elevated CO₂ (Dubbs and Whalen submitted) and here report the influence of plant exudates on atmospheric CH₄ consumption in soils from the same study site. We examine the effect of root exudate acids and primary or secondary metabolites from plant exudates (throughfall, duff and leaf leachates) on CH₄ consumption. Plant exudates from forest plots exposed to elevated CO₂ since 1996 (~580 mL L⁻¹ CO₂) or from control plots subjected to ambient conditions and acid root exudates from loblolly pines (*Pinus taeda*) grown under elevated CO₂ were applied to soils. Duff leachates occasionally inhibited CH₄ consumption regardless of CO₂ treatment, and levulinic acid inhibited CH₄ consumption at a concentration of 100 μmol L⁻¹, but not at 50 μmol L⁻¹. All other tested exudates had no effect on rates of CH₄ consumption. While plant exudates may only assert transient and secondary control on CH₄ consumption under elevated CO₂, identification of temporal and spatial patterns of influence warrant further study because they otherwise confound the correlation between the primary drivers of CH₄ consumption and measured rates of net CH₄ consumption.

Introduction

The global atmospheric CO₂ concentration has more than doubled since the Industrial Revolution (Forster et al. 2007). Little is understood about how increasing atmospheric CO₂ will affect the biogeochemical cycling of other greenhouse gases, including CH₄, but we recently reported a sustained decrease in net atmospheric CH₄ consumption under elevated CO₂ in a temperate loblolly pine (*Pinus taeda*) forest (Dubbs and Whalen submitted). Our previous work showed that a sustained CO₂-induced negative feedback on forest soil CH₄ consumption could lead to a 15% reduction (3.6 Tg CH₄ yr⁻¹) in the current forest soil sink of 24 Tg yr⁻¹ (Ridgwell et al. 1999). This negative feedback to increasing CO₂ is of concern because consumption by upland soils is the only terrestrial sink for atmospheric CH₄, which is a greenhouse gas with radiative forcing second only to CO₂ among trace atmospheric gases (Forster et al. 2007).

The reasons for the observed decline in net atmospheric CH₄ consumption by these soils under elevated CO₂ were not entirely clear. However, we postulated several pathways by which changes in a temperate forest ecosystem exposed to elevated CO₂ could lead to decreased CH₄ consumption by methanotrophic bacteria and increased CH₄ production by methanogenesis (Fig. 3.1). Rates of CH₄ exchange between upland soils and the atmosphere are dependent upon the balance between methanotrophy in largely oxic soils and methanogenesis in anoxic microzones. The resultant net CH₄ consumption accounts for the observed sink strength of upland soils in the global CH₄ budget (Forster et al. 2007).

Pathway A (Fig. 3.1) depicts a mechanism by which changes in the chemistry of forest plant tissues may contribute to the observed reduction in net CH₄ consumption.

Ecosystem-scale elevated CO₂ enrichment experiments, or free air carbon exchange (FACE) experiments, indicate that temperate forest responses to elevated CO₂ include increased net primary production (DeLucia et al. 1999, Hamilton et al. 2002, DeLucia et al. 2005, Norby et al. 2005, Finzi et al. 2006a), litter fall (Allen et al. 2000, Lichter et al. 2005, Liu et al. 2005, Lichter et al. 2008), and fine-root production (Norby et al. 2004). Changes in plant productivity, tissue chemistry, C allocation and plant-microbe interactions under elevated CO₂ in turn impact the quantity and quality of C in the ecosystem. For example, Lichter et al. (2000a) observed an increase in labile dissolved organic C in throughfall, and Matamala and Schlesinger (2000) observed a 5.6% increase in the storage of C in forest soils under elevated CO₂. Norby et al. (1987) found that elevated CO₂ enhances root exudation of organic acids in a pot study of *Pinus echinata* seedlings. Elevated CO₂-induced changes in plants also increase the abundance of secondary C compounds in tissues and root exudates relative to plants exposed to ambient CO₂ (Peñuelas and Estiarte 1998, Verburg et al. 1999, Tuchman et al. 2002, Billings and Ziegler 2005, Wetzler and Tuchman 2005a). Secondary C compounds, such as phenolics and terpenes, inhibit metabolism and growth by broad groups of soil bacteria (Souto et al. 2000), and specifically, methanotrophs (Amaral and Knowles 1997, 1998).

Here we extend previous research, which indicated that reduced net CH₄ consumption by a temperate forest soil is a sustained response to elevated CO₂ (Dubbs and Whalen submitted), by examining the influence of plant exudates on rates of CH₄ consumption in soils from the same study site (Fig. 3.1, Pathway A). We investigate the effect of organic acids, found to be the most abundant organic component of photosynthates released from roots in the rhizosphere (Smith 1976), and the effects of

primary or secondary metabolites from exudates (throughfall, duff and leaf leachates) on CH₄ consumption. Plant exudates were collected from forest plots exposed to elevated CO₂ since 1996 or from plots subjected to ambient conditions (~380 mL L⁻¹ CO₂; control), and applied to soils. Additionally, organic acids identified to be major components of root exudates (Phillips and Bernhardt unpublished) from loblolly pine laboratory-grown under elevated CO₂ were applied to soils individually and in mixed cocktails to evaluate the effects of these root exudate acids on CH₄ consumption. Effects of plant exudates on rates of CH₄ production were also determined and deemed negligible, and thus, are not discussed further in this manuscript.

Methods

Field site

Field measurements were conducted at the Duke Forest (North Carolina; USA) Free-Air CO₂ Enrichment (FACE) experiment sited in an even-aged stand of loblolly pine (*Pinus taeda* L.) planted in 1983. Soils are clay loam, Ultic Hapludalf's of the Enon Series (Oh and Richter 2005). Average normal air temperature ranges from 3.6 ° C in January to 25.3 ° C in July and annual precipitation averages 1209 mm (State Climate Office of North State Climate Office of North Carolina 2003-2009).

Site characteristics are fully documented in Hendrey et al. (1999a) and briefly described here. The experiment consists of eight circular 30-m diameter plots. Four treatment plots (referred to as “CO₂-enriched”) are fumigated with CO₂ to maintain atmospheric CO₂ concentrations 200 mL L⁻¹ above ambient levels, while three additional treatment plots are fumigated with ambient air to replicate micrometeorological effects

associated with CO₂ addition. A fourth is subjected to ambient air without fumigation. The latter four plots are referred to as “controls”. Continuous (24 h d⁻¹) fumigation was initiated in August 1996, but was reduced to daylight hours only from 2003 to present.

Plant exudate collection

Throughfall

Throughfall collectors consisted of 4 L amber acid-washed glass bottles. The necks of the bottles were plugged by rubber stoppers, which were penetrated by acid-washed glass funnels (60° angle bowl and 100 mm stem). The funnel stems were stuffed with Pyrex glass wool to exclude large particles. One throughfall collector was randomly placed within each of the experimental plots (n=4 for each CO₂-enriched and control treatments) within 48 h of a predicted precipitation event in June and November of 2004, and June of 2005. Throughfall samples were transferred to amber HDPE wide-mouth bottles within 6 h of the conclusion of each discrete rainfall, and the contents were frozen at -10 °C. Samples were thawed and applied to soils within 10 d of collection.

Fresh leaf litter and duff collection and leaching

Approximately 5 g (wet weight) of freshly fallen *Acer rubrum* (red maple), *Liquidambar styraciflua* (sweetgum), *Pinus taeda* (loblolly pine), and *Ulmus alata* (winged elm) leaves were collected from each CO₂-enriched and control plot (n=3 for each treatment) in June and October of 2005. These species were chosen because there was at least one individual tree of each of these species in each experimental plot. Freshly fallen leaves were identified as green leaves lying on the boardwalks that divide

six of the eight experimental plots into quadrants. Upon returning to the laboratory (~1 h after collection), wet mass was determined and leaves were placed in acid-washed 30 mL glass vials. Leaves (by species) were submerged in 20 mL deionized water (DIW) while a vial filled with DIW served as a control throughout the leaching process and soil incubation. Vials were covered by parafilm and leaves were allowed to leach in the dark at ~24 °C for 24 h (Mann and Wetzel 1996).

Duff was randomly collected from the forest floor of CO₂-enriched and control plots (n=4 for each treatment) in October of 2006, and October, November, and December of 2007. Duff is identified as partially decaying plant material on the forest floor surface. Upon returning to the laboratory (~1 h after collection), 10 g of duff (wet mass) from each plot was placed in a 118 mL acid-washed glass jar. Duff was submerged in DIW (60 mL), and one jar without duff was filled with 60 mL of DIW, to serve as a control throughout the leaching process and soil incubation. Jars were covered by parafilm, and duff was allowed to leach in the dark at ~24 °C for 24 h (Mann and Wetzel 1996).

Root exudate preparation

Several organic acids were identified as primary root exudates from loblolly pine trees grown under elevated CO₂ in a glass bead rooting substitute by the Bernhardt lab at Duke University. The primary root acid exudates that were identified included citric, malic, oxalic, maleic, fumaric, levulinic, succinic, shikimic, and protocatechuic acids. Solutions of individual acids and a cocktail of all acids were prepared at 100 μmol L⁻¹ concentrations in DIW used for incubation experiments in April and October of 2006.

Selected organic acid exudates were additionally prepared at $10 \mu\text{mol L}^{-1}$, $50 \mu\text{mol L}^{-1}$, $100 \mu\text{mol L}^{-1}$, $500 \mu\text{mol L}^{-1}$, and $1000 \mu\text{mol L}^{-1}$ and used in a companion experiment intended to identify a threshold concentration for inhibition of CH_4 consumption.

Soil assays and incubation

Soils from the 0 to 20 cm depth interval were collected at the research site from outside of the experimental plots with a hand trowel 1 d prior to initiation of experimentation. Soils were collected from outside of experimental plots because destructive sampling within experimental plots is highly restricted. Upon return to the lab, soils were immediately homogenized by sieving (4.75 mm mesh), and 10 g subsamples of field moist soil were placed into 120 mL glass serum bottles. Twice the number of soil-filled bottles were prepared as were needed ($n=3$ for each treatment in leaf leachate and root exudates experiments, $n=4$ in throughfall and duff leachate experiments). Bottles were allowed to equilibrate with laboratory air ($\sim 1.8 \text{ mL L}^{-1} \text{ CH}_4$) for 1 h before being capped with butyl rubber stoppers and crimp-sealed. Headspace CH_4 concentrations were determined immediately upon sealing and 12 h later by removing 3 mL of headspace gas with 5 mL plastic syringes. The jars in which CH_4 was consumed at the most similar rates were used for further experimentation.

Throughfall, leaf and duff leachates, and the organic acids identified to be primary root exudates were administered to chosen soil aliquots. The amount of liquid added to soils depended upon extant soil moisture as liquid additions were intended to achieve a water holding capacity of approximately 50%. An equal volume of DIW was added to soil samples as a control in all experiments. Following liquid addition, soils were allowed to

equilibrate at the lab atmosphere for 12 to 15 h before jars were sealed and sampled as described above, except that at least 5 samples were collected at evenly-spaced intervals over 48 h incubation periods. Headspace pressure was maintained at 1 atm by replacing removed headspace gas with an equivalent volume of ultrapure N₂.

Gas samples were analyzed for CH₄ by flame ionization detection gas chromatography (Shimadzu model GC 8 A; precision expressed as the coefficient of variation for 10 replicate injections of a 0.94 mL CH₄ L⁻¹ standard was < 3%) within 10 h of collection, well within our predetermined holding time of 24 h. Sample separation was accomplished on a 1-m length x 0.32 cm diameter molecular sieve 5A column with an ultrahigh purity N₂ carrier gas (33 mL min⁻¹). Injector and detector temperatures were set at 90 °C and 140 °C. Headspace measurements from replicate bottles without soil ensured that changes in headspace CH₄ concentrations did not result from gas exchange with butyl rubber stoppers.

Statistical Analysis

Soil dry mass-based rates of CH₄ consumption were calculated from the log-linear change of CH₄ concentration in jar headspaces. Rates or rate constants for CH₄ consumption were compared by paired t-tests. A significance level of $\alpha=0.05$ was used for all statistical comparisons.

Results

Throughfall and leaf leachates

Rate constants (k ; d^{-1}) for CH_4 consumption by soils following application of throughfall or leaf leachates from several individual species of trees collected from control or CO_2 -enriched plots were not significantly different on any of the three dates tested (Fig. 3.2). Similarly, values of k for CH_4 consumption by soils to which plant exudates were added also were not significantly different from those of soils to which DIW was applied on either of the two dates tested (Fig. 3.3).

Duff leachates

Application of duff leachates from either CO_2 -enriched or control plots in fall of 2006 significantly reduced rates of CH_4 consumption, by 34% (CO_2 -enriched) and 38% (control), relative to rates of CH_4 consumption by soils to which DIW was added (Fig. 3.4). The rates of CH_4 consumption for CO_2 -enriched and control plot treatments were not significantly different from each other. This pattern of reduced CH_4 consumption by soils to which duff leachate from both CO_2 -enriched and control treatment plots was added was not repeatable, however, in similar experiments conducted three times in the fall and winter of 2007 (Fig. 3.5). In all cases, rates of CH_4 consumption were not significantly different for any treatment in soils amended with DIW or duff leachate from control or CO_2 -enriched plots.

Organic acids from root exudates

In general, rates of CH_4 consumption in soils following the addition of individual organic acids or a cocktail of organic acids identified as primary components of root

exudates from loblolly pine trees grown under elevated CO₂ were not significantly different from that of soils to which DIW was added (Fig. 3.6). Levulinic acid was the exception as it significantly inhibited rates of CH₄ consumption. A 100 μmol L⁻¹ solution of levulinic acid reduced CH₄ consumption in soils by 63% and 91% relative to DIW-treated soils in October and April 2006 trials, respectively. An experiment conducted to identify the concentration threshold for inhibition of CH₄ consumption by levulinic acid revealed that CH₄ consumption was not significantly reduced at concentrations below 100 μmol L⁻¹ (Fig. 3.7). Rates of CH₄ consumption were, however, significantly reduced at levels above 100 μmol L⁻¹. Further, CH₄ consumption was completely inhibited when levulinic acid was added to soils at concentrations of 500 μmol L⁻¹ or 1000 μmol L⁻¹.

Discussion

Plants grown under elevated CO₂ contain increased tissue concentrations of secondary C compounds (Gebauer et al. 1997, Peñuelas and Estiarte 1998, Wetzel and Tuchman 2005b; Fig. 3.1, pathway A), which have the potential to impact CH₄ dynamics because they inhibit metabolism and growth by methanotrophs (Amaral and Knowles 1997, 1998). Indeed, we found evidence of transient inhibition of CH₄ consumption by duff collected from CO₂-enriched plots, as well as from control plots, and from an organic acid identified to be a primary root exudate of loblolly pine trees grown under elevated CO₂. However, neither throughfall nor leaf leachates from the four dominant tree species at the study site affected rates of CH₄ consumption by forest soils, regardless of the CO₂ treatment origins of the plant exudates.

The inhibition of CH₄ consumption by duff leachate from both CO₂-enriched and control treatments on one occasion suggests that some chemical(s) released from fresh autumnal duff inhibits methanotrophy, but the inhibitory substances are independent of CO₂ level. While we leached the same wet mass of duff from both CO₂ treatments to conduct our leaching experiment, there is actually greater litter fall under elevated CO₂ at our site (Allen et al. 2000, Lichter et al. 2005, Lichter et al. 2008), thus, perhaps higher concentrations of the inhibitory substances are leached from the larger mass of duff to the mineral soil in CO₂-enriched plots, which would result in a stronger inhibitory affect.

There are several possible reasons why duff collected on one occasion in the fall of 2006 was inhibitory to CH₄ consumption, while that collected on other occasions in the fall and winter of 2007 was not. For instance, losses of secondary C compounds from leaf litter occurs rapidly (Yavitt and Fahey 1986, Amaral and Knowles 1997, Schofield et al. 1998, Kainulainen and Holopainen 2002), and the concentrations of secondary C compounds leached from leaf litter are influenced by environmental conditions (Harris and Safford 1996). Yavitt and Fahey (1986) found that > 80% of the soluble phenolics and carbohydrates were lost from leaf litter in a lodgepole pine ecosystem in less than a year, and Amaral and Knowles (1997) reported that forest soil extracts only inhibited CH₄ consumption for 3 to 5 d. We may have collected duff in 2006 soon enough after leaf fall that inhibitory compounds in leachates were sufficiently concentrated to significantly reduce CH₄ consumption, yet we may have missed the window between leaf fall and leachate losses in the 2007 experiments. Timing of freeze/thaw cycles may have influenced the availability of inhibitory substances. Harris and Safford (1996) observed that repeated freeze/thaw cycles pre- and post- leaf fall, among other factors, increased

the amount of water-soluble carbon leached from fallen leaves from a temperate forest. Indeed, the first freeze/thaw cycle at our study site in 2006 occurred the day before duff collection, whereas the first freeze/thaw cycle in 2007 did not occur until 2 d after duff collection for the November 2007 experiment and 25 d before duff collection for the December 2007 experiment. This suggests that duff collection may have coincided with the maximum potential for leaching of inhibitory compounds in 2006, but not in subsequent experiments. Nonetheless, the degree of inhibition was apparently independent of the CO₂ level under which trees were grown.

Levulinic acid, a primary root exudate acid released from loblolly pines exposed to elevated CO₂, was found here to inhibit CH₄ consumption. The threshold concentration for inhibition by levulinic acid lies within the range of 50 to 100 μmol L⁻¹, which far exceeds the concentration of 0.5 μmol L⁻¹ for all phenolic compounds found in pore water from centrifuged samples of the top 25 cm of soil from a coniferous forest (Gallet and Pellissier 1997). Little is known about the presence and persistence of levulinic acid in forest soils. However, its increased release by tree roots under elevated CO₂ and significant and even complete inhibition of methanotrophy at concentrations between 50 and 100 μmol L⁻¹ suggest that this compound could inhibit methanotrophy in the rhizosphere, and warrants further attention in attempts to understand the feedback between an increasing atmospheric CO₂ concentration and a reduction in the forest soil sink strength for CH₄.

Despite some transient inhibition of net CH₄ consumption by forest soils by plant exudates reported here, the spatial and temporal patterns of *in situ* net CH₄ consumption observed in our previous study indicate that the secondary compounds in plant exudates

produced under elevated CO₂ are not the primary reason for the observed decrease in net CH₄ consumption by temperate forest soils under elevated CO₂ (Dubbs and Whalen submitted). The quantity and quality of plant exudates vary among plant species (Smith 1976, Ström et al. 1994). Likewise, the quantity and chemistry of plant exudates from roots or leaf litter are typically seasonal (Kuzyakov and Cheng 2001, Muscolo and Sidari 2006, Phillips et al. 2008). Saerte and Bååth (2000) reported “spatial patterns of the microbial community to be related to the positions of trees” in a mixed Norway spruce-birch stand in Finland. We previously reported high spatial and temporal variability in net CH₄ consumption at permanently established sampling locations in a temperate forest where there was not any specific site that consistently exhibited higher or lower rates of net CH₄ consumption relative to other sites (Dubbs and Whalen submitted). Since the patterns in net CH₄ consumption in the temperate forest at our study site do not correspond to specific locations or periods of time, it is only reasonable to conclude that then plant exudates do not exert the primary control on methanotrophy or methanogenesis. Consequently, our previous and present research indicates that despite transient inhibition of net CH₄ consumption in forest soils by plant exudates, Pathway A (Fig. 1) is not the primary driver for reduced net CH₄ consumption in soils under elevated CO₂. However, it does deserve further consideration since the transient influences of chemical inhibitors may weaken the correlation between standard influences on CH₄ consumption (soil moisture) and measured rates of net CH₄ consumption.

Future work should focus on identifying inhibitory compounds in bulk leachates that show enhanced production by plants under elevated CO₂ by high performance liquid chromatography. Focus should also be placed on identifying the temporal and spatial

patterns of influence of these compounds on *in situ* net CH₄ consumption. Research in this area will help to refine models aimed predicting the upland soil sink strength for CH₄.

References

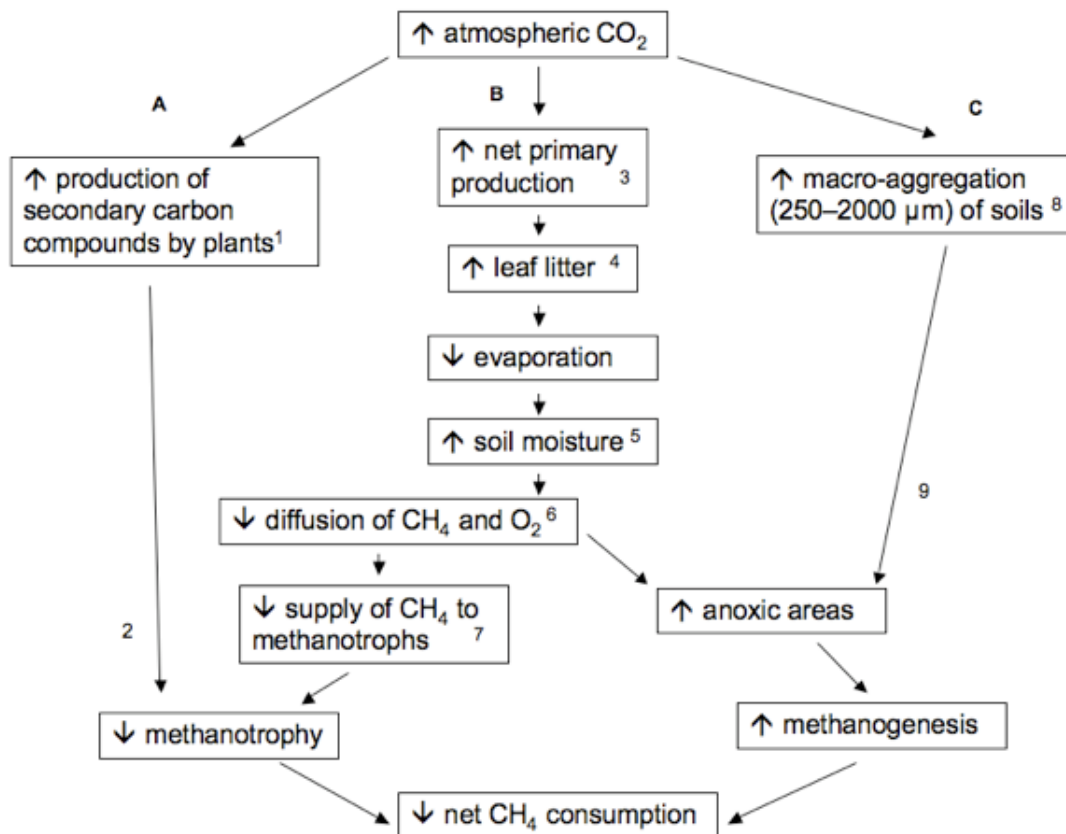
- Allen, A. S., J. A. Andrews, A. C. Finzi, R. Matamala, D. D. Richter, and W. H. Schlesinger. 2000. Effects of free-air CO₂ enrichment (FACE) on belowground processes in a *Pinus taeda* forest. *Ecological Applications* **10**:437-448.
- Amaral, J. A. and R. Knowles. 1997. Inhibition of methane consumption in forest soils and pure cultures of methanotrophs by aqueous forest soil extracts. *Soil Biology and Biochemistry* **29**:1713-1720.
- Amaral, J. A. and R. Knowles. 1998. Inhibition of methane consumption in forest soils by monoterpines. *Journal of Chemical Ecology* **24**:723-734.
- Billings, S. A. and S. E. Ziegler. 2005. Linking microbial activity and soil organic matter transformations in forest soils under elevated CO₂. *Global Change Biology* **11**:203-212.
- Castro, M. S., P. A. Steudler, J. M. Melillo, J. D. Aber, and R. D. Bowden. 1995. Factors controlling atmospheric methane consumption by temperate forest soils. *Global Biogeochemical Cycles* **9**:1-10.
- DeLucia, E. H., J. G. Hamilton, S. L. Naidu, R. B. Thomas, J. A. Andrews, A. Finzi, M. Lavine, R. Matamala, J. E. Mohan, G. R. Hendrey, and W. H. Schlesinger. 1999. Net primary production of a forest ecosystem with experimental CO₂ enrichment. *Science* **284**:1177-1179.
- DeLucia, E. H., D. J. Moore, and R. J. Norby. 2005. Contrasting responses of forest ecosystems to rising atmospheric CO₂: Implications for the global C cycle. *Global Biogeochemical Cycles* **19**: GB3006.
- Finzi, A. C., D. J. P. Moore, E. H. DeLucia, J. Lichter, K. S. Hofmockel, R. B. Jackson, H.-S. Kim, R. Matamala, H. R. McCarthy, R. Oren, J. S. Pippin, and W. H. Schlesinger. 2006. Progressive nitrogen limitation of ecosystem processes under elevated CO₂ in a warm-temperate forest *Ecology* **87**:15-25.
- Forster, P., V. Ramaswamy, P. Artaxo, T. Berntsen, R. Betts, D. W. Fahey, J. Haywood, J. Lean, D. C. Lowe, G. Myhre, J. R. Nganga, R. Prinn, G. Raga, M. Schulz, and R. Van Dorland. 2007. Changes in Atmospheric Constituents and in Radiative Forcing. *in* S. Solomon, D. Qin, M. Manning, J. Chen, M. Marquis, K. B. Averyt, M. Tignor, and A. J. Miller, editors. *Climate Change 2007: The Physical Sciences Basis. Contribution of the Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Gallet, C. and F. Pellissier. 1997. Phenolic compounds in natural solutions of a coniferous forest. *Journal of Chemical Ecology* **23**:2401-2412.

- Gebauer, R. L. E., B. R. Strain, and J. F. Reynolds. 1997. The effect of elevated CO₂ and N availability on tissue concentrations and whole plant pools of carbon-based secondary compounds in loblolly pine (*Pinus taeda*). *Oecologia* **113**:29-36.
- Hamilton, J., E. DeLucia, K. George, S. Naidu, A. Finzi, and W. Schlesinger. 2002. Forest carbon balance under elevated CO₂. *Oecologia* **131**:250-260.
- Harris, M. M. and L. O. Safford. 1996. Effects of season and four tree species on soluble carbon content in fresh and decomposing litter of temperate forests. *Soil Science* **161**:130-135.
- Hendrey, G. R., D. S. Ellsworth, K. F. Lewin, and J. Nagy. 1999. A free-air enrichment system for exposing tall forest vegetation to elevated atmospheric CO₂. *Global Change Biology* **5**:293-309.
- Hoosbeek, M. and G. Scarascia-Mugnozza. 2009. Increased litter build up and soil organic matter stabilization in a poplar plantation after 6 years of atmospheric CO₂ enrichment (FACE): final results of POP-EuroFACE compared to other forest FACE experiments. *Ecosystems* **12**:220-239.
- Kainulainen, R. and J. K. Holopainen. 2002. Concentrations of secondary compounds in Scots pine needles at different stages of decomposition. *Soil Biology and Biochemistry* **34**:37-42.
- Kuzyakov, Y. and W. Cheng. 2001. Photosynthesis controls of rhizosphere respiration and organic matter decomposition. *Soil Biology and Biochemistry* **33**:1915-1925.
- Lichter, J., S. H. Barron, C. E. Bevacqua, A. C. Finzi, K. F. Irving, E. M. Stemmler, and W. Schlesinger. 2005. Soil carbon sequestration and turnover in a pine forest after six years of atmospheric CO₂ enrichment. *Ecology* **86**:1835-1847.
- Lichter, J., S. A. Billings, S. E. Ziegler, D. Indh, R. Ryals, A. C. Finzi, R. B. Jackson, E. M. Stemmler, and W. H. Schlesinger. 2008. Soil carbon sequestration in a pine forest after 9 years of atmospheric CO₂ enrichment. *Global Change Biology* **14**:1-13.
- Lichter, J., M. Lavine, K. A. Mace, D. D. Richter, and W. H. Schlesinger. 2000. Throughfall Chemistry in a Loblolly Pine Plantation under Elevated Atmospheric CO₂ Concentrations. *Biogeochemistry* **50**:73-93.
- Liu, L., J. S. King, F. L. Booker, C. P. Giardina, H. L. Allen, and S. Hu. 2009. Enhanced litter input rather than changes in litter chemistry drive soil carbon and nitrogen cycles under elevated CO₂: a microcosm study. *Global Change Biology* **15**:441-453.

- Liu, L., J. S. King, and C. P. Giardina. 2005. Effects of elevated concentrations of atmospheric CO₂ and tropospheric O₃ on leaf litter production and chemistry in trembling aspen and paper birch communities. *Tree Physiology* **25**:1511-1522.
- Mann, C. J. and R. G. Wetzel. 1996. Loading and utilization of dissolved organic carbon from emergent macrophytes. *Aquatic Botany* **53**:61-72.
- Matamala, R. and W. H. Schlesinger. 2000. Effects of elevated atmospheric CO₂ on fine root production and activity in an intact temperate forest ecosystem. *Global Change Biology* **6**:967-979.
- Muscolo, A. and M. Sidari. 2006. Seasonal fluctuations in soil phenolics of a coniferous forest: effects on seed germination of different coniferous species. *Plant Soil* **284**:305-318.
- Norby, R. J., E. H. DeLucia, B. Gielen, C. Calfapietra, C. P. Giardina, J. S. King, J. Ledford, H. R. McCarthy, D. J. P. Moore, R. Ceulemans, P. De Angelis, A. C. Finzi, D. F. Karnosky, M. E. Kubiske, M. Lukac, K. S. Pregitzer, G. E. Scarascia-Mugnozza, W. H. Schlesinger, and R. Oren. 2005. Forest response to elevated CO₂ is conserved across a broad range of productivity. *Proceedings of the National Academy of Sciences of the United States of America* **102**:18052-18056.
- Norby, R. J., J. Ledford, C. D. Reilly, N. E. Miller, and E. G. O'Neill. 2004. Fine-root production dominates response of a deciduous forest to atmospheric CO₂ enrichment. *Proceedings of the National Academy of Sciences of the United States of America* **101**:9689-9693.
- Norby, R. J., E. G. O'Neill, W. G. Hood, and R. J. Luxmoore. 1987. Carbon allocation, root exudation, and mycorrhizal colonization of *Pinus echinata* seedlings grown under CO₂ enrichment. *Tree Physiology* **3**:203-210.
- Oh, N. H. and D. D. Richter. 2005. Elemental translocation and loss from three highly weathered soil-bedrock profiles in the southeastern United States. *Geoderma* **126**:5-25.
- Peñuelas, J. and M. Estiarte. 1998. Can elevated CO₂ affect secondary metabolism and ecosystem function? *Trees* **13**:20-24.
- Phillips, R. P., Y. Erlitz, R. Bier, and E. S. Bernhardt. 2008. New approach for capturing soluble root exudates in forest soils. *Functional Ecology* **22**:990-999.
- Ridgwell, A. J., S. J. Marshall, and K. Gregson. 1999. Consumption of atmospheric methane by soils: a process-based model. *Global Biogeochemical Cycles* **13**: 59-70.

- Saetre, P. and E. Bååth. 2000. Spatial variation and patterns of soil microbial community structure in a mixed spruce-birch stand. *Soil Biology and Biochemistry* **32**:909-917.
- Schäfer, K. V. R., R. Oren, D. S. Ellsworth, C.-T. Lai, J. D. Herrick, A. C. Finzi, D. D. Richter, and G. G. Katul. 2003. Exposure to an enriched CO₂ atmosphere alters carbon assimilation and allocation in a pine forest ecosystem. *Global Change Biology* **9**:1378-1400.
- Schlesinger, W. H. and J. Lichter. 2001. Limited carbon storage in soil and litter of experimental forest plots under increased atmospheric CO₂. *Nature* **411**:466-469.
- Schofield, J. A., A. E. Hagerman, and A. Harold. 1998. Loss of tannins and other phenolics from willow leaf litter. *Journal of Chemical Ecology* **24**:1409-1421.
- Sexstone, A. J., N. P. Revsbech, T. B. Parkin, and J. M. Tiedje. 1985. Direct measurement of oxygen profiles and denitrification rates in soil aggregates. *Soil Science Society of America Journal* **49**:645-651.
- Smith, W. H. 1976. Character and significance of forest tree root exudates. *Ecology* **57**:324-331.
- Souto, X. C., G. Chiapusio, and F. Pellissier. 2000. Relationships between phenolics and soil microorganisms in spruce forests: significance for natural regeneration. *Journal of Chemical Ecology* **26**:2025-2034.
- State Climate Office of North Carolina. 2003-2009. NC Climate Retrieval and Observations Network of the Southeast Database (NCCRONOS).
- Striegl, R. G. 1993. Diffusional limits to the consumption of atmospheric methane by soils. *Chemosphere* **26**:715-720.
- Ström, L., T. Olsson, and G. Tyler. 1994. Differences between calcifuge and acidifuge plants in root exudation of low-molecular organic acids. *Plant and Soil* **167**:239-245.
- Suwa, M., G. G. Katul, R. Oren, J. Andrews, J. Pippen, A. Mace, and W. H. Schlesinger. 2004. Impact of elevated atmospheric CO₂ on forest floor respiration in a temperate pine forest. *Global Biogeochemical Cycles* **18**: GB2013.
- Tuchman, N. C., R. G. Wetzel, S. T. Rier, K. A. Wahtera, and J. A. Teeri. 2002. Elevated atmospheric CO₂ lowers leaf litter nutritional quality for stream ecosystem food webs. *Global Change Biology* **8**:163-170.

- Verburg, P. S. J., W. K. P. Van Loon, and A. L¹/₄kewille. 1999. The CLIMEX soil-heating experiment: soil response after 2 years of treatment. *Biology and Fertility of Soils* **28**:271-276.
- Wetzel, R. G. and N. C. Tuchman. 2005a. Effects of atmospheric CO₂ enrichment and sunlight on degradation of plant particulate and dissolved organic matter and microbial utilization. *Archiv für Hydrobiologie* **162**:287-308.
- Wetzel, R. G. and N. C. Tuchman. 2005b. Effects of elevated CO₂ on the lignin and total phenolic concentrations of cattail and trembling aspen leaves. *Archiv für Hydrobiologie* **162**:287-308.
- Whalen, S. C. and W. S. Reeburgh. 1996. Moisture and temperature sensitivity of CH₄ oxidation in boreal soils. *Soil Biology and Biochemistry* **28**:1271-1281.
- Yavitt, J. B. and T. J. Fahey. 1986. Litter decay and leaching from the forest floor in *Pinus contorta* (Lodgepole Pine) ecosystems *The Journal of Ecology* **74**:525-545.
- Zausig, J., W. Stepniewski, and R. Horn. 1993. Oxygen concentration and redox potential gradients in unsaturated model soil aggregates. *Soil Science Society of America Journal* **57**:908-916.



¹ Gebauer et al.1997; Peñuelas and Estiarte 1998; Wetzel and Tuchman 2005
² Amaral and Knowles 1997, 1998
³ DeLucia et al. 1999; Hamilton et al. 2002; DeLucia et al. 2005; Norby et al. 2005; Finzi et al. 2006
⁴ Allen et al. 2000; Schlesinger and Lichter 2001

⁵ Schäfer et al. 2002
⁶ Suwa et al. 2004
⁷ Striegl 1993; Castro et al. 1995; Whalen and Reeburgh 1996
⁸ Hoosbeek and Scarascia-Mugnozza 2009
⁹ Sextone et al.1985; Zausig et al. 1993

Figure 3.1. Conceptual model of the impact of forest ecosystem responses to elevated CO₂ that influence soil CH₄ cycling dynamics. Up and down arrows within each response function indicate a positive or negative impact, respectively, of that factor on net atmospheric CH₄ consumption. Response functions are either documented or hypothesized by the associated references.

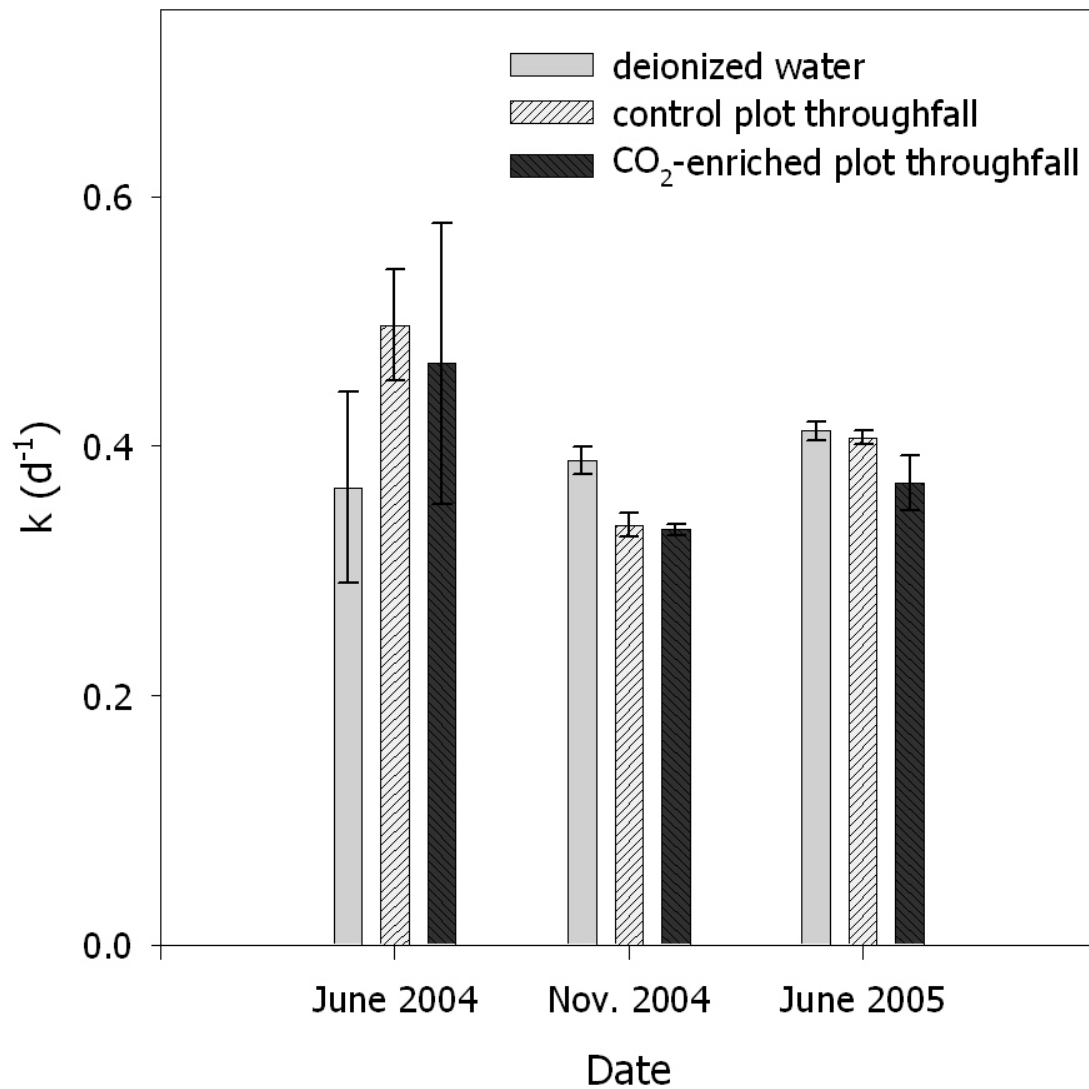


Figure 3.2. Mean first order rate constants (k ; d^{-1}) for CH_4 consumption in temperate forest soils amended with deionized water or throughfall from CO_2 -enriched ($n=3$) or control ($n=3$) plots. Error bars represent one standard error of the mean.

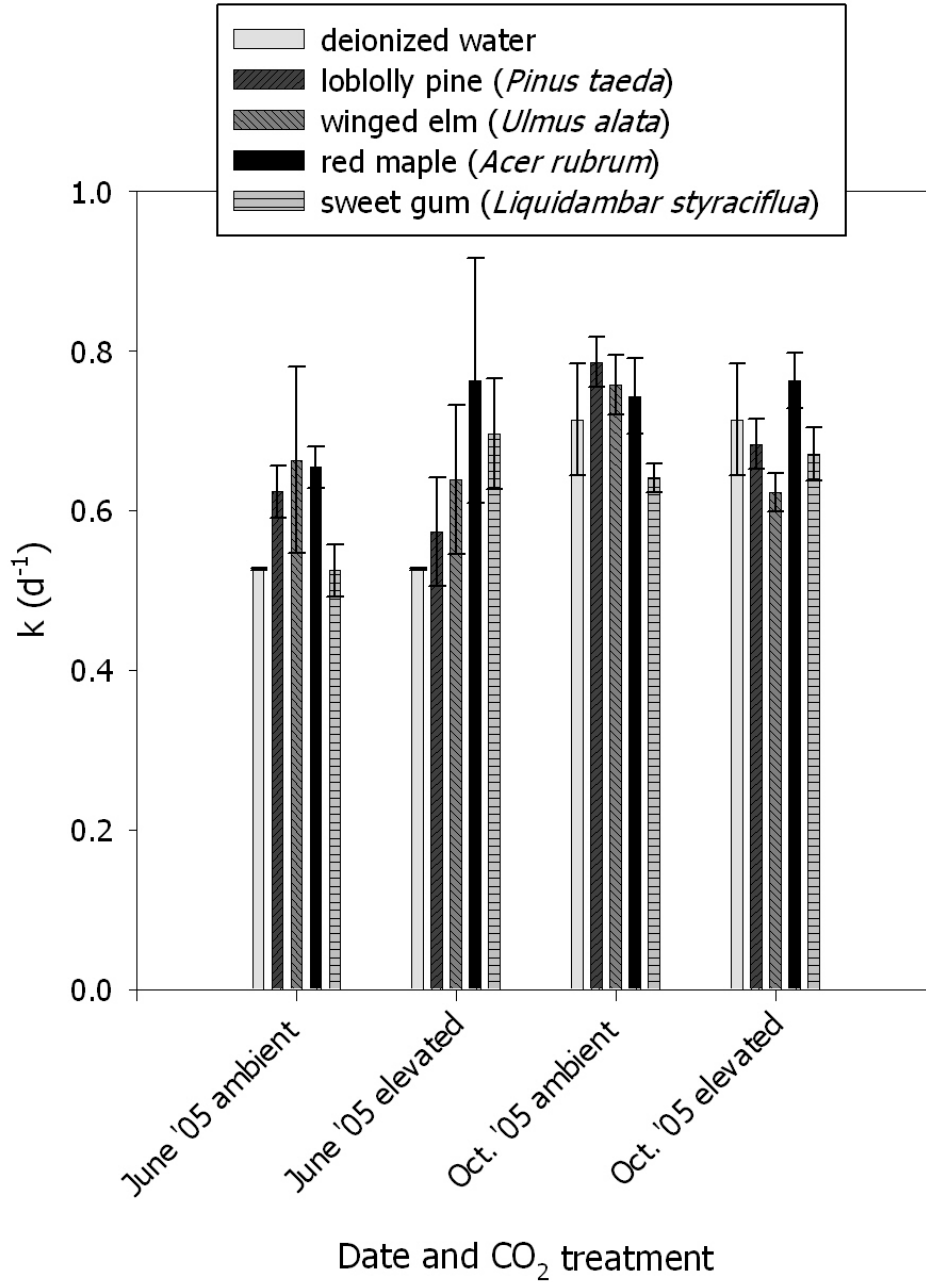


Figure 3.3. Mean first order rate constants (k ; d^{-1}) for CH_4 consumption by temperate forest soils amended with leaf leachate from the four most dominant trees within the control ($n=3$) and CO_2 -enriched ($n=3$) plots. Error bars represent one standard error of the mean.

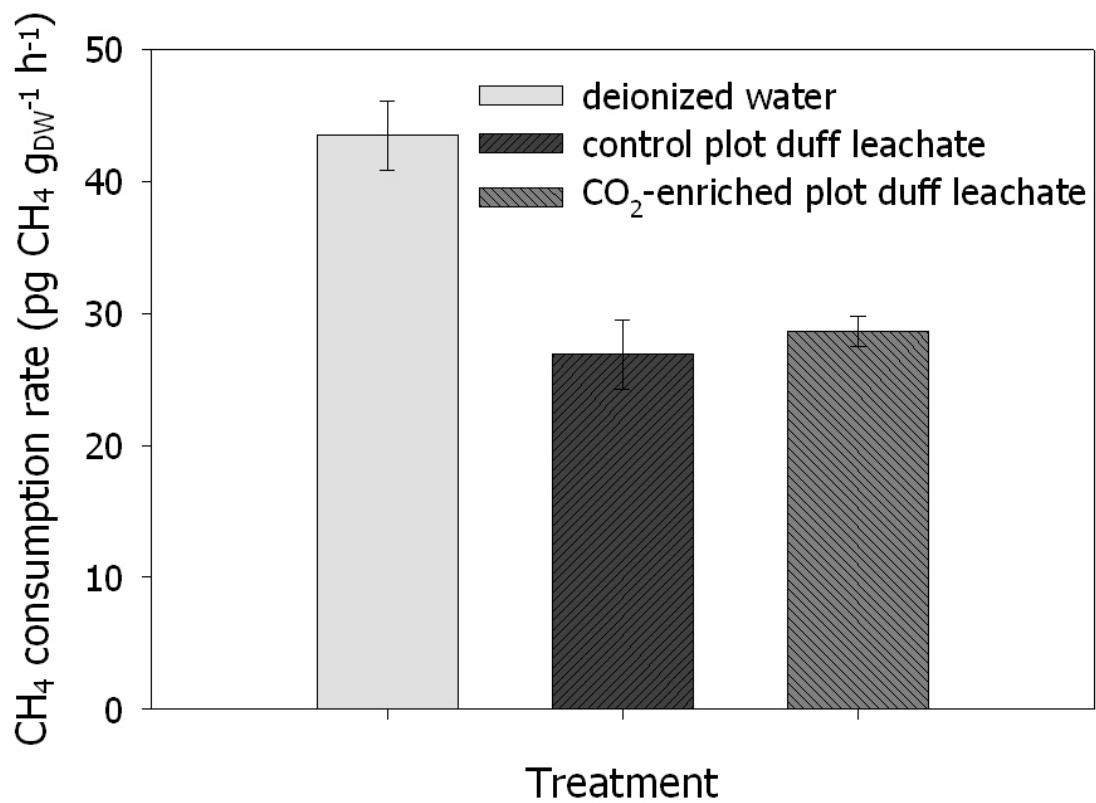


Figure 3.4. Rates of CH₄ consumption by forest soils amended with deionized water or duff leachates from CO₂-enriched (n=4) or control (n=4) plots. Error bars represent one standard error of the mean.

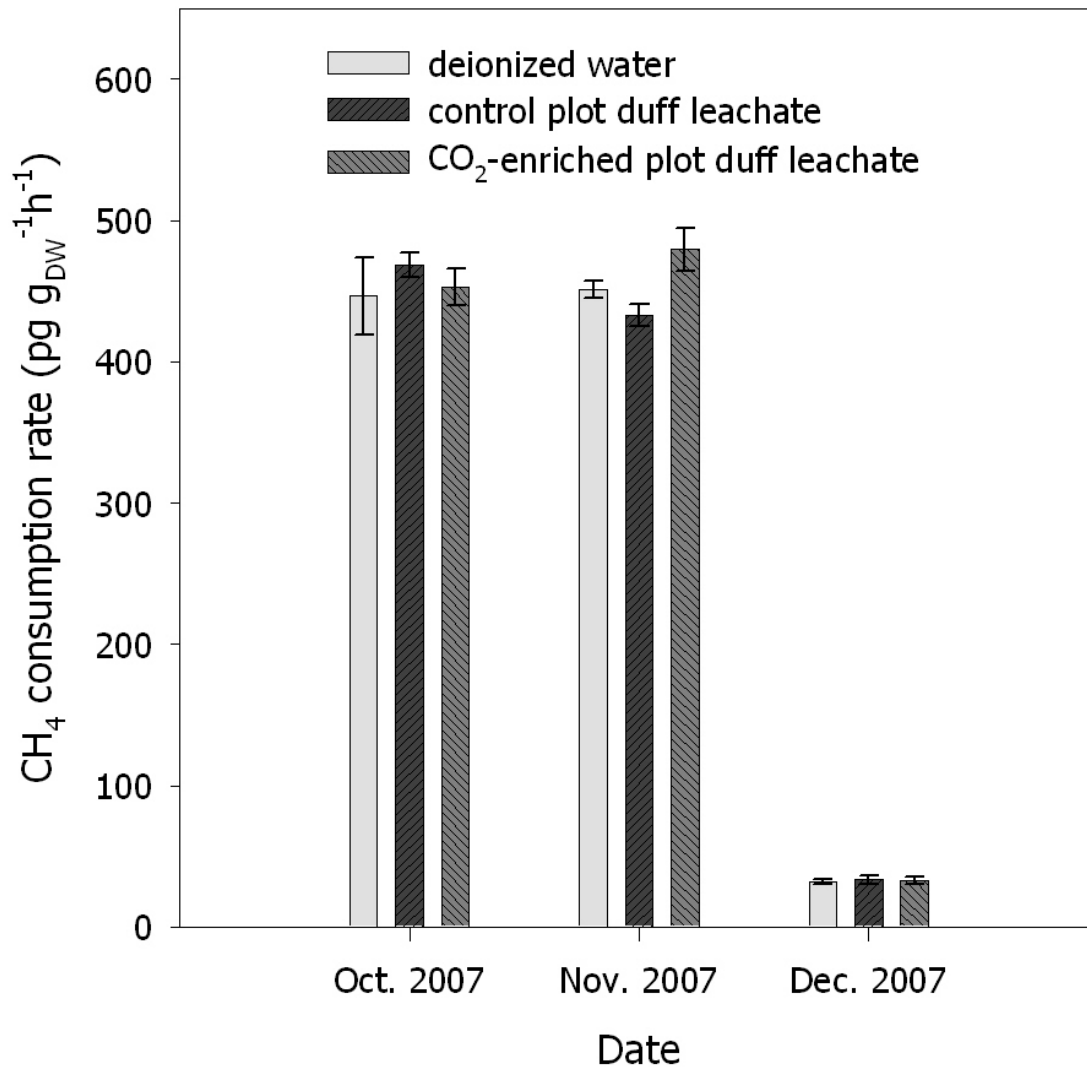


Figure 3.5. Rates of CH₄ consumption by forest soils amended with duff leachates or deionized water. Error bars represent one standard error of the mean (n=3).

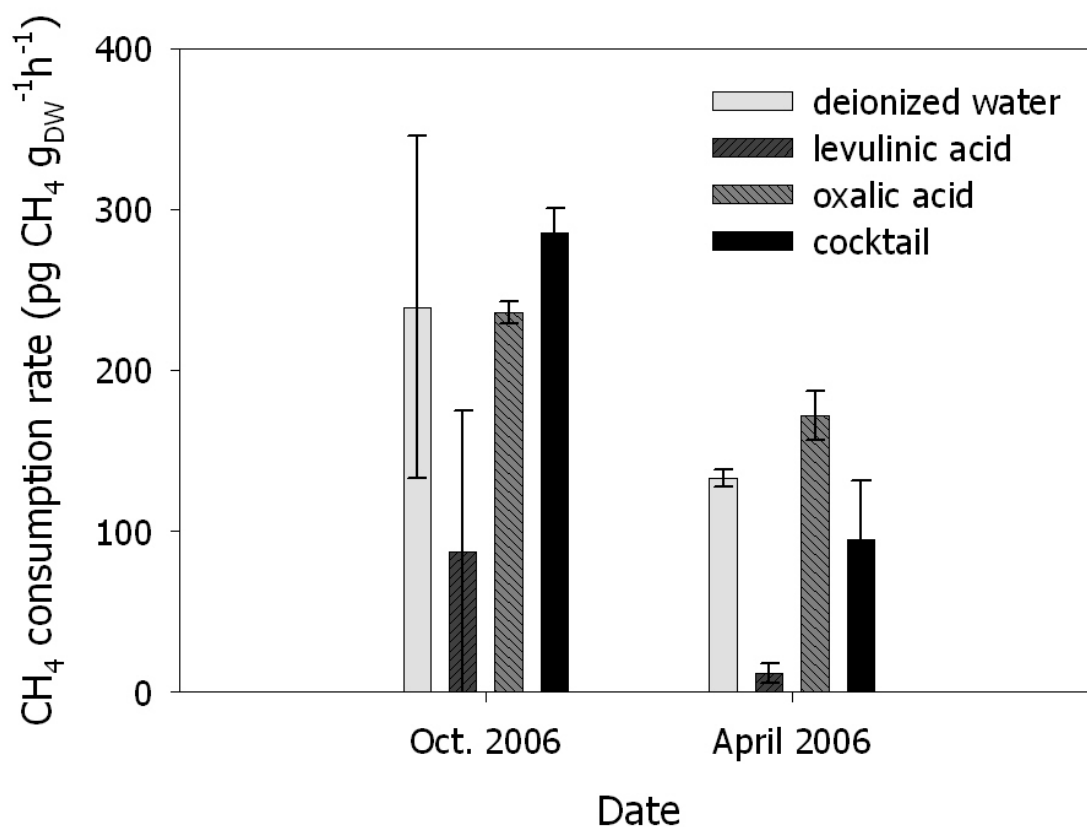


Figure 3.6. Rates of CH₄ consumption by forest soils amended with deionized water or representative organic acids (100 μmol L⁻¹) determined to be primary root exudates from loblolly pine (*Pinus taeda*) trees grown under elevated CO₂. Rates of CH₄ consumption by soils to which individual organic acids not shown here (citric, malic, maleic, fumaric, succinic, shikimic, and protocatecuic acids) was similar to that of soils to which oxalic acid and the cocktail of organic acids was added. Error bars represent one standard error of the mean (n=3).

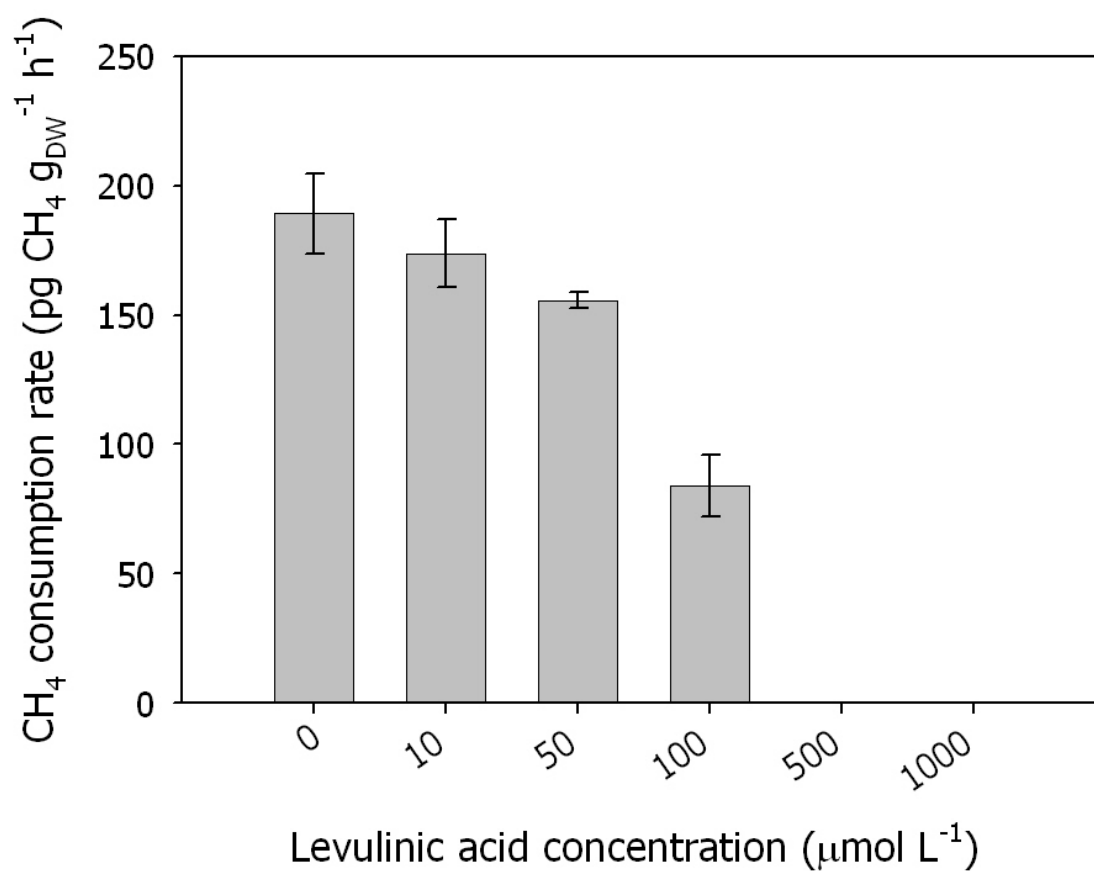


Figure 3.7. Rates of CH_4 consumption by forest soils amended with deionized water or levulinic acid, an organic acid determined to be a primary root exudate from loblolly pine (*Pinus taeda*) trees grown under elevated CO_2 for a February 2009 experiment. Error bars represent one standard error of the mean ($n=3$).

CHAPTER 4: REDUCED NET CH₄ CONSUMPTION CAUSED BY CHANGES IN THE SOURCES AND TRANSPORT OF SOIL GASES IN A TEMPERATE FOREST EXPOSED TO ELEVATED CO₂

Abstract

We previously reported a sustained reduction in net atmospheric CH₄ consumption by temperate forest soils exposed to elevated CO₂ since 1996 (~580 mL L⁻¹ CO₂; Dubbs and Whalen submitted). Changes in the transport and supply of atmospheric gases within the soil profile under elevated CO₂, and subsequent changes in locus or activity of the CH₄ oxidizing and producing communities, may help to explain the decrease in net CH₄ consumption. We examined the depth distribution of CH₄ in the soil profile, the effective diffusivity of CH₄ through the soil, and the extent and activity of CH₄ consuming and CH₄ producing communities in CO₂-enriched and control (ambient atmospheres) plots at the same study site. High spatial and temporal variability in net CH₄ consumption and CH₄ production rates and high error in diffusivity measurements, along with limited ability to collect soil samples, largely resulted in the inability to detect significant differences between CO₂ treatments in rates of net CH₄ consumption or CH₄ production, depth profile CH₄ concentrations, or effective diffusivity. However, qualitative trends of low overall diffusivity and increased incidence and rates of CH₄ production in elevated CO₂ plots, supported by a long-term record of significantly higher soil moisture in CO₂ plots, indicate that increased soil moisture along

with increased activity of methanogens under elevated CO₂ in soils with low diffusivity at our study site contribute to the observed decline in CH₄ oxidation under elevated CO₂.

Introduction

The atmospheric CH₄ concentration has more than doubled from a pre-industrial level of about 750 μL L⁻¹ to a present day concentration of about 1780 μL L⁻¹ (NOAA 2008). Although CH₄ is less abundant than CO₂, additions of CH₄ to the tropospheric reservoir cause more direct warming than CO₂, both on a per molecule and a mass basis (Wuebbles and Hayhoe 2002). Methane also indirectly contributes to global warming because of its role in the stratospheric chemistry of ozone and water vapor formation (Wuebbles and Hayhoe 2002). Thus, a complete understanding of the CH₄ cycle as well as the feedbacks and interactions with other biogeochemical cycles are important to the accurate prediction of future climates.

Known sinks for tropospheric CH₄ include reaction with the hydroxyl radical, which removes approximately 445 Tg of CH₄ from the atmosphere annually; and mixing of tropospheric CH₄ with the stratosphere accounts for another 40 Tg of CH₄ removal annually (Forster et al. 2007). Upland soils are the only known biological sink for atmospheric CH₄, accounting for approximately 38 Tg of CH₄. This biological sink results from the net balance of CH₄ consumption by methanotrophic bacteria in the largely oxic soil profile, and production by methanogenic bacteria in anoxic microsites (reviewed by Conrad 1996).

We recently reported a sustained reduction of ~15% in net atmospheric CH₄ consumption by temperate forest soils under elevated CO₂ relative to plots exposed to ambient levels of CO₂ (Dubbs and Whalen submitted). We also proposed several pathways

whereby changes in other aspects of forest ecosystem function in response to elevated CO₂ could impact net CH₄ consumption by these soils (Fig. 4.1). Reduced gas diffusivity has been demonstrated (Dörr et al. 1993) to control rates of CH₄ supply to the usual subsurface locus of CH₄ oxidation (e.g. Whalen and Reeburgh 1992), which is itself substrate-limited in well-drained forest soils, based on kinetic considerations (Bradford et al. 2001). Thus, factors that introduce diffusion resistance or increase the diffusional path will reduce rates of atmospheric CH₄ consumption (Fig. 4.1, pathway B). Similarly, reduced gas diffusivity slows the transport of O₂ from the atmosphere and, paired with respiratory consumption of O₂ within the soil matrix, may result in the formation of anoxic microsites, supporting methanogenesis (Fig. 4.1; pathway C). In this circumstance, methantrophs are supported not only by atmospheric CH₄, but also by endogenously produced substrate.

Soil moisture increases diffusional resistance because gases diffuse 10³ to 10⁴ times more slowly through water than air. Schäfer et al. (2003) and more recently, we (Dubbs and Whalen submitted) reported higher soil moisture in CO₂-enriched plots at a temperate forest study site, relative to plots exposed to ambient atmospheres. Schäfer et al. (2003) attributed the higher soil moisture in elevated CO₂ plots to increased leaf litter depth (Allen et al. 2000, Schlesinger and Lichter 2001) and topographic convergence. Increased leaf litter depth is a manifestation of increased net primary production in temperate forests in response to elevated CO₂ (DeLucia et al. 1999, Hamilton et al. 2002, DeLucia et al. 2005, Norby et al. 2005, Finzi et al. 2006a) that ultimately inhibits evaporation from the soil surface and results in higher soil moisture (Fig. 4.1, pathway B) while topographic convergence is an inherent difference in the lateral flow of soil pore water unrelated to CO₂ treatment.

Here we examine the transport of atmospheric CH₄ in the soil profile, the effective diffusivity of CH₄ through the soil, as well as the extent and activity of CH₄ consuming and producing communities in a temperate forest exposed to elevated CO₂. Changes in the transport and source of atmospheric gases within the soil profile under elevated CO₂, or a change in the locus of the CH₄ oxidizing community, may help to explain the observed persistent decrease in net CH₄ consumption under elevated CO₂ and provide information useful to modeling efforts aimed at forecasting future climates.

Methods

Field site

Field measurements were conducted at the Duke Forest (North Carolina) Free-Air CO₂ Enrichment (FACE) experiment sited in an even-aged stand of loblolly pine (*Pinus taeda* L.) planted in 1983. Soils are clay loam, Ultic Hapludalf's of the Enon Series (Oh and Richter 2005). Average air temperature ranges from 3.6 °C in January to 25.3 °C in July and annual precipitation averages 1209 mm (State Climate Office of North State Climate Office of North Carolina 2003-2009).

Site characteristics are fully documented in Hendrey et al. (1999a) and briefly described here. The experiment consists of eight circular 30-m diameter plots. Four treatment plots (referred to as “CO₂-enriched”) are fumigated with CO₂ to maintain atmospheric CO₂ concentrations 200 mL L⁻¹ above ambient levels, while three additional treatment plots are fumigated with ambient air to replicate micrometeorological effects associated with CO₂ addition. A fourth is subjected to ambient air without fumigation. The latter four plots are referred to as “controls”. Each plot is divided into quadrats by a

boardwalk that minimizes the impact of foot traffic during sampling. Continuous (24 h d⁻¹) fumigation was initiated in August 1996, but was reduced to daylight hours only from 2003 to present.

Soil physical characteristics are similar between CO₂ treatment plots, with the exception of soil organic matter, which averaged 4.6% in CO₂-enriched plots, and only 3.4% in control plots. Averages for all control and elevated CO₂ plots (0 to 20 cm depth zone) for soil particle density, bulk density, and pH were 2.5 g cm⁻³, 1.2 g cm⁻³, and 5.7 units, respectively. Soil texture was 9% clay, 42% silt, and 49% sand.

Soil gas sampling

Sets of soil gas wells were installed in 2005 within 30 cm of permanently emplaced static chambers utilized for soil-atmosphere CH₄ exchange determinations made in another aspect of this research (Dubbs and Whalen submitted). Wells were located at 5 cm depth intervals from 5 to 25 cm below the soil surface. There were a total of five wells per set located in two quadrants of each of the eight FACE plots for a total of 16 well sets per treatment. Each well consisted of 1 cm ID stainless steel tube, open and perforated at the bottom, and topped with Swagelock reducing union fitted with a septum for syringe sampling. The sampling wells were installed vertically such that the open and perforated bottom allowed diffusion of gases only from the prescribed depth.

Soil gas wells were sampled 31 times between July 2005 and July 2007. On each sampling date, wells were initially evacuated with a hand-operated vacuum pump (Handivac) and then allowed to equilibrate with soil air for approximately 0.5 to 1 h before sampling. Headspace samples were collected from each soil gas well in 10 ml glass syringes.

The atmosphere above the soil surface adjacent to wells was additionally sampled with similar syringes on each date.

Soil cores

Two soil cores (5.5 cm diameter by 25 cm length) were collected randomly from within each of the eight experimental plots in July 2005, September 2006, and April 2007 using a stainless steel soil core sampler (AMS, Inc) fitted with a slide hammer (AMS, Inc) and stainless steel liners (AMS, Inc). For each core, soil was extracted from the liner and divided into 2 depth increments from 0 to 15 cm, and from 15 to 25 cm, in the field. Soil core sections were then transported to the laboratory (<1 h) in Ziploc bags, sieved (4.75 mm mesh), and mixed.

One 10 g field moist aliquot of homogenized soil from each depth increment of each core was placed into a 120 mL glass serum bottle and allowed to equilibrate with laboratory air for 1 h. Serum bottles were then sealed with butyl rubber stoppers, crimp sealed, and incubated in the dark at approximately 20 °C. Headspace samples for net CH₄ consumption measurements were withdrawn into 10 mL gastight glass syringes at zero time and every 2 to 4 h interval thereafter for up to 3 d (n ≥ 4). Atmospheric pressure was maintained in the serum bottles by replacing removed headspace gas with an equivalent volume of ultrapure N₂. Replicate bottles were also sealed without soil and sampled in conjunction with experimental vessels to ensure that changes in headspace CH₄ concentrations did not result from exchange with butyl rubber stopper. Upon completion of net CH₄ consumption measurements, rates of CH₄ production were determined on the same samples. This was accomplished by addition of 50 Pa of difluoromethane (CH₂F₂), an inhibitor of

methanotrophy (Miller et al. 1998), to serum bottles, following the time course for CH₄ consumption in the serum vial headspace as described above.

Methane sample measurements

All gas samples were analyzed for CH₄ by flame ionization detection gas chromatography (Shimadzu model GC 8 A; precision expressed as the coefficient of variation for 10 replicate injections of a 0.94 mL CH₄ L⁻¹ standard was < 3%) within 10 h of collection, well within our predetermined holding time of 24 h. Sample separation was accomplished on a 1-m length x 0.32 cm diameter molecular sieve 5A column with an ultrahigh purity N₂ carrier gas (33 mL min⁻¹). Injector and detector temperatures were set at 90 °C and 140 °C.

Diffusivity

We employed a ²²²Rn-based method (Born et al. 1990) to estimate effective diffusivity at soil collars within each of the eight experimental plots that we have used in previous research (Phillips et al. 2001a, Dubbs and Whalen unpublished). The ²²²Rn-based method involves the simultaneous measurement of ²²²Rn flux from soil collars, using the static chamber method (Whalen et al. 1992), and measurement of soil air ²²²Rn and CH₄ concentrations at the soil surface and at a depth of 25 cm from our gas sampling wells. Two static chamber and well sets from each of the eight experimental plot were used for October 2008 and February 2009 diffusivity experiments while one static chamber and well set from each plot was used for April 2008 and July 2008 diffusivity experiments.

To initiate ^{222}Rn and CH_4 flux determinations, polyvinyl chloride covers fitted with a sampling port and capillary bleed were emplaced on soil collars. Radon-222 samples were withdrawn into 50 mL syringes from the soil surface, the 25 cm gas sampling well, and the static chamber. These samples were then used to fill evacuated 170 mL (volume) counting cells (Lucas) through quick connect fittings equipped with Teflon septa. Immediately following ^{222}Rn sample collection, a 10 mL chamber headspace sample and a 5 mL sample from each gas sampling well, from 5 cm to 25 cm depths, were also withdrawn into 10 mL glass syringes for CH_4 analysis. Additional static chamber headspace samples were similarly collected ~24 h later for ^{222}Rn and CH_4 analysis. Radon-222 activity was determined by scintillation counting of gas samples contained in Lucas cells using a portable radon monitor (Pylon Model AB-5). Gas samples were analyzed for CH_4 as described above.

Environmental measurements

Soil temperature and soil moisture were measured on each sampling date. Soil temperature was measured at 3 cm intervals from 1 cm to 19 cm depth with a multithermistors temperature probe. Volumetric soil moisture ($\text{mL H}_2\text{O cm}^{-3}$ soil) was measured by time domain reflectometry on each sampling occasion using a hand-held portable soil reflectometry sensor (Campbell Scientific 620 with 20 cm-long probe rods).

Calculations and Statistical Analysis

Rates of net CH_4 consumption and CH_4 production of core sections were calculated from the headspace volume of the bottles and log-linear or time-linear changes of CH_4

concentrations, respectively. Rates of net CH₄ consumption and CH₄ production in core sections from the soil surface to 15 cm, and those from 15 cm to 25 cm below the soil surface, were averaged for comparison of CO₂ treatments. Area-based rates of net CH₄ consumption determined commensurate to diffusivity observations were calculated from static chamber geometry and the log-linear change in the CH₄ concentration in static chamber headspaces.

Soil ²²²Rn profiles and ²²²Rn chamber flux measurements were used to calculate the effective diffusivity of CH₄ in the soil (P_{CH4}) according to Dörr and Münnich (1990):

$$P_{CH4} = D_{0,CH4} / D_{0,Rn} * P_{Rn}$$

where D_{0,CH4} and D_{0,Rn} are the diffusion coefficients of CH₄ (0.194 cm² s⁻¹ Lerman 1979) and Rn (0.1 cm² s⁻¹; Tanner 1964) in air and P_{Rn} is the permeability of Rn, which is the quotient of Rn flux divided by the concentration gradient of Rn in the soil profile. The effective diffusivity of CH₄ was then used to calculate the flux of CH₄ (J_{CH4}):

$$J_{CH4} = P_{CH4} * \Delta C_{CH4} / \Delta z_{CH4}$$

where $\Delta C_{CH4} / \Delta z_{CH4}$ is the linear change in CH₄ concentration (C_{CH4}) with depth (z_{CH4}).

Paired t-tests were used to analyze for statistical differences between CO₂ treatment means for CH₄ concentration. Paired t-tests were likewise used to analyze for statistical differences between CO₂ treatment averages of rates of net CH₄ consumption or CH₄ production. Differences in diffusivity between CO₂ treatment plots were compared by student t-tests for each of the four observations. All statistical analyses were performed at $\alpha=0.05$.

Results

Depth profiles of CH₄ concentrations

Soil CH₄ concentrations decreased sharply with depth from 5 to 20 cm below the soil surface. The rate of decline in CH₄ concentrations with depth decreased from 20 to 25 cm (Fig. 4.2). The depth profiles of average CH₄ concentrations in control and CO₂-enriched plots were similar. The average CH₄ concentration in control plots was slightly higher (0.1 mL L⁻¹) than in CO₂-enriched plots at 5 cm, but the average CH₄ concentration was slightly higher (0.01 to 0.05 mL L⁻¹) in CO₂-enriched plots relative to control plots at all other depths. However, there was not any significant difference in CH₄ concentrations between CO₂ treatments at any depth, nor for the whole soil profile, for any sampling date or for the entire data.

Depth profiles of net CH₄ consumption and CH₄ production

Net CH₄ consumption was observed in all core sections before addition of the CH₂F₂, at which point CH₄ production or zero flux of CH₄ was observed for the remainder of the observational period. Rates of net CH₄ consumption were similar in soils from both CO₂ treatments at each depth increment, ranging from 150 to 300 pg g_{DW}⁻¹ h⁻¹, (Fig. 4.3a and b). Methane production was more variable, spanning almost three orders of magnitude in soils from both the 0 to 15 cm, (Fig. 4.4a; 0.5 to 450 pg g_{DW}⁻¹ h⁻¹) and 15 to 25 cm depth intervals (Fig. 4.4b; 0 to 880 pg g_{DW}⁻¹ h⁻¹).

The average rates of net CH₄ consumption in control plot soils from 0 to 15 cm were up to 14% higher than, or nearly equivalent to, the average rates of net CH₄ consumption in

soils from the CO₂-enriched plots from the same depth (Fig. 4.3a). In contrast, the average rates of CH₄ production in CO₂-enriched plots from 0 to 15 cm depths were up to two orders of magnitude higher than, or nearly equivalent to, the average rates of CH₄ production from the same depth (Fig. 4.4a). The patterns in rates of net CH₄ consumption and CH₄ production in soils from 15 to 25 cm depths are less clear with regard to differences between CO₂ treatments. The experiment with the overall highest average rate of CH₄ production among the 15 to 25 cm depth increments showed a value for soils from CO₂-enriched plots that exceeded that for control plots (July 2005; Fig. 4.4b). This corresponded with a higher average rate of net CH₄ consumption in control plot soils (Fig. 4.3b). The rates of CH₄ production in soils from 15 to 25 cm core sections on both other sampling dates (Oct. 2006 and April 2007; Fig. 4.4b) were between 40 and 100% lower than the corresponding rates of net CH₄ consumption, and rates in soils from CO₂-enriched plots were higher than rates in control plots. The differences in rates of net CH₄ consumption and CH₄ production between CO₂ treatments were not significant for either depth interval or for the whole core on any date.

Effective diffusivity

There was not any consistent or significant difference in P_{CH₄} between CO₂ treatments. The effective diffusivity of CH₄ was higher in control plots on two of four dates (Table 4.1), which also corresponded with higher overall P_{CH₄} values. There was not a clear relationship between P_{CH₄} and soil moisture, or between P_{CH₄} and J_{CH₄} (Table 4.1). The calculated flux of CH₄ (J_{CH₄}) was more than 25% lower than measured CH₄ flux for three out of four observations (Table 4.1). The exception was in April 2008 when predicted (J_{CH₄}) and

measured CH₄ fluxes were only different by 20 and 24 % in CO₂-enriched and control plots, respectively. Relatively good agreement between measured and predicted rates of net CH₄ flux in April corresponded with the highest P_{CH₄} value. Soil moisture was higher in control plots relative to CO₂-enriched plots on each date when diffusivity measurements were made (Table 4.1).

Discussion

Methane concentrations at depth within the soil profile are determined by relative rates of diffusion from the atmosphere, consumption by methanotrophs, and/or production by methanogens. Our values for effective diffusivity are on the low end of reported values for a multitude of European (Born et al. 1990; Dörr et al. 1993) and boreal forest soils (Whalen et al. 1992), which range from 2 to 1504 cm² h⁻¹. The low effective diffusivity may in part be due to the clay loam texture of our soils as soil texture influences diffusivity (Ball et al. 1997). For example, clay soils show net CH₄ consumption rates an order of magnitude lower than sandy soils (Dörr et al. 1993). The low diffusivity of our soils was evident during the collection and measurement of ²²²Rn samples, as it was difficult to flush sample wells and subsequently collect sufficient sample for ²²²Rn analysis. Further, soils produced ²²²Rn at such low rates that accurate zero time ²²²Rn determination (t_{1/2} = 3.85 d) was problematic for stored, synoptically collected samples after the first few samples had been assayed because each assay required 6 h of counting. Therefore, the error associated with diffusivity measurements may overwhelm any treatment effect.

Nonetheless, decreased soil diffusivity, as a result of increased leaf litter depth and/or higher soil moisture, would reduce the substrate supply for methanotrophs, and could explain

the observed decline in net CH₄ consumption under elevated CO₂. The deeper duff in CO₂-enriched plots at our study site (Allen et al. 2000, Schlesinger and Lichter 2001, Lichter et al. 2005, Lichter et al. 2008; Fig. 4.1; pathway B) may be responsible for the observed reduction in net consumption of CH₄ under elevated CO₂. Duff has been shown to reduce diffusion of atmospheric gases to the mineral soil occupied by methanotrophs (Borken and Brumme 1997, Saari et al. 1997, Dong et al. 1998). Dong et al. (1998) observed that the removal of the leaves and humus layer from the soil surface resulted in 17% higher rates of CH₄ consumption by temperate forest soils. Similarly, we observed an increase of 6% in net CH₄ consumption when leaf litter was removed (data not shown), suggesting that a CO₂ treatment effect on leaf litter depth may contribute to reduced rates of net CH₄ consumption under elevated CO₂. Further, a thicker duff layer can slow evaporation from the soil surface thereby causing higher soil moisture in CO₂-enriched plots (Schäfer et al. 2003). Schäfer et al. (2003) observed significantly higher soil moisture in CO₂-enriched plots at our study site through 2002, although they proposed that the difference between CO₂ treatment plots did not necessarily reflect a treatment effect. We (Dubbs and Whalen unpublished) also found higher soil moisture in CO₂-enriched plots during biweekly determination of soil-atmosphere CH₄ exchange in 2004 and 2006, and that soil moisture explained 34% of the variability in rates of net CH₄ consumption. Higher soil moisture and associated reduction in diffusivity (Suwa et al. 2004) in CO₂-enriched plots likely contributes to the observed decrease in rates of net CH₄ consumption under elevated CO₂, regardless of the cause of increased soil moisture.

Low overall effective diffusivity and high soil moisture in CO₂-enriched plots would also slow the diffusion of atmospheric O₂ into and within the mineral soil. This reduced

supply of O₂, paired with increased respiratory O₂ consumption within the soil matrix under elevated CO₂ (Bernhardt et al. 2006) may result in the formation of anoxic microsites. Independent reports of anoxic microzones (Sexstone et al. 1985, Zausig et al. 1993) and methanogenic activity in macroscopically oxygenated soils (Yavitt et al. 1995, Saari et al. 1997, von Fischer and Hedin 2002, Teh et al. 2005) indicate that simultaneous CH₄ production and consumption are occurring in well-drained upland soils, with anoxic soil aggregates supporting localized zones of methanogenesis and oxic sites supporting methanotrophy. In fact, Hoosbeek and Scarascia-Munozza (2009) saw increased macro-aggregation (250–2000 μm) of soils under elevated CO₂ in a temperate *Populus x euramericana* plantation (Fig. 4.4; pathway C), and further found that the soil aggregates contained higher concentrations of C and N. Horn and Smucker (2005) found that the redox potential decreased rapidly, and thus the propensity for anoxia increased, when such soil aggregates were saturated by water. While not explicitly determined here, such loci of microbial activity where respiratory consumption of O₂ is enhanced and the development of anaerobic conditions are stimulated may explain the qualitative trend of increased CH₄ production under elevated CO₂ observed here. We previously reported episodic net CH₄ efflux from the soil (indicated net CH₄ production) under both CO₂ treatments, with nearly double the observations in CO₂-enriched plots (Dubbs and Whalen unpublished). We additionally measured gross CH₄ production in all soils from a 0 to 15 cm depth increment in laboratory experiments where an inhibitor of methanotrophy was administered (Fig. 4.4). Further, in the 15 to 25 cm depth increment, high rates of CH₄ production were clearly manifested by reduced rates of net CH₄ consumption (July 2005; Figs. 4.3 and 4.4). Finally, the depth profiles (Fig. 4.2) provide further evidence of methanogenesis at depth since there

were only 15 occasions out of 2480 observations, where the CH₄ concentration was drawn below the widely acknowledged threshold of about 0.2 μl L⁻¹ for high affinity methanotrophs (Bender and Conrad 1995), indicating a soil source of CH₄ augments the atmospheric supply to methanotrophs.

While reduced diffusivity of CH₄ in CO₂-enriched plots was not expressed in depth profiles of CH₄ concentrations, which showed no significant differences in CH₄ concentrations at any depth (Fig. 4.2), the depth profiles did suggest slightly higher CH₄ production in CO₂-enriched plots. Methane concentrations at depths between 10 and 20 cm in CO₂-enriched plots were slightly higher than those in control plots. Additionally, there was not any difference in CH₄ concentrations between CO₂ treatments at any depth (Fig. 4.2), nor was there a significant difference in net CH₄ oxidizing activity between CO₂ treatments at any depth (Fig. 4.3). A down-profile shift in the locus of the CH₄ oxidizing community in response to elevated CO₂ would increase the diffusional path of atmospheric CH₄ to the locus of CH₄ oxidation. Thus, this lack of difference in CH₄ concentrations between CO₂ treatments indicate that the long-term pattern of reduced net CH₄ consumption in soils exposed to elevated CO₂ (Phillips et al. 2001a, b, McLain et al. 2002, Whalen unpublished, Dubbs and Whalen unpublished) is not the result of such a downprofile shift in the CH₄ oxidizing community.

High spatial variability in net CH₄ consumption and CH₄ production rates and diffusivity measurements, along with limited ability to collect soil samples hampered our ability to detect significant differences between CO₂ treatments. However, trends indicating low overall diffusivity and increased incidence and rates of CH₄ production in elevated CO₂ plots relative to control plots. This conclusion is supported by long-term repeated field

measures of soil moisture and net atmospheric CH₄ consumption where significant, quantitative differences between CO₂ treatments were observed with CO₂-enriched plots showing higher soil moisture and lower rates of net atmospheric CH₄ consumption (Dubbs and Whalen, submitted). Thus, global changes that impact soil hydrology directly or through biological feedbacks (Denman et al. 2007) are useful predictors of the direction and rates of CH₄ flux in upland soils. Factors that increase soil aggregation can also be expected to influence CH₄ dynamics, although more research is needed regarding this pathway (Fig. 4.1, Pathway C).

References

- Allen, A. S., J. A. Andrews, A. C. Finzi, R. Matamala, D. D. Richter, and W. H. Schlesinger. 2000. Effects of free-air CO₂ enrichment (FACE) on belowground processes in a *Pinus taeda* forest. *Ecological Applications* **10**:437-448.
- Amaral, J. A. and R. Knowles. 1997. Inhibition of methane consumption in forest soils and pure cultures of methanotrophs by aqueous forest soil extracts. *Soil Biology and Biochemistry* **29**:1713-1720.
- Amaral, J. A. and R. Knowles. 1998. Inhibition of methane consumption in forest soils by monoterpines. *Journal of Chemical Ecology* **24**:723-734.
- Ball, B. C., K. E. Dobbie, J. P. Parker, and K. A. Smith. 1997. The influence of gas transport and porosity on methane oxidation in soils. *Journal of Geophysical Research* **102**:23301-23308.
- Bender, M. and R. Conrad. 1995. Effect of CH₄ concentrations and soil conditions on the induction of CH₄ oxidation activity. *Soil Biology and Biochemistry* **27**:1517-1527.
- Bernhardt, E., J. Barber, J. Pippen, L. Taneva, J. Andrews, and W. Schlesinger. 2006. Long-term effects of free air CO₂ enrichment (FACE) on soil respiration. *Biogeochemistry* **77**:91-116.
- Borken, W. and R. Brumme. 1997. Liming practice in temperate forest ecosystems and the effects on CO₂, N₂O and CH₄ fluxes. *Soil Use and Management* **13**:251-257.
- Born, M., H. Dorr, and I. Levin. 1990. Methane consumption in aerated soils of the temperate zone. *Tellus B* **42**:2-8.
- Bradford, M. A., P. Ineson, P. A. Wookey, and H. M. Lappin-Scott. 2001. Role of CH₄ oxidation, production and transport in forest soil CH₄ flux. *Soil Biology and Biochemistry* **33**:1625-1631.
- Castro, M. S., P. A. Steudler, J. M. Melillo, J. D. Aber, and R. D. Bowden. 1995. Factors controlling atmospheric methane consumption by temperate forest soils. *Global Biogeochemical Cycles* **9**:1-10.
- Conrad, R. 1996. Soil microorganisms as controllers of atmospheric trace gases (H₂, CO, CH₄, OCS, N₂O, and NO). *Microbiological Reviews* **60**:609-640.
- DeLucia, E. H., J. G. Hamilton, S. L. Naidu, R. B. Thomas, J. A. Andrews, A. Finzi, M. Lavigne, R. Matamala, J. E. Mohan, G. R. Hendrey, and W. H. Schlesinger. 1999. Net primary production of a forest ecosystem with experimental CO₂ enrichment. *Science* **284**:1177-1179.

- DeLucia, E. H., D. J. Moore, and R. J. Norby. 2005. Contrasting responses of forest ecosystems to rising atmospheric CO₂: Implications for the global C cycle. *Global Biogeochemical Cycles* **19**: GB3006.
- Denman, K. L., G. Brasseur, A. Chidthaisong, P. Ciais, P. Cox, R. E. Dickinson, D. Hauglustaine, C. Heinze, E. Holland, D. Jacob, D. Lohmann, S. Ramachandran, P. L. da Silva Dias, S. C. Wofsy, and X. Zhang. 2007. Couplings Between Changes in the Climate System and Biogeochemistry. *in* S. Solomon, D. Qin, M. Manning, J. Chen, and M. Marquis, editors. *Climate Change 2007: The Physical Sciences Basis. Contributions of the Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Dong, Y., D. Scharffe, J. M. Lobert, P. J. Crutzen, and E. Sanhueza. 1998. Fluxes of CO₂, CH₄, and N₂O from a temperate forest soil: the effects of leaves and humus layers. *Tellus* **50B**:243-252.
- Dörr, H., L. Katruff, and I. Levin. 1993. Soil texture parameterization of the methane uptake in aerated soils. *Chemosphere* **26**:697-713.
- Dörr, H. and K. O. Münnich. 1990. ²²²Rn flux and soil air concentration profiles in West Germany. Soil ²²²Rn as tracer for gas transport in the unsaturated soil zone. *Tellus* **42B**:20-28.
- Finzi, A. C., D. J. P. Moore, E. H. DeLucia, J. Lichter, K. S. Hofmockel, R. B. Jackson, H.-S. Kim, R. Matamala, H. R. McCarthy, R. Oren, J. S. Phippen, and W. H. Schlesinger. 2006. Progressive nitrogen limitation of ecosystem processes under elevated CO₂ in a warm-temperate forest *Ecology* **87**:15-25.
- Forster, P., V. Ramaswamy, P. Artaxo, T. Berntsen, R. Betts, D. W. Fahey, J. Haywood, J. Lean, D. C. Lowe, G. Myhre, J. R. Nganga, R. Prinn, G. Raga, M. Schulz, and R. Van Dorland. 2007. Changes in Atmospheric Constituents and in Radiative Forcing. *in* S. Solomon, D. Qin, M. Manning, J. Chen, M. Marquis, K. B. Averyt, M. Tignor, and A. J. Miller, editors. *Climate Change 2007: The Physical Sciences Basis. Contribution of the Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
- Gebauer, R. L. E., B. R. Strain, and J. F. Reynolds. 1997. The effect of elevated CO₂ and N availability on tissue concentrations and whole plant pools of carbon-based secondary compounds in loblolly pine (*Pinus taeda*). *Oecologia* **113**:29-36.
- Hamilton, J., E. DeLucia, K. George, S. Naidu, A. Finzi, and W. Schlesinger. 2002. Forest carbon balance under elevated CO₂. *Oecologia* **131**:250-260.

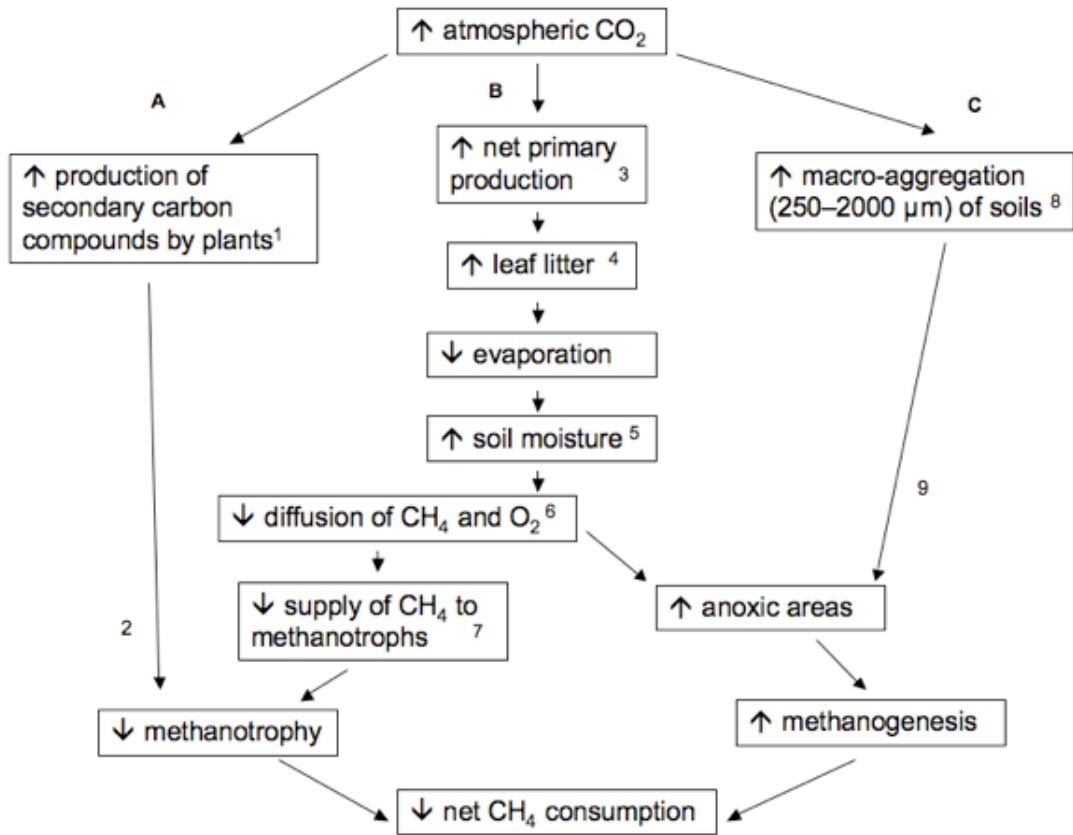
- Hendrey, G. R., D. S. Ellsworth, K. F. Lewin, and J. Nagy. 1999. A free-air enrichment system for exposing tall forest vegetation to elevated atmospheric CO₂. *Global Change Biology* **5**:293-309.
- Hoosbeek, M. and G. Scarascia-Mugnozza. 2009. Increased litter build up and soil organic matter stabilization in a poplar plantation after 6 years of atmospheric CO₂ enrichment (FACE): final results of POP-EuroFACE compared to other forest FACE experiments. *Ecosystems* **12**:220-239.
- Horn, R. and A. Smucker. 2005. Structure formation and its consequences for gas and water transport in unsaturated arable and forest soils. *Soil and Tillage Research* **82**:5-14.
- Lerman, A. 1979. *Geochemical processes: water and sediment environments*. Wiley, NY.
- Lichter, J., S. H. Barron, C. E. Bevacqua, A. C. Finzi, K. F. Irving, E. M. Stemmler, and W. Schlesinger. 2005. Soil carbon sequestration and turnover in a pine forest after six years of atmospheric CO₂ enrichment. *Ecology* **86**:1835-1847.
- Lichter, J., S. A. Billings, S. E. Ziegler, D. Indh, R. Ryals, A. C. Finzi, R. B. Jackson, E. M. Stemmler, and W. H. Schlesinger. 2008. Soil carbon sequestration in a pine forest after 9 years of atmospheric CO₂ enrichment. *Global Change Biology* **14**:1-13.
- McLain, J. E. T., T. B. Keppeler, and D. M. Ahnman. 2002. Belowground factors mediating changes in methane consumption in a forest soil under elevated CO₂. *Global Biogeochemical Cycles* **16**:1050-1064.
- Miller, L. G., C. Sasson, and R. S. Oremland. 1998. Difluoromethane, a new and improved inhibitor of methanotrophy. *Applied and Environmental Microbiology* **64**:4357-4362.
- NOAA. 2008. Carbon dioxide, methane rise sharply. Page http://www.noaa.gov/stories2008/20080423_methane.html. NOAA News.
- Norby, R. J., E. H. DeLucia, B. Gielen, C. Calfapietra, C. P. Giardina, J. S. King, J. Ledford, H. R. McCarthy, D. J. P. Moore, R. Ceulemans, P. De Angelis, A. C. Finzi, D. F. Karnosky, M. E. Kubiske, M. Lukac, K. S. Pregitzer, G. E. Scarascia-Mugnozza, W. H. Schlesinger, and R. Oren. 2005. Forest response to elevated CO₂ is conserved across a broad range of productivity. *Proceedings of the National Academy of Sciences of the United States of America* **102**:18052-18056.
- Oh, N. H. and D. D. Richter. 2005. Elemental translocation and loss from three highly weathered soil-bedrock profiles in the southeastern United States. *Geoderma* **126**:5-25.

- Peñuelas, J. and M. Estiarte. 1998. Can elevated CO₂ affect secondary metabolism and ecosystem function? *Trees* **13**:20-24.
- Phillips, R. L., S. C. Whalen, and W. H. Schlesinger. 2001a. Influence of atmospheric CO₂ enrichment on methane consumption in a temperate forest soil. *Global Change Biology* **7**:557-563.
- Phillips, R. L., S. C. Whalen, and W. H. Schlesinger. 2001b. Response of soil methanotrophic activity to carbon dioxide enrichment in a North Carolina coniferous forest. *Soil Biology and Biochemistry* **33**:793-800.
- Saari, A., P. J. Martikainen, A. Ferm, J. Ruuskanen, W. De Boer, S. R. Troelstra, and H. J. Laanbroek. 1997. Methane oxidation in soil profiles of Dutch and Finnish coniferous forests with different soil texture and atmospheric nitrogen deposition. *Soil Biology and Biochemistry* **29**:1625-1632.
- Schäfer, K. V. R., R. Oren, D. S. Ellsworth, C.-T. Lai, J. D. Herrick, A. C. Finzi, D. D. Richter, and G. G. Katul. 2003. Exposure to an enriched CO₂ atmosphere alters carbon assimilation and allocation in a pine forest ecosystem. *Global Change Biology* **9**:1378-1400.
- Schlesinger, W. H. and J. Lichter. 2001. Limited carbon storage in soil and litter of experimental forest plots under increased atmospheric CO₂. *Nature* **411**:466-469.
- Sexstone, A. J., N. P. Revsbech, T. B. Parkin, and J. M. Tiedje. 1985. Direct measurement of oxygen profiles and denitrification rates in soil aggregates. *Soil Science Society of America Journal* **49**:645-651.
- State Climate Office of North Carolina. 2003-2009. NC Climate Retrieval and Observations Network of the Southeast Database (NCCRONOS).
- Striegl, R. G. 1993. Diffusional limits to the consumption of atmospheric methane by soils. *Chemosphere* **26**:715-720.
- Suwa, M., G. G. Katul, R. Oren, J. Andrews, J. Pippen, A. Mace, and W. H. Schlesinger. 2004. Impact of elevated atmospheric CO₂ on forest floor respiration in a temperate pine forest. *Global Biogeochemical Cycles* **18**: GB2013.
- Tanner, A. 1964. Radon migration in the ground: A review. Pages 161-190 *The Natural Radiation Environment*. University of Chicago Press, Chicago, IL.
- Teh, Y. A., W. L. Silver, and R. Conrad. 2005. Oxygen effects on methane production and oxidation in humid tropical forest soils. *Global Change Biology* **11**:1283-1297.

- von Fischer, J. C. and L. O. Hedin. 2002. Separating methane production and consumption with a field-based isotope pool dilution technique. *Global Biogeochemical Cycles* **16**: 1034.
- Wetzel, R. G. and N. C. Tuchman. 2005. Effects of elevated CO₂ on the lignin and total phenolic concentrations of cattail and trembling aspen leaves. *Archiv für Hydrobiologie* **162**:287-308.
- Whalen, S. C. and W. S. Reeburgh. 1992. Interannual variations in tundra methane emission: a 4-year time series at fixed sites. *Global Biogeochemical Cycles* **6**:139-159.
- Whalen, S. C. and W. S. Reeburgh. 1996. Moisture and temperature sensitivity of CH₄ oxidation in boreal soils. *Soil Biology and Biochemistry* **28**:1271-1281.
- Whalen, S. C., W. S. Reeburgh, and V. A. Barber. 1992. Oxidation of methane in boreal forest soils: a comparison of seven measures. *Biogeochemistry* **16**:181-211.
- Wuebbles, D. J. and K. Hayhoe. 2002. Atmospheric methane and global change. *Earth-Science Reviews* **57**:177-210.
- Yavitt, J. B., T. J. Fahey, and J. A. Simmons. 1995. Methane and carbon dioxide dynamics in a northern hardwood ecosystem. *Soil Science Society of America Journal* **59**:796-804.
- Zausig, J., W. Stepniewski, and R. Horn. 1993. Oxygen concentration and redox potential gradients in unsaturated model soil aggregates. *Soil Science Society of America Journal* **57**:908-916.

Table 4.1. Effective diffusivity (P_{CH_4}) and corresponding calculated net CH_4 flux (J_{CH_4} ; $\mu g m^{-2} h^{-1}$), measured net CH_4 flux ($\mu g m^{-2} h^{-1}$), and soil moisture in control and CO_2 enriched plots (n=3, each treatment for July 2008 and April 2008; n=6, each treatment for Oct.2008 and Feb. 2009) at the Duke FACE site.

Date	CO_2 treatment	P_{CH_4} ($cm^2 h^{-1}$)	J_{CH_4} ($\mu g m^{-2} hr^{-1}$)	Measured net CH_4 flux ($\mu g m^{-2} hr^{-1}$)	Soil moisture (mL $H_2O cm^{-3}$ soil)
July '08	Control	2.8	1.3	1.7	0.32
	Enriched	2.4	1.1	1.5	0.27
April '08	Control	1.8	0.7	1.6	0.33
	Enriched	1.3	0.5	1.6	0.31
Oct. '08	Control	0.9	0.4	1.6	0.24
	Enriched	1.1	0.4	1.8	0.24
Feb. '09	Control	0.0	0.0	0.8	0.39
	Enriched	0.5	0.1	0.6	0.34



¹ Gebauer et al. 1997; Peñuelas and Estiarte 1998; Wetzel and Tuchman 2005

² Amaral and Knowles 1997, 1998

³ DeLucia et al. 1999; Hamilton et al. 2002; DeLucia et al. 2005; Norby et al. 2005; Finzi et al. 2006

⁴ Allen et al. 2000; Schlesinger and Lichter 2001

⁵ Schäfer et al. 2002

⁶ Suwa et al. 2004

⁷ Striegl 1993; Castro et al. 1995; Whalen and Reeburgh 1996

⁸ Hoosbeek and Scarascia-Mugnozza 2009

⁹ Sextone et al. 1985; Zausig et al. 1993

Figure 4.1. Conceptual model of the impact of forest ecosystem responses to elevated CO₂ that influence soil CH₄ cycling dynamics. Up and down arrows within each response function indicate a positive or negative impact, respectively, of that factor on net atmospheric CH₄ consumption. Response functions are either documented or hypothesized by the associated references.

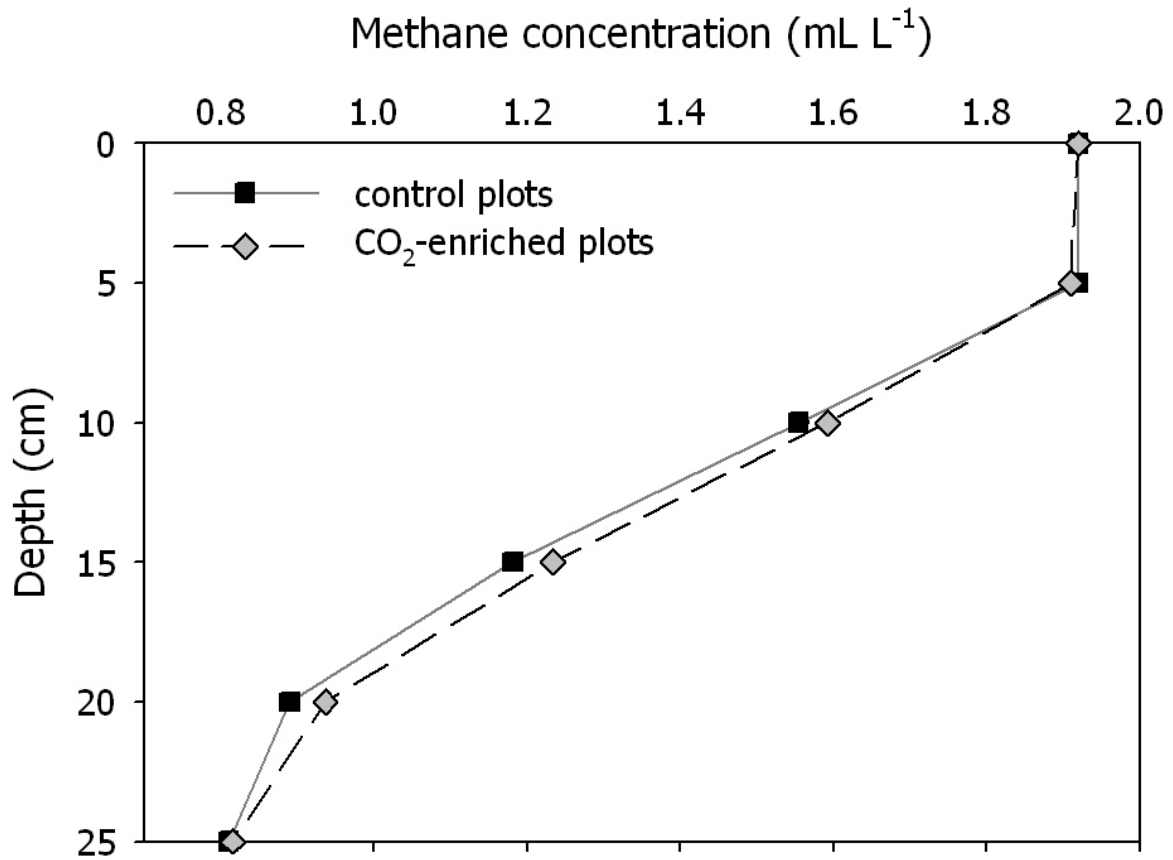


Figure 4.2. Composite depth profiles of CH_4 in soils in forest plots exposed to elevated CO_2 or the ambient atmosphere (control). Data for each depth represent the mean from 8 soil gas wells for each treatment over 31 dates. Error bars are eliminated for clarity

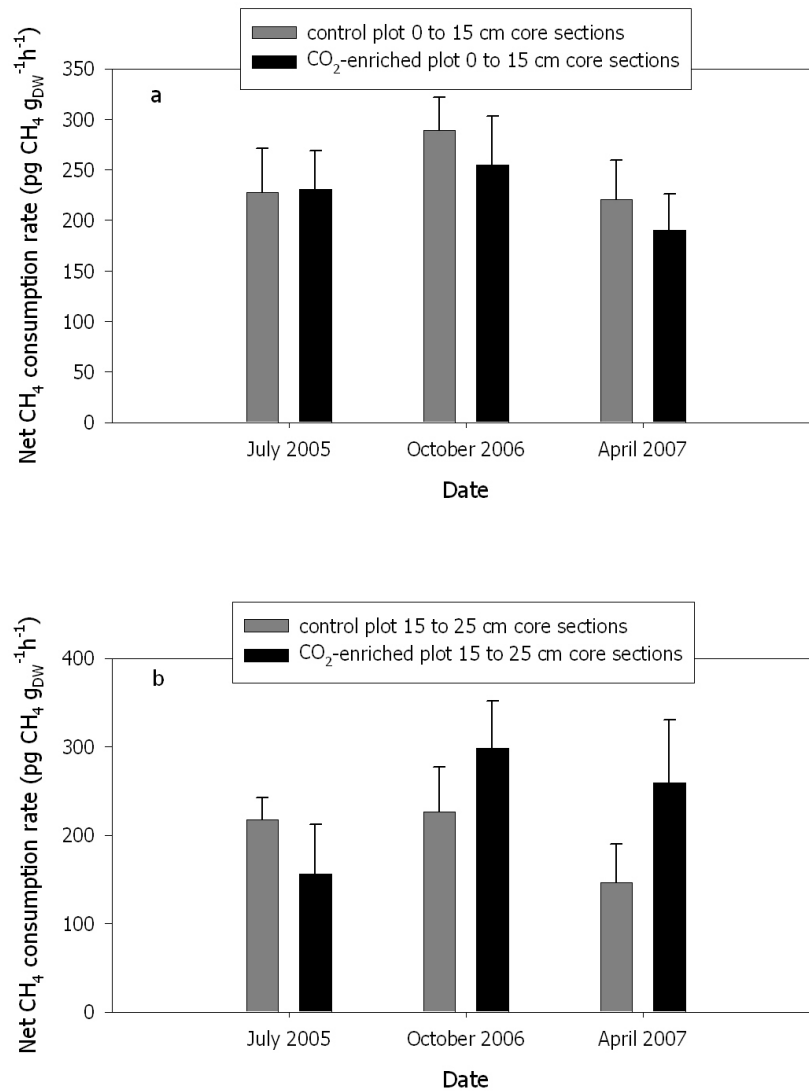


Figure 4.3. Mean rates of net CH₄ consumption in forest soils from plots exposed to elevated CO₂ or the ambient atmosphere (control). Data are mean rates for: a) 0 to 15 cm; and b) 15 to 25 cm depth increments from 16 cores. Error bars represent 1 standard error of the mean (n=8).

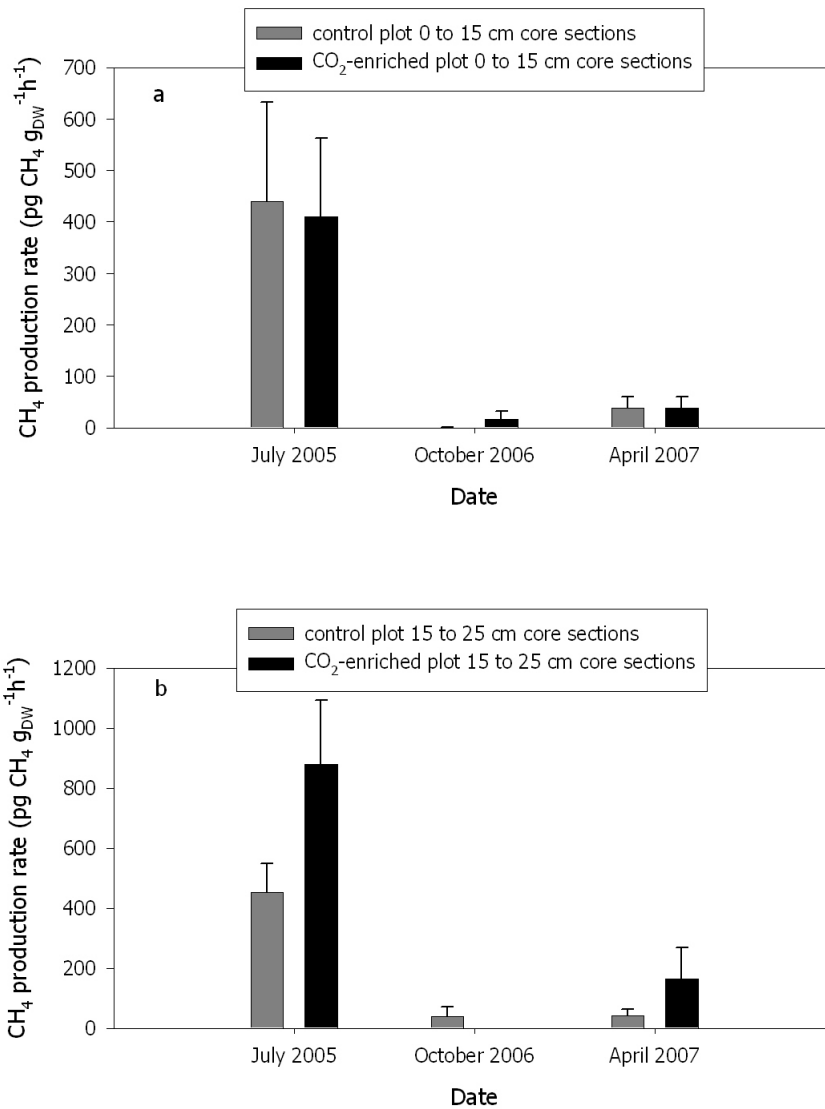


Figure 4.4. Mean rates of CH₄ production in forest soils from plots exposed to elevated CO₂ or the ambient atmosphere (control). Data are mean rates for: a) 0 to 15 cm; and b) 15 to 25 cm depth increments from 16 cores. Error bars represent 1 standard error of the mean (n=8).

CHAPTER 5: CONCLUSION

My research suggests that reduced net CH₄ consumption is a sustained, equilibrium response of this temperate forest soil to elevated CO₂. I observed lower annualized rates of net CH₄ consumption in soils from CO₂-enriched plots relative to controls for all 3 y of my study, extending the record from permanently emplaced soil collars at my study site (Phillips *et al.* 2001; Whalen and Fischer unpublished) to 8 nearly continuous years. The average decrease in net CH₄ consumption under elevated CO₂ for all annual observations was ~15% and there was not any consistent change in the magnitude of the CO₂ enrichment effect on the CH₄ sink strength over the extended record.

A decline in soil CH₄ consumption of the magnitude observed here (~15%) across all forest biomes with an estimated aggregated sink of 24 Tg CH₄ y⁻¹ (Ridgwell *et al.* 1999), gives a decrease of 3.6 Tg CH₄ y⁻¹. This reduction represents 10% of the model estimate (Ridgwell *et al.* 1999) of 38 Tg CH₄ y⁻¹ for the total soil sink.

Causative factors for the observed decrease in net CH₄ consumption under elevated CO₂ are difficult to identify, as the destructive sampling necessary for process-level investigations is limited to maintain ecosystem integrity at the Duke FACE site. However, the modeled soil moisture x CO₂ treatment interaction for 3 y of field measurements of net CH₄ flux and corresponding environmental variables was significant, indicating that site-wise differences in net CH₄ consumption are at least in part moisture-related. The observation that soil moisture explains 34% of the variability in net CH₄ measurements further supports soil

moisture control of net CH₄ consumption where soil moisture is higher in elevated CO₂ plots. However, soil moisture does not appear to be the only driver of the observed decline in net CH₄ consumption under elevated CO₂.

I found that some plant exudates from this forest ecosystem inhibit CH₄ consumption, including levulinic acid, an organic acid that is released from plant roots in greater quantities under elevated CO₂; and duff leachates from the duff of both CO₂ treatment plots, but which is thicker under elevated CO₂. These leachates do not exert consistent control over rates of atmospheric CH₄ consumption. However, their temporal and spatial influence on net CH₄ consumption under elevated CO₂ deserve further consideration since their transient influences may weaken the correlation between well-studied influences on CH₄ consumption and measured rates of net CH₄ consumption.

While high spatial variability and high error, along with limited ability to collect soil samples largely resulted in the inability to detect significant differences in rates of CH₄ consumption and CH₄ production and soil diffusivity between CO₂ treatments, qualitative trends showed low overall effective diffusivity of these soils and increased incidence and rates of CH₄ production in elevated CO₂ plots. When these trends are viewed together with the contributions of soil moisture to explaining reduced net CH₄ consumption under elevated CO₂, it is apparent that increased activity of methanogens under elevated CO₂ contribute to the observed decline in CH₄ oxidation at this study site.

My research has identified several research needs. These include further investigation of the spatial and temporal inhibitory influences of plant compounds produced under elevated CO₂ that may influence rates of CH₄ consumption, as well as determination of the factors that contribute to formation of anoxic microsites in upland soils under elevated

CO₂. If my results can be broadly extrapolated, my research also suggests that a 200 ml L⁻¹ increase in present-day atmospheric CO₂ concentrations can be expected to reduce the forest soil sink for CH₄ of ~24 Tg y⁻¹ by approximately 15%. Further, the observed relationship between increasing soil moisture and the reduction in the forest sink for CH₄ indicates that climate forecasting models can constrain the predicted upland sink for CH₄ by relating it to soil hydrology.

References

Phillips, R. L., S. C. Whalen, and W. H. Schlesinger. 2001. Influence of atmospheric CO₂ enrichment on methane consumption in a temperate forest soil. *Global Change Biology* **7**:557-563.

Ridgwell, A. J., S. J. Marshall, and K. Gregson. 1999. Consumption of atmospheric methane by soils: a process-based model. *Global Biogeochemical Cycles* **13**:59-70.