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Stimulation of Anterior Thalamic Nucleus Protects Hippocampus Neural Injury in Kainic Acid-Induced Epileptic Rats

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Abstract

Anterior nucleus of thalamus (ANT) stimulation has been proved to be effective in the treatment of refractory epilepsy, but the underlying mechanisms remain to be elucidated. We examined the role of ANT stimulation on hippocampal neuron death after seizures induced by kainic acid (KA). Our data showed that ANT stimulation could significantly rescue neurons from death induced by seizures, by reducing the release of cytochrome c (cyto c) and also via apoptosis-inducing factor (AIF) induced by seizures through inhibiting the activated caspase-9 and caspase-3. Our data suggest that ANT stimulation may protect against neuronal loss and reduce neuronal injury in the ipsilateral CA3 region of the hippocampus in the KA-induced epileptic rats, and the underlying mechanism may be mediated by inhibiting mitochondrial caspase-dependent (cyto c release and the subsequent cleavage of caspase-) and caspase-independent (nuclear translocation of AIF) apoptosis pathways.

Keywords: Epilepsy; Anterior nucleus of thalamus; Stimulation; Apoptosis; Neuroprotection; Cytochrome c; Apoptosis-inducing factor

Introduction

Epilepsy is a progressive neurodegenerative disorder characterized by an enduring predisposition to generate seizures [1]. Temporal lobe epilepsy (TLE) is the most common type of pharmaco-resistant epilepsy in adults, and one third patients is poorly controlled or their seizures are refractory to drug treatment [2]. The resection of mesial temporal structures can lead to the absence of seizures in more than 70% of patients [3]. However, such surgery is not suitable for up to 30% TLE patients [4]. High frequency stimulation (HFS) has become an alternative treatment for intractable epilepsy, as HFS can be tailored to address the specific requirements of the patients. Although several small-scale studies on patients with anterior nucleus of thalamus (ANT) stimulation showed a similar significant reduction of approximately 50% in the number of seizures [5], the underlying mechanism of how ANT stimulation decreases seizure frequency is still unknown. Apoptotic cell death has emerged as an important factor in brain injury produced by prolonged epileptic seizures or status epilepticus [6]. Our previous study has demonstrated that hippocampal high frequency stimulation has protective on hippocampal neurons by inhibition of apoptosis in epilepsy macaques [7]. Furthermore, we have proposed hypotheses that ANT-DBS could have neuroprotective effects in epilepsy patients by regulating the metabolism of amino acids [8]. Yet, it is still unclear whether ANT stimulation has protection on hippocampal neurons. Kainic acid (KA), a potent neuroexcitatory amino acid has been widely used to establish TLE models, which closely imitates the pathophysiological alterations in patients with TLE [9,10]. Intra-hippocampal KA injection in rats has been widely used a model to study the mechanisms of seizure-induced neuronal cell death in the hippocampus [11,12]. Therefore, in the present study, we examined the protective effects and potential mechanisms of ANT stimulation on hippocampal neurons in kainic acid epileptic rats.

Materials and Methods

Animals

Male Sprague Dawley rats weighing 280 g-320 g were provided by Experimental Animal Center of Military Medical Sciences, China. Animals were individually housed in a cage (constant temperature, 22-

25°C; humidity, 60-70%; illumination, 12/12 h cycles) with free access to food and water. All experiments were performed during the light phase of the cycle. The care and handling of animals were conducted in compliance with the Chinese Animal Welfare Act and was approved by the responsible governmental agency at Capital Medical University. Sixty-four rats were randomized into four groups: (1) control group (n=16), underwent administration of saline; (2) KA model group (n=16), underwent administration of KA; (3) sham stimulation group (n=16), received KA injections with implantation of electrodes into the ipsilateral ANT but without stimulation; (4) stimulation group (n=16), received KA injections with implantation of electrodes into the ipsilateral ANT and stimulation.

Seizure model

All rats were anesthetized with chloral hydrate (350 mg/kg, i.p.) and mounted in a stereotaxic frame (David Kopf Instruments, USA). Right injections of KA into the CA3 region of the hippocampus were injected at the following coordinate: A/P=5.6 mm, R=4.5 mm, V=-5.5mm [13]. KA, at a dose of 0.5 µg/0.2 µL (in phosphate buffer) for a 250 g rat (and adjusted according to body weight), was injected slowly during 10 min with a 0.5 µL Hamilton micro syringe in KA group, sham stimulation group and stimulation group [14]. Rats of the control group were stereotaxically injected with equivalent saline instead. The rats were continuously monitored for behavioral seizures for 24 h after injection.

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Implantation of electrode

Three weeks later, the concentric bipolar stimulation electrodes (outer diameter 200 μ m, inner diameter 50 μ m, CBBPF50, FHC, USA) were glued to the guide cannula with a specially made device and the electrode tips were 2 mm below the end of the guide cannula. One cannula was implanted for right ANT (AP=-2.0 mm, R=1.8 mm, V=4.7 mm) within sham stimulation group and stimulation group [13]. The cannulas were cemented in place using dental cement to affix them to three stainless steel screws fastened to the skull. Penicillin (6 \times 10⁷ U) was administered intraperitoneally three times a day for the first three days after surgery. After a recovery of seven days, continuous unilateral ANT stimulation was then performed using a stimulator system (Master-8 and ISO-flex, Israel). Parameters were: 500 μ A, 60 μ s and 130 Hz. However, the sham stimulation group was manipulated in the same way but without electrical stimulation.

Fixation and processing of tissue

After three months, the rats were harvested and ipsilateral hippocampus resection was then performed. A portion of each resected sample was immediately placed in liquid nitrogen for subsequent western blot analysis; another portion was fixed in 10% buffered formalin and 2.5% glutaraldehyde for paraffin embedding, Nissl staining, apoptosis detection and electron microscope.

Nissl staining and apoptosis detection

Tissues were post-fixed for 24 h and then placed for another 24 h in 30% sucrose in 0.1 M phosphate buffer. Coronal sections (12 μ m) underwent Nissl staining with Toluidine Blue and terminal deoxynucleotidyl transferase-mediated DUTP nick-end labeling (Tunel) staining with a peroxidase in situ apoptosis detection kit (Roche Molecular Biochemicals, Indianapolis, IN). Tunel staining was performed as recommended by the manufacturer. Cell counts were performed on n=6 brain sections for the entire hippocampal CA3 subfields. And the numbers of surviving hippocampal CA3 region neurons or the Tunel-positive neurons at 100 μ m in diameter were automatically counted in blind manner.

Preparation of cytosolic fraction and mitochondrial fractions

The cytosolic fraction was generated according to previous reported methods [15]. Briefly, tissue was homogenized in buffer containing 100 mM sucrose, 1 mM EGTA, 20 mM MOPS, pH 7.4 with 5% Ficoll, 0.01% digitonin, 1 μ g/ml pepstatin, 2 μ g/ml leupeptin and 100 μ g/ml PMSF and incubated for 15 min on ice. Samples were then centrifuged at 2500x g for 10 min to precipitate the nuclei and cellular debris. The supernatant was centrifuged at 15,000x g for 15 min to pellet mitochondria. Mitochondria were collected at the 1-1.5 M interphase by lateral suction through the tube, washed in four volumes of MS buffer with centrifugation at 15,000x g between washes, and resuspended in a final volume of 200 μ l MS buffer. The supernatant was subsequently centrifuged at 100,000x g for 60 min to obtain the cytosol (supernatant) [15].

Western blotting

Protein was extracted from samples in 100 μ l lysis buffer and concentration was determined using the bicinchoninic acid protein assay kit (Pierce Biotechnology, Inc., Rockford, IL, USA). Samples of 4-10 μ g protein were resolved by 10% SDS-PAGE, and transferred to a polyvinylidene fluoride membrane (Bio-Rad, Hercules, CA, USA).

These were blocked with 5% nonfat dry milk in Tris-buffered saline and 0.1% Tween 20, and probed with antibodies against actin (1:5000, Abcam), apoptosis-inducing factor (AIF, 1:2000, Sigma), cytochrome c (cyto c) (1:1000, Sigma), caspase-9 (1:1000, Sigma) cleaved caspase-3 (1:1000, Sigma). The band was detected using a chemiluminescent peroxidase substrate followed by exposure to X-ray film. The density of bands on the membrane were scanned and analyzed using an image software (Alpha Innotech, San Leandro, CA, USA).

Transmission electron microscopy

For electron microscopy, the samples were fixed with 1% w/v osmium tetroxide, dehydrated in a graded ethanol and embedded in Epon. Ultrathin sections (70 nm) were cut with a microtome (Bromma, Sweden), counterstained with uranyl acetate and lead citrate and viewed using an electron microscope (Hitachi H7650, Tokyo, Japan).

Statistical Analysis

Statistical analysis was performed using SPSS 13.0 (SPSS Inc., Chicago, IL, USA). Data were expressed as mean \pm SD. Data were analyzed using one-way analysis of variance (ANOVA) with a post hoc test. A P value <0.05 was considered statistically significant.

Results

Effects of ANT stimulation on hippocampal neuronal cells

Nissl staining revealed that seizures led to significant neuronal loss in the hippocampal CA3 regions of the KA group. The number of surviving neurons was decreased significantly compared with the control (P<0.05), but ANT stimulation in stimulation group significantly attenuated the neuronal loss induced by seizure compared with KA group (Figure 1). However, no significant difference in neuronal loss were found between KA group and sham stimulation group (P>0.05). A large number of Tunel positive neurons were found in ipsilateral CA3 of the KA group compared with that of the control group. In contrast, stimulation group with ANT stimulation show remarkably decreased Tunel positive cells compared with KA group (Figure 2). However, there was no significant difference in the number of CA3 Tunel positive cells between sham stimulation group and KA group. These results demonstrated that KA-induced seizures caused prominent neuronal loss in the hippocampus and ANT stimulation could rescue neurons from death induced by seizures. Electron microscopically, loss of nucleolus and granularly coagulated chromosome aggregated under the membrane with high electron density were observed in the KA group and the sham stimulation group (Figure S1). However, no damaged neural cells were found in the controls and in the stimulation group.

ANT stimulation reduces mitochondrial release of cyto c, AIF following seizures

To investigate the effect of ANT stimulation on the mitochondrial release of cyto c and AIF induced by seizures, we detected the appearance of cyto c and AIF in the cytosolic fraction from the hippocampus samples. Western blot analysis revealed that the levels of cyto c and AIF largely increased following KA injection in the KA group compared with that of the control group whereas ANT stimulation reduced the levels of cyto c and AIF in the stimulation group compared with the KA group (P<0.05) (Figure 3). There were no significant difference in cyto c and AIF between the KA group and the sham stimulation group (P>0.05). Moreover, a significant decrease of cyto c and AIF in the mitochondrial fraction was noted in the KA group compared the control group and stimulation group, respectively.

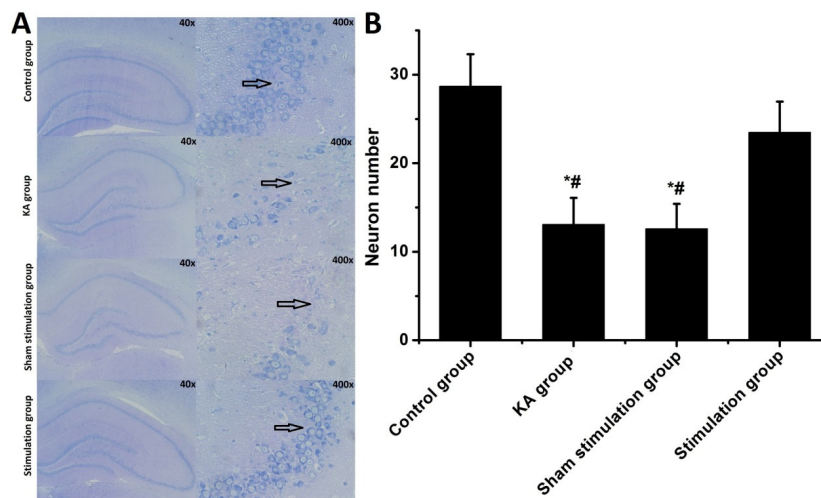


Figure 1: Morphology and number of hippocampal CA3 neurons in rats receiving KA injection and ANT stimulation. (A) Nissl staining of hippocampal CA3 neurons. (B) Quantification of surviving hippocampal CA3 neurons in each group.

Histogram data are shown as mean ± SD

*P<0.05, compared to control group; #P<0.05, compared to ANT stimulation group

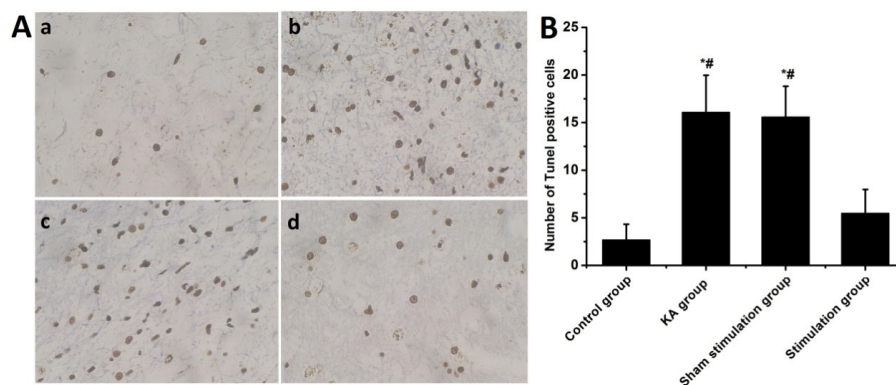


Figure 2: Hippocampal apoptosis in rats receiving KA injection and HFS. A: TUNEL staining of hippocampal CA3 neurons, (a) control group, (b) KA group, (c) sham stimulation group, (d) stimulation group; B: Quantification of TUNEL positive cells in each group.

Histogram data are shown as mean ± SD

*P<0.05, compared to control group; #P<0.05, compared to ANT stimulation group

ANT stimulation inhibits caspase-9 and caspase-3 activation triggered by seizures

To further investigate the underlying neuroprotective mechanisms of ANT stimulation on brain injury induced by seizures, we utilized Western Blotting to measure the levels of caspase-9 and active cleavage product of caspase-3. The levels of caspase 9 and the active cleavage product of caspase-3 were significantly increased in the KA group compared to the control (P<0.05) while ANT stimulation reduced the levels of caspase-9 and cleaved caspase-3 compared to the KA group (P<0.05) (Figure 4). Nevertheless, there was no significant difference in caspase-3 and caspase-9 activity between KA group and sham stimulation group (P>0.05). These results illustrate a possible neuroprotective mechanism against seizures by ANT stimulation through the inhibition of caspase-9 and caspase-3.

Electron microscopic observation

An examination by electron microscopy revealed that KA

treatment caused mitochondrial swelling as well as malformation and disintegration of cristae in the KA group and the sham stimulation group (Figure 5). However, mitochondria in the control group were round and had numerous transverse cristae with parallel alignment. And mitochondrial structures in the stimulation group were similar to that of the control group, providing direct evidence that ANT stimulation mitigate KA-induced neuronal apoptosis by limiting mitochondrial injury.

Discussion

In the present study, neuronal death in the hippocampus was clearly observed following KA-induced seizures in rats, which was consistent with previous report [6]. Our results illustrated that ANT stimulation could significantly decrease the numbers of TUNEL positive cells and increase the number of surviving cells, indicating that ANT stimulation could rescue neurons from death induced by seizures and has a protective effect on hippocampal neuronals in KA-induced

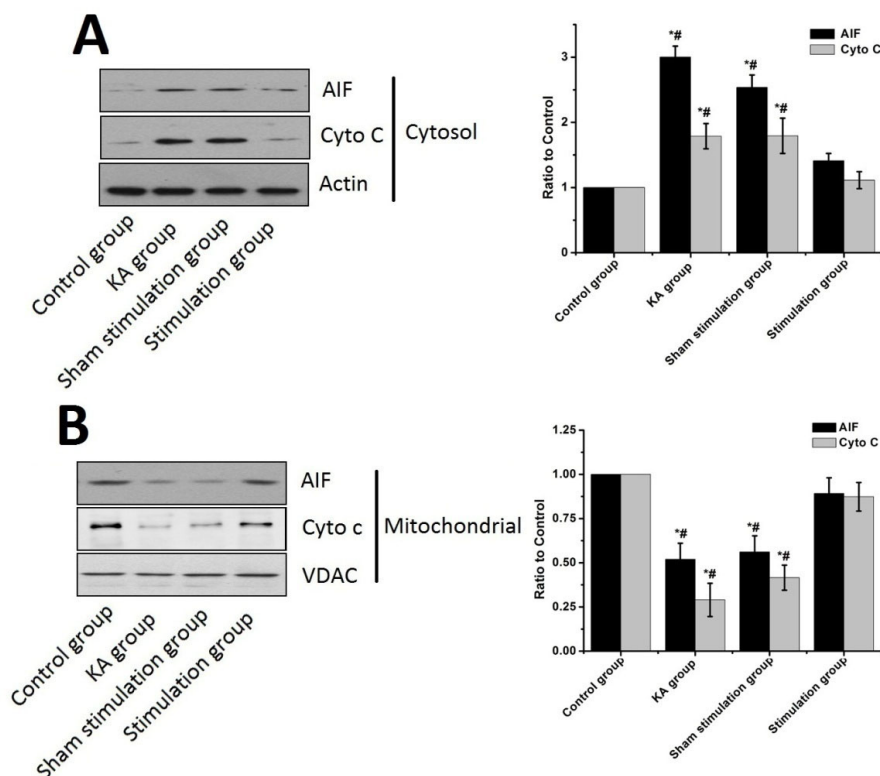


Figure 3: Effect of ANT stimulation on the mitochondrial release of AIF and cyto c release following seizures. (A) Representative western blots from ipsilateral hippocampal cytosolic fraction in control group, KA group, sham stimulation group and stimulation group, respectively. Compared to KA group or sham stimulation group, ANT stimulation reduced the presence of AIF and cyto c in the cytosolic fraction. (B) Representative western blots from ipsilateral hippocampal mitochondrial fraction in control group, KA group, sham stimulation group and stimulation group, respectively.

Histogram data are shown as mean \pm SD from three independent experiments

* $P < 0.05$, compared to control group; # $P < 0.05$, compared to ANT stimulation group

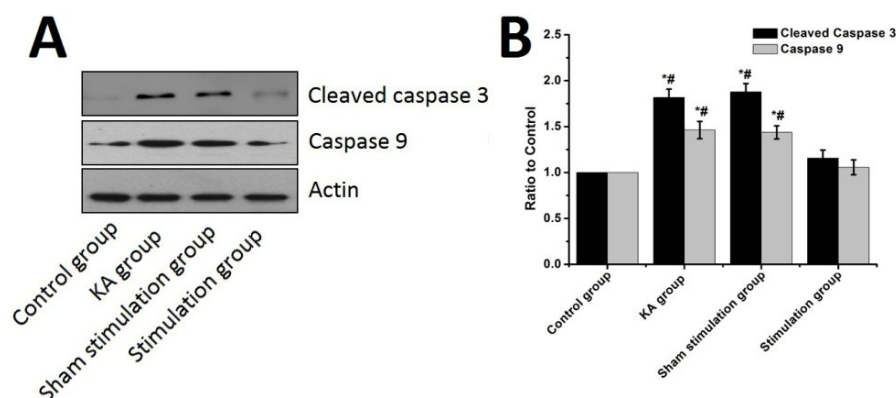


Figure 4: Effect of ANT stimulation on caspase activation following seizures. (A) Caspase-9 and cleaved caspase-3 activity was detected in control group, KA group, sham stimulation group and stimulation group, respectively. Compared to KA group or sham stimulation group, ANT stimulation reduced the levels of caspase-9 and cleaved caspase-3. (B) Histogram data are shown as mean \pm SD from three independent experiments.

* $P < 0.05$, compared to control group; # $P < 0.05$, compared to ANT stimulation group

seizures of rats. In addition, ANT stimulation reduced the release of cyto c and AIF induced by seizures while inhibiting the activated caspase-9 and caspase-3. Our results suggested that the neuroprotection of ANT stimulation against hippocampal cell death after KA treatment is mediated through the inhibition of caspase-dependent (cyto c release

and the subsequent cleavage of caspase-) and caspase-independent (nuclear translocation of AIF) apoptosis pathways, which may be an important mechanism for ANT stimulation in epilepsy treatment.

Apoptosis is a complex process in which individual cells undergo self-

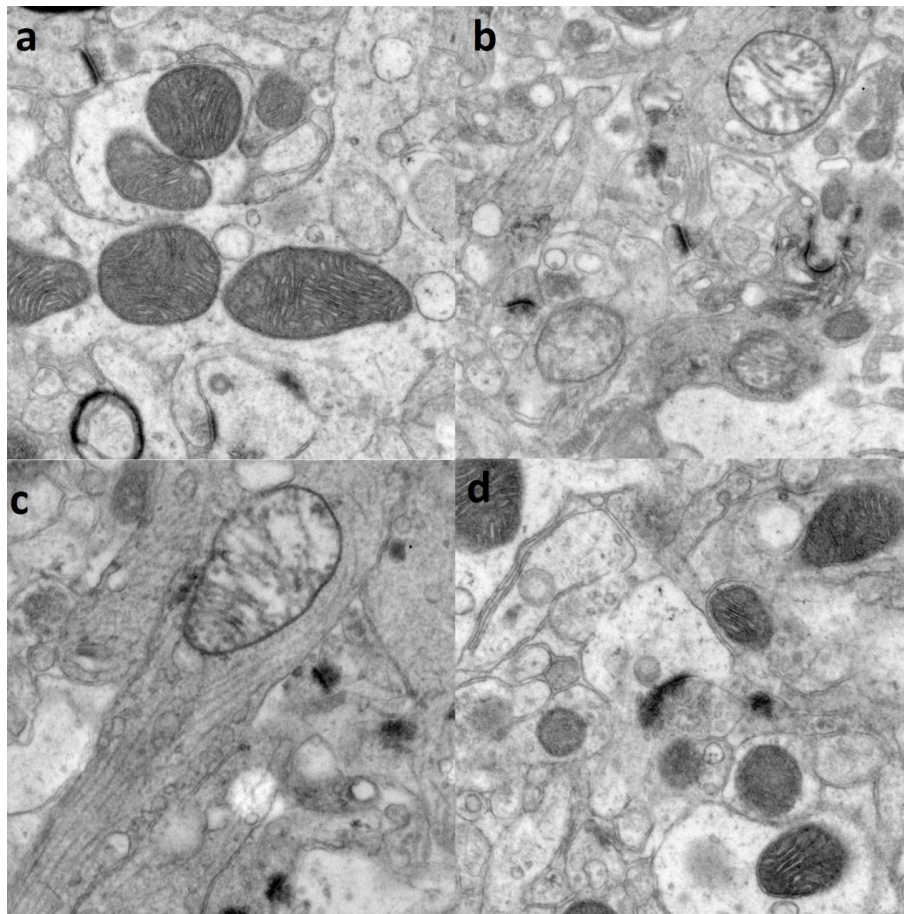


Figure 5: Electron micrographs showing swelling and ultrastructural damage to mitochondria in KA-treated rats, which was abrogated by ANT stimulation. (a) control group, (b) KA group, (c) sham stimulation group, (d) stimulation group.

destruction without inducing an inflammatory response. Several groups have reported that seizures induced by KA in adult rats caused ipsilateral death of CA3 neurons in the hippocampus [16,17]. There is increasing evidence that neuronal cell death induced by seizures activates mitochondria pathway [18]. Mitochondria play an important role in cell death by realys of signals for both caspase-dependent and caspase-independent death pathways [19]. The mitochondrion subserve important cellular functions to control of neuronal cell death, which is critical site for the initiation and/or reinforcement of cell death pathways [20]. It is well known that the release of cyto c by mitochondria is directly linked to induction of apoptosis [21]. Cyto c could interact with Apaf-1 and Datp, forming the apoptosome and causing the activation of caspase-9, which activates caspase-3 leading to chromation condensation, DNA fragmentation and apoptotic cell death in the caspse-dependent apoptosis pathways [15,22]. Previous reports demonstrated that the levels of cyto c, caspase-9 and cleaved caspase-3 were elevated and inhibition of the mitochondria release of cyto c could attenuate apoptosis after seizures [23,24]. In the current study, we also observed the similar results of release cyto c from mitochondria and activation of caspase-9 and caspase-3 following seizures and in contrast, decrease of motochondria release of cyto c and inhibitionof caspase-9 and caspase-3 activation by ANT stimulation. These results suggested that the ANT stimulation could exert neuroprotective effects against neuronal death by inhibiting mitochondrial caspase-dependent apoptosis pathways activation following KA-induced seizures.

Recent reports showed that AIF is involved in seizure induced apoptotic cell death in the mitochondrial caspase-independent apoptosis pathways [19]. AIF is a key factor in caspase-independent death pathway when it translocates from mitochondria to the nucleus, where it results in peripheral chormatin condensation and high-molecular weight DNA fragmentation that culminates in cell death [25,26]. Heo et al. reported that intra-hippocampal KA injection results in AIF translocation from mitochondria to nucleus in mouse hippocampal neurons as well as DNA cleavage [27]. Our study observed the similar results that the release of AIF from mitochondria following seizures, indicating that AIF could actually contribute to cell death induced by seizures. In addition, it has been reported that neurons with reduced AIF are protected against excitotoxicity specifically through the activation of kainic acid receptors and Hq mice with a mutation reducing AIF expression by 80% [28] have significantly less hippocampal damage than wild-type littermates in an in vivo seizure model induced by kainic treatment [19]. Meanwhile, we also found that ANT stimulation decrease mitochondria release of AIF illustrating that ANT stimulation could exert neuroprotective effects against neuronal death by inhibiting mitochondrial caspase-independent apoptosis pathways activation following KA-induced seizures.

However, the mechanism responsible of ANT stimulation to prevent mitochondria release of cyto c and AIF remains obscure. Recent work has suggested that mitochondria are highly dynamic organelles and the

changes in mitochondrial morphology could affect the susceptibility of cell to apoptosis in epilepsy [29]. The ultrastructural damage of mitochondria in the hippocampus may be a key to the pathogenesis of status epilepticus [30]. The mitochondrial dysfunction could cause the release of cyto c and AIF. The electron microscopic examination of mitochondrial ultrastructure in hippocampal neurons in this study demonstrated that swelling and disruption of mitochondrion in KA and sham stimulation group with ANT stimulation maintain most mitochondrial structural integrity in stimulation group. One possible mechanism of ANT stimulation to prevent mitochondria release of cyto c and AIF is that ANT stimulation mitigates KA-induced neuronal apoptosis by limiting mitochondrial injury.

Conclusion

We provided evidence that ANT stimulation attenuates neuronal death in hippocampus induced by seizures and exerts neuroprotective effect on hippocampal neurons. The underlying mechanisms may be mediated by inhibiting mitochondrial caspase-dependent (cyto c release and the subsequent cleavage of caspase-) and caspase-independent (nuclear translocation of AIF) apoptosis pathways.

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