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# Genetic determinants of rates of cognitive decline in preclinical Alzheimer's Disease

This thesis is presented for the degree of

**Doctor of Philosophy** 

**Tenielle Louise Sandra Porter** 

Edith Cowan University

School of Medical and Health Sciences

2018

## Genetic determinants of rates of cognitive decline in

## preclinical Alzheimer's Disease

**Tenielle Louise Sandra Porter** 

**Doctoral Thesis** 

April 2018

School of Medical and Health Sciences

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### Abstract

#### Background

In 2015 the number of people worldwide living with Dementia was 46.8 million, with approximately 50-75% of these cases being clinically defined as Alzheimer's disease (AD). Despite extensive efforts, clinical trials have so far failed to yield a treatment that successfully addresses the underlying cause of AD. This lack of treatment has been suggested, in part, to be a result of late stage of intervention in current clinical trial design. For this reason, greater focus has been placed on preclinical trials and in turn both the identification of individuals at-risk for AD and, amongst these, those that are expected to decline over the course of a trial. While brain imaging to determine A $\beta$ -amyloid burden has utility in identifying individuals with preclinical AD, further work needs to be conducted to determine what influences rates of change during these early disease stages. Of particular focus is the rate of decline in cognitive performance, as it is the primary outcome measure of efficacy in clinical trials. A number of genetic variants have been associated with cognitive performance, however additional research needs to be conducted to accurately understand the influence that genetic variation has on cognition in preclinical AD.

#### Aims

Initially the aim of this thesis was to assess the combined genetic influence of established AD risk genetic variants on preclinical cognitive performance, specifically using AD-risk effect-size weighted polygenic risk scores (PRSs) (Chapter 2). It was then aimed to evaluate the effects on cognitive rates of change in preclinical AD of genes with *a priori* evidence for association with cognition, both individually (Chapter

3) and then when combined (Chapter 4). The results of the preceding chapters informed the final aim which was to determine a novel method of weighting individual variants in genes associated with AD-risk and/or cognition, for use in a genetic risk score that would improve the prediction of preclinical cognitive rates of change (Chapter 5).

#### Methods

All studies presented in this thesis utilised data from the highly characterised Australian Imaging, Biomarkers and Lifestyle Study of Aging (AIBL). The AIBL study is a longitudinal cohort study collecting data at 18-monthly intervals, currently consisting of 7.5 years of follow up. Individuals investigated in this thesis had been Positron Emission Tomography (PET) imaged to determine neocortical amyloid burden. Further, all individuals were classified as  $A\beta^{high}$  or  $A\beta^{low}$  based on tracer specific cut offs. In addition, a subset of these samples underwent lumbar puncture for CSF collection at the study baseline, and  $A\beta_{42}$ , total-tau and phospho-tau were quantified. Finally, based on the AIBL neuropsychological test battery, three cognitive composites previously developed were calculated for all participants. The cognitive composites investigated were; verbal episodic memory, a statistically driven global cognition composite, and the Pre-Alzheimer's Cognitive Composite.

The AD-risk weighted PRS (Chapter 2) consisted of 22 genetic variants associated with AD classification, and was calculated by weighting individual variants based on their previously published associations with risk for AD. A statistically derived Cognitive Genetic Risk Profile (*Cog-GRP*), specifically driven by verbal episodic memory, was developed using a decision tree analysis (Chapter 4). Finally, a 27 genetic variant cognition weighted PRS (*cw*PRS), was developed and tested in a preclinical AD sample

(Chapter 5). For the *cw*PRS, effect sizes for decline in a verbal episodic memory were determined individually for all variants in a reference sample. The resulting effect sizes were then used to calculate the *cw*PRS for each participant in a test sample (Chapter 5). For both the AD-risk weighted PRS (Chapter 2) and the *cw*PRS (Chapter 5), PRS calculations were conducted with both the inclusion and exclusion of the major genetic risk factor for, Apolipoprotein E (*APOE*).

In all studies, linear mixed models were used to investigate associations between genetic factors, independent or in combination, and longitudinal rates of cognitive performance.

#### Results

In CN older adults the AD-risk weighted PRS, both including and excluding *APOE*, was positively correlated with brain and blood biomarkers, specifically; brain A $\beta$  burden, CSF total-tau and phospho-tau (Chapter 2). When investigating cognitive performance, specifically in CN A $\beta^{high}$  participants, significant associations with baseline and longitudinal cognition were only observed in the AD-risk weighted PRS with *APOE* (Chapter 2).

When investigating gene variants previously reported to influence cognition, in CN  $A\beta^{high}$  participants, no independent associations were observed for any variant (Chapter 3). However, in the same sample, after interaction with *APOE*  $\epsilon$ 4, significant associations were observed for variants in the Kidney Brain Expressed Protein (*KIBRA*) and Spondin-1 (*SPON1*) genes (Chapter 3). The combination of variants investigated in Chapter 3, with additional variants, resulted in the development of the *Cog-GRP* 

(Chapter 4). The *Cog-GRP* was able to delineate four groups: *APOE*  $\epsilon$ 4+ Risk, *APOE*  $\epsilon$ 4+ Resilient, *APOE*  $\epsilon$ 4- Risk, *APOE*  $\epsilon$ 4- Resilient, with the  $\epsilon$ 4+ Risk group reporting significantly faster decline in cognition than all other groups (Chapter 4).

Finally, a PRS encompassing a combination of AD-risk genes (Chapter 2) and cognitive-risk genes (Chapters 3 and 4), weighted by episodic memory (*cw*PRS), was reported to be associated with preclinical longitudinal cognitive performance (Chapter 5). Further, these associations were observed irrespective of the presence or absence of *APOE* in the calculation of the *cw*PRS (Chapter 5).

#### Conclusions

The work presented in this thesis provides an in depth investigation of genetic influences in preclinical AD, particularly on cognitive performance. Importantly, it supports the hypothesis that there is are differences between the genetic architectures of AD-risk and AD progression. The results presented here support the use of combinatory approaches when investigating genetic influence. Finally, reported here is a novel method for PRS weighting, with the ability to predict preclinical cognitive performance in the presence and absence of *APOE*. Further investigation is required in cohorts with comparable data to the AIBL study, to validate the methods explored in this thesis, allowing for their eventual use in a clinical setting.

## Acknowledgements

The work presented in this thesis is dedicated to my poppy (Angus Hugh Porter), I know how proud you would have been to see this.

This thesis would not have been possible without the help and support of my supervisors, colleagues, friends and family.

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## Statement of Contribution of others

The thesis "Genetic determinants of rates of cognitive decline in preclinical Alzheimer's Disease" is submitted as a series of publications for the Degree of Doctor of Philosophy, by Tenielle Porter, School of Medical and Health Sciences, Edith Cowan University, 2018. Below is a statement of Contribution of others for the publications included.

#### **Publication Reference**

#### **Contribution by Tenielle Porter (%)**

#### **Chapter 1**

Porter, T., Gozt, A.K., Mastaglia, F.L., Laws S.M. Genetics of TP 51% (literature review, Neurodegenerative Disease: Role of genetics in Alzheimer's and Parkinson's Disease. In: Martins, RN and Brenan, C (eds) Factors that Modify Risk of Alzheimer's Disease: The role of Genes, Diabetes, Diet, Exercise and Hormones (2018) Wiley, UK. Submitted 21.03.2018

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#### Chapter 2

Porter T, Burnham SC, Milicic L, Savage G, Maruff P, Lim YY, Li QX, Ames D, Masters CL, Rainey-Smith SR, Rowe CC, Salvado O, Groth D, Verdile G, Villemagne V, Laws SM., for the AIBL research group. Utility of an Alzheimer's risk-weighted polygenic risk score for predicting rates of cognitive decline in preclinical Alzheimer's disease: a prospective longitudinal study. Submitted 03.04.2018; Alzheimer's Research and Therapy

#### TP 60% (literature review, study design, genetic data collection, data analysis, manuscript preparation)

#### Chapter 3

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**Porter T,** Burnham SC, Milicic L, Savage G, Maruff P, Lim YY, Ames D, Masters CL, Rainey-Smith SR, Rowe CC, Salvado O, Groth D, Verdile G, Villemagne V, Laws SM., for the AIBL research group. *COMT* val158met is not associated with Aβ-amyloid and *APOE* ε4 related cognitive decline in cognitively normal older adults. *Final draft for submission; Neurobiology of Aging (Genetic Report)\** 

**Porter T,** Burnham SC, Milicic L, Savage G, Maruff P, Lim YY, Ames D, Masters CL, Rainey-Smith SR, Rowe CC, Salvado O, Groth D, Verdile G, Villemagne V, Laws SM., for the AIBL research group. Klotho allele status is not associated with  $A\beta$  and *APOE*  $\epsilon$ 4 related cognitive decline in preclinical Alzheimer's disease. *Final draft for submission; Neurobiology of Aging (Genetic Report)\**  *TP 65%* (literature review, study design, genetic data collection, data analysis, manuscript preparation)

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*TP 65%* (literature review, study design, genetic data collection, data analysis, manuscript preparation)

#### **Chapter 4**

**Porter T,** Villemagne V, Savage G, Milicic L, Lim YY, Maruff P, Masters CL, Ames D, Bush AI, Rainey-Smith SR, Rowe CC, Groth D, Taddei K, Martins RN, Verdile G, Burnham SC, Laws SM., for the AIBL research group. Cognitive Gene Risk Profile for the Prediction of Cognitive Decline in Presymptomatic Alzheimer's Disease. *Personalized Medicine in Psychiatry March 2018* 7-8:14-20. *doi:10.1016/j.pmip.2018.03.001* 

*TP 60%* (literature review, study design, genetic data collection, data analysis, manuscript preparation)

#### Chapter 5

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*TP 65%* (literature review, study design, genetic data collection, data analysis, manuscript preparation)

I, Tenielle Porter, contributed to the above listed publications at the stated level.

Signed:

Date: 30/04/2018

\* At time of submission of the thesis these manuscripts were not yet submitted, but are hereby certified as being at a stage approved by co-authors to be ready for submission for peer-review

 Date:

 Assoc. Prof. Simon M Laws (Principle Supervisor and Corresponding Author)

I, as a co-author, endorse that this level of contribution by the candidate indicated above is appropriate.

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## **Publications**

#### Accepted publications contributing to the thesis

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**Porter T,** Villemagne V, Savage G, Milicic L, Lim YY, Maruff P, Masters CL, Ames D, Bush AI, Rainey-Smith SR, Rowe CC, Groth D, Taddei K, Martins RN, Verdile G, Burnham SC, Laws SM., for the AIBL research group. Cognitive Gene Risk Profile for the Prediction of Cognitive Decline in Presymptomatic Alzheimer's Disease. *Personalized Medicine in Psychiatry 2018 7-8:14-20. doi:10.1016/j.pmip.2018.03.001* 

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**Porter T,** Bharadwaj P, Groth D, Paxman A, Laws SM, Martins RN, Verdile G. The effects of latrepirdine on amyloid beta aggregation and toxicity. *J Alzheimers Dis 2016 50(3): 895-905. doi: 10.3233/JAD-150790. PMID: 26836170* 

Lim YY Laws SM, Villemagne VL, Pietrzak RH, **Porter T,** Ames D, Fowler C, Rainey-Smith SR, Martins RN, Salvado O, Bourgeat P, Rowe CC, Masters CL, Maruff P. Aβ-related memory decline in *APOE* ε4 non-carriers: Implications for Alzheimer's disease. *Neurology 2016 86(17):1635-42. doi: 10.1212/WNL.00000000002604. PMID: 27029632* 

Pietrzak RH\* Laws SM\*, Lim YY, Bender SJ, **Porter T,** Doecke J, Ames, D, Fowler C, Masters, CL, Milicic L, Rainey-Smith SR, Villemagne, VL, Rowe, CC, Martins, RN, Maruff, P. Plasma cortisol, amyloid-beta, and cognitive decline in preclinical Alzheimer's disease: A 6-year prospective cohort study. *Biol Psychiatry: Cog Neurosci and Neuroimaging 2017 2(1):45-52. doi: 10.1016/j.bpsc.2016.08.006. PMID: 29560886* 

Lim YY\*, Rainey-Smith S\*, Lim Y, Laws SM, Gupta V, **Porter T,** Bourgeat P, Ames D, Fowler C, Salvado O, Villemagne VL, Rowe CC, Masters CL, Zhou XF, Martins RN, Maruff P. *BDNF* Val66Met in preclinical Alzheimer's disease is associated with short-term changes in episodic memory and hippocampal volume but not serum mBDNF. *International Psychogeriatrics 2017 29(11):1825-1834. doi: 10.1017/S1041610217001284. PMID: 28720165* 

Laws SM, Gaskin S, Woodfield A, Srikanth V, Bruce D, Fraser PE, **Porter T**, Newsholme P, Wijesekara N, Burnham S, Doré V, Li QX, Maruff P, Masters CL, Rainey-Smith SR, Rowe CC, Salvado O, Villemagne VL, Martins RN, Verdile G. Insulin resistance is associated with reductions in specific cognitive domains and increases in CSF tau in cognitively normal adults. *Scientific Reports 2017 7(1):9766. doi: 10.1038/s41598-017-09577-4. PMID: 28852028* 

Hollands S, Lim, YY, Laws, SM, Villemagne, VL, Pietrzak, RH, Harrington K, **Porter T,** Snyder P, Ames, D, Fowler C, Rainey-Smith SR, Martins, RN, Salvado O, Robertson J, Rowe CC, Masters, CL, Maruff, P., for the AIBL research group. *APOE* ɛ4 genotype, amyloid and clinical disease progression in cognitively normal older adults. *J. Alzheimer Dis 2017* 57(2):411-422. *doi:* 10.3233/JAD-161019. *PMID:* 27029632

Rainey-Smith SR, Mazzucchelli GN, Villemagne VL, Brown BM, **Porter T,** Weinborn M, Bucks RS, Milicic L, Sohrabi HR, Taddei K, Ames D, Maruff P, Masters CL, Rowe CC, Salvado O, Martins RN, Laws SM, for the AIBL research group. Genetic variation in Aquaporin-4 moderates the relationship between sleep and brain Aβ-amyloid burden. *Transl Psychiatry 2018 8(1):47. doi: 10.1038/s41398-018-0094-x. PMID: 29479071* 

#### International conference proceedings

**Porter T,** Garin S, Wilson AC, Verdile G, Groth D, Villemagne VL, Ames D, Bush A, Ellis KA, Macaulay SL, Masters CL, Rainey-Smith SR, Rembach A, Rowe CC, Taddei K, Martins RN, Laws SM, for the AIBL Research Group. Genetic analysis of the steroidogenesis pathway: Associations with Alzheimer's disease risk and related phenotypes. *Poster presentation to the Alzheimer's Association International Conference for Alzheimer's disease, July 2014, Copenhagen, Denmark*  **Porter T,** Verdile G, Li QX, Dore V, Lim YY, Villemagne VL, Ames D, Bush A, Ellis KA, Groth D, Macaulay SL, Maruff P, Masters CL, Rainey-Smith SR, Rembach A, Rowe CC, Taddei K, Wilson AC, Martins RN, Laws SM, for the AIBL Research Group Tau haplotypes and their association with Alzheimer's disease endophenotypes in the Australian Imaging Biomarkers and Lifestyle Study of Aging. *Poster presentation to the Alzheimer's disease Parkinson's disease Conference for Alzheimer's disease, March 2015, Nice, France* 

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## List of Abbreviations

AA	Alzheimer's Assocation
Αβ	Amyloid β
ABCA7	ATP Binding Cassette Subfamily A Member 7 Gene
AD	Alzheimer's Disease
ADAD	Autosomal Dominant AD
ADPD	Autosomal Dominant PD
ADAM10	A Disintegrin And Metalloprotease 10 Gene
ADAS-COG	Alzheimer's Disease Assessment Scale – Cognitive Section
ADNI	Alzheimer's Disease Neuroimaging Initiative
ADRDA	Alzheimer's Disease and Related Disorders Association
AIBL	Australian Imaging, Biomarkers and Lifestyle
AICD	APP Intracellular Domain
ANOVA	Analysis of Variance
APOE	Apolipoprotein E Gene
АроЕ	Apolipoprotein E Protein
ApoJ	Apolipoprotein J Protein/Clusterin Protein
AP2	Adaptor Protein Complex 2
APP	Amyloid Precursor Protein Gene
APP	Amyloid Precursor Protein
ATP	Adenosine Triphosphate
ATP13A2	Probable Cation-Transporting ATPase 13A2 Gene
AUC	Area Under the Curve
BACE-1	β-Site APP Cleaving Enzyme

BDNF	Brain Derived Neurotropic Factor Gene
BeCKeT	Before the Centiloid Kernel Transformation
BINI	Bridging Integrator 1 Gene
Bin1	Bridging Integrator 1 Protein
BNT	Boston Naming Test
BST1	Bone Marrow Stromal Cell Antigen 1
C99	C-Terminal of APP
CAA	Cerebral Amyloid Angiopathy
CASS4	Cas Scaffolding Protein Family Member 4 Gene
CD2AP	CD2-Associated Protein Gene
CD33	Myeloid Cell Surface Antigen CD33 Gene
CELF1	CUGBP, Elav-Like Family Member 1 Gene
CDR	Clinical Dementia Rating
CDR <sub>SB</sub>	CDR Sum of Boxes
CHMP2B	Charged Multivesicular Body Protein 2B Gene
CI	Confidence Interval
CLU	Clusterin Gene
CLU	Clusterin Protein
CN	Cognitively Normal
CNS	Central Nervous System
Cog-GRP	Cognitive Genetic Risk Profile
COMT	Catechol-O-Methyltransferase Gene
COWA	Controlled Oral Word Association Test
CR1	Complement Receptor 1 Gene
CSF	Cerebrospinal Fluid

CSMD1	CUB and Sushi Multiple Domains 1
CVLT-II	California Verbal Learning Test-Second Edition
CVLT <sub>FP</sub> /CVLT <sub>LDFR</sub>	CVLT False Positives and Long Delay Free Recall
cwPRS	Cognitive Weighted PRS
DGKQ	Diacylglycerol Kinase Theta Gene
DJ-1	Protein Deglycase DJ-1 Gene
D-KEFS	Delis-Kaplan Executive Function System
DNA	Deoxyribonucleic Acid
DSM-5	Diagnostic and Statistical Manual of Mental Disorders 5th
Edition	
ECM	Extracellular Matrix
EEG	Electroencephalography
EIF4G1	Eukaryotic Translation Initiation Factor 4 Gamma 1 Gene
ELISA	Enzyme-Linked Immunosorbent Assays
EOAD	Early Onset AD
EOFAD	Early Onset Familial AD
EOPD	Early Onset PD
EPHA1	EPH Receptor A1 Gene
FBXO7	F-Box Protein 7 Gene
FDG	<sup>18</sup> F-Fluorodeoxyglucose
FDR	False Discovery Rate
FERMT2	Fermitin Family Member 2 Gene
fMRI	Functional MRI
FSIQ	WAIS-III Full Scale Intelligence Quotient
FTLD	Frontotemporal Lobar Degeneration

GAK	Cyclin G-Associated Kinase Gene
GBA	Glucocerebroside Gene
GERARD1	Genetic and Environment Risk in AD Consortium 1
GDS	Geriatric Depression Scale
GIGYF2	PERQ Amino Acid-Rich with GYF Domain-Containing
Protein 2 Gene	
GSK3	Glycogen Synthase Kinase 3
GWA	Genome Wide Association
GWAS	Genome-Wide Association Study
HABS	Harvard Aging Brain Study
HLA	Human Leukocyte Antigen
HLA-DRA	HLA Class II Histocompatibility Antigen, DR Alpha Chain
Gene	
HLA-DRB1	Major Histocompatibility Complex Class II, DR Beta 1 Gene
HLA- DRB5	Major Histocompatibility Complex Class II, DR Beta 5 Gene
HLA-DQB1	Major Histocompatibility Complex, Class II, DQ Beta 1 Gene
HTRA2	HtrA Serine Peptidase 2 Gene
IGAP	International Genomics of Alzheimer's Project
IGF	Insulin-Like Growth Factor
IL-1β	Interleukin 1 β
IL-6	Interleukin 6
IL-8	Interleukin 8
IL-10	Interleukin 10
IL-18	Interleukin 18
INPP5D	Inositol Polyphosphate-5-Phosphatase Gene
IQ	Intelligence Quotient
----------	--
JPD	Juvenile PD
KIBRA	Kidney Brain Expressed Protein Gene
KIBRA	Kidney Brain Expressed Protein
KL	Klotho Gene
LDL	Low Density Lipoprotein
LDLR	Low Density Lipoprotein Receptor
LME	Linear Mixed Effects
LMI/LMII	Logical Memory I and II
LOAD	Late Onset AD
LOPD	Late Onset PD
LRP1	Low Density Lipoprotein Receptor-Related Protein-1
LRRK2	Leucine-Rich Repeat Kinase 2 Gene
MAF	Minor Allele Frequency
МАРК	Mitogen-Activated Protein Kinase
MAPT	Microtubule-Associated Protein Tau Gene
MCI	Mild Cognitive Impairment
MeDi	Mediterranean Diet
MEF2C	Myocyte Enhancer Factor 2C Gene
МНС	Major Histocompatibility Complex
MMSE	Mini-Mental State Examination
MR	Magnetic Resonance
MRI	Magnetic Resonance Imaging
mRNA	Messenger Ribonucleic Acid
MS4A	Membrane Spanning 4 Domains, Subfamily A

MS4A4A	Membrane Spanning 4 Domains, A4A Gene
MS4A6A	Membrane Spanning 4 Domains, A6A Gene
mtDNA	Mitochondrial DNA
NADH	Nicotinamide Adenine Dinucleotide
NART	North American National Adult Reading Test
NCD	Neurocognitive Disorder
ND3	NADH Dehydrogenase 3
NDPK	Nucleoside Diphosphate Kinase
NFT	Neurofibrillary Tangles
NGS	Next Generation Sequencing
NIA	National Institute on Aging
NINCDS	National Institute of Neurological and Communicative
	Disorders and Stroke
NME8	NME/NM23 Family Member 8 Gene
OR	Odd Ratio
PACC	Pre-Alzheimer's Cognitive Composite
PCR	Polymerase Chain Reaction
PET	Positron Emission Tomography
PD	Parkinson's Disease
PiB	<sup>11</sup> C-Pittsburgh Compound B
PICALM	Phosphatidylinositol Binding Clathrin Assembly Lymphoid
	Myeloid Protein Gene
PINK-1	PTEN-Induced Putative Kinase 1 Gene
РКСζ	Protein Kinase C Isoform ζ
ΡΚΜζ	Protein Kinase M Isoform ζ

PLA2G6	Phospholipase A2 Group 6 Gene
PLD3	Phospholipase D3 Gene
PLOSL	Polycystic Lipomembranous Osteodysplasia with Sclerosing
	Leukoencephalopathy
PSEN1	Presenilin 1 Gene
PSEN2	Presenilin 2 Gene
PRKN	Parkin Gene
PRS	Polygenic Risk Score
РТС	Premature Termination Codon
PTK2B	Protein Tyrosine Kinase 2β Gene
RA	Rheumatoid Arthritis
RAVLT	Rey Auditory Verbal Learning Test
RCFT	Rey Complex Figure Test
RIN3-	Rab Interactor 3 Gene
RIT2	Ras Like Without CAAX 2 Gene
ROC	Receiver Operating Characteristic Curve
ROCF	Rey-Osterrieth Complex Figure Test
SAD	Sporadic AD
SETD1A	SET Domain Containing 1A
SD	Standard Deviation
SLC24A4	Solute Carrier Family 24 Sodium/Potassium/Calcium
	Exchanger, Member 4 Gene
SLE	Systemic Lupus Erythematosus
SNARE	SNAP (Soluble NSF(N-ethylmaleimide-sensitive factor)
	Attachment Protein) Receptor

SNCA	α-Synuclein Gene
SNP	Single Nucleotide Polymorphism
SORL1	Sortilin-Related Receptor LDLR Class A Repeats Containing
Gene	
SPON1	Spondin 1 Gene
SUVR	Standardized Uptake Value
SUVR	Standardized Uptake Value Ratios
ΤΝΓα	Tumor Necrosis Factor α
TREM2	Triggering Receptor Expressed on Myeloid Cells 2 Gene
UCHL1	Ubiquitin Carboxy-Terminal Hydrolase L1 Gene
UPS	Ubiquitin Proteasome System
USD	United States Dollar
VAMP2	Vesicle Associated Membrane Protein 2
VLDL	Very Low Density Lipoprotein
VPS35	Vacuolar Protein Sorting 35 Gene
WTAR	Wechsler Test of Adult Reading
WAIS-III	Wechsler Adult Intelligence Scale-Third Edition
WMS	Wechsler Memory Scale
WWC1	WW Domain-Containing Protein 1
ZCWPW1	Zinc Finger, CW Type With PWWP Domain 1 Gene

# Chapter 1

Please note that this online copy of the thesis does not contain the complete version of Chapter 1

# **CHAPTER 1: General Introduction**

# 1.1 Dementia and Alzheimer's Disease

In 2015 there were almost 900 million people over the age of 60 [1]. As the global population continues to age there is a growing focus on age-associated diseases such as Dementia. Dementia is described generally as progressive decline in a patient's cognitive functioning greater than what is expected to occur in normal aging. The term "Dementia" does not define a single disease, but describes several diseases typified by the detrimental changes in brain function. Domains in which these changes occur include language, memory, perception, personality and cognitive skills [2].

In 2015 46.8 million people globally were living with dementia, with this number expected to double every 20 years [1]. It was estimated that there are 9.9 million new cases a year, or one new case every 3.2 seconds [1]. In Australia alone there are approximately 410,000 people with Dementia [3]. Further, in 2015 dementia was the second leading cause of death in Australia [4].

In addition to the human cost of dementia there is a significant economic cost. The global cost of dementia increased by USD\$214 billion dollars between 2010 and 2015, rising from USD\$604 to USD\$818 billion [1]. These costs include social care, professional and volunteer, and medical care [1]. It has been estimated that dementia will cost USD\$1 trillion by 2018 [1]. It has been predicted that by delaying the onset of dementia by 5 years, the number of people with the condition in Australia could be

reduced by around one third [5]. Additionally, in America, a study was conducted that reported delaying the onset of Alzheimer's disease (AD) by 5 years would result in an economic saving of USD\$935 billion over 10 years [6].

It has been reported that 50-75% of dementia cases are clinically defined as Alzheimer's Disease (AD) [2, 7]. AD is characterised by neuronal loss, abnormal protein deposition in the brain, and the deterioration of both cognitive function and the ability to perform activities of daily living.

#### **1.1.1 Pathological Features**

The pathological features of AD can be divided into macroscopic structural changes to the brain, and the presence in increased concentrations of extra- and intracellular fibrous protein deposits. Importantly, it is generally increased concentrations of these proteins, due to inefficient clearance or over production, which results in pathogenicity [8]. It has previously been observed that low concentrations have a non-pathogenic effect in cognitively normal older adults (CN) [9].

#### 1.1.1.1 Cerebral Atrophy

The major macroscopic hallmark of AD is the progressive loss of brain volume, termed cerebral atrophy (Figure 1.1.1). The brain regions most significantly affected by atrophy have been reported to change as the disease develops [10, 11]. It has been observed that hippocampal atrophy occurs earliest in the disease process, followed by atrophy of the temporal parietal lobes, and in the late disease stages the frontal lobe [10]. Significant atrophy of the medial parietal lobe has been observed throughout the disease [10, 12]. A number of studies have reported hippocampal volume significantly

reduced in Mild Cognitive Impairment (MCI) [13] and AD [13] when compared to cognitively normal (CN) individuals. Additionally, longitudinal rates of hippocampal atrophy have been associated with progression to AD (from MCI) [14], and classification of AD [15].





Representation of the brain of a cognitively normal older adult when compared to that of an Alzheimer's patient, displaying the gross structural changes associated with the disease. Image sourced from the BrightFocus Foundation (2000).

#### 1.1.1.2 Cerebral Amyloid Angiopathy

The accumulation of Amyloid Beta ( $A\beta$ -amyloid; hereby abbreviated to  $A\beta$ ) in the cerebral and meningeal blood vessels, Cerebral Amyloid Angiopathy (CAA), is a prominent hallmark in AD (Figure 1.1.2, [16]). The most common, and most severely affected, brain region affected by CAA is the occipital lobe, followed by the parietal, frontal and temporal [17-19]. Sporadic age-related CAA has been observed in the brains of healthy elderly individuals [20, 21], however it is more prevalent in those of AD patients [21, 22]. Further, CAA has been reported occurring more frequently in pathology confirmed AD than in other diseases leading to dementia [18, 21].

### 1.1.1.3 Senile Amyloid Plaques

Senile amyloid plaques, or neuritic plaques, are the most commonly associated pathological hallmark of AD (Figure 1.1.3). Senile amyloid plaques have been used in the post-mortem diagnosis of AD, due to their occurrence being uncommon in other neurodegenerative disorders. The initial deposition of amyloid plaques occurs through the frontal, parietal, temporal, or occipital neocortex [24]. These areas are followed by the entorhinal region and addition subcortical regions, and eventually the cerebellum and brainstem [24]. A number of protein components make up amyloid plaques including; a core of insoluble A $\beta$  [25, 26], surrounded by apolipoprotein E (ApoE),  $\alpha$ 2-macroglobulin, interleukins,  $\alpha$ 2-macroglobulin receptor, and low-density lipoprotein receptor-related protein [27-30].



### Figure 1.1.2. Cerebral Amyloid Angiopathy

(A) Mild, (B) moderate, and (C) severe cerebral amyloid angiopathy (CAA) in the meningeal vessels of the temporal lobe, immunohistochemistry with anti-amyloid antibody 4G8, scale bar: 50 μm. Image sourced from [23].



## Figure 1.1.3. Senile Amyloid Plaques

Advanced stage AD patient's temporal cortex with senile amyloid plaques evident (brown), immunochemistry with anti-amyloid antibody 4G8,  $100 \times$  magnification. Image sourced from [31].

#### 1.1.1.3.1 Amyloid Beta

AB, the major component of senile amyloid plaques, is a small 4-5kDa protein found in the brain, CSF and blood of CN older adults and AD patients [26]. Proteolytic cleavage of the amyloid precursor protein (APP), a 110-135kDa transmembrane glycoprotein, results in the generation of A $\beta$  and other peptides. The cleavage of APP to produce A $\beta$  is via the amyloidogenic pathway (Figure 1.1.4A, E-G), which occurs in healthy CN individuals but is favoured in AD. APP is first cleaved at the N-terminus of the A $\beta$  domain by  $\beta$ -site APP cleaving enzyme (BACE-1) (Figure 1.1.4E), producing soluble APPB and the C-terminal of APP (C99; Figure 1.1.4F). C99 is then cleaved within the transmembrane domain by  $\gamma$ -secretase, releasing AB and the APP intracellular domain (AICD; Figure 1.1.4F, G). Cleavage of APP via the nonamyloidogenic pathway involves cleavage within the A $\beta$  domain by  $\alpha$ -secretase, precluding AB production (Figure 1.1.4A-D). Due to differences in cleavage sites several A $\beta$  isoforms are produced including; A $\beta_{1-42}$ , A $\beta_{4-42}$ , A $\beta_{1-40}$  and pGluA $\beta_{3-42}$ . It has been shown that in the AD brain, there are higher concentrations of A $\beta_{1-42}$ , the more toxic A $\beta$  isoform. This toxicity is due to its increased ability to aggregate, which results from two additional hydrophobic amino acids at the C terminus of the peptide.

Aβ accumulation occurs by the aggregation of monomeric Aβ into soluble aggregates (dimers, trimers and tetramers), collectively termed oligomers. Evidence suggests these soluble oligomers are the toxic species associated with AD [32-35]. Oligomers further aggregate into protofibrils and fibrils, with the aggregation of fibrils leading to the senile plaque formation. Proposed mechanisms of oligomer toxicity include; mitochondrial dysfunction [36, 37], synaptic toxicity [38], membrane depolarisation [39], oxidative stress [40], and inhibition of long-term potentiation [41, 42].



## Figure 1.1.4. APP Processing

Non-amyloidogenic (A, B, C, D) and amyloidogenic (A, E, F, G) Amyloid Precursor Protein (APP) processing. In the non-amyloidogenic pathway, APP (A) is cleaved by  $\alpha$ -secretase within the A $\beta$  domain (B), releasing  $\alpha$ -APPs and C83 (C). C83 is then cleaved by  $\gamma$ -secretase within the transmembrane or lipid membrane (C), releasing p3 and the APP intracellular domain (AICD; D). The amyloidogenic pathway involves APP (A) being cleaved by  $\beta$ -site APP Cleaving Enzyme (BACE-1) within at the Nterminus of the A $\beta$  domain (E), releasing  $\alpha$ -APPs and C99 (F). C99 is then cleaved by  $\gamma$ -secretase within the lipid membrane (F), releasing A $\beta$  and AICD (G).

#### 1.1.1.4 Neurofibrillary Tangles

Neurofibrillary tangles (NFT) are described as bundles of paired, helically wound filaments present in the cytoplasm of neurons (Figure 1.1.5, [43]). It has been observed that the density of NFTs is correlated with the severity of AD [44]. NFTs have been shown to be concentrated in the entorhinal cortex, hippocampus, amygdala, and frontal, temporal, and parietal lobes [45]. The main component of NFTs is aggregated hyperphosphorylated insoluble microtubule-associated protein tau (MAPT, Tau) [46-48].

#### 1.1.1.4.1 Tau

Tau, encoded by the *MAPT* gene located on chromosome 17, is expressed in 6 isoforms ranging from 45-65kDa, which are produced by the alternative splicing of the mRNA. The main function of the tau protein is the stabilisation of microtubules which constitute the neuronal cytoskeleton [50]. Phosphorylation of tau occurs normally as a method of microtubule binding regulation [51]. However, hyperphosphorylation is proposed to be, in some cases, a result of up-regulation of kinases that interact with proteins involved in APP processing [52]. The hyperphosphorylated tau is hypothesised to alter the binding of microtubules and result in aggregation [51]. This aggregation leads to the reduction in tau's ability to stabilise dendrite and axon branches, leading to synaptic loss.



Figure 1.1.5. Neurofibrillary Tangles

Sections of an Alzheimer's disease patient's hippocampus, immunochemistry with anti-tau antibody. (A) A number of neurofibrillary tangles (NFT) are observed with examples highlighted by red arrows (180× magnification). (B) Two NFTs present (red arrows), as well as two neurons with low levels of tau immunoreactivity in the 'pre-tangle' stage (black arrows; 360× magnification). Image sourced from [49].

#### **1.1.2 Clinical Features**

The deterioration of memory, cognition and the ability to perform functions required for daily living characterises the clinical presentation of AD [53]. Initially, patients find difficulty in learning and retaining new information with little impact on older memories. A patient in the early disease stages will also begin to struggle when organising complex tasks of daily living [53]. Further, neuropsychological tests will be able to observe in some patients subtle decline in vocabulary and speech fluency [54]. Finally, patients can experience some disorientation and issues with navigation during the early disease stages [55].

This progresses gradually to the loss of recent memories, obvious difficulties when verbally expressing themselves, and the inability to perform functions of daily life without supervision [53]. Additionally, disease progression is accompanied by the deterioration of facets of visual processing including; disorientation, impaired recognition of known faces and delusions [56].

Most cognitive functions are impacted in severely demented AD patients. In particular, loss of early memories and the ability to verbally communicate characterise this stage [54]. Additionally, misunderstandings of carer's actions can lead to aggressive behaviour by patients [57, 58]. During the late disease stage, impairment of daily living tasks becomes so severe it results in a significant reduction in life expectancy [59].

#### 1.1.3 Diagnosis and Monitoring

Definitive AD diagnosis requires post-mortem identification of neuropathological hallmarks in the brain of the patient [60]. However, clinical assessment, cerebrospinal fluid (CSF) and blood biomarkers, and brain imaging can provide tentative diagnoses. Further, these techniques, in particular brain imaging, have the ability to monitor disease progression from preclinical to symptomatic AD.

#### 1.1.3.1 Neuropsychological Testing

In recent years there have been revisions of the criteria previously used in the diagnosis of AD to reflect the increased understanding of the disease. The National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's Disease and Related Disorders Association (NINCDS-ADRDA) criteria published in 1984, initially characterised a patient's likelihood of having AD (definite, probable, possible, unlikely) based on the number of cognitive domains (memory, language, perceptual skills, attention, constructive abilities, orientation, problem solving, functional abilities) impaired [61]. The National Institute on Aging/Alzheimer's Association (NIA/AA) updated guidelines included the addition of measurements of changes occurring in the brain as measured by biomarkers [62].

The Diagnostic and Statistical Manual of Mental Disorders, 5th Edition (DSM-5) 2013 update included the renaming of Dementia to 'major neurocognitive disorder' (NCD) and the recognition of early cognitive decline (mild NCD) [63]. The DSM-5 diagnoses NCD by the observing cognitive impairment as the defining feature of a patient's impairment. Further, the cognitive symptoms observed must impair the patient's ability to function in daily life [63]. The cognitive domains affected as presented in the DSM- 5 include; complex attention, executive function, learning and memory, language, perceptual-motor function and social cognition. Mild NCD is diagnosed by the modest impairment of the previously mentioned cognitive domains from previous performance, while not interfering with independence in daily activity. The diagnosis of NCD or mild NCD is followed by the identification of the causative disorder (e.g. AD) based on the domains affected and the order in which they are affected [63].

There are a number of neuropsychological assessments used to evaluate cognitive function for the diagnostic criteria above. These assessments have also been used to monitor disease progression and symptoms in patients and decline in at risk or healthy individuals. Table 1.1.1 lists the commonly utilised cognitive assessments in AD as well as the cognitive domains they aim to measure. The combination of results from these assessments into domain specific composite scores has been shown to improve the measurement of subtle preclinical cognitive decline [64-67].

#### 1.1.3.2 Neuroimaging

Neuropsychological testing alone is not accurate in diagnosing early stages of AD, where clinical features are not pronounced. For this reason, much research focus has been on the observation of biological changes occurring as the disease progresses. The ability to monitor pathological changes early is favourable due to the extensive preclinical stage in AD. Brain imaging techniques have proven successful in evaluating these changes, allowing for the observation of gross structural changes and accumulation of disease associated proteins.

Neuropsychological Assessments	<b>Cognitive Domain</b>	Reference
Alzheimer's Disease Assessment Scale – Cognitive section (ADAS-COG)	Memory, Language, Attention	[68]
Boston Naming Test (BNT)	Language	[69]
Brief Visuospatial Memory Test—Revised	Memory, Visuospatial Ability	[70]
California Verbal Learning Test (CVLT)	Memory	[71]
Clinical Dementia Rating (CDR)	Clinical Progression of Dementia	[72]
Clock Test	Visuospatial Ability	[73]
Controlled Oral Word Association Test (COWA)	Language	[74]
Delis-Kaplan Executive Function System (D-KEFS)	Executive Function	[75]
Mini-Mental State Examination (MMSE)	General Cognition	[76]
North American National Adult Reading Test (NART)	Intelligence	[77]
Rey Auditory Verbal Learning Test (RAVLT)	Memory	[78]
Rey Complex Figure Test And Recognition Trial (RCFT)	Visuospatial Ability	[79]
Rey-Osterrieth Complex Figure Test (ROCF)	Memory	[80, 81]
Stroop Task	Executive Function	[82]
Trail Making Test	Attention, Problem- Solving	[83]
Wechsler Adult Intelligence Scale (WAIS)	Intelligence	[84]
Wechsler Memory Scale (WMS)	Memory, Attention	[85]
Wechsler Test of Adult Reading (WTAR)	Intelligence	[86]

# Table 1.1.1. Neuropsychological Assessments in AD

Neuropsychological assessments used to diagnose and monitor progression and risk for

AD, including the cognitive domains they aim to measure.

#### 1.1.3.2.1 Magnetic Resonance Imaging

The most widely utilised Magnetic Resonance Imaging (MRI) technique facilitates the observation of structural brain changes, in particular brain atrophy, and is considered important in AD diagnosis. In addition to allowing for the investigation of both the whole brain structure and specific areas, MRI is widely available, relatively inexpensive and non-invasive.

MRI global and regional brain volume measures have previously been shown to have the ability to discriminate between AD, MCI and CN classifications (Figure 1.1.6, [13, 87-90]). Cross-sectional and longitudinal MRI measures have been associated with conversion from MCI to AD [14, 88, 89, 91-95], and the severity of disease [96]. Further, in studies investigating ongoing disease progression and cognitive decline, the addition of MRI measures significantly improved the prediction power of the models containing age, gender and baseline memory scores [97-99]. More recently, measures of regional brain volume by MRI have been used in healthy elderly populations as a way of predicting the development of AD before clinical symptomology [100].

#### 1.1.3.2.2 Positron Emission Tomography

Positron emission tomography (PET) is an imaging technique based on the detection of positron-emitting radioisotopes. Appropriate ligands and radiolabelled isotopes constitute imaging agents, or tracers, in PET scanning. Commonly used ligands in AD studies utilise the structure of the hallmark protein aggregates for binding and detection. Aggregated A $\beta$  and Tau form  $\beta$ -sheet secondary structures [102, 103] within which the aromatic tracers bind [104].



#### Figure 1.1.6. Magnetic Resonance Imaging

T1-weighted volume Magnetic Resonance Images (MRIs), in (A) cognitively normal, (B) mildly cognitively impaired, and (C) Alzheimer's disease older adults ( $\geq$  70 years old). Image sourced from [101].

A $\beta$ , as discussed previously, is the main component of one of the defining hallmarks of AD. Additionally, it is relatively abundant in the diseased brain [105], has regional distribution specific to AD, and begins accumulation well before clinical diagnosis [106]. For these reasons there has been much research into the development and use of A $\beta$  specific PET imaging agents. The first A $\beta$  tracer developed, Pittsburgh Compound B (PiB), consisted of modified thioflavin-T and a carbon-11 (<sup>11</sup>C) label [107]. Due to the short half-life of <sup>11</sup>C (20 minutes) research has since focused on tracers labelled with fluorine-18 (<sup>18</sup>F, 110 minutes, [108]). <sup>18</sup>F tracers approved for clinical use include; florbetapir [109], flutemetamol [110] and florbetaben [111].

Neocortical A $\beta$ -amyloid burden as measured by PET has been consistently correlated with both post-mortem brain A $\beta$  burden [112-117] and CSF A $\beta_{42}$  [118-123]. Further, brain regions previously associated with increase A $\beta$  plaque load (by autopsy or biopsy), were replicated in PET studies by levels of tracer retention [117, 124-130]. The extent of neocortical A $\beta$  burden has been shown to be significantly different between clinical classifications of CN, MCI and AD (Figure 1.1.7, [110, 131-133]). Longitudinal studies have reported rates of A $\beta$  accumulation from CN or MCI to AD [134, 135] consistent with the accepted timelines of AD development [106]. Increased amounts of neocortical A $\beta$  has been associated with cognitive decline and brain atrophy in CN and MCI [136-141].



### Figure 1.1.7. Positron Emission Tomography

Positron Emission Tomography (PET) images utilising a <sup>11</sup>C Pittsburgh Compound B (PiB) tracer in cognitively normal, mildly cognitively impaired (MCI), and Alzheimer's disease participants. Colour intensity is correlated with the concentration of deposited amyloid. Image sourced from [142].

While major research initially focused on the development of tracers for A $\beta$  detection, there has been increased interest in the development of Tau specific tracers. There are numerous difficulties associated with the development of tracers for Tau including; the location of Tau aggregation requiring the tracer to cross the blood brain barrier, the low concentration of Tau aggregates, and Tau aggregation not being specific to AD [143]. Despite these difficulties, a number of Tau tracers have been developed and used in human trials [144-146]. These tracers have demonstrated the ability to bind to hyperphosphorylated Tau in the brains of patients with AD [147, 148], and have been associated with levels of cognitive impairment [147]. Further, there are a number of tracers in development currently [149, 150]. Like A $\beta$  tracers, those used for Tau detection use both <sup>11</sup>C [146] and <sup>18</sup>F [144, 145, 149, 150] isotope tags.

#### **1.1.4 Preclinical Disease**

The notion of the preclinical AD stage has been investigated since the 1970s when, through autopsy, neuropathological changes were observed in the brains of asymptomatic individuals [151, 152]. Broadly, preclinical AD is the long period in which abnormal neuropathological features accrue while the individual is considered cognitively normal. In 2013, Villemagne *et al.* reported that the deposition of A $\beta$  occurs for ~20 years before the clinical diagnosis of AD (Figure 1.1.8, [106]). Further, it has been observed in both the Harvard Aging Brain Study (HABS) and the Australian Imaging Biomarker and Lifestyle Study of Aging (AIBL), that ~30% of CN individuals have significant A $\beta$  deposition [153, 154].



Figure 1.1.8. A B Deposition in Alzheimer's Disease

(A) The distribution of Aβ-amyloid (Aβ) burden based on clinical classification and(B) the timeline of Aβ deposition. Image sourced from [106]. Healthy control (HC),Mild cognitive impairment (MCI), Alzheimer's disease (AD).

Clinically normal individuals with high  $A\beta$  burden are more likely to present with neurodegeneration as measured by hippocampal volume or glucose metabolism [106, 139, 155]. Further, individuals with high  $A\beta$  burden and/or neurodegeneration are reported to have impaired cross-sectional and longitudinal cognitive performance [106, 155-157]. A meta-analysis of 38 studies investigating relationships between  $A\beta$  levels (CSF or neocortical) and cognition found  $A\beta$ -related impairment in global cognition, visuospatial function, processing speed, episodic memory, and executive function [158]. Additionally, this meta-analysis reported observable decline in global cognition, semantic memory, visuospatial function, and episodic memory related to  $A\beta$  burden [158].

Finally, there have been a number of genetic factors, in particular Apolipoprotein E (*APOE*) and Brain Derived Neurotropic Factor (*BDNF*; both discussed in detail below), shown to influence preclinical decline. *APOE* and *BDNF* have been reported to increase the rates of cognitive decline and hippocampal atrophy in CN older adults with high levels of neocortical A $\beta$ , both individually (*APOE* or *BDNF*; [159-164]) and in combination (*APOE*×*BDNF*; [165]).

Due to these developments in the understanding of preclinical disease, in addition to the updated diagnostic guidelines, the NIA/AA also published recommended criteria for different stages of preclinical AD (Figure 1.1.9, [166]). These stages precede MCI and AD and aim to represent the progression of asymptomatic individuals. Stage 1 includes individuals that demonstrate A $\beta$ -accumulation (amyloidosis), as measured by



#### Figure 1.1.9. Stages of Preclinical AD

Preclinical AD stages represented graphically. Not all individuals once entering a preclinical AD stage are expected to progress to the following stage. Imaged sourced from [166].

CSF biomarkers or PET, in the absence of any additional change in neuropathological or cognitive changes [166]. Stage 2 involves amyloidosis and measures of neurodegeneration, including; structural MRI, <sup>18</sup>F-fluorodeoxyglucose (FDG) PET (a measure of glucose metabolism), or elevated levels of CSF tau or phospho-tau [166]. Finally, stage 3 consists of amyloidosis, neurodegeneration, and subtle cognitive decline that does not yet meet criteria for the diagnosis of MCI [166].

#### 1.1.5 Risk Factors

Risk factors associated with the development of AD can be separated into 2 categories. Autosomal Dominant AD (ADAD), is characterised by the inheritance of autosomal dominant mutations, and accounts for ~1% of all AD cases. ADAD commonly presents a more aggressive course of disease, and has an early age of onset, usually younger than 65 years. Alternatively, Sporadic AD (SAD), with an age at onset generally older than 65 years, is a complex disease believed to result from the combination of genetic (nonmodifiable), environment and lifestyle (modifiable) factors.

#### 1.1.5.1 Lifestyle

There has been a wide range of modifiable lifestyle risk factors that have been associated with the development of SAD. These factors include; smoking, diet, sleep and physical activity. It has previously been reported that smoking is associated with increased risk of AD [167], and results in increased rates of brain atrophy [168, 169] and cognitive decline [168]. A risk of AD [170-172] and increased rates of cognitive decline [172, 173] have been associated with poor adherence to healthy dietary patterns, a principle example of which is the Mediterranean Diet (MeDi). Briefly, the MeDi can be characterised by an increased consumption of fruits and vegetables, legumes and cereals, fish, and unsaturated fatty acids, and decreased consumption of dairy, meat and poultry, and saturated fatty acids. Further, moderate but regular alcohol consumption, mostly in the form of wine, also typifies the MeDi. Sleep disturbance has been observed to increase the risk of AD development and cognitive decline in CN [174, 175]. Finally, higher levels of physical activity have been associated with a reduction in risk for AD [176-178], as well as improved longitudinal and cross-sectional cognitive function [179-181], memory [179] and attention [182].

## 1.1.5.2 Genetics

The causative genes implicated in ADAD encode proteins involved in APP processing, and are associated with alterations in A $\beta$  production and aggregation. Alternatively, genes associated with increased risk of developing SAD have been generally implicated in a number of biological pathways involved in A $\beta$  clearance and processing. The following book chapter provides in depth information into the genes that have been implicated in ADAD, SAD and Parkinson's disease (PD).

# 1.2 Genetics of Neurodegenerative Disease: Role of genetics in

# Alzheimer's and Parkinson's Disease

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# **1.3 Polygenic Risk Scores**

As discussed in the preceding book chapter, there have been a number of GWAS identifying variants associated with AD [192, 200, 609-611]. These studies have reported that individually, the variants identified have small effect sizes for the risk of developing the late-onset form of AD when compared to the major genetic risk factor, *APOE*. To account for their nominal effects, the combination of these gene variants into a polygenic risk score (PRS) has been employed in various forms.

In general, PRS are calculated by the sum of the risk alleles for all variants, or the sum of risk alleles weighted by odds ratios or effect sizes [612]. A popular approach in AD research is the calculation of PRS using the gene variants identified in the aforementioned GWAS [192, 200, 609-611]. These PRS contain between 3 and 23 variants and have been associated with a range of cognitive and biological disease markers. Studies investigating un-weighted PRS have evaluated baseline and longitudinal cognitive change in cognitively normal older adults, and have failed to identify any significant associations [613-616]. In contrast, when weighting risk alleles by previously published odds ratios, or beta coefficients, a number of associations with overall disease risk have been observed. In cohorts investigating CN to AD participants, PRS weighted by GWAS reported odds ratios, were associated with; incidence of dementia [443], age of disease onset [617], age related structural brain changes [618], memory decline [619], and measures of CSF A $\beta_{42}$  [617, 620]. Additionally, in a CN to AD cohort evaluating a PRS weighted by beta coefficients, associations with diagnostic status and the severity of MRI measures were identified [324]. Significant associations between PRS and disease measures have also been identified in preclinical and

cognitively normal cohorts. In MCI cohorts, PRS weighted by odds ratios were associated with cognitive decline [621], measures of CSF Tau and phosphorylated Tau [621], and accelerated progression to AD [622]. Finally, when investigating CN participants, odds ratio weighted PRS were associated with baseline and longitudinal cognition [613, 623], hippocampal volume [390, 624], and cortex thickness [625].

In addition to the PRS reported above, others have been reported with altered methods of variant inclusion and weightings. While focusing on AD-risk genome-wide significant SNPs is important, it has two potential limitations. The first of these is the resultant loss of much genetic variance, i.e. the exclusion of genetic variants that are statistically significant but fail to reach stringent genome-wide significance cut-offs. To address this first potential limitation, PRS have been developed using an expanded selection of SNPs that are identified through decreasing the stringency of genetic association with AD risk. In a study by Mormino et al. an AD-risk weighted PRS, which set the criteria for inclusion at p=0.01 resulted in the inclusion of ~16,000 SNPs. This "conservative" PRS, was reported to be associated with baseline and longitudinal memory and executive function, baseline hippocampal volume, progression to MCI or AD, and neocortical A $\beta$  [626]. Lupton *et al.* also reported an association between hippocampal volume and an odds ratio weighted PRS employing a less stringent ADrisk association threshold (p<1×10<sup>-4</sup>; n(SNPs)=~1000; [627]). Finally, Escott-Price et al. reported a PRS consisting of ~200,000 SNPs ( $p \le 0.5$ ) with an 84% prediction accuracy for pathology confirmed AD [628]. Further, different methods of risk evaluation and weighting in PRS have been investigated for the prediction of AD phenotypes. A PRS of AD-associated variants, weighted by a combination of AD risk and population-based rates of AD, has been associated with individuals age of AD onset [629].

The second potential limitation of focusing on AD-risk genetic variants is the exclusion of SNPs that are more associated with pathological or symptomatic (i.e. cognition) changes than AD-risk. This potential limitation has been far-less explored to date. However, the combination of two genes previously associated with cognitive decline has been investigated, namely APOE and Brain Derived Neurotrophic Factor (BDNF). While APOE is the main genetic risk factor in AD, it is also significantly associated with rates of cognitive decline [159, 162]. On the other hand, whilst BDNF has been associated with cognition [161] the AlzGene meta-analysis suggests that it is not risk factor for AD [630]. The combination of these genetic variants has been shown to significantly impact Aβ-induced cognitive decline. In a cohort of CN older adults with high levels of brain A $\beta$ , those carrying both an *APOE*  $\epsilon$ 4 and the *BDNF* met allele (rs6265) declined cognitively significantly faster when compared to all other groups of allele combinations [165]. These studies highlight the potential importance of expanding the genes included in AD PRS from only those associated with AD risk to those gene variants that have been previously associated with disease phenotypes. This notion provides the principle theoretical framework for the research undertaken through the course of this doctoral thesis.

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## **1.4 Theoretical Framework**

In 2014 it was reported by Cummings *et al.* that of the 413 AD clinical trials performed between 2002 and 2012, there was only a 0.4% overall success rate [631]. It has been reported that these failures are due partly to the late disease stage of trial implementation. This, in addition to the increasing understanding of the extended preclinical stage of AD, has resulted in greater focus placed on clinical trials being conducted in the earlier pre-symptomatic disease stages. These trials involve the identification of at-risk individuals who are expected to decline over the course of the trial based on brain imaging and CSF biomarkers. However, these biomarkers do not inform on rates of cognitive change, which remains in most cases the primary outcome evaluated in trials. It has been observed that in biomarker-positive AD at-risk individuals there is much variability between individuals in longitudinal measures of cognition. This results in uncertainty as to whether at-risk individuals selected for trial inclusion will decline cognitively at rates appropriate for short clinical trials.

In addition to the genetic influence on AD risk outlined in the review above, cognition has been reported to be highly heritable and polygenic. Combinations of genes associated with AD-risk based on the large GWAS studies have been associated with cognitive decline. Further, combinations of genes associated with cognition, namely *APOE* and *BDNF*, have been reported to be associated with longitudinal cognition in CN at-risk cohorts. As such, genetics could be utilised, in combination with brain imaging and CSF biomarkers, for the identification of individuals appropriate for enrolment in clinical trials The aim of this thesis was to confirm previously reported associations between ADrisk genes and cognition associated genes with sensitive cognitive composite scores in a preclinical subset of a highly characterised longitudinal cohort. Further it was aimed to combine the effects of AD-risk genes and genes associated with cognition in a novel method that had the ability to appropriately weight genes based on their influence on the preclinical endophenotypes being tested rather than late stage measures of disease risk.

# 1.5 Hypothesis and Aims

The overarching hypothesis of this thesis is that *genetic factors in combination influence cognitive rates of change in preclinical Alzheimer's disease.* 

#### Aim 1 (Chapter 2):

Assess the impact of genes previously associated with AD risk on measures of cognition at a preclinical stage.

## Aim 2 (Chapter 3):

Assess the effects of genes with *a priori* evidence for association with cognition on cognitive rates of change in preclinical AD.

### Aim 3 (Chapter 4):

Investigate whether there is a synergistic effect of genes previously associated with cognition, and further what the best combination of these genes would be.

#### Aim 4 (Chapter 5):

Determine a method of weighting genes associated with both AD-risk and cognition, for use in a genetic risk score to improve the prediction of preclinical cognitive rates of change.
### **1.6 References**

- 1. Alzheimer's Disease International, World Alzheimer Report. 2015, ADI: London.
- Australian Institute of Health and Welfare, *Dementia in Australia*. 2012, AIHW: Canberra.
- Modelling, N.C.f.S.a.E., *Economic Cost of Dementia in Australia 2016-2056*. 2017, Institute for Governance and Policy Analysis: Canberra.
- 4. Australian Bureau of Statistics, *Causes of Death, Australia, 2015.* 2016: Canberra.
- 5. Vickland, V., et al., Modelling the impact of interventions to delay the onset of dementia in Australia. A report for Alzheimer's Australia. 2012.
- 6. Alzheimer's Association, *Changing the Trajectory of Alzheimer's Disease: How a Treatment by 2025 Saves Lives and Dollars*. 2015, Alzheimer's Association: USA.
- 7. Alzheimer's Disease International, *World Alzheimer Report*. 2009, ADI: London.
- Hardy, J.A. and G.A. Higgins, *Alzheimer's disease: the amyloid cascade hypothesis*.
   Science, 1992. 256(5054): p. 184-5.
- 9. Arriagada, P.V., K. Marzloff, and B.T. Hyman, *Distribution of Alzheimer-type* pathologic changes in nondemented elderly individuals matches the pattern in *Alzheimer's disease*. Neurology, 1992. **42**(9): p. 1681-8.
- Scahill, R.I., et al., *Mapping the evolution of regional atrophy in Alzheimer's disease:* unbiased analysis of fluid-registered serial MRI. Proc Natl Acad Sci U S A, 2002.
   99(7): p. 4703-7.
- Gili, T., et al., *Regional brain atrophy and functional disconnection across Alzheimer's disease evolution*. J Neurol Neurosurg Psychiatry, 2011. 82(1): p. 58-66.

- Cavedo, E., et al., *Medial temporal atrophy in early and late-onset Alzheimer's disease*. Neurobiol Aging, 2014. 35(9): p. 2004-12.
- Convit, A., et al., Specific hippocampal volume reductions in individuals at risk for Alzheimer's disease. Neurobiol Aging, 1997. 18(2): p. 131-8.
- 14. Jack, C.R., Jr., et al., *Prediction of AD with MRI-based hippocampal volume in mild cognitive impairment*. Neurology, 1999. **52**(7): p. 1397-403.
- Lehericy, S., et al., *Magnetic resonance imaging of Alzheimer's disease*. Eur Radiol, 2007. 17(2): p. 347-62.
- Vinters, H.V., *Cerebral amyloid angiopathy. A critical review.* Stroke, 1987. 18(2): p. 311-24.
- Ellis, R.J., et al., *Cerebral amyloid angiopathy in the brains of patients with Alzheimer's disease: the CERAD experience, Part XV.* Neurology, 1996. 46(6): p. 1592-6.
- Attems, J., et al., *Topographical distribution of cerebral amyloid angiopathy and its effect on cognitive decline are influenced by Alzheimer disease pathology*. J Neurol Sci, 2007. 257(1-2): p. 49-55.
- Biffi, A. and S.M. Greenberg, *Cerebral amyloid angiopathy: a systematic review*. J Clin Neurol, 2011. 7(1): p. 1-9.
- 20. Tomonaga, M., *Cerebral amyloid angiopathy in the elderly*. J Am Geriatr Soc, 1981.
  29(4): p. 151-7.
- 21. Yamada, M., et al., *Cerebral amyloid angiopathy in the aged.* J Neurol, 1987. 234(6):p. 371-6.

- 22. Zekry, D., et al., *Cerebral amyloid angiopathy in the elderly: vessel walls changes and relationship with dementia.* Acta Neuropathol, 2003. **106**(4): p. 367-73.
- 23. Kovari, E., et al., *The relationship between cerebral amyloid angiopathy and cortical microinfarcts in brain ageing and Alzheimer's disease*. Neuropathol Appl Neurobiol, 2013. 39(5): p. 498-509.
- 24. Thal, D.R., et al., *Phases of A beta-deposition in the human brain and its relevance for the development of AD*. Neurology, 2002. **58**(12): p. 1791-800.
- Glenner, G.G. and C.W. Wong, *Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein*. Biochem Biophys Res Commun, 1984. **120**(3): p. 885-90.
- Masters, C.L., et al., *Amyloid plaque core protein in Alzheimer disease and Down syndrome*. Proc Natl Acad Sci U S A, 1985. 82(12): p. 4245-9.
- 27. Griffin, W.S., et al., Brain interleukin 1 and S-100 immunoreactivity are elevated in Down syndrome and Alzheimer disease. Proc Natl Acad Sci U S A, 1989. 86(19): p. 7611-5.
- 28. Thal, D.R., R. Schober, and G. Birkenmeier, *The subunits of alpha2-macroglobulin receptor/low density lipoprotein receptor-related protein, native and transformed alpha2-macroglobulin and interleukin 6 in Alzheimer's disease*. Brain Res, 1997.
  777(1-2): p. 223-7.
- 29. Namba, Y., et al., *Apolipoprotein E immunoreactivity in cerebral amyloid deposits* and neurofibrillary tangles in Alzheimer's disease and kuru plaque amyloid in *Creutzfeldt-Jakob disease*. Brain Res, 1991. **541**(1): p. 163-6.
- McGeer, P.L., et al., *Involvement of microglia in Alzheimer's disease*. Neuropathol Appl Neurobiol, 1994. 20(2): p. 191-2.

- Perl, D.P., Neuropathology of Alzheimer's disease. Mt Sinai J Med, 2010. 77(1): p. 32-42.
- 32. Lambert, M.P., et al., *Diffusible, nonfibrillar ligands derived from Abeta1-42 are potent central nervous system neurotoxins*. Proc Natl Acad Sci U S A, 1998. 95(11): p. 6448-53.
- Lesne, S., et al., A specific amyloid-beta protein assembly in the brain impairs memory. Nature, 2006. 440(7082): p. 352-7.
- Maji, S.K., et al., Amino acid position-specific contributions to amyloid beta-protein oligomerization. J Biol Chem, 2009. 284(35): p. 23580-91.
- 35. Tomic, J.L., et al., Soluble fibrillar oligomer levels are elevated in Alzheimer's disease brain and correlate with cognitive dysfunction. Neurobiol Dis, 2009. 35(3): p. 352-8.
- Lustbader, J.W., et al., *ABAD directly links Abeta to mitochondrial toxicity in Alzheimer's disease*. Science, 2004. **304**(5669): p. 448-52.
- 37. Reddy, P.H., M. Manczak, and X. Yin, *Mitochondria-Division Inhibitor 1 Protects Against Amyloid-beta induced Mitochondrial Fragmentation and Synaptic Damage in Alzheimer's Disease*. J Alzheimers Dis, 2017. **58**(1): p. 147-162.
- Smith, L.M. and S.M. Strittmatter, *Binding Sites for Amyloid-beta Oligomers and Synaptic Toxicity*. Cold Spring Harb Perspect Med, 2017. 7(5).
- 39. Muller, W.E., C. Kirsch, and G.P. Eckert, *Membrane-disordering effects of betaamyloid peptides*. Biochem Soc Trans, 2001. **29**(Pt 4): p. 617-23.

- 40. Martins, R.N., et al., *Increased cerebral glucose-6-phosphate dehydrogenase activity in Alzheimer's disease may reflect oxidative stress.* J Neurochem, 1986. **46**(4): p. 1042-5.
- Wang, H.W., et al., Soluble oligomers of beta amyloid (1-42) inhibit long-term potentiation but not long-term depression in rat dentate gyrus. Brain Res, 2002.
  924(2): p. 133-40.
- 42. Walsh, D.M., et al., *Naturally secreted oligomers of amyloid beta protein potently inhibit hippocampal long-term potentiation in vivo*. Nature, 2002. 416(6880): p. 535-9.
- 43. Grundke-Iqbal, I., et al., *Microtubule-associated protein tau. A component of Alzheimer paired helical filaments.* J Biol Chem, 1986. **261**(13): p. 6084-9.
- 44. Bierer, L.M., et al., *Neocortical neurofibrillary tangles correlate with dementia severity in Alzheimer's disease*. Arch Neurol, 1995. **52**(1): p. 81-8.
- 45. Delacourte, A., et al., *The biochemical pathway of neurofibrillary degeneration in aging and Alzheimer's disease*. Neurology, 1999. **52**(6): p. 1158-65.
- Wood, J.G., et al., Neurofibrillary tangles of Alzheimer disease share antigenic determinants with the axonal microtubule-associated protein tau (tau). Proc Natl Acad Sci U S A, 1986. 83(11): p. 4040-3.
- 47. Ihara, Y., et al., *Phosphorylated tau protein is integrated into paired helical filaments in Alzheimer's disease*. J Biochem, 1986. **99**(6): p. 1807-10.
- 48. Iqbal, K., et al., *Defective brain microtubule assembly in Alzheimer's disease*. Lancet, 1986. 2(8504): p. 421-6.

- 49. Brion, J.P., *Neurofibrillary tangles and Alzheimer's disease*. Eur Neurol, 1998. 40(3):
  p. 130-40.
- 50. Drubin, D.G. and M.W. Kirschner, *Tau protein function in living cells*. J Cell Biol, 1986. 103(6 Pt 2): p. 2739-46.
- 51. Avila, J., *Tau aggregation into fibrillar polymers: taupathies*. FEBS Lett, 2000.
  476(1-2): p. 89-92.
- 52. Vincent, I., et al., Aberrant expression of mitotic cdc2/cyclin B1 kinase in degenerating neurons of Alzheimer's disease brain. J Neurosci, 1997. 17(10): p. 3588-98.
- Forstl, H. and A. Kurz, *Clinical features of Alzheimer's disease*. Eur Arch Psychiatry Clin Neurosci, 1999. 249(6): p. 288-90.
- 54. Chobor, K.L. and J.W. Brown, *Semantic deterioration in Alzheimer's: the patterns to expect.* Geriatrics, 1990. **45**(10): p. 68-70, 75.
- 55. Vlcek, K. and J. Laczo, *Neural correlates of spatial navigation changes in mild cognitive impairment and Alzheimer's disease*. Front Behav Neurosci, 2014. **8**: p. 89.
- 56. Pal, S., et al., Visual manifestations in Alzheimer's disease: a clinic-based study from India. Am J Alzheimers Dis Other Demen, 2013. 28(6): p. 575-82.
- 57. Lyketsos, C.G., et al., *Mental and behavioral disturbances in dementia: findings from the Cache County Study on Memory in Aging.* Am J Psychiatry, 2000. 157(5): p. 708-14.
- Bidzan, L., M. Bidzan, and M. Pachalska, *Aggressive and impulsive behavior in Alzheimer's disease and progression of dementia*. Med Sci Monit, 2012. 18(3): p. CR182-9.

- 59. Burns, A., et al., *Factors affecting survival in Alzheimer's disease*. Psychol Med, 1991. 21(2): p. 363-70.
- 60. Mirra, S.S., et al., *The Consortium to Establish a Registry for Alzheimer's Disease* (CERAD). Part II. Standardization of the neuropathologic assessment of Alzheimer's disease. Neurology, 1991. **41**(4): p. 479-86.
- 61. McKhann, G., et al., *Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease.* Neurology, 1984. **34**(7): p. 939-44.
- McKhann, G.M., et al., *The diagnosis of dementia due to Alzheimer's disease:* recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement, 2011. 7(3): p. 263-9.
- 63. American Psychiatric Association, *Diagnostic and statistical manual of mental disorders*. 5th ed. 2013, Arlington, VA: American Psychiatric Publishing.
- 64. Burnham, S.C., et al., Novel Statistically-Derived Composite Measures for Assessing the Efficacy of Disease-Modifying Therapies in Prodromal Alzheimer's Disease Trials: An AIBL Study. J Alzheimers Dis, 2015. **46**(4): p. 1079-89.
- 65. Lim, Y.Y., et al., Sensitivity of composite scores to amyloid burden in preclinical Alzheimer's disease: Introducing the Z-scores of Attention, Verbal fluency, and Episodic memory for Nondemented older adults composite score. Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring, 2016. 2: p. 19-26.
- 66. Donohue, M.C., et al., *The preclinical Alzheimer cognitive composite: measuring amyloid-related decline*. JAMA Neurol, 2014. **71**(8): p. 961-70.

- 67. Harrington, K.D., et al., *The association of Abeta amyloid and composite cognitive measures in healthy older adults and MCI*. Int Psychogeriatr, 2013. 25(10): p. 1667-77.
- Rosen, W.G., R.C. Mohs, and K.L. Davis, *A new rating scale for Alzheimer's disease*.
   Am J Psychiatry, 1984. 141(11): p. 1356-64.
- 69. Kaplan, E., H. Goodglass, and S. Weintraub, *Boston Naming Test.* 1983, Philadelphia, PA: Lea & Febiger.
- Benedict, R.H., Brief Visuospatial Memory Test—Revised. 1997, Odessa, FL:
   Psychological Assessment Resources, Inc.
- 71. Delis, D.C., et al., *California Verbal Learning Test second edition. Adult version. Manual.* 2000, San Antonio, TX: Psychological Corporation.
- Morris, J.C., *The Clinical Dementia Rating (CDR): current version and scoring rules*.
  Neurology, 1993. 43(11): p. 2412-4.
- 73. Tuokko, H., et al., *The Clock Test: a sensitive measure to differentiate normal elderly from those with Alzheimer disease.* J Am Geriatr Soc, 1992. **40**(6): p. 579-84.
- 74. Benton, A.L., K.d.S. Hamsher, and A.B. Sivan, *Multilingual Aphasia Examination*.1994, Iowa City, IA: AJA Associates.
- Delis, D.C., E. Kaplan, and J.H. Kramer, *Delis-Kaplan Executive Function System* (*D-KEFS*). 2001, San Antonio, TX: Psychological Corporation.
- 76. Folstein, M.F., S.E. Folstein, and P.R. McHugh, "Mini-mental state". A practical method for grading the cognitive state of patients for the clinician. J Psychiatr Res, 1975. 12(3): p. 189-98.

- 77. Grober, E., M. Sliwinsk, and S.R. Korey, *Development and Validation of a Model for Estimating Premorbid Verbal Intelligence in the Elderly*. Journal of Clinical and Experimental Neuropsychology, 1991. 13(6): p. 933-949.
- Rey, A., *L'examen clinique en psychologie*. 1964, Paris, France: Presses Universitaires de France.
- Meyers, J. and K. Meyers, *Rey Complex Figure Test and Recognition Trial. Professional Manual.* 1995, Lutz, FL: Psychological Assessment Resource, Inc.
- Rey, A., *L'examen psychologique dans les cas d'escephalopathie traumatique*.
  Archives de Psychologie, 1941. 28: p. 286-340.
- 81. Osterrieth, P.A., *Le test de copie d'une figure complexe*. Archives de Psychologie, 1944. 30: p. 206-356.
- 82. Stroop, R.J., *Studies of interference in serial verbal reactions*. J. Exp. Psychol, 1935.
  18(6): p. 643-662.
- 83. Army Individual Test: Manual of Directions and Scoring. 1944, Washington, DC:
   War Department, Adjutant General's Office.
- 84. Wechsler, D., Wechsler Adult Intelligence Scale. 4th ed. 2008, San Antonio, TX: Pearson
- Wechsler, D., Wechsler Memory Scale Fourth Edition (WMS-IV). 2009, San Antonio, TX: Pearson.
- Wechsler, D., Wechsler Test of Adult Reading: Examiner's Manual. 2001, San Antonio, TX: Psychological Corporation.

- 87. Morra, J.H., et al., *Automated 3D mapping of hippocampal atrophy and its clinical correlates in 400 subjects with Alzheimer's disease, mild cognitive impairment, and elderly controls.* Hum Brain Mapp, 2009. **30**(9): p. 2766-88.
- Eckerstrom, C., et al., Small baseline volume of left hippocampus is associated with subsequent conversion of MCI into dementia: the Goteborg MCI study. J Neurol Sci, 2008. 272(1-2): p. 48-59.
- 89. Devanand, D.P., et al., *MRI hippocampal and entorhinal cortex mapping in predicting conversion to Alzheimer's disease*. Neuroimage, 2012. **60**(3): p. 1622-9.
- 90. Henneman, W.J., et al., *Hippocampal atrophy rates in Alzheimer disease: added value over whole brain volume measures.* Neurology, 2009. **72**(11): p. 999-1007.
- 91. de Toledo-Morrell, L., et al., *From healthy aging to early Alzheimer's disease: in vivo detection of entorhinal cortex atrophy.* Ann N Y Acad Sci, 2000. **911**: p. 240-53.
- 92. Chetelat, G., et al., Using voxel-based morphometry to map the structural changes associated with rapid conversion in MCI: a longitudinal MRI study. Neuroimage, 2005. 27(4): p. 934-46.
- 93. Costafreda, S.G., et al., *Automated hippocampal shape analysis predicts the onset of dementia in mild cognitive impairment*. Neuroimage, 2011. **56**(1): p. 212-9.
- 94. Leung, K.K., et al., *Cerebral atrophy in mild cognitive impairment and Alzheimer disease: rates and acceleration.* Neurology, 2013. **80**(7): p. 648-54.
- 95. Apostolova, L.G., et al., *Conversion of mild cognitive impairment to Alzheimer disease predicted by hippocampal atrophy maps.* Arch Neurol, 2006. **63**(5): p. 693-9.

- 96. Wolf, H., et al., *Hippocampal volume discriminates between normal cognition; questionable and mild dementia in the elderly*. Neurobiol Aging, 2001. 22(2): p. 177-86.
- 97. Visser, P.J., et al., Medial temporal lobe atrophy and memory dysfunction as predictors for dementia in subjects with mild cognitive impairment. J Neurol, 1999.
  246(6): p. 477-85.
- 98. Visser, P.J., et al., *Medial temporal lobe atrophy predicts Alzheimer's disease in patients with minor cognitive impairment*. J Neurol Neurosurg Psychiatry, 2002.
  72(4): p. 491-7.
- 99. Convit, A., et al., Atrophy of the medial occipitotemporal, inferior, and middle temporal gyri in non-demented elderly predict decline to Alzheimer's disease.
  Neurobiol Aging, 2000. 21(1): p. 19-26.
- 100. Achterberg, H.C., et al., *Hippocampal shape is predictive for the development of dementia in a normal, elderly population.* Hum Brain Mapp, 2014. **35**(5): p. 2359-71.
- 101. Jack, C.R., Jr., *Alzheimer disease: new concepts on its neurobiology and the clinical role imaging will play.* Radiology, 2012. **263**(2): p. 344-61.
- Mukrasch, M.D., et al., Sites of tau important for aggregation populate {beta}structure and bind to microtubules and polyanions. J Biol Chem, 2005. 280(26): p. 24978-86.
- 103. Nabers, A., et al., Amyloid-beta-Secondary Structure Distribution in Cerebrospinal Fluid and Blood Measured by an Immuno-Infrared-Sensor: A Biomarker Candidate for Alzheimer's Disease. Anal Chem, 2016. 88(5): p. 2755-62.
- 104. Naiki, H., et al., *Fluorometric determination of amyloid fibrils in vitro using the fluorescent dye, thioflavin T1*. Anal Biochem, 1989. **177**(2): p. 244-9.

- 105. Roberts, B.R., et al., *Biochemically-defined pools of amyloid-beta in sporadic Alzheimer's disease: correlation with amyloid PET.* Brain, 2017.
- 106. Villemagne, V.L., et al., *Amyloid beta deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study.* Lancet Neurol, 2013. 12(4): p. 357-67.
- 107. Mathis, C.A., et al., *A lipophilic thioflavin-T derivative for positron emission tomography (PET) imaging of amyloid in brain.* Bioorg Med Chem Lett, 2002. 12(3): p. 295-8.
- 108. Mistur, R., et al., Current Challenges for the Early Detection of Alzheimer's Disease:
   Brain Imaging and CSF Studies. J Clin Neurol, 2009. 5(4): p. 153-66.
- 109. Wong, D.F., et al., *In vivo imaging of amyloid deposition in Alzheimer disease using the radioligand 18F-AV-45 (florbetapir [corrected] F 18).* J Nucl Med, 2010. 51(6): p. 913-20.
- 110. Vandenberghe, R., et al., *18F-flutemetamol amyloid imaging in Alzheimer disease and mild cognitive impairment: a phase 2 trial.* Ann Neurol, 2010. **68**(3): p. 319-29.
- 111. Villemagne, V.L., et al., *Amyloid imaging with (18)F-florbetaben in Alzheimer disease and other dementias*. J Nucl Med, 2011. 52(8): p. 1210-7.
- 112. Ikonomovic, M.D., et al., *Post-mortem correlates of in vivo PiB-PET amyloid imaging in a typical case of Alzheimer's disease*. Brain, 2008. **131**(Pt 6): p. 1630-45.
- 113. Clark, C.M., et al., Cerebral PET with florbetapir compared with neuropathology at autopsy for detection of neuritic amyloid-beta plaques: a prospective cohort study. Lancet Neurol, 2012. 11(8): p. 669-78.

- 114. Curtis, C., et al., *Phase 3 trial of flutemetamol labeled with radioactive fluorine 18 imaging and neuritic plaque density.* JAMA Neurol, 2015. **72**(3): p. 287-94.
- Sabri, O., et al., *Florbetaben PET imaging to detect amyloid beta plaques in Alzheimer's disease: phase 3 study.* Alzheimers Dement, 2015. 11(8): p. 964-74.
- 116. Ikonomovic, M.D., et al., *Post-mortem histopathology underlying beta-amyloid PET imaging following flutemetamol F 18 injection*. Acta Neuropathol Commun, 2016.
  4(1): p. 130.
- Seo, S.W., et al., *Regional correlations between [11C]PIB PET and post-mortem burden of amyloid-beta pathology in a diverse neuropathological cohort*. Neuroimage Clin, 2017. 13: p. 130-137.
- 118. Fagan, A.M., et al., *Inverse relation between in vivo amyloid imaging load and cerebrospinal fluid Abeta42 in humans*. Ann Neurol, 2006. **59**(3): p. 512-9.
- 119. Landau, S.M., et al., *Comparing positron emission tomography imaging and cerebrospinal fluid measurements of beta-amyloid*. Ann Neurol, 2013. 74(6): p. 826-36.
- Li, Q.X., et al., Alzheimer's Disease Normative Cerebrospinal Fluid Biomarkers
   Validated in PET Amyloid-beta Characterized Subjects from the Australian Imaging,
   Biomarkers and Lifestyle (AIBL) study. J Alzheimers Dis, 2015. 48(1): p. 175-87.
- 121. Leuzy, A., et al., *Pittsburgh compound B imaging and cerebrospinal fluid amyloidbeta in a multicentre European memory clinic study*. Brain, 2016. **139**(Pt 9): p. 2540-53.
- 122. Pannee, J., et al., *Reference measurement procedure for CSF amyloid beta (Abeta)1-*42 and the CSF Abeta1-42 /Abeta1-40 ratio a cross-validation study against amyloid PET. J Neurochem, 2016. 139(4): p. 651-658.

- 123. Schipke, C.G., et al., *Correlation of florbetaben PET imaging and the amyloid peptide Ass42 in cerebrospinal fluid.* Psychiatry Res, 2017. **265**: p. 98-101.
- 124. Bacskai, B.J., et al., *Molecular imaging with Pittsburgh Compound B confirmed at autopsy: a case report.* Arch Neurol, 2007. **64**(3): p. 431-4.
- 125. Leinonen, V., et al., Assessment of beta-amyloid in a frontal cortical brain biopsy specimen and by positron emission tomography with carbon 11-labeled Pittsburgh Compound B. Arch Neurol, 2008. 65(10): p. 1304-9.
- 126. Sojkova, J., et al., *In vivo fibrillar beta-amyloid detected using [11C]PiB positron emission tomography and neuropathologic assessment in older adults*. Arch Neurol, 2011. 68(2): p. 232-40.
- 127. Kadir, A., et al., Positron emission tomography imaging and clinical progression in relation to molecular pathology in the first Pittsburgh Compound B positron emission tomography patient with Alzheimer's disease. Brain, 2011. **134**(Pt 1): p. 301-17.
- 128. Wolk, D.A., et al., *Association between in vivo fluorine 18-labeled flutemetamol amyloid positron emission tomography imaging and in vivo cerebral cortical histopathology*. Arch Neurol, 2011. **68**(11): p. 1398-403.
- 129. Clark, C.M., et al., Use of florbetapir-PET for imaging beta-amyloid pathology. JAMA, 2011. 305(3): p. 275-83.
- 130. Wong, D.F., et al., *An in vivo evaluation of cerebral cortical amyloid with* [18F]flutemetamol using positron emission tomography compared with parietal biopsy samples in living normal pressure hydrocephalus patients. Mol Imaging Biol, 2013. 15(2): p. 230-7.

- 131. Fleisher, A.S., et al., Using positron emission tomography and florbetapir F18 to image cortical amyloid in patients with mild cognitive impairment or dementia due to Alzheimer disease. Arch Neurol, 2011. 68(11): p. 1404-11.
- Klunk, W.E., et al., *Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B*. Ann Neurol, 2004. 55(3): p. 306-19.
- 133. Rowe, C.C., et al., *Imaging beta-amyloid burden in aging and dementia*. Neurology, 2007. 68(20): p. 1718-25.
- 134. Villemagne, V.L., et al., *Longitudinal assessment of Abeta and cognition in aging and Alzheimer disease*. Ann Neurol, 2011. **69**(1): p. 181-92.
- 135. Okello, A., et al., *Conversion of amyloid positive and negative MCI to AD over 3 years: an 11C-PIB PET study.* Neurology, 2009. **73**(10): p. 754-60.
- 136. Sperling, R.A., et al., Amyloid deposition detected with florbetapir F 18 ((18)F-AV45) is related to lower episodic memory performance in clinically normal older
  individuals. Neurobiol Aging, 2013. 34(3): p. 822-31.
- 137. Doraiswamy, P.M., et al., *Florbetapir F 18 amyloid PET and 36-month cognitive decline: a prospective multicenter study.* Mol Psychiatry, 2014. **19**(9): p. 1044-51.
- 138. Chetelat, G., et al., *Relationship between atrophy and beta-amyloid deposition in Alzheimer disease*. Ann Neurol, 2010. **67**(3): p. 317-24.
- 139. Bourgeat, P., et al., *Beta-amyloid burden in the temporal neocortex is related to hippocampal atrophy in elderly subjects without dementia*. Neurology, 2010. 74(2): p. 121-7.
- 140. Pike, K.E., et al., *Beta-amyloid imaging and memory in non-demented individuals: evidence for preclinical Alzheimer's disease.* Brain, 2007. **130**(Pt 11): p. 2837-44.

- 141. Villemagne, V.L., et al., *Abeta deposits in older non-demented individuals with cognitive decline are indicative of preclinical Alzheimer's disease*. Neuropsychologia, 2008. 46(6): p. 1688-97.
- Mathis, C.A., B.J. Lopresti, and W.E. Klunk, *Impact of amyloid imaging on drug development in Alzheimer's disease*. Nucl Med Biol, 2007. 34(7): p. 809-22.
- 143. Bischof, G.N., et al., *Tau-imaging in neurodegeneration*. Methods, 2017.
- 144. Josephs, K.A., et al., [18F]AV-1451 tau-PET uptake does correlate with quantitatively measured 4R-tau burden in autopsy-confirmed corticobasal degeneration. Acta Neuropathol, 2016. **132**(6): p. 931-933.
- 145. Tago, T., et al., *Structure-Activity Relationship of 2-Arylquinolines as PET Imaging Tracers for Tau Pathology in Alzheimer Disease.* J Nucl Med, 2016. **57**(4): p. 608-14.
- 146. Wang, M., et al., Synthesis of a PET tau tracer [(11)C]PBB3 for imaging of Alzheimer's disease. Bioorg Med Chem Lett, 2015. 25(20): p. 4587-92.
- 147. Johnson, K.A., et al., *Tau positron emission tomographic imaging in aging and early Alzheimer disease*. Ann Neurol, 2016. **79**(1): p. 110-9.
- 148. Chien, D.T., et al., *Early clinical PET imaging results with the novel PHF-tau radioligand [F-18]-T807.* J Alzheimers Dis, 2013. **34**(2): p. 457-68.
- Hostetler, E.D., et al., Preclinical Characterization of 18F-MK-6240, a Promising PET Tracer for In Vivo Quantification of Human Neurofibrillary Tangles. J Nucl Med, 2016. 57(10): p. 1599-1606.
- Declercq, L., et al., Preclinical Evaluation of 18F-JNJ64349311, a Novel PET Tracer for Tau Imaging. J Nucl Med, 2017. 58(6): p. 975-981.

- 151. Dayan, A.D., *Quantitative histological studies on the aged human brain. I. Senile plaques and neurofibrillary tangles in "normal" patients.* Acta Neuropathol, 1970.
  16(2): p. 85-94.
- 152. Katzman, R., et al., Clinical, pathological, and neurochemical changes in dementia: a subgroup with preserved mental status and numerous neocortical plaques. Ann Neurol, 1988. 23(2): p. 138-44.
- Sperling, R., E. Mormino, and K. Johnson, *The evolution of preclinical Alzheimer's disease: implications for prevention trials.* Neuron, 2014. 84(3): p. 608-22.
- 154. Rowe, C.C., et al., *Amyloid imaging results from the Australian Imaging, Biomarkers and Lifestyle (AIBL) study of aging.* Neurobiol Aging, 2010. **31**(8): p. 1275-83.
- Mormino, E.C., et al., Synergistic effect of beta-amyloid and neurodegeneration on cognitive decline in clinically normal individuals. JAMA Neurol, 2014. 71(11): p. 1379-85.
- 156. Lim, Y.Y., et al., *Effect of amyloid on memory and non-memory decline from* preclinical to clinical Alzheimer's disease. Brain, 2014. **137**(Pt 1): p. 221-31.
- 157. Lim, Y.Y., et al., Abeta and cognitive change: examining the preclinical and prodromal stages of Alzheimer's disease. Alzheimers Dement, 2014. 10(6): p. 743-751 e1.
- 158. Baker, J.E., et al., Cognitive impairment and decline in cognitively normal older adults with high amyloid-beta: A meta-analysis. Alzheimers Dement (Amst), 2017. 6:
  p. 108-121.
- Lim, Y.Y., et al., Abeta amyloid, cognition, and APOE genotype in healthy older adults. Alzheimers Dement, 2013. 9(5): p. 538-45.

- 160. Lim, Y.Y., et al., *Effect of BDNF Val66Met on memory decline and hippocampal atrophy in prodromal Alzheimer's disease: a preliminary study.* PLoS One, 2014.
  9(1): p. e86498.
- 161. Lim, Y.Y., et al., *BDNF Val66Met, Abeta amyloid, and cognitive decline in preclinical Alzheimer's disease*. Neurobiol Aging, 2013. **34**(11): p. 2457-64.
- 162. Lim, Y.Y., et al., APOE epsilon4 moderates amyloid-related memory decline in preclinical Alzheimer's disease. Neurobiol Aging, 2015. 36(3): p. 1239-44.
- 163. Lim, Y.Y., et al., *Effect of APOE Genotype on Amyloid Deposition, Brain Volume,* and Memory in Cognitively Normal Older Individuals. J Alzheimers Dis, 2017. 58(4): p. 1293-1302.
- 164. Lim, Y.Y., et al., *Abeta-related memory decline in APOE epsilon4 noncarriers: Implications for Alzheimer disease*. Neurology, 2016. 86(17): p. 1635-42.
- 165. Lim, Y.Y., et al., APOE and BDNF polymorphisms moderate amyloid beta-related cognitive decline in preclinical Alzheimer's disease. Mol Psychiatry, 2015. 20(11): p. 1322-8.
- Sperling, R.A., et al., Toward defining the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement, 2011. 7(3): p. 280-92.
- 167. Rusanen, M., et al., *Heavy smoking in midlife and long-term risk of Alzheimer disease and vascular dementia*. Arch Intern Med, 2011. **171**(4): p. 333-9.
- Almeida, O.P., et al., 24-month effect of smoking cessation on cognitive function and brain structure in later life. Neuroimage, 2011. 55(4): p. 1480-9.

- 169. Durazzo, T.C., et al., *Greater regional brain atrophy rate in healthy elderly subjects with a history of cigarette smoking*. Alzheimers Dement, 2012. **8**(6): p. 513-9.
- 170. Gardener, S., et al., *Adherence to a Mediterranean diet and Alzheimer's disease risk in an Australian population*. Transl Psychiatry, 2012. **2**: p. e164.
- 171. Scarmeas, N., et al., *Mediterranean diet and risk for Alzheimer's disease*. Ann Neurol, 2006. 59(6): p. 912-21.
- 172. Lourida, I., et al., *Mediterranean diet, cognitive function, and dementia: a systematic review.* Epidemiology, 2013. **24**(4): p. 479-89.
- 173. Valls-Pedret, C., et al., *Mediterranean Diet and Age-Related Cognitive Decline: A Randomized Clinical Trial.* JAMA Intern Med, 2015. **175**(7): p. 1094-103.
- 174. Lim, A.S., et al., *Sleep Fragmentation and the Risk of Incident Alzheimer's Disease and Cognitive Decline in Older Persons.* Sleep, 2013. **36**(7): p. 1027-1032.
- 175. Potvin, O., et al., *Sleep quality and 1-year incident cognitive impairment in community-dwelling older adults*. Sleep, 2012. **35**(4): p. 491-9.
- 176. Larson, E.B., et al., *Exercise is associated with reduced risk for incident dementia among persons 65 years of age and older*. Ann Intern Med, 2006. **144**(2): p. 73-81.
- 177. Scarmeas, N., et al., *Physical activity, diet, and risk of Alzheimer disease*. JAMA, 2009. **302**(6): p. 627-37.
- 178. Buchman, A.S., et al., *Total daily physical activity and the risk of AD and cognitive decline in older adults*. Neurology, 2012. **78**(17): p. 1323-9.
- Weuve, J., et al., *Physical activity, including walking, and cognitive function in older women.* JAMA, 2004. 292(12): p. 1454-61.

- Middleton, L.E., et al., *Changes in cognition and mortality in relation to exercise in late life: a population based study*. PLoS One, 2008. 3(9): p. e3124.
- 181. Lautenschlager, N.T., et al., *Effect of physical activity on cognitive function in older adults at risk for Alzheimer disease: a randomized trial.* JAMA, 2008. **300**(9): p. 1027-37.
- 182. Barnes, D.E., et al., *Cognition in older women: the importance of daytime movement.*J Am Geriatr Soc, 2008. 56(9): p. 1658-64.
- 609. Hollingworth, P., et al., *Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease*. Nat Genet, 2011. 43(5): p. 429-35.
- 610. Naj, A.C., et al., *Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease.* Nat Genet, 2011. **43**(5): p. 436-41.
- 611. Harold, D., et al., *Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease*. Nat Genet, 2009. **41**(10): p. 1088-93.
- 612. Che, R. and A.A. Motsinger-Reif, *Evaluation of genetic risk score models in the presence of interaction and linkage disequilibrium*. Front Genet, 2013. **4**: p. 138.
- 613. Andrews, S.J., et al., *Association of genetic risk factors with cognitive decline: the PATH through life project.* Neurobiol Aging, 2016. **41**: p. 150-8.
- 614. Bressler, J., et al., *Genetic variants associated with risk of Alzheimer's disease contribute to cognitive change in midlife: The Atherosclerosis Risk in Communities Study.* Am J Med Genet B Neuropsychiatr Genet, 2017. **174**(3): p. 269-282.
- 615. Gui, H., et al., *Influence of Alzheimer's disease genes on cognitive decline: the Guangzhou Biobank Cohort Study*. Neurobiol Aging, 2014. **35**(10): p. 2422 e3-8.

- 616. Harrison, T.M., et al., An Alzheimer's Disease Genetic Risk Score Predicts
  Longitudinal Thinning of Hippocampal Complex Subregions in Healthy Older Adults.
  eNeuro, 2016. 3(3).
- 617. Sleegers, K., et al., A 22-single nucleotide polymorphism Alzheimer's disease risk score correlates with family history, onset age, and cerebrospinal fluid Abeta42. Alzheimers Dement, 2015. 11(12): p. 1452-60.
- 618. Habes, M., et al., Advanced brain aging: relationship with epidemiologic and genetic risk factors, and overlap with Alzheimer disease atrophy patterns. Transl Psychiatry, 2016. 6: p. e775.
- 619. Marden, J.R., et al., Using an Alzheimer Disease Polygenic Risk Score to Predict Memory Decline in Black and White Americans Over 14 Years of Follow-up.
  Alzheimer Dis Assoc Disord, 2016. 30(3): p. 195-202.
- 620. Martiskainen, H., et al., *Effects of Alzheimer's disease-associated risk loci on cerebrospinal fluid biomarkers and disease progression: a polygenic risk score approach.* J Alzheimers Dis, 2015. **43**(2): p. 565-73.
- 621. Louwersheimer, E., et al., *Alzheimer's disease risk variants modulate endophenotypes in mild cognitive impairment*. Alzheimers Dement, 2016. **12**(8): p. 872-81.
- 622. Rodriguez-Rodriguez, E., et al., *Genetic risk score predicting accelerated progression from mild cognitive impairment to Alzheimer's disease*. J Neural Transm (Vienna), 2013. 120(5): p. 807-12.
- 623. Carrasquillo, M.M., et al., *Late-onset Alzheimer's risk variants in memory decline, incident mild cognitive impairment, and Alzheimer's disease*. Neurobiol Aging, 2015.
  36(1): p. 60-7.

- 624. Foley, S.F., et al., Multimodal Brain Imaging Reveals Structural Differences in Alzheimer's Disease Polygenic Risk Carriers: A Study in Healthy Young Adults. Biol Psychiatry, 2017. 81(2): p. 154-161.
- 625. Sabuncu, M.R., et al., *The association between a polygenic Alzheimer score and cortical thickness in clinically normal subjects*. Cereb Cortex, 2012. 22(11): p. 2653-61.
- 626. Mormino, E.C., et al., *Polygenic risk of Alzheimer disease is associated with earlyand late-life processes*. Neurology, 2016. **87**(5): p. 481-8.
- 627. Lupton, M.K., et al., *The effect of increased genetic risk for Alzheimer's disease on hippocampal and amygdala volume*. Neurobiol Aging, 2016. **40**: p. 68-77.
- 628. Escott-Price, V., et al., *Polygenic risk score analysis of pathologically confirmed alzheimer disease*. Ann Neurol, 2017.
- 629. Desikan, R.S., et al., Genetic assessment of age-associated Alzheimer disease risk: Development and validation of a polygenic hazard score. PLoS Med, 2017. 14(3): p. e1002258.
- 630. Bertram, L., et al., *Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database.* Nat Genet, 2007. **39**(1): p. 17-23.
- 631. Cummings, J.L., T. Morstorf, and K. Zhong, *Alzheimer's disease drug-development pipeline: few candidates, frequent failures.* Alzheimers Res Ther, 2014. **6**(4): p. 37.

## Chapter 2

Please note that this online copy of the thesis does not contain the complete version of Chapter 2

# CHAPTER 2: Association of a priori candidate, AD-risk associated, genes with cognitive rates of change in preclinical AD

### **2.1 Prologue**

As discussed in Chapter 1 (Section 1.3), there have been numerous Polygenic Risk Scores (PRSs) calculated and subsequently associated with Alzheimer's disease (AD) related phenotypes, including AD-related imaging and fluid biomarkers and cognition. In most such studies, the PRSs that have been calculated have combined AD risk associated genetic variants, previously identified through genome wide association (GWA) studies (GWAS). These individual variants are invariably weighted by a measure of AD risk, typically the respective odds ratios or effect sizes generated from large GWAS, the most common being the International Genomics of Alzheimer's Project (IGAP) meta-analysis of GWA data [1]. Presented in this chapter is a replication of this form of PRS in the Australian Imaging, Biomarkers and Lifestyle (AIBL) Study of Aging. As no AIBL samples were included in the IGAP meta-analysis, presented here is an independent validation of previous studies. Specifically, it investigates cognitively normal (CN) individuals in the preclinical stages of AD.

Previous studies investigating associations between PRS and measures of cognitive decline have produced varying results, particularly in CN cohorts. The lack of PRS-cognition associations reported by some studies could be attributed to sample heterogeneity within the CN cohort, or the use of cognitive measures not sensitive enough for early in the disease process. The ensuing study attempts to overcome these limitations, by focusing specifically on CN older adults who are biomarker positive based on Aβ-amyloid (Aβ) imaging. Further, three cognitive composite scores are utilised that measure the first cognitive changes occurring in AD. Through this approach, the first aim of this thesis, to *assess the impact of genes previously associated with AD risk on measures of cognition at a preclinical stage*, is addressed.

### **Prologue References:**

 Lambert, J.C., et al., *Meta-analysis of 74,046 individuals identifies 11 new* susceptibility loci for Alzheimer's disease. Nat Genet, 2013. 45(12): p. 1452-8.

# 2.2 Utility of an Alzheimer's risk-weighted polygenic risk score for predicting rates of cognitive decline in preclinical Alzheimer's

## disease: a prospective longitudinal study

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## Chapter 3

Please note that this online copy of the thesis does not contain the complete version of Chapter 3

# CHAPTER 3: Association of *a priori* candidate, cognition associated, genes with cognitive rates of change in preclinical AD

### **3.1 Prologue**

The results presented in Chapter 2 evidence the potential lack of utility of Alzheimer's disease (AD) risk weightings when investigating the association of polygeneic risk scores (PRSs) with preclinical disease outcomes, such as decline in cognition. When investigating the combinatory influence of gene variants most commonly associated with the development of AD, significant associations with rates of cognitive decline in a preclinical cognitively normal population were only observed with the inclusion of *APOE*  $\varepsilon$ 4 weighting.

While it was observed that the PRS both with and without *APOE* were associated with neocortical amyloid beta ( $A\beta$ ) burden and CSF-tau, the same was not seen when investigating longitudinal cognition. This discrepancy in results between disease biomarkers and cognition could be due to the disease-risk weighting applied to the genetic variants differing from the actual influence that these variants have on cognitive performance. Further, limiting the inclusion of single nucleotide polymorphisms (SNPs) in the PRS to those with association with clinical diagnosis of AD is biased against those variants which may influence the rate of decline in the preclinical stages of the disease. For example, the non-synonymous rs6265 (Val66Met) SNP in brain derived neurotropic factor (*BDNF*) has been reported by the Australian Imaging, Biomarkers and Lifestyle (AIBL) Study of Aging and others, to be associated with altered rates of cognitive decline [1-4] but is not amongst the leading AD genetic risk factors and thus has

been excluded from PRS calculations. A number of genes have previously been associated with cognitive performance in both AD and CN populations. The studies presented in this chapter aimed to *assess the effects of genes with a priori evidence for association with cognition on cognitive rates of change in preclinical AD*, and determine their potential viability for inclusion into a polygenic measure.

A review of the literature was conducted to identify genes and genetic variants that have previously been associated with cognitive performance. The genes that were selected for inclusion are described below:

- *KIBRA*: Kidney and Brain expressed protein, variants within this gene have been associated with memory performance
- *SPON1*: Spondin1, expression of this gene has been associated with improved learning and cognition
- *COMT*: Catechol-O-methyltransferase, non-synonymous variants involved in the expression of this gene are associated with cognition
- *KL*: Klotho, variants within this gene controlling it's expression have been associated with aging phenotypes and cognitive performance

#### **Prologue references:**

- Lim, Y.Y., et al., *BDNF Val66Met*, *Abeta amyloid, and cognitive decline in preclinical Alzheimer's disease*. Neurobiol Aging, 2013. **34**(11): p. 2457-64.
- 2. Lim, Y.Y., et al., *APOE and BDNF polymorphisms moderate amyloid beta-related cognitive decline in preclinical Alzheimer's disease*. Mol Psychiatry, 2015. **20**(11): p. 1322-8.
- 3. Kennedy, K.M., et al., *BDNF val66met polymorphism affects aging of multiple types of memory*. Brain Res, 2015. **1612**: p. 104-17.

 Cathomas, F., et al., *Fine-mapping of the brain-derived neurotrophic factor (BDNF) gene* supports an association of the Val66Met polymorphism with episodic memory. Int J Neuropsychopharmacol, 2010. 13(8): p. 975-80.

### 3.2 KIBRA is associated with accelerated cognitive decline and

### hippocampal atrophy in APOE ɛ4-positive cognitively normal

### adults with high Aβ-amyloid burden

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#### 3.2.1 Abstract

A single nucleotide polymorphism, rs17070145, in the KIdney and BRAin expressed protein (*KIBRA*) gene has been associated with cognition and hippocampal volume in cognitively normal (CN) individuals. However, the impact of rs17070145 on longitudinal cognitive decline and hippocampal atrophy in CN adults at greatest risk of developing Alzheimer's disease is unknown. We investigated the impact rs17070145 has on the rate of cognitive decline and hippocampal atrophy over six years in 602 CN adults, with known brain Aβ-amyloid levels and whether there is an interactive effect with *APOE* genotype. We reveal that whilst limited independent effects of *KIBRA* genotype were observed, there was an interaction with *APOE* in CN adults who presented with high Aβ-amyloid levels across study duration. In comparison to *APOE*  $\varepsilon$ 4-ve individuals carrying the rs17070145-T allele, significantly faster rates of cognitive decline (global, p=0.006; verbal episodic memory, p=0.004;), and hippocampal atrophy (p=0.04) were observed in individuals who were *APOE*  $\varepsilon$ 4+ve and did not carry the rs17070145-T allele, in the presence of high Aβ-amyloid suggest that carriers of the rs17070145-T allele are conferred a level of resilience to the detrimental effects of high Aβ-amyloid and *APOE*  $\varepsilon$ 4.

#### **3.2.2 Introduction**

In cognitively normal older individuals, high levels of neocortical amyloid- $\beta$  (A $\beta$ -amyloid) are associated with subtle but detectable cognitive decline [1] and hippocampal atrophy [2]. This observation is consistent with models of Alzheimer's disease (AD) which propose a protracted preclinical phase that could take up to 20 years [3]. This provides a period of opportunity for understanding, and even interfering with, AD pathogenesis and thus the identification of biological factors, or trait characteristics, that themselves can influence AD progression has become of increased importance.

Several genes have been associated with cognitive performance, particularly episodic memory, and hippocampal atrophy. Previous studies have associated genetic polymorphisms, in particular apolipoprotein E (*APOE*)  $\varepsilon 2/\varepsilon 3/\varepsilon 4$  genotype (see review [4, 5]) and the nonsynonymous rs6265 (Val66Met) SNP in brain derived neurotropic factor (*BDNF*) [6-9], with altered rates of episodic memory decline and hippocampal atrophy. Decline in measures of episodic memory, modified by genetic variation, have been reported in both the healthy elderly [10] and those predicted to be in the early stages of AD based on neocortical Aβ-amyloid imaging [6, 7, 11]. These findings raise the potential that other genetic factors may also moderate the toxic effects of Aβ-amyloid early in AD and contribute to altered rates of cognitive decline and hippocampal atrophy.

One such candidate is the gene encoding the KIdney and BRAin expressed protein (*KIBRA*; sometimes referred to as WW domain-containing protein 1 (*WWC1*)) [12]. KIBRA is a cytoplasmic, signal transducer protein expressed mainly in the kidney and brain [13] and *in vitro* experiments suggest that, through reduction in postsynaptic levels, it mediates tau induced memory loss and disruption of synaptic plasticity [14]. This *in vitro* data is supported through
genetic studies that report the association of allelic variation in the *KIBRA* gene with memory performance, hippocampal atrophy and measurable differences in brain activation. Specifically, a substitution of C for T in the 9<sup>th</sup> intron (rs17070145), was initially identified through a GWAS of verbal episodic memory performance and replicated in two additional independent cohorts [12]. Episodic memory is one of the earliest cognitive domains to decline, with previous studies observing decline 4-8 years prior to executive function and up to 7-10 years prior to other cognitive domains [15-17].

However, there is a lack of consensus in subsequent studies that attempted to replicate these genetic associations with memory performance. Cross-sectional studies of cognitively normal (CN) older adults, carriage of the rs17070145-T allele has been associated with better performance in episodic memory [18-22], delayed recall [23-25] and spatial learning [26] and increased hippocampal volume [20] and activity [19, 24]. Conversely, several studies have either associated the absence of rs17070145-T with better semantic [27] and long-term [28] memory, executive function [29] and overall cognitive performance [30] or were unable to show any association of the SNP with cross sectional episodic memory [29, 31-33] and hippocampal volume [31] or longitudinal decline in episodic memory and hippocampal volume [31]. However, common to all these studies is the lack of inclusion of A $\beta$ -amyloid imaging, which may contribute to the lack of consensus due to the impact of underlying A $\beta$ -amyloid burden on cognition not being considered [1, 6, 7, 11].

To address this conjecture requires the availability of comprehensive longitudinal data from the prospective cohort studies of AD, such as the Australian Imaging, Biomarkers and Lifestyle (AIBL) Study, which offers the opportunity to retrospectively evaluate candidate biological factors (e.g. genetic variation) to determine the impact on progression of AD related phenotypes, such as cognitive decline and hippocampal atrophy. The AIBL Study has now more than six years of serial cognitive and neuroimaging assessments, including Aβ-amyloid and structural imaging, in a group of CN adults collected at 18-month intervals. Therefore, the aim of this study was to characterise, through reporting on 6-years of longitudinal data, the role of *KIBRA* rs17070145 allelic variation in this highly characterised CN adult sample and examine the extent to which this allelic variation is associated with Aβ-amyloid related cognitive decline and atrophy of the hippocampus. The hypothesis was that CN adults who carry the rs17070145-T allele would show a slower rate of memory decline and hippocampal atrophy than those not carrying this allele, though this relationship would be dependent on the presence of a high brain Aβ-amyloid burden and interact with *APOE* genotype.

#### 3.2.3 Methods

#### 3.2.3.1 Participants

This study included 602 CN Caucasian adults enrolled in the AIBL Study, a prospective longitudinal study of ageing. Information regarding the AIBL Study's design, enrolment process, neuropsychological assessments, and diagnostic criteria has been previously described [34]. The clinical classification of CN, MCI or AD was determined, after clinical review, by a panel of old age psychiatrists, geriatricians, neurologists, and neuropsychologists who were blinded to Aβ-amyloid status. Individuals were classified as CN if they did not meet the clinical criteria for diagnosis of MCI [35] or dementia [36], as described previously [34]. Approval of the AIBL Study has been granted by each of the ethics committees of each of the member institutions; Austin Health, St Vincent's Health, Hollywood Private Hospital, and Edith Cowan University, and informed written consent was given by all volunteers. All clinical investigations were conducted in accord with the principles expressed in the Declaration of Helsinki 1975. All participants were assessed every 18-months. Cognitive, neuroimaging and laboratory assessment were acquired within 3-months of each other.

#### 3.2.3.2 Cognitive Measures

The neuropsychological test battery administered in the AIBL study has been described in detail previously [34]. Briefly, it incorporates at each 18-month follow-up, the Mini-Mental State Examination (MMSE), Clock Drawing Test, California Verbal Learning Test-Second edition (CVLT-II), Logical Memory I and II (LMI; LMII; Story A only), D-KEFS verbal fluency, a 30-item version of the Boston Naming Test (BNT), Wechsler Test of Adult Reading (WTAR) for premorbid IQ, Digit Span and Digit Symbol-Coding subtests of the Wechsler Adult Intelligence Scale-Third edition (WAIS-III), the Stroop task (Victoria version), and the Rey Complex Figure Test (RCFT). Resultant data from this battery, in addition to the Clinical

Dementia Rating (CDR), have been previously used to statistically derive cognitive composites as previously described [37]. In this study, a verbal episodic memory composite (CDR sum of boxes (CDR<sub>SB</sub>), LMII, CVLT false positives (CVLT<sub>FP</sub>) and long delay free recall (CVLT<sub>LDFR</sub>)), and a statistically driven global composite (CDR<sub>SB</sub>, MMSE, LMII, CVLT<sub>FP</sub> and Clock), aimed as a sensitive measure for longitudinal decline in individuals predisposed to AD [37], were investigated across five study time points: baseline, 18, 36, 54 and 72 months. A correction for age, gender, years of education, WTAR-estimated premorbid IQ (WAIS-III Full Scale Intelligence Quotient (FSIQ)) and depressive symptoms (Geriatric Depression Scale (GDS)) was incorporated in the calculation of the cognitive composites [38].

#### 3.2.3.3 Brain Imaging

The 602 CN adults included in this study had undergone Aβ-amyloid imaging, at varying time points, with PET using <sup>11</sup>C-Pittsburgh Compound B (PiB), <sup>18</sup>F-florbetapir or <sup>18</sup>F-flutemetamol as previously described [39-41]. PET standardized uptake value (SUV) ratio (SUVR) data was determined for all tracers using using CapAIBL, a web based freely availably MR-less methodology [42]. Briefly, SUVs were summed and normalized to either the cerebellar cortex SUV (PiB), whole cerebellum SUV (florbetapir) or pons SUV (flutemetamol) to yield the target-region to reference-region SUVR. These SUVRs were then classified as either low (Aβ<sup>low</sup>) or high (Aβ<sup>high</sup>) Aβ-amyloid burden, based on a tracer-specific SUVR threshold;  $\geq 1.5$ ,  $\geq 1.10$  and  $\geq 0.62$  for PiB, florbetapir and flutemetamol, respectively, as previously described [43]. Of these 602 participants, 548 also underwent clinical magnetic resonance imaging (MRI) for clinical screening and co-registration with PET images. MRI parameters have been described in detail previously [44]. Briefly, a 3T T1-weighted MRI was performed using the ADNI magnetization-prepared rapid gradient echo protocol, with an in-plane resolution of 1×1 mm and a slice thickness of 1.2 mm. Hippocampal volume was calculated after correcting for age in years and intracranial volume, defined as the sum of grey matter, white matter and cerebrospinal fluid volumes, as previously described [45].

#### 3.2.3.4 Genotyping

DNA extraction from 5mL of whole blood was performed using QIAamp DNA Blood Maxi Kits (Qiagen, Hilden, Germany) according to manufacturer's instructions. TaqMan® genotyping assays were used to determine *APOE* (rs7412, assay ID: C\_\_\_904973\_10; rs429358, assay ID: C\_\_\_3084793\_20) and *KIBRA* (rs17070145, assay ID: C\_\_33286269\_10) genotypes (Life Technologies, Carlsbad, CA). All TaqMan® genotyping assays were performed on a QuantStudio 12K Flex<sup>TM</sup> Real-Time-PCR systems (Applied Biosystems, Foster City, CA) using the TaqMan® GTXpress<sup>TM</sup> Master Mix (Life Technologies) methodology as per manufacturer instructions. *KIBRA* genotype was observed not depart from Hardy-Weinberg equilibrium. For the purpose of this study *APOE* carrier status is defined by the presence (1 or 2 copies) or absence (0 copies) of the *APOE*  $\varepsilon$ 4 allele, henceforth referred to as *APOE*  $\varepsilon$ 4-ve, respectively.

#### 3.2.3.5 Statistical Analyses

All statistical analyses were performed using Rstudio (Rstudio Team 2015) Version 0.98.1103 for Macintosh [46]. All analyses were performed based on a dominant model for the *KIBRA* rs17070145-T (minor) allele, i.e. T carrier (i.e. C\_T and T\_T) compared with non-T carrier (i.e. C\_C), as per previous studies [12, 18-21, 24]. Baseline demographic data analyses provided means, standard deviations, and percentages across the entire PET imaged cognitively normal sample and stratified by *KIBRA* rs17070145-T allele carrier (*KIBRA*-T) and non-carrier (*KIBRA* non-T) status. ANOVA (age, premorbid IQ, depressive symptoms) and chi-squared

tests (gender, years of education, *APOE*  $\epsilon$ 4+ve, high Aβ-amyloid burden) were used to determine the significance of differences between allelic groups. To determine differences in rates of cognitive change and hippocampal atrophy random intercepts linear mixed-effects (LME) models were performed using the "nlme" package in R. LMEs were performed due to their ability to model fixed and random effects, and their robustness when dealing with missing data [47].

After the inclusion of main effects within the model, i.e. *KIBRA* genotype, interaction terms and covariates were included and modelled as described here. Specifically, to investigate the effect of *KIBRA* on the rate of cognitive decline and hippocampal atrophy, initially a *KIBRA*×Time interaction was modelled across the entire sample, covarying for *APOE*  $\varepsilon$ 4 carrier and Aβ-amyloid status, with the cognitive composites and hippocampal volume as the dependent variables. The effect of Aβ status in combination with *KIBRA* was then investigated by separately modelling an Aβ×*KIBRA*×Time interaction, co-varying for *APOE*  $\varepsilon$ 4 carrier status. The third analysis focused on only Aβ<sup>high</sup> participants, with *APOE* included within an *APOE*×*KIBRA*×Time interaction. In addition, all analyses for hippocampal atrophy co-varied for gender. Graphical representations of all models are presented with time dependent standard error. Further, for all analyses correction for the False Discovery Rate (FDR) using Q-Value (bootstrap method) was performed [48]. Finally, chi-squared analyses were performed between groups to ascertain that group differences in rates of decline were not due to disproportionate rates of clinical conversion over the course of the study.

#### 3.2.4 Results

## 3.2.4.1 The effect of KIBRA on cognition and hippocampal atrophy in cognitively normal adults

A total of 602 CN older adults, defined through the AIBL battery of clinical and neuropsychological assessments [34] were included in this study. As shown in Table 3.2.1 there were no significant differences or trends between rs17070145 (henceforth referred to simply as KIBRA) T carriers and non-T carriers at baseline with respect to demographic variables, premorbid intellect, depressive symptoms, or genotype. In the initial analysis, covaried for APOE  $\varepsilon 4$  carrier and A $\beta$ -amyloid status (classified by being above (A $\beta^{high}$ ) or below  $(A\beta^{low})$  Positron Emission Tomography (PET) A $\beta$ -amyloid tracer-specific thresholds) there were no significant differences in the trajectories between T carriers and non-carriers for measures of global cognition or episodic memory amongst CN adults (Supplementary Data; Figure S3.2.3, Table S3.2.4). However, there was a trend towards T-carriers having a mild improvement (0.028 standard deviations (SD)/year) in both global cognition (non-T carriers, -0.025SD/year; p=0.051) and verbal episodic memory (non-T carriers, -0.019SD/year; p=0.085), likely due to a practice effect. When evaluating the effect of *KIBRA* on hippocampal atrophy in all cases, and co-varying for APOE ε4 carrier and Aβ-amyloid status, no significant difference (p=0.242) was observed between T carriers (-0.017 cm<sup>3</sup>/year), and non-T carriers (-0.026 cm<sup>3</sup>/year) over six years (Supplementary Data; Figure S3.2.3, Table S3.2.4). Further, no significant differences were observed at baseline in any measures of cognition or hippocampal volume.

		Overall n = 602	<i>KIBRA</i> T carrier n = 335	<i>KIBRA</i> non-T carrier n = 267	р
Age (years)		70.79 (6.55)	70.73 (6.49)	70.72 (6.41)	0.9788
Female (%)		334 (55.48)	188 (56.12)	146 (54.68)	0.7871
Years of Education	0-8	48 (8.00)	27 (8.08)	21 (7.89)	0.9419
	9-12	222 (37.00)	127 (38.02)	95 (35.71)	
	13-15	126 (21.00)	69 (20.66)	57 (21.43)	
	15+	204 (34.00)	111 (33.23)	93 (34.96)	
Premorbid IQ	(FSIQ)	107.86 (7.23)	107.66 (7.28)	108.14 (7.30)	0.4311
Depressive Symptoms (GDS)		1.05 (1.28)	1.05 (1.35)	1.04 (1.18)	0.9156
APOE ɛ4 carriage (%)		165 (27.97)	84 (25.53)	81 (31.03)	0.1655
High Aβ-amyloid burden (%)		145 (24.09)	76 (22.69)	69 (25.84)	0.4215
MRI (n)		548	301	247	NA

## Table 3.2.1 Demographic Information

Baseline demographic and clinical characteristics of all imaged cognitively normal adults in the AIBL study, and based on *KIBRA* rs17070145 T carriage (T\_T and C\_T) and non-carriage (C\_C). p values represent statistical significance when comparing T carriage and non-carriage. GDS, Geriatric Depression Scale; FSIQ, Wechsler Adult Intelligence Scale 3<sup>rd</sup> Edition (WAIS-III) Full Scale Intelligence Quotient.

No significant differences were observed at baseline in either measure of cognition or hippocampal volume when investigating the  $A\beta \times KIBRA \times Time$  interaction. Relative to  $A\beta^{low}/KIBRA$  T carriers, the  $A\beta^{high}/KIBRA$  non-T carrier group showed a significantly greater rate of decline in global cognition (0.037 SD/year; -0.085 SD/year; p=0.008, q=0.036), and the verbal episodic memory (0.033 SD/year; -0.080SD/year; p=0.012, q=0.042) (Figure 3.2.1, Table 3.2.2). However, no statistical difference was seen between  $A\beta^{high}/KIBRA$  T carriers and  $A\beta^{low}/KIBRA$  non-T carriers. Analysis of hippocampal atrophy revealed that relative to  $A\beta^{low}/KIBRA$  T carriers (-0.015 cm<sup>3</sup>/year), the  $A\beta^{high}/KIBRA$  non-T carrier group (-0.055 cm<sup>3</sup>/year) showed a significantly greater rate of hippocampal atrophy (p=0.002, q=0.034) over six years (Figure 3.2.1, Table 3.2.2). Likewise, this trajectory of hippocampal atrophy was also significantly different (p=0.009, q=0.034) relative to  $A\beta^{low}/KIBRA$  non-T carriers (-0.017 cm<sup>3</sup>/year). In contrast,  $A\beta^{high}/KIBRA$  T carriers' rate of atrophy did not differ from the  $A\beta^{low}$  groups.



Figure 3.2.1 Rates of change in cognitively normal adults based on KIBRA T carriage and Aβ-amyloid status.

Rates of change are presented for (a) a statistically driven global composite, (b) a verbal episodic memory composite, and (c) hippocampal atrophy (n=548) in cognitively normal adults (n=602 unless otherwise stated).  $A\beta^{low}$ , low  $A\beta$ -amyloid burden;  $A\beta^{high}$ , high  $A\beta$ -amyloid burden.  $A\beta^{low}/KIBRA$  T carriers (green),  $A\beta^{low}/KIBRA$  non-T carriers (blue),  $A\beta^{high}/KIBRA$  T carriers (orange),  $A\beta^{high}/KIBRA$  non-T carriers (red), controlling for *APOE*  $\epsilon$ 4 carrier status. Hippocampal atrophy analysis also controlled for gender (shading represents time dependent standard error, \*p<0.05 when comparing to the  $A\beta^{low}/KIBRA$  T carrier group,  $^p$ <0.05 when comparing to the  $A\beta^{high}/KIBRA$  T carrier).

	Αβ <sup>low</sup> <i>KIBRA</i> T carrier n=259	Αβ <sup>low</sup> <i>KIBRA</i> non-T carrier n=198	Αβ <sup>high</sup> KIBRA T carrier n=76	Αβ <sup>high</sup> <i>KIBRA</i> non-T carrier n=69
	β	β	β	β
Global	0.037	-0.006	-0.012	-0.085*
Verbal Episodic Memory	0.033	0.0004	0.005	-0.080*
Hippocampal Atrophy	-0.015	-0.017	-0.026	-0.055*^

Table 3.2.2 Group slopes for cognitive composites and hippocampal atrophy in all imaged cognitively normal participants by KIBRA carrier and Aβ-amyloid status Group slopes for cognitive composites (presented in SD/year; n=602) and hippocampal atrophy (presented in cm<sup>3</sup>/year; n=548) in all imaged cognitively normal participants, controlling for *APOE* ε4 carrier status. Aβ<sup>low</sup>, low Aβ-amyloid burden; Aβ<sup>high</sup>, high Aβ-amyloid burden. \*p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA T carrier (T\_T and C\_T) group, ^p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA non-T carrier group,  $\varphi$  p<0.05 when comparing to the Aβ<sup>low</sup>/KIBRA T carrier

# 3.2.4.2 The effect of KIBRA on cognition and hippocampal atrophy in cognitively normal adults with high Aβ-amyloid

No significant differences were observed in  $A\beta^{high}$  CN adults at baseline in either measure of cognition or hippocampal volume when investigating the APOE×KIBRA×Time interaction. Relative to APOE £4-ve/KIBRA T carriers, the APOE £4+ve/KIBRA non-T carrier group showed a significantly greater rate of decline in global cognition (p=0.006, q=0.034) and verbal episodic memory (p=0.004, q=0.034) over six years (Figure 3.2.2, Table 3.2.3). Further, relative to APOE £4+ve/KIBRA T carriers, the APOE £4+ve/KIBRA non-T carrier group showed a nominally significantly greater rate of decline on the verbal episodic memory composite, however after FDR correction this remained only a strong trend (p=0.018, q=0.055) over six years (Figure 3.2.2, Table 3.2.3). Hippocampal atrophy analysis revealed that relative to APOE £4-ve/KIBRA T carriers (-0.016 cm<sup>3</sup>/year), the APOE £4+ve/KIBRA non-T carrier group (-0.067 cm<sup>3</sup>/year) had nominally significantly different rates of hippocampal atrophy however did not survive correction for multiple testing (p=0.040, q=0.107) over six years (Figure 3.2.2, Table 3.2.3). This trajectory of hippocampal atrophy was suggestive of being different to APOE £4-ve/KIBRA non-T carriers (-0.006 cm<sup>3</sup>/year), however this did not reach significance (p=0.125), even though this trajectory showed negligible difference to APOE  $\varepsilon$ 4ve/KIBRA T carriers. APOE ɛ4+ve/KIBRA T carriers' rate of atrophy did not differ from the APOE E4-ve groups. To ascertain that these differences in rates of decline were not due to disproportionate rates of clinical conversion, the frequency of individuals who converted to Mild Cognitive Impairment (MCI) or AD over the course of the study was investigated. Within the APOE  $\varepsilon$ 4+ve group there was no significant difference (p=0.43) between KIBRA non-T carriers (0.294, 15 out of 41) and KIBRA T carriers (0.294, 10 out of 34) in terms of clinical conversion.



Figure 3.2.2 Rates of change in cognitively normal adults with high Aβ-amyloid burden

Rates of change are presented for (a) a statistically driven global composite, (b) a verbal episodic memory composite, (c) hippocampal atrophy in cognitively normal adults with high A $\beta$ -amyloid (n=145). *APOE*  $\epsilon$ 4-negative/*KIBRA* T carriers (green), *APOE*  $\epsilon$ 4-ve/*KIBRA* non-T carriers (blue), *APOE*  $\epsilon$ 4+ve/*KIBRA* T carriers (orange), *APOE*  $\epsilon$ 4+ve/*KIBRA* non-T carriers (red). Hippocampal atrophy analysis controlled for gender (shading represents time dependent standard error, \*p<0.05 when comparing to the *APOE*  $\epsilon$ 4-ve/*KIBRA* T carrier group, ^p<0.05 when comparing to the *APOE*  $\epsilon$ 4+ve/*KIBRA* T carrier).

	<i>APOE</i> ε4-ve <i>KIBRA</i> T carrier n=38	<i>APOE</i> ε4-ve <i>KIBRA</i> non-T carrier n=27	<i>APOE</i> ε4+ve <i>KIBRA</i> T carrier n=34	<i>APOE</i> ε4+ve <i>KIBRA</i> non-T carrier n=40
	β	β	β	β
Global	-0.016	-0.014	-0.063	<b>-0.163*</b> ^†
Verbal Episodic Memory	-0.008	-0.019	-0.031	-0.146*^φ†
Hippocampal Atrophy	-0.016	-0.006	-0.034	-0.067*

Table 3.2.3 Group slopes for cognitive composites and hippocampal atrophy in imaged cognitively normal adults with high  $A\beta$ -amyloid.

Group slopes for cognitive composites (presented in SD/year) and hippocampal atrophy (presented in cm<sup>3</sup>/year) in imaged cognitively normal adults with high Aβ-amyloid (n=145). \*p<0.05 when comparing to the *APOE* ε4-ve/*KIBRA* T carrier group, ^p<0.05 when comparing to the *APOE* ε4-ve/*KIBRA* T carrier group,  $\phi$  p<0.05 when comparing to the *APOE* ε4-ve/*KIBRA* T carrier group,  $\phi$  p<0.05 when comparing to the *APOE* ε4-ve/*KIBRA* T carrier group,  $\phi$  p<0.05 when comparing to the *APOE* ε4-ve/*KIBRA* T carrier group,  $\phi$  p<0.05 when comparing to the *APOE* ε4-ve/*KIBRA* T carrier. †q<0.05 for those reporting nominal significance at p<0.05.

#### **3.2.5 Discussion**

The data reported here support the hypothesis that *KIBRA* genotype, in combination with *APOE*  $\varepsilon$ 4 and Aβ-amyloid, affects rates of memory decline and hippocampal atrophy in cognitively normal adults. In those CN adults with high Aβ-amyloid burden at baseline, *KIBRA* non-T carriers showed significantly faster decline in the statistically driven global composite, and verbal episodic memory when compared to T carriers with low Aβ-amyloid burden. Within the subset of CN adults with high Aβ-amyloid burden, we showed that those who are *APOE*  $\varepsilon$ 4+ve and *KIBRA* non-T carriers had significantly faster rates of decline in verbal episodic memory over 6 years, compared to *APOE*  $\varepsilon$ 4+ve/*KIBRA* T carrier and both *APOE*  $\varepsilon$ 4-ve groups. Importantly, minimal decline was also observed in the *APOE*  $\varepsilon$ 4+ve/*KIBRA* T carrier group, suggesting that carriage of the *KIBRA* T allele imparts a level of resilience to negative effects of *APOE*  $\varepsilon$ 4 and Aβ-amyloid on memory performance. Further, between group comparisons of the rates of clinical conversion (CN>MCI/AD) over the course of the study revealed no significant differences, suggesting that the faster rates of decline were not due to a higher rate of clinical conversion.

This is further supported by the observations that rates of hippocampal atrophy in this study also differ based on *KIBRA* genotype. In CN adults Aβ-amyloid has been previously reported to be associated with increased hippocampal atrophy [2, 45, 49], however in this study this was only observed in those individuals who did not possess the *KIBRA* T-allele, whilst in contrast *KIBRA* T-carriers' rate of atrophy did not significantly differ from the Aβ<sup>low</sup> groups. In a metaanalysis of *APOE* neuroimaging studies, hippocampal atrophy has been shown to be increased in *APOE* ε4 carriers [5]. Here we report that this association, in a group of Aβ<sup>high</sup> CN individuals, was again only observed in those individuals who did not possess the *KIBRA* Tallele, whilst in contrast *APOE* ε4+ve/*KIBRA* T-carriers' rate of atrophy did not differ from the *APOE*  $\varepsilon$ 4-ve groups. Taken together, we propose that the *KIBRA* T allele affords carriers a level of resilience to the detrimental effects of A $\beta$ -amyloid and *APOE*  $\varepsilon$ 4 allele on neurodegeneration, specifically hippocampal atrophy.

The findings presented herein are in line with the original study [12] and subsequent reports linking the *KIBRA* T allele with resilience in episodic memory performance [18-21, 24]. The absence of replication by other studies [27-29, 31-33] may be in part due to the lack of consistency in the measures of memory decline, whereby varying single neuropsychological tests, aiming to measure a certain feature of memory or cognition, were used. The use in this current study of a combination of global and episodic memory composite scores, which encompass several different tests best associated with a cognitive construct, could also have contributed to the ability to detect associations with the *KIBRA* genotype. However, the lack of inclusion of an assessment of underlying Aβ-amyloid burden in the previous studies may in fact be the more telling contributor to the lack of consensus on the association of *KIBRA* with cognitive performance. The level of neocortical Aβ-amyloid is associated with differential rates of cognitive decline [1, 50], and this is further altered by genetic factors, in particular *APOE* [10, 11] and *BDNF* [6, 7]. Accounting for the underlying Aβ-amyloid burden in the current study may have further contributed to the detection of differences in rates of cognitive decline and hippocampal atrophy reported with *APOE* ε4 and *KIBRA*.

Whilst the incorporation of cognitive composites and accounting for underlying A $\beta$ -amyloid burden is considered a strength of this study, the following limitations of the study are acknowledged. Firstly, the use of different cognitive tests individually or in combination for the calculation of domain composites, then those specifically described in this study and using the methodology described herein, may yield different results. Second, this study included 6years of longitudinal follow-up and validation in other longitudinal cohorts, not undertaken herein, over longer durations of follow-up, may result in different findings. Third, the cognitively normal participants in this study were volunteers and not selected at random from the community, they were generally well educated and performed well on cognitive assessments and as such the findings presented herein may be applicable only to similar cohorts. Fourth, there is an overlap between those who are  $A\beta^{high}$  and those who are *APOE*  $\epsilon$ 4+ve, which could confound the results when looking at their interaction. Finally, the *KIBRA* T-allele's previously reported association with altered brain activation using functional MRI (fMRI) [12, 19] could not be tested due to the lack of fMRI data, under a non-resting state, in the AIBL Study.

Studies have previously demonstrated the main areas of *KIBRA* expression in the brain are those also that are implicated in memory function, the hippocampus and temporal cortex [12, 51]. Furthermore, increased *KIBRA* gene expression in the temporal cortex [52] and hippocampus [22] has been associated with late onset AD. However, in a recent post-mortem brain transcriptomic study in neuropathogically normal individuals by Piras and colleagues a trend towards increased *KIBRA* gene expression was observed in *KIBRA* T homozygotes [53]. Further quantitative PCR analysis reported an over-expression in T-homozygotes compared to C-homozygotes in the hippocampus [53]. Further, the transcriptomic analysis revealed differential activation of the mitogen-activated protein kinase (MAPK) pathway [53], a pathway important in learning and memory processes, suggesting a potential mechanism underpinning a decline in memory performance reported in this study. It has also been shown that there is increased hippocampal activity in episodic memory performance tasks in *KIBRA* T carriers when compared with non-T carriers [19], consistent with the notion of protection from memory decline. *KIBRA* T allele carriers have also been shown to have a decreased levels

of brain activation compared to non-T allele carriers in several hippocampal regions activated during memory retrieval [12]. The authors hypothesised that individuals who do not carry the T allele require a greater level of hippocampal activation for memory retrieval [12].

In addition to the association studies described above, recent in vivo evidence provides molecular insights into mechanisms by which KIBRA is involved in memory performance. Synaptic plasticity, which is altered in AD, is modulated by dendrin, which in turn binds to the protein that KIBRA encodes (KIBRA; see review [54]). Further, KIBRA protein contains a protein kinase C (isoform  $\zeta$ ; PKC $\zeta$ ) binding domain [55] and has been reported to co-localise with protein kinase M (isoform  $\zeta$ ; PKM $\zeta$ ) [56], a brain specific variant of PKC $\zeta$ , which plays important roles in memory formation and long-term potentiation. Johannsen et al have shown the function of the KIBRA protein to be regulated by its C2 domain [51], which is required for Ca<sup>2+</sup> binding and is therefore involved in signal transduction in the neurons. This regulation is hypothesised to mediate the effect of the KIBRA protein on memory formation [51]. In a recent study, Tracy and colleagues have proposed a novel mechanism by which acetylated tau associated memory loss and disruption of synaptic plasticity is mediated by a reduction in postsynaptic KIBRA protein [14]. This finding links the previous reports of reduced KIBRA gene expression in AD with a biological mechanism mediated by acetylated *tau*. Whether the KIBRA T allele affords a level of resilience to this loss of synaptic plasticity remains to be determined.

Our findings indicate that *KIBRA* rs17070145 genotype, when combined with high brain Aβamyloid burden and *APOE*  $\varepsilon$ 4 carriage, modifies longitudinal rates of decline in verbal episodic memory, a global cognitive composite and hippocampal volume. We propose that early in the disease process of AD, carriers of the *KIBRA* T-allele are conferred a level of resilience to Aβamyloid and *APOE*  $\varepsilon$ 4 driven decline. The potential mechanisms by which *KIBRA* contributes to synaptic plasticity, and AD progression warrant further investigation, including the potential impact on Aβ-amyloid accumulation, and may reveal novel pathways contributing to neuroprotection/neurodegeneration. Our results also highlight the potential application of genetics for risk stratification when designing clinical trials, particularly those that employ Aβamyloid imaging for screening. The nature of the effects of genetic variations, specifically assessing the combined effect(s) of additional genes affecting cognitive performance would have merit in such settings and requires further investigation.

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#### **3.2.7** Competing Financial Interests

CLM is an advisor to Prana Biotechnology Ltd and a consultant to Eli Lilly. PM is a full-time employee of Cogstate Ltd. DA has served on scientific advisory boards for Novartis, Eli Lilly, Janssen, and Pfizer Inc. RNM is a consultant to Alzhyme. SML has previously been a paid consultant to Alzhyme. CCR has served on scientific advisory boards for Bayer Pharma, Elan Corporation, GE Healthcare and AstraZeneca; has received speaker honoraria from Bayer Pharma and GE Healthcare; and has received research support from Bayer Pharma, GE Healthcare, Piramal Lifesciences and Avid Radiopharmaceuticals. VLV served as a consultant for Bayer Pharma; and received research support from a NEDO grant from Japan. All other authors have nothing to disclose.

#### 3.2.8 Author Contribution Statement

TP contributed to acquisition of genetic data, statistical analysis, interpretation of findings, drafting the manuscript. SCB contributed to specific study concept and design, study supervision, statistical analysis, interpretation of findings, and drafting of the manuscript. VD, PB contributed to acquisition and analysis of imaging data and revising the manuscript. GS contributed to AIBL study design, obtaining funding, interpretation of findings. KB, LM contributed to acquisition of genetic data. DA, AIB, CLM, CCR, RNM contributed to AIBL study design, obtaining funding and revising the manuscript. PM contributed to AIBL study design, obtaining funding, revising the manuscript. SRS contributed to revising the manuscript. DG, GV contributed to study supervision and revising the manuscript. VLV contributed to current study concept and design, obtaining funding, study supervision, acquisition of data, interpretation of findings and drafting of the manuscript. SML contributed to current study concept and design, obtaining funding, study supervision, acquisition of data, interpretation of findings and drafting of the manuscript. All authors read and approved the final manuscript.

#### 3.2.9 Data Availability

All data and samples used in this study are derived from the Australian Imaging, Biomarkers and Lifestyle (AIBL) Study of Ageing. All AIBL data, and that specific to this study, is publically accessible to all interested parties through an Expression of Interest procedure and is governed by the AIBL Data Use Agreement, for more information please see https://aibl.csiro.au/awd/.

#### 3.2.11 Supplementary Data



Figure S3.2.3 Rates of change in cognitively normal adults based on KIBRA T carriage.

Rates of change are presented for (a) a statistically driven global composite, (b) a verbal episodic memory composite, (c) hippocampal atrophy (n=548) in cognitively normal adults (n=602 unless otherwise stated). *KIBRA* T carriers (grey) and non-T carriers (black), controlling for *APOE*  $\epsilon$ 4 carrier and Aβ-amyloid status. Hippocampal atrophy analysis also controlled for gender (shading represents time dependent standard error, \* p<0.05).

	KIBRA T carrier	KIBRA non-T carrier	
	β	β	р
Global	0.028	-0.025	0.051
Verbal Episodic Memory	0.028	-0.019	0.085
Hippocampal Atrophy	-0.017	-0.026	0.242

Table S3.2.4 Group slopes for cognitive composites and hippocampal atrophy in all imaged cognitively normal participants by KIBRA carrier status

Group slopes for cognitive composites (presented in SD/year; n=602) and hippocampal atrophy (presented in cm<sup>3</sup>/year; n=548) in all imaged cognitively normal participants, controlling for *APOE*  $\epsilon$ 4 carrier and A $\beta$ -amyloid status. \*Represents a nominally statistically significant difference in slope of the *KIBRA* non-T carrier (C\_C) group when compared to the *KIBRA* T carrier (T\_T and C\_T) group.

#### 3.2.10 References

- 1. Villemagne, V.L., et al., *Longitudinal assessment of Abeta and cognition in aging and Alzheimer disease*. Ann Neurol, 2011. **69**(1): p. 181-92.
- Andrews, K.A., et al., *Atrophy rates in asymptomatic amyloidosis: implications for Alzheimer prevention trials.* PLoS One, 2013. 8(3): p. e58816.
- Villemagne, V.L., et al., *Amyloid beta deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study.* Lancet Neurol, 2013. 12(4): p. 357-67.
- 4. El Haj, M., et al., *Apolipoprotein E (APOE) epsilon4 and episodic memory decline in Alzheimer's disease: A review.* Ageing Res Rev, 2016. **27**: p. 15-22.
- Liu, Y., et al., APOE genotype and neuroimaging markers of Alzheimer's disease: systematic review and meta-analysis. J Neurol Neurosurg Psychiatry, 2015. 86(2): p. 127-34.
- 6. Lim, Y.Y., et al., *BDNF Val66Met, Abeta amyloid, and cognitive decline in preclinical Alzheimer's disease*. Neurobiol Aging, 2013. **34**(11): p. 2457-64.
- Lim, Y.Y., et al., APOE and BDNF polymorphisms moderate amyloid beta-related cognitive decline in preclinical Alzheimer's disease. Mol Psychiatry, 2015. 20(11): p. 1322-8.
- 8. Kennedy, K.M., et al., *BDNF val66met polymorphism affects aging of multiple types of memory*. Brain Res, 2015. **1612**: p. 104-17.
- 9. Cathomas, F., et al., *Fine-mapping of the brain-derived neurotrophic factor (BDNF)* gene supports an association of the Val66Met polymorphism with episodic memory.
  Int J Neuropsychopharmacol, 2010. 13(8): p. 975-80.

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- Lim, Y.Y., et al., *Abeta amyloid, cognition, and APOE genotype in healthy older adults*. Alzheimers Dement, 2013. 9(5): p. 538-45.
- 11. Lim, Y.Y., et al., *APOE epsilon4 moderates amyloid-related memory decline in preclinical Alzheimer's disease*. Neurobiol Aging, 2015. **36**(3): p. 1239-44.
- Papassotiropoulos, A., et al., *Common Kibra alleles are associated with human memory performance*. Science, 2006. **314**(5798): p. 475-8.
- Kremerskothen, J., et al., *Characterization of KIBRA, a novel WW domain-containing protein*. Biochem Biophys Res Commun, 2003. **300**(4): p. 862-7.
- 14. Tracy, T.E., et al., *Acetylated Tau Obstructs KIBRA-Mediated Signaling in Synaptic Plasticity and Promotes Tauopathy-Related Memory Loss.* Neuron, 2016.
- 15. Elias, M.F., et al., *The preclinical phase of Alzheimer disease: a 22-year prospective study of the Framingham Cohort.* Archives of neurology, 2000. **57**(6): p. 808-813.
- Grober, E., et al., *Memory impairment, executive dysfunction, and intellectual decline in preclinical Alzheimer's disease*. Journal of the International Neuropsychological Society, 2008. 14(2): p. 266.
- 17. Derby, C.A., et al., *Screening for predementia AD Time-dependent operating characteristics of episodic memory tests.* Neurology, 2013. **80**(14): p. 1307-1314.
- Almeida, O.P., et al., *KIBRA genetic polymorphism influences episodic memory in later life, but does not increase the risk of mild cognitive impairment.* J Cell Mol Med, 2008. 12(5A): p. 1672-6.
- Kauppi, K., et al., *KIBRA polymorphism is related to enhanced memory and elevated hippocampal processing*. J Neurosci, 2011. **31**(40): p. 14218-22.

- 20. Witte, A.V., et al., *Impact of KIBRA Polymorphism on Memory Function and the Hippocampus in Older Adults*. Neuropsychopharmacology, 2016. **41**(3): p. 781-90.
- 21. Yasuda, Y., et al., *Association study of KIBRA gene with memory performance in a Japanese population*. World J Biol Psychiatry, 2010. **11**(7): p. 852-7.
- 22. Corneveaux, J.J., et al., *Evidence for an association between KIBRA and late-onset Alzheimer's disease*. Neurobiol Aging, 2010. **31**(6): p. 901-9.
- Bates, T.C., et al., Association of KIBRA and memory. Neurosci Lett, 2009. 458(3): p. 140-3.
- 24. Muse, J., et al., *WWC1 genotype modulates age-related decline in episodic memory function across the adult life span.* Biol Psychiatry, 2014. **75**(9): p. 693-700.
- 25. Schaper, K., et al., *KIBRA gene variants are associated with episodic memory in healthy elderly*. Neurobiol Aging, 2008. **29**(7): p. 1123-5.
- 26. Schuck, N.W., et al., *Aging and KIBRA/WWC1 genotype affect spatial memory processes in a virtual navigation task.* Hippocampus, 2013. **23**(10): p. 919-30.
- Laukka, E.J., et al., *Genetic effects on old-age cognitive functioning: a population*based study. Psychol Aging, 2013. 28(1): p. 262-74.
- 28. Nacmias, B., et al., *KIBRA gene variants are associated with episodic memory performance in subjective memory complaints.* Neurosci Lett, 2008. **436**(2): p. 145-7.
- Wersching, H., et al., *Impact of common KIBRA allele on human cognitive functions*.
   Neuropsychopharmacology, 2011. 36(6): p. 1296-304.
- 30. Liu, J.J., et al., *KIBRA genetic polymorphism and cognitive dysfunction in depression*.
   Psychiatry Res, 2015. 226(1): p. 405-6.

- 31. Boraxbekk, C.J., et al., *Investigating the influence of KIBRA and CLSTN2 genetic* polymorphisms on cross-sectional and longitudinal measures of memory performance and hippocampal volume in older individuals. Neuropsychologia, 2015. **78**: p. 10-7.
- 32. Franks, K.H., M.J. Summers, and J.C. Vickers, *KIBRA gene polymorphism has no association with verbal or visual episodic memory performance*. Front Aging Neurosci, 2014. **6**: p. 270.
- 33. Need, A.C., et al., *Failure to replicate effect of Kibra on human memory in two large cohorts of European origin*. Am J Med Genet B Neuropsychiatr Genet, 2008.
  147B(5): p. 667-8.
- 34. Ellis, K.A., et al., *The Australian Imaging, Biomarkers and Lifestyle (AIBL) study of aging: methodology and baseline characteristics of 1112 individuals recruited for a longitudinal study of Alzheimer's disease.* Int Psychogeriatr, 2009. **21**(4): p. 672-87.
- 35. Winblad, B., et al., *Mild cognitive impairment--beyond controversies, towards a consensus: report of the International Working Group on Mild Cognitive Impairment.*Journal of Internal Medicine, 2004. 256(3): p. 240-246.
- 36. McKhann, G., et al., *Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease.* Neurology, 1984. **34**(7): p. 939-944.
- Burnham, S.C., et al., Novel Statistically-Derived Composite Measures for Assessing the Efficacy of Disease-Modifying Therapies in Prodromal Alzheimer's Disease Trials: An AIBL Study. J Alzheimers Dis, 2015. 46(4): p. 1079-89.
- Burnham, S.C., et al., *Comparision of three normative data correction approaches: A cross-sectional evaluation in the AIBL study*. Alzheimer's & Dementia, 2014. 10(4):
  p. P4-293.

- 39. Rowe, C.C., et al., *Amyloid imaging results from the Australian Imaging, Biomarkers and Lifestyle (AIBL) study of aging.* Neurobiol Aging, 2010. **31**(8): p. 1275-83.
- 40. Clark, C.M., et al., *Use of florbetapir-PET for imaging beta-amyloid pathology*.
  JAMA, 2011. **305**(3): p. 275-83.
- 41. Vandenberghe, R., et al., *18F-flutemetamol amyloid imaging in Alzheimer disease and mild cognitive impairment: a phase 2 trial.* Ann Neurol, 2010. **68**(3): p. 319-29.
- 42. Bourgeat, P., et al., *Comparison of MR-less PiB SUVR quantification methods*. Neurobiol Aging, 2015. 36 Suppl 1: p. S159-66.
- 43. Rowe, C.C., et al., *Predicting Alzheimer disease with beta-amyloid imaging: results from the Australian imaging, biomarkers, and lifestyle study of ageing.* Ann Neurol, 2013. 74(6): p. 905-13.
- 44. Bourgeat, P., et al., *Beta-amyloid burden in the temporal neocortex is related to hippocampal atrophy in elderly subjects without dementia*. Neurology, 2010. 74(2): p. 121-7.
- 45. Dore, V., et al., *Cross-sectional and longitudinal analysis of the relationship between Abeta deposition, cortical thickness, and memory in cognitively unimpaired individuals and in Alzheimer disease.* JAMA Neurol, 2013. **70**(7): p. 903-11.
- 46. RStudio Team, *RStudio: Integrated Development for R*, I. RStudio, Editor. 2015: Boston, MA.
- 47. Gueorguieva, R. and J.H. Krystal, *Move over ANOVA: progress in analyzing* repeated-measures data and its reflection in papers published in the Archives of General Psychiatry. Arch Gen Psychiatry, 2004. **61**(3): p. 310-7.

- Storey, J.D., *A direct approach to false discovery rates*. Journal of the Royal Statistical Society: Series B (Statistical Methodology), 2002. 64(3): p. 479-498.
- 49. Andrews, K.A., et al., *Acceleration of hippocampal atrophy rates in asymptomatic amyloidosis*. Neurobiol Aging, 2016. **39**: p. 99-107.
- 50. Lim, Y.Y., et al., *Stronger effect of amyloid load than APOE genotype on cognitive decline in healthy older adults*. Neurology, 2012. **79**(16): p. 1645-52.
- 51. Johannsen, S., et al., *Temporal-spatial expression and novel biochemical properties of the memory-related protein KIBRA*. Neuroscience, 2008. **155**(4): p. 1165-73.
- 52. Burgess, J.D., et al., *Association of common KIBRA variants with episodic memory and AD risk.* Neurobiol Aging, 2011. **32**(3): p. 557 e1-9.
- 53. Piras, I.S., et al., *Whole transcriptome profiling of the human hippocampus suggests an involvement of the KIBRA rs17070145 polymorphism in differential activation of the MAPK signaling pathway.* Hippocampus, 2017. **27**(7): p. 784-793.
- 54. Schwab, L.C., et al., *Effects of the KIBRA Single Nucleotide Polymorphism on Synaptic Plasticity and Memory: A Review of the Literature*. Curr Neuropharmacol, 2014. 12(3): p. 281-8.
- Buther, K., et al., *KIBRA is a novel substrate for protein kinase Czeta*. Biochem Biophys Res Commun, 2004. **317**(3): p. 703-7.
- 56. Yoshihama, Y., et al., *KIBRA Co-localizes with protein kinase Mzeta (PKMzeta) in the mouse hippocampus.* Biosci Biotechnol Biochem, 2009. **73**(1): p. 147-51.

## 3.3 SPON1 rs11023139 is associated with Aβ-amyloid and APOE

## ε4 related cognitive decline in cognitively normal adults.

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## **3.4** *COMT* val158met is not associated with Aβ-amyloid and

## APOE ε4 related cognitive decline in cognitively normal older

### adults

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## 3.5 Klotho allele status is not associated with Aβ and APOE ε4

## related cognitive decline in preclinical Alzheimer's disease

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## **3.6 Epilogue**

The preceding studies presented in this chapter aimed to investigate the effect of variants within a targeted set of genes, with *a priori* evidence of association with cognitive performance, on cognition in a preclinical AD cohort. When investigating the *KIBRA* and *SPON1* variations, no independent associations were observed with cognitive composite measures. However, after interaction with *APOE*  $\varepsilon$ 4 in individuals with high levels of neocortical amyloid beta (Aβ) significant associations were observed. No significant associations were observed, independently or with interaction, when investigating *KL* or *COMT*. Taken together the studies presented in this chapter, along with previous associations of *BDNF* with preclinical cognitive decline, provide strong evidence to support the inclusion of genetic variants over and above those associated with the clinical diagnosis of AD in polygenic risk scores (PRSs). Further, the biased inclusion of AD risk variants may potentially be at the detriment of the performance of the PRS, particularly in the prediction of cognitive decline in preclinical AD.

Although significant associations were observed for a subset of variants studied, these analyses were in isolation and as such does not completely discount the influence of these variants. Specifically, the contribution of a genetic variant to influencing preclinical cognitive decline may be apparent when considered in combination with other variants rather than as an independent effect. The next chapters explore different methods of combining these variants. In Chapter 4, genetic variants associated with cognitive decline are combined. Then, in Chapter 5, an approach is taken to combine both cognitive performance and AD risk associated variants such that it would allow for the accurate prediction of cognitive decline in preclinical AD.
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# CHAPTER 4: Assessing the utility of combining a priori candidate, cognition associated, genes for predicting cognitive decline in preclinical AD.

# **4.1 Prologue**

The studies presented in Chapter 3 provided the characterisation of a targeted selection of genes with *a priori* evidence of association with cognitive performance. Specifically, the independent and interactional effects (with *APOE*  $\epsilon$ 4) of the cognitive gene variants on rates of cognitive decline were explored in cognitively normal older adults in the Australian Imaging Biomarkers and Lifestyle (AIBL) Study of Aging. These studies were the first to investigate the influence of these genetic variants on cognition in a preclinical Alzheimer's disease (AD) cohort, defined using Aβ-Amyloid (Aβ) brain imaging.

Of the four candidates investigated in Chapter 3, two reported to have significant associations with longitudinal cognitive performance. That is, in cognitively normal individuals with high levels of neocortical A $\beta$  and carrying at least one copy of the *APOE*  $\varepsilon$ 4 allele, variants within *SPON1* and *KIBRA* were associated with significantly difference rates of cognitive decline. Conversely, when investigated *KL* and *COMT* gene variants were not associated with differences in cognitive performance, even after interaction with A $\beta$  burden and *APOE*  $\varepsilon$ 4 carriage. Whilst the preceding studies did not present significant associations across all genes investigated, it is still plausible that individual genes may have subtle effects on cognitive decline in the preclinical stages of AD that are more apparent when studied in combination with other genes.

The ensuing study presented in this chapter hypothesized that cognitive genes could have an additive influence that is obviously not observed when investigating variants independently. To address this hypothesis, the study aimed to address the third aim of the thesis, being to *investigate whether there is a synergistic effect of genes previously associated with cognition, and further what the best combination of these genes would be.* 

To achieve this aim it was proposed to combine genes associated with cognitive performance using a method that would be simple to use and replicate, allowing for ease of use clinically. In addition to the genes investigated in Chapter 3, the Val66Met variant in *BDNF*, previously studied in the AIBL cohort [1, 2], and a variant within the "CUB and Sushi Multiple Domains 1" (*CSMD1*) gene were included. *CSMD1* is involved in complement regulation and variants within it have been associated with cognitive performance in healthy individuals [3, 4]. A decision tree approach was undertaken to derive groups based on rates of cognitive decline, specifically decline in a composite measure of verbal episodic memory in cognitively normal individuals with high levels of brain A $\beta$ .

# **Prologue References:**

- 1. Lim, Y.Y., et al., *BDNF Val66Met, Abeta amyloid, and cognitive decline in preclinical Alzheimer's disease.* Neurobiol Aging, 2013. **34**(11): p. 2457-64.
- 2. Lim, Y.Y., et al., *APOE and BDNF polymorphisms moderate amyloid beta-related cognitive decline in preclinical Alzheimer's disease*. Mol Psychiatry, 2015. **20**(11): p. 1322-8.
- Kraus, D.M., et al., CSMD1 is a novel multiple domain complement-regulatory protein highly expressed in the central nervous system and epithelial tissues. J Immunol, 2006. 176(7): p. 4419-30.
- 4. Athanasiu, L., et al., *A genetic association study of CSMD1 and CSMD2 with cognitive function*. Brain Behav Immun, 2017. **61**: p. 209-216.

# 4.2 Cognitive Gene Risk Profile for the Prediction of Cognitive

# **Decline in Presymptomatic Alzheimer's Disease**

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### 4.2.1 Abstract

INTRODUCTION: In cognitively normal (CN) older adults, high levels of Aβ-amyloid are associated with significant decline in cognition, especially episodic memory. Several genes have previously been associated with cognition, including *APOE*, *KIBRA*, *KLOTHO*, *BDNF*, *COMT*, *SPON1* and *CSMD1*. While some of this variation has been attributed to some of these genes individually, the combined effects of these genes on rates of cognitive decline, particularly in preclinical Alzheimer's Disease remain largely unknown.

METHODS: To elucidate if risk alleles within these genes can be suitably combined to predict cognitive decline 127 CN older adults with elevated PET-ascertained A $\beta$ -amyloid were included in a decision tree analysis to define a "cognitive gene risk profile" for decline in a verbal episodic memory composite.

RESULTS: The episodic memory-derived cognitive gene risk profile defined four groups: *APOE*  $\varepsilon$ 4+ Risk,  $\varepsilon$ 4+ Resilient,  $\varepsilon$ 4- Risk,  $\varepsilon$ 4- Resilient, with the  $\varepsilon$ 4+ Risk group declining significantly faster than all other groups ( $\varepsilon$ 4+ Resilient, p=0.0008;  $\varepsilon$ 4- Risk, p=0.025;  $\varepsilon$ 4- Resilient, p=0.0006). The  $\varepsilon$ 4+ Risk group also declined significantly faster than all other groups on Global, Clinical Progression and Pre-Alzheimer's cognitive composites. DISCUSSION: The defined cognitive gene risk profile has potential utility in participant selection/stratification for preclinical AD trials that incorporate A $\beta$ -amyloid and where decline in cognition is essential to determine therapeutic effectiveness.

**KEY WORDS:** genetic risk profiles, cognitive decline, Alzheimer's disease, episodic memory, Aβ-amyloid

**NON-STANDARD ABBREVIATIONS:** AIBL, Australian Imaging Biomarkers and Lifestyle study of Ageing; CN, Cognitive Normal; Cog-GRP, Cognitive Gene Risk Profile; PACC, Pre-Alzheimer's Cognitive Composite

### 4.2.2 Introduction

Evidence from prospective longitudinal cohort studies suggests that the pathological changes in Alzheimer's Disease (AD) commence decades before the onset of clinical symptomology [1]. Further, it has been established that higher levels of A $\beta$ -amyloid (A $\beta$ ) in cognitively normal (CN) older adults is associated with accelerated decline in cognition [2]. As such, cerebrospinal fluid (CSF) and imaging biomarkers of A $\beta$  are used to define the preclinical stage of AD [3, 4]. However, at the preclinical stage of AD there is considerable interpersonal variability in the rate of cognitive decline, suggesting that while A $\beta$  is a necessary condition for AD, other factors influence the relationship between this biomarker and clinical disease progression. Cognition has been shown to be both highly heritable and highly polygenic [5] and allelic variation in several genes associated with cognition has been shown to explain some variation in cognitive function in older adults and in A $\beta$  related cognitive decline in early AD [6-8]. Thus suggesting that genetics could help inform and predict rates of cognitive decline, and identify groups of CN older adults that are at a higher risk of a more rapid decline in cognition.

There have been several individual genes associated with cognitive performance and decline. The major genetic risk factor for AD, the  $\epsilon$ 4 allele of apolipoprotein E (*APOE*) [9], has been consistently associated with accelerated rates of episodic memory decline and hippocampal atrophy (reviewed in [10]). The non-synonymous rs6265 (Val66Met) single nucleotide polymorphism (SNP) in the brain derived neurotropic factor (*BDNF*), has been linked with altered rates of decline in several cognitive domains, and hippocampal atrophy [7, 8]. A further non-synonymous SNP that regulates dopamine availability in the central nervous system, rs4680 (Val158Met) within Catechol-O-methyltransferase (*COMT*), has also been associated with cognitive performance [11]. The Klotho gene (*KL*), initially discovered in transgenic mice

with a phenotype resembling human aging [12], has a functional variant, *KL*-VS that has been associated with life expectancy [13], global cognition [14], processing speed [14], and brain volume [15].

A further gene, *KIBRA*, that encodes the KIdney and BRAin expressed protein has recently been shown to be involved in the mediation of tau-induced memory loss and synaptic plasticity [16]. Allelic variation in the *KIBRA* gene, specifically a substitution of C for T in the 9<sup>th</sup> intron (rs17070145), has been reported to be associated with memory performance [17], hippocampal atrophy [18] and measurable differences in brain activation [17]. We have described recently how this gene contributes to moderating A $\beta$  driven cognitive decline [19]. Additionally, several SNPs in the *CSMD1* (CUB and Sushi Multiple Domains 1) gene, involved in the regulation of complement and inflammation [20], have been associated with episodic memory and general cognition in a cognitively normal sample [21]. Finally, multiple SNPs within the Spondin 1 (*SPON1*) gene, involved in the processing of amyloid precursor protein (APP) [22], have been associated with disease severity [23] and rates of cognitive decline [24], though only in AD individuals.

Several studies have investigated the extent to which combinations of genes can influence cognitive decline and clinical progression in AD [25-28]. However, most of these studies focused on genes shown previously to be associated with risk for AD, with gene weighting based on AD risk [25, 26]. Thus these polygenic approaches may have resulted in exclusion of genes associated with cognitive performance, or if included, their influence diluted due to a disease risk based weighting [26]. Further, few studies have taken brain A $\beta$  burden into consideration and investigated combining genes associated with cognitive performance in preclinical AD [8, 29].

This study hypothesised that combining genes shown to be associated with cognition would explain variance in  $A\beta$  related cognitive decline in preclinical AD. This study aimed to combine these genes into a straightforward profile able to discriminate individuals based on cognition, and particularly episodic memory, which is one of the earliest cognitive domains to decline [30]. The profile was created in CN older adults, signified at risk of cognitive decline based on brain imaging biomarkers, enrolled in the Australian Imaging, Biomarkers and Lifestyle (AIBL) Study. Extensive 18-monthly assessment, including cognitive and neuroimaging, within the AIBL Study allows for the longitudinal evaluation of this profile. Such a genetic profile could be easily implemented for the identification of individuals with accelerated rates of cognitive decline, which could have utilisation for clinical trial design, leading to more efficient clinical trials and secondary prevention studies.

### 4.2.3 Materials and Methods

# 4.2.3.1 Study Participants

One hundred and thirty-three CN biomarker positive (based on brain imaging) older adults enrolled in the AIBL Study, a prospective longitudinal study of ageing, were included in this study. The study design, enrolment process, neuropsychological assessments, and diagnostic criteria of the AIBL Study have been previously described [31]. Approval of the AIBL Study has been granted by each of the ethics committees of each of the member institutions: Austin Health, St Vincent's Health, Hollywood Private Hospital, and Edith Cowan University, and all volunteers gave informed written consent. Assessments occurred every 18 months, with cognitive, neuroimaging and laboratory assessment achieved within 3 months of each other.

#### 4.2.3.2 Cognitive Measures

Burnham *et al.* previously calculated cognitive composite scores using the AIBL neuropsychological test battery and the Clinical Dementia Rating (CDR) scale [32]. These composite scores were used in this study to assess cognitive performance. The AIBL neuropsychological test battery consists of Mini-Mental State Examination (MMSE), Clock Drawing Test, California Verbal Learning Test-Second edition (CVLT-II), Logical Memory I and II (LMI; LMII; Story A only), D-KEFS verbal fluency, a 30-item version of the Boston Naming Test (BNT), Wechsler Test of Adult Reading (WTAR), Digit Span and Digit Symbol-Coding subtests of the Wechsler Adult Intelligence Scale-Third edition (WAIS-III), the Stroop task (Victoria version), and the Rey Complex Figure Test (RCFT) [31]. Briefly, a verbal episodic memory composite (CDR sum of boxes (CDR<sub>SB</sub>), LMII, CVLT-II recognition false positives ( $CVLT_{FP}$ ) and long delay free recall ( $CVLT_{LDFR}$ )) was used as the primary cognitive measure for defining groups with different rates of decline. Groups defined by decline in episodic memory were also assessed against a global cognition composite (CDR<sub>SB</sub>, MMSE,

LMII, CVLT<sub>FP</sub> and Clock), and a composite measure of clinical progression (CDR<sub>SB</sub>, MMSE) [32]. In addition, the Pre-Alzheimer's cognitive composite (PACC) previously calculated by Donohue *et al.* was also investigated [33]. In the calculation of the statistically driven composites there were corrections for age, sex, years of education, premorbid IQ (WTAR-estimated WAIS-III Full Scale Intelligence Quotient (FSIQ)) and depressive symptoms (Geriatric Depression Scale (GDS)) [34]. Five cognitive assessment time points were used: baseline, 18, 36, 54 and 72 months.

# 4.2.3.3 Brain Imaging

Aβ imaging with positron emission tomography (PET) using <sup>11</sup>C-Pittsburgh Compound B (PiB), <sup>18</sup>F-florbetapir or <sup>18</sup>F-flutemetamol was performed on the 133 cognitively normal adults included in this study as previously described [35-37]. The same region of interest template was used to determine PET standardized uptake value (SUV) ratio (SUVR) data for all tracers [38]. Briefly, SUVs were summed and scaled based on tracers PiB, florbetapir, and flutemetamol, to the cerebellar cortex, whole cerebellum or pons, respectively, to yield the target-region to reference-region SUVR. This study classified participants as high (Aβ<sup>high</sup>) Aβ burden, based on a tracer-specific SUVR threshold; ≥1.5, ≥1.10 and ≥0.62 for PiB, florbetapir and flutemetamol, respectively, as previously described [39]. For cross-sectional comparison of Aβ burden with multiple tracers a linear regression transformation was applied to <sup>18</sup>Flabbelled tracers to generate PiB-like SUVR units termed the "Before the Centiloid Kernel Transformation" (BeCKeT) scale [40].

# 4.2.3.4 Genotyping

We have previously described methods of DNA extraction and SNP genotyping [41]. Briefly, manufacturer's instructions were followed to extract DNA from 5mL of whole blood using QIAamp DNA Blood Maxi Kits (Qiagen, Hilden, Germany). TaqMan® genotyping assays were used to determine *APOE* (rs7412, assay ID: C\_\_\_904973\_10; rs429358, assay ID: C\_\_\_3084793\_20), *BDNF* (rs6265, assay ID: C\_\_11592758\_10), *KIBRA* (rs17070145, assay ID: C\_\_33286269\_10), *COMT* (rs4680, assay ID: C\_\_25746809\_50), *KL* (*KL*-VS; rs9536314, assay ID: C\_\_\_2983037\_20; rs9527025, assay ID: C\_\_2983036\_20), *SPON1* (rs11023139, assay ID: C\_\_\_55174\_30), and *CSMD1* (rs2740931, custom designed assay) genotypes (Life Technologies, Carlsbad, CA). TaqMan® genotyping assays were performed on a QuantStudio 12K Flex<sup>TM</sup> Real-Time-PCR systems (Applied Biosystems, Foster City, CA) using the TaqMan® GTXpress<sup>TM</sup> Master Mix (Life Technologies) as per manufacturer instructions. *APOE* carrier status is defined by the presence (1 or 2 copies) or absence (0 copies) of the *APOE* ɛ4 allele, henceforth referred to as *APOE* ɛ4+ve or *APOE* ɛ4-ve, respectively. Further *KL*-VS homozygotes (n=6) were excluded from all analyses resulting in the inclusion of 127 CN adults.

#### 4.2.3.5 Statistical Analysis

All statistical analyses were performed using Rstudio (Rstudio Team 2015) Version 0.98.1103 for Macintosh [42]. Baseline demographic data analyses, using the generic functions of the R "base" package, provided means, standard deviations, and percentages across the cognitively normal sample. The first stage of analysis was the definition of the individual slopes for verbal episodic memory decline in the  $A\beta^{high}$  sample (n=127), which would then be included in the subsequent decision tree analysis. These individual slopes were created using the "nlme" package in R using random intercepts linear mixed-effects (LME) models, which model fixed and random effects, and deal with missing data robustly [43]. In this analysis, a verbal episodic memory  $\times$  time interaction was modelled to generate per person  $\beta$  values (slopes). As the calculation of the verbal episodic memory composite is controlled for age, sex, years of education, premorbid IQ and depressive symptoms, no further covariates were included in the LME models. The second stage of analysis utilised these slopes (dependent variables) in combination with the seven genes of interest (APOE, BDNF, KIBRA, KL, COMT, SPON1 and CSMDI; independent variables), in a decision tree model using the "rpart" package in R, to define the "Cognitive Gene Risk Profile" (Cog-GRP) groups to be used in subsequent analyses. The final stage of analysis was to assess the performance of the defined Cog-GRP groups. To achieve this differences in rates of cognitive change between these groups were assessed using random intercepts LME models, using the "nlme" package in R. Specifically, a Cog-GRP group × Time interaction was modelled across the entire sample, with the cognitive composites as the dependent variables. With the exception of analyses for the AIBL-PACC, which covaried for age, no additional covariates were included due to their inclusion in the generation of the cognitive composites. All LME models were presented graphically with time point dependent standard error. Effect sizes were calculated based on cognitive performance at the sixth year of follow-up using the "effsize" package in R.

### 4.2.4 Results

# 4.2.4.1 $A\beta^{high}$ cognitively normal adults baseline demographics, genotype frequencies and cognitive slopes

Table 4.2.1 shows the demographics, genotype frequencies and cognitive slopes of the 127  $A\beta^{high}$  CN older adults included in the study. The statistically driven global composite (-0.0901 SD/year), clinical progression (-0.0484 SD/year), and verbal episodic (-0.0774 SD/year) composites all presented with a negative rate of change when investigating  $A\beta^{high}$  CN older adults.

# 4.2.4.2 Defining the Cognitive Gene Risk Profile (Cog-GRP) and group stratification

The "Rpart" package in R was used to calculate the decision tree that defined the *Cog-GRP*. The decision tree was constructed using 7 gene variants (*APOE*  $\varepsilon$ 4+/ $\varepsilon$ 4-, *BDNF* Met+/Val/Val, *KIBRA* T-/T+, *COMT* Val+/Met/Met, *KLOTHO* VS-/VS+, *SPONI* A-/A+, and *CSMD1* G-/G+) against a composite score of verbal episodic memory in an A $\beta^{high}$  sample. The analysis resulted in the selection of six of the seven genes (*COMT* falling out of the analysis, Figure 4.2.1a), which were used to classify the participants into 8 groups. Due to small sample sizes in the resultant 8 groups, groups were collapsed at the end of the respective  $\varepsilon$ 4+ and  $\varepsilon$ 4- branches. This was based on their same directions, and similar rates, of change (SD/year) in verbal episodic memory over the assessed 6-years. The resulting 4 groups were then classified as "at risk" or "resilient" based on carriage of the  $\varepsilon$ 4 allele and differences in decline on the verbal episodic composite for these collapsed groups were reconfirmed (Figure 4.2.1b). The  $\varepsilon$ 4+ Risk (-0.1891 SD/year) group had a significantly faster rate of decline than  $\varepsilon$ 4+ Resilient (0.0014 SD/year; p=0.0008; At 6<sup>th</sup> year: Cohen's *d*=1.14, 95% CI 0.67-1.59) and  $\varepsilon$ 4- Resilient (0.0097 SD/year; p=0.0006; At 6<sup>th</sup> year: Cohen's *d*=2.37, 95% CI 1.63-3.04) groups and reached

clinically significant thresholds of cognitive impairment (performance at 1.5 standard deviations below controls, dashed line Figure 4.2.1b) after 5 years. For comparison purposes, Figure 4.2.1b, shows that this threshold is crossed at approximately 9.6 years when only carriage of the *APOE*  $\varepsilon$ 4 allele is considered, which had a decline of -0.110 SD/year.

		Aβ <sup>high</sup> CN older adults
Age (years)		73.17 (6.50)
Female (%)		66 (51.97)
Years of Education (%)	0-8	11 (8.66)
	9-12	51 (40.16)
	13-15	31 (24.41)
	15+	34 (26.77)
Premorbid IQ (FSIQ)		108.54 (6.87)
Depressive Symptoms (GDS)		1 (1.26)
<i>APOE</i> (% ε4+ve)		66 (51.97)
BDNF (% Met+ve)		43 (33.86)
KIBRA (% T-ve)		62 (48.82)
<i>KL</i> (%VS-ve)		99 (77.95)
COMT (% Val+ve)		103 (81.10)
<i>CSMD1</i> (% G–ve)		42 (33.07)
SPON1 (% A-ve)		112 (88.19)
Global Cognition*		-0.0901
Clinical Progression*		-0.0484
Verbal Episodic Memory*		-0.0774
AIBL-PACC†		-0.1144

# Table 4.2.1 Baseline demographic information

Baseline demographic, genotype frequencies and cognitive composites slopes for cognitively normal (CN) older adults with high A $\beta$ -amyloid (A $\beta^{high}$ ) in the AIBL Study (n=127). \*Cognitive composites presented in SD/year, and the †Pre-Alzheimer's Cognitive Composite (AIBL-PACC) presented in 4×SD/year. AIBL-PACC controlled for age. *KL-VS* homozygotes excluded (n=6). GDS, Geriatric Depression Scale; FSIQ, WTAR-estimated FSIQ.



# Figure 4.2.1 Cognitive Gene Risk Profile (Cog-GRP)

(a) Derivation of the cognitive gene risk profile (*Cog-GRP*) within cognitively normal adults high Aβ-amyloid (A $\beta^{high}$ ; n=127) using decision tree analysis. Defining four groups  $\varepsilon$ 4+ Risk (red, n=40),  $\varepsilon$ 4+ Resilient (orange, n=26),  $\varepsilon$ 4– Risk (blue, n=43),  $\varepsilon$ 4– Resilient (green, n=18). Values represent SD/year change in the verbal episodic memory composite. All analyses were corrected for age, sex, years of education, premorbid IQ and depressive symptoms. (b) Confirmation of performance of collapsed groups identified by the *Cog-GRP* in A $\beta^{high}$ cognitively normal adults (n=127) and comparative performance of *APOE* when considered independently.  $\varepsilon$ 4– Resilient group (green line),  $\varepsilon$ 4– Risk group (blue line),  $\varepsilon$ 4+ Resilient group (orange line),  $\varepsilon$ 4+ Risk group (red line), *APOE*  $\varepsilon$ 4-ve (grey dotted line), *APOE*  $\varepsilon$ 4+ve (black dotted line). Broken black line represents 1.5 SD of decline. Error bars represent time dependent standard error, \*p<0.05 when comparing to the  $\varepsilon$ 4+ Risk group

#### 4.2.4.3 Performance of Cog-GRP groups on cognition in cognitively normal adults

When investigating the association between *Cog-GRP* groups and cognition in  $A\beta^{high}$  CN older adults, no significant differences were observed at baseline in any composite measures. The ε4+ Risk group showed a significantly greater rate of decline on the global composite (p=0.00009), clinical progression composite (p=0.0003) and AIBL-PACC (p=0.0022), over six years when compared to  $\varepsilon$ 4- Resilient group (Table 4.2.2, Figure 4.2.2). At the 6<sup>th</sup> year of follow-up there was a large effect observed for the global (Cohen's d=2.57, 95% CI 1.82-3.26) and clinical progression composites (Cohen's d=1.87, 95% CI 1.20-2.50) and AIBL-PACC (Cohen's d=2.32, 95% CI 1.60-2.99). Further, relative to the  $\varepsilon$ 4- Risk and  $\varepsilon$ 4+ Resilient groups, the  $\varepsilon$ 4+ Risk group also showed a significantly greater rate of decline on the global composite (p=0.020, p=0.001), and clinical progression (p=0.023, p=0.015) over six years (Table 4.2.2, Figure 4.2.2). Large effects were again observed at the 6<sup>th</sup> year of follow-up for both the global (e4- Risk: Cohen's d=1.18, 95% CI 0.70-1.63; e4+ Resilient: Cohen's d=1.52, 95% CI 0.94-2.06) and clinical progression composites ( $\varepsilon$ 4- Risk: Cohen's d=0.95, 95% CI 0.48-1.39;  $\varepsilon$ 4+ Resilient: Cohen's d=1.27, 95% CI 0.72-1.79). The  $\varepsilon$ 4+ Risk group also declined significantly faster on the AIBL-PACC (p=0.040) when compared to the ɛ4– Risk group (at 6<sup>th</sup> year, Cohen's d=1.18,95% CI 0.70-1.63), though only a trend toward significance, (p=0.073), when comparing to the  $\varepsilon$ 4+ Resilient group. Across all cognitive composites there was no significant difference between groups in terms of baseline cognitive performance, with the exception of the extremes of ɛ4+ Risk compared to ɛ4- Resilient groups in the Global cognitive composite (Table 4.2.2). Finally, mean A<sup>β</sup> burden was observed to be significantly different between Cog-GRP groups (ε4+ Risk, 2.02±0.35; ε4+ Resilient, 1.85±0.21; ε4- Risk, 1.82±0.24; ε4- Resilient 1.97 $\pm$ 0.46; F= 3.41, p=0.020); though this was driven only by a difference between  $\varepsilon$ 4+ Risk and ɛ4- Risk groups (Post-hoc Bonferroni, p=0.026). Finally, when analyses were repeated in 397 A $\beta^{low}$  CN older adults from the AIBL study, to determine whether the defined Cog-GRP

had utility in defining cognitive decline in in biomarker negative CN older adults, no significant differences at baseline or between slopes in any composite measures were observed (see Section 4.2.8 Supplementary Data, for full sample demographics and analysis outcome measures in  $A\beta^{low}$  CN older adults; Tables S4.2.3 and S4.2.4).

	ε4+ Risk group		ε4+ Resilient group		ε4– Risk group		ε4– Resilient group	
	α	β	α	β	α	β	α	β
Global	-0.578	-0.218	-0.385	-0.019*	-0.284	-0.089*	0.291*	0.030*
<b>Clinical Progression</b>	-0.087	-0.100	0.048	-0.032*	-0.029	-0.042*	0.079	0.006*
AIBL-PACC	7.168	-0.334	8.349	-0.072	7.982	-0.066*	8.603	0.126*

Table 4.2.2 Group intercepts and slopes for cognitive composites in  $A\beta^{high}$  CN older adults Group intercepts ( $\alpha$ ; as SD) and slopes ( $\beta$ ; as SD/year) for cognitive composites (presented in SD/year) and AIBL-PACC (presented in SD×4/year) in imaged cognitively normal (CN) older adults with high A $\beta$ -amyloid ( $A\beta^{high}$ ; n=127). \*p<0.05 when comparing to the  $\epsilon$ 4+ at Risk group.



# Figure 4.2.2 Cognitive rates of change in $A\beta^{high}$ CN older adults

Cognitive rates of change are presented for a (a) global composite, (b) clinical progression composite, and (c) Pre-Alzheimer's Cognitive Composite (AIBL-PACC) in cognitively normal (CN) older adults with high A $\beta$ -amyloid (A $\beta^{high}$ ; n=127).  $\epsilon$ 4– Resilient group (green line),  $\epsilon$ 4– Risk group (blue line),  $\epsilon$ 4+ Resilient group (orange line),  $\epsilon$ 4+ Risk group (red). AIBL-PACC controlled for age. Error bars represent time dependent standard error, \*p<0.05 when comparing to the  $\epsilon$ 4+ Risk group.

# 4.2.5 Discussion

Results from this study support the hypothesis that combining genes previously associated with cognitive performance allows for the identification of groups of individuals with accelerated rates of cognitive decline. In CN older adults with high AB burden at baseline a decision tree was created driven by decline in a composite score of verbal episodic memory to define a Cog-GRP. This profile combined the effects of APOE, BDNF, KIBRA, KLOTHO, SPON1 and CSMD1. COMT dropped out of the model due to lack of influence in discriminating the cognitive change within the sample. There is no association between *COMT* individually and rates of cognitive decline in this population (data not shown), so it is unsurprising that it has not contributed to the genetic risk profile created. Due to the lack of significant difference in decline and the small sample size between the groups at either extreme of the *Cog-GRP* these groups were collapsed to 4 overall classifications. These classifications were described based on carriage of the APOE ɛ4 allele; ɛ4+ Risk, ɛ4+ Resilient, ɛ4- Risk and ɛ4- Resilient. The  $\varepsilon 4$ + Risk group showed significantly faster decline in the global composite and the composite of clinical progression when compared to all other groups. Further, the ɛ4+ Risk group showed significantly faster decline in the AIBL-PACC when compared to the  $\varepsilon$ 4- Risk and  $\varepsilon$ 4- Resilient groups, however the difference between the  $\varepsilon$ 4+ Resilient group only trended towards significance. Finally, we report that the defined Cog-GRP has no utility in defining cognitive decline in  $A\beta^{low}$  CN older adults.

In  $A\beta^{high}$  CN older adults, where the *Cog-GRP* was able to differentiate rates of cognitive decline, no significant differences in baseline cognition was apparent, whilst  $A\beta$  burden was only observed to be different between the two "at risk" groups. Suggesting that the observed cognitive outcomes were more dependent upon the *Cog*-

*GRP* than driven by either of these factors. However, the lack of observable impact of the *Cog-GRP* in in  $A\beta^{low}$  CN older adults suggests that above threshold levels of brain A $\beta$  burden is required for observable cognitive decline. This has been reported previously, where it is suggested that the absence of above threshold level of A $\beta$  burden, even in the presence of neurodegeneration, does not confer an increased risk for cognitive decline [44]. Taken together these observations have potential implication for the design of clinical trials. Specifically, trials would likely benefit from the inclusion of both a measure of A $\beta$  burden and risk stratification through, for example, a genetic risk profile as presented in this study.

We have previously reported the ability of genetic factors to discriminate individuals with accelerated rates of cognitive decline in the AIBL Study, above and beyond the effects of *APOE* alone [8]. In the present study, the defined *Cog-GRP* polygenic approach, showed that the  $\epsilon$ 4+ Risk group would reach clinically significant thresholds of cognitive impairment in episodic memory (performance at 1.5 standard deviations below controls, illustrated by the dashed line in Figure 4.2.1b) after 5 years, compared to approximately 14 years in the  $\epsilon$ 4- Risk group and indeterminate years for the remaining groups (due to positive slopes), including the  $\epsilon$ 4+ Resilient group. A slightly shorter period of approximately 4 years was estimated for a clinically significant decline in the global composite. The period of time to cross threshold in the  $\epsilon$ 4+ Risk group is almost twice as fast as that when considering *APOE* alone ( $\epsilon$ 4+ Risk, 5 years;  $\epsilon$ 4+ only, 9.6 years) whilst the  $\epsilon$ 4+ Resilient group shows no decline, suggesting that the additional genes affected trajectories of cognitive decline above and beyond the effects of *APOE* alone and are able to clearly define  $\epsilon$ 4 who decline or remain stable . This is further emphasised by the negligible differences in cognitive performance at

baseline in these groups. Finally, the defined  $\epsilon$ 4+ Risk group captures twice the number of individuals (n=40) as *APOE* and *BDNF* alone (n=20). Taken together, these finding suggest that broadening the scope in terms of genetic variants may provide more clinical utility for implementation in clinical trials where cognitive decline is a primary endpoint.

APOE, BDNF, KIBRA, KL, SPON1 and CSMD1 have all previously been associated with both cross-sectional and longitudinal measures of cognitive change [7, 14, 17, 21], whereby the independent influences of these genes have been investigated thoroughly. In contrast, there is minimal research focused on the combined effects of these genes, with the research that has been conducted focusing on combination effects with either APOE or BDNF [8, 29]. In AD, polygenic investigations of disease progression and cognition have focused broadly on those genes identified in case-control GWAS studies [26-28]. While there have been a number of genetic risk scores that have been developed and are associated with longitudinal and cross-sectional cognition, these scores have had limited validation in at-risk preclinical AD cohorts. The decision tree derived Cog-GRP reported in this study is novel in its use of cognitively associated genes to predict decline in preclinical disease. Whilst the significant differences of large effect observed at the 6<sup>th</sup> year of follow-up, which would likely be considered to be clinically meaningful [45] (Cohen's d > 1.0 across all cognitive composites), suggests the strong potential for translation into clinical practice. Decision trees have been widely investigated in neurodegenerative disease research, typically for diagnosis of disease or disease stage. Investigators have used a range of approaches to achieve this: neuropsychologically-framed interview questions to discriminate dementia, MCI and controls [46], or between neurodegenerative disorders [46, 47], gene expression to

diagnose AD [48], demographic variables to determine cognitive and functional change [49], fMRI, behavioural and demographic information for diagnosis of AD [50], MMSE, neurofibrillary tangles and gene expression to classify disease stages [51]. Similarly, the decision tree reported in this study was developed for possible clinical use making the method's ease of utility favourable.

The combination of genes associated with cognitive decline is a strength of this study as it allows for the discrimination of rates of cognitive decline at preclinical disease stages, however, the authors do acknowledge the limitations in the study. The decision tree created within this study was statistically driven based on an episodic memory composite derived from specific cognitive assessments, and the use of different neuropsychological tests to create these composite scores could result in the creation of a different genetic risk profile. Secondly, participants in the AIBL Study are not randomly selected: they volunteer for involvement, likely accounting for a typically slightly higher than average level of performance in cognitive assessments, which may not represent the general population and might complicate replication in other cohorts. In addition to these limitations, the small sample sizes of the groups after discrimination by the *Cog-GRP* created could influence the results, and it will be important to replicate these findings in other cohorts that are conducive to cross validation of comparable cognitive endpoints.

Overall, this study reports a genetic risk profile, derived from *a priori* gene candidates previously associated with cognitive performance, that can partition CN older adults into groups that differ significantly in rates of cognitive decline. With the later disease stage intervention strategy of previous AD clinical trials generally considered to have contributed to their lack of success and decline on clinical endpoints (cognitive tests) still essential to assess efficacy, there is a now a focus on preclinical AD trials and the appropriate means to select participants. The ease of clinical utility of the presented *Cog-GRP* would not only readily allow its employment in clinical trial design for group stratification but also for use in the retrospective analysis of prior clinical trial data. Furthermore, *Cog-GRP* also supports the investigation of additional genes beyond those associated with AD risk in GWAS, for defining polygenic risk scores for cognitive decline in presymptomatic biomarker positive individuals.

# 4.2.6 Acknowledgements

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# **4.2.7 Competing Financial Interests**

CLM is an advisor to Prana Biotechnology Ltd and a consultant to Eli Lilly. PM is a full-time employee of Cogstate Ltd. DA has served on scientific advisory boards for Novartis, Eli Lilly, Janssen, and Pfizer Inc. RNM is a consultant to Alzhyme. SML has previously been a paid consultant to Alzhyme. CCR has served on scientific advisory boards for Bayer Pharma, Elan Corporation, GE Healthcare and AstraZeneca; has received speaker honoraria from Bayer Pharma and GE Healthcare; and has received research support from Bayer Pharma, GE Healthcare, Piramal Lifesciences and Avid Radiopharmaceuticals. VLV served as a consultant for Bayer Pharma; and received research support from a NEDO grant from Japan. All other authors have nothing to disclose.

# 4.2.8 Supplementary Data

# 4.2.8.1 Supplementary Materials and Methods

#### 4.2.8.1.1 Study Participants

In addition to the 133 biomarker positive individuals, 406 CN biomarker negative older adults enrolled in the AIBL Study, a prospective longitudinal study of ageing, were investigated. Participants were classified as low ( $A\beta^{low}$ ; n=406) A $\beta$  burden, based on a tracer-specific SUVR threshold; <1.5, <1.10 and <0.62 for PiB, florbetapir and flutemetamol, respectively, as previously described [39]. Nine individuals were excluded from further analysis based on *KL*-VS homozygosity resulting in the inclusion 397  $A\beta^{low}$  CN adults.

# 4.2.8.1.1 Statistical Analysis

All statistical analyses were performed using Rstudio (Rstudio Team 2015) Version 0.98.1103 for Macintosh [42]. Baseline demographic data analyses provided means, standard deviations, and percentages across the cognitively normal sample. Differences in rates of cognitive change between the groups defined by the *Cog-GRP* were assessed in  $A\beta^{low}$  group using random intercepts linear mixed-effects (LME) models and were performed using the "nlme" package in R. A *Cog-GRP* group × Time interaction was modelled across the entire sample, with the cognitive composites as the dependent variables. All analyses for the AIBL-PACC co-varied for age.

# 4.2.8.2 Supplementary Results

# 4.2.8.2.1 $A\beta^{low}$ cognitively normal adults baseline demographics, genotype frequencies and cognitive slopes

Table S4.2.3 shows the demographics, genotype frequencies and cognitive slopes of the 406 A $\beta^{low}$  CN older adults included in the study. All composites presented with positive rates of change in A $\beta^{low}$  CN adults, likely reflecting a practice effect in these participants.

# 4.2.8.2.2 Performance of Cog-GRP groups on cognition in cognitively normal $A\beta$ low adults

No significant differences at baseline or between slopes in any composite measures were observed when investigating cognitively normal  $A\beta^{low}$  older adults (Table S4.2.4).

		Aβ <sup>low</sup> CN older adults
Age (years)		69.81 (6.20)
Female (%)		224 (56.42)
Years of Education (%)	30 (7.56)	11 (8.66)
	150 (37.78)	51 (40.16)
	72 (18.14)	31 (24.41)
	145 (36.52)	34 (26.77)
Premorbid IQ (FSIQ)		107.80 (7.09)
Depressive Symptoms (GDS)		1 (1.21)
<i>APOE</i> (% ε4+ve)		79 (19.90)
BDNF (% Met+ve)		144 (36.27)
KIBRA (% T-ve)		172 (43.32)
<i>KL</i> (%VS–ve)		293 (73.80)
COMT (% Val+ve)		302 (76.07)
<i>CSMD1</i> (% G–ve)		115 (28.97)
SPONI (% A-ve)		358 (90.18)
Global Cognition*		0.0218
Clinical Progression*		0.0010
Verbal Episodic Memory*		0.0248
AIBL-PACC†		0.0213

*Table S4.2.3 Baseline demographic information for*  $A\beta^{low}$  *CN older adults* Baseline demographic, genotype frequencies and cognitive composites slopes for cognitively normal (CN) older adults with low Aβ-amyloid (A $\beta^{low}$ ) in the AIBL Study (n=397). \*Cognitive composites presented in SD/year, and the †Pre-Alzheimer's Cognitive Composite (AIBL-PACC) presented in 4×SD/year. AIBL-PACC controlled for age. *KL-VS* homozygotes excluded (n=9). GDS, Geriatric Depression Scale; FSIQ, WTAR-estimated FSIQ.

	ε4+ Risk group	ε4+ Resilient group	ε4– Risk group	ε4– Resilient group
	β	β	β	β
Global	0.014	0.033	0.015	0.034
<b>Clinical Progression</b>	0.004	0.012	0.003	-0.008
Verbal Episodic Memory	0.026	0.028	0.017	0.038
AIBL-PACC	0.028	0.083	0.009	0.020

*Table S4.2.4 Mean slopes for cognitive composites in A\beta^{low} CN older adults* Mean slopes for cognitive composites (presented in SD/year) and AIBL-PACC (presented in SD×4/year) in imaged cognitively normal (CN) older adults with low A $\beta$  (A $\beta^{low}$  n=397). \*p<0.05 when comparing to the  $\epsilon$ 4+ at Risk group

# 4.2.9 References

- Villemagne, V.L., et al., *Amyloid beta deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study.* Lancet Neurol, 2013. **12**(4): p. 357-67.
- 2. Villemagne, V.L., et al., *Longitudinal assessment of Abeta and cognition in aging and Alzheimer disease*. Ann Neurol, 2011. **69**(1): p. 181-92.
- 3. Habib, M., et al., *Functional neuroimaging findings in healthy middle-aged adults at risk of Alzheimer's disease*. Ageing Res Rev, 2017. **36**: p. 88-104.
- Kulic, L. and P.G. Unschuld, *Recent advances in cerebrospinal fluid* biomarkers for the detection of preclinical Alzheimer's disease. Curr Opin Neurol, 2016. 29(6): p. 749-755.
- 5. Kirkpatrick, R.M., et al., *Results of a "GWAS plus:" general cognitive ability is substantially heritable and massively polygenic.* PLoS One, 2014. **9**(11): p. e112390.
- Lim, Y.Y., et al., *Abeta-related memory decline in APOE epsilon4 noncarriers: Implications for Alzheimer disease*. Neurology, 2016. 86(17): p. 1635-42.
- Lim, Y.Y., et al., *BDNF Val66Met, Abeta amyloid, and cognitive decline in preclinical Alzheimer's disease*. Neurobiol Aging, 2013. 34(11): p. 2457-64.
- Lim, Y.Y., et al., APOE and BDNF polymorphisms moderate amyloid betarelated cognitive decline in preclinical Alzheimer's disease. Mol Psychiatry, 2015. 20(11): p. 1322-8.

- 9. Corder, E.H., et al., *Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families.* Science, 1993. **261**(5123): p. 921-3.
- Liu, Y., et al., APOE genotype and neuroimaging markers of Alzheimer's disease: systematic review and meta-analysis. J Neurol Neurosurg Psychiatry, 2015. 86(2): p. 127-34.
- Sheldrick, A.J., et al., *Effect of COMT val158met genotype on cognition and personality*. Eur Psychiatry, 2008. 23(6): p. 385-9.
- 12. Kuro-o, M., et al., *Mutation of the mouse klotho gene leads to a syndrome resembling ageing*. Nature, 1997. **390**(6655): p. 45-51.
- Arking, D.E., et al., Association of human aging with a functional variant of klotho. Proc Natl Acad Sci U S A, 2002. 99(2): p. 856-61.
- 14. de Vries, C.F., et al., *Klotho, APOEepsilon4, cognitive ability, brain size, atrophy, and survival: a study in the Aberdeen Birth Cohort of 1936.*Neurobiol Aging, 2017. 55: p. 91-98.
- 15. Yokoyama, J.S., et al., *Variation in longevity gene KLOTHO is associated with greater cortical volumes.* Ann Clin Transl Neurol, 2015. **2**(3): p. 215-30.
- Tracy, T.E., et al., Acetylated Tau Obstructs KIBRA-Mediated Signaling in Synaptic Plasticity and Promotes Tauopathy-Related Memory Loss. Neuron, 2016. 90(2): p. 245-60.
- 17. Kauppi, K., et al., *KIBRA polymorphism is related to enhanced memory and elevated hippocampal processing*. J Neurosci, 2011. **31**(40): p. 14218-22.

- Witte, A.V., et al., Impact of KIBRA Polymorphism on Memory Function and the Hippocampus in Older Adults. Neuropsychopharmacology, 2016. 41(3): p. 781-90.
- Porter, T., et al., *KIBRA is associated with accelerated cognitive decline and hippocampal atrophy in APOE epsilon4-positive cognitively normal adults with high Abeta-amyloid burden.* Sci Rep, 2018. 8(1): p. 2034.
- 20. Kraus, D.M., et al., *CSMD1 is a novel multiple domain complement*regulatory protein highly expressed in the central nervous system and epithelial tissues. J Immunol, 2006. **176**(7): p. 4419-30.
- 21. Athanasiu, L., et al., *A genetic association study of CSMD1 and CSMD2 with cognitive function*. Brain Behav Immun, 2017. **61**: p. 209-216.
- Ho, A. and T.C. Sudhof, *Binding of F-spondin to amyloid-beta precursor protein: a candidate amyloid-beta precursor protein ligand that modulates amyloid-beta precursor protein cleavage.* Proc Natl Acad Sci U S A, 2004.
  101(8): p. 2548-53.
- Jahanshad, N., et al., Genome-wide scan of healthy human connectome discovers SPON1 gene variant influencing dementia severity. Proc Natl Acad Sci U S A, 2013. 110(12): p. 4768-73.
- 24. Sherva, R., et al., *Genome-wide association study of the rate of cognitive decline in Alzheimer's disease*. Alzheimers Dement, 2014. **10**(1): p. 45-52.
- 25. Sleegers, K., et al., *A 22-single nucleotide polymorphism Alzheimer's disease risk score correlates with family history, onset age, and cerebrospinal fluid Abeta42.* Alzheimers Dement, 2015. **11**(12): p. 1452-60.

- 26. Andrews, S.J., et al., *Association of genetic risk factors with cognitive decline: the PATH through life project*. Neurobiol Aging, 2016. **41**: p. 150-8.
- 27. Mormino, E.C., et al., *Polygenic risk of Alzheimer disease is associated with early- and late-life processes*. Neurology, 2016. **87**(5): p. 481-8.
- Desikan, R.S., et al., *Genetic assessment of age-associated Alzheimer disease risk: Development and validation of a polygenic hazard score*. PLoS Med, 2017. 14(3): p. e1002258.
- 29. Das, D., et al., *Cognitive ability, intraindividual variability, and common* genetic variants of catechol-O-methyltransferase and brain-derived neurotrophic factor: a longitudinal study in a population-based sample of older adults. Psychol Aging, 2014. **29**(2): p. 393-403.
- 30. Derby, C.A., et al., Screening for predementia AD: time-dependent operating characteristics of episodic memory tests. Neurology, 2013. 80(14): p. 1307-14.
- 31. Ellis, K.A., et al., *The Australian Imaging, Biomarkers and Lifestyle (AIBL)* study of aging: methodology and baseline characteristics of 1112 individuals recruited for a longitudinal study of Alzheimer's disease. Int Psychogeriatr, 2009. 21(4): p. 672-87.
- Burnham, S.C., et al., Novel Statistically-Derived Composite Measures for Assessing the Efficacy of Disease-Modifying Therapies in Prodromal Alzheimer's Disease Trials: An AIBL Study. J Alzheimers Dis, 2015. 46(4): p. 1079-89.
- Donohue, M.C., et al., *The preclinical Alzheimer cognitive composite: measuring amyloid-related decline*. JAMA Neurol, 2014. **71**(8): p. 961-70.

- Burnham, S.C., et al., Comparision of three normative data correction approaches: A cross-sectional evaluation in the AIBL study. Alzheimer's & Dementia, 2014. 10(4): p. P4-293.
- 35. Rowe, C.C., et al., *Amyloid imaging results from the Australian Imaging, Biomarkers and Lifestyle (AIBL) study of aging.* Neurobiol Aging, 2010.
  31(8): p. 1275-83.
- 36. Clark, C.M., et al., *Use of florbetapir-PET for imaging beta-amyloid pathology*. JAMA, 2011. **305**(3): p. 275-83.
- 37. Vandenberghe, R., et al., 18F-flutemetamol amyloid imaging in Alzheimer disease and mild cognitive impairment: a phase 2 trial. Ann Neurol, 2010.
  68(3): p. 319-29.
- Bourgeat, P., et al., *Comparison of MR-less PiB SUVR quantification methods*.
   Neurobiol Aging, 2015. 36 Suppl 1: p. S159-66.
- 39. Rowe, C.C., et al., Predicting Alzheimer disease with beta-amyloid imaging: results from the Australian imaging, biomarkers, and lifestyle study of ageing. Ann Neurol, 2013. 74(6): p. 905-13.
- 40. Villemagne, V.L., et al., *En Attendant Centiloid*. Advances in Research, 2014.
- 41. Brown, B.M., et al., *Influence of BDNF Val66Met on the relationship between physical activity and brain volume*. Neurology, 2014. **83**(15): p. 1345-52.
- 42. RStudio Team, *RStudio: Integrated Development for R*, I. RStudio, Editor.2015: Boston, MA.

- 43. Gueorguieva, R. and J.H. Krystal, *Move over ANOVA: progress in analyzing repeated-measures data and its reflection in papers published in the Archives of General Psychiatry.* Arch Gen Psychiatry, 2004. **61**(3): p. 310-7.
- Burnham, S.C., et al., *Clinical and cognitive trajectories in cognitively healthy elderly individuals with suspected non-Alzheimer's disease pathophysiology (SNAP) or Alzheimer's disease pathology: a longitudinal study.* Lancet Neurol, 2016. 15(10): p. 1044-53.
- 45. Keefe, R.S., et al., *Defining a clinically meaningful effect for the design and interpretation of randomized controlled trials*. Innov Clin Neurosci, 2013.
  10(5-6 Suppl A): p. 4S-19S.
- 46. Foy, C.M., et al., *Diagnosing Alzheimer's disease--non-clinicians and computerised algorithms together are as accurate as the best clinical practice.*Int J Geriatr Psychiatry, 2007. 22(11): p. 1154-63.
- 47. Hogervorst, E., et al., *The validity and reliability of 6 sets of clinical criteria to classify Alzheimer's disease and vascular dementia in cases confirmed post-mortem: added value of a decision tree approach.* Dement Geriatr Cogn Disord, 2003. 16(3): p. 170-80.
- Kumar, A. and T.R. Singh, A New Decision Tree to Solve the Puzzle of Alzheimer's Disease Pathogenesis Through Standard Diagnosis Scoring System. Interdiscip Sci, 2017. 9(1): p. 107-115.
- Hochstetler, H., et al., *Empirically Defining Trajectories of Late-Life Cognitive and Functional Decline*. J Alzheimers Dis, 2016. 50(1): p. 271-82.
- 50. Tripoliti, E.E., et al., *A six stage approach for the diagnosis of the Alzheimer's disease based on fMRI data.* J Biomed Inform, 2010. **43**(2): p. 307-20.

51. Mestizo Gutiérrez, S.L., et al., *Decision trees for the analysis of genes involved in Alzheimer's disease pathology*. Journal of Theoretical Biology, 2014. 357: p. 21-25.
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### 4.3 Epilogue

The study presented in this chapter aimed to investigate whether there is a synergistic effect of genes previously associated with cognition, and further what the best combination of these genes would be. To successfully achieve this aim, a decision tree approach was implemented utilising genetic variants in seven genes, *APOE*, *KIBRA*, *KLOTHO*, *BDNF*, *COMT*, *SPON1* and *CSMD1*. The resultant episodic memory-derived cognitive gene risk profile included all variants apart from *COMT* and defined four groups: *APOE*  $\varepsilon$ 4+ Risk,  $\varepsilon$ 4+ Resilient,  $\varepsilon$ 4- Risk,  $\varepsilon$ 4- Resilient. In measures of verbal episodic memory, global cognition, and clinical progression the *APOE*  $\varepsilon$ 4+ Risk group declined significantly faster that all other groups. This defined cognitive gene risk profile supports the notion that combining genetic variants associated with cognition has utility for prediction of cognitive decline at the preclinical stages of AD, even if independently they do not. It also provides further weight for such variants to be considered for inclusion, along with AD risk associated variants, in polygenic risk score PRS development.

With this view, and considering the lack of utility of AD risk weighted PRS and the overriding impact that *APOE* ɛ4 has, the study presented in Chapter 5 attempts to undertake a novel approach to combine both cognitive performance and AD risk associated variants into a single PRS weighted by a phenotype more suited to the desired outcome, being the prediction of cognitive decline in preclinical AD. Employing such a phenotype specific approach to PRS weighting is hypothesised to yield a PRS that would allow for a more accurate prediction of cognitive decline in preclinical AD.

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## Chapter 5

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# CHAPTER 5: Assessing the utility of a novel method of weighting a priori candidate, cognition and ADrisk associated, genes for predicting cognitive decline in preclinical AD.

### **5.1 Prologue**

The preceding chapters in this thesis have explored several approaches for the prediction of cognitive decline in the preclinical stages of Alzheimer's disease (AD). In Chapter 2 a polygenic risk score (PRS) weighted by AD risk was analyzed that as a whole was significantly associated with cognitive decline. However, after the removal of Apolipoprotein E (APOE) this association was lost, suggesting it had negligible utility above and beyond APOE. The following chapters aimed to broaden the scope for selection of genetic variants to be included in polygenic approaches to predict longitudinal cognitive performance. The studies presented in Chapter 3 provided evidence that a targeted selection of genes with a priori evidence of association with cognitive performance had utility. Whilst Chapter 4 suggested that a genetic risk profile combining the effects of the cognitive risk genes examined in Chapter 3 could be used to define a population of individuals declining at significantly accelerated rates. Chapters 3 and 4 thus suggest that broadening the scope of genetic variants included in a PRS may provide increased utility. However, as suggested by the study presented in Chapter 2, the choice of weighting to apply to such a PRS needs to be carefully considered.

These preceding chapters have therefore laid the foundation and provided the supporting evidence for the study presented in this chapter, which hypothesizes that through the use of an endophenotype weighting of genetic variants previously associated with AD risk and cognitive decline, improved prediction of preclinical rates of cognitive decline would be possible. To address this hypothesis, this study will address the fourth and final aim of the thesis, being to *determine a method of weighting genes associated with both AD-risk and cognitive rates of cognitive in a genetic risk score to improve the prediction of preclinical cognitive rates of change.* 

To achieve this aim, it was proposed to combine genes and variants studied in this thesis, that have previously been associated with either an increased risk for AD (Chapter 2) or associated with cognitive performance (Chapter 3 and 4). These variants would be weighted by effect sizes for decline in verbal episodic memory, one of the earliest cognitive domains to decline, [1-3], in a reference sample of cognitively normal individuals with high brain A $\beta$ -burden. The resultant effect sizes allowed for the calculation of a cognitively weighted PRS (*cw*PRS), the performance of which was then assessed in a further test sample with respect to decline in performance across multiple cognitive composites.

#### **Prologue References:**

- 1. Elias, M.F., et al., *The preclinical phase of Alzheimer disease: a 22-year prospective study of the Framingham Cohort.* Archives of neurology, 2000. **57**(6): p. 808-813.
- Grober, E., et al., *Memory impairment, executive dysfunction, and intellectual decline in preclinical Alzheimer's disease*. Journal of the International Neuropsychological Society, 2008. 14(2): p. 266.

3. Derby, C.A., et al., *Screening for predementia AD Time-dependent operating characteristics of episodic memory tests.* Neurology, 2013. **80**(14): p. 1307-1314.

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## 5.2 A polygenic risk score derived from episodic memory

### weighted genetic variants is associated with cognitive decline

## in preclinical Alzheimer's disease

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## **CHAPTER 6: Overall Discussion**

The limited success of clinical trials for effective disease modifying treatments for Alzheimer's disease (AD) has, in part, been attributed to targeting the disease at symptomatic clinical stages where underlying neurodegeneration is well established [1]. In recognition of this there has been an increased research focus on characterising the preclinical disease stages of AD thereby enabling the implementation of disease modifying interventions at stages where the disease is pre-symptomatic [2]. To achieve this requires the accurate identification of pre-symptomatic individuals who will progress to develop AD (i.e. preclinical AD) and subsequently those who are most likely to present with the fastest rates of change in clinical end-points. Together, the identification of these individuals is critical for the design of preventative or early intervention clinical trials.

The accuracy of identification of individuals at the preclinical stages of AD has improved greatly through the advances in brain imaging, i.e. A $\beta$ -amyloid (A $\beta$ ). This is reflected in both the current diagnostic criteria, which incorporates high brain A $\beta$ burden [2], and the use of A $\beta$  imaging in pre-trial screening of participants. The major limitation of imaging techniques is the high cost involved in serial imaging, deceasing its feasibility as a primary outcome measure in large clinical trials. At present, the efficacy of clinical trials is predominantly determined through the use of a primary endpoint of arresting decline in cognitive performance. This places importance on the ability to identify those individuals whose cognition declines at an observable rate within the trial time frames (typically up to 2 and 4 years for Phase 2 and 3, respectively [3]). A major difficulty with this approach is the variability known to exist in rates of cognitive performance at these earliest stages of the disease process between individuals [4].

As outlined in detail in Chapter 1, while the imaging of  $A\beta$  has increased the accuracy of preclinical AD identification, and thus identifying those who will eventually exhibit cognitive decline, it is less sensitive at predicting the rate of this decline. This suggests that, while  $A\beta$  is a necessary condition for AD, other factors (e.g. imaging, fluid biomarkers or genetics) influence the relationship between this biomarker and clinical disease progression. Recently, there has been an increased focus on tau imaging, the results showing it is associated with cognitive performance [5]. Cerebrospinal Fluid (CSF) biomarkers have been shown to predict risk of AD and cognitive performance. The collection of CSF biomarkers is invasive and so, as with imaging, does not present a viable option for serial collections. At present, there is limited information regarding the genetic factors defining the rates of cognitive performance in the preclinical stages of AD. However, if genetic factors could be established, a combinatory approach with genetics augmenting preclinical AD identification (through  $A\beta$  imaging), could present a practical method to predict the rates of cognitive decline at the early stages of the disease.

Cognitive performance has been observed to be heritable and highly polygenic [6]. In addition to genetic variants identified as conferring risk for AD (as reviewed in [7]), a number of single nucleotide polymorphisms (SNPs) have been associated with cognitive performance independent of an association with AD risk [8-16]. Several methods are currently used for the investigation of combined genetic effects on disease risk and phenotypes [17]. In AD, polygenic influences have been reported for disease risk [18, 19], pathological biomarkers [19-22], and cognitive performance [23]. However, studies reporting on polygenic effects in cognitively normal elderly, particularly on cognitive performance, have had varying outcomes [23-32]. The discrepancy in outcomes could be partly due to a lack of knowledge regarding participants'  $A\beta$  status and thus whether they are truly representative of the preclinical stages of AD. This thesis aimed to provide clarity concerning the inconsistent results previously reported, and further, to improve the methods by which combinations of genetic variants are investigated, particularly with respect to preclinical cognitive performance in AD. The overarching hypothesis of this thesis was that *a combination of genetic factors will influence cognitive rates of change in preclinical Alzheimer's disease*.

To clearly define factors, genetic or otherwise, that contribute to changes in cognition at the earliest pre-clinical stages of AD, highly characterised longitudinal cohorts are required. This thesis benefits from access to a world leading longitudinal cohort, the Australian Imaging, Biomarkers and Lifestyle Study of Aging (AIBL). The AIBL study has collected extensive data from participants at 18-monthly intervals including cognitive, neuroimaging and biological assessments. Currently, the study consists of 7.5 years of longitudinal follow-up, allowing for a detailed analysis of preclinical performance.

Initially, I implemented established strategies to assess combined genetic influence of established AD risk genetic variants on preclinical cognitive performance (Chapter 2). AD-risk effect-size weightings, derived from large AD-risk GWAS [33], were applied and resultant PRSs assessed with respect to cognitive performance. In order to address

the potential biased selection of genetic variants and thus encompass a wider range of gene variants, I then investigated genetic variants previously associated with cognitive performance and assessed their influence in preclinical AD, both independently (Chapter 3) and in combination (Chapter 4). Finally, a novel method of phenotype derived effect-size weighting was applied to both AD-risk and cognition associated gene variants, which allowed for them to be appropriately combined into a PRS and used to predict preclinical cognitive performance (Chapter 5).

# Polygenic risk scores weighted by a measure of AD-risk have limited utility for the prediction of preclinical cognitive performance.

The most common approach currently utilised for investigating the influence of genetic variant combinations in AD, is through the calculation of AD-risk weighted Polygenic Risk Scores (PRSs). Using the calculation methods previously described [19], I *assessed the impact of genetic variants previously associated with AD risk on measures of cognition at a preclinical stage* (Chapter 2). In addition to this aim, associations between an AD-risk weighted PRS, and AD fluid and imaging biomarkers were also investigated. Further, the PRS was investigated both with (PRS<sup> $\overline{c}APOE$ </sup>) and without (PRS<sup> $\overline{s}APOE$ </sup>) Apolipoprotein E (*APOE*), to understand the dependence of these associations on carriage of the *APOE*  $\varepsilon$ 4 allele. The results presented in this chapter confirmed previous reports of significant associations between PRS<sup> $\overline{c}APOE$ </sup> and pathological biomarkers, particularly increased neocortical Aβ [30], and reduced levels of CSF Aβ [19, 21, 28] and Aβ:total-tau ratios. The PRS<sup> $\overline{s}APOE$ </sup> was significantly associated with increased levels of CSF total-tau [27] and phospho-tau [27]. It could be speculated that the contrasting results when including or excluding *APOE* could be explained by differences in genetic associations with specifically different aspects of

AD pathology. It is currently well accepted that *APOE* is strongly associated with brain A $\beta$  burden [34], and it can be hypothesised that the additional genes included in the PRS calculation are more closely associated with Tau. Several studies have observed an influence of a number of these genes on *in vivo* pathological changes in Tau and its propagation [35-37].

As previously established, the use of brain imaging data available within the AIBL study allows for the true identification of individuals in the preclinical stages of AD. When specifically investigating the association between PRSs and cognitive performance in a preclinical AD sample (defined as cognitively normal older adults with high neocortical A $\beta$ ) the following findings were reported. Significant associations were reported between PRS<sup> $\bar{c}APOE$ </sup> and composite measures of cognition, specifically global cognition, verbal episodic memory, and the AIBL Pre Alzheimer's Cognitive Composite (AIBL-PACC). These associations were observed at both study baseline and in terms of longitudinal change. However, after the removal of *APOE* from the calculation of the PRS<sup> $\bar{s}APOE$ </sup> no significant associations were recorded.

These results confirm previous findings reported in the literature, particularly the loss of association after removal of *APOE* [24, 25]. Additional investigations found that individuals within the upper quartile of PRS<sup>cAPOE</sup> scores declined cognitively at rates significantly faster than individuals within all other quartiles. However, closer examination of the distribution of *APOE*  $\epsilon$ 4 carriage amongst these quartiles revealed that this association is most likely driven by the influence of this distribution. It was observed that the upper quartile included all of the *APOE*  $\epsilon$ 4 homozygote individuals and a significant proportion of *APOE*  $\epsilon$ 4 heterozygotes. Further, the proportion of

*APOE*  $\varepsilon$ 4 heterozygotes within quartiles declined with reducing risk. The chapter described above represents one of a small number of studies investigating the utility of AD-risk weighted PRSs for the prediction of preclinical cognitive performance in AD. To the best of the author's knowledge it is the first to thoroughly investigate the influence of *APOE* within a PRS. Specifically, the saturating effect of *APOE*, and the spread of  $\varepsilon$ 4 alleles over the range of the PRS, particularly influencing investigations concerning cognitive measures.

Results from this study suggest that while PRSs calculated in this manner can be utilised for the prediction of AD and pathological AD biomarkers, they lack utility when predicting rates of preclinical cognitive decline. Particularly, this is supported by the observed *APOE* dependent cognitive association. These results, combined with the hypothesis that the genetic architecture of AD-risk likely differs from that of AD progression, informed the decision to investigate genes with *a priori* evidence of influence over broad cognitive performance (Chapter 3).

## Genetic variants previously associated with broad cognitive performance influence rates of cognitive decline in a preclinical AD sample.

A perceived weakness of previous efforts that have investigated polygenic risk in AD is the biased selection of genetic variants which have focused largely on those associated with AD diagnosis. A number of genes and gene variants have been associated with cognitive performance and have been speculated to influence decline in preclinical AD. However, due to a lack of association with clinical AD diagnosis have been largely overlooked or excluded [24] from previous polygenic approaches.

One such example is the non-synonymous variant (rs6265; Val66Met) within brain derived neurotropic factor (*BDNF*). Our group has extensively reported its influence on rates of cognitive performance [9, 10], yet it has negligible influence on AD risk [38]. In previous studies in AIBL, the effect of *BDNF* Val66Met on longitudinal cognitive performance is seen above that of *APOE*  $\varepsilon$ 4 in preclinical AD, specifically in cognitively normal older adults having increased neocortical A $\beta$  burden [9, 10]. In addition to *BDNF*, a number of other genetic variants have been associated with cognitive performance that are independent of AD risk. For these reasons, it was the aim of Chapter 3 to assess the effects of genes with a priori evidence for association with cognition on cognitive rates of change in preclinical AD.

Previous studies have shown associations between the gene variants investigated in Chapter 3 (Kidney Brain expressed protein, *KIBRA* [39]; F-Spondin, *SPON1* [16]; Catechol-O-methyltransferase, *COMT* [40]; Klotho, *KL* [41]) and cognitive performance. No such associations were observed here when assessing the independent influence of each variant in the current studies. However, in the presence of an elevated brain A $\beta$  burden and *APOE*  $\varepsilon$ 4 carriage a number of significant associations are found. It should be noted that the studies presented in this thesis are the first to investigating the interactional effects of the gene variants of interest with A $\beta$  burden and *APOE*  $\varepsilon$ 4 carriage.

Investigation of *KIBRA* rs17070145, in combination with A $\beta$  burden and *APOE*  $\epsilon$ 4 carriage, showed an influence of the gene over memory performance and hippocampal atrophy[14]. Specifically, individuals who had high A $\beta$  burden, carried an *APOE*  $\epsilon$ 4

allele, and homozygote for the *KIBRA* C allele declined significantly faster than those carrying at least one copy of the *KIBRA* T allele [14].

The findings reported in Chapter 3 support those from previous studies reporting a protective influence of the *KIBRA* T allele over cognitive performance [13, 39, 42-45]. Gene expression [39, 46-48], brain activity [13, 39], and functional studies [46, 49, 50] investigating *KIBRA* support the hypothesis that the *KIBRA* T allele promotes resilience to cognitive decline. *KIBRA* expression occurs mainly in those areas responsible for memory function [39, 46], with those carrying a T allele reporting increased hippocampal activity during memory tasks [13, 39]. More recently, the postsynaptic KIBRA protein has been reported to mediate *tau* associated memory decline [50].

Assessment of *SPON1* rs11023139 resulted in trends towards significance when investigating the independent effect of this variant on cognitive performance, particularly in measures of global cognition and verbal episodic memory. As in the analysis of *KIBRA*, after interaction with A $\beta$  burden and *APOE*  $\varepsilon$ 4 carriage, significant associations between SPON1 rs11023139 and cognitive performance were observed. High A $\beta$  burden, carriage of an *APOE*  $\varepsilon$ 4 allele, and a *SPON1* A allele resulted in cognitive decline at a significantly faster rate compared to those with the same A $\beta$ burden and *APOE*  $\varepsilon$ 4 carriage carrying no *SPON1* A alleles. These findings related to this variant are the first to build upon those initially reported in the Alzheimer's disease neuroimaging initiative (ADNI) [16].

Biologically, functions of the protein (Spondin-1) encoded by *SPON1* support the genetic variant findings described above. Specifically, Spondin-1 is involved in

neuronal development in embryos [51], regeneration of axons [52], and regulation of amyloid precursor protein (APP) cleavage by beta-secretase [53]. The specific effects of identified genetic variation within *SPON1* have not been assessed, and further such investigation will be valuable.

In contrast with the analysis of the two previous gene variants, analysis of *COMT* Val158Met and *KL*-VS variations revealed no significant associations with cognitive performance were observed, even after interaction with A $\beta$  burden and *APOE*  $\varepsilon$ 4 carriage. While the current study assessing *COMT* Val158Met was unable to replicate previous reported associations with cognitive performance [54-58], it is not the first to report no effect of the variant [59-61]. Likewise, previous reports investigating the influence of *KL*-VS have differed with a number reporting that heterozygosity was associated with improved cognitive performance or reduced decline [12, 41, 62, 63], and a further study similarly reporting no associations [62].

While significant associations were not observed for all variants investigated here, it cannot be assumed that they confer no influence over preclinical cognitive decline. As observed in *KIBRA* and *SPON1*, where significant associations were only observed after interaction with *APOE*, it could be hypothesised that variants within *KL* and *COMT* require combination with additional genetic factors before their impact is observable. As such, studies outlined in Chapters 4 and 5 were undertaken that aimed to determine the influence on cognitive performance of the combination of the cognition associated genes studied in Chapter 3 (Chapter 4) and then the combination of both AD-risk and cognition associated genes (Chapter 5).

# Combining genes previously associated with cognition can define profiles of risk for, and resilience to, preclinical decline in cognition.

The Initial aim was to focus on those genes previously associated with cognitive performance, specifically, *investigate whether there is a synergistic effect of genes previously associated with cognition, and further what the best combination of these genes would be* (Chapter 4). Investigating a reduced number of genes known to influence cognition was proposed to be optimal for use in a simple method for routine clinical use.

Clinically, decision tree based risk profiles have been utilised for risk triage and disease differentiation in neurodegenerative disorders [64-68]. Thus, a cognitive genetic risk profile (*Cog-GRP*) was developed using a statistically calculated decision tree driven by longitudinal change in verbal episodic memory. This profile was developed in cognitively normal individuals with high neocortical A $\beta$  burden. In addition to the genes investigated in the previous chapter (*KIBRA*, *SPON1*, *COMT*, *KL*), *APOE* [8], *BDNF* [9, 10] and *CSMDI* (CUB and Sushi Multiple Domains 1) [15, 69] were also included in the definition of the *Cog-GRP* based on their associations with cognitive performance in other studies.

Calculation of the *Cog-GRP* resulted in the utilisation of all genes with the exception of *COMT*. No significant associations were observed independently between *COMT* and cognition, as presented in Chapter 3. This finding supports the results of a number of previous studies including a large meta-analysis [70]. Four classifications were determined based on the *Cog-GRP* after the groups were collapsed based on sample size and similarities in rates of cognitive change;  $\varepsilon$ 4+ Risk,  $\varepsilon$ 4+ Resilient,  $\varepsilon$ 4- Risk and  $\epsilon$ 4- Resilient. The most at risk group,  $\epsilon$ 4+ Risk, was reported to decline significantly faster on additional measures of cognition including; global cognition and clinical progression, compared to all other groups. Further, it was observed that when comparing to *APOE* alone, those individuals in the  $\epsilon$ 4+ Risk group, declined to a clinically significant threshold (1.5 standard deviations lower in cognitive performance than controls) twice as fast, 5 years compared to 9.6 years. While the decision tree method is not novel, the combination of the genes presented here in the *Cog-GRP* is the first to the author's knowledge to be published.

# A phenotypically relevant weighting of genetic variants can define a polygenic risk score for preclinical cognitive performance in the presence and absence of APOE.

The PRS<sup> $\overline{c}APOE$ </sup> and PRS<sup> $\overline{s}APOE$ </sup> described in Chapter 2 were calculated through applying an AD-risk weighting. The admixture of additional genetic variants was observed to have no effect above that of *APOE* alone, despite these genes having been previously associated with AD-risk and cognitive performance [24, 71-76]. In additional, despite being previously associated with cognition, the genes presented in Chapters 3 and 4 have been, at best, weakly associated with AD risk. For these reasons, when combining AD-risk and cognitive-risk genes the current most utilised method of polygenic risk score calculation, AD-risk weighting, was deemed inappropriate. As such, it was aimed to *determine a method of weighting genes associated with both AD-risk and cognition, for use in a genetic risk score to improve the prediction of preclinical cognitive rates of change* (Chapter 5).

While previously published PRSs, with conservative SNP inclusions, have largely not included variants associated with cognitive performance, or have excluded them based

on lack of influence [24], results from the aforementioned *Cog-GRP* provides evidence for their inclusion. To accurately account for the impact of the included genetic variants on preclinical cognitive performance, each was weighted by an effect size associated with verbal episodic memory performance over 7.5 years. As in the development and testing of previous genetic associations, these weightings were created and tested in a preclinical AD sample as defined by cognitive normality and high A $\beta$  burden. The resulting PRS (*cw*PRS) was tested in an additional sample and found to be associated with cognitive performance, specifically, verbal episodic memory, global cognition and the AIBL-PACC. The *cw*PRS was also associated with cognitive performance after the exclusion of *APOE* from score calculation. This thesis therefore presents, to the best of this researcher's knowledge, the first cognitively weighted PRS developed in preclinical AD. It is also one of a small number of PRSs with the ability to predict longitudinal cognitive performance in a cognitively normal sample.

### 6.1 Limitations

Despite attempts to overcome weaknesses within the studies presented, the following limitations of the results reported in this thesis are acknowledged. Limitations exist which are specifically related to the cohort utilised in the studies. The Australian Imaging, Biomarkers and Lifestyle Study of Aging (AIBL) cohort, data from which was utilised in all studies presented here, represents a Caucasian population which is not representative of the wider community in Australia or globally. The voluntary recruitment of AIBL participants has led to high levels of education observed in the cohort which results in cognitive performances above expectations [77]. As of 2018 the AIBL Study has an extensive 7.5-year follow-up period. Whilst this is valuable when investigating the AIBL study independently, it has the potential to hinder the ability to

validate these results in similar studies with reduced follow-up periods. Despite AIBL being a relatively large longitudinal cohort, at times reduced sample sizes are reported due to genetic stratification. Some studies presented in this thesis and therefore the reference profiles and measures developed may be difficult to replicate when investigated in smaller or less comprehensive cohorts.

This thesis focused on rates of cognitive performance in preclinical AD. It aimed to measure performance in cognitive domains known to be impacted in the early stages of AD (particularly verbal episodic memory). For this reason, statistically derived cognitive composites previously developed in the AIBL study were utilised. This is considered a limitation as the cognitive composite scores were developed based on the AIBL neuropsychological test battery, which differs from batteries administered in other studies. The absence of similar scores for the precise measurement of cognitive performance could impact on the ability to validate the results in other cohorts. At the time of completing this thesis there was a concerted effort underway amongst the cognitive arms of large prospective longitudinal cohorts to address this and define cognitive measures which would allow for future ease of cross-validation.

Finally, it is widely accepted that the *APOE*  $\varepsilon$ 4 allele is the strongest genetic risk factor for AD, with carriage of one increasing an individual's risk for AD by four times and carriage of two by twenty times [78]. Within the studies presented here, and in the wider community, there is an overlap observed between increased neocortical Aβ-amyloid burden and carriage of an *APOE*  $\varepsilon$ 4 allele. This could confound the investigations of genetic interactions in those with high Aβ burden. That being said, in Chapter 5, this thesis presents a novel PRS that shows utility independent of *APOE*. There was an observable reduction in association with the exclusion of *APOE* and so further studies are recommended to address this potential confounder.

#### **6.2 Future Directions**

Published studies investigating polygenic risk in AD currently focus on AD-risk as a weighting measure [18-28, 31, 32, 79-87]. Only recently have studies been presented taking into account addition phenotypic weighting factors [88]. Further, even in these recent studies, a lack of understanding of polygenic risk in the prediction of preclinical cognitive performance in AD remains.

The research presented here confirms the *APOE* dependent nature of associations between AD-risk weighted PRSs and cognition [24, 25], and further describes the influence of specific genetic variants on cognitive performance in preclinical AD. To build upon these findings, a phenotype weighted PRS was developed and found to be associated with cognitive decline over 7.5 years. Further validation of the methods investigated in this thesis is required in other independent longitudinal studies with similar phenotypic information. This will increase the ability to assess clinical utility of the work presented. Outlined here are future directions of the work currently presented, including those which would possibly allow for its transition into a clinical setting.

Presented in all studies in the current thesis are results based on the assessment of previously developed cognitive composite scores [89]. These scores were statistically driven to best represent the verbal episodic memory domain, global cognition, and changes that occur in preclinical AD (AIBL-PACC) [90]. As described in the limitations above, while these composite scores strengthen the study by specifically

measuring domains impacted early in AD, there is a lack of comparable scores in the other large cohort studies. A number of these studies exist, most namely ADNI, however differences in the cognitive battery undertaken present difficulties for replication. In order to replicate the results presented in this thesis, the development of comparable cognitive composite scores in additional large cohort studies would be required, followed by development of reference measures for the weighting of genetic variants. Presented here are studies reporting on weighting of genetic variants in relatively small sample sizes (~150-600 participants). Currently the most utilised reference measure for the weighting of PRSs, is the odds ratio for AD-risk as calculated in a meta-analysis from the International Genomics of Alzheimer's Project (IGAP) consisting of >50,000 participants [33]. For wide scale utility of the methods discussed in this thesis, larger reference cohorts would be required to ensure wider validations.

Once the results presented in the current study are adequately validated, the aim would be for the methods and scores described to transition from research use into a clinical setting, with particular utility in patient selection for clinical trials. Clinical trials utilising genetics are currently occurring in a number of diseases including in AD. The TOMORROW Study aimed to focus drug treatment on individuals based on their *APOE* genotype and Translocase of Outer Mitochondrial Membrane 40 (*TOMM40*) repeat status, although it has recently failed due to lack of treatment effect.

As discussed previously, neocortical amyloid imaging alone is unable to predict individuals decline. The polygenic approaches developed here are targeted at individuals with high  $A\beta$  which needs to be identified through amyloid imaging. The recent development of plasma  $A\beta$  biomarkers being reported to predict brain  $A\beta$  burden, could mean this can be utilised in the identification of individuals at risk of decline [91]. The combination of plasma biomarkers and genetic testing would represent an inexpensive and relatively non-invasive screening method for clinical trials, as a single blood sample would be sufficient for both tests.

Multimodal approaches for the development of risk profiles in preclinical AD are valuable. In addition to genetic variation, amyloid imaging and CSF biomarkers, recent developments in the mapping of brain iron have been shown to predict cognitive performance [92]. The use of multiple methods to predict cognitive performance will increase the likelihood of selecting appropriate clinical trial participants. Genetic testing would not only allow for the selection of appropriate trial cohorts after the identification of those with high brain A $\beta$  burden, but could also reduce the number of individuals initially requiring amyloid imaging for confirmation of their preclinical AD status.

Due to the biological actions of drug candidates for AD treatments, many of these result in adverse side effects [93]. The polygenic approaches described here could assist in the movement towards personalised medicine [94], wherein individuals are prescribed medications, including adjusted doses, dependent on their expected rate of preclinical decline. Individuals expected to progress towards disease at an increased rate could require more aggressive levels of treatment to halt decline. Conversely, tailoring treatments based on rates of preclinical decline could also reduce side effects in those patients not declining at accelerated rates, by possibly lowering required dosages. Finally, while outcomes from the current study aim to play a role in the enrolment of appropriate individuals for clinical trials going forward, they could also have utility in the analysis of historical clinical trial data. Particularly in AD, there is a wealth of information from clinical trials which is being further investigated to better understand the reasons for their failures [95]. Improving the understanding of previous clinical trials and the reasons for their failures is important in moving forward with new treatments and targets. Retrospective trial analyses and a more complete understanding of the natural history of AD have resulted in the current changes to focus clinical trials on the preclinical disease phase. Having a better understanding of individuals' genetic composition could also assist in understanding the possible reasons for the prior trial failures. This could include determining whether, based on the time frames and individuals investigated, any change in cognitive performance could have been expected.

### 6.3 General Conclusion

This thesis provided a thorough investigation of genetic influence over rates of cognitive performance in preclinical AD. The work highlights the importance of polygenic approaches in association studies and the limitations of the current methods, particularly in preclinical disease. The findings provide evidence that cognitive performance in preclinical AD is genetically influenced and that the genetic architecture of cognitive decline does not mirror that of AD-risk. This understanding of the genetic influences over rates of preclinical cognitive performance has significant implications in clinical trial design. Combined genetic approaches may assist the selection of those individuals that are likely to show rapid cognitive decline for inclusion in preclinical AD trials, allowing these trials to be conducted in feasible time

frames. Whilst further study is required to validate and build on the results presented here for their transition into a clinical setting, appropriate participant inclusion in AD preclinical trials would improve the likelihood of identifying an appropriate treatment for AD, reducing the enormous global impact of the disease.

#### **6.4 References**

- Cummings, J.L., T. Morstorf, and K. Zhong, *Alzheimer's disease drugdevelopment pipeline: few candidates, frequent failures*. Alzheimers Res Ther, 2014. 6(4): p. 37.
- Sperling, R.A., et al., Toward defining the preclinical stages of Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease.
   Alzheimers Dement, 2011. 7(3): p. 280-92.
- U.S. Food & Drug Administration. *Clinical Research*. 2018 [cited 2018 20/04/2018]; Available from: https://www.fda.gov/ForPatients/Approvals/Drugs/ucm405622.htm.
- 4. Wilson, R.S., et al., *Individual differences in rates of change in cognitive abilities of older persons*. Psychol Aging, 2002. **17**(2): p. 179-93.
- 5. Johnson, K.A., et al., *Tau positron emission tomographic imaging in aging and early Alzheimer disease*. Ann Neurol, 2016. **79**(1): p. 110-9.
- Gatz, M., et al., *Role of genes and environments for explaining Alzheimer disease*. Arch Gen Psychiatry, 2006. 63(2): p. 168-74.
- 7. Karch, C.M. and A.M. Goate, *Alzheimer's disease risk genes and mechanisms of disease pathogenesis.* Biol Psychiatry, 2015. **77**(1): p. 43-51.
- Liu, Y., et al., APOE genotype and neuroimaging markers of Alzheimer's disease: systematic review and meta-analysis. J Neurol Neurosurg Psychiatry, 2015. 86(2): p. 127-34.

- 9. Lim, Y.Y., et al., *BDNF Val66Met, Abeta amyloid, and cognitive decline in preclinical Alzheimer's disease*. Neurobiol Aging, 2013. **34**(11): p. 2457-64.
- Lim, Y.Y., et al., APOE and BDNF polymorphisms moderate amyloid betarelated cognitive decline in preclinical Alzheimer's disease. Mol Psychiatry, 2015. 20(11): p. 1322-8.
- Sheldrick, A.J., et al., *Effect of COMT val158met genotype on cognition and personality*. Eur Psychiatry, 2008. 23(6): p. 385-9.
- de Vries, C.F., et al., *Klotho, APOEepsilon4, cognitive ability, brain size, atrophy, and survival: a study in the Aberdeen Birth Cohort of 1936.*Neurobiol Aging, 2017. 55: p. 91-98.
- 13. Kauppi, K., et al., *KIBRA polymorphism is related to enhanced memory and elevated hippocampal processing*. J Neurosci, 2011. **31**(40): p. 14218-22.
- Porter, T., et al., *KIBRA is associated with accelerated cognitive decline and hippocampal atrophy in APOE epsilon4-positive cognitively normal adults with high Abeta-amyloid burden.* Sci Rep, 2018. 8(1): p. 2034.
- 15. Athanasiu, L., et al., *A genetic association study of CSMD1 and CSMD2 with cognitive function*. Brain Behav Immun, 2017. **61**: p. 209-216.
- 16. Sherva, R., et al., *Genome-wide association study of the rate of cognitive decline in Alzheimer's disease*. Alzheimers Dement, 2014. **10**(1): p. 45-52.
- Maher, B.S., *Polygenic Scores in Epidemiology: Risk Prediction, Etiology,* and Clinical Utility. Curr Epidemiol Rep, 2015. 2(4): p. 239-244.

- Adams, H.H., et al., *Genetic risk of neurodegenerative diseases is associated with mild cognitive impairment and conversion to dementia*. Alzheimers Dement, 2015. 11(11): p. 1277-85.
- Sleegers, K., et al., A 22-single nucleotide polymorphism Alzheimer's disease risk score correlates with family history, onset age, and cerebrospinal fluid Abeta42. Alzheimers Dement, 2015. 11(12): p. 1452-60.
- Habes, M., et al., Advanced brain aging: relationship with epidemiologic and genetic risk factors, and overlap with Alzheimer disease atrophy patterns.
   Transl Psychiatry, 2016. 6: p. e775.
- Martiskainen, H., et al., *Effects of Alzheimer's disease-associated risk loci on cerebrospinal fluid biomarkers and disease progression: a polygenic risk score approach.* J Alzheimers Dis, 2015. 43(2): p. 565-73.
- 22. Biffi, A., et al., *Genetic variation and neuroimaging measures in Alzheimer disease*. Arch Neurol, 2010. **67**(6): p. 677-85.
- Marden, J.R., et al., Using an Alzheimer Disease Polygenic Risk Score to Predict Memory Decline in Black and White Americans Over 14 Years of Follow-up. Alzheimer Dis Assoc Disord, 2016. 30(3): p. 195-202.
- 24. Andrews, S.J., et al., *Association of genetic risk factors with cognitive decline: the PATH through life project*. Neurobiol Aging, 2016. **41**: p. 150-8.
- 25. Carrasquillo, M.M., et al., Late-onset Alzheimer's risk variants in memory decline, incident mild cognitive impairment, and Alzheimer's disease.
  Neurobiol Aging, 2015. 36(1): p. 60-7.
- 26. Mormino, E.C., et al., *Polygenic risk of Alzheimer disease is associated with early- and late-life processes*. Neurology, 2016. **87**(5): p. 481-8.

- 27. Louwersheimer, E., et al., *Alzheimer's disease risk variants modulate endophenotypes in mild cognitive impairment*. Alzheimers Dement, 2016.
  12(8): p. 872-81.
- 28. Sabuncu, M.R., et al., *The association between a polygenic Alzheimer score and cortical thickness in clinically normal subjects*. Cereb Cortex, 2012.
  22(11): p. 2653-61.
- 29. Bressler, J., et al., Genetic variants associated with risk of Alzheimer's disease contribute to cognitive change in midlife: The Atherosclerosis Risk in Communities Study. Am J Med Genet B Neuropsychiatr Genet, 2017. 174(3): p. 269-282.
- 30. Gui, H., et al., *Influence of Alzheimer's disease genes on cognitive decline: the Guangzhou Biobank Cohort Study*. Neurobiol Aging, 2014. 35(10): p. 2422
  e3-8.
- Harrison, T.M., et al., An Alzheimer's Disease Genetic Risk Score Predicts Longitudinal Thinning of Hippocampal Complex Subregions in Healthy Older Adults. eNeuro, 2016. 3(3).
- 32. Darst, B.F., et al., *Pathway-Specific Polygenic Risk Scores as Predictors of Amyloid-beta Deposition and Cognitive Function in a Sample at Increased Risk for Alzheimer's Disease.* J Alzheimers Dis, 2017. **55**(2): p. 473-484.
- Lambert, J.C., et al., *Meta-analysis of 74,046 individuals identifies 11 new* susceptibility loci for Alzheimer's disease. Nat Genet, 2013. 45(12): p. 1452-8.
- 34. Liu, C.C., et al., *ApoE4 Accelerates Early Seeding of Amyloid Pathology*.
  Neuron, 2017. 96(5): p. 1024-1032 e3.

- 35. Chapuis, J., et al., *Increased expression of BIN1 mediates Alzheimer genetic risk by modulating tau pathology*. Mol Psychiatry, 2013. **18**(11): p. 1225-34.
- Dourlen, P., et al., Functional screening of Alzheimer risk loci identifies PTK2B as an in vivo modulator and early marker of Tau pathology. Mol Psychiatry, 2017. 22(6): p. 874-883.
- Moreau, K., et al., *PICALM modulates autophagy activity and tau accumulation*. Nat Commun, 2014. 5: p. 4998.
- 38. Bertram, L., et al., *Systematic meta-analyses of Alzheimer disease genetic association studies: the AlzGene database*. Nat Genet, 2007. **39**(1): p. 17-23.
- 39. Papassotiropoulos, A., et al., *Common Kibra alleles are associated with human memory performance*. Science, 2006. **314**(5798): p. 475-8.
- 40. Egan, M.F., et al., *Effect of COMT Val108/158 Met genotype on frontal lobe function and risk for schizophrenia*. Proc Natl Acad Sci U S A, 2001. 98(12): p. 6917-22.
- 41. Dubal, D.B., et al., *Life extension factor klotho enhances cognition*. Cell Rep, 2014. 7(4): p. 1065-76.
- 42. Almeida, O.P., et al., *KIBRA genetic polymorphism influences episodic memory in later life, but does not increase the risk of mild cognitive impairment.* J Cell Mol Med, 2008. **12**(5A): p. 1672-6.
- 43. Muse, J., et al., WWC1 genotype modulates age-related decline in episodic memory function across the adult life span. Biol Psychiatry, 2014. 75(9): p. 693-700.

- Witte, A.V., et al., Impact of KIBRA Polymorphism on Memory Function and the Hippocampus in Older Adults. Neuropsychopharmacology, 2016. 41(3): p. 781-90.
- 45. Yasuda, Y., et al., *Association study of KIBRA gene with memory performance in a Japanese population*. World J Biol Psychiatry, 2010. **11**(7): p. 852-7.
- 46. Johannsen, S., et al., *Temporal-spatial expression and novel biochemical properties of the memory-related protein KIBRA*. Neuroscience, 2008. 155(4):
  p. 1165-73.
- Burgess, J.D., et al., Association of common KIBRA variants with episodic memory and AD risk. Neurobiol Aging, 2011. 32(3): p. 557 e1-9.
- 48. Corneveaux, J.J., et al., *Evidence for an association between KIBRA and lateonset Alzheimer's disease*. Neurobiol Aging, 2010. **31**(6): p. 901-9.
- 49. Piras, I.S., et al., Whole transcriptome profiling of the human hippocampus suggests an involvement of the KIBRA rs17070145 polymorphism in differential activation of the MAPK signaling pathway. Hippocampus, 2017.
  27(7): p. 784-793.
- Tracy, T.E., et al., Acetylated Tau Obstructs KIBRA-Mediated Signaling in Synaptic Plasticity and Promotes Tauopathy-Related Memory Loss. Neuron, 2016.
- 51. Feinstein, Y., et al., *F-spondin and mindin: two structurally and functionally related genes expressed in the hippocampus that promote outgrowth of embryonic hippocampal neurons.* Development, 1999. **126**(16): p. 3637-48.

- Burstyn-Cohen, T., et al., *Accumulation of F-spondin in injured peripheral nerve promotes the outgrowth of sensory axons*. J Neurosci, 1998. 18(21): p. 8875-85.
- 53. Bai, Y., et al., *The in vivo brain interactome of the amyloid precursor protein*.Mol Cell Proteomics, 2008. 7(1): p. 15-34.
- 54. Bellander, M., et al., Lower baseline performance but greater plasticity of working memory for carriers of the val allele of the COMT Val(1)(5)(8)Met polymorphism. Neuropsychology, 2015. 29(2): p. 247-54.
- Starr, J.M., et al., *COMT genotype and cognitive ability: a longitudinal aging study*. Neurosci Lett, 2007. 421(1): p. 57-61.
- 56. Papenberg, G., et al., *COMT polymorphism and memory dedifferentiation in old age*. Psychol Aging, 2014. **29**(2): p. 374-83.
- 57. Nagel, I.E., et al., *Human aging magnifies genetic effects on executive functioning and working memory*. Front Hum Neurosci, 2008. **2**: p. 1.
- Degen, C., et al., *The COMTp. Val158Met Polymorphism and Cognitive Performance in Adult Development, Healthy Aging and Mild Cognitive Impairment*. Dement Geriatr Cogn Disord, 2016. 41(1-2): p. 27-34.
- 59. Stuart, K., et al., *BDNF and COMT polymorphisms have a limited association with episodic memory performance or engagement in complex cognitive activity in healthy older adults.* Neurobiol Learn Mem, 2014. **110**: p. 1-7.
- 60. O'Hara, R., et al., *COMT genotype, gender and cognition in communitydwelling, older adults*. Neurosci Lett, 2006. **409**(3): p. 205-9.
- 61. de Frias, C.M., et al., *Influence of COMT gene polymorphism on fMRIassessed sustained and transient activity during a working memory task.* J Cogn Neurosci, 2010. **22**(7): p. 1614-22.
- 62. Morar, B., et al., *The longevity gene Klotho is differentially associated with cognition in subtypes of schizophrenia*. Schizophr Res, 2017.
- 63. Deary, I.J., et al., *KLOTHO genotype and cognitive ability in childhood and old age in the same individuals*. Neurosci Lett, 2005. **378**(1): p. 22-7.
- 64. Foy, C.M., et al., *Diagnosing Alzheimer's disease--non-clinicians and computerised algorithms together are as accurate as the best clinical practice*. Int J Geriatr Psychiatry, 2007. 22(11): p. 1154-63.
- 65. Hogervorst, E., et al., *The validity and reliability of 6 sets of clinical criteria to classify Alzheimer's disease and vascular dementia in cases confirmed post-mortem: added value of a decision tree approach.* Dement Geriatr Cogn Disord, 2003. **16**(3): p. 170-80.
- Kumar, A. and T.R. Singh, A New Decision Tree to Solve the Puzzle of Alzheimer's Disease Pathogenesis Through Standard Diagnosis Scoring System. Interdiscip Sci, 2017. 9(1): p. 107-115.
- 67. Tripoliti, E.E., et al., *A six stage approach for the diagnosis of the Alzheimer's disease based on fMRI data.* J Biomed Inform, 2010. **43**(2): p. 307-20.
- Mestizo Gutiérrez, S.L., et al., *Decision trees for the analysis of genes involved in Alzheimer's disease pathology*. Journal of Theoretical Biology, 2014. 357: p. 21-25.

- 69. Kraus, D.M., et al., *CSMD1 is a novel multiple domain complement*regulatory protein highly expressed in the central nervous system and epithelial tissues. J Immunol, 2006. **176**(7): p. 4419-30.
- Barnett, J.H., L. Scoriels, and M.R. Munafo, *Meta-analysis of the cognitive effects of the catechol-O-methyltransferase gene Val158/108Met polymorphism.* Biol Psychiatry, 2008. 64(2): p. 137-44.
- 71. Chung, S.J., et al., *CR1, ABCA7, and APOE genes affect the features of cognitive impairment in Alzheimer's disease.* J Neurol Sci, 2014. 339(1-2): p. 91-6.
- 72. Pedraza, O., et al., *Evaluation of memory endophenotypes for association with CLU, CR1, and PICALM variants in black and white subjects.* Alzheimers Dement, 2014. **10**(2): p. 205-13.
- Greenbaum, L., et al., Potential contribution of the Alzheimer's disease risk locus BIN1 to episodic memory performance in cognitively normal Type 2 diabetes elderly. Eur Neuropsychopharmacol, 2016. 26(4): p. 787-95.
- Liu, Y., et al., Association between NME8 locus polymorphism and cognitive decline, cerebrospinal fluid and neuroimaging biomarkers in Alzheimer's disease. PLoS One, 2014. 9(12): p. e114777.
- 75. Davies, G., et al., Genetic contributions to variation in general cognitive function: a meta-analysis of genome-wide association studies in the CHARGE consortium (N=53949). Mol Psychiatry, 2015. 20(2): p. 183-92.
- 76. Nettiksimmons, J., et al., *Gene-based aggregate SNP associations between candidate AD genes and cognitive decline*. Age (Dordr), 2016. **38**(2): p. 41.

- Filis, K.A., et al., *The Australian Imaging, Biomarkers and Lifestyle (AIBL)* study of aging: methodology and baseline characteristics of 1112 individuals recruited for a longitudinal study of Alzheimer's disease. Int Psychogeriatr, 2009. 21(4): p. 672-87.
- Farrer, L.A., et al., *Effects of age, sex, and ethnicity on the association* between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer Disease Meta Analysis Consortium. JAMA, 1997.
  278(16): p. 1349-56.
- 79. Chouraki, V., et al., Evaluation of a Genetic Risk Score to Improve Risk
   Prediction for Alzheimer's Disease. J Alzheimers Dis, 2016. 53(3): p. 921-32.
- Foley, S.F., et al., Multimodal Brain Imaging Reveals Structural Differences in Alzheimer's Disease Polygenic Risk Carriers: A Study in Healthy Young Adults. Biol Psychiatry, 2017. 81(2): p. 154-161.
- 81. Lacour, A., et al., Genome-wide significant risk factors for Alzheimer's disease: role in progression to dementia due to Alzheimer's disease among subjects with mild cognitive impairment. Mol Psychiatry, 2017. 22(1): p. 153-160.
- 82. Lupton, M.K., et al., *The effect of increased genetic risk for Alzheimer's disease on hippocampal and amygdala volume*. Neurobiol Aging, 2016. 40: p. 68-77.
- Rodriguez-Rodriguez, E., et al., *Genetic risk score predicting accelerated progression from mild cognitive impairment to Alzheimer's disease*. J Neural Transm (Vienna), 2013. **120**(5): p. 807-12.

- 84. Chauhan, G., et al., *Association of Alzheimer's disease GWAS loci with MRI markers of brain aging*. Neurobiol Aging, 2015. **36**(4): p. 1765 e7-16.
- 85. Escott-Price, V., et al., *Polygenic risk score analysis of pathologically confirmed alzheimer disease*. Ann Neurol, 2017.
- Escott-Price, V., et al., Common polygenic variation enhances risk prediction for Alzheimer's disease. Brain, 2015. 138(Pt 12): p. 3673-84.
- 87. Voyle, N., et al., *Genetic Risk as a Marker of Amyloid-beta and Tau Burden in Cerebrospinal Fluid.* J Alzheimers Dis, 2017. **55**(4): p. 1417-1427.
- 88. Desikan, R.S., et al., *Genetic assessment of age-associated Alzheimer disease risk: Development and validation of a polygenic hazard score*. PLoS Med, 2017. 14(3): p. e1002258.
- Burnham, S.C., et al., Novel Statistically-Derived Composite Measures for Assessing the Efficacy of Disease-Modifying Therapies in Prodromal Alzheimer's Disease Trials: An AIBL Study. J Alzheimers Dis, 2015. 46(4): p. 1079-89.
- 90. Donohue, M.C., et al., *The preclinical Alzheimer cognitive composite: measuring amyloid-related decline*. JAMA Neurol, 2014. **71**(8): p. 961-70.
- 91. Nakamura, A., et al., *High performance plasma amyloid-beta biomarkers for Alzheimer's disease*. Nature, 2018. **554**(7691): p. 249-254.
- 92. Ayton, S., et al., *Cerebral quantitative susceptibility mapping predicts amyloid-beta-related cognitive decline*. Brain, 2017. **140**(8): p. 2112-2119.
- Hung, S.Y. and W.M. Fu, *Drug candidates in clinical trials for Alzheimer's disease*. J Biomed Sci, 2017. 24(1): p. 47.

- 94. Vogenberg, F.R., C. Isaacson Barash, and M. Pursel, *Personalized medicine: part 1: evolution and development into theranostics*. P T, 2010. **35**(10): p. 560-76.
- 95. Cummings, J., Lessons Learned from Alzheimer Disease: Clinical Trials with Negative Outcomes. Clin Transl Sci, 2018. **11**(2): p. 147-152.

## Appendices