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Ranking candidate genes of esophageal squamous cell carcinomas based on differentially expressed genes and the topological properties of the co-expression network

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Abstract

Background: The aim of this study was to identify the candidate genes of esophageal squamous cell carcinoma (ESCC).

Methods: Gene expression profiling of 17 ESCC samples and 17 adjacent normal samples, GSE20347, was downloaded from Gene Expression Omnibus database. The raw data were preprocessed, and the differentially expressed genes (DEGs) between ESCC and normal samples were identified by using SAM software (false discovery rate <0.001). Then, the co-expression network of DEGs was constructed based on Pearson's correlation test (r -value ≥ 0.8). Furthermore, the topological properties of the co-expression network were analyzed through NetworkAnalyzer (default settings) of Cytoscape. The expression fold changes of DEGs and topological properties were utilized to identify the candidate genes of ESCC (Cri_n score >4), which were further analyzed based on DAVID functional enrichment analysis (P -value <0.05).

Results: A total of 1,063 DEGs were identified, including 490 up-regulated and 573 down-regulated DEGs. Then, the co-expression network of DEGs was constructed, containing 999 nodes and 46,323 edges. Based on the expression fold changes of DEGs and the topological properties of the co-expression network, DEGs were ranked, and the top 24 genes were candidate genes of ESCC, such as *CRISP3*, *EREG*, *CXCR2*, and *CRNN*. Furthermore, the 24 genes were significantly enriched in bio-functions regarding cell differentiation, glucan biosynthetic process and immune response.

Conclusion: The present study suggested that *CRISP3*, *EREG*, *CXCR2*, and *CRNN* might be causative genes of ESCC, and play vital roles in the development of ESCC. However, further experimental studies are needed to confirm our results.

Keywords: Esophageal squamous cell carcinomas, Differentially expressed genes, Topological properties, Co-expression network, Candidate genes

Background

Esophageal squamous cell carcinoma (ESCC) is the predominant histologic subtype of esophageal cancer, which is characterized by high mortality rate and geographic differences in incidence [1]. ESCC is a common malignant cancer worldwide, especially in China [2]. Recently, clinical therapies have been used to treat ESCC, such as neoadjuvant chemoradiotherapy [3,4], surgery [5], and combination therapy [6]. However, these approaches do not increase survival rate of patients with ESCC. To

investigate novel therapies, a clear understanding of the molecular pathogenesis of ESCC is required, to which much effort has been made.

Microarray analysis has been widely used to investigate gene expression in various diseases, such as breast tumors [7], brain cancer [8], endometrial cancer [9], and renal cell carcinoma [10]. In previous studies of ESCC, loss of heterozygosity (LOH) and copy number alteration in ESCC were identified by using microsatellite markers and low- and high-density SNP arrays [11,12]. The differentially expressed genes (DEGs) and microRNAs between ESCC and normal squamous epithelia have been identified based on microarray analysis [13,14]. Moreover, a comprehensive survey of commonly inactivated tumor suppressor genes

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in ESCC was performed based on microarray analysis and functional reactivation of silenced tumor suppressor genes by 5-aza-2'-deoxycytidine and trichostatin [15]. Thus, microarray analysis is a useful approach to identify key genes involved in ESCC.

In our study, microarrays were utilized to identify the DEGs between human ESCC samples and adjacent normal tissues samples. Then, the co-expression network of DEGs was constructed, and the topological properties of the co-expression network were analyzed. Additionally, we built an integrated index to rank DEGs and identify the candidate genes of ESCC. Furthermore, the relevant functions of candidate genes were investigated. We anticipate that our work can find key genes related with ESCC, and provide new insights for target therapies.

Methods

Microarray data and preprocessing

The raw gene expression profile GSE20347 [2] was downloaded from the public functional genomics database Gene Expression Omnibus (GEO, <http://www.ncbi.nlm.nih.gov/geo/>). In total, 34 specimens were available, including 17 human ESCC samples and 17 matched adjacent normal tissues samples. None of the patients with ESCC had received prior therapy, and informed consent was obtained. Demographic and clinical information were matched. The corresponding platform was GPL571, [HG-U133A_2] Affymetrix Human Genome U133A 2.0 Array. The background correction and normalization of microarray data among chips were conducted based on RMA (Robust Multiarray Averaging) method [16]. When several probes were mapped to one gene, their mean value was taken as the gene expression value of this gene.

DEG screening

SAM 4.0 software [17] was used to screen out the DEGs between ESCC and normal samples. SAM is a widely used tool for DEG screening. The criterion for this analysis was false discovery rate (FDR) <0.001.

Hierarchical clustering analysis of DEGs

To determine the specificity of DEGs between ESCC and adjacent normal samples, the heatmap package in R was utilized to perform bidirectional hierarchical clustering analysis (BHCA) [18,19]. Rationally, after BHCA, ESCC and adjacent normal samples are supposed to be distinguished clearly by DEGs.

Constructing the co-expression network of DEGs

In organisms, biological functions are often based on the interaction of several genes, and significantly co-expressed genes are usually co-regulated and involved in the same or similar biological processes and pathways [20]. In order

to identify the co-expressed DEGs in ESCC tissues, the expression values of DEGs in each sample were abstracted, and Pearson's correlation test was used to calculate the correlation coefficient (r-value) of DEGs. Higher *P*-value represents stronger correlation between node 'n' and its adjacent nodes, and only the co-expressed DEG pairs with r-value ≥ 0.8 were utilized to construct the co-expression network of DEGs, which was visualized by using Cytoscape [21].

Analyzing the topological properties of the co-expression network

The topological properties (node degree and clustering coefficient) of DEGs in the co-expression network were analyzed based on NetworkAnalyzer of Cytoscape software [22]. Node degree and clustering coefficient are important topological properties of network. Node degree ' k_n ' is the number of nodes connected to node 'n', displaying the local centrality of this node in network. Higher node degree represents stronger importance of a node for the stability of network. Clustering coefficient of node 'n' is defined as:

$$C_n = 2e_n / [k_n(k_n - 1)]$$

In this formula, ' k_n ' is the number of adjacent nodes, and ' e_n ' is the number of interconnections among adjacent nodes. Representing the clustering degree of node 'n', C_n is between 0 and 1, and higher C_n represents that the adjacent nodes of node 'n' connected with each other more closely.

DEG ranking and candidate gene identification

Candidate genes of ESCC could be identified by ranking DEGs based on expression fold changes, node degrees, and clustering coefficients in the co-expression network. In this study, firstly, Z-transformation was performed to transform three sets of data into Z-scores, which are common standardized scores in statistics [23]. Secondly, the ranking index of each DEG was calculated based on the formula as follows:

$$Cri_n = FC_n + degree_n + C \quad (1)$$

Here, Cri_n is the ranking index of gene 'n', FC_n is the expression fold change of gene 'n', $degree_n$ is the node degree of gene 'n' in the co-expression network, and C_n is the clustering coefficient of gene 'n' in the co-expression network. Thirdly, the DEGs with Cri_n score >4 were defined as candidate genes of ESCC.

Functional enrichment analysis

To reveal the biological process associated with ESCC, gene ontology (GO) functional enrichment analysis was performed for the candidate genes of ESCC based on

the Database for Annotation, Visualization and Integrated Discovery (DAVID) [24]. DAVID provides exploratory visualization tools to promote functional classification. In this study, the criterion for this analysis was a P -value <0.05 .

Results

Identification of DEGs and hierarchical clustering analysis

After preprocessing and DEG screening, 1,063 DEGs (FDR <0.001) were identified between ESCC and adjacent normal samples, including 490 up-regulated and 573

down-regulated DEGs. After BHCA, ESCC and adjacent normal samples could be distinguished clearly by DEGs (Figure 1).

Construction of the co-expression network

The r -values of DEG pairs were calculated based on Pearson's correlation test, and the co-expression network of DEGs with r -value ≥ 0.8 were constructed (Additional file 1). The co-expression network involved 999 nodes (DEGs) and 46,323 edges (co-expression relationships),

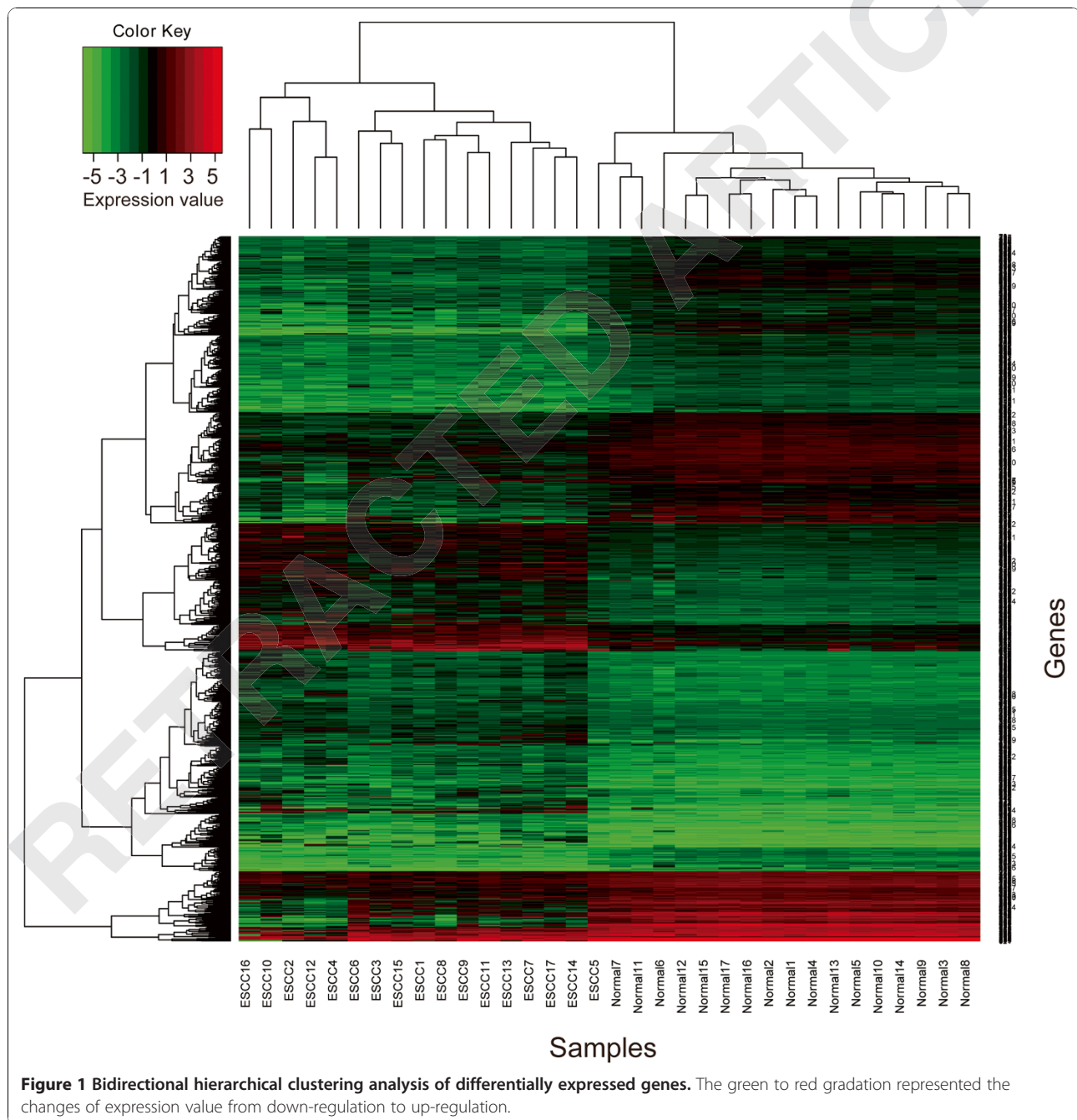


Figure 1 Bidirectional hierarchical clustering analysis of differentially expressed genes. The green to red gradation represented the changes of expression value from down-regulation to up-regulation.

and it was further divided into two closely connected large sub-networks.

DEG ranking and candidate gene identification

To identify the candidate genes of ESCC, we established an integrated ranking index by integrating the expression fold changes, node degrees, and clustering coefficients of DEGs in the co-expression network. Consequently, a total of 24 genes were candidate genes (Cri_n score >4) of ESCC (Table 1), such as cysteine-rich secretory protein 3 (*CRISP3*), epiregulin (*EREG*), chemokine receptor 2 (*CXCR2*), and cornulin (*CRNN*).

Functional enrichment analysis

To understand the biological processes involving candidate genes, GO functional enrichment analysis (P -value <0.05) was performed. It was found that the 24 candidate genes were significantly enriched in bio-functions regarding cell differentiation, glucan biosynthetic process and immune response, including epidermal cell differentiation, epithelial cell differentiation, epidermis development, keratinocyte differentiation, glucan biosynthetic process, and regulation of immune response (Table 2).

Discussion

In the present study, we identified 1,063 DEGs between ESCC and adjacent normal samples, including 490 up-regulated and 573 down-regulated genes. Furthermore, the co-expression network of DEGs was constructed, consisting of 2 large sub-networks, 999 nodes, and 46,323 edges. Then, the expression fold changes, node degrees, and clustering coefficients of DEGs in the co-expression network were comprehensively analyzed to rank DEGs, and 24 candidate genes of ESCC were identified.

Among the 24 candidate genes, the highest ranking was *CRISP3*. It was originally discovered in human

Table 1 The 24 candidate genes of esophageal squamous cell carcinomas

Rank	Symbol	Rank	Symbol
1	<i>CRISP3</i>	13	<i>CYP4B1</i>
2	<i>EREG</i>	14	<i>SLURP1</i>
3	<i>ENDOU</i>	15	<i>MAL</i>
4	<i>CXCR2</i>	16	<i>TGM3</i>
5	<i>CRCT1</i>	17	<i>CWH43</i>
6	<i>FLG</i>	18	<i>TMPRSS11E</i>
7	<i>CLIC3</i>	19	<i>CLCA4</i>
8	<i>CRNN</i>	20	<i>PPP1R3C</i>
9	<i>MMP1</i>	21	<i>BBOX1</i>
10	<i>GYS2</i>	22	<i>HPGD</i>
11	<i>EPB41L3</i>	23	<i>SPINK5</i>
12	<i>SCEL</i>	24	<i>FCERIA</i>

Table 2 Functional enrichment analysis of the 24 candidate genes

GO ID	GO term	P-value
GO:0009913	Epidermal cell differentiation	2.43E-06
GO:0030855	Epithelial cell differentiation	3.14E-05
GO:0008544	Epidermis development	9.93E-05
GO:0030216	Keratinocyte differentiation	1.10E-04
GO:0008544	Epidermis development	1.21E-04
GO:0007398	Ectoderm development	1.34E-04
GO:0060429	Epithelium development	2.23E-04
GO:0009888	Tissue development	1.15E-02
GO:0009250	Glucan biosynthetic process	1.70E-02
GO:0005978	Glycogen biosynthetic process	1.70E-02
GO:0033692	Cellular polysaccharide biosynthetic process	3.54E-02
GO:0050776	Regulation of immune response	4.14E-02
GO:0044042	Glucan metabolic process	4.87E-02
GO:0006073	Cellular glucan metabolic process	4.87E-02
GO:0009913	Epidermal cell differentiation	2.43E-06
GO:0030855	Epithelial cell differentiation	3.14E-05

GO: gene ontology; ID: identifier.

neutrophilic granulocytes, and is a glycoprotein that belongs to a family of cysteine-rich secretory proteins (CRISPs) [25]. In previous studies, *CRISP3* is found to be overexpressed in prostate adenocarcinoma by using quantitative real-time reverse-transcription-PCR [26]. Additionally, Su *et al.* reveal that *CRISP3* is significantly down-regulated in ESCC, and may be the biomarker of ESCC [27]. Furthermore, *CRISP3* was reported to be down-regulated in oral squamous cell carcinoma (OSCC), and the loss of its DNA copy number was observed in two of the five OSCC-derived cell lines [28].

EREG was the second highest ranking of 24 candidate genes whose protein product, EREG, induces cell growth by binding to the epidermal growth factor receptor (EGFR) [29]. It is reported that *EREG* is epigenetically silenced in gastric cancer cells by aberrant DNA methylation and histone modification [29]. Moreover, EREG is involved in the invasion and metastasis of esophageal carcinoma by combining with sphingosine kinase-1 (SPHK1) [30].

CXCR2 codes a receptor of ELR + CXC chemokines, which are potent promoters of angiogenesis [31]. It is reported that GROA-C XCR2 and GROB-C XCR2 signaling contribute significantly to esophageal cancer cell proliferation, and this autocrine signaling pathway may be involved in esophageal tumorigenesis [32,33].

CRNN codes cornulin, a Ca^{2+} - binding protein that presents in the upper layer of squamous epithelia [34]. It has been shown that the large majority of ESCC cases have little or no expression of cornulin in carcinoma or stromal cells [35]. These evidences suggested that *CRISP3*,

EREG, *CXCR2*, and *CRNN* may play crucial roles in ESCC, as well as other candidate genes.

In addition, GO functional enrichment analysis was performed, and some biological processes were enriched significantly, such as epidermal cell differentiation, epithelial cell differentiation, epidermis development, keratinocyte differentiation, and regulation of the immune response. It has been shown that proliferation and development of esophageal epithelial cells are associated with the development of ESCC [36]. Moreover, ESCC-related gene modules are significantly enriched in epidermal cell differentiation, epithelial cell differentiation, epidermis development, and keratinocyte differentiation [37]. Additionally, keratinocytes migrate from the basal to the superficial layers of the epidermis, and undergo morphological and biochemical changes during terminal differentiation, which are involved in the development of ESCC [38,39]. Our results were consistent with these evidences.

Conclusions

In conclusion, the DEGs between ESCC and adjacent normal tissues were screened out, and the co-expression network was constructed, consisting of 2 large sub-networks, 999 nodes, and 46,323 edges. After analyzing the gene expression and topological properties of DEGs in the co-expression network, DEGs were ranked, and 24 candidate genes of ESCC were identified. Candidate genes, such as *CRISP3*, *EREG*, *CXCR2*, and *CRNN*, were identified as potentially playing key roles in the development of ESCC. Furthermore, functional enrichment analysis revealed that the 24 genes were mainly enriched in epithelial cell differentiation, epidermis development, and keratinocyte differentiation. These results provided us with candidate genes and demonstrated their potential functions in the development of ESCC. However, more experimental studies are needed to confirm these results.

Additional file

Additional file 1: The co-expression network of differentially expressed genes.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

YS carried out the design and coordinated the study, participated in most of the experiments and prepared the manuscript. JT provided assistance in the design of the study, coordinated and carried out all the experiments and participated in manuscript preparation. HZ provided assistance for all experiments. All authors have read and approved the content of the manuscript.

Authors' information

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References

1. Lam AKY: **Molecular biology of esophageal squamous cell carcinoma.** *Crit Rev Oncol Hematol* 2000, **33**:71–90.
2. Hu N, Clifford R, Yang H, Wang C, Goldstein A, Ding T, Taylor P, Lee M: **Genome wide analysis of DNA copy number neutral loss of heterozygosity (CNNLOH) and its relation to gene expression in esophageal squamous cell carcinoma.** *BMC Genomics* 2010, **11**:576.
3. Wieder HA, Brücher BL, Zimmermann F, Becker K, Lordick F, Beer A, Schwaiger M, Fink U, Siewert JR, Stein HJ: **Time course of tumor metabolic activity during chemoradiotherapy of esophageal squamous cell carcinoma and response to treatment.** *J Clin Oncol* 2004, **22**:900–908.
4. Brücher BL, Weber W, Bauer M, Fink U, Avril N, Stein HJ, Werner M, Zimmermann F, Siewert JR, Schwaiger M: **Neoadjuvant therapy of esophageal squamous cell carcinoma: response evaluation by positron emission tomography.** *Ann Surg* 2001, **233**:300.
5. Stahl M, Stuschke M, Lehmann N, Meyer H-J, Walz MK, Seeber S, Klump B, Budach W, Teichmann R, Schmitt M: **Chemoradiation with and without surgery in patients with locally advanced squamous cell carcinoma of the esophagus.** *J Clin Oncol* 2005, **23**:2310–2317.
6. Tepper J, Krasna MJ, Niedzwiecki D, Hollis D, Reed CE, Goldberg R, Kiel K, Willett C, Sugarbaker D, Mayer R: **Phase III trial of trimodality therapy with cisplatin, fluorouracil, radiotherapy, and surgery compared with surgery alone for esophageal cancer: CALGB 9781.** *J Clin Oncol* 2008, **26**:1086–1092.
7. Pollack JR, Sørlie T, Perou CM, Rees CA, Jeffrey SS, Lonning PE, Tibshirani R, Botstein D, Børresen-Dale A-L, Brown PO: **Microarray analysis reveals a major direct role of DNA copy number alteration in the transcriptional program of human breast tumors.** *Proc Natl Acad Sci* 2002, **99**:12963–12968.
8. Mischel PS, Cloughesy TF, Nelson SF: **DNA-microarray analysis of brain cancer: molecular classification for therapy.** *Nat Rev Neurosci* 2004, **5**:782–792.
9. Risinger JI, Maxwell GL, Chandramouli G, Jazaeri A, Aprelikova O, Patterson T, Berchuck A, Barrett JC: **Microarray analysis reveals distinct gene expression profiles among different histologic types of endometrial cancer.** *Cancer Res* 2003, **63**:6–11.
10. Moch H, Schraml P, Bubendorf L, Mirlacher M, Kononen J, Gasser T, Mihatsch MJ, Kallioniemi OP, Sauter G: **High-throughput tissue microarray analysis to evaluate genes uncovered by cDNA microarray screening in renal cell carcinoma.** *Am J Pathol* 1999, **154**:981–986.
11. Hu N, Roth MJ, Emmert-Buck MR, Tang Z-Z, Polymeropoulos M, Wang Q-H, Goldstein AM, Han X-Y, Dawsey SM, Ding T, Giffen C, Taylor PR: **Allelic loss in esophageal squamous cell carcinoma patients with and without family history of upper gastrointestinal tract cancer.** *Clin Cancer Res* 1999, **5**:3476–3482.
12. Hu N, Wang C, Hu Y, Yang HH, Kong L-H, Lu N, Su H, Wang Q-H, Goldstein AM, Buetow KH: **Genome-wide loss of heterozygosity and copy number alteration in esophageal squamous cell carcinoma using the Affymetrix GeneChip mapping 10 K array.** *BMC Genomics* 2006, **7**:299.
13. Luo A, Kong J, Hu G, Liew C-C, Xiong M, Wang X, Ji J, Wang T, Zhi H, Wu M: **Discovery of Ca²⁺-relevant and differentiation-associated genes downregulated in esophageal squamous cell carcinoma using cDNA microarray.** *Oncogene* 2003, **23**:1291–1299.
14. Kimura S, Naganuma S, Susuki D, Hirano Y, Yamaguchi A, Fujieda S, Sano K, Itoh H: **Expression of microRNAs in squamous cell carcinoma of human head and neck and the esophagus: miR-205 and miR-21 are specific markers for HNSCC and ESCC.** *Oncol Rep* 2010, **23**:1625–1633.
15. Yamashita K, Upadhyay S, Osada M, Hoque MO, Xiao Y, Mori M, Sato F, Meltzer SJ, Sidransky D: **Pharmacologic unmasking of epigenetically silenced tumor suppressor genes in esophageal squamous cell carcinoma.** *Cancer Cell* 2002, **2**:485–495.
16. Irizarry RA, Hobbs B, Collin F, Beazer-Barclay YD, Antonellis KJ, Scherf U, Speed TP: **Exploration, normalization, and summaries of high density oligonucleotide array probe level data.** *Biostatistics* 2003, **4**:249–264.

17. Tusher VG, Tibshirani R, Chu G: **Significance analysis of microarrays applied to the ionizing radiation response.** *Proc Natl Acad Sci* 2001, **98**:5116–5121.
18. Deza M, Deza E: **Encyclopedia of Distances.** In *Encyclopedia of Distances.* Berlin Heidelberg: Springer; 2009:94.
19. Szekely GJ, Rizzo ML: **Hierarchical clustering via joint between-within distances: extending Ward's minimum variance method.** *J Classif* 2005, **22**:151–183.
20. Allocco DJ, Kohane IS, Butte AJ: **Quantifying the relationship between co-expression, co-regulation and gene function.** *BMC Bioinformatics* 2004, **5**:18.
21. Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B, Ideker T: **Cytoscape: a software environment for integrated models of biomolecular interaction networks.** *Genome Res* 2003, **13**:2498–2504.
22. Assenov Y, Ramirez F, Schelhorn SE, Lengauer T, Albrecht M: **Computing topological parameters of biological networks.** *Bioinformatics* 2008, **24**:282–284.
23. Quackenbush J: **Microarray data normalization and transformation.** *Nat Genet* 2002, **32**:496–501.
24. Sherman BT, Lempicki RA: **Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists.** *Nucleic Acids Res* 2009, **37**:1–13.
25. Udby L, Cowland JB, Johnsen AH, Sørensen OE, Borregaard N, Kjeldsen L: **An ELISA for SGP28/CRISP-3, a cysteine-rich secretory protein in human neutrophils, plasma, and exocrine secretions.** *J Immunol Methods* 2002, **263**:43–55.
26. Kosari F, Asmann YW, Cheville JC, Vasmataz G: **Cysteine-rich secretory protein-3: a potential biomarker for prostate cancer.** *Cancer Epidemiol Biomark Prev* 2002, **11**:1419–1426.
27. Su H, Hu N, Yang HH, Wang C, Takikita M, Wang Q-H, Giffen C, Clifford R, Hewitt SM, Shou J-Z, Goldstein AM, Lee MP, Taylor PR: **Global gene expression profiling and validation in esophageal squamous cell carcinoma and its association with clinical phenotypes.** *Clin Cancer Res* 2011, **17**:2955–2966.
28. Ko W-C, Sugahara K, Sakuma T, Yen C-Y, Liu S-Y, Liaw G-A, Shibahara T: **Copy number changes of CRISP3 in oral squamous cell carcinoma.** *Oncol Lett* 2012, **3**:75–81.
29. Yun J, Song S-H, Park J, Kim H-P, Yoon Y-K, Lee K-H, Han S-W, Oh D-Y, Im S-A, Bang Y-J: **Gene silencing of EREG mediated by DNA methylation and histone modification in human gastric cancers.** *Lab Invest* 2012, **92**:1033–1044.
30. Pan J, Tao Y-F, Zhou Z, Cao B, Wu S-Y, Zhang Y-L, Hu S-Y, Zhao W-L, Wang J, Lou G-L: **An novel role of sphingosine kinase-1 (SPHK1) in the invasion and metastasis of esophageal carcinoma.** *J Transl Med* 2011, **9**:157.
31. Belperio JA, Keane MP, Arenberg DA, Addison CL, Ehlert JE, Burdick MD, Strieter RM: **CXC chemokines in angiogenesis.** *J Leukoc Biol* 2000, **68**:1–8.
32. Wang B, Hendricks DT, Wamunyokoli F, Parker MI: **A growth-related oncogene/CXC chemokine receptor 2 autocrine loop contributes to cellular proliferation in esophageal cancer.** *Cancer Res* 2006, **66**:3071–3077.
33. Wang B, Khachigian LM, Esau L, Birrer MJ, Zhao X, Parker MI, Hendricks DT: **A key role for early growth response-1 and nuclear factor-κB in mediating and maintaining GRO/CXCR2 proliferative signaling in esophageal cancer.** *Mol Cancer Res* 2009, **7**:755–764.
34. Contzler R, Favre B, Huber M, Hohl D: **Cornulin, a new member of the 'fused gene' family, is expressed during epidermal differentiation.** *J Invest Dermatol* 2005, **124**:990–997.
35. Pawar H, Maharudraiah J, Kashyap MK, Sharma J, Srikanth SM, Choudhary R, Chavan S, Sathe G, Manju HC, Kumar KV, Vijayakumar M, Sirdeshmukh R, Harsha HC, Prasad TS, Pandey A, Kumar RV: **Downregulation of cornulin in esophageal squamous cell carcinoma.** *Acta Histochem* 2013, **115**:89–99.
36. Zhang H, Chen W, Duan C-J, Zhang C-F: **Overexpression of HSPA2 is correlated with poor prognosis in esophageal squamous cell carcinoma.** *World J Surg Oncol* 2013, **11**:141.
37. Gao H, Wang L, Cui S, Wang M: **Combination of meta-analysis and graph clustering to identify prognostic markers of ESCC.** *Genet Mol Biol* 2012, **35**:530–537.
38. Kadowaki Y, Fujiwara T, Fukazawa T, Shao J, Yasuda T, Itoshima T, Kagawa S, Hudson LG, Roth JA, Tanaka N: **Induction of differentiation-dependent apoptosis in human esophageal squamous cell carcinoma by adenovirus-mediated p21^{sd1} gene transfer.** *Clin Cancer Res* 1999, **5**:4233–4241.
39. Banks-Schlegel SP, Quintero J: **Growth and differentiation of human esophageal carcinoma cell lines.** *Cancer Res* 1986, **46**:250–258.

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