

Research Article

Oligonol Ameliorates CCl₄-Induced Liver Injury in Rats via the NF-Kappa B and MAPK Signaling Pathways

Jeonghyeon Bak,¹ Nam Kyung Je,¹ Hae Young Chung,¹ Takako Yokozawa,² Sik Yoon,³ and Jeon-Ok Moon¹

¹College of Pharmacy, Pusan National University, Busan 609-735, Republic of Korea

²Graduate School of Science and Engineering for Research, University of Toyama, Toyama 930-0194, Japan

³Department of Anatomy, College of Medicine, Pusan National University, Yangsan 626-870, Republic of Korea

Correspondence should be addressed to Jeon-Ok Moon; mjo@pusan.ac.kr

Received 24 July 2015; Accepted 27 September 2015

Academic Editor: Pablo Muriel

Copyright © 2016 Jeonghyeon Bak et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Oxidative stress is thought to be a key risk factor in the development of hepatic diseases. Blocking or retarding the reactions of oxidation and the inflammatory process by antioxidants could be a promising therapeutic intervention for prevention or treatment of liver injuries. Oligonol is a low molecular weight polyphenol containing catechin-type monomers and oligomers derived from lychee fruit. In this study, we investigated the anti-inflammatory effect of oligonol on carbon tetrachloride- (CCl₄-) induced acute hepatic injury in rats. Oral administration of oligonol (10 or 50 mg/kg) reduced CCl₄-induced abnormalities in liver histology and serum AST and serum ALT levels. Oligonol treatment attenuated the CCl₄-induced production of inflammatory mediators, including TNF- α , IL-1 β , cyclooxygenase-2 (COX-2), and inducible nitric oxide synthase (iNOS) mRNA levels. Western blot analysis showed that oligonol suppressed proinflammatory nuclear factor-kappa B (NF- κ B) p65 activation, phosphorylation of extracellular signal-regulated kinase (ERK), c-Jun NH₂-terminal kinase (JNK), and p38 mitogen-activated protein kinases (MAPKs) as well as Akt. Oligonol exhibited strong antioxidative activity *in vitro* and *in vivo*, and hepatoprotective activity against *t*-butyl hydroperoxide-induced HepG2 cells. Taken together, oligonol showed antioxidative and anti-inflammatory effects in CCl₄-intoxicated rats by inhibiting oxidative stress and NF- κ B activation via blockade of the activation of upstream kinases including MAPKs and Akt.

1. Introduction

Liver inflammation is a common response to various types of chronic liver injury. In the initial stages of inflammation, hepatocytes, Kupffer cells, platelets, and leukocytes are activated and produce reactive oxygen species (ROS) and inflammatory mediators such as platelet-derived growth factor, transforming growth factor- β (TGF- β), connective tissue growth factor, and tissue necrosis factor- α (TNF- α). These factors probably act as paracrine mediators to activate quiescent hepatic stellate cells (HSCs) that are localized in the perisinusoidal space, resulting in abnormal quantity and composition of extracellular matrix [1], which in turn leads to hepatitis, liver fibrosis, and cirrhosis. Thus, it is important to suppress hepatic inflammation in the early stages of liver fibrosis.

Oxidative stress, in particular, lipid peroxidation, has been considered one of the major causes of liver damage and has been reported to be associated with HSC activation [2]. Lipid peroxidation may provoke liver damage by compromising the integrity of membranes and by inducing covalent binding of reactive intermediates to important antioxidants such as glutathione [3]. Antioxidants are potent free radical scavengers and have been documented to protect hepatocytes from lipid peroxidation in the carbon tetrachloride- (CCl₄-) or dimethylnitrosamine- (DMN-) induced liver injury models [4, 5]. Therefore, blocking or retarding the reactions of lipid peroxidation and the inflammatory process by antioxidants could be a promising therapeutic intervention for prevention or treatment of liver injuries.

Dietary phytochemicals of fruits, vegetables, whole grains, and other plant foods were shown to have potent

antioxidant activity, and the mixture or combination of phytochemicals was proposed to be responsible for their strong antioxidant activity [6]. In most cases, the poor absorption rate of polyphenolic substances limits their use as dietary supplements in human. Particularly, many polyphenols exist in polymeric forms of high molecular weight that may further decrease their bioavailability [7]. Oligonol is a phenolic product derived from lychee fruit (*Litchi chinensis* Sonn.) extract by a manufacturing process that converts polyphenol polymers into oligomers. Oligonol comprises 16.0% monomers (catechin, epicatechin, epicatechin gallate, and epigallocatechin gallate), 13.9% dimers (procyanidin A1, A2, B1, and B2), and oligomers of proanthocyanidins [8]. Oligonol delivers higher levels of oligomeric proanthocyanidins compared with fruit and plant sources that contain high molecular weight proanthocyanidins [7].

There is a growing evidence that oligonol can elicit some physiological and biochemical alterations *in vitro* and *in vivo*, such as inhibition of adipogenesis in 3T3-L1 adipocytes [9], improvement of memory and cognition under an amyloid beta-induced Alzheimer's mouse model [10], the induction of apoptosis in MCF-7 and MDA-MB-231 human breast cancer cell lines [11], and antioxidant and anti-inflammatory effects in ultraviolet B- (UVB-) irradiated mouse skin [12]. However, the potential protective activity and mechanism of oligonol on acute liver inflammation induced CCl_4 have not yet been conducted.

Here, we report that oral administration of oligonol exerts an antioxidant, anti-inflammatory, and hepatoprotective effect in CCl_4 -induced acute liver injury in rats. The possible molecular mechanism of action of oligonol was explored by analyzing the expression of nuclear factor-kappa B (NF- κ B), TNF- α , IL-1 β , cyclooxygenase-2 (COX-2), inducible nitric oxide synthase (iNOS), Akt, and mitogen-activated protein kinases (MAPKs), including extracellular signal-regulated kinase (ERK), c-Jun NH₂-terminal kinase (JNK), and p38.

2. Materials and Methods

2.1. Chemicals. Bicinchoninic acid (BCA) solution, bovine serum albumin (BSA), *t*-butyl hydroperoxide (*t*-BHP), butylated hydroxytoluene (BHT), CCl_4 , 1,1-diphenyl-2-picrylhydrazyl (DPPH), formalin, sodium carboxymethylcellulose (CMC), 2',7'-dichlorofluorescein diacetate (DCFDA), phosphoric acid, trichloroacetic acid (TCA), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromides (MTT), and Trolox were obtained from Sigma-Aldrich Co. 2-Thiobarbituric acid (TBA) was obtained from Tokyo Chemical Industry Co. Malondialdehyde (MDA) tetrabutylammonium salt was obtained from Fluka. Oligonol is commercially available (Amino Up Chemical Co., Ltd., Sapporo, Japan).

2.2. DPPH Assay. The scavenging activity of oligonol was measured using the stable DPPH free radical, according to a published method [13] with slight modifications. The reaction mixture contained 50 mM phosphate buffer at pH 7.4 (80 μL), 100 μM DPPH dissolved in ethanol (100 μL), and

the indicated concentrations of oligonol and Trolox (20 μL). Triplicate reaction tubes were wrapped in aluminum foil and placed at room temperature for 30 min in the dark. Spectrophotometric readings were taken at 517 nm using a microplate reader (Apollo-LB913, Berthold Technologies). The percent inhibition of free radical production was calculated from $([A_0 - (A - A_b)]/A_0) \times 100$, where A_0 is the absorbance of the control, A is the absorbance of the sample, and A_b is the absorbance of the blank sample (containing all reagents except DPPH). IC_{50} values were obtained from the inhibition curves.

2.3. Antioxidative Activity against Lipid Peroxidation Induced $\text{FeSO}_4/\text{H}_2\text{O}_2$ in Rat Liver Homogenates. Lipid peroxidation in rat liver homogenates induced by the Fenton reaction, comprising 0.1 mM FeSO_4 , 3 mM H_2O_2 , various concentrations of the tested substances, and liver homogenates (7.5 mg protein/mL), was measured by the method of Buege and Aust [14] with some modifications. The reaction was started by the addition of FeSO_4 and H_2O_2 and then incubated at 37°C for 10 min. The reaction was stopped by mixing with 3 mL of a stock solution of 15% (w/v) TCA, 0.375% (w/v) TBA, 0.125 M hydrochloric acid, and 0.6 mM BHT. The combination of reaction mixture and stock solution was heated for 30 min in a boiling water bath. After cooling, the flocculent precipitate was removed by centrifugation at 1,250 g for 20 min. The absorbance of the supernatant was determined at 532 nm, and the MDA concentration was calculated using MDA tetrabutylammonium salt as a standard. Protein concentrations were determined by the BCA assay using BSA as the reference standard.

2.4. Protective Effect of Oligonol on Cell Damage Induced by *t*-BHP. The human hepatocellular carcinoma cell line HepG2 was purchased from the Korean Cell Line Bank. The cells were cultured in a complete medium composed of Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS), 1% glutamine, penicillin (100 $\mu\text{g}/\text{mL}$), and streptomycin (100 $\mu\text{g}/\text{mL}$) at 37°C in a 5% CO_2 humidified incubated environment. The cells were placed in 96-well plates at a density of 2.0×10^4 cells per well. After 24 h cultivation, the complete medium of the plates was replaced with serum-free medium, and various concentrations of oligonol (0.5, 2, 5, and 10 $\mu\text{g}/\text{mL}$) were added to the cells. Four hours later, the cells were exposed to 300 μM *t*-BHP for 3 h. After incubation, 100 μL of MTT solution (1 mg/mL in phosphate buffered saline) was added to each well and incubated for another 2 h. After the culture medium had been removed, 100 μL of DMSO was added and mixed to dissolve the MTT formazan crystals. The plates were read on a microplate reader (Apollo-LB913, Berthold Technologies) using a wavelength of 540 nm. The survival values, used to examine the protective effects of the compounds against cell damage by *t*-BHP, were expressed as a percentage of the absorbance of the normal cells.

2.5. Cell Lysis. Cells were washed by phosphate buffered saline (PBS), and then 1 mL of ice-cold PBS was added.

Pellets were harvested at 1,000 g at 4°C for 3 min. The pellets were suspended in ProEX CETi Lysis Buffer (TransLab), incubated on ice for 20 min and then centrifuge at 14,000 g at 4°C for 10 min. The supernatants were used as total protein extraction.

2.6. Animals. Male Sprague-Dawley rats were obtained from Samtako (Osan, Korea). Animals were provided standard rat chow with free access to tap water and were maintained at a controlled temperature ($23 \pm 3^\circ\text{C}$) and humidity ($50 \pm 20\%$) with a 12 h light-dark cycle. With respect to ethical issues and scientific care, the animal protocol used in this study was reviewed and approved by the Pusan National University-Institutional Animal Care and Use Committee (PNU-IACUC; Approval number PNU 2008-0541).

2.7. Induction of Acute Hepatic Inflammation with CCl_4 . Twenty-four rats weighing 140–160 g and 5–6 weeks in age were assigned to 4 groups ($n = 6$): control, CCl_4 , Oli10, and Oli50. Animals in the control group received olive oil (CCl_4 vehicle) by intraperitoneal (i.p.) injection and CMC (oligonol vehicle) by oral gavage; the CCl_4 group received CCl_4 and CMC, while the Oli10 and Oli50 group received CCl_4 and oligonol at 10 and 50 mg/kg/day, respectively. Liver injury was induced by a single i.p. injection of 25% (w/v) CCl_4 (0.6 g/kg body weight) in olive oil. Oligonol was suspended in 0.5% CMC solution to a concentration of 10 and 50 mg/mL and administered by oral gavage twice, once at 16 h and once at 30 min before CCl_4 intoxication. Twenty-four hours after the CCl_4 injection, all rats were euthanized by ether anesthesia, and the livers were excised and weighed. Blood samples for biochemical analyzes were obtained from the inferior vena cava.

2.8. Liver Homogenate Preparation. The remaining liver tissue was rapidly cut into small pieces and homogenized with two volumes (w/v) of ice-cold potassium phosphate buffer (pH 7.4) using an IKA T10 basic Ultra-Tur Rax homogenizer. Debris and nuclei were removed from the homogenate by centrifugation at $700 \times g$ at 4°C for 10 min and stored at -80°C for further analysis.

2.9. Histology. Liver specimens were fixed by immersion in 10% neutral buffered formaldehyde solution (NBF) for 24 h and then washed overnight. The samples from each group ($n = 6$) were dehydrated in a graded series of ethanol solutions, cleared in xylene, and embedded in paraffin. Eight to ten tissue sections (6 μm thick) were cut and stained with hematoxylin and eosin (H&E) to assess the architectural alterations. The degree of liver damage was evaluated semiquantitatively using the Ishak system under a light microscope [15].

2.10. Biochemical Analysis of Liver Enzymes. Serum aspartate transaminase (AST) and alanine transaminase (ALT) activities were measured using the method described by Reitman and Frankel [16], using AST/ALT kits (Asan Chemical Co.).

2.11. Determination of MDA Content. MDA levels in the liver tissue were measured by a published method with modifications [17]. Standards were prepared via serial dilution of a stock solution of 10 μM MDA tetrabutylammonium salt in distilled water. For the assay, 10 mg of liver tissue was homogenized with 1 mL of solution containing 26 mM TBA, 0.64 mM BHT, 0.93 M TCA, and 11 mM hydrochloric acid. The homogenates were heated for 1 h in a boiling water bath. After cooling, tubes were centrifuged for 15 min at $2,000 \times g$. The absorbance of supernatant was determined at 532 nm and the MDA concentration was calculated using MDA tetrabutylammonium salt as a standard.

2.12. Measurement of ROS Level. A fluorometric assay was used to determine levels of ROS, such as $\cdot\text{O}_2^-$, $\cdot\text{OH}$, and H_2O_2 . Nonfluorescent DCFDA was oxidized to the highly fluorescent 2',7'-dichlorofluorescein (DCF) in the presence of esterases and ROS, including lipid peroxides [18]. For the assay, 50 μM DCFDA was added to liver homogenates for 250 μL of final volume. Changes in fluorescence intensity were measured every 5 min for 30 min on a fluorescence plate reader, GENios (Tecan Instrument, Salzburg, Austria), with excitation and emission wavelengths set at 485 and 530 nm, respectively.

2.13. RNA Extraction and Reverse Transcriptase-Polymerase Chain Reaction (RT-PCR). Total RNA was extracted from samples of 100 mg of frozen liver by homogenization in Trizol reagent (Invitrogen). RNA purity was assessed by the absorbance ratio at 260 nm and 280 nm. cDNA was prepared from samples of 1 μg of RNA with iScript cDNA Synthesis Kit (Bio-Rad, Hercules, CA, USA) according to the protocol provided by the manufacturer. PCR was performed in 20 μL of reaction solution containing 1 μg of cDNA and the appropriate primers from Bioneer (Daejeon, Korea) (Table 1) using a Promega GoTaq Flexi DNA Polymerase PCR kit. PCR conditions were as follows: denaturation at 95°C for 10 min, 35 cycles of 30 s at 95°C , 90 s at 60°C , 60 s at 72°C , and a final extension at 72°C for 5 min. GAPDH was measured as an internal control for normalization of mRNA levels. The amplified products were analyzed by 1.5% agarose gel electrophoresis and visualized by ethidium bromide staining under UV light illumination (Gel Doc/ChemiDoc imager, Azure). All reactions were performed in triplicate.

2.14. Western Blotting Analysis. Nuclear extracts, cytosol extracts, or total proteins of liver tissue were prepared as a published method with modification [19]. The protein concentration was measured by the BCA assay. Aliquots of protein (30 μg) were denatured at 95°C for 5 min before electrophoresis on 10% SDS-polyacrylamide gel. After transfer to a polyvinylidene difluoride (PVDF) membrane (Millipore), the blot was blocked with 5% nonfat milk solution for 1 h at room temperature and then incubated with a 1 : 1,000 dilution of primary antibodies selective against either NF- κB p65, total JNK, p-JNK, total ERK, p-ERK, total Akt, p-Akt, total p38, p-p38, histone, or β -actin (Santa Cruz Biotechnology) in Tris-buffered saline Tween-20 (TBST) at 4°C overnight,

TABLE 1: Oligonucleotide sequences used in RT-PCR analysis.

Group	Direction	Sequence
TNF- α	Forward	TTCTGTCTACTGAACTTGGGGGTGATCGGTCC
	Reverse	GTATGAGATAGCAAATCGGCTGACGGTGTGGG
IL-1 β	Forward	ATGGCAACTGTTTCCTGAACTCAACT
	Reverse	CAGGACAGGTATAGATTCTTTTCCTTT
COX-2	Forward	CCAGAGCAGAGAGATGAAATACCA
	Reverse	GCAGGGCGGGATACAGTTC
iNOS	Forward	GATTCAGTGGTCCAACCTGCA
	Reverse	CGACCTGATGTTGCCACTGTT
GAPDH	Forward	GACAACCTTTGGCATCGTGGA
	Reverse	ATGCAGGGATGATGTTCTGG

TNF- α : tumor necrosis factor-alpha; IL-1 β : interleukin-1 beta; COX-2: cyclooxygenase-2; iNOS: inducible nitric oxide synthase; GAPDH: glyceraldehyde-3-phosphate dehydrogenase.

TABLE 2: Antioxidative activities of oligonol against DPPH radical.

Substances	Concentration	Inhibition (%)	IC ₅₀
Oligonol ($\mu\text{g/mL}$)	5	10.7 \pm 0.002	50.5
	10	28.5 \pm 0.001	
	25	38.9 \pm 0.001	
	50	49.9 \pm 0.001	
	100	61.1 \pm 0.006	
Trolox (μM)	5	19.2 \pm 0.008	18.6
	10	26.1 \pm 0.003	
	25	50.4 \pm 0.007	
	50	65.1 \pm 0.001	
	100	71.3 \pm 0.006	

The reaction mixture consisted of 0.5 mL of 60 μM ethanolic solution of DPPH and 0.5 mL of various concentrations of sample solution. After allowing the mixture to stand at room temperature for 30 min, the absorbance of the remaining DPPH was determined at 517 nm. Trolox was used as a positive control.

followed by 1 h at room temperature. The membrane was washed 3 times for 5 min each with TBST solution. The membranes were incubated with 1:10,000 dilution of horseradish peroxidase-conjugated rabbit or mouse secondary antibodies (Santa Cruz Biotechnology) at room temperature for 1 h. The transferred proteins were visualized with an enhanced chemiluminescence (ECL) detection system and the band intensities were determined using a Gel Doc/Chemidoc imager (Azure). The protein concentration was determined with a BCA protein assay kit (Pierce, Rockford, IL, USA).

2.15. Statistical Analyses. All results are expressed as the mean \pm SE of the indicated number of replicates. Data were analyzed for statistical differences by one-way analysis of variance (ANOVA). A *p* value of 0.05 or less was considered statistically significant.

3. Results

3.1. Antioxidative Activities of Oligonol against the Lipid Peroxidation of Rat Liver Homogenates Induced by FeSO₄ and H₂O₂ and against DPPH Radical. The antioxidative activities

TABLE 3: Antioxidative activities of oligonol against the lipid peroxidation of rat liver homogenates induced by FeSO₄ and H₂O₂.

Substances	Concentration	Inhibition (%)	IC ₅₀
Oligonol ($\mu\text{g/mL}$)	10	47.52 \pm 0.06	15.16
	50	62.01 \pm 0.01	
	100	74.74 \pm 0.06	
	500	93.02 \pm 0.01	
	5	16.51 \pm 0.04	
BHT (μM)	10	43.43 \pm 0.03	15.01
	20	59.82 \pm 0.01	
	50	89.61 \pm 0.05	
	50	89.61 \pm 0.05	

Results are expressed as mean \pm SE (*n* = 3). The reaction mixture was composed of the rat liver homogenate, 0.1 mM FeSO₄, 3 mM H₂O₂, and various concentrations of oligonol or BHT. After incubation at 37°C for 10 min, the amount of MDA formation was measured by the method of Buege and Aust. Inhibition (%) of MDA formation in oligonol or BHT was calculated based on the amount of MDA formation of the FeSO₄/H₂O₂-treated control after subtracting the normal.

of oligonol were investigated by the examination of the inhibitory effect against FeSO₄/H₂O₂-induced lipid peroxidation in rat liver homogenates (Table 2) and the DPPH radical scavenging effect (Table 3). As positive control for the inhibition of lipid peroxidation, a well-known antioxidant BHT was tested. Under the reaction condition which allows the IC₅₀ of BHT to be 15.01 μM , IC₅₀ of oligonol was 15.15 $\mu\text{g/mL}$. Oligonol showed DPPH free radical scavenging activity (IC₅₀ = 50.5 μg) and Trolox was tested as a positive control (IC₅₀ = 18.6 μM)

3.2. Hepatoprotective Effect of Oligonol against Cell Damage Induced by *t*-BHP. *t*-BHP is a cytotoxic agent that is metabolized to free radicals including *t*-butoxyl, *t*-butylperoxyl, or methyl radical that interfere with cellular functions. The protective effect of oligonol on cell damage induced by *t*-BHP was examined (Figure 1). By exposing the cells to 300 μM *t*-BHP for 3 h, cell viability decreased to 46%. However, oligonol was found to protect *t*-BHP-induced cell damage dose-dependently, and the EC₅₀ was calculated to be 0.25 μg . In addition, we investigated the effect of oligonol

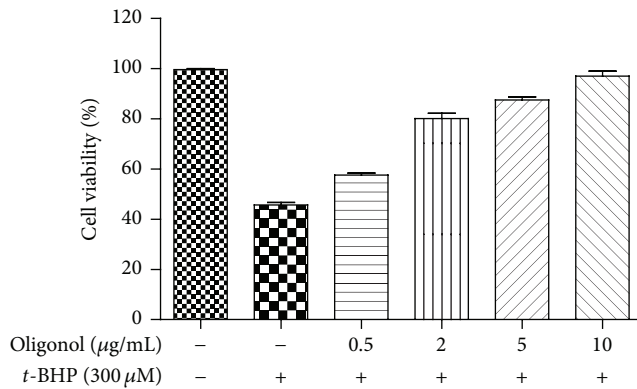


FIGURE 1: Effect of oligonol on HepG2 cell damage induced by *t*-BHP. Cell viability was assessed using MTT assays. Data shown represent means \pm standard deviation of triplicate experiments.

on the induction of proinflammatory mediator COX-2 in *t*-BHP-treated HepG2 cells at the protein level by western blot analysis. As shown in Figure 2, oligonol suppressed the *t*-BHP-induced COX-2 induction.

3.3. Changes in Body and Liver Weight and Serum Parameters in Rats Intoxicated by CCl₄. Treatment with CCl₄ caused a slight increase in the ratio of body weight/liver weight when compared with the control group (Table 4). In contrast, animals injected with CCl₄ and orally administered with oligonol showed significantly reduced ratios of liver weight to body weight, compared to untreated animals injected with CCl₄. These results indicate that oligonol reduces the ratios of body to liver weight induced by CCl₄ intoxication. Biochemical analyses of serum AST and serum ALT activities were performed to determine whether oligonol protected the liver from CCl₄-induced injury (Figures 3(a) and 3(b)). Serum AST and serum ALT levels were significantly higher in rats injected with CCl₄ (475.1 ± 330.3 and 160.0 ± 120.6 U/L, resp.) than in the control rats (35.5 ± 5.2 and 28.4 ± 4.3 U/L, resp.). Serum AST and serum ALT activities were significantly reduced by oral administration of oligonol at both 10 and 50 mg/kg doses; levels of AST and ALT in the Oli10 group were 193.7 ± 61.7 and 72.9 ± 26.0 U/L, respectively, and levels in the Oli50 group were 121.1 ± 29.4 and 44.2 ± 17.6 U/L, respectively.

3.4. Prevention of ROS Production and Lipid Peroxidation by Oligonol. To assess the overall oxidative status, total ROS was measured with DCFDA probe in the liver homogenates. Results show that increased ROS levels with CCl₄ intoxication were suppressed by the administration of oligonol (Figure 4(a)). Induction of lipid peroxidation by CCl₄ was measured by the production of MDA in liver tissues (Figure 4(b)). The MDA content in the livers of CCl₄-treated rats was significantly higher than that in the control animals but significantly reduced in the livers of rats treated with oligonol in a dose-dependent manner, which is consistent with the results of ROS production and the liver function tests.

TABLE 4: Effects of oligonol on body and liver weights of rats treated with CCl₄.

	Body weight (g)	Liver weight (g)	Ratio (%) ^a
Control	154.02 \pm 7.40	5.96 \pm 1.25	3.85
CCl ₄	158.01 \pm 7.29	7.69 \pm 0.66**	4.87
Oli10	158.16 \pm 4.62	6.98 \pm 0.34 [#]	4.42
Oli50	156.75 \pm 10.05	6.71 \pm 0.29 [#]	4.29

CCl₄: CCl₄ alone treated group; Oli10: oligonol (10 mg/kg) with CCl₄; Oli50: oligonol (50 mg/kg) with CCl₄. ^aValues are expressed as the ratios of liver weight to body weight. Data are the mean \pm SE of $n = 6$ rats/group. ** $p < 0.01$ compared with the control group and [#] $p < 0.05$ compared with the CCl₄ group.

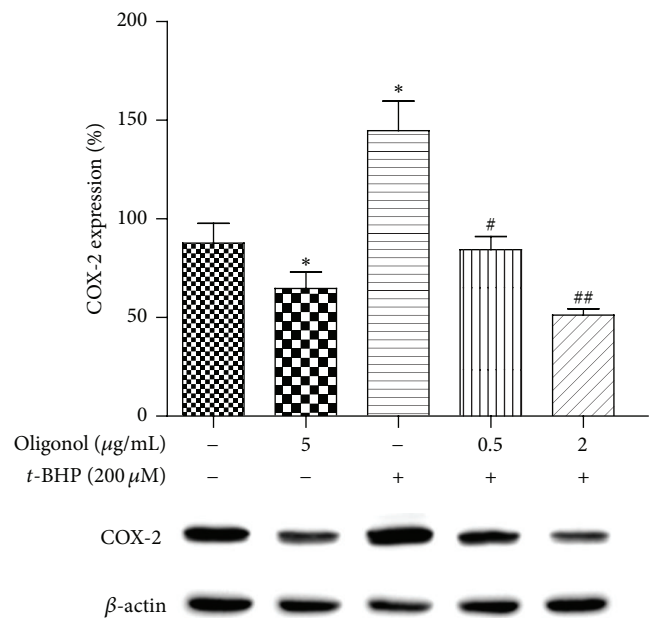


FIGURE 2: Effect of oligonol on *t*-BHP-induced COX-2 expression on HepG2 cells. HepG2 cells were treated with different concentrations of oligonol (0, 0.5, and 2 µg/mL) for 4 h before being exposed to *t*-BHP (200 µM) for 24 h. Western blotting was performed to detect COX-2 in whole protein (30 µg) from HepG2 cell. One representative blot of each protein is shown from three experiments that yielded similar result, respectively. Values are normalized as percentage of β -actin. Values are mean \pm SE of $n = 3$. * $p < 0.05$ compared with the control group, and [#] $p < 0.05$ and ^{##} $p < 0.01$ compared with the *t*-BHP group.

3.5. Liver Histopathology. The effect of oligonol on CCl₄-induced histopathological changes in the liver was evaluated on H&E stained liver sections (Table 5). Livers of the control group showed normal lobular architecture with central veins and radiating hepatic cords. No histological abnormalities were observed (Figure 5(a)). In contrast, the liver sections from CCl₄-treated animals showed distorted tissue architecture, submassive necrosis, vacuolization, and macrovesicular fatty changes of hepatocytes (Figure 5(b)). Notably, these pathologic changes were markedly reduced dose-dependently in the livers of animals treated with oligonol (Figures 5(c) and 5(d)).

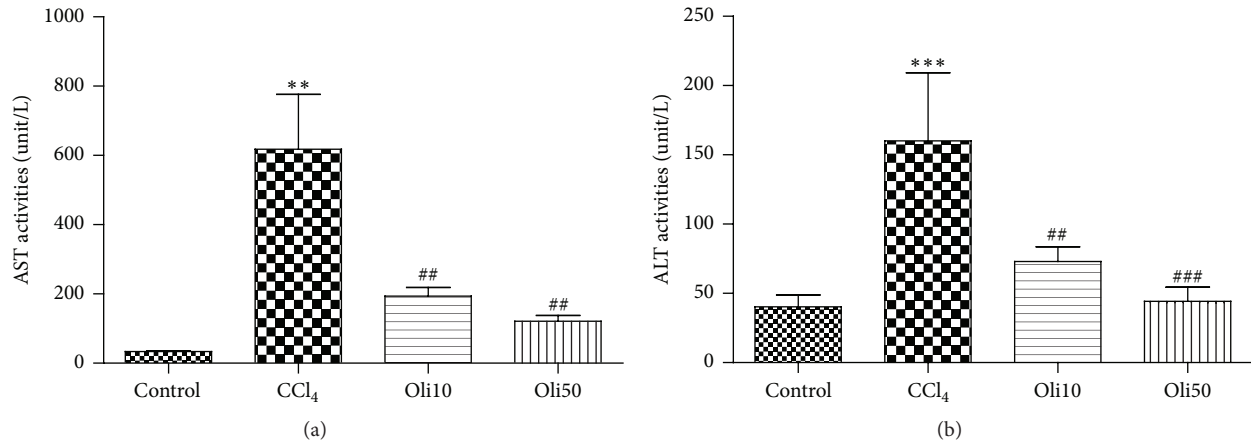


FIGURE 3: Measurements of serum ALT and serum AST levels. Groups are as described in “Methods.” Values are mean \pm SE of $n = 6$ rats/group. ** $p < 0.01$ and *** $p < 0.001$ compared with the control group and ## $p < 0.01$ and ### $p < 0.001$ compared with the CCl₄ group.

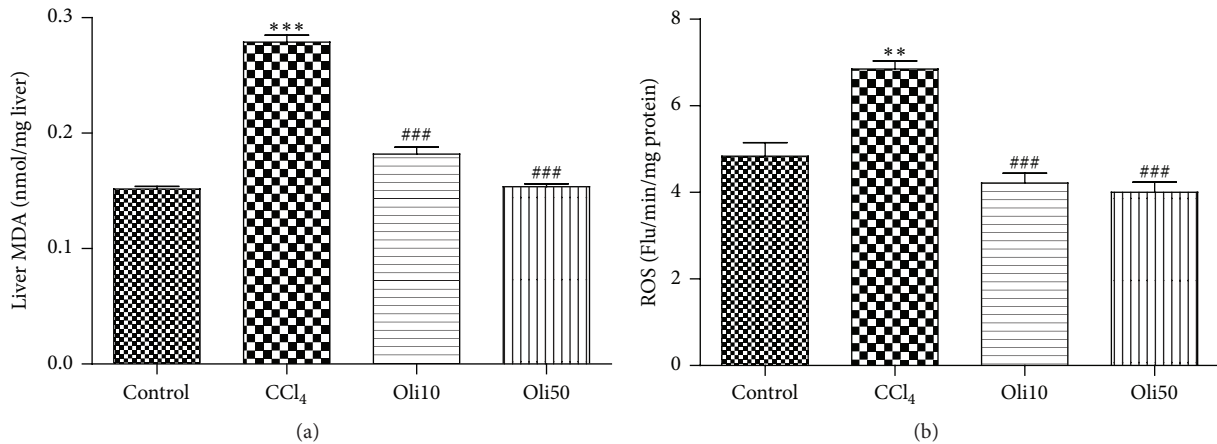


FIGURE 4: Effect of oligonol on ROS and MDA levels in rat liver intoxicated with CCl₄. ROS generation was measured by DCF formation with a fluorescent probe, DCFDA. Effect of oligonol on CCl₄-induced lipid peroxidation activity from the liver was measured by the method of Buege and Aust. Values are mean \pm SE of $n = 6$ rats/group. ** $p < 0.01$ and *** $p < 0.001$ compared with the control group. ### $p < 0.001$ compared with the CCl₄ group.

TABLE 5: Effects of oligonol on the histopathological score of liver of CCl₄-treated rats.

Group	Number	Histopathological score of liver				
		0	1	2	3	4
Control	6	6	0	0	0	0
CCl ₄	6	0	0	0	1	5
Oli10	6	1	2	1	2	0
Oli50	6	3	1	1	1	0

3.6. *Expression of TNF- α , IL-1 β , COX-2, and iNOS mRNA.* Expression of mRNA of proinflammatory cytokines TNF- α and IL-1 β and proinflammatory proteins COX-2 and iNOS in the liver was measured by RT-PCR. Following agarose gel electrophoresis of reaction products, mRNA levels were quantified by normalization against the expression of the housekeeping gene, GAPDH. CCl₄ treatment increased the

expression of TNF- α , IL-1 β , COX-2, and iNOS mRNA in the liver, but these were dose-dependently and significantly reduced by pretreatment of rats with oligonol (Figures 6 and 7).

3.7. *NF- κ B Translocation to the Nucleus.* The protein level of transcription factor NF- κ B was examined. Activation of NF- κ B was based on the detection of its translocation into cell nuclei from its initial location in the cytoplasm where it exists in an inactive form. Western blotting of NF- κ B p65 protein in nuclear and cytosolic fractions of the liver tissues indicates that CCl₄ treatment exhibited an enhancement of nuclear NF- κ B and a reduction of cytosolic NF- κ B (Figure 8). Oligonol treatment to CCl₄-intoxicated rat markedly inhibited CCl₄-induced increase of NF- κ B in the nuclear fraction of the liver. It also abrogated the reduction of NF- κ B in the cytosolic fraction. The relative level of NF- κ B p65 in the nuclear and cytosol is compared with β -actin and histon, respectively, and quantified by image analysis.

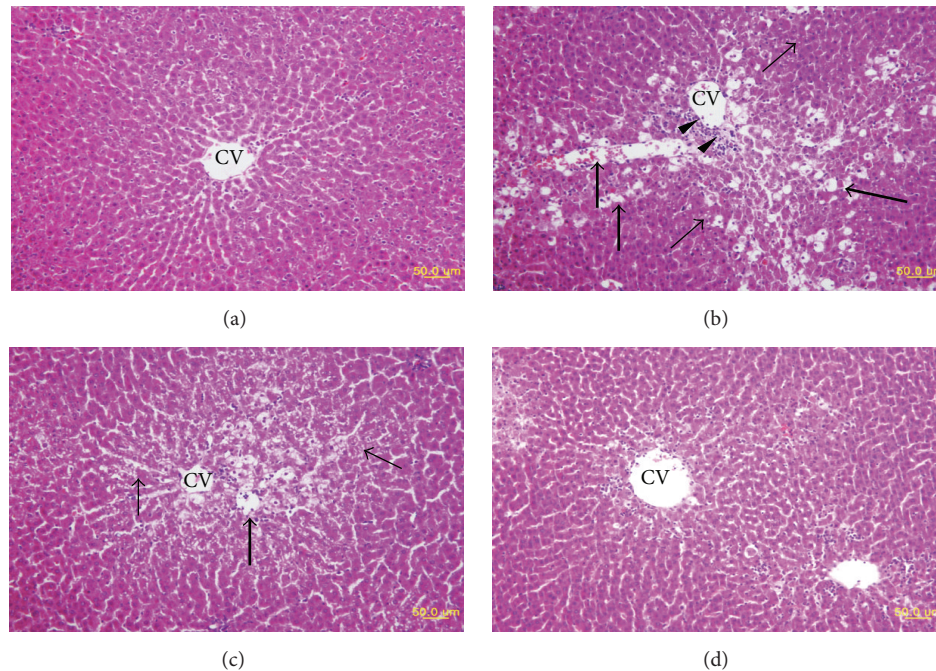


FIGURE 5: Effects of oligonol on CCl_4 -induced histopathological changes in rat livers. Representative H&E stained sections of livers of rats treated as described in methods. (a) Liver section of control group shows normal liver architecture, intact hepatocytes, and radiating hepatic cords from the central vein (CV); (b) CCl_4 -induced damage is indicated by distortion of the tissue architecture, submassive necrosis (thick arrows), fatty changes (thin arrows) in hepatocytes, and aggregations of inflammatory cells (arrowheads); (c) oligonol (10 mg/kg) plus CCl_4 ; and (d) oligonol (50 mg/kg) plus CCl_4 ; oligonol treatment reduced the pathological alterations induced by CCl_4 . All images are original magnification $\times 400$.

It showed that CCl_4 -treated rats had significantly increased expression of NF- κ B p65 compared with the control animals. Moreover, the expression of NF- κ B p65 in the oligonol-treated rats was reduced significantly in a dose-dependent manner. These results demonstrate that oligonol treatment in CCl_4 -intoxicated rats strongly inhibited the translocation of NF- κ B p65 from the cytosol to the nuclear fraction.

3.8. MAPKs and Akt Signaling Pathways Involved in NF- κ B Activation. To investigate the molecular mechanism of NF- κ B activation in the CCl_4 -intoxicated rat, we measured the expression levels of ERK1/2, JNK, and p38 MAPKs as well as Akt by using western blot analysis. The phosphorylation of MAPKs and Akt was increased in rats treated with CCl_4 alone as compared with the control group. However, treatment with low (10 mg/kg/day) and high (50 mg/kg/day) dose of oligonol in CCl_4 -intoxicated rats significantly decreased the expression levels of phosphorylated ERK1/2, JNK, and p38 MAPKs as well as Akt, in a dose-dependent manner (Figure 9).

4. Discussion

Oxidative stress, caused by the increased production of reactive oxygen species (ROS), is thought to be a key risk factor in the development of liver disease [20]. CCl_4 intoxication has been widely used as an experimental model of liver injury. CCl_4 is a substrate for cytochrome P450 2E1 (CYP2E1). It is converted to a CCl_3 radical, which generates CCl_3OO^*

by reacting with molecular oxygen. Since CCl_3OO^* reacts with microsomal membranes and induces lipid peroxidation, membrane damage by free radical chain reaction has been postulated to be the primary cause of hepatocellular injury by this compound [21]. In CCl_4 -induced injury, antioxidants are widely known to be able to protect against hepatocyte necrosis because they intercept the CCl_4 -induced oxidative stress in hepatocytes by scavenging $^*\text{CCl}_3$ and lipid peroxy radicals [22].

In this study, a single dose of CCl_4 induced distorted tissue architecture, submassive centrilobular necrosis, fatty changes, and inflammatory cell infiltration. However, these pathologic changes were significantly reduced by pretreatment of 10 and 50 mg/kg of oligonol dose-dependently. Treatment of oligonol effectively improved the CCl_4 -induced elevation in serum AST and serum ALT levels, indicating the hepatoprotective effects of oligonol against CCl_4 intoxication. CCl_4 treatment caused high levels of liver oxidative damage, as evidenced by a significant elevation of ROS production and MDA concentration in liver homogenates. However, oligonol markedly inhibits CCl_4 -induced oxidative stress in liver of rats in a dose-dependent manner.

Antioxidant activity of oligonol is well established in many studies [23], and the activities were consistent with the results that oligonol exerts the inhibitory effect against $\text{FeSO}_4/\text{H}_2\text{O}_2$ -induced lipid peroxidation in rat liver homogenates and against the DPPH radical scavenging effects. In addition, the protective effect of oligonol via antioxidative

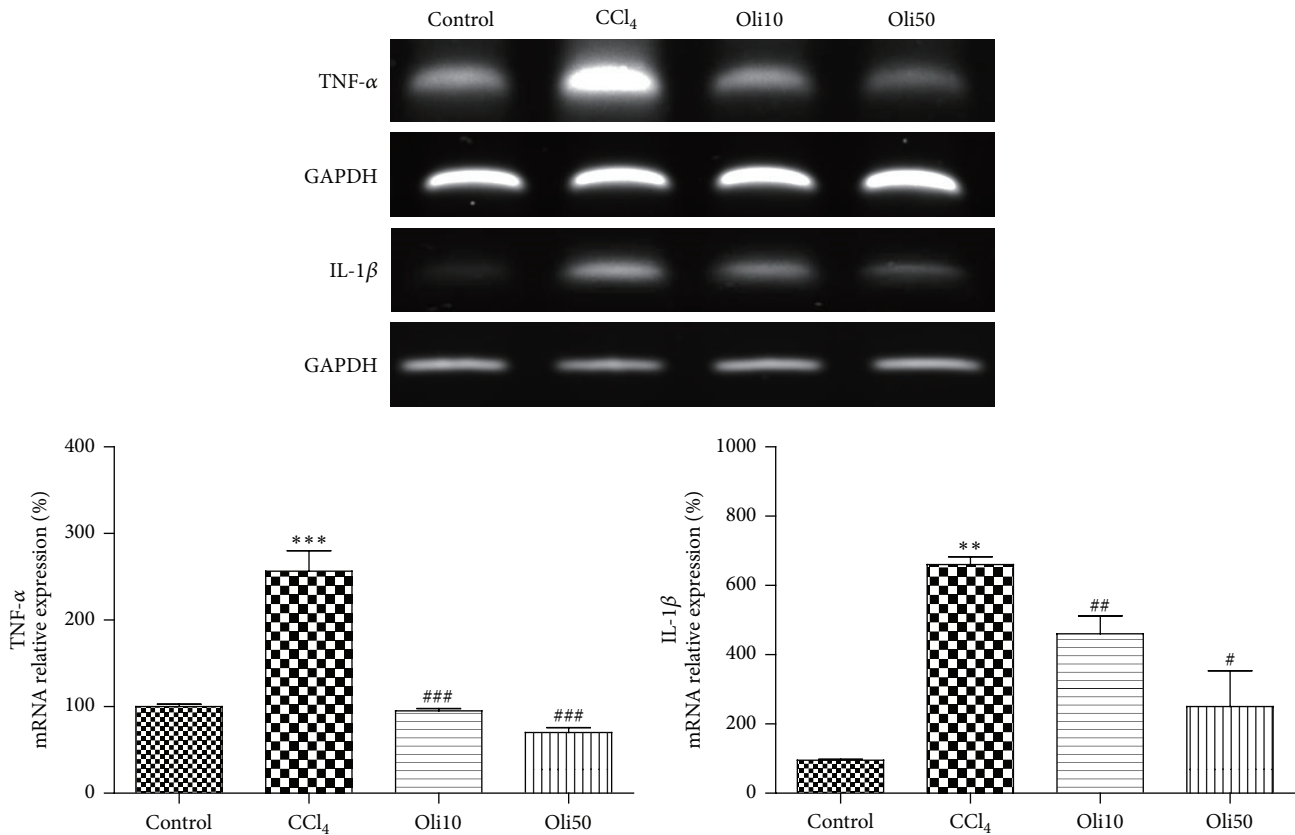


FIGURE 6: Effects of oligonol on TNF- α and IL-1 β mRNA expression after CCl₄ administration. RT-PCR was performed to measure TNF- α and IL-1 β mRNA expression in the liver tissues. Values are mean \pm SE of $n = 6$. ** $p < 0.01$ and *** $p < 0.001$ compared with the control group and ## $p < 0.01$ and ### $p < 0.001$ compared with the CCl₄ group.

activity was assessed in *t*-BHP-induced HepG2 cell damage. HepG2 cells are considered a reasonable model for studying *in vitro* xenobiotic metabolism and liver toxicity since they maintain a majority of specialized functions similar to normal human hepatocytes [24]. The oxidant *t*-BHP is well known to induce oxidative stress [25]. Oligonol showed hepatoprotective effects against *t*-BHP-induced oxidative stress. Taken together, hepatoprotective effects of oligonol in CCl₄-intoxicated rat model and in *t*-BHP-induced HepG2 cells may be due to the potent antioxidant and free radical scavenging activities of oligonol.

Proinflammatory cytokines such as TNF- α , IL-1 β , and IL-6 have been the focus of investigations of inflammatory organ injury because the uncontrolled and prolonged action of these proteins is potentially harmful [26]. Considerable evidence suggests that TNF- α and IL-1 β contribute to the pathogenesis of liver inflammatory diseases by activating the NF- κ B signaling pathway [27], suggesting that it may be important to monitor proinflammatory cytokines when studying liver injury. In our study, we focused on the anti-inflammatory effect of oligonol by analyzing the expression of NF- κ B p65, TNF- α , and IL-1 β . Our data show that the production of TNF- α and IL-1 β were significantly increased by CCl₄-induced hepatotoxicity, which is consistent with the findings of Reyes-Gordillo et al. [28]. We found that the translocation of NF- κ B p65 protein into nucleus and TNF- α

and IL-1 β mRNA expression were inhibited in rat pretreated oligonol, suggesting that oligonol acts, at least in part, by inhibition of NF- κ B activity.

Beside proinflammatory cytokines, NF- κ B regulates the expression of the inflammatory proteins COX-2 and iNOS. This study showed a significant increase of COX-2 and iNOS mRNA expression levels in the liver treated CCl₄. These increases were attenuated by treatment with oligonol. In the study with HepG2 cells, the increased COX-2 protein levels in *t*-BHP-treated cells were also reduced by pretreatment of oligonol in a dose-dependent manner. These results suggest that oligonol exerts effects in suppressing inflammatory responses caused by CCl₄ or *t*-BHP.

A series of upstream kinases including MAPKs and Akt are involved in a relay in transmitting stimuli-induced signals to the downstream transcription factors like NF- κ B by regulating transcriptional activation of a variety of genes encoding COX-2, iNOS, TNF- α , and IL-1 β . MAPKs mediate intracellular signaling associated with a variety of cellular activities including cell proliferation, differentiation, survival, death, and transformation [29]. The mammalian MAPK family consists of ERK, JNK, and p38. The MAPK pathways are activated by diverse extracellular and intracellular stimuli including cytokines and various cellular stressors such as oxidative stress caused ROS and endoplasmic reticulum stress. Activated MAPKs phosphorylate various substrate

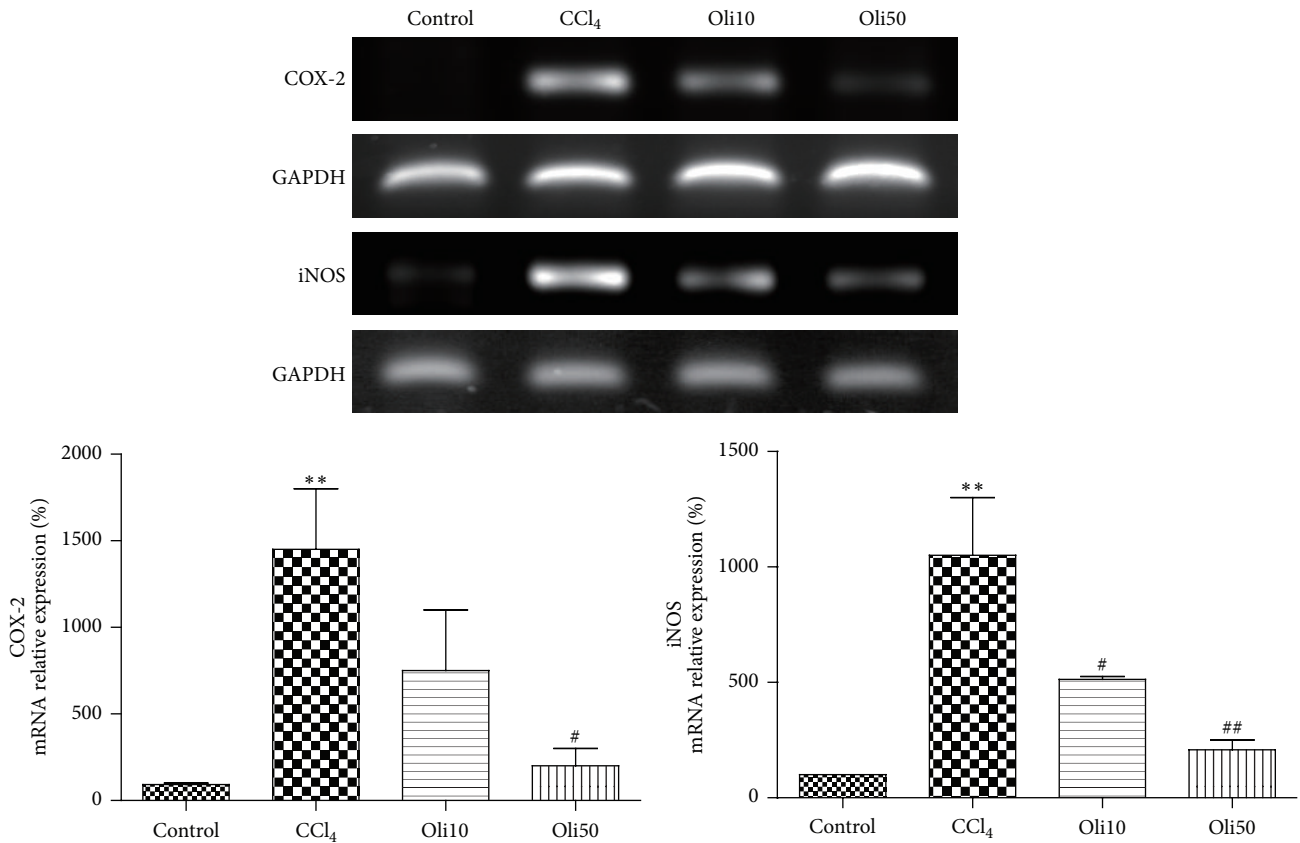


FIGURE 7: Effects of oligonol on COX-2 and iNOS mRNA expression after CCl₄ administration. RT-PCR was performed to measure COX-2 and iNOS mRNA expression in the liver tissues. Values are mean ± SE of *n* = 6. ** *p* < 0.01 compared with the control group and # *p* < 0.05 and ## *p* < 0.01 compared with the CCl₄ group.

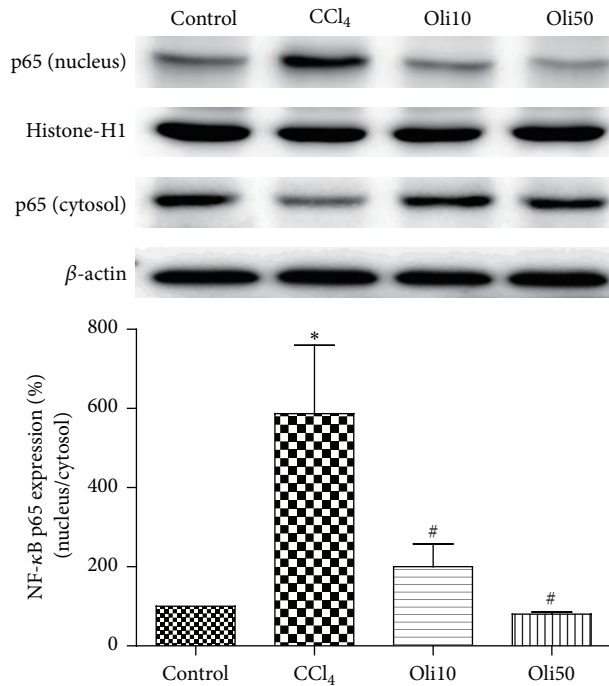


FIGURE 8: Effects of oligonol on CCl₄-induced NF-κB p65 activation. Western blotting was performed to detect nuclear and cytoplasmic localization of NF-κB p65 in the livers tissues. The relative level of NF-κB p65 in the nuclear and cytosol compared with β-actin and histon, respectively, and quantified by image analysis. Values are mean ± SE of *n* = 6. * *p* < 0.05 compared with the control group and # *p* < 0.05 compared with the CCl₄ group.

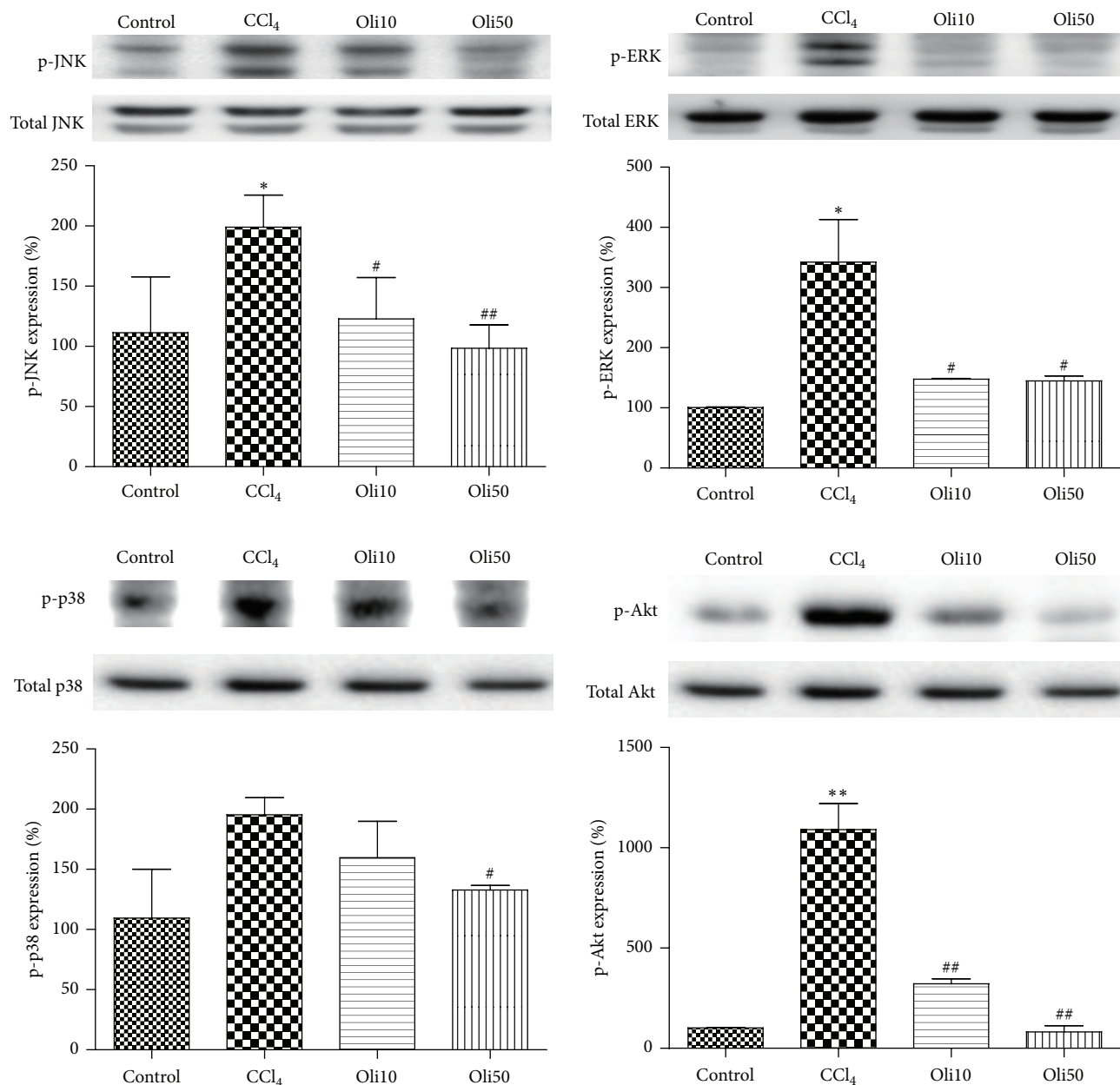


FIGURE 9: Effects of oligonol on CCl₄-induced MAPKs and Akt activation. Western blotting was performed to detect ERK, JNK, and p38 MAPK, as well as Akt, in liver tissues. Activation of these kinases was detected using their specific phosphorylated antibody. The band intensity of phosphorylated forms of MAPKs and Akt was quantified by densitometry and normalized to total forms of MAPKs and Akt, respectively. Values are mean \pm SE of $n = 6$. * $p < 0.05$ and ** $p < 0.01$ compared with the control group and # $p < 0.05$ and ## $p < 0.01$ compared with the CCl₄ group.

proteins including transcription factors like NF- κ B. Our results also showed that CCl₄ exposure induced activation of MAPKs in rat liver, and oligonol inhibited CCl₄-induced JNK, ERK, and p38 phosphorylation. In addition, in order to activate NF- κ B in response to specific stimuli, NF- κ B needs to first be liberated from its inhibitory I κ B partner [30]. On phosphorylation by I κ B kinase (IKK), I κ B is degraded by the proteasome and NF- κ B is set free. IKK can be activated by Akt. In this study, phosphorylation of Akt was increased in CCl₄-intoxicated rats, while oligonol suppressed this Akt activation. This suggests that oligonol is involved with the Akt/NF- κ B pathway.

5. Conclusion

The findings of the present study indicate that oligonol is highly effective in preventing CCl₄-induced acute liver damage, which is most likely mediated by its activity to suppress oxidative stress and lipid peroxidation as an antioxidant. It has capacity to inhibit NF- κ B p65 activation and the expression of the proinflammatory cytokines, TNF- α and IL-1 β , and proinflammatory proteins, such as COX-2 and iNOS. The underlying mechanisms for this NF- κ B inactivation may be due to inhibition of the activation of upstream kinases including ERK, JNK, and p38 MAPKs as well as Akt. Our data

suggests that oligonol may be useful as a therapeutic agent for the suppression of hepatic inflammation.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This work was supported by the National Research Foundation of Korea (NRF) grant funded by the Korean government (MSIP) (no. 2009-0083538). The authors thank the Aging Tissue Bank (Busan, Korea) for supplying research materials.

References

- [1] J. Wu and M. A. Zern, "Hepatic stellate cells: a target for the treatment of liver fibrosis," *Journal of Gastroenterology*, vol. 35, no. 9, pp. 665–672, 2000.
- [2] K. S. Lee, S. J. Lee, H. J. Park et al., "Oxidative stress effect on the activation of hepatic stellate cells," *Yonsei Medical Journal*, vol. 42, no. 1, pp. 1–8, 2001.
- [3] P. Muriel, "Peroxidation of lipids and liver damage: in Baskin," in *Antioxidants, Oxidants, and Free Radicals*, S. I. Baskin and H. Salem, Eds., Washington, DC, USA, pp. 237–257, Taylor and Francis, 1997.
- [4] D.-S. Shin, K. W. Kim, H. Y. Chung, S. Yoon, and J.-O. Moon, "Effect of sinapic acid against dimethylnitrosamine-induced hepatic fibrosis in rats," *Archives of Pharmacal Research*, vol. 36, no. 5, pp. 608–618, 2013.
- [5] D.-S. Shin, K. W. Kim, H. Y. Chung, S. Yoon, and J.-O. Moon, "Effect of sinapic acid against carbon tetrachloride-induced acute hepatic injury in rats," *Archives of Pharmacal Research*, vol. 36, no. 5, pp. 626–633, 2013.
- [6] R. H. Liu, "Health-promoting components of fruits and vegetables in the diet," *Advances in Nutrition*, vol. 4, no. 3, pp. 384–392, 2013.
- [7] M.-H. Li, J.-H. Jang, B. Sun, and Y.-J. Surh, "Protective effects of oligomers of grape seed polyphenols against beta-amyloid-induced oxidative cell death," *Annals of the New York Academy of Sciences*, vol. 1030, pp. 317–329, 2004.
- [8] J. S. Noh, H. Y. Kim, C. H. Park, H. Fujii, and T. Yokozawa, "Hypolipidaemic and antioxidative effects of oligonol, a low-molecular-weight polyphenol derived from lychee fruit, on renal damage in type 2 diabetic mice," *British Journal of Nutrition*, vol. 104, no. 8, pp. 1120–1128, 2010.
- [9] J. Y. Park, Y. Kim, J. A. Im, S. You, and H. Lee, "Inhibition of adipogenesis by Oligonol through Akt-mTOR inhibition in 3T3-L1 adipocytes," *Evidence-Based Complementary and Alternative Medicine*, vol. 2014, Article ID 895272, 11 pages, 2014.
- [10] Y. Y. Choi, T. Maeda, H. Fujii et al., "Oligonol improves memory and cognition under an amyloid β_{25-35} -induced Alzheimer's mouse model," *Nutrition Research*, vol. 34, no. 7, pp. 595–603, 2014.
- [11] E.-H. Jo, S.-J. Lee, N.-S. Ahn et al., "Induction of apoptosis in MCF-7 and MDA-MB-231 breast cancer cells by Oligonol is mediated by Bcl-2 family regulation and MEK/ERK signaling," *European Journal of Cancer Prevention*, vol. 16, no. 4, pp. 342–347, 2007.
- [12] J. K. Kundu, E.-J. Chang, H. Fujii, B. Sun, and Y.-J. Surh, "Oligonol inhibits UVB-induced COX-2 expression in HR-1 hairless mouse skin—AP-1 and C/EBP as potential upstream targets," *Photochemistry and Photobiology*, vol. 84, no. 2, pp. 399–406, 2008.
- [13] W. Brand-Williams, M. E. Cuvelier, and C. Berset, "Use of a free radical method to evaluate antioxidant activity," *LWT—Food Science and Technology*, vol. 28, no. 1, pp. 25–30, 1995.
- [14] J. A. Buege and S. D. Aust, "Microsomal lipid peroxidation," *Methods in Enzymology*, vol. 52, pp. 302–310, 1978.
- [15] K. Ishak, A. Baptista, L. Bianchi et al., "Histological grading and staging of chronic hepatitis," *Journal of Hepatology*, vol. 22, no. 6, pp. 696–699, 1995.
- [16] S. Reitman and S. Frankel, "A colorimetric method for the determination of serum glutamic oxalacetic and glutamic pyruvic transaminases," *American Journal of Clinical Pathology*, vol. 28, no. 1, pp. 56–63, 1957.
- [17] R. Agarwal and S. D. Chase, "Rapid fluorimetric-liquid chromatographic determination of malondialdehyde in biological samples," *Journal of Chromatography B: Analytical Technologies in the Biomedical & Life Sciences*, vol. 775, no. 1, pp. 121–126, 2002.
- [18] C. P. LeBel, H. Ischiropoulos, and S. C. Bondy, "Evaluation of the probe 2',7'-dichlorofluorescein as an indicator of reactive oxygen species formation and oxidative stress," *Chemical Research in Toxicology*, vol. 5, no. 2, pp. 227–231, 1992.
- [19] A. Ejarque-Ortiz, M. G. Medina, J. M. Tusell, A. P. Pérez-González, J. Serratos, and J. Saura, "Upregulation of CCAAT/enhancer binding protein β in activated astrocytes and microglia," *Glia*, vol. 55, no. 2, pp. 178–188, 2007.
- [20] W. Zhu and P. C. W. Fung, "The roles played by crucial free radicals like lipid free radicals, nitric oxide, and enzymes NOS and NADPH in CCl₄-induced acute liver injury of mice," *Free Radical Biology and Medicine*, vol. 29, no. 9, pp. 870–880, 2000.
- [21] J. A. Castro and M. I. Diaz Gomez, "Studies on the irreversible binding of 14CCl₄ to microsomal lipids in rats under varying experimental conditions," *Toxicology and Applied Pharmacology*, vol. 23, no. 4, pp. 541–552, 1972.
- [22] J. A. Castro, H. A. Sasame, H. Sussman, and J. R. Gillette, "Diverse effects of SKF 525-A and antioxidants on carbon tetrachloride-induced changes in liver microsomal P-450 content and ethylmorphine metabolism," *Life Sciences*, vol. 7, no. 3, pp. 129–136, 1968.
- [23] J. H. Ahn, J. W. Choi, J. M. Choi et al., "Protective role of oligonol from oxidative stress-induced inflammation in C6 glial cell," *Nutrition Research and Practice*, vol. 9, no. 2, pp. 123–128, 2015.
- [24] R. Krithika, R. Mohankumar, R. J. Verma et al., "Isolation, characterization and antioxidative effect of phyllanthin against CCl₄-induced toxicity in HepG2 cell line," *Chemico-Biological Interactions*, vol. 181, no. 3, pp. 351–358, 2009.
- [25] D.-S. Lee, K.-S. Kim, W. Ko et al., "The cytoprotective effect of sulfuretin against tert-butyl hydroperoxide-induced hepatotoxicity through Nrf2/ARE and JNK/ERK MAPK-mediated heme oxygenase-1 expression," *International Journal of Molecular Sciences*, vol. 15, no. 5, pp. 8863–8877, 2014.
- [26] P. Muriel, "Cytokines in liver diseases," in *Hepatotoxicity: From Genomics to In Vitro and In Vivo Models*, S. Sahu, Ed., pp. 371–389, John Wiley & Sons, West Sussex, UK, 2007.
- [27] P. Muriel, "NF- κ B in liver diseases: a target for drug therapy," *Journal of Applied Toxicology*, vol. 29, no. 2, pp. 91–100, 2009.

- [28] K. Reyes-Gordillo, J. Segovia, M. Shibayama, P. Vergara, M. G. Moreno, and P. Muriel, "Curcumin protects against acute liver damage in the rat by inhibiting NF κ B, proinflammatory cytokines production and oxidative stress," *Biochimica et Biophysica Acta—General Subjects*, vol. 1770, no. 6, pp. 989–996, 2007.
- [29] E. K. Kim and E.-J. Choi, "Pathological roles of MAPK signaling pathways in human diseases," *Biochimica et Biophysica Acta (BBA)—Molecular Basis of Disease*, vol. 1802, no. 4, pp. 396–405, 2010.
- [30] T. Luedde and R. F. Schwabe, "NF- κ B in the liver—linking injury, fibrosis and hepatocellular carcinoma," *Nature Reviews Gastroenterology & Hepatology*, vol. 8, no. 2, pp. 108–118, 2011.




Hindawi

Submit your manuscripts at
<http://www.hindawi.com>

