

REVIEW

Open Access

Diet in irritable bowel syndrome

Magdy El-Salhy^{1,2,3*} and Doris Gundersen⁴

Abstract

Irritable bowel syndrome (IBS) is a common chronic gastrointestinal disorder that is characterized by intermittent abdominal pain/discomfort, altered bowel habits and abdominal bloating/distension. This review aimed at presenting the recent developments concerning the role of diet in the pathophysiology and management of IBS. There is no convincing evidence that IBS patients suffer from food allergy/intolerance, and there is no evidence that gluten causes the debated new diagnosis of non-coeliac gluten sensitivity (NCGS). The component in wheat that triggers symptoms in NCGS appears to be the carbohydrates. Patients with NCGS appear to be IBS patients who are self-diagnosed and self-treated with a gluten-free diet. IBS symptoms are triggered by the consumption of the poorly absorbed fermentable oligo-, di-, monosaccharides and polyols (FODMAPs) and insoluble fibre. On reaching the distal small intestine and colon, FODMAPs and insoluble fibre increase the osmotic pressure in the large-intestine lumen and provide a substrate for bacterial fermentation, with consequent gas production, abdominal distension and abdominal pain or discomfort. Poor FODMAPs and insoluble fibres diet reduces the symptom and improve the quality of life in IBS patients. Moreover, it changes favourably the intestinal microbiota and restores the abnormalities in the gastrointestinal endocrine cells. Five gastrointestinal endocrine cell types that produce hormones regulating appetite and food intake are abnormal in IBS patients. Based on these hormonal abnormalities, one would expect that IBS patients to have increased food intake and body weight gain. However, the link between obesity and IBS is not fully studied. Individual dietary guidance for intake of poor FODMAPs and insoluble fibres diet in combination with probiotics intake and regular exercise is to be recommended for IBS patients.

Keywords: Appetite, Non-coeliac gluten sensitivity, Endocrine cells

Introduction

Irritable bowel syndrome (IBS) is a common chronic gastrointestinal disorder that affects 5–20% of the general population [1–15]. IBS is usually diagnosed at a young age (i.e. <50 years of age) and is more common in females than males [1,3,5,6,16]. This condition reduces considerably the patients' quality of life, although it is not known to progress to a more serious disease or to cause death [1,17–19].

IBS patients suffer from intermittent abdominal pain/discomfort, altered bowel habits and abdominal bloating/distension [1,2]. Patients believe that their symptoms are triggered by certain food items such as milk and milk products, wheat products, caffeine, cabbage, onion,

peas, beans, hot spices, and fried and smoked food [20–23]. Some IBS patients avoiding several foodstuffs, but there does not appear to be any difference between them and the general population regarding the intake of energy, carbohydrates, proteins and fats [23–29]. However, one study found that 62% of IBS patients had either limited or excluded certain food items from their daily diet, and of these 12% were at risk of long-term nutritional deficiencies [30].

The role of diet in the development of IBS symptoms and dietary management as a tool for controlling these symptoms has been the subject of several reviews [20,29,31–36]. The aim of this review was to present the recent developments concerning the role of diet in the pathophysiology and management of IBS.

Diet and the pathophysiology of IBS

It is generally accepted that diet plays an important role in the pathophysiology of IBS [27,36–45]. Several factors

* Correspondence: magdy.el-salhy@helse-fonna.no

¹Department of Medicine, Section for Gastroenterology, Stord Hospital, Stord, Norway

²Department of Clinical Medicine, Section for Gastroenterology, University of Bergen, Box 4000, 54 09 Stord, Norway

Full list of author information is available at the end of the article

have been proposed for explaining how diet influences IBS, such as food allergy/intolerance, poorly absorbed carbohydrates and fibre, and the comorbidity of obesity and IBS.

Food allergy/intolerance

Food allergy occurs in 6–8% of children and 1–4% of adults [46]. The food allergy reaction, which is mediated by immunoglobulin E, occurs within 2 hours of ingesting the offending food item, and manifests as swelling, itching, hives, wheezing, nausea, vomiting, diarrhoea, abdominal pain and collapse. There is no evidence that such an allergic reaction takes place in IBS [47–54]. A large proportion of IBS patients complain of subjective intolerance to various foods [20,21,52,55–60]. Food intolerance is a non-toxic, non-immune-mediated reaction to bioactive chemicals in food such as histamines, sulphites and monosodium glutamate, with symptoms usually manifesting outside the gastrointestinal tract. There is no documented proof that such intolerance occurs in IBS [42,54].

Non-coeliac gluten sensitivity (NCGS) has recently received attention from the mass media and the general public, and has become confused with the popular speculation that the high carbohydrate content of wheat is responsible for negative health aspects such as obesity [61]. NCGS is defined as having gastrointestinal and extra-gastrointestinal IBS-like symptoms without coeliac disease or wheat allergy, but with the symptoms being relieved by a gluten-free diet (GFD) and relapsing on gluten challenge [62–69]. The prevalence of NCGS has been reported as 0.55–6% of the USA population [64,70].

For more than 3 decades, patients with abdominal pain, diarrhoea and small-intestine biopsy findings with no significant changes have experienced symptom relief on a GFD with return of the symptoms after a gluten challenge [71,72]. Similar results have been reported in patients with non-coeliac IBS-like symptoms [73–75]. This was confirmed by double-blind randomized placebo-controlled studies [76,77]. There is disagreement as to whether or not NCGS patients have low-grade inflammation and abnormal intestinal permeability [74,77–82].

It is noteworthy that in studies showing beneficial effects on symptoms in NCGS [74,76,77], those effects were actually the result of wheat withdrawal rather than withdrawal of gluten [83]. In a placebo-controlled, cross-over study of patients with IBS-like symptoms on a self-imposed GFD [84], the gastrointestinal symptoms consistently and significantly improved when consuming a diet with reduced fermentable oligo-, di-, monosaccharides and polyols (FODMAPs), and these symptoms were not worsened by either a low- or high-dose challenge with gluten. It therefore seems that the carbohydrate

content (fructans and galactans) of wheat rather than gluten is responsible for triggering NCGS symptoms. Furthermore, in those who believed that they had NCGS, 24% had uncontrolled symptoms despite consuming a GFD, 27% did not follow a GFD alone, and 65% avoided other foods that contain high levels of FODMAPs [85]. These findings lend further support to the idea that NCGS symptoms are not triggered by the gluten in wheat.

The basic description of NCGS [67] is the same as that of IBS. Both NCGS and IBS patients have the same gastrointestinal and extra-gastrointestinal symptoms that are triggered by wheat consumption. NCGS patients have been reported to have high frequency of immunoglobulin G (IgG)/immunoglobulin A (IgA) antigliadin antibodies (AGA) and a stronger association with human leucocyte antigen-DQ2 (DQ2) and -DQ8 (DQ8) [76]. The prevalence of positivity for IgG/IgA AGA, with negative tissue transglutaminase or deamidated gliadin peptide antibodies, in the blood of IBS patients has been reported to be 5–17% [86–88] or as high as about 50% [65,89]. Serum AGA has been reported to have a good sensitivity but a low specificity for coeliac disease [90], and 12–15% of serum samples from healthy subjects are positive for AGA [86,87,90,91]. Moreover, DQ2 and DQ8 are common in the general population. It appears that NCGS patients are IBS patients with a self-diagnosis and who self-treat with a GFD. It is noteworthy in this context that 20–70% of IBS patients complain of subjective intolerance to various foods [20,21,52,55–60].

Poorly absorbed carbohydrates and fibre

The triggering of symptoms in IBS patients by certain foodstuff has been attributed to indigestible and poorly absorbed short-chain carbohydrates, FODMAPs [42,92–94] and FODMAPs intake is hypothesized to be one factor among others for IBS aetiology. These short-chain sugars include fructose, lactose, sugar alcohols (sorbitol, maltitol, mannitol, xylitol and isomalt), fructans and galactans [42]. FODMAPs occur in a wide range of foods, including wheat, rye, vegetables, fruits and legumes [95–97]. A significant proportion of these carbohydrates enter the distal small intestine and colon, where they exert osmotic effects in the large-intestine lumen, increasing its water content and providing a substrate for bacterial fermentation, with consequent gas production [92,95,98]. The produced gas causes abdominal distension and abdominal pain/discomfort. FODMAPs have been found to trigger gastrointestinal symptoms in IBS, and a low-FODMAPs diet reduces the symptoms and improves the patient's quality of life [23,29,93,94,99–102]. Recent studies have shown that the mechanisms by which FODMAPs exert their effects are more complicated than originally thought. A low-FODMAPs diet appears to induce favourable changes

in the intestinal microbiota [103] and gastrointestinal endocrine cells [104-107].

It has been reported that changing from typical Australian food to a low-FODMAPs diet changed the intestinal microbiota [103]. Thus, a low-FODMAPs diet in healthy subjects and IBS patients reduced the total bacterial abundance, while a typical Australian diet increased the relative abundance of butyrate-producing *Clostridium cluster XIVa* and the mucus-associated *Akkermansia muciniphila*, and reduced *Ruminococcus torques* [103].

Several types of endocrine cell in all segments of the gastrointestinal tract of IBS patients are abnormal [108-129]. The gastrointestinal endocrine cells interact and integrate with each other, with the enteric nervous system and with the afferent and efferent nerve fibres of the central nervous system, in particular the autonomic nervous system [42,130-132]. These cells regulate several functions of the gastrointestinal tract, including sensation, motility, secretion, absorption, local immune defence and food intake (by affecting appetite) [42,131-134]. The abnormalities in the gastrointestinal endocrine cells are considered to play a major role in the development of symptoms in IBS, and therefore represent future targets for treatment [43,135]. Switching from a typical Norwegian diet to a low-FODMAPs diet was shown to lead to normalization of the endocrine cells in the stomach and large intestines [104-107].

A low intake of dietary fibre was initially believed to be the cause of IBS [136]. In clinical settings the increase in dietary fibre intake in IBS patients has been found to increase abdominal pain, bloating and abdominal distension. A meta-analysis of 12 trials revealed that IBS patients treated with increased fibre intake had no improvement in symptoms compared to placebo or a low-fibre diet [137]. However, it has been reported that

water-soluble fibre—but not insoluble fibre—improves the symptoms [138,139].

Obesity and IBS

As mentioned above, IBS patients tend to avoid certain food items that they associate with the onset of their symptoms. There has been some concern that the onset of IBS symptoms upon ingesting certain foods would reduce the amount of food consumed and thereby lead to malnutrition [30]. However, whereas an association between low BMI and IBS in 367 patients with IBS has been reported [140], in another report most of the 330 IBS patients examined were either normal or overweight [20]. In a recent comprehensive review, the association between IBS and obesity was found to be controversial, and the author concluded that obesity and IBS might be linked [141].

Appetite is regulated by a large number of hormones, several of which are secreted by gastrointestinal endocrine cells [142]. The gastrointestinal hormones exert their effects by acting upon the appetite control centre in the hypothalamus [142]. The arcuate nucleus (ARC) lies in the median eminence, which lacks a complete blood barrier, making the ARC particularly susceptible to hormones circulating in the blood [142-145]. The ARC is the centre that integrates the neurological and blood-borne signals [142-145]. The brain reward system in the midbrain controls hedonic feeding (i.e. the consumption of palatable food), which is modulated by blood-borne signals [145].

The following five gastrointestinal endocrine cell types that secrete hormones that regulate appetite are abnormal in patients with IBS: ghrelin, cholecystokinin (CCK), peptide YY (PYY), enteroglucagon (oxyntomodulin) and

Table 1 Abnormalities in the gastrointestinal endocrine cells that regulate appetite in IBS patients

Gastrointestinal segment	Hormone	Cell density			Hormone function
		IBS-D	IBS-M	IBS-C	
Stomach	Ghrelin	Increased	Unchanged	Decreased	Orexigenic (increases appetite)
	Serotonin	Increased	Unchanged	Decreased	Anorexigenic (decreases appetite)
Duodenum	CCK	Decreased	Unchanged	Unchanged	Anorexigenic (decreases appetite)
	Serotonin	Unchanged	Unchanged	Unchanged	See above
Ileum	PYY	Unchanged	Unchanged	Increased	Anorexigenic (decreases appetite)
	Serotonin	Decreased	Decreased	Decreased	See above
Colon	PYY	Decreased	Unknown	Decreased	See above
	Serotonin	Decreased	Unknown	Decreased	See above
Rectum	PYY	Decreased	Decreased	Decreased	See above
	Enteroglucagon	Decreased	Unknown	Decreased	Anorexigenic (decreases appetite)
	Serotonin	Unchanged	Unknown	Unchanged	See above

IBS-D, IBS patients with diarrhoea as the predominant symptom; IBS-M, IBS patients with alternating diarrhea and constipation; IBS-C, IBS patients with constipation as the predominant symptom.

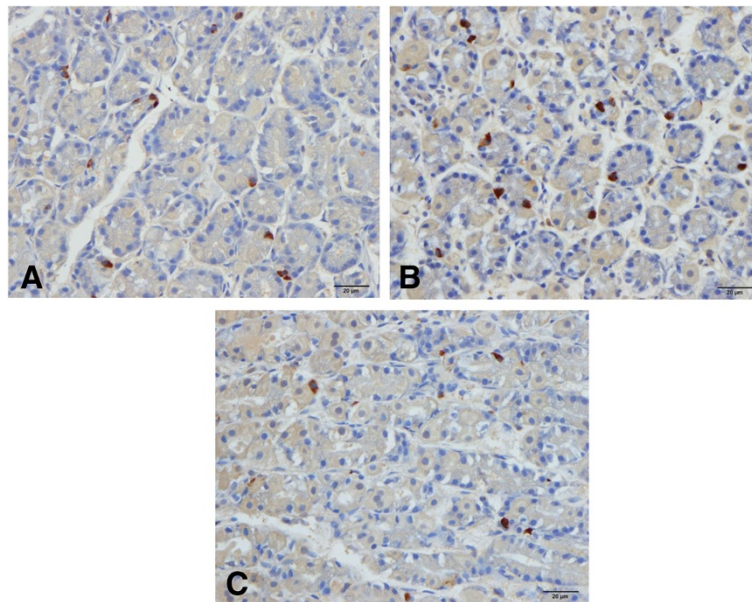


Figure 1 Ghrelin-immunoreactive cells in the oxyntic mucosa of a healthy subject (A), in a patient with IBS-D (B) and in a patient with IBS-C (C).

serotonin (Table 1 and Figures 1, 2, 3, 4 and 5) [108-110,112-115,132,146-149]. The endocrine cells in the oxyntic mucosa are the main source of circulating ghrelin, although small amounts do occur in the small and large intestines as well as in the ARC of the hypothalamus [144,150-153]. Ghrelin has several roles, such as regulating the release of pituitary growth hormone and accelerating gastric and intestinal motility [150-166]. Moreover, ghrelin increases appetite and feeding; central or peripheral administration of ghrelin stimulates the consumption of food and body weight gain [150]. CCK stimulates gallbladder contraction, intestinal motility and pancreatic exocrine secretion, and inhibits gastric motility and food consumption [131]. The anorexigenic action of CCK occurs via the CCK-B (CCK-2) receptor, which is the predominant receptor type in the brain [167-183]. PYY release is proportional to the composition (including calorie content) of a particular meal, and its infusion reduces the consumption of food

[184,185]. PYY binds to Y2 receptors localized on the presynaptic terminals of neuropeptide Y and to agouti-related protein neurons in the hypothalamus, which causes inactivation of these neurons, resulting in anorexia [186]. Moreover, PYY is the main regulator of the ileal brake and consequently inhibits the consumption of further food once nutrients reach the terminal ileum [187-194]. Enteroglucagon reduces gastric motility and secretion [195-200] and, similar to PYY, the amount released into the bloodstream is proportional to the calories ingested [201,202]; however, enteroglucagon seems to have only a modest anorexigenic effect [145]. Serotonin has also been reported to have an anorexigenic effect [203].

Whereas the ghrelin cell density is increased in IBS patients with diarrhoea as the predominant symptom (IBS-D), it is reduced in IBS patients with constipation as the predominant symptom (IBS-C). It is therefore reasonable to assume that IBS-D patients would have a

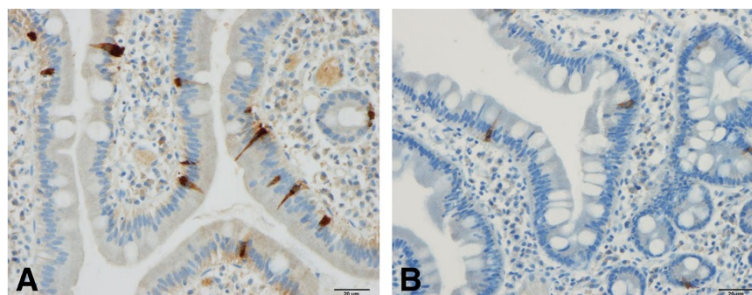


Figure 2 Duodenal CCK cells in a healthy subject (A) and in a patient with IBS (B).

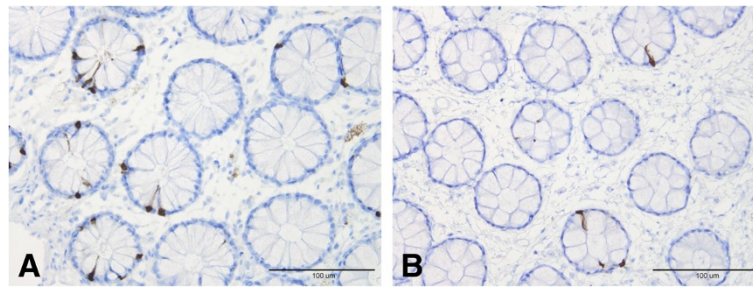


Figure 3 PYY cells in the colon of a healthy subject (A) and in a patient with IBS (B).

greater appetite and food intake than those with IBS-C. In both IBS subtypes, the densities of the four endocrine cell types that produce anorexigenic hormones—namely CCK, PYY, enteroglucagon and serotonin—are reduced. Thus, the changes in the gastrointestinal endocrine cells regulating appetite in IBS patients favour an increase in food consumption. BMI and appetite in IBS patients have not been studied in detail, and the available data are controversial. It is not clear whether IBS patients have a greater appetite, which is opposed by the avoidance of eating because of worsening of symptoms upon eating. Further studies are needed to clarify this issue.

Diet and management of IBS

As mentioned above, it is recommended that IBS patients consume a diet that is poor in FODMAPs and insoluble fibre. IBS patients make a conscious choice to avoid certain food items, some of which are rich in FODMAPs, but they may also consume large amounts of other food items that are rich in FODMAPs and avoid food sources that are important to health maintenance [23]. Dietary guidance is therefore important for IBS patients [23,99]. Furthermore, in our experience individual dietary guidance is preferable due to the wide variety of individual tolerances to different FODMAPs-rich food items, probably due to the intestinal microbiota differing between individuals.

Consuming probiotics increases the tolerance for FODMAPs-rich foodstuffs, and adding regular exercise amplifies the beneficial effects of such a diet [1,204]. People in numerous countries (including Norway) rely on bread and wheat products for a substantial part of their diet [205]. Gluten-free bread (mostly made of rice/corn) contains 0.19 g/100 g fructans, and bread made with spelt flour contains 0.14 g/100 g [96]. Either gluten-free products or spelt products can be consumed. Many of our patients consume spelt bread and spelt products rather than gluten-free products, with satisfactory results. The protein content of spelt flour is also 16% lower in terms of gluten than that in wheat [206].

Conclusion

Diet plays a major role in the pathophysiology of IBS and is a powerful tool for managing IBS. There is no convincing evidence that IBS patients suffer from food allergy/intolerance, and there is evidence that NCGS is caused by the fructans in wheat, rather than by gluten. NCGS patients appear to be IBS patients who have self-diagnosed and self-treated with a GFD. Food items that are rich in poorly absorbed short-chain carbohydrates (FODMAPs) and insoluble fibre trigger IBS symptoms. There appear to be several mechanisms by which these food items exert this effect (Figure 6):

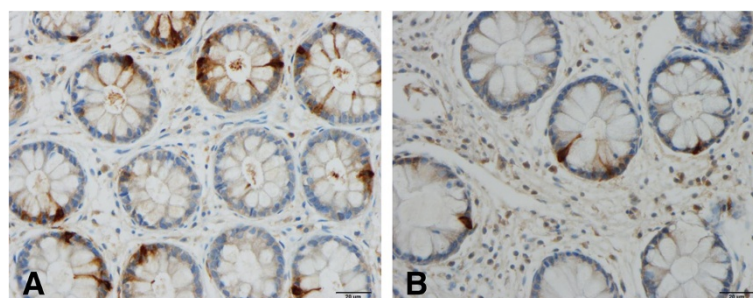


Figure 4 Rectal enteroglucagon cells in a healthy subject (A) and in a patient with IBS (B).

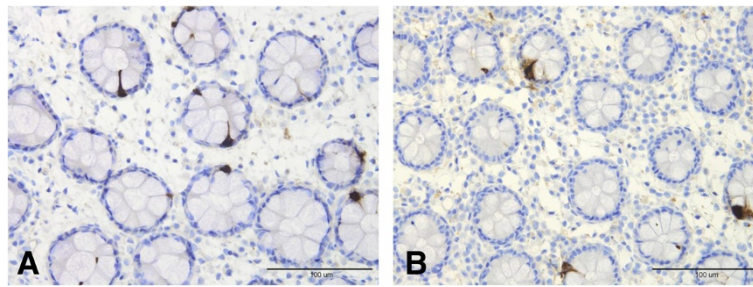


Figure 5 Serotonin cells in the colon of a healthy subject (A) and in a patient with IBS (B).

1. Upon entering the distal small intestine and colon, they increase the osmotic pressure and provide a substrate for bacterial fermentation, resulting in gas production, abdominal distension and abdominal pain/discomfort.
2. They are prebiotics that favour the colonization of the large intestine with *Clostridium* bacteria, which produce gas on fermentation.

3. They affect the gastrointestinal endocrine cells that regulate gastrointestinal sensation, motility, secretion and absorption, as well as local immune defence and food consumption.

The link between obesity and IBS is an interesting area that needs to be explored further. This is of particular interest since IBS patients have an increased density of

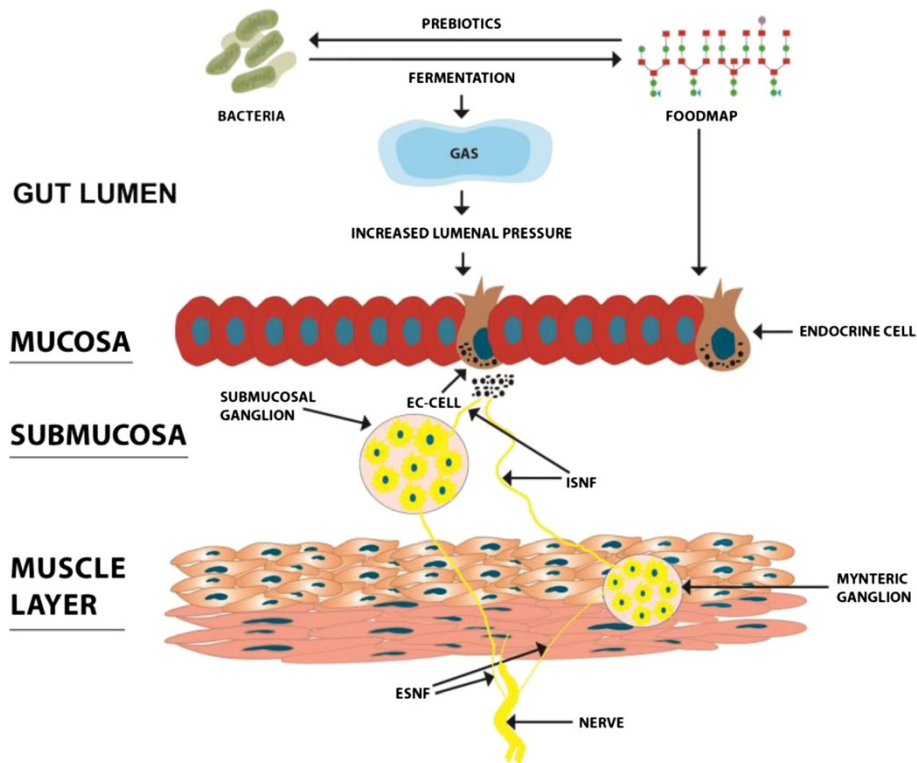


Figure 6 Schematic illustration for the possible mechanisms by which FODMAPs can trigger IBS symptoms. Upon reaching the large intestine FODMAPs can exert direct or indirect effect on the intestinal endocrine cells. They act as prebiotics and change the intestinal flora and they are fermented by the intestinal microbiota with gas production. The production of gas increases the luminal pressure and stimulates the release of serotonin from serotonin (EC) cells. Serotonin act on the intrinsic sensory nerve fibres (ISNF) of the submucosal and myenteric ganglia, which in turn convey the activation to the extrinsic sensory nerve fibres (ESNF) to the central nervous system.

ghrelin cells, which increases appetite, stimulates the reconsumption of food and body weight gain, and have decreased densities of the four endocrine cells that produce anorexigenic hormones, namely CCK, PYY, enteroglucagon and serotonin.

A diet that is poor in FODMAPs and insoluble fibre reduces the symptoms and improves the quality of life of IBS patients. Individual dietary guidance is necessary to identify a suitable diet to which the patient is likely to adhere to in the long term. Combining this diet with probiotics and regular exercise will amplify the effect of such a diet.

Abbreviations

AGA: Anti-gliadin antibodies; ARC: Arcuate nucleus; FODMAPs: Fermentable oligo-, di-, monosaccharides and polyols; GFD: Gluten-free diet; IBS: Irritable bowel syndrome; IBS-D: IBS patients with diarrhoea as the predominant symptom; IBS-C: IBS patients with constipation as the predominant symptom; NCGS: Non-coeliac gluten sensitivity.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MES delimited the topics, performed the bibliographic search and drafted the manuscript. GD contributed to the planning of the review and made comments that improved the manuscript. Both authors read and approved the final manuscript.

Acknowledgements

This work was supported by grants from Helse-Vest and Helse-Fonna.

Author details

¹Department of Medicine, Section for Gastroenterology, Stord Hospital, Stord, Norway. ²Department of Clinical Medicine, Section for Gastroenterology, University of Bergen, Box 4000, 54 09 Stord, Norway. ³Department of Medicine, National Centre for Functional Gastrointestinal Disorders, Haukeland University Hospital, Bergen, Norway. ⁴Department of Research, Helse-Fonna, Haugesund Hospital, Haugesund, Norway.

Received: 24 January 2015 Accepted: 31 March 2015

Published online: 14 April 2015

References

1. El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T. Irritable bowel syndrome: diagnosis, pathogenesis and treatment options. New York: Nova Science Publishers, Inc.; 2012.
2. Thompson WG. A World View of IBS. In: Camilleri M, Spiller RC, editors. Irritable Bowel Syndrome. Philadelphia and London: Saunders; 2002. p. 17–26.
3. Agreus L, Svardsudd K, Nyren O, Tibblin G. Irritable bowel syndrome and dyspepsia in the general population: overlap and lack of stability over time. *Gastroenterology*. 1995;109:671–80.
4. Thompson WG, Heaton KW. Functional bowel disorders in apparently healthy people. *Gastroenterology*. 1980;79:283–8.
5. Kennedy TM, Jones RH, Hungin AP, O'Flanagan H, Kelly P. Irritable bowel syndrome, gastro-oesophageal reflux, and bronchial hyper-responsiveness in the general population. *Gut*. 1998;43:770–4.
6. Drossman DA, Li Z, Andruzzi E, Temple RD, Talley NJ, Thompson WG, et al. U.S. household survey of functional gastrointestinal disorders. Prevalence, sociodemography, and health impact. *Dig Dis Sci*. 1993;38:1569–80.
7. Talley NJ, Gabriel SE, Harmsen WS, Zinsmeister AR, Evans RW. Medical costs in community subjects with irritable bowel syndrome. *Gastroenterology*. 1995;109:1736–41.
8. Hungin AP, Whorwell PJ, Tack J, Mearin F. The prevalence, patterns and impact of irritable bowel syndrome: an international survey of 40,000 subjects. *Aliment Pharmacol Ther*. 2003;17:643–50.
9. Jones R, Lydeard S. Irritable bowel syndrome in the general population. *BMJ*. 1992;304:87–90.
10. Bordie AK. Functional disorders of the colon. *J Indian Med Assoc*. 1972;58:451–6.
11. O'Keefe EA, Talley NJ, Zinsmeister AR, Jacobsen SJ. Bowel disorders impair functional status and quality of life in the elderly: a population-based study. *J Gerontol A Biol Sci Med Sci*. 1995;50:M184–9.
12. Everhart JE, Renault PF. Irritable bowel syndrome in office-based practice in the United States. *Gastroenterology*. 1991;100:998–1005.
13. Wilson S, Roberts L, Roalfe A, Bridge P, Singh S. Prevalence of irritable bowel syndrome: a community survey. *Br J Gen Pract*. 2004;54:495–502.
14. Quigley EM, Locke GR, Mueller-Lissner S, Paulo LG, Tytgat GN, Helfrich I, et al. Prevalence and management of abdominal cramping and pain: a multinational survey. *Aliment Pharmacol Ther*. 2006;24:411–9.
15. Harvey RF, Salih SY, Read AE. Organic and functional disorders in 2000 gastroenterology outpatients. *Lancet*. 1983;1:632–4.
16. Thompson WG, Irvine EJ, Pare P, Ferrazzi S, Rance L. Functional gastrointestinal disorders in Canada: first population-based survey using Rome II criteria with suggestions for improving the questionnaire. *Dig Dis Sci*. 2002;47:225–35.
17. Miller V, Whitaker K, Morris JA, Whorwell PJ. Gender and irritable bowel syndrome: the male connection. *J Clin Gastroenterol*. 2004;38:558–60.
18. Whitehead WE, Burnett CK, Cook 3rd EW, Taub E. Impact of irritable bowel syndrome on quality of life. *Dig Dis Sci*. 1996;41:2248–53.
19. Gralnek IM, Hays RD, Kilbourne A, Naliboff B, Mayer EA. The impact of irritable bowel syndrome on health-related quality of life. *Gastroenterology*. 2000;119:654–60.
20. Simren M, Mansson A, Langkilde AM, Svedlund J, Abrahamsson H, Bengtsson U, et al. Food-related gastrointestinal symptoms in the irritable bowel syndrome. *Digestion*. 2001;63:108–15.
21. Nanda R, James R, Smith H, Dudley CR, Jewell DP. Food intolerance and the irritable bowel syndrome. *Gut*. 1989;30:1099–104.
22. Bohn L, Storsrud S, Tornblom H, Bengtsson U, Simren M. Self-reported food-related gastrointestinal symptoms in IBS are common and associated with more severe symptoms and reduced quality of life. *Am J Gastroenterol*. 2013;108:634–41.
23. Ostgaard H, Hausken T, Gundersen D, El-Salhy M. Diet and effects of diet management on quality of life and symptoms in patients with irritable bowel syndrome. *Mol Med Report*. 2012;5:1382–90.
24. Jarrett M, Heitkemper MM, Bond EF, Georges J. Comparison of diet composition in women with and without functional bowel disorder. *Gastroenterol Nurs*. 1994;16:253–8.
25. Saito YA, Locke 3rd GR, Weaver AL, Zinsmeister AR, Talley NJ. Diet and functional gastrointestinal disorders: a population-based case-control study. *Am J Gastroenterol*. 2005;100:2743–8.
26. Williams EA, Nai X, Corfe BM. Dietary intakes in people with irritable bowel syndrome. *BMC Gastroenterol*. 2011;11:9.
27. Bohn L, Storsrud S, Simren M. Nutrient intake in patients with irritable bowel syndrome compared with the general population. *Neurogastroenterol Motil*. 2013;25:23–e21.
28. Ligaarden SC, Lydersen S, Farup PG. Diet in subjects with irritable bowel syndrome: a cross-sectional study in the general population. *BMC Gastroenterol*. 2012;12:61.
29. El-Salhy M, Ostgaard H, Gundersen D, Hatlebakk JG, Hausken T. The role of diet in the pathogenesis and management of irritable bowel syndrome (Review). *Int J Mol Med*. 2012;29:723–31.
30. Monsbakken KW, Vandvik PO, Farup PG. Perceived food intolerance in subjects with irritable bowel syndrome—etiology, prevalence and consequences. *Eur J Clin Nutr*. 2006;60:667–72.
31. Wald A, Rakel D. Behavioral and complementary approaches for the treatment of irritable bowel syndrome. *Nutr Clin Pract*. 2008;23:284–92.
32. Heizer WD, Southern S, McGovern S. The role of diet in symptoms of irritable bowel syndrome in adults: a narrative review. *J Am Diet Assoc*. 2009;109:1204–14.
33. Morcos A, Dinan T, Quigley EM. Irritable bowel syndrome: role of food in pathogenesis and management. *J Dig Dis*. 2009;10:237–46.
34. Eswaran S, Tack J, Chey WD. Food: the forgotten factor in the irritable bowel syndrome. *Gastroenterol Clin North Am*. 2011;40:141–62.
35. Austin GL, Dalton CB, Hu Y, Morris CB, Hankins J, Weinland SR, et al. A very low-carbohydrate diet improves symptoms and quality of life in diarrhea-predominant irritable bowel syndrome. *Clin Gastroenterol Hepatol*. 2009;7:706–8.

36. El-Salhy M, Gilja OH, Gundersen D, Hatlebakk JG, Hausken T. Interaction between ingested nutrients and gut endocrine cells in patients with irritable bowel syndrome (Review). *Int J Mol Med*. 2014;34:363–71.
37. Lee YJ, Park KS. Irritable bowel syndrome: Emerging paradigm in pathophysiology. *World J Gastroenterol*. 2014;20:2456–69.
38. Asare F, Storsrud S, Simren M. Meditation over medication for irritable bowel syndrome? On exercise and alternative treatments for irritable bowel syndrome. *Curr Gastroenterol Rep*. 2012;14:283–9.
39. Gibson PR. Food intolerance in functional bowel disorders. *J Gastroenterol Hepatol*. 2011;26 Suppl 3:128–31.
40. Gibson PR, Shepherd SJ. Food choice as a key management strategy for functional gastrointestinal symptoms. *Am J Gastroenterol*. 2012;107:657–66. quiz 667.
41. Soares RL. Irritable bowel syndrome: A clinical review. *World J Gastroenterol*. 2014;20:12144–60.
42. El-Salhy M. Irritable bowel syndrome: diagnosis and pathogenesis. *World J Gastroenterol*. 2012;18:5151–63.
43. El-Salhy M, Gundersen D, Gilja OH, Hatlebakk JG, Hausken T. Is irritable bowel syndrome an organic disorder? *World J Gastroenterol*. 2014;20:384–400.
44. El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T. Diet and Irritable Bowel Syndrome, with a Focus on Appetite-Regulating Hormones. In: Watson RR, editor. *Nutrition in the Prevention and Treatment of Abdominal Obesity*. San Diego: Elsevier; 2014. p. 5–16.
45. El-Salhy M, Hatlebakk JG, Gilja OH, Hausken T. Irritable bowel syndrome: recent developments in diagnosis, pathophysiology, and treatment. *Expert Rev Gastroenterol Hepatol*. 2014;8:435–43.
46. Stefanini GF, Saggiaro A, Alvisi V, Angelini G, Capurso L, di Lorenzo G, et al. Oral cromolyn sodium in comparison with elimination diet in the irritable bowel syndrome, diarrheic type. Multicenter study of 428 patients. *Scand J Gastroenterol*. 1995;30:535–41.
47. Whorwell PJ. The growing case for an immunological component to irritable bowel syndrome. *Clin Exp Allergy*. 2007;37:805–7.
48. Zar S, Benson MJ, Kumar D. Food-specific serum IgG4 and IgE titers to common food antigens in irritable bowel syndrome. *Am J Gastroenterol*. 2005;100:1550–7.
49. Park MI, Camilleri M. Is there a role of food allergy in irritable bowel syndrome and functional dyspepsia? A systematic review. *Neurogastroenterol Motil*. 2006;18:595–607.
50. Uz E, Turkay C, Aytac S, Bavbek N. Risk factors for irritable bowel syndrome in Turkish population: role of food allergy. *J Clin Gastroenterol*. 2007;41:380–3.
51. Dainese R, Galliani EA, De Lazzari F, Di Leo V, Naccarato R. Discrepancies between reported food intolerance and sensitization test findings in irritable bowel syndrome patients. *Am J Gastroenterol*. 1999;94:1892–7.
52. Bischoff S, Crowe SE. Gastrointestinal food allergy: new insights into pathophysiology and clinical perspectives. *Gastroenterology*. 2005;128:1089–113.
53. McKee AM, Prior A, Whorwell PJ. Exclusion diets in irritable bowel syndrome: are they worthwhile? *J Clin Gastroenterol*. 1987;9:526–8.
54. Boettcher E, Crowe SE. Dietary proteins and functional gastrointestinal disorders. *Am J Gastroenterol*. 2013;108:728–36.
55. Young E, Stoneham MD, Petrukevitch A, Barton J, Rona R. A population study of food intolerance. *Lancet*. 1994;343:1127–30.
56. Locke 3rd GR, Zinsmeister AR, Talley NJ, Fett SL, Melton LJ. Risk factors for irritable bowel syndrome: role of analgesics and food sensitivities. *Am J Gastroenterol*. 2000;95:157–65.
57. Bischoff SC, Herrmann A, Manns MP. Prevalence of adverse reactions to food in patients with gastrointestinal disease. *Allergy*. 1996;51:811–8.
58. Jones JG, Elmes ME. The measurement of mucosal non-myelinated nerve fibre area and endocrine cell area in coeliac disease using morphometric analysis. *Diagn Histopathol*. 1982;5:183–8.
59. Bhat K, Harper A, Gorard DA. Perceived food and drug allergies in functional and organic gastrointestinal disorders. *Aliment Pharmacol Ther*. 2002;16:969–73.
60. Bijkerk CJ, de Wit NJ, Stalman WA, Knottnerus JA, Hoes AW, Muris JW. Irritable bowel syndrome in primary care: the patients' and doctors' views on symptoms, etiology and management. *Can J Gastroenterol*. 2003;17:363–8. quiz 405–366.
61. Davis W. *Wheat belly: lose the wheat, lose the weight and find your path back to health*. New York: Rodale; 2011.
62. Lundin KE. Non-coeliac gluten sensitivity - why worry? *BMC Med*. 2014;12:86.
63. Aziz I, Sanders DS. Emerging concepts: from coeliac disease to non-coeliac gluten sensitivity. *Proc Nutr Soc*. 2012;71:576–80.
64. Mansueto P, Seidita A, D'Alcamo A, Carroccio A. Non-coeliac gluten sensitivity: literature review. *J Am Coll Nutr*. 2014;33:39–54.
65. Volta U, De Giorgio R. New understanding of gluten sensitivity. *Nat Rev Gastroenterol Hepatol*. 2012;9:295–9.
66. Sapone A, Bai JC, Ciacci C, Dolinsek J, Green PH, Hadjivassiliou M, et al. Spectrum of gluten-related disorders: consensus on new nomenclature and classification. *BMC Med*. 2012;10:13.
67. Catassi C, Bai JC, Bonaz B, Bouma G, Calabro A, Carroccio A, et al. Non-Celiac Gluten sensitivity: the new frontier of gluten related disorders. *Nutrients*. 2013;5:3839–53.
68. Sestak K, Fortgang I. Celiac and non-coeliac forms of gluten sensitivity: shifting paradigms of an old disease. *Br Microbiol Res*. 2013;3:585–9.
69. Czaja-Bulsa G. Non coeliac gluten sensitivity - a new disease with gluten intolerance. *Clin Nutr*. 2015;34:189–94.
70. Piston F, Gil-Humanes J, Barro F. Integration of promoters, inverted repeat sequences and proteomic data into a model for high silencing efficiency of coeliac disease related gliadins in bread wheat. *BMC Plant Biol*. 2013;13:136.
71. Ellis A, Linaker BD. Non-coeliac gluten sensitivity? *Lancet*. 1978;1:1358–9.
72. Cooper BT, Holmes GK, Ferguson R, Thompson RA, Allan RN, Cooke WT. Gluten-sensitive diarrhea without evidence of celiac disease. *Gastroenterology*. 1980;79:801–6.
73. Campanella J, Biagi F, Bianchi PI, Zanellati G, Marchese A, Corazza GR. Clinical response to gluten withdrawal is not an indicator of coeliac disease. *Scand J Gastroenterol*. 2008;43:1311–4.
74. Vazquez-Roque MI, Camilleri M, Smyrk T, Murray JA, Marietta E, O'Neill J, et al. A controlled trial of gluten-free diet in patients with irritable bowel syndrome-diarrhea: effects on bowel frequency and intestinal function. *Gastroenterology*. 2013;144:903–11.
75. Kaukinen K, Turjanmaa K, Maki M, Partanen J, Venalainen R, Reunala T, et al. Intolerance to cereals is not specific for coeliac disease. *Scand J Gastroenterol*. 2000;35:942–6.
76. Carroccio A, Mansueto P, Iacono G, Soresi M, D'Alcamo A, Cavataio F, et al. Non-coeliac wheat sensitivity diagnosed by double-blind placebo-controlled challenge: exploring a new clinical entity. *Am J Gastroenterol*. 2012;107:1898–906. quiz 1907.
77. Biesiekierski JR, Newnham ED, Irving PM, Barrett JS, Haines M, Doecke JD, et al. Gluten causes gastrointestinal symptoms in subjects without celiac disease: a double-blind randomized placebo-controlled trial. *Am J Gastroenterol*. 2011;106:508–14. quiz 515.
78. Verdu EF, Armstrong D, Murray JA. Between celiac disease and irritable bowel syndrome: the "no man's land" of gluten sensitivity. *Am J Gastroenterol*. 2009;104:1587–94.
79. Carroccio A, Brusca I, Mansueto P, Pirrone G, Barrale M, Di Prima L, et al. A cytologic assay for diagnosis of food hypersensitivity in patients with irritable bowel syndrome. *Clin Gastroenterol Hepatol*. 2010;8:254–60.
80. Carroccio A, Brusca I, Mansueto P, D'Alcamo A, Barrale M, Soresi M, et al. A comparison between two different in vitro basophil activation tests for gluten- and cow's milk protein sensitivity in irritable bowel syndrome (IBS)-like patients. *Clin Chem Lab Med*. 2013;51:1257–63.
81. Bucci C, Zingone F, Russo I, Morra I, Tortora R, Pogna N, et al. Gliadin does not induce mucosal inflammation or basophil activation in patients with nonceliac gluten sensitivity. *Clin Gastroenterol Hepatol*. 2013;11:1294–9.
82. Sapone A, Lammers KM, Casolaro V, Cammarota M, Giuliano MT, De Rosa M, et al. Divergence of gut permeability and mucosal immune gene expression in two gluten-associated conditions: celiac disease and gluten sensitivity. *BMC Med*. 2011;9:23.
83. Nijeboer P, Bontkes HJ, Mulder CJ, Bouma G. Non-coeliac gluten sensitivity. Is it in the gluten or the grain? *J Gastrointest Liver Dis*. 2013;22:435–40.
84. Biesiekierski JR, Peters SL, Newnham ED, Rosella O, Muir JG, Gibson PR. No effects of gluten in patients with self-reported non-coeliac gluten sensitivity after dietary reduction of fermentable, poorly absorbed, short-chain carbohydrates. *Gastroenterology*. 2013;145:320–8.
85. Biesiekierski JR, Newnham ED, Shepherd SJ, Muir JG, Gibson PR. Characterization of Adults With a Self-Diagnosis of Nonceliac Gluten Sensitivity. *Nutr Clin Pract*. 2014;29:504–9.
86. Sanders DS, Carter MJ, Hurlstone DP, Pearce A, Ward AM, McAlindon ME, et al. Association of adult coeliac disease with irritable bowel syndrome: a

- case-control study in patients fulfilling ROME II criteria referred to secondary care. *Lancet*. 2001;358:1504–8.
87. Sanders DS, Patel D, Stephenson TJ, Ward AM, McCloskey EV, Hadjivassiliou M, et al. A primary care cross-sectional study of undiagnosed adult coeliac disease. *Eur J Gastroenterol Hepatol*. 2003;15:407–13.
 88. Elloumi H, El Assoued Y, Ghedira I, Ben Abdelaziz A, Yacoobi MT, Ajmi S. [Immunological profile of coeliac disease in a subgroup of patients with symptoms of irritable bowel syndrome]. *Tunis Med*. 2008;86:802–5.
 89. Volta U, Tovoli F, Cicola R, Parisi C, Fabbri A, Piscaglia M, et al. Serological tests in gluten sensitivity (nonceliac gluten intolerance). *J Clin Gastroenterol*. 2012;46:680–5.
 90. Ruuskanen A, Kaukinen K, Collin P, Huhtala H, Valve R, Maki M, et al. Positive serum anti gliadin antibodies without celiac disease in the elderly population: does it matter? *Scand J Gastroenterol*. 2010;45:1197–202.
 91. Hadjivassiliou M, Gibson A, Davies-Jones GA, Lobo AJ, Stephenson TJ, Milford-Ward A. Does cryptic gluten sensitivity play a part in neurological illness? *Lancet*. 1996;347:369–71.
 92. Barrett JS, Geary RB, Muir JG, Irving PM, Rose R, Rosella O, et al. Dietary poorly absorbed, short-chain carbohydrates increase delivery of water and fermentable substrates to the proximal colon. *Aliment Pharmacol Ther*. 2010;31:874–82.
 93. Barrett JS, Gibson PR. Fermentable oligosaccharides, disaccharides, monosaccharides and polyols (FODMAPs) and nonallergic food intolerance: FODMAPs or food chemicals? *Therap Adv Gastroenterol*. 2012;5:261–8.
 94. Gibson PR, Shepherd SJ. Evidence-based dietary management of functional gastrointestinal symptoms: The FODMAP approach. *J Gastroenterol Hepatol*. 2010;25:252–8.
 95. Shepherd SJ, Lomer MC, Gibson PR. Short-chain carbohydrates and functional gastrointestinal disorders. *Am J Gastroenterol*. 2013;108:707–17.
 96. Biesiekierski JR, Rosella O, Rose R, Liels K, Barrett JS, Shepherd SJ, et al. Quantification of fructans, galacto-oligosaccharides and other short-chain carbohydrates in processed grains and cereals. *J Hum Nutr Diet*. 2011;24:154–76.
 97. Muir JG, Rose R, Rosella O, Liels K, Barrett JS, Shepherd SJ, et al. Measurement of short-chain carbohydrates in common Australian vegetables and fruits by high-performance liquid chromatography (HPLC). *J Agric Food Chem*. 2009;57:554–65.
 98. Marcason W. What is the FODMAP diet? *J Acad Nutr Diet*. 2012;112:1696.
 99. Mazzawi T, Hausken T, Gundersen D, El-Salhy M. Effects of dietary guidance on the symptoms, quality of life and habitual dietary intake of patients with irritable bowel syndrome. *Mol Med Rep*. 2013;8:845–52.
 100. Halmos EP, Power VA, Shepherd SJ, Gibson PR, Muir JG. A diet low in FODMAPs reduces symptoms of irritable bowel syndrome. *Gastroenterology*. 2014;146:67–75.
 101. Ong DK, Mitchell SB, Barrett JS, Shepherd SJ, Irving PM, Biesiekierski JR, et al. Manipulation of dietary short chain carbohydrates alters the pattern of gas production and genesis of symptoms in irritable bowel syndrome. *J Gastroenterol Hepatol*. 2010;25:1366–73.
 102. Gibson PR, Shepherd SJ. Personal view: food for thought—western lifestyle and susceptibility to Crohn's disease. *FODMAP hypothesis Aliment Pharmacol Ther*. 2005;21:1399–409.
 103. Halmos EP, Christophersen CT, Bird AR, Shepherd SJ, Gibson PR, Muir JG. Diets that differ in their FODMAP content alter the colonic luminal microenvironment. *Gut*. 2015;64:93–100.
 104. Mazzawi T, Hausken T, Gundersen D, El-Salhy M. Effect of dietary management on the gastric endocrine cells in patients with irritable bowel syndrome. *Eur J Clin Nutr*. 2014;69:519–24.
 105. Mazzawi T, Gundersen D, Hausken T, El-Salhy M. Increased gastric chromogranin A cell density following changes to diets of patients with irritable bowel syndrome. *Mol Med Rep*. 2014;10:2322–6.
 106. Mazzawi T, Gundersen D, Hausken T, El-Salhy M. Increased chromogranin A cell density in the large intestine of patients with irritable bowel syndrome after receiving dietary guidance. *Gastroenterol Res Pract*. 2015;ID 269831:8.
 107. Mazzawi T, Hausken T, Gundersen D, El-Salhy M. Normalization of large intestinal endocrine cells following dietary management in patients with irritable bowel syndrome. *Eur J Clin Nutr*. in press 2015.
 108. El-Salhy M, Gilja OH, Gundersen D, Hatlebakk JG, Hausken T. Duodenal chromogranin A cell density as a biomarker for the diagnosis of irritable bowel syndrome. *Gastroenterol Res Pract*. 2014;2014:462856.
 109. El-Salhy M, Gilja OH, Gundersen D, Hatlebakk JG, Hausken T. Endocrine cells in the ileum of patients with irritable bowel syndrome. *World J Gastroenterol*. 2014;20:2383–91.
 110. El-Salhy M, Gilja OH, Gundersen D, Hausken T. Endocrine cells in the oxyntic mucosa of the stomach in patients with irritable bowel syndrome. *World J Gastrointest Endosc*. 2014;6:176–85.
 111. El-Salhy M, Gilja OH, Hatlebakk JG, Hausken T. Stomach antral endocrine cells in patients with irritable bowel syndrome. *Int J Mol Med*. 2014;34:967–74.
 112. El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T. Abnormal rectal endocrine cells in patients with irritable bowel syndrome. *Regul Pept*. 2014;188:60–5.
 113. El-Salhy M, Gundersen D, Ostgaard H, Lomholt-Beck B, Hatlebakk JG, Hausken T. Low densities of serotonin and peptide YY cells in the colon of patients with irritable bowel syndrome. *Dig Dis Sci*. 2012;57:873–8.
 114. El-Salhy M, Hatlebakk JG, Gilja OH, Hausken T. Densities of rectal peptide YY and somatostatin cells as biomarkers for the diagnosis of irritable bowel syndrome. *Peptides*. 2015;67:12–9.
 115. El-Salhy M, Vaali K, Dizdar V, Hausken T. Abnormal small-intestinal endocrine cells in patients with irritable bowel syndrome. *Dig Dis Sci*. 2010;55:3508–13.
 116. El-Salhy M, Wendelbo I, Gundersen D. Serotonin and serotonin transporter in the rectum of patients with irritable bowel disease. *Mol Med Rep*. 2013;8:451–5.
 117. El-Salhy M, Wendelbo IH, Gundersen D. Reduced chromogranin A cell density in the ileum of patients with irritable bowel syndrome. *Mol Med Rep*. 2013;7:1241–4.
 118. Wendelbo I, Mazzawi T, El-Salhy M. Increased serotonin transporter immunoreactivity intensity in the ileum of patients with irritable bowel disease. *Mol Med Rep*. 2014;9:180–4.
 119. Wang SH, Dong L, Luo JY, Gong J, Li L, Lu XL, et al. Decreased expression of serotonin in the jejunum and increased numbers of mast cells in the terminal ileum in patients with irritable bowel syndrome. *World J Gastroenterol*. 2007;13:6041–7.
 120. Park JH, Rhee PL, Kim G, Lee JH, Kim YH, Kim JJ, et al. Enteroendocrine cell counts correlate with visceral hypersensitivity in patients with diarrhoea-predominant irritable bowel syndrome. *Neurogastroenterol Motil*. 2006;18:539–46.
 121. Coates MD, Mahoney CR, Linden DR, Sampson JE, Chen J, Blaszyk H, et al. Molecular defects in mucosal serotonin content and decreased serotonin reuptake transporter in ulcerative colitis and irritable bowel syndrome. *Gastroenterology*. 2004;126:1657–64.
 122. Wang LH, Fang XC, Pan GZ. Bacillary dysentery as a causative factor of irritable bowel syndrome and its pathogenesis. *Gut*. 2004;53:1096–101.
 123. Dizdar V, Spiller R, Singh G, Hanevik K, Gilja OH, El-Salhy M, et al. Relative importance of abnormalities of CCK and 5-HT (serotonin) in Giardia-induced post-infectious irritable bowel syndrome and functional dyspepsia. *Aliment Pharmacol Ther*. 2010;31:883–91.
 124. Coleman NS, Foley S, Dunlop SP, Wheatcroft J, Blackshaw E, Perkins AC, et al. Abnormalities of serotonin metabolism and their relation to symptoms in untreated celiac disease. *Clin Gastroenterol Hepatol*. 2006;4:874–81.
 125. Dunlop SP, Coleman NS, Blackshaw E, Perkins AC, Singh G, Marsden CA, et al. Abnormalities of 5-hydroxytryptamine metabolism in irritable bowel syndrome. *Clin Gastroenterol Hepatol*. 2005;3:349–57.
 126. Dunlop SP, Jenkins D, Neal KR, Spiller RC. Relative importance of enterochromaffin cell hyperplasia, anxiety, and depression in postinfectious IBS. *Gastroenterology*. 2003;125:1651–9.
 127. Lee KJ, Kim YB, Kim JH, Kwon HC, Kim DK, Cho SW. The alteration of enterochromaffin cell, mast cell, and lamina propria T lymphocyte numbers in irritable bowel syndrome and its relationship with psychological factors. *J Gastroenterol Hepatol*. 2008;23:1689–94.
 128. Kim HS, Lim JH, Park H, Lee SI. Increased immunoendocrine cells in intestinal mucosa of postinfectious irritable bowel syndrome patients 3 years after acute Shigella infection—an observation in a small case control study. *Yonsei Med J*. 2010;51:45–51.
 129. Spiller RC, Jenkins D, Thornley JP, Hebden JM, Wright T, Skinner M, et al. Increased rectal mucosal enteroendocrine cells, T lymphocytes, and increased gut permeability following acute Campylobacter enteritis and in post-dysenteric irritable bowel syndrome. *Gut*. 2000;47:804–11.
 130. Gershon MD. 5-Hydroxytryptamine (serotonin) in the gastrointestinal tract. *Curr Opin Endocrinol Diabetes Obes*. 2013;20:14–21.

131. El-Salhy M, Seim I, Chopin L, Gundersen D, Hatlebakk JG, Hausken T. Irritable bowel syndrome: the role of gut neuroendocrine peptides. *Front Biosci (Elite Ed)*. 2012;4:2783–800.
132. Seim I, El-Salhy M, Hausken T, Gundersen D, Chopin L. Ghrelin and the brain-gut axis as a pharmacological target for appetite control. *Curr Pharm Des*. 2012;18:768–75.
133. May CL, Kaestner KH. Gut endocrine cell development. *Mol Cell Endocrinol*. 2010;323:70–5.
134. Gunawardene AR, Corfe BM, Staton CA. Classification and functions of enteroendocrine cells of the lower gastrointestinal tract. *Int J Exp Pathol*. 2011;92:219–31.
135. Camilleri M. Physiological underpinnings of irritable bowel syndrome: neurohormonal mechanisms. *J Physiol*. 2014;592:2967–80.
136. Ford AC, Talley NJ, Spiegel BM, Foxx-Orenstein AE, Schiller L, Quigley EM, et al. Effect of fibre, antispasmodics, and peppermint oil in the treatment of irritable bowel syndrome: systematic review and meta-analysis. *BMJ*. 2008;337:a2313.
137. Francis CY, Whorwell PJ. Bran and irritable bowel syndrome: time for reappraisal. *Lancet*. 1994;344:39–40.
138. Bijkerk CJ, Muris JW, Knottnerus JA, Hoes AW, de Wit NJ. Systematic review: the role of different types of fibre in the treatment of irritable bowel syndrome. *Aliment Pharmacol Ther*. 2004;19:245–51.
139. Bijkerk CJ, de Wit NJ, Muris JW, Whorwell PJ, Knottnerus JA, Hoes AW. Soluble or insoluble fibre in irritable bowel syndrome in primary care? Randomised placebo Controlled Trial *BMJ*. 2009;339:b3154.
140. Kubo M, Fujiwara Y, Shiba M, Kohata Y, Yamagami H, Tanigawa T, et al. Differences between risk factors among irritable bowel syndrome subtypes in Japanese adults. *Neurogastroenterol Motil*. 2011;23:249–54.
141. Pickett-Blakely O. Obesity and irritable bowel syndrome: a comprehensive review. *Gastroenterologist Hepatology*. 2014;10:411–216.
142. Chaudhri O, Small C, Bloom S. Gastrointestinal hormones regulating appetite. *Philos Trans R Soc Lond B Biol Sci*. 2006;361:1187–209.
143. Peruzzo B, Pastor FE, Blazquez JL, Schobitz K, Pelaez B, Amat P, et al. A second look at the barriers of the medial basal hypothalamus. *Exp Brain Res*. 2000;132:10–26.
144. Cone RD, Cowley MA, Butler AA, Fan W, Marks DL, Low MJ. The arcuate nucleus as a conduit for diverse signals relevant to energy homeostasis. *Int J Obes Relat Metab Disord*. 2001;25 Suppl 5:S63–7.
145. Yu JH, Kim MS. Molecular mechanisms of appetite regulation. *Diabetes Metab J*. 2012;36:391–8.
146. El-Salhy M. Ghrelin in gastrointestinal diseases and disorders: a possible role in the pathophysiology and clinical implications (review). *Int J Mol Med*. 2009;24:727–32.
147. El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T. Abnormal endocrine cells in the ileum of patients with irritable bowel syndrome. *World J Gastroenterol*. 2013;20:2383–91.
148. El-Salhy M, Lillebo E, Reinemo A, Salmelid L. Ghrelin in patients with irritable bowel syndrome. *Int J Mol Med*. 2009;23:703–7.
149. El-Salhy M, Mazzawi T, Gundersen D, Hatlebakk JG, Hausken T. The role of peptide YY in gastrointestinal diseases and disorders (Review). *Int J Mol Med*. 2013;31:275–82.
150. Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H, Kangawa K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature*. 1999;402:656–60.
151. Kojima M, Hosoda H, Kangawa K. Purification and distribution of ghrelin: the natural endogenous ligand for the growth hormone secretagogue receptor. *Horm Res*. 2001;56 Suppl 1:93–7.
152. Kojima M, Hosoda H, Matsuo H, Kangawa K. Ghrelin: discovery of the natural endogenous ligand for the growth hormone secretagogue receptor. *Trends Endocrinol Metab*. 2001;12:118–22.
153. Date Y, Kojima M, Hosoda H, Sawaguchi A, Mondal MS, Suganuma T, et al. Ghrelin, a novel growth hormone-releasing acylated peptide, is synthesized in a distinct endocrine cell type in the gastrointestinal tracts of rats and humans. *Endocrinology*. 2000;141:4255–61.
154. Masuda Y, Tanaka T, Inomata N, Ohnuma N, Tanaka S, Itoh Z, et al. Ghrelin stimulates gastric acid secretion and motility in rats. *Biochem Biophys Res Commun*. 2000;276:905–8.
155. Fujino K, Inui A, Asakawa A, Kihara N, Fujimura M, Fujimiya M. Ghrelin induces fasted motor activity of the gastrointestinal tract in conscious fed rats. *J Physiol*. 2003;550:227–40.
156. Asakawa A, Ataka K, Fujino K, Chen CY, Kato I, Fujimiya M, et al. Ghrelin family of peptides and gut motility. *J Gastroenterol Hepatol*. 2011;26 Suppl 3:73–4.
157. Dornon ville de laCour C, Lindstrom E, Norlen P, Hakanson R. Ghrelin stimulates gastric emptying but is without effect on acid secretion and gastric endocrine cells. *Regul Pept*. 2004;120:23–32.
158. Fukuda H, Mizuta Y, Isomoto H, Takeshima F, Ohnita K, Ohba K, et al. Ghrelin enhances gastric motility through direct stimulation of intrinsic neural pathways and capsaicin-sensitive afferent neurones in rats. *Scand J Gastroenterol*. 2004;39:1209–14.
159. Edholm T, Levin F, Hellstrom PM, Schmidt PT. Ghrelin stimulates motility in the small intestine of rats through intrinsic cholinergic neurons. *Regul Pept*. 2004;121:25–30.
160. Levin F, Edholm T, Schmidt PT, Gryback P, Jacobsson H, Degerblad M, et al. Ghrelin stimulates gastric emptying and hunger in normal-weight humans. *J Clin Endocrinol Metab*. 2006;91:3296–302.
161. Tack J, Depoortere I, Bisschops R, Delpoort C, Coulie B, Meulemans A, et al. Influence of ghrelin on interdigestive gastrointestinal motility in humans. *Gut*. 2006;55:327–33.
162. Ariga H, Tsukamoto K, Chen C, Mantyh C, Pappas TN, Takahashi T. Endogenous acyl ghrelin is involved in mediating spontaneous phase III-like contractions of the rat stomach. *Neurogastroenterol Motil*. 2007;19:675–80.
163. Tumer C, Oflazoglu HD, Obay BD, Kelle M, Tasdemir E. Effect of ghrelin on gastric myoelectric activity and gastric emptying in rats. *Regul Pept*. 2008;146:26–32.
164. Tebbe JJ, Mronga S, Tebbe CG, Ortmann E, Arnold R, Schafer MK. Ghrelin-induced stimulation of colonic propulsion is dependent on hypothalamic neuropeptide Y1- and corticotrophin-releasing factor 1 receptor activation. *J Neuroendocrinol*. 2005;17:570–6.
165. Fujimiya M, Ataka K, Asakawa A, Chen CY, Kato I, Inui A. Regulation of gastroduodenal motility: acyl ghrelin, des-acyl ghrelin and obestatin and hypothalamic peptides. *Digestion*. 2012;85:90–4.
166. Ogiso K, Asakawa A, Amitani H, Inui A. Ghrelin: a gut hormonal basis of motility regulation and functional dyspepsia. *J Gastroenterol Hepatol*. 2011;26 Suppl 3:67–72.
167. Corwin RL, Gibbs J, Smith GP. Increased food intake after type A but not type B cholecystokinin receptor blockade. *Physiol Behav*. 1991;50:255–8.
168. Gibbs J, Falasco JD, McHugh PR. Cholecystokinin-decreased food intake in rhesus monkeys. *Am J Physiol*. 1976;230:15–8.
169. Gibbs J, Smith GP. Cholecystokinin and satiety in rats and rhesus monkeys. *Am J Clin Nutr*. 1977;30:758–61.
170. Gibbs J, Smith GP. Satiety: the roles of peptides from the stomach and the intestine. *Fed Proc*. 1986;45:1391–5.
171. Smith GP, Falasco J, Moran TH, Joyner KM, Gibbs J. CCK-8 decreases food intake and gastric emptying after pylorotomy or pyloroplasty. *Am J Physiol*. 1988;255:R113–6.
172. Smith GP, Gibbs J. Role of CCK in satiety and appetite control. *Clin Neuropharmacol*. 1992;15(1):Pt A:476A.
173. Smith GP, Gibbs J. Gut peptides and postprandial satiety. *Fed Proc*. 1984;43:2889–92.
174. Smith GP, Gibbs J. Cholecystokinin: a putative satiety signal. *Pharmacol Biochem Behav*. 1975;3:135–8.
175. Smith GP, Tyrka A, Gibbs J. Type-A CCK receptors mediate the inhibition of food intake and activity by CCK-8 in 9- to 12-day-old rat pups. *Pharmacol Biochem Behav*. 1991;38:207–10.
176. Stallone D, Nicolaidis S, Gibbs J. Cholecystokinin-induced anorexia depends on serotonergic function. *Am J Physiol*. 1989;256:R1138–41.
177. Weller A, Smith GP, Gibbs J. Endogenous cholecystokinin reduces feeding in young rats. *Science*. 1990;247:1589–91.
178. Woods SC, Gibbs J. The regulation of food intake by peptides. *Ann N Y Acad Sci*. 1989;575:236–43.
179. Huppi K, Siwarski D, Pisegna JR, Wank S. Chromosomal localization of the gastric and brain receptors for cholecystokinin (CCKAR and CCKBR) in human and mouse. *Genomics*. 1995;25:727–9.
180. Silvente-Poirot S, Wank SA. A segment of five amino acids in the second extracellular loop of the cholecystokinin-B receptor is essential for selectivity of the peptide agonist gastrin. *J Biol Chem*. 1996;271:14698–706.
181. Tarasova NI, Stauber RH, Choi JK, Hudson EA, Czerwinski G, Miller JL, et al. Visualization of G protein-coupled receptor trafficking with the aid of the

- green fluorescent protein. Endocytosis and recycling of cholecystokinin receptor type A. *J Biol Chem*. 1997;272:14817–24.
182. Tarasova NI, Wank SA, Hudson EA, Romanov VI, Czerwinski G, Resau JH, et al. Endocytosis of gastrin in cancer cells expressing gastrin/CCK-B receptor. *Cell Tissue Res*. 1997;287:325–33.
 183. Wank SA. Cholecystokinin receptors. *Am J Physiol*. 1995;269:G628–46.
 184. Batterham RL, Cohen MA, Ellis SM, Le Roux CW, Withers DJ, Frost GS, et al. Inhibition of food intake in obese subjects by peptide YY3-36. *N Engl J Med*. 2003;349:941–8.
 185. Batterham RL, Cowley MA, Small CJ, Herzog H, Cohen MA, Dakin CL, et al. Gut hormone PYY(3-36) physiologically inhibits food intake. *Nature*. 2002;418:650–4.
 186. Michel MC, Beck-Sickinger A, Cox H, Doods HN, Herzog H, Larhammar D, et al. XVI. International Union of Pharmacology recommendations for the nomenclature of neuropeptide Y, peptide YY, and pancreatic polypeptide receptors. *Pharmacol Rev*. 1998;50:143–50.
 187. Lin HC, Zhao XT, Wang L. Intestinal transit is more potently inhibited by fat in the distal (ileal brake) than in the proximal (jejunal brake) gut. *Dig Dis Sci*. 1997;42:19–25.
 188. Lin HC, Zhao XT, Wang L. Jejunal brake: inhibition of intestinal transit by fat in the proximal small intestine. *Dig Dis Sci*. 1996;41:326–9.
 189. Van Citters GW, Lin HC. The ileal brake: a fifteen-year progress report. *Curr Gastroenterol Rep*. 1999;1:404–9.
 190. Ohtani N, Sasaki I, Naito H, Shibata C, Matsuno S. Mediators for fat-induced ileal brake are different between stomach and proximal small intestine in conscious dogs. *J Gastrointest Surg*. 2001;5:377–82.
 191. Pironi L, Stanghellini V, Miglioli M, Corinaldesi R, De Giorgio R, Ruggeri E, et al. Fat-induced ileal brake in humans: a dose-dependent phenomenon correlated to the plasma levels of peptide YY. *Gastroenterology*. 1993;105:733–9.
 192. Maljaars J, Peters HP, Masclee AM. Review article: The gastrointestinal tract: neuroendocrine regulation of satiety and food intake. *Aliment Pharmacol Ther*. 2007;26 Suppl 2:241–50.
 193. Maljaars PW, Peters HP, Mela DJ, Masclee AA. Ileal brake: a sensible food target for appetite control. *A Rev Physiol Behav*. 2008;95:271–81.
 194. Maljaars PW, Symersky T, Kee BC, Haddeman E, Peters HP, Masclee AA. Effect of ileal fat perfusion on satiety and hormone release in healthy volunteers. *Int J Obes (Lond)*. 2008;32:1633–9.
 195. Dubrasquet M, Bataille D, Gespach C. Oxyntomodulin (glucagon-37 or bioactive enteroglucagon): a potent inhibitor of pentagastrin-stimulated acid secretion in rats. *Biosci Rep*. 1982;2:391–5.
 196. Dubrasquet M, Roze C, Ling N, Florencio H. Inhibition of gastric and pancreatic secretions by cerebroventricular injections of gastrin-releasing peptide and bombesin in rats. *Regul Pept*. 1982;3:105–12.
 197. Schjoldager BT, Baldissera FG, Mortensen PE, Holst JJ, Christiansen J. Oxyntomodulin: a potential hormone from the distal gut. Pharmacokinetics and effects on gastric acid and insulin secretion in man. *Eur J Clin Invest*. 1988;18:499–503.
 198. Schjoldager B, Mortensen PE, Myhre J, Christiansen J, Holst JJ. Oxyntomodulin from distal gut. Role in regulation of gastric and pancreatic functions. *Dig Dis Sci*. 1989;34:1411–9.
 199. Dakin CL, Small CJ, Batterham RL, Neary NM, Cohen MA, Patterson M, et al. Peripheral oxyntomodulin reduces food intake and body weight gain in rats. *Endocrinology*. 2004;145:2687–95.
 200. Wynne K, Park AJ, Small CJ, Patterson M, Ellis SM, Murphy KG, et al. Subcutaneous oxyntomodulin reduces body weight in overweight and obese subjects: a double-blind, randomized, controlled trial. *Diabetes*. 2005;54:2390–5.
 201. Le Quellec A, Kervran A, Blache P, Ciurana AJ, Bataille D. Oxyntomodulin-like immunoreactivity: diurnal profile of a new potential enterogastrone. *J Clin Endocrinol Metab*. 1992;74:1405–9.
 202. Ghatei MA, Uttenthal LO, Christofides ND, Bryant MG, Bloom SR. Molecular forms of human enteroglucagon in tissue and plasma: plasma responses to nutrient stimuli in health and in disorders of the upper gastrointestinal tract. *J Clin Endocrinol Metab*. 1983;57:488–95.
 203. Fang XL, Shu G, Yu JJ, Wang LN, Yang J, Zeng QJ, et al. The Anorexigenic Effect of Serotonin Is Mediated by the Generation of NADPH Oxidase-Dependent ROS. *PLoS One*. 2013;8, e53142.
 204. El-Salhy M, Lilbo E, Reinemo A, Salmøed L, Hausken T. Effects of a health program comprising reassurance, diet management, probiotic administration and regular exercise on symptoms and quality of life in patients with irritable bowel syndrome. *Gastroenterology Insights*. 2010;2:21–6.
 205. Shewry PR. Wheat. *J Exp Bot*. 2009;60:1537–53.
 206. Pattison AL, Appelbee M, Trethowan RM. Characteristics of modern triticale quality: glutenin and secalin subunit composition and mixograph properties. *J Agric Food Chem*. 2014;62:4924–31.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

