

Review Article

Oxidative Stress in Neurodegenerative Diseases: From Molecular Mechanisms to Clinical Applications

Zewen Liu,^{1,2} Tingyang Zhou,^{1,3} Alexander C. Ziegler,⁴ Peter Dimitrion,⁴ and Li Zuo^{1,3}

¹Radiologic Sciences and Respiratory Therapy Division, School of Health and Rehabilitation Sciences, The Ohio State University College of Medicine, The Ohio State University Wexner Medical Center, Columbus, OH 43210, USA

²Department of Anesthesiology, Affiliated Ezhou Central Hospital, Wuhan University, Ezhou 436000, China

³Interdisciplinary Biophysics Program, The Ohio State University, Columbus, OH 43210, USA

⁴Department of Biotechnology, Johns Hopkins University, Baltimore, MA 21218, USA

Correspondence should be addressed to Li Zuo; zuo.4@osu.edu

Received 29 January 2017; Revised 26 May 2017; Accepted 7 June 2017; Published 12 July 2017

Academic Editor: James Duce

Copyright © 2017 Zewen Liu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Increasing numbers of individuals, particularly the elderly, suffer from neurodegenerative disorders. These diseases are normally characterized by progressive loss of neuron cells and compromised motor or cognitive function. Previous studies have proposed that the overproduction of reactive oxygen species (ROS) may have complex roles in promoting the disease development. Research has shown that neuron cells are particularly vulnerable to oxidative damage due to their high polyunsaturated fatty acid content in membranes, high oxygen consumption, and weak antioxidant defense. However, the exact molecular pathogenesis of neurodegeneration related to the disturbance of redox balance remains unclear. Novel antioxidants have shown great potential in mediating disease phenotypes and could be an area of interest for further research. In this review, we provide an updated discussion on the roles of ROS in the pathological mechanisms of Alzheimer's disease, Huntington's disease, Parkinson's disease, amyotrophic lateral sclerosis, and spinocerebellar ataxia, as well as a highlight on the antioxidant-based therapies for alleviating disease severity.

1. Introduction

Neurodegenerative diseases are characterized by progressive damage in neural cells and neuronal loss, which lead to compromised motor or cognitive function. Common neurodegenerative diseases include Alzheimer's disease (AD), Parkinson's disease (PD), Huntington's disease (HD), amyotrophic lateral sclerosis (ALS), and spinocerebellar ataxia (SCA) [1–4]. These diseases represent a primary health problem especially in the aging population [3, 5]. For example, AD ranks as the sixth leading causes of death in the United States. PD, the second most prevalent neurodegenerative disease, affects 1 to 2% of the population above age of 65 [2, 6–9]. Reactive oxygen species (ROS) are chemically reactive molecules that have been implicated in the pathogenesis of neurodegenerative diseases. They are naturally generated within the biological system, playing important roles in mediating cellular activities such as inflammation,

cell survival, and stressor responses as well as many diseases including cardiovascular disorders, muscle dysfunction, allergy, and cancers [10–13]. Due to their reactivity, high concentrations of ROS can lead to cell death or oxidative stress (OS), which is defined as the disruption of balance between pro-oxidant and antioxidant levels [11]. The complex pathogenesis of the neurodegenerative diseases remain largely unknown; however, mounting evidence suggests that ROS may play a critical role as high levels of OS are commonly observed in the brain of patients with neurodegenerative conditions [3, 14].

Numerous studies have been performed to investigate the roles of ROS in neurodegenerative progression and some positive results are yielded [15–17]. Although ROS may not be the triggering factor for neurodegenerative diseases, they are likely to exacerbate disease progression through oxidative damage and interaction with mitochondria (Figure 1) [14]. Of note is that neuron cells are particularly vulnerable to

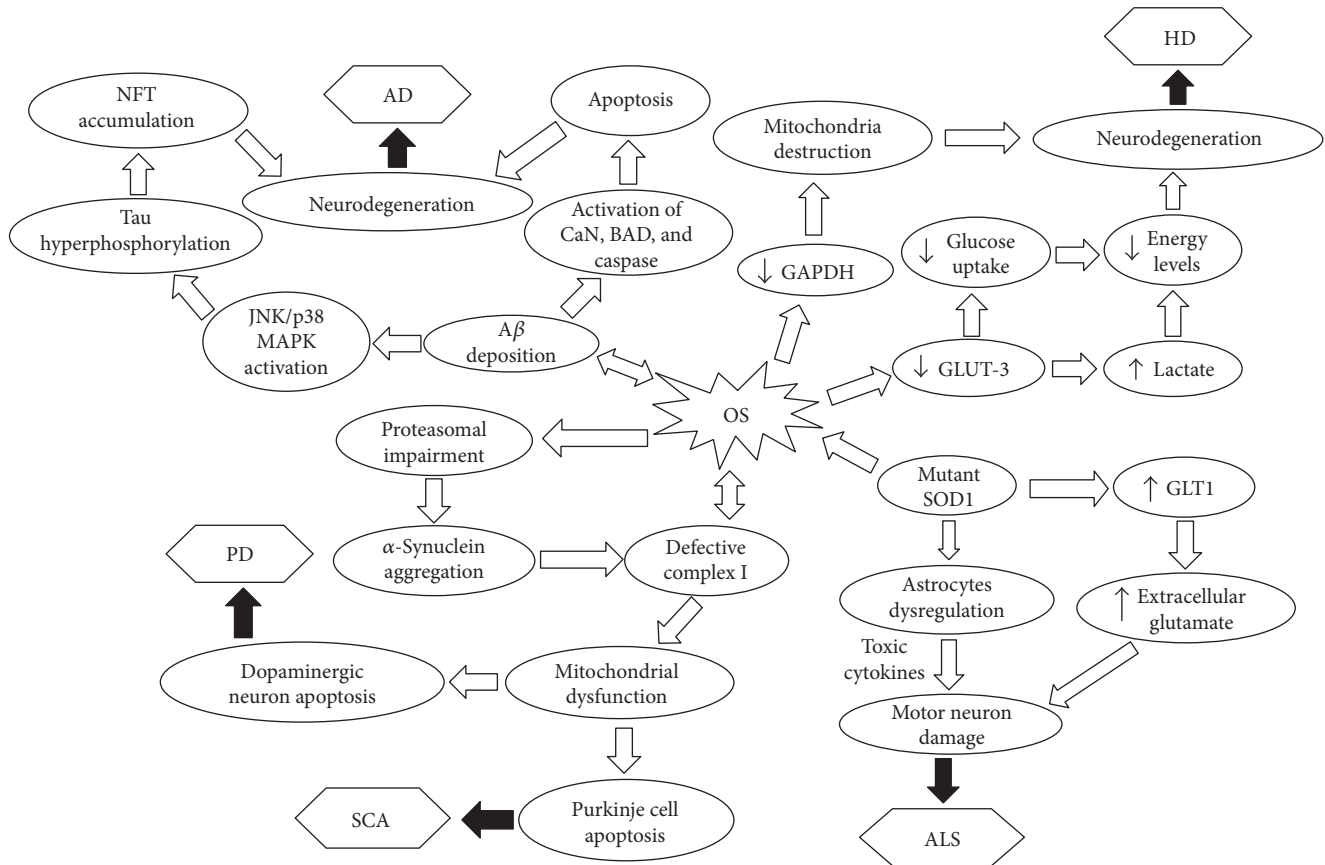


FIGURE 1: Schematic illustrating the key roles of OS in the development of AD, HD, PD, ALS, and SCA. AD: Alzheimer's disease; ALS: amyotrophic lateral sclerosis; $A\beta$: amyloid beta; BAD: Bcl-2-associated death promoter; CaN: calcineurin; GAPDH: glyceraldehyde-3-phosphate dehydrogenase; HD: Huntington's disease; JNK: c-Jun N-terminal kinase; MAPK: mitogen-activated protein kinase; NFT: neurofibrillary tangle; OS: oxidative stress; PD: Parkinson's disease; SCA: spinocerebellar ataxia; SOD: superoxide dismutase.

oxidative damage because of their high polyunsaturated fatty acid content in membranes, high oxygen consumption, and weak antioxidant defense [17]. Under physiological conditions, ROS generated from mitochondria, NADPH oxidase (Nox), and xanthine oxidase (XO), are maintained at relatively low levels by endogenous antioxidants [11]. However, the redox balance can be disturbed by neural inflammation or abnormal mitochondrial function [17, 18]. The pathogenesis of several neurodegenerative disorders such as AD and PD is associated with the accumulation of misfolded proteins. The aggregation of these modified proteins can in turn trigger inflammatory response in the brain, which induces marked ROS release and subsequent OS [11, 19]. Mitochondrial dysfunction, which often accompanies aberrant ROS production, is tightly linked with neurodegenerative disorders [3, 20–22]. For example, in HD, mutant huntingtin (mHTT) may directly interact with mitochondria, leading to compromised energy supply and increased ROS levels [23].

Considering the pivotal roles of OS in neurodegenerative diseases, the manipulation of ROS levels may represent a promising treatment option to slow down neurodegeneration and alleviate associated symptoms. In this regard, many compounds that possess antioxidant properties including glutathione (GSH), vitamin C, vitamin E, and coenzyme Q10 have been examined for their potential to attenuate

neurodegenerative symptoms; however, mixed results are yielded. For instance, treatment with vitamin E at a dose of 2000 IU per day for two years has been shown to attenuate the functional decline in patients with moderate AD [24]. Vitamin E supplementation early in life potentially decreases the risk of PD [25]. However, another study by Pappert et al. indicated that oral intake of vitamin E for five months had no effect on the elevation of vitamin E levels in ventricular cerebrospinal fluid in PD patients [26]. This suggests the drawbacks of oral administration of antioxidants associated with limited entry across blood–brain barrier and rapid antioxidant metabolism [26]. To counteract this, some small antioxidant molecules, such as ferrostatin-1 (Fer-1), have been developed, which show efficacy in attenuating neurodegenerative symptoms [27, 28]. Furthermore, the time, the method, and the dose of antioxidant administration as well as the stage of the disease are all key factors that should be taken into account when investigating the effects of antioxidant therapies in future studies [29, 30]. Current understanding of the molecular pathogenesis of neurodegeneration is incomplete. Future studies that explore the implications of ROS in various neurodegenerative diseases may be critical to discover novel and reliable therapies.

Currently, no updated systemic review has been performed that specifically focuses on the critical role of ROS

and oxidative damage in major neurodegenerative diseases including AD, PD, HD, ALS, and SCA. Thus, there is an urgent need for such comprehensive review. In this article, we have thoroughly discussed the most recent and novel advance in the field of neurodegenerative diseases. Particularly, we highlighted the molecular mechanisms of ROS and the importance of antioxidant therapy in these important neuronal diseases. These critical integrations of various neurodegenerative disease models may shed light into the discovery of novel therapy in the future.

2. Oxidative Stress and Alzheimer's Disease (AD)

AD, the most prevalent neurodegenerative disorder, is characterized by the progressive deterioration of behavior, cognition and functionality, which significantly impairs daily living activities [31]. The pathophysiology of AD is mainly associated with the extracellular deposition of amyloid beta ($A\beta$) plaques and the accumulation of intracellular tau neurofibrillary tangles (NFT) [32, 33]. $A\beta$ plaques can deplete calcium ions (Ca^{2+}) storage in endoplasmic reticulum (ER), resulting in cytosolic Ca^{2+} overload. In response to cytosolic Ca^{2+} increase, endogenous GSH levels are reduced and ROS can be overaccumulated inside the cells [34]. ROS-induced OS is emerging as an important factor in pathogenesis of AD as ROS overproduction is thought to play a critical role in the accumulation and deposition of $A\beta$ in AD (Figure 1) [35]. Mitochondrial dysfunction can lead to ROS misregulation, reduced production of adenosine triphosphate (ATP), altered Ca^{2+} homeostasis, and excitotoxicity. All these alterations may be implicated in the progression of AD [36]. The severe OS observed in AD patients could be a result of overactivating N-methyl-D-aspartate-type glutamate receptors (NMDARs). NMDAR activation has been shown to lead to excessive influx of Ca^{2+} by promoting cell permeability and subsequent generation of neurotoxic levels of ROS/reactive nitrogen species (RNS) [37, 38]. ROS potentially play a role in mediating JNK/stress-activated protein kinase pathways. The activation of these cascades has been implicated in the hyperphosphorylation of tau proteins and $A\beta$ -induced cell death [39]. In addition, $A\beta$ proteins can directly initiate free radical formation via the activation of NADPH oxidase [40]. $A\beta$ -induced ROS overproduction modifies cellular signaling pathways and initiates tau hyperphosphorylation via the activation of p38 mitogen-activated protein kinase (MAPK). An abnormal accumulation of hyperphosphorylated tau proteins can lead to intracellular NFT formation (Figure 1) [41, 42]. Furthermore, $A\beta$ has been shown to play an important role in mediating cellular apoptotic cascades [43]. Specifically, $A\beta$ can enhance the activity of calcineurin, which in turn triggers Bcl-2-associated death promoter, leading to cytochrome *c* release from mitochondria [44]. $A\beta$ may also directly associate with caspases, and eventually induce neuron apoptosis (Figure 1) [44].

Aging, inflammation, environmental stress, and some nutritional factors (e.g., redox-active metals) can induce additional OS leading to an increased $A\beta$ production [45–48]. Elderly individuals are more prone to OS, which

partially accounts for AD susceptibility in aging populations [45, 49]. Inflammation is responsible for increased expression of cytokines, ROS levels, and cellular toxicity, thereby exacerbating AD progression [50, 51]. $A\beta$ deposition results in microglial activation [52]. It is becoming increasingly evident that prolonged activation of microglia leads to the release of proinflammatory cytokines, initiating a proinflammatory cascade and subsequently contributing to neuronal damage and loss [53]. OS can be induced by environmental factors such as pollutants, chemicals, and radiation [47, 54]. For instance, ROS formation increases when excess iron deposits are present [54]. $A\beta$ itself can interact with metal ions to generate free radicals, and methionine 35 plays a critical role in these reactions [54, 55]. Interestingly, Cu^{2+}/Zn^{2+} -bound $A\beta$ has been shown to possess a structure similar to superoxide dismutase (SOD), with potential antioxidant properties [56]. Therefore, the supplementation of Cu^{2+} and Zn^{2+} has been suggested as a novel strategy to decrease $A\beta$ -induced ROS generation and metal-catalyzed $A\beta$ deposition [56].

Therapeutic approaches for AD are focused on decreasing $A\beta$ oligomers and phosphorylated tau levels, reducing OS, and controlling epigenetic changes [57, 58]. AD treatments mostly rely on compounds that have neuroprotective, anti-inflammatory, and antioxidant properties [59]. Medications (e.g., rutin, resveratrol, and vitamin E) that target ROS-mediated cascades such as JNK and NF- κ B have produced some positive results both in vitro and in vivo [31]. Important factors such as reaction kinetics and bioavailability (absorption, retention in the targeted area, distribution, and transport) must be considered when using antioxidants [59]. Several ROS-related neuroprotective therapeutic strategies continue to show great potential in treating AD. For instance, the nuclear factor erythroid 2-related factor 2 (Nrf2)-mediated antioxidant response element (ARE) pathway is known as an essential protective mechanism against OS [60]. The binding of Nrf2 to ARE activates the expression of several antioxidant genes in a synchronized manner that can work together for oxidative detoxification. Weakened Nrf2-ARE pathways were observed in the brains of transgenic mice with AD symptoms, while the enhancement of Nrf2-ARE cascades using adenoviral Nrf2 gene transfer has shown protective effects against the toxicity of $A\beta$ deposition [60]. Therefore, the transcriptional manipulation of the endogenous antioxidants may hold great promises in relieving AD symptoms [60]. In addition, traditional Chinese medicine and Ayurvedic treatments, metal ion chelators, and histone deacetylase inhibitors have also demonstrated potentials in AD treatment [61–64].

3. Huntington's Disease (HD) and Reactive Oxygen Species

HD is a progressive neurodegenerative disease linked with unstable expansion of cytosine, adenine, guanine (CAG) repeats in the HTT gene [65, 66]. The expansion of CAG repeats within the exon1 of the HTT gene gives rise to a mutation that leads to the elongation of polyglutamine tract, resulting in a HTT protein product that is susceptible to

aggregation [65]. The mHTT aggregates are accumulated throughout the brain of the affected individuals, which can interrupt protein quality control and transcription process. Those alterations are potentially responsible for the aberrant motor and cognitive problems in HD [65]. Existing HD treatments help to suppress the severity of symptoms; however, no treatment directly cures or significantly halts the disease progression [65, 66]. mHTT is a well-studied mutant of HD that contributes toward the buildups of cytoplasmic plaque and neuronal nuclear inclusion in HD [67–69]. mHTT has been shown to reduce the expression of peroxisome proliferator-activated receptor- γ coactivator-1 α , altering the level of antioxidant enzymes and lowering the concentration of striatal mitochondria [66, 70, 71]. Interestingly, despite the established connection of the role of OS in HD, trials attempting to treat the disease using classic antioxidants have largely been ineffective [72].

The effect of HD on energy levels in the brain has been an area of focus for researchers. Initial studies support the theory that HD lowers energy levels, as reduced glucose usage and increased lactate levels are observed in HD patients [67, 73]. Recent research shows that oxidative damage is related to decreased expression of glucose transporter (GLUT)-3, resulting in inhibited glucose uptake and accumulation of lactate (Figure 1) [73, 74]. The majority of ATP synthesis occurs through the generation of the proton motive force during operation of the electron transport chain [75]. mHtt has been shown to play a key role in mitochondrial dysfunction. Under electron microscopy, Panov et al. identified that the N-terminal of mHtt can interact with mitochondrial membranes, causing mitochondrial calcium abnormalities. In addition, mHtt has direct inhibitory effects on respiratory complex II [76, 77]. The inhibition of the mitochondrial electron transport can result in higher levels of ROS, as well as decreased ATP production [20, 77]. In 2015, a new mechanism underlying mitochondrial damage in HD was proposed, which suggested that OS could inactivate the catalytic activity of glyceraldehyde-3-phosphate dehydrogenase (GAPDH). The inactive GAPDH (iGAPDH) associates with damaged mitochondria and acts as a signaling molecule in order to drive the impaired mitochondria towards selective degradation via lysosome engulfment. Nevertheless, in the presence of mHtt, iGAPDH can abnormally interact with the long polyglutamine of mHTT at the mitochondrial outer membrane, which blocks the iGAPDH signaling for degradation. Consequently, damaged mitochondria cannot be engulfed by lysosome and unfavorably accumulate in the mHtt-expressed cells, leading to cell death (Figure 1) [78]. Both mitochondrial alterations and ROS can promote positive feedback loops, resulting in more OS and neuronal loss in the striatum and cortex [66]. Although the involvement of mitochondrial alterations and ROS overproduction in HD progression is well evident, it is not clear which event occurs initially [66, 79, 80].

Numerous studies illustrate the connection between increased levels of oxidative markers and irreversible neuronal damage [65, 81, 82]. One study aimed to establish the merits of neurorehabilitation exercise by tracking the concentrations of 8-hydroxy-2-deoxyguanosine (8-OHdG) and

neuron-specific enolase (NSE), both of which are known biomarkers of oxidative damage in HD [83]. In addition, Cu/Zn-SOD (SOD1) was indicated as a potential peripheral biomarker of oxidative damage in neurons and the level was significantly increased in HD patients compared to control, representing a compensatory response to increased oxidative levels in HD [83]. However, whether SOD1 can be considered as an oxidative biomarker in HD remains undetermined because mixed results have been yielded regarding the concentrations and activities of SOD in HD [72, 84]. Upon completion of the neurorehabilitation program (an exercise regimen lasts for three weeks), markedly decreased levels of NSE and 8-OHdG were observed while levels of SOD1 remained high, suggesting the possible neuroprotective role of SOD1 in scavenging free radicals [83]. Overall, physical exercise is recommended for individuals with HD as it can potentially improve redox homeostasis and help to combat disease progression [83, 85, 86].

Other commonly used oxidative biomarkers in HD models include protein carbonyls, thiobarbituric acid reactive substances (TBARS), and 3-nitrotyrosine [72, 87, 88]. In addition, increased F₂-isoprostane (F₂-IsoP) levels have been observed in the brain tissue and cerebrospinal fluid of several neurodegenerative disorders including HD and AD. Therefore, the measurement of F₂-IsoPs provides a potential approach to evaluate the significance of OS in HD patients. Of note, in the early stage of HD progression, there might be an overlap of F₂-IsoP levels between the HD and control groups [89]. Therefore, the alterations of oxidative biomarkers should be interpreted with caution because OS can also be correlated with other particular factors such as aging or pathological conditions. Moreover, levels of oxidative modifications may not provide sufficient information on whether the oxidative modifications play a causal role in disease progression or directly resulted from cell death [11, 72, 89, 90]. Despite the presence of such limitations, progress has been made in understanding the relationship between OS and HD [72]. The use of robust biomarkers will be crucial to help clarify the contribution of OS in neurodegenerative diseases and to improve the treatment therapies for HD.

4. Oxidative Stress and Parkinson's Disease (PD)

PD is the second most prevalent neurodegenerative disease characterized by dopaminergic neuron loss in the substantia nigra pars compacta of the brain [7, 91, 92]. About 1-2% of the population over 65 is affected by PD and this rate increases to 4% in individuals above 85 years of age [6–8]. The pathological mechanisms underlying the degeneration of dopaminergic neurons have been correlated to overaccumulation of ROS or other free radicals. As mentioned previously, excessive ROS production can be caused by mitochondrial dysfunction or inflammation [14]. The maintenance of redox homeostasis is critical for the proper function of redox-sensitive signaling proteins in neuron cells as well as neuronal survival [93, 94]. In the brain, the primary sites of ROS generation include mitochondria in the neurons and glia [14]. The production of these free

radicals is exacerbated in PD due to neuroinflammation, dopamine degradation, mitochondrial dysfunction, aging, GSH depletion, and high levels of iron or Ca^{2+} [14, 95]. Furthermore, ROS accumulation in PD may be worsened when the individuals are exposed to environmental factors such as pesticides, neurotoxins, and dopamine [96]. This is evidenced by the significant association between increased risk of PD occurrence and pesticides exposure [96]. ROS have been shown to contribute significantly to dopaminergic neuronal loss (Figure 1) [14, 95]. Other studies suggested that the loss of dopaminergic neurons could also be associated with the presence of neuromelanin since highly pigmented neurons are more susceptible to damages [97]. The formation of neuromelanin appears to be related to dopamine auto-oxidation, a process induced by ROS overproduction [97].

Neurodegeneration-induced ROS can damage key cellular proteins and disrupt lipid membranes, promoting OS. Mitochondrial dysfunction leads to increased free radical production in the respiratory chain [14]. Particularly, mitochondrial complex I deficiency has been identified to strongly associate with PD. Indeed, a large amount of the unfavorable neural apoptosis observed in PD can be attributed to the complex I defect [98, 99]. This defect is correlated with the mutation of PTEN-induced putative kinase 1 (PINK1). PINK1 protein is universally expressed in human tissues and plays a critical role in fighting against OS and maintaining mitochondrial membrane potential [98, 100]. The mutation of PINK1 is related to PD onset [98]. In addition to PINK1, other mutations have been shown to implicate in the development of PD, such as DJ-1, parkin, α -synuclein, and leucine-rich repeat kinase 2 (LRRK2). These mutations can potentially impair mitochondrial function, leading to aggravation of ROS generation and increased susceptibility to OS. For instance, mutated parkin may contribute to the pathogenesis of autosomal recessive PD due to its critical roles in suppressing ROS and preventing the formation of neurotoxic proteins caused by ubiquitination [98, 101]. Furthermore, the aggregation of α -synuclein has been shown to interrupt mitochondrial complex I activities, thus resulting in compromised ATP synthesis and mitochondrial dysfunction [102]. Several mechanisms may account for the accumulation of α -synuclein in PD including decreased efficiency of protein degradation caused by proteasomal impairment as well as translation and posttranslation-associated protein overexpression [102]. Proteasomal impairment, which is partially caused by dopamine-derived ROS, has been shown to play a critical role in the neurodegeneration in PD (Figure 1) [102].

Even though there are currently no effective means for curing PD, the understanding of ROS-related mechanisms in disease progression provides important insight to possible treatments that alleviate PD symptoms. Various neuroprotective strategies have been identified to diminish mitochondrial OS within dopaminergic neurons. Fruits and antioxidants are known to attenuate the damaging effects of free radicals [11, 103]. Vitamins C, E, and GSH are essential antioxidants that can be recycled by antioxidant lipoic acid (LA). LA protects neurons against OS and

OS-induced mitochondrial dysfunction by GSH generation and stimulation of lipid peroxide depletion [98, 104]. It has been shown that treatment with LA leads to neuroprotection in an animal study, by improving ATP efficiency and motor coordination [105]. Specifically, rotenone-parkinsonian rats administered with LA showed reduced lipid peroxide in the brain and exhibited improved motor performance [105]. Substances with antioxidant properties such as coenzyme Q10, vitamin C, tocopherol (vitamin E), docosahexaenoic acid (DHA), *Ginkgo biloba*, and polyphenols have been tested but have not demonstrated legitimate evidence of any efficacy in neuroprotectivity [14, 106–109]. Failures from those antioxidant treatments should provide future guidance for using combination therapies targeting mitochondrial function enhancement and blocking ROS generation to PD patients [110].

5. Molecular Mechanisms of ROS in Amyotrophic Lateral Sclerosis

Amyotrophic lateral sclerosis (ALS) is characterized by progressive loss of motor neurons in the anterior horn of the spinal cord [111, 112]. It is classified as either familial or sporadic depending on whether there is a clearly defined, inherited genetic element. Sporadic ALS (sALS) typically emerges between 50 and 60 years old [113]. The onset of sALS is unknown, and thus the identification of causal genes and environmental factors remains elusive. In familial ALS, about 20% of the cases resulted from mutations in SOD1 [114]. The functions of SOD1 are diverse and include scavenging excessive superoxide radical ($\text{O}_2^{\bullet-}$), modulating cellular respiration, energy metabolism, and posttranslational modification [115]. Although SOD dysfunction leads to a loss of antioxidant capability, evidence has shown that the genetic ablation of SOD1 in mice does not lead to neurodegenerative conditions [11, 16]. In contrast, the gain-of-function of mutant SOD1 protein has been significantly implicated in the motor neuron diseases [16]. For instance, a recent study by Bastow et al. showed that mutant SOD1 can disturb the amino acid biosynthesis of cells in a yeast model and mediate cellular destruction, accounting for the neural degeneration in ALS (Figure 1) [116].

SOD1 regulates Rac1 directly via the association within endosomes, which subsequently activates Nox. Nox-containing endosomes, also called redoxosomes, are important in regulating proinflammatory signals through NF- κ B. Nox forms $\text{O}_2^{\bullet-}$, from molecular oxygen, which plays important roles in cell signaling, enzyme regulation, and antibacterial effect [11, 117]. Under physiological conditions, the ratio of ROS to anti-oxidative molecules is balanced. However, during disease states, rapid changes in ROS levels and disruptions in antioxidant capability can result in increased DNA damage, lipid peroxidation, and apoptosis [11]. SOD1 is responsible for the conversion of $\text{O}_2^{\bullet-}$ into hydrogen peroxide (H_2O_2) and molecular oxygen. SOD1 mutants enhance the production of Nox2-dependent ROS, which is thought to be the cause of motor neuron death in ALS [117]. Interestingly, oxidized or misfolded wild-type SOD1 has been shown

to lead to mitochondrial dysfunction contributing to the pathogenesis of sALS [118].

Mutant SOD1 can contribute to the progression of familial ALS through the dysregulation of signal transduction pathways in motor neurons and in the activity of supportive glial cells [117, 119]. For example, SOD1 is thought to be an important cell-signaling molecule with neuromodulatory properties. Studies *in vitro* and in transgenic mouse models show that SOD1 is secreted via the microvesicular secretory pathway. Secreted SOD1 binds to muscarinic receptors on neighboring neurons and increases ERK/AKT signaling, and intracellular Ca^{2+} concentration [120–122]. Whether the secretion of SOD1 is induced by ROS or is constitutive remains as a topic of debate [120]. SOD1 maintains the integrity of motor neurons through activating ERK/AKT signaling, and it has been shown that SOD1 secretion can be increased in neurons under OS [123, 124]. In rats, propofol conditioning treatment was shown to prevent ischemia–reperfusion damage of the spinal cord by increasing PI3K/AKT signaling mediated potentially through increased SOD1 activity [125]. Additionally, OS can induce neuron cell death by inhibiting neuroprotective IGF-I/AKT pathway, prompting further investigation on the role of AKT signaling in neurodegeneration [126].

Like many of the other ALS genes, SOD1 is also expressed in cells other than motor neurons [119]. SOD1 mutant potentially alters astrocytes and microglia activities, contributing to motor neuron degeneration. This pathological mechanism has been termed the non-cell autonomous pathway [119]. The importance of the interaction between astrocytes and motor neurons can be observed using coculture models, where primary or embryonic stem cell derived motor neurons are cultured in the presence of astrocytes expressing mutant SOD1. These cocultured motor neurons, as well as motor neurons treated with conditioned cell culture medium from astrocytes expressing mutant SOD1, experienced increased cell damage [127]. Studies also indicate the excretion of specific soluble factors that are toxic to motor neurons and lead to an inflammatory response, as a direct cause of disease progression [127, 128]. Furthermore, astrocytes expressing mutant SOD1 reduce the expression of glutamate transporter (GLT) 1, leading to increased extracellular concentration of glutamate and glutamate toxicity in motor neurons (Figure 1) [129, 130]. However, the overexpression of GLT1 via adenoviral vectors is not effective in reducing motor neuron death, supporting the multifaceted role of SOD1 in disease progression of ALS [128].

6. Oxidative Stress in Spinocerebellar Ataxia Disease

Spinocerebellar ataxia (SCA) is an autosomal dominant disease characterized by progressive neurodegeneration. Common symptoms associated with SCA are ataxic gait, oculomotor disorders, dysarthria and cognitive impairment, which can ultimately cause death. Over 20 types of SCAs have been identified based on genetic descriptions [131–133]. An expansion of repeated CAG trinucleotides accounts for the major pathogenic mutation in SCA [132]. This mutation

leads to the overexpression of mutant ataxin 1 (ATXN1) protein that has an expanded polyglutamine. Mutant ATXN1 can affect the stability of RAR-related orphan receptor alpha ($\text{ROR}\alpha$), which plays an important role in Purkinje cell functions. Decreased gene expression of $\text{ROR}\alpha$ has been associated with ataxia and cerebellar hypoplasia [134, 135].

While most types of SCAs are suggested as genetic diseases correlated with ATXN mutation, other pathogenic mechanisms that involve the dysfunction of mitochondria have been proposed [134, 135]. Hakonen et al. observed respiratory complex I deficiency and mitochondrial DNA depletion in the brain of infantile-onset SCA individuals [136]. Although low levels of ROS play a critical role in cell signaling, the overaccumulation of ROS can be neurotoxic and have been implicated in neurodegeneration [11]. A study conducted by Stucki et al. found elevated OS and significant mitochondrial alterations in the Purkinje cells of SCA1. They suspect that there is a potential correlation between the progression of SCA and mitochondrial impairments caused by OS (Figure 1) [135]. Correspondingly, the study examined the effects of MitoQ (a mitochondrial antioxidant) using a SCA mouse model. It was found that long-term administration of MitoQ significantly restores mitochondrial morphology and functions in Purkinje cells as well as alleviates SCA1-related symptoms such as motor incoordination [135]. Their findings show the potential of using mitochondria-targeted antioxidants as a treatment for SCA1.

Like many neural disorders mentioned previously, SCA is linked with mitochondrial dysfunction [21, 137]. A typical example is the Friedreich ataxia, which is characterized by a loss of frataxin, an iron transporter protein located on mitochondrial inner membrane. With the reduction of the frataxin, iron concentration increases in the mitochondrial matrix, which promotes the conversion of H_2O_2 to $\cdot\text{OH}$ through the Fenton reaction. The highly reactive $\cdot\text{OH}$ molecules can cause oxidative damage to mitochondria and compromise the efficiency of energy production in neuron cells [21, 22]. As a result, antioxidant treatment including vitamin E and coenzyme Q10 supplementation has been shown to improve energy generation for some Friedreich ataxia patients by restoring mitochondrial function and attenuating OS [138].

Due to the high mitochondrial content in the brain, mitochondrial dysfunction can result in significant negative effects on the neural system. ROS are naturally produced from the mitochondrial respiratory chain and play an important role in maintaining mitochondrial function as well as the robustness of neural cells. However, scant research has been done so far to resolve the potential roles of ROS on SCA diseases and develop optimal therapeutic strategies. Further studies are needed to understand the redox mechanisms underlying different types of SCAs, with a focus on ROS-targeted therapies.

7. Summary and Prospective

Numerous studies have been performed to investigate the therapeutic effects of antioxidants on neurodegenerative

disorders but they have yielded mixed results [14, 106, 110]. This is potentially due to inadequate dosage or timing of administration, or the unsuitable duration of therapy [30, 139]. Novel antioxidant compounds have been developed that show great potential in mediating disease phenotypes. For example, Fer-1, as a potent antioxidant, can effectively prevent neuronal cell death in HD and PD, via the inhibition of lipid peroxidation and the attenuation of glutamate toxicity [27, 28]. Antioxidants protect the redox balance of neural cells by targeting both the upstream and downstream of OS [140]. The upstream protective effects rely on the prevention of ROS overaccumulation and lipid or protein oxidation [11]. Common dietary antioxidants such as vitamins C and E can interact with oxyradicals directly to diminish their toxic effects [140]. Alternatively, supplemental fatty acids can replace oxidized membrane lipids thus reducing oxidative damage [140]. By contrast, antioxidant therapeutics that work downstream of OS focuses on reducing the toxicity from mutant protein aggregation and alleviating neural inflammation [140]. For example, *Ginkgo biloba*, a Chinese herb with strong antioxidant properties, has been shown to be able to improve cognitive conditions in AD by alleviating toxicity of A β plaques [141].

Presumably, there is more than one type of ROS involved in the pathophysiology of disease progression [30, 72]. ROS production is subtly regulated by complex antioxidant defense systems within the biological system [11]. Therefore, single exogenous antioxidant supplementation may not be sufficient to combat OS induced under pathophysiological conditions and may even result in a disturbance of redox balance in the body. In this regard, a combination of various antioxidants should be considered for treating neurodegenerative diseases in the future studies. Furthermore, the optimized time, dose and duration of antioxidant treatments remain to be determined for optimal clinical outcomes [30, 72].

In this review, we summarized the complex roles of ROS in common neurodegenerative disorders including AD, PD, HD, ALS, and SCA. High levels of OS have been implicated in many neurodegenerative conditions. Accumulating evidence suggests that ROS may be generated via various mechanisms and have complex roles in promoting disease development. Particularly, mitochondria dysfunction is linked with sustained OS in neurodegenerative disorders. Multiple studies have been performed to explore the effectiveness of antioxidants in attenuating neurodegenerative symptoms. Although there has been no convincing evidence showing their neuroprotective efficacy, research has certainly achieved encouraging outcomes. Further research is paramount in addressing the exact roles of ROS in various neurodegenerative conditions and developing antioxidant-based therapeutic interventions.

Conflicts of Interest

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

The authors thank Chia-Chen Chuang, Denethi Wijegunawardana, Evan R. Prather, and David C. Sypert for their assistance during the manuscript preparation.

References

- [1] Alzheimer's Association, "2016 Alzheimer's disease facts and figures," *Alzheimer's & Dementia*, vol. 12, pp. 459–509, 2016.
- [2] H. R. Silvade, N. L. Khan, and N. W. Wood, "The genetics of Parkinson's disease," *Current Opinion in Genetics & Development*, vol. 10, pp. 292–298, 2000.
- [3] D. S. Albers and M. F. Beal, "Mitochondrial dysfunction and oxidative stress in aging and neurodegenerative disease," *Journal of Neural Transmission Supplementum*, vol. 59, pp. 133–154, 2000.
- [4] A. Matilla-Duenas, T. Ashizawa, A. Brice et al., "Consensus paper: pathological mechanisms underlying neurodegeneration in spinocerebellar ataxias," *Cerebellum*, vol. 13, pp. 269–302, 2014.
- [5] M. Hamer and Y. Chida, "Physical activity and risk of neurodegenerative disease: a systematic review of prospective evidence," *Psychological Medicine*, vol. 39, pp. 3–11, 2009.
- [6] M. C. Rijkde, L. J. Launer, K. Berger et al., "Prevalence of Parkinson's disease in Europe: a collaborative study of population-based cohorts," *Neurology*, vol. 54, pp. S21–S33, 2000.
- [7] L. M. Bekris, I. F. Mata, and C. P. Zabetian, "The genetics of Parkinson disease," *Journal of Geriatric Psychiatry and Neurology*, vol. 23, pp. 228–242, 2010.
- [8] M. J. Farrer, "Genetics of Parkinson disease: paradigm shifts and future prospects," *Nature Reviews Genetics*, vol. 7, pp. 306–318, 2006.
- [9] Alzheimer's Association, "2011 Alzheimer's disease facts and figures," *Alzheimer's & Dementia*, vol. 7, pp. 208–244, 2011.
- [10] L. Zuo, M. S. Koozechian, and L. L. Chen, "Characterization of reactive nitrogen species in allergic asthma," *Annals of Allergy Asthma & Immunology*, vol. 112, pp. 18–22, 2014.
- [11] L. Zuo, T. Zhou, B. K. Pannell, A. C. Ziegler, and T. M. Best, "Biological and physiological role of reactive oxygen species—the good, the bad and the ugly," *Acta Physiologica*, vol. 214, pp. 329–348, 2015.
- [12] F. He and L. Zuo, "Redox roles of reactive oxygen species in cardiovascular diseases," *International Journal of Molecular Sciences*, vol. 16, pp. 27770–27780, 2015.
- [13] L. Zuo, A. Shiah, W. J. Roberts, M. T. Chien, P. D. Wagner, and M. C. Hogan, "Low Po₂ conditions induce reactive oxygen species formation during contractions in single skeletal muscle fibers," *American Journal of Physiology - Regulatory, Integrative and Comparative Physiology*, vol. 304, pp. R1009–R1016, 2013.
- [14] V. Dias, E. Junn, and M. M. Mouradian, "The role of oxidative stress in Parkinson's disease," *Journal of Parkinson's Disease*, vol. 3, pp. 461–491, 2013.
- [15] J. St-Pierre, S. Drori, M. Uldry et al., "Suppression of reactive oxygen species and neurodegeneration by the PGC-1 transcriptional coactivators," *Cell*, vol. 127, pp. 397–408, 2006.
- [16] K. Hensley, M. Mhatre, S. Mou et al., "On the relation of oxidative stress to neuroinflammation: lessons learned from the G93A-SOD1 mouse model of amyotrophic lateral

- sclerosis," *Antioxidants & Redox Signaling*, vol. 8, pp. 2075–2087, 2006.
- [17] A. C. Rego and C. R. Oliveira, "Mitochondrial dysfunction and reactive oxygen species in excitotoxicity and apoptosis: implications for the pathogenesis of neurodegenerative diseases," *Neurochemical Research*, vol. 28, pp. 1563–1574, 2003.
- [18] R. L. Mosley, E. J. Benner, I. Kadiu et al., "Neuroinflammation, oxidative stress and the pathogenesis of Parkinson's disease," *Clinical Neuroscience Research*, vol. 6, pp. 261–281, 2006.
- [19] T. Wyss-Coray and L. Mucke, "Inflammation in neurodegenerative disease—a double-edged sword," *Neuron*, vol. 35, pp. 419–432, 2002.
- [20] M. T. Lin and M. F. Beal, "Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases," *Nature*, vol. 443, pp. 787–795, 2006.
- [21] V. Campuzano, L. Montermini, Y. Lutz et al., "Frataxin is reduced in Friedreich ataxia patients and is associated with mitochondrial membranes," *Human Molecular Genetics*, vol. 6, pp. 1771–1780, 1997.
- [22] D. C. Wallace, "Mitochondrial diseases in man and mouse," *Science*, vol. 283, pp. 1482–1488, 1999.
- [23] C. A. Ross and S. J. Tabrizi, "Huntington's disease: from molecular pathogenesis to clinical treatment," *Lancet Neurology*, vol. 10, pp. 83–98, 2011.
- [24] M. Sano, C. Ernesto, R. G. Thomas et al., "A controlled trial of selegiline, alpha-tocopherol, or both as treatment for Alzheimer's disease," *The New England Journal of Medicine*, vol. 336, pp. 1216–1222, 1997.
- [25] L. I. Golbe, T. M. Farrell, and P. H. Davis, "Case-control study of early life dietary factors in Parkinson's disease," *Archives of Neurology*, vol. 45, pp. 1350–1353, 1988.
- [26] E. J. Pappert, C. C. Tangney, C. G. Goetz et al., "Alpha-tocopherol in the ventricular cerebrospinal fluid of Parkinson's disease patients: dose-response study and correlations with plasma levels," *Neurology*, vol. 47, pp. 1037–1042, 1996.
- [27] R. Skouta, "Neuroprotective effect of antioxidant compounds," *Neural Regeneration Research*, vol. 11, pp. 566–567, 2016.
- [28] R. Skouta, S. J. Dixon, J. Wang et al., "Ferrostatins inhibit oxidative lipid damage and cell death in diverse disease models," *Journal of the American Chemical Society*, vol. 136, pp. 4551–4556, 2014.
- [29] Y. Gilgun-Sherki, E. Melamed, and D. Offen, "Oxidative stress induced-neurodegenerative diseases: the need for antioxidants that penetrate the blood brain barrier," *Neuropharmacology*, vol. 40, pp. 959–975, 2001.
- [30] G. H. Kim, J. E. Kim, S. J. Rhie, and S. Yoon, "The role of oxidative stress in neurodegenerative diseases," *Experimental Neurobiology*, vol. 24, pp. 325–340, 2015.
- [31] L. Zuo, B. T. Hemmelgarn, C. C. Chuang, and T. M. Best, "The role of oxidative stress-induced epigenetic alterations in amyloid-beta production in Alzheimer's disease," *Oxidative Medicine and Cellular Longevity*, vol. 2015, Article ID 604658, 13 pages, 2015.
- [32] H. W. Querfurth and F. M. LaFerla, "Alzheimer's disease," *The New England Journal of Medicine*, vol. 362, pp. 329–344, 2010.
- [33] D. A. Butterfield, "The 2013 SFRBM discovery award: selected discoveries from the butterfield laboratory of oxidative stress and its sequela in brain in cognitive disorders exemplified by Alzheimer disease and chemotherapy induced cognitive impairment," *Free Radical Biology & Medicine*, vol. 74, pp. 157–174, 2014.
- [34] E. Ferreiro, C. R. Oliveira, and C. M. Pereira, "The release of calcium from the endoplasmic reticulum induced by amyloid-beta and prion peptides activates the mitochondrial apoptotic pathway," *Neurobiology of Disease*, vol. 30, pp. 331–342, 2008.
- [35] D. J. Bonda, X. Wang, G. Perry et al., "Oxidative stress in Alzheimer disease: a possibility for prevention," *Neuropharmacology*, vol. 59, pp. 290–294, 2010.
- [36] W. J. Huang, X. Zhang, and W. W. Chen, "Role of oxidative stress in Alzheimer's disease," *Biomedical Reports*, vol. 4, pp. 519–522, 2016.
- [37] T. Nakamura and S. A. Lipton, "Redox modulation by S-nitrosylation contributes to protein misfolding, mitochondrial dynamics, and neuronal synaptic damage in neurodegenerative diseases," *Cell Death and Differentiation*, vol. 18, pp. 1478–1486, 2011.
- [38] T. Nakamura and S. A. Lipton, "Preventing Ca²⁺-mediated nitrosative stress in neurodegenerative diseases: possible pharmacological strategies," *Cell Calcium*, vol. 47, pp. 190–197, 2010.
- [39] D. A. Patten, M. Germain, M. A. Kelly, and R. S. Slack, "Reactive oxygen species: stuck in the middle of neurodegeneration," *Journal of Alzheimer's Disease*, vol. 20, Supplement 2, pp. S357–S367, 2010.
- [40] P. B. Shelat, M. Chalimoniuk, J. H. Wang et al., "Amyloid beta peptide and NMDA induce ROS from NADPH oxidase and AA release from cytosolic phospholipase A₂ in cortical neurons," *Journal of Neurochemistry*, vol. 106, pp. 45–55, 2008.
- [41] E. Giraldo, A. Lloret, T. Fuchsberger, and J. Vina, "Abeta and tau toxicities in Alzheimer's are linked via oxidative stress-induced p38 activation: protective role of vitamin E," *Redox Biology*, vol. 2, pp. 873–877, 2014.
- [42] N. Bulat and C. Widmann, "Caspase substrates and neurodegenerative diseases," *Brain Research Bulletin*, vol. 80, pp. 251–267, 2009.
- [43] P. Agostinho, J. P. Lopes, Z. Velez, and C. R. Oliveira, "Overactivation of calcineurin induced by amyloid-beta and prion proteins," *Neurochemistry International*, vol. 52, pp. 1226–1233, 2008.
- [44] A. Awasthi, Y. Matsunaga, and T. Yamada, "Amyloid-beta causes apoptosis of neuronal cells via caspase cascade, which can be prevented by amyloid-beta-derived short peptides," *Experimental Neurology*, vol. 196, pp. 282–289, 2005.
- [45] A. Hamilton and C. Holscher, "The effect of ageing on neurogenesis and oxidative stress in the APP_{swe}/PS1_{deltaE9} mouse model of Alzheimer's disease," *Brain Research*, vol. 1449, pp. 83–93, 2012.
- [46] M. L. Block, "NADPH oxidase as a therapeutic target in Alzheimer's disease," *BMC Neuroscience*, vol. 9, Supplement 2, p. S8, 2008.
- [47] G. S. Aseervatham, T. Sivasudha, R. Jeyadevi, and A. D. Arul, "Environmental factors and unhealthy lifestyle influence oxidative stress in humans—an overview," *Environmental Science and Pollution Research International*, vol. 20, pp. 4356–4369, 2013.

- [48] M. A. Smith, X. Zhu, M. Tabaton et al., "Increased iron and free radical generation in preclinical Alzheimer disease and mild cognitive impairment," *Journal of Alzheimer's Disease*, vol. 19, pp. 363–372, 2010.
- [49] E. R. Stadtman, "Protein oxidation in aging and age-related diseases," *Annals of the New York Academy of Sciences*, vol. 928, pp. 22–38, 2001.
- [50] C. Holmes, C. Cunningham, E. Zotova et al., "Systemic inflammation and disease progression in Alzheimer disease," *Neurology*, vol. 73, pp. 768–774, 2009.
- [51] M. Johnstone, A. J. Gearing, and K. M. Miller, "A central role for astrocytes in the inflammatory response to beta-amyloid; chemokines, cytokines and reactive oxygen species are produced," *Journal of Neuroimmunology*, vol. 93, pp. 182–193, 1999.
- [52] T. J. Seabrook, L. Y. Jiang, M. Maier, and C. A. Lemere, "Minocycline affects microglia activation, a beta deposition, and behavior in APP-tg mice," *Glia*, vol. 53, pp. 776–782, 2006.
- [53] W. Y. Wang, M. S. Tan, J. T. Yu, and L. Tan, "Role of pro-inflammatory cytokines released from microglia in Alzheimer's disease," *Annals of Translational Medicine*, vol. 3, p. 136, 2015.
- [54] M. Nizzari, S. Thellung, A. Corsaro et al., "Neurodegeneration in Alzheimer disease: role of amyloid precursor protein and presenilin 1 intracellular signaling," *Journal of Toxicology*, vol. 2012, Article ID 187297, 13 pages, 2012.
- [55] D. A. Butterfield and D. Boyd-Kimball, "The critical role of methionine 35 in Alzheimer's amyloid beta-peptide (1-42)-induced oxidative stress and neurotoxicity," *Biochimica et Biophysica Acta*, vol. 1703, pp. 149–156, 2005.
- [56] C. C. Curtain, F. Ali, I. Volitakis et al., "Alzheimer's disease amyloid-beta binds copper and zinc to generate an allosterically ordered membrane-penetrating structure containing superoxide dismutase-like subunits," *The Journal of Biological Chemistry*, vol. 276, pp. 20466–20473, 2001.
- [57] M. W. Dysken, P. D. Guarino, J. E. Vertrees et al., "Vitamin E and memantine in Alzheimer's disease: clinical trial methods and baseline data," *Alzheimer's & Dementia*, vol. 10, pp. 36–44, 2014.
- [58] D. J. Selkoe, "Alzheimer's disease: genes, proteins, and therapy," *Physiological Reviews*, vol. 81, pp. 741–766, 2001.
- [59] M. Dumont and M. F. Beal, "Neuroprotective strategies involving ROS in Alzheimer disease," *Free Radical Biology and Medicine*, vol. 51, pp. 1014–1026, 2011.
- [60] K. Kanninen, T. M. Malm, H. K. Jyrkkanen et al., "Nuclear factor erythroid 2-related factor 2 protects against beta amyloid," *Molecular and Cellular Neurosciences*, vol. 39, pp. 302–313, 2008.
- [61] S. Jeon, J. Hur, H. J. Jeong, B. S. Koo, and S. C. Pak, "SuHeXiang wan essential oil alleviates amyloid beta induced memory impairment through inhibition of tau protein phosphorylation in mice," *The American Journal of Chinese Medicine*, vol. 39, pp. 917–932, 2011.
- [62] F. Yang, G. P. Lim, A. N. Begum et al., "Curcumin inhibits formation of amyloid beta oligomers and fibrils, binds plaques, and reduces amyloid in vivo," *The Journal of Biological Chemistry*, vol. 280, pp. 5892–5901, 2005.
- [63] L. Lannfelt, K. Blennow, H. Zetterberg et al., "And group PEs. Safety, efficacy, and biomarker findings of PBT2 in targeting Aβeta as a modifying therapy for Alzheimer's disease: a phase IIa, double-blind, randomised, placebo-controlled trial," *Lancet Neurology*, vol. 7, pp. 779–786, 2008.
- [64] J. Graff and L. H. Tsai, "The potential of HDAC inhibitors as cognitive enhancers," *Annual Review of Pharmacology and Toxicology*, vol. 53, pp. 311–330, 2013.
- [65] J. Labbadia and R. I. Morimoto, "Huntington's disease: underlying molecular mechanisms and emerging concepts," *Trends in Biochemical Sciences*, vol. 38, pp. 378–385, 2013.
- [66] J. Gil-Mohapel, P. S. Brocardo, and B. R. Christie, "The role of oxidative stress in Huntington's disease: are antioxidants good therapeutic candidates?," *Current Drug Targets*, vol. 15, pp. 454–468, 2014.
- [67] F. Mochel and R. G. Haller, "Energy deficit in Huntington disease: why it matters," *The Journal of Clinical Investigation*, vol. 121, pp. 493–499, 2011.
- [68] J. G. Hodgson, N. Agopyan, C. A. Gutekunst et al., "A YAC mouse model for Huntington's disease with full-length mutant huntingtin, cytoplasmic toxicity, and selective striatal neurodegeneration," *Neuron*, vol. 23, pp. 181–192, 1999.
- [69] S. W. Davies, M. Turmaine, B. A. Cozens et al., "Formation of neuronal intranuclear inclusions underlies the neurological dysfunction in mice transgenic for the HD mutation," *Cell*, vol. 90, pp. 537–548, 1997.
- [70] L. Cui, H. Jeong, F. Borovecki, C. N. Parkhurst, N. Tanese, and D. Krainc, "Transcriptional repression of PGC-1α by mutant huntingtin leads to mitochondrial dysfunction and neurodegeneration," *Cell*, vol. 127, pp. 59–69, 2006.
- [71] P. Weydt, V. V. Pineda, A. E. Torrence et al., "Thermoregulatory and metabolic defects in Huntington's disease transgenic mice implicate PGC-1α in Huntington's disease neurodegeneration," *Cell Metabolism*, vol. 4, pp. 349–362, 2006.
- [72] A. Kumar and R. R. Ratan, "Oxidative stress and Huntington's disease: the good, the bad, and the ugly," *Journal of Huntington's Disease*, vol. 5, pp. 217–237, 2016.
- [73] A. Covarrubias-Pinto, P. Moll, M. Solis-Maldonado et al., "Beyond the redox imbalance: oxidative stress contributes to an impaired GLUT3 modulation in Huntington's disease," *Free Radical Biology and Medicine*, vol. 89, pp. 1085–1096, 2015.
- [74] L. P. Reagan, A. M. Magarinos, D. K. Yee et al., "Oxidative stress and HNE conjugation of GLUT3 are increased in the hippocampus of diabetic rats subjected to stress," *Brain Research*, vol. 862, pp. 292–300, 2000.
- [75] M. Bonora, M. R. Wieckowski, C. Chinopoulos et al., "Molecular mechanisms of cell death: central implication of ATP synthase in mitochondrial permeability transition," *Oncogene*, vol. 34, p. 1608, 2015.
- [76] A. V. Panov, C. A. Gutekunst, B. R. Leavitt et al., "Early mitochondrial calcium defects in Huntington's disease are a direct effect of polyglutamines," *Nature Neuroscience*, vol. 5, pp. 731–736, 2002.
- [77] E. Bossy-Wetzel, A. Petrilli, and A. B. Knott, "Mutant huntingtin and mitochondrial dysfunction," *Trends in Neurosciences*, vol. 31, pp. 609–616, 2008.
- [78] G. Liot, J. Valette, J. Pepin, J. Flament, and E. Brouillet, "Energy defects in Huntington's disease: why "in vivo" evidence matters," *Biochemical and Biophysical Research Communications*, vol. 483, no. 4, pp. 1084–1095, 2017.
- [79] C. Zuccato, M. Valenza, and E. Cattaneo, "Molecular mechanisms and potential therapeutical targets in Huntington's disease," *Physiological Reviews*, vol. 90, pp. 905–981, 2010.

- [80] J. P. Bolanos, M. A. Moro, I. Lizasoain, and A. Almeida, "Mitochondria and reactive oxygen and nitrogen species in neurological disorders and stroke: therapeutic implications," *Advanced Drug Delivery Reviews*, vol. 61, pp. 1299–1315, 2009.
- [81] I. Tunez, F. Sanchez-Lopez, E. Aguera, R. Fernandez-Bolanos, F. M. Sanchez, and I. Tasset-Cuevas, "Important role of oxidative stress biomarkers in Huntington's disease," *Journal of Medicinal Chemistry*, vol. 54, pp. 5602–5606, 2011.
- [82] N. Klepac, M. Relja, R. Klepac, S. Hecimovic, T. Babic, and V. Trkulja, "Oxidative stress parameters in plasma of Huntington's disease patients, asymptomatic Huntington's disease gene carriers and healthy subjects : a cross-sectional study," *Journal of Neurology*, vol. 254, pp. 1676–1683, 2007.
- [83] I. Ciancarelli, D. AmicisDe, C. MassimoDi et al., "Influence of intensive multifunctional neurorehabilitation on neuronal oxidative damage in patients with Huntington's disease," *Functional Neurology*, vol. 30, pp. 47–52, 2015.
- [84] M. A. Sorolla, G. Reverter-Branchat, J. Tamarit, I. Ferrer, J. Q. Ros, and E. Cabiscol, "Proteomic and oxidative stress analysis in human brain samples of Huntington disease," *Free Radical Biology and Medicine*, vol. 45, pp. 667–678, 2008.
- [85] T. Y. C. Pang, N. C. Stam, J. Nithianantharajah, M. L. Howard, and A. J. Hannan, "Differential effects of voluntary physical exercise on behavioral and brain-derived neurotrophic factor expression deficits in Huntington's disease transgenic mice," *Neuroscience*, vol. 141, pp. 569–584, 2006.
- [86] R. M. Arida, F. A. Scorza, and E. A. Cavalheiro, "Role of physical exercise as complementary treatment for epilepsy and other brain disorders," *Current Pharmaceutical Design*, vol. 19, pp. 6720–6725, 2013.
- [87] S. E. Browne, R. J. Ferrante, and M. F. Beal, "Oxidative stress in Huntington's disease," *Brain Pathology*, vol. 9, pp. 147–163, 1999.
- [88] M. C. Polidori, P. Mecocci, S. E. Browne, U. Senin, and M. F. Beal, "Oxidative damage to mitochondrial DNA in Huntington's disease parietal cortex," *Neuroscience Letters*, vol. 272, pp. 53–56, 1999.
- [89] E. Miller, A. Morel, L. Saso, and J. Saluk, "Isoprostanes and neuroprostanes as biomarkers of oxidative stress in neurodegenerative diseases," *Oxidative Medicine and Cellular Longevity*, vol. 2014, Article ID 572491, 10 pages, 2014.
- [90] R. S. Sohal and R. Weindruch, "Oxidative stress, caloric restriction, and aging," *Science*, vol. 273, pp. 59–63, 1996.
- [91] A. L. McCormack, M. Thiruchelvam, A. B. Manning-Bog et al., "Environmental risk factors and Parkinson's disease: selective degeneration of nigral dopaminergic neurons caused by the herbicide paraquat," *Neurobiology of Disease*, vol. 10, pp. 119–127, 2002.
- [92] L. X. Qin, J. Q. Tan, H. N. Zhang et al., "BAG5 interacts with DJ-1 and inhibits the neuroprotective effects of DJ-1 to combat mitochondrial oxidative damage," *Oxidative Medicine and Cellular Longevity*, vol. 2017, Article ID 5094934, 10 pages, 2017.
- [93] K. Apel and H. Hirt, "Reactive oxygen species: metabolism, oxidative stress, and signal transduction," *Annual Review of Plant Biology*, vol. 55, pp. 373–399, 2004.
- [94] S. J. Chinta and J. K. Andersen, "Redox imbalance in Parkinson's disease," *Biochimica et Biophysica Acta*, vol. 1780, pp. 1362–1367, 2008.
- [95] J. Meiser, D. Weindl, and K. Hiller, "Complexity of dopamine metabolism," *Cell Communication and Signaling: CCS*, vol. 11, p. 34, 2013.
- [96] S. Gangemi, E. Gofita, C. Costa et al., "Occupational and environmental exposure to pesticides and cytokine pathways in chronic diseases (review)," *International Journal of Molecular Medicine*, vol. 38, pp. 1012–1020, 2016.
- [97] R. Perfeito, T. Cunha-Oliveira, and A. C. Rego, "Revisiting oxidative stress and mitochondrial dysfunction in the pathogenesis of Parkinson disease—resemblance to the effect of amphetamine drugs of abuse," *Free Radical Biology and Medicine*, vol. 53, pp. 1791–1806, 2012.
- [98] L. Zuo and M. S. Motherwell, "The impact of reactive oxygen species and genetic mitochondrial mutations in Parkinson's disease," *Gene*, vol. 532, pp. 18–23, 2013.
- [99] V. Adam-Vizi, "Production of reactive oxygen species in brain mitochondria: contribution by electron transport chain and non-electron transport chain sources," *Antioxidants & Redox Signaling*, vol. 7, pp. 1140–1149, 2005.
- [100] E. M. Valente, P. M. Abou-Sleiman, V. Caputo et al., "Hereditary early-onset Parkinson's disease caused by mutations in PINK1," *Science*, vol. 304, pp. 1158–1160, 2004.
- [101] H. Jiang, Y. Ren, J. Zhao, and J. Feng, "Parkin protects human dopaminergic neuroblastoma cells against dopamine-induced apoptosis," *Human Molecular Genetics*, vol. 13, pp. 1745–1754, 2004.
- [102] G. Ganguly, S. Chakrabarti, U. Chatterjee, and L. Saso, "Proteinopathy, oxidative stress and mitochondrial dysfunction: cross talk in Alzheimer's disease and Parkinson's disease," *Drug Design, Development and Therapy*, vol. 11, pp. 797–810, 2017.
- [103] N. A. Mazo, V. Echeverria, R. Cabezas et al., "Medicinal plants as protective strategies against Parkinson's disease," *Current Pharmaceutical Design*, vol. 23, 2017.
- [104] P. I. Moreira, X. Zhu, X. Wang et al., "Mitochondria: a therapeutic target in neurodegeneration," *Biochimica et Biophysica Acta*, vol. 1802, pp. 212–220, 2010.
- [105] S. A. Zaitone, D. M. Abo-Elmatty, and A. A. Shaalan, "Acetyl-L-carnitine and alpha-lipoic acid affect rotenone-induced damage in nigral dopaminergic neurons of rat brain, implication for Parkinson's disease therapy," *Pharmacology, Biochemistry, and Behavior*, vol. 100, pp. 347–360, 2012.
- [106] I. Shoulson, "DATATOP: a decade of neuroprotective inquiry. Parkinson study group. Deprenyl and tocopherol Antioxidative therapy of parkinsonism," *Annals of Neurology*, vol. 44, pp. S160–S166, 1998.
- [107] S. Fahn, "A pilot trial of high-dose alpha-tocopherol and ascorbate in early Parkinson's disease," *Annals of Neurology*, vol. 32, Supplement, pp. S128–S132, 1992.
- [108] M. Etminan, S. S. Gill, and A. Samii, "Intake of vitamin E, vitamin C, and carotenoids and the risk of Parkinson's disease: a meta-analysis," *Lancet Neurology*, vol. 4, pp. 362–365, 2005.
- [109] C. W. Shults, D. Oakes, K. Kieburtz et al., "Effects of coenzyme Q10 in early Parkinson disease: evidence of slowing of the functional decline," *Archives of Neurology*, vol. 59, pp. 1541–1550, 2002.
- [110] M. H. Yan, X. Wang, and X. Zhu, "Mitochondrial defects and oxidative stress in Alzheimer disease and Parkinson disease," *Free Radical Biology and Medicine*, vol. 62, pp. 90–101, 2013.

- [111] M. C. Kiernan, S. Vucic, B. C. Cheah et al., "Amyotrophic lateral sclerosis," *Lancet*, vol. 377, pp. 942–955, 2011.
- [112] J. P. Taylor, R. H. Brown Jr., and D. W. Cleveland, "Decoding ALS: from genes to mechanism," *Nature*, vol. 539, pp. 197–206, 2016.
- [113] C. Ingre, P. M. Roos, F. Piehl, F. Kamel, and F. Fang, "Risk factors for amyotrophic lateral sclerosis," *Clinical Epidemiology*, vol. 7, pp. 181–193, 2015.
- [114] J. Gamez, M. Corbera-Bellalta, G. Nogales et al., "Mutational analysis of the cu/Zn superoxide dismutase gene in a Catalan ALS population: should all sporadic ALS cases also be screened for SOD1?," *Journal of the Neurological Sciences*, vol. 247, pp. 21–28, 2006.
- [115] R. A. Saccon, R. K. Bunton-Stasyshyn, E. M. Fisher, and P. Fratta, "Is SOD1 loss of function involved in amyotrophic lateral sclerosis?," *Brain*, vol. 136, pp. 2342–2358, 2013.
- [116] E. L. Bastow, A. R. Peswani, D. S. J. Tarrant et al., "New links between SOD1 and metabolic dysfunction from a yeast model of amyotrophic lateral sclerosis," *Journal of Cell Science*, vol. 129, pp. 4118–4129, 2016.
- [117] Q. Li, N. Y. Spencer, N. J. Pantazis, and J. F. Engelhardt, "Alsin and SOD1(G93A) proteins regulate endosomal reactive oxygen species production by glial cells and proinflammatory pathways responsible for neurotoxicity," *The Journal of Biological Chemistry*, vol. 286, pp. 40151–40162, 2011.
- [118] E. D'Amico, P. Factor-Litvak, R. M. Santella, and H. Mitsumoto, "Clinical perspective on oxidative stress in sporadic amyotrophic lateral sclerosis," *Free Radical Biology and Medicine*, vol. 65, pp. 509–527, 2013.
- [119] J. Lee, S. J. Hyeon, H. Im, H. Ryu, Y. Kim, and H. Ryu, "Astrocytes and microglia as non-cell autonomous players in the pathogenesis of ALS," *Experimental Neurobiology*, vol. 25, pp. 233–240, 2016.
- [120] P. Mondola, S. Damiano, A. Sasso, and M. Santillo, "The cu, Zn superoxide dismutase: not only a dismutase enzyme," *Frontiers in Physiology*, vol. 7, p. 594, 2016.
- [121] P. Mondola, M. Santillo, R. Seru et al., "Cu,Zn superoxide dismutase increases intracellular calcium levels via a phospholipase C-protein kinase C pathway in SK-N-BE neuroblastoma cells," *Biochemical and Biophysical Research Communications*, vol. 324, pp. 887–892, 2004.
- [122] P. Mondola, G. Ruggiero, R. Seru et al., "The Cu,Zn superoxide dismutase in neuroblastoma SK-N-BE cells is exported by a microvesicles dependent pathway," *Brain Research Molecular Brain Research*, vol. 110, pp. 45–51, 2003.
- [123] P. Mondola, T. Annella, R. Seru et al., "Secretion and increase of intracellular CuZn superoxide dismutase content in human neuroblastoma SK-N-BE cells subjected to oxidative stress," *Brain Research Bulletin*, vol. 45, pp. 517–520, 1998.
- [124] S. Damiano, T. Petrozziello, V. Ucci, S. Amente, M. Santillo, and P. Mondola, "Cu-Zn superoxide dismutase activates muscarinic acetylcholine M1 receptor pathway in neuroblastoma cells," *Molecular and Cellular Neurosciences*, vol. 52, pp. 31–37, 2013.
- [125] Q. J. Yu and Y. Yang, "Function of SOD1, SOD2, and PI3K/AKT signaling pathways in the protection of propofol on spinal cord ischemic reperfusion injury in a rabbit model," *Life Sciences*, vol. 148, pp. 86–92, 2016.
- [126] D. Davila and I. Torres-Aleman, "Neuronal death by oxidative stress involves activation of FOXO3 through a two-arm pathway that activates stress kinases and attenuates insulin-like growth factor I signaling," *Molecular Biology of the Cell*, vol. 19, pp. 2014–2025, 2008.
- [127] M. Nagai, D. B. Re, T. Nagata et al., "Astrocytes expressing ALS-linked mutated SOD1 release factors selectively toxic to motor neurons," *Nature Neuroscience*, vol. 10, pp. 615–622, 2007.
- [128] A. Gustin, M. Kirchmeyer, E. Koncina et al., "NLRP3 Inflammation is expressed and functional in mouse brain microglia but not in astrocytes," *PLoS One*, vol. 10, article e0130624, 2015.
- [129] E. Foran, A. Bogush, M. Goffredo et al., "Motor neuron impairment mediated by a sumoylated fragment of the glial glutamate transporter EAAT2," *Glia*, vol. 59, pp. 1719–1731, 2011.
- [130] K. Li, T. J. Hala, S. Seetharam, D. J. Poulsen, M. C. Wright, and A. C. Lepore, "GLT1 overexpression in SOD1(G93A) mouse cervical spinal cord does not preserve diaphragm function or extend disease," *Neurobiology of Disease*, vol. 78, pp. 12–23, 2015.
- [131] M. Rossi, S. Perez-Lloret, L. Doldan et al., "Autosomal dominant cerebellar ataxias: a systematic review of clinical features," *European Journal of Neurology*, vol. 21, pp. 607–615, 2014.
- [132] M. U. Manto, "The wide spectrum of spinocerebellar ataxias (SCAs)," *Cerebellum*, vol. 4, pp. 2–6, 2005.
- [133] Y. M. Sun, C. Lu, and Z. Y. Wu, "Spinocerebellar ataxia: relationship between phenotype and genotype - a review," *Clinical Genetics*, vol. 90, pp. 305–314, 2016.
- [134] H. Y. Zoghbi and H. T. Orr, "Pathogenic mechanisms of a polyglutamine-mediated neurodegenerative disease, spinocerebellar ataxia type 1," *The Journal of Biological Chemistry*, vol. 284, pp. 7425–7429, 2009.
- [135] D. M. Stucki, C. Ruegsegger, S. Steiner et al., "Mitochondrial impairments contribute to spinocerebellar ataxia type 1 progression and can be ameliorated by the mitochondria-targeted antioxidant MitoQ," *Free Radical Biology and Medicine*, vol. 97, pp. 427–440, 2016.
- [136] A. H. Hakonen, S. Goffart, S. Marjavaara et al., "Infantile-onset spinocerebellar ataxia and mitochondrial recessive ataxia syndrome are associated with neuronal complex I defect and mtDNA depletion," *Human Molecular Genetics*, vol. 17, pp. 3822–3835, 2008.
- [137] A. Federico, E. Cardaioli, P. PozzoDa, P. Formichi, G. N. Gallus, and E. Radi, "Mitochondria, oxidative stress and neurodegeneration," *Journal of the Neurological Sciences*, vol. 322, pp. 254–262, 2012.
- [138] R. Lodi, P. E. Hart, B. Rajagopalan et al., "Antioxidant treatment improves in vivo cardiac and skeletal muscle bioenergetics in patients with Friedreich's ataxia," *Annals of Neurology*, vol. 49, pp. 590–596, 2001.
- [139] M. P. Murphy, "Antioxidants as therapies: can we improve on nature?" *Free Radical Biology and Medicine*, vol. 66, pp. 20–23, 2014.
- [140] B. Uttara, A. V. Singh, P. Zamboni, and R. T. Mahajan, "Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options," *Current Neuropharmacology*, vol. 7, pp. 65–74, 2009.
- [141] Z. X. Yao, K. Drieu, and V. Papadopoulos, "The *Ginkgo biloba* extract EGb 761 rescues the PC12 neuronal cells from beta-amyloid-induced cell death by inhibiting the formation of beta-amyloid-derived diffusible neurotoxic ligands," *Brain Research*, vol. 889, pp. 181–190, 2001.



Hindawi
Submit your manuscripts at
<https://www.hindawi.com>

