



Review Article African Orphan Crops under Abiotic Stresses: Challenges and Opportunities

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A changing climate, a growing world population, and a reduction in arable land devoted to food production are all problems facing the world food security. The development of crops that can yield under uncertain and extreme climatic and soil growing conditions can play a key role in mitigating these problems. Major crops such as maize, rice, and wheat are responsible for a large proportion of global food production but many understudied crops (commonly known as "orphan crops") including millets, cassava, and cowpea feed millions of people in Asia, Africa, and South America and are already adapted to the local environments in which they are grown. The application of modern genetic and genomic tools to the breeding of these crops can provide enormous opportunities for ensuring world food security but is only in its infancy. In this review, the diversity and types of understudied crops will be introduced, and the beneficial traits of these crops as well as their role in the socioeconomics of Africa will be discussed. In addition, the response of orphan crops to diverse types of abiotic stresses is investigated. A review of the current tools and their application to the breeding of enhanced orphan crops will also be described. Finally, few examples of global efforts on tackling major abiotic constraints in Africa are presented.

1. Introduction

Every year, natural disasters such as drought, flooding, storms, and earthquakes cause considerable damage to animals, plants, and nature in general. According to the United Nations Food and Agriculture Organization, the agricultural sector absorbs 22% of the economic impact caused by natural hazards and disasters in the developing world as agriculture contributes up to 30% of the GDP of vulnerable countries [1]. Within agriculture, the crop sector suffers the most. From 78 disasters occurring from the years 2003 to 2013, the highest damage and losses to crops were 42% followed by those to livestock (36%) [1]. According to the same report, the three most important hazards to crops are floods (60%), storms (23%), and drought (15%). The 2004 Tsunami that impacted several Asian countries, especially Thailand, and the 2011 Tsunami in Japan contributed to large losses of the agricultural land. The Japanese Tsunami alone brought high levels of salt from the sea to the agricultural land, thereby considerably affecting rice (*Oryza sativa* L.) cultivation on over twenty thousand hectares of land due to high levels of soil salinity [2].

Changes in the amount and pattern of the annual rainfall have been negatively affecting the productivity of crops in African countries especially in Ethiopia [3, 4]. The periodically occurring El Niño weather system, formed by local warming of surface water temperatures across the Central and East-Central Equatorial Pacific Ocean, is the cause of most droughts in the Horn of Africa [5, 6]. The intensity of El Niño fluctuates from year to year and is periodically severe. For instance, the El Niño in 1991/92 caused drought in about 350 million ha of land while that of 1997/98 caused drought on only 80 million ha [6]. Since the peak of El Niño development coincides with the growing season of crops in Africa [5], the effect on food security is high. As drought is usually accompanied by famine, especially in the Least Developed Countries (LDCs), the occurrence of a single natural disaster does not only affect crop and livestock production but also

claims the lives of thousands or even millions of people depending on its intensity. A study in West Africa showed that the frequency of dry spells in a given year could be predicted from the annual rainfall in the previous years; hence, informed decisions can be made on the type of crop or cultivar to be cultivated and the appropriate management practices implemented [7].

Environmental stresses that cause damage to crops are broadly grouped into biotic and abiotic based on the cause of damage. While biotic stresses refer to constraints caused by living things such as pests, diseases, and weeds, abiotic stresses are those caused by climate and soil-related problems. This particular review focuses on the types and extent of damage caused by abiotic stresses and on the tools or techniques available to enhance the tolerance of indigenous crops to these environmental calamities and as a consequence to increase the productivity of crops. It gives due attention to African indigenous or orphan crops since these crops have not received much attention from the global scientific community. A changing climate, a growing world population, and a reduction in arable land devoted to food production are problems facing the world food security in the near future. The development of crops that can yield under uncertain and extreme climatic and soil growing conditions can play a key role in mitigating these problems. African indigenous crops can play a key role in this quest.

2. Significance of Orphan Crops in Africa

Orphan crops are also known as "neglected and underutilized species (NUS)" [60], "underutilized crops" [61], or "crops for the future" [62]. These crops are important for food security, nutrition, and income generation in many developing countries, but they have not been significantly researched [63–66]. They are often well-adapted to local growing conditions and fulfill the social and economic needs of local people and are often tolerant to many abiotic stresses compared to the world's major crops like wheat (*Triticum aestivum* L.) and maize (*Zea mays* L.) [66, 67].

2.1. Orphan Crops Are Extensively Cultivated in Africa. Orphan crops are staple foods for millions of people in the developing world, particularly in Africa. These crops are annually cultivated on large tracts of land and play a key role in the livelihood of the resource-poor farmers and consumers. Table 1 shows important orphan crops, including major crops and orphan crops, in terms of the total amount of area on which they are cultivated in Africa and the total production in Africa compared to the area and production in the entire world [8]. In addition, the distribution of land and production between major and orphan crop is shown.

These crops belong to the major categories of crops such as cereals, legumes, vegetables, root crops, and fruits and have diverse centers of origin. Although similar types of crops are cultivated in the rest of the world, Africa has unique crops which are cultivated and consumed solely on this continent. Africa's unique crops include cereals such as fonio [*Digitaria exilis* (Kippist) Stapf and *D. iburua* Stapf] in western Africa, and tef [*Eragrostis tef* (Zucc.) Trotter] in the Horn of Africa, and a legume called bambara groundnut [*Vigna subterranean* (L.) Verdc.] in southern and western Africa [8].

A great deal of diversity has been maintained by farmers for diverse types of orphan or underutilized crops. Efforts have been made by national and international institutions to collect from representative locations these valuable germplasms of diverse crops. Collected germplasms are conserved at facilities in national and/or international organizations including the Svalbard Global Seed Vault in Norway where 860,00 samples of different crop species are available [68]. The Southern African Development Community (SADC) Plant Genetic Resources Center based in Lusaka, Zambia, preserved over 18,000 collections of diverse crops from 12 of its 15 member states [69]. These large germplasm collections can be used as a reliable source for screening for traits of interest. The study of only 10% of the 5000 tef collections at the Ethiopian Biodiversity Institute shows a huge diversity in several important agronomic traits which includes grain yield, harvest index, and lodging percentage [70]. Although these and other landraces possess several desirable agronomic traits, only few traits in limited crop types have so far been harnessed.

Among the root crops grown in Africa, cassava (manioc; Manihot esculenta Crantz), yam (Dioscorea spp.), sweet potato [Ipomoea batatas (L.) Lam.], and enset [Ensete ventricosum (Welw.) Cheeseman] are the source of food for a large number of people. Cassava is a staple food for about 600 million people worldwide and for more than 200 million people in Sub-Saharan Africa [71]. In 2012, yam was cultivated globally on about five million hectares of which over 95% was in Africa [8]. Banana and plantain (Musa spp.) are among the major fruit crops grown in Africa. In the year 2012, about 16 million tons of banana and 27 million tons of plantain were produced in the continent [8]. Although a substantial amount of land in Africa is devoted to indigenous crops, the production constitutes a considerably low percentage of global production. For example, 10.1% of the global land devoted to barley (Hordeum vulgare L.) cultivation is in Africa, but the continent contributes only 4.5% of the world barley production (Table 1). This is because of the use of nonimproved cultivars as well as lack or minimal use of other agricultural inputs such as fertilizers and pesticides. The production per unit area of chickpea (Cicer arietinum L.), however, is slightly higher in Africa than in the rest of the world.

2.2. Orphan Crops Are Tolerant to Environmental Stresses. Table 2 shows some benefits of indigenous crops in terms of agronomy, nutrition, and health. Millets especially finger millet (*Eleusine coracana* Gaertn.), pearl millet [*Pennisetum* glaucum (L.) R. Br.], fonio, and tef, a tiny-seeded staple food in Ethiopia, are dominantly cultivated in semiarid areas of Asia and Africa due to their extreme tolerance to moisture deficit [72, 73]. While finger millet and pearl millet are cultivated in both continents, fonio is exclusively cultivated in West Africa. African rice (*Oryza glaberrima*), grown in West Africa, matures early allowing it to escape terminal drought. It is also resistant to diseases and pests and tolerates

ction. **	ha ⁻¹)	Africa		1.2
s of total produ	Yield (t ha ⁻¹)	Global		2.7
ortant crops in terms	Total production in Africa	(000' tons) % of global		4.5
top 10 globally impo	Total product	(000' tons)		6,022
vity. * indicates the	Area cultivated in Africa	000' ha) % of global		10.1
iction, and producti	Area cultiv	(000' ha)		4,997
l area of cultivation, total production, and productivity. * indicates the top 10 globally important crops in terms of total production. nd others. Source: [8, 9].	Contor of onivin	Cerner of origin		Ethiopia
ABLE 1: Major crops grown in Africa with their global area of efers to finger millet, pearl millet, and proso millet and others	Crop	Botanical name		Hordeum vulgare
TABLE 1: Major crops refers to finger millet		Common name	Cereals	Barley*

	Crop	Center of origin	Area cultive	Area cultivated in Africa	Total production in Africa	ion in Africa	Yield (t ha ⁻¹)	t ha ⁻¹)
Common name	Botanical name		(000' ha)	% of global	(000 ⁷ tons)	% of global	Global	Africa
<i>Cereals</i> Barlev [*]	Hordeum vulgare	Ethiopia	4,997	10.1	6,022	4.5	2.7	1.2
Fonio	Digitaria exilis	· · · J - · · · · ·	566	100	587	100	1.0	1.0
Maize*	Zea mavs	Mexico & Central America	33,710	19.0	69,636	2.9	4.9	2.1
Millets ^{**}	Diverse species	Ethiopia	19,977	62.9	16,083	53.8	0.9	0.8
Rice*	Orvza sativa	India	10,538	6.4	26,823	3.7	4.4	2.5
Sorøhum	Sorohum hicolor	Ethionia	23.169	60.7	23.312	40.9	L.	1.0
Tef	Eraorostis tef	Ethionia	3.000	100	4.700	100	1.6	1.6
Wheat*	Triticum spp.	Middle East	10,216	4.7	24,710	3.7	3.1	2.4
Food legumes (pulses)								
Bambara beans	Vigna subterranea		232	100	159	100	0.7	0.7
Beans, dry	Diverse species	Diverse	7,829	26.7	5,069	21.5	0.8	0.6
Chickpea	Cicer arietinum	Central Asia	593	4.8	696	5.9	0.9	1.2
Cowpea	Vigna unguiculata	India	10,402	92.1	5,402	94.5	0.5	0.5
Pigeon pea	Cajanus cajan	India	754	14.1	632	14.9	0.8	0.8
Vegetables								
Cabbages	Brassica spp.	China	124	5.2	2,805	4.0	29.3	22.6
Okra	Abelmoschus esculentus	Ethiopia	516	47.4	1,833	21.9	7.7	3.5
Tomato*	Lycopersicon lycopersicum	Mexico & Central America	1,010	21.0	17,937	1.11	33.7	17.8
Un seeas					6		,	
Castor seed	Kicinus communis	Ethiopia	097	14.8	06	4.6	1.1	0.3
Sesame	Sesamum indicum	Central Asia	3,283	41.5	1,708	42.3	0.5	0.5
Soybean*	Glycine max	China	1,637	1.5	1,986	0.8	2.3	1.2
Sunflower	Helianthus annuus	North America	1,790	7.2	2,223	5.9	1.5	1.2
Roots & tubers								
Cassava*	Manihot esculentum		13,662	67.0	149,479	52	12.9	10.9
Enset	Ensete ventricosum		312	100	728	100	2.3	2.3
Potato*	Solanum tuberosum	North America	1,894	9.6	28,169	7.7	18.9	14.8
Sweet potato*	Ipomoea batatas	Mexico & Central America	3,506	43.3	17,999	17.4	12.7	5.1
Taro	Colocasia esculenta	India	1,127	85.7	7,367	73.8	7.6	6.5
Yam	Dioscorea spp.	India	4,801	95.3	56,500	96.1	11.7	11.7
Fruits								
Banana*	Musa spp.	India	1,493	30.1	15,863	15.5	20.6	10.6
Mango	Mangifera indica	India	691	13.4	7,135	16.9	8.1	10.3
Papaya	Carica papaya	Mexico & Central America	134	30.8	1,330	10.7	28.5	9.8
Plantain	Musa spp.		4,368	80.8	26,545	71.4	6.9	6.1
Others								
Cocoa	Тнеоbroma сасао	Mexico & Central America	6,337	63.8	3,289	65.7	0.5	0.5
Coffee	Coffea snn	Ethionia	1 000	10.0	1 000	11 2		ц С

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Beneficial trait	Crops	Reference
	Agronomy-related	
Drought-tolerant	African rice, bambara groundnut, cassava, cowpea, enset, Ethiopian mustard, grass pea, pearl millet, tef, yam	[10-16]
Waterlogging tolerant	Tef	[17]
Salinity-tolerant	Sorghum (moderately tolerant)	[18]
Heat-tolerant	Cow pea, pearl millet	[19]
Adaptation to poor soil	African rice, cassava, cowpea	[12, 14, 19]
Early maturing (drought escape)	African rice, amaranth, fonio	[12, 13]
Erosion control (fast ground cover)	Cowpea	[20]
Disease tolerance	African rice, Ethiopian mustard, tef	[12, 16, 17]
Pest tolerance	African rice, Ethiopian mustard, tef	[12, 16, 17]
	Nutrition-related	
Complete food (protein, carbohydrate, fate)	bambara groundnut	[13]
Rich in essential amino acids	Amaranth, bambara groundnut, cowpea, finger millet, fonio, grass pea	[10, 12, 13, 21]
Rich in iron	Banana, fonio, tef	[12, 22]
High vitamin A content	Banana	[23]
High oil content	Noug [Guizotia abyssinica (L.f.) Cass.]	[24]
	Health-related	
Gluten-free	Tef, millets	[25, 26]
Low glycemic index	Finger millet	[27]
Anticancer property	Millets	[28]

TABLE 2: Diverse types of benefits related to orphan African crops.

fluctuations in growing conditions [45]. Tef is moderately tolerant to moisture scarcity and, in addition, it resists poorly drained soils, conditions that crops such as maize and wheat cannot withstand [74]. Noug (*Guizotia abyssinica* Cass.), an oilseed crop related to sunflower, grows best on poorly drained, heavy clay soils [75].

Legumes such as cowpea [*Vigna unguiculata* (L.) Walp.], bambara groundnut, and grass pea (*Lathyrus sativus* L.) tolerate extremely low soil moisture [10, 76]. Cowpea is the most widely cultivated food legume in Africa. In the year 2012, it was cultivated globally on 11.2 million ha of land of which 98% was in Africa, particularly in three countries (i.e., Niger; Nigeria; and Burkina Faso), which account for 82% of the total land devoted to this crop [8]. Cow pea is tolerant to drought, heat, and soils with low levels of organic matter and phosphorus [19]. Grass pea is considered as an insurance crop in Ethiopia as it produces reliable yields when all other crops fail.

Among the root crops grown in Africa, cassava, yam, sweet potato, and enset are the source of food for millions of people. Cassava is tolerant to drought and also performs better than other crops on poor soils. Enset, which is related to banana but has an underground corm as the edible part, is a major food source for over 10 million people in the densely populated regions of Ethiopia due to its extreme tolerance to drought and adaptation to diverse soil types [11].

2.3. Orphan Crops Provide Nutritious and Healthy Food

2.3.1. Nutrition-Related Benefits. Nutritional benefits of some orphan crops are indicated in Table 2. Orphan crops often provide more nutrients than the foods that dominate global production. African cereals, particularly millets, contain high amounts of vitamins, calcium, iron, potassium, magnesium, and zinc [77]. The seeds of finger millet contain valuable amino acids especially methionine [12], which is lacking in the diets of hundreds of millions of Africans who live on starchy staples such as cassava. The seeds of fonio are nutritious especially in amino acids such as methionine and cysteine [78] which are essential for human health but deficient in major cereals such as wheat, rice, and maize [21]. Amaranth (Amaranthus spp.) which is largely consumed in Togo, Liberia, Guinea, Benin, and Sierra Leone, has a relatively high protein content and a balanced composition of essential amino acids and minerals [79].

While the grain is the main source of food for humans, crop residues, particularly the straw from cereals, are an invaluable source of livestock feed. The straw from tef is more palatable and nutritious than those of wheat and barley and, hence, it fetches higher prices [80].

Legumes such as cow pea, bambara groundnut, and grass pea are the major source of protein for consumers. Due to their ability to fix atmospheric nitrogen and convert it into a form usable for plants, legumes also contribute towards improving soil fertility. Bambara groundnut is grown for human consumption and is the third most important grain legume in Africa after cowpea and groundnut (*Arachis hypogaea* L.) [76]. The seeds of bambara groundnut are considered to be a complete food because they contain adequate quantities of protein (19%), carbohydrate (63%), and fat (6.5%) [13].

Banana, especially the orange-pulped type with high carotenoid and iron content, could reduce Iron Deficiency Anemia (IDA) by over 50% and also Vitamin A Deficiency (VAD) in East Africa, where both IDA and VAD affect a large number of people [22]. The seed oil content of noug ranges from 39.8% to 46.9% oil [75]. The fatty acid content of the oil is similar to the oils of the sunflower family with linoleic acid being the dominant oil.

2.3.2. Health-Related Benefits. The benefits of some orphan crops as health-food are shown in Table 2. Finger millet is a popular food among diabetic patients because of its low glycemic index and slow digestion [27]. Finger and pearl millets were shown to have anticancer properties and might have potential in the prevention of cancer initiation [28]. This antiproliferative property is associated with the presence and content of phenolic extracts. Tef, sorghum, and millets are considered to be a healthy food since the grain does not contain gluten [25, 81, 82], the cause of celiac disease.

2.4. Orphan Crops Are Compatible with the Socioeconomic Conditions. Agriculture provides more than just food production—it affects nonfood products, environmental and land management, economic development, employment opportunities, social stability, and maintenance of cultural tradition and identity. Recognition of the many social and economic functions of agriculture has resulted in increased efforts of developing land use practices that support the functions in an integrated manner [83].

In southern Senegal, African rice is thought to have been domesticated 2000–3000 years ago. The Jola people of Senegal still cultivate this rice and choose the varieties to be planted based on maturation time (fast or slow maturing or both to stagger the harvest time), plant height (a tall or short variety based on harvest considerations), and soil type (the soil composition and the amount of irrigation which is particular to each field) [45]. Likewise, some fonio varieties produce grain in only 6 to 8 weeks after planting. Such varieties extend the period of time for which food is available and can bridge the gap between the last and the next harvest. Planting crops with varying maturation times increases the chances of harvesting under challenging environmental conditions [12].

In addition to adaptation to local environmental conditions, orphan or indigenous crops are preferred because of their use in preparing local recipes. For instance, tef, which is the staple food for over 60 million people in Ethiopia, is the most preferred grain since it makes the best quality spongy bread called *injera*. Compared to *injera* made from tef, those made from sorghum and maize have poor consistency and short shelf-life and thus have low acceptance by consumers. Due to these desirable properties, the grain of tef fetches higher price than that of other crops.

Similarly, indigenous crops are compatible to the agroecology of the region in which they are cultivated. The study in Northwestern Ethiopia showed that when a widely cultivated indigenous tef crop was replaced by exotic maize, the incidence of malaria increased by about 10-fold [84–86]. This was mainly due to a higher survival rate of mosquito larvae fed on maize pollen than of those fed on tef pollen.

3. Challenges of Abiotic Stresses to Orphan Crops

Although indigenous crops possess a variety of desirable traits, they are under continuous pressure to detect and adapt to the environmental constraints prevalent on the continent [65]. Orphan crops are normally cultivated in marginal environments in terms of climate and soil. For instance, millets are crops of dry and hot climate while tef is cultivated on poorly drained Vertisols in the highlands of Ethiopia where other crops fail to survive. Hence, harvests from these crops are mostly suboptimal. These constraints are broadly grouped into biotic and abiotic stresses. Biotic factors such as weeds, diseases, and insect pests cause significant crop damage in Africa due to environments conducive to these crop nuisances. The current review, however, focuses on abiotic stresses, which cause huge losses to the biomass of these crops, especially in relation to the types and extent of damage they cause and the tools developed to enhance the tolerance of crops against this damage.

Yield loss estimates have been documented for major crops. According to a website tabulating losses to these crops, yield losses due to abiotic stresses range from 50% for sugar beet to 82% for wheat [87]. However, information on orphan crops is limited.

3.1. Drought. A considerable amount of water is used in both rain-fed and irrigated agriculture. Irrigated agriculture accounts for about 60 percent of production in the developing countries. Although the global water withdrawal from rivers, lakes, and aquifers for agriculture is estimated at 2,000 to 2,500 km³ per year, only 900 km³ of this is used by food crops while the remaining is lost to evaporation, deep infiltration, or the growth of weeds [88]. Countries with an extensive area under irrigation utilize large amount of water. For instance, from the total water consumption in Egypt 85.9% is for irrigation, with 2.6% for industrial and the remaining 11.5% for municipal uses [89]. Since data for the majority of African countries is not available, it is difficult to know the countries which utilize small proportion of their water for agriculture. Although agriculture requires a large amount of water, the scarcity of moisture as it frequently occurs in most African countries is critical to increasing production and productivity of crops.

Semiarid and arid regions in Africa are fragile to sustain the ecosystem mainly due to the scarcity of moisture. About 40% of the population in the Sub-Saharan Africa live in arid to dry environment and the population is expected to double in the dry areas by 2050 [90]. According to these authors, crop failure in Africa is mostly not due to scarcity of precipitation but due to dry spells which occur at critical growth stages particularly during flowering.

A study on proso millet (*Panicum miliaceum*), little millet (*Panicum sumatrense*), foxtail millet (*Setaria italica*), and wild millet (*Setaria glauca*) showed that drought treatment before flowering contributed to a significant yield reduction in these four millets [91]. A similar study on two finger millet landraces indicated that drought occurring four weeks after sowing resulted in a complete yield loss [92].

In pearl millet, terminal drought occurring between the flowering and the crop maturity stages is the most critical as it resulted in 60% yield loss [93]. Yield losses due to drought were estimated to be 40% in tef [94], 51% in pearl millet, and 57% in bambara groundnut [95] at 50% moisture treatment.

Most orphan cereal crops in Africa such as millet and tef are C_4 plants which efficiently utilize moisture; hence, they generally outperform C_3 plants in hot and dry climates. A study in the Sahel region where the annual precipitation is as low as 200 mm shows that sorghum and millet had similar water use and biomass production, hence similar biomass per unit of evapotranspiration [96].

Although reports are not available for indigenous crops, the study on wheat indicated that moisture scarcity affects not only crop productivity in terms of grain yield but also the characteristics of grain including protein quality, gluten level, and dough strength [97, 98].

3.2. Waterlogging. Waterlogging is among the major constraints to increasing crop productivity particularly in areas which receive high precipitation and have agriculturally problematic soils. Since soil pores during waterlogging are filled with water, the diffusion of gases is hampered resulting in anaerobic conditions which might also be accompanied by the accumulation of toxic compounds. As a result, the normal functioning of stomata, photosynthesis, and roots are severely affected [99].

Waterlogging especially on *Vertisols*, the black clay soils with high water-holding capacity, restricts the cultivation of most major crops such as maize and wheat. About 43 million ha of *Vertisols* exists in 28 countries in Africa [100]. A study on millets indicated that waterlogging treatment from two weeks after sowing to crop maturity reduced the grain yield of proso millet and wild millet *(Setaria glauca)* by 16% and 18%, respectively [101].

In addition to hindering the aeration of the root systems due to poor drainage, *Vertisols* are less suitable for the cultivation of root crops such as groundnut since pulling of the pods from the soil during harvesting is hindered by the hardy nature of the soil upon drying.

3.3. Soil Acidity. An estimate of the total amount of land with acidic soil in the world is about 30% with about 50% of the arable land being acidic [102]. In acidic soils, aluminum toxicity is the most important factor affecting crop productivity since aluminum under a soil pH below 5 is

solubilized into Al³⁺ which is toxic for plant roots, inhibiting their function and development [102]. A general rule is that aluminum concentrations between 2 and 5 ppm are toxic to aluminum-sensitive plants while concentrations above 5 ppm are toxic to tolerant species. Toxic levels of aluminum affect root growth dramatically and stunted growth, small grain size, and poor yield are the results of the consequent lack of nutrition [103]. The effects of Al-toxicity on cereal crops were recently reviewed [104].

3.4. Soil Salinity. Soil salinity, characterized by a high concentration of soluble salts, affects crop productivity although some indigenous crops have some level of tolerance to this abiotic stress. Soil salinity is a broad term which refers to three types of agriculturally problematic soils. These are (i) saline soil: with electrical conductivity, ECe > 4 dS/m; exchangeable sodium percentage, ESP < 15; and pH < 8.5; (ii) sodic soil: ECe < 4 dS/m, ESP > 15, and pH > 8.5; and (iii) sodic-saline soil: ECe > 4 dS/m, ESP > 15, and pH < 8.5. Chloride and sulphate salts of sodium, magnesium, and calcium are the most common soluble salts.

It is estimated that approximately 4.8% of the land in Africa is considered to be affected by salinity or sodicity [105]. Irrigated lands in the Mediterranean region have been converted into extensive salt accumulation where 60% of the farm area in Egypt is affected by saline and sodic-saline conditions [106]. Although salinity occurs due to diverse causes, the main cause of the elevated salinity level in the Mediterranean area is the use of highly saline irrigation water in the hot and dry environment where evapotranspiration is high [106].

Irrigation in arid and semiarid climates is associated with saline soils as almost all water contains some salts and, after high evapotranspiration, the salt remains on the surface of the soil and/or root zone of crops. The accumulation of salts depends on the quality of the irrigation water, the irrigation management, and the drainage of the soil. Often salinity problems are associated with poorly drained and waterlogged soils as excess water allows the salt to rise to the root zone via capillary action [107]. Once salt has accumulated, only additional water can leach the salts away from the root zone. The study in tef showed that saline soil causes up to 93% yield loss in tef [108].

3.5. Heat (High Temperature). Heat causes a multitude of molecular, cellular, and physiological changes to plants. Among these, effects on respiration and photosynthesis are the major ones affecting crop productivity [109]. Extraordinary heat waves have already caused significant yield losses across the globe and pose a dire threat to global food security. Climatological extremes in general have a negative effect on plant growth and can lead to catastrophic losses in crop yields. Studies on major crops showed that, with heat, yields increase until a crop-dependent threshold (29°C for maize, 30°C for soybeans, and 32°C for cotton) but temperatures above these thresholds become very harmful [110]. Once the threshold is passed, the potential for adaptation within the crop is limited. It is estimated that a temperature increase of 3 to 4°C could

cause crop yields to fall by 15–35% in Africa and Asia [109]. Modeling suggests that cereal production in Southern Africa and Southern Asia is most likely to be affected by climate change particularly by global warming [111].

3.6. Cold and Frost. One-third of the total global land is icefree while 42% of land has temperatures below -20° C [112]. In these cold regions, plants have specialized adaptations that help them to survive freezing temperatures. The amount of damage caused by freezing depends on the temperature and duration of the cold stress as well as the developmental stage of the plant. Damage from cold particularly from frost is common in the highlands of the eastern Africa especially when it coincides with the growing season of crops. Although climate change is warming rather than cooling the planet, warm winters can lead to early blooming of perennials and consequent frost damage when cold conditions occur in late spring [113].

4. Mechanism of Crop Tolerance to Abiotic Stresses

Understanding the response of plants to resist and tolerate abiotic stresses is important for devising breeding strategies to ensure productivity under all conditions. Diverse types of mechanisms are involved in the abiotic stress tolerance of plants [114, 115]. Mechanisms employed by some indigenous crops towards adaptation and/or tolerance to diverse types of abiotic stresses are shown in Table 3. Some of these mechanisms against major abiotic stresses which affect crop productivity in Africa are briefly indicated below.

4.1. Drought Tolerance. Major strategies and/or mechanisms of drought tolerance or adaptation of millets that are mostly cultivated in drought-prone areas of the world were recently reviewed [72]. Four of these mechanisms are as follows: (i) *drought escape* which refers to the condition in which plants reach maturity before the drought occurs; (ii) *drought avoidance* which refers to the ability of the plant to maintain a favorable water balance under moisture stress in order to avoid water deficit in the plant tissue; (iii) *drought tolerance* which refers to the ability of the plant to produce some yield by withstanding low water potential; and (iv) *drought recovery* which refers to a condition in which plants recover from the adverse effects of drought in order to provide some yield and/or biomass.

Most African orphan crops are cultivated in a marginal environmental where moisture is limited. However, they are adapted to efficiently utilize the scarce moisture in order to sustain their growth and produce at least some yield for farmers desperately waiting for them. These neglected crops guarantee future food security either directly as alternative crops in drought-prone areas or indirectly as germplasm resources for crop improvement [116].

Several morphological and physiological traits are involved in drought tolerance of plants [117]. In little millet (*Panicum sumatrense* Roth), osmotic adjustment, which refers to the maintenance of water potential during moisture stress, and ROS (reactive oxygen species) scavenging systems are employed to protect the plant from drought [31]. Drought tolerance is known to be associated with the tensile strength of the stem of *Eragrostis* species of which tef is a member [32]. Roots also contribute to drought tolerance, and especially the efficiency of the uptake of water during reproduction and grain filling is critical since moisture scarcity during these developmental stages is detrimental to the survival of the plant [118]. The expression of dehydrin in wheat has been correlated with the acquisition of drought tolerance at early growth stage [119].

4.2. Waterlogging Tolerance. Plants respond to waterlogging with a variety of mechanisms [120]. In waterlogged soil, the diffusion of gases to and from the roots is severely limited. Oxygen at the root surface is severely decreased and toxic byproducts are unable to diffuse away from the plant. Many aspects of the plant response are a direct consequence of anoxia, a complete lack of oxygen or hypoxia, and reduced oxygen. One of the responses of plants to low oxygen is to change from aerobic to anaerobic metabolism as has been found for finger millet [37]. Though much less efficient than aerobic metabolism, the production of ATP by glycolysis can provide the plant with ATP for a short period of time. This requires an abundant supply of soluble sugars and thus changes in carbohydrate metabolism are often found in tolerant species. Increases in antioxidants are also observed in tolerant plants such as pigeon pea [36].

Changes in the cellular structure of the plant are also a response to waterlogging. For example, aerenchyma, air spaces in spongy tissue that let gases diffuse from the stem to the roots, is often formed in plants tolerant to waterlogging stress. These air spaces are generated either with (lysigenous) or without (schizogenous) cell death. Waterlogged sunflower developed lysigenous aerenchyma within two days of the application of stress [33]. Another strategy found in waterlogging tolerant species is the formation of adventitious roots when stressed [120]. Both sorghum [34] and finger millet [35] have been shown to form adventitious roots when waterlogged. A strategy to escape the flood via fast growth has been shown for mung bean [38]. In addition, plants tolerant to waterlogging possess high amount of soluble sugars [121]. In tef, the occurrence of nitrogen reductase activity in the shoots rather than in the roots has been implicated in its waterlogging tolerance.

4.3. Soil Acidity Tolerance. Two general mechanisms of aluminum (Al) tolerance exist in plants. These are (i) Al exclusion, which refers to the prevention of Al from entering the root apex, and (ii) Al tolerance, in which Al is allowed to enter the plant but detoxified [122]. The Al exclusion mechanism is also described by the exudation of organic acids or the release of phenolic compounds as well as related products which have the ability to chelate Al^{3+} ions in the rhizosphere [122].

The MATE family of proteins are transporters of metabolic and xenobiotic organic cations. Members of this family responsible for malate and citrate efflux have been

Abiotic stress	Trait	Response to stress	Crop	Reference
	Seed yield	High yield for drought-tolerant genotypes	Pearl millet	[29]
	Flowering time	Adjust phenology to rainfall pattern	Pearl millet	[30]
	Shoot length	Decreased under drought	Little millet	[31]
Drought	Root length	Increased under drought	Little millet	[31]
	Leaf tensile strength	Increased in drought-tolerant plants	Tef	[32]
	Water extraction	Less extraction before flowering; more extraction after flowering	Pearl millet	[29]
	Chlorophyll content	Decreased under drought	Little millet	[31]
	Coll[Aerenchyma formation	Sunflower	[33]
	Cellular structure	Formation of adventitious roots	Sorghum, finger millet	[34, 35]
Waterlogging	Matchelican	Generation of antioxidants	Pigeon pea	[36]
	INTELADOIDSIII	Change to anaerobic metabolism	Finger millet	[37]
	Plant height	Fast growth	Mung bean	[38]
Uroot	Growth	Growth and nutrient uptake	Chickpea, Pearl millet	[39, 40]
ITCAL	Yield	Unknown	Cowpea	[41]
Cold and funct	0.000+h	Early flowering, protracted rosette	Faba bean	[42]
COIN AIIU II USI	TIDWOID	Early flowering	Sorghum	[43]

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implicated in aluminum resistance in wheat [123], sorghum [124], and barley [125]. An aluminum susceptible cultivar of barley was made tolerant after transformation with the TaALMT1 gene from wheat [126]. Nine barley transformant lines with the TaALMT1 gene also showed increased malate efflux and aluminum resistance and some transformants surpassed the original line in aluminum toxicity resistance [127].

4.4. Soil Salinity Tolerance. Three mechanisms of plant tolerance to soil salinity were reported. These are (i) tolerance to osmotic stress which reduces cell expansion in root tips and young leaves which results in stomatal closure; (ii) Na⁺ exclusion from leaf blades in order to remove the toxic effect of Na+; and (iii) tissue tolerance to accumulated Na⁺ [128].

Management of saline soils includes the provision of appropriate drainage, controlling irrigation, and the use of deep-rooted plants, which maintain the water table at a level appropriate for the land. Considerable improvements in growth and yield have been obtained by either priming seed with chemicals or extreme temperatures before planting or by applying exogenous chemicals to plants growing under salt stress. Efforts in breeding salt-tolerant cultivars are being pursued [129]. The use of saline tolerant crops both increases the amount of land available for growing and reduces the quality of the water that must be used for irrigation. Crops described as salt tolerant include date palm, asparagus, barley, and salt grass, while moderately tolerant species include olive, beet, sorghum, wheat, and rice [18].

Plant microRNAs are known to be involved in diverse abiotic stresses. A list of microRNAs from model and crop plants with differential expression under saline condition was recently documented [130].

4.5. Heat Tolerance. Heat stress is a major concern for crop production and much work is being done to develop plants that can still maintain yield under hot conditions. It is known that heat stress interferes with photosystem (PS) II, increases the amount of reactive oxygen species (ROS), and reduces electron transport [131]. Plants have evolved a number of adaptive, avoidance, and acclimation mechanisms to cope with heat stress.

Tolerance mechanisms involve heat shock proteins, ion transporters, osmoprotectants, antioxidants, transcription factors, and signaling molecules [132, 133]. Lipid membranes are susceptible to lipid peroxidation, and membrane thermal stability can be a selection criterion for heat stress tolerance as has been used in wheat [134]. Reactive oxygen species (ROS) are normal byproducts of cell metabolism that can be toxic at high concentrations. Many plants have systems containing antioxidants to combat ROS as protection against a wide variety of abiotic stresses.

Heat tolerant accessions have been found for pearl millet [40], chickpea [39], and cowpea [41]. A study in chickpea identified 18 SNPs from five genes (ERECTA, ASR, DREB, CAP2 promoter, and AMDH) that were significantly associated with different traits under heat and drought stress [39].

4.6. Cold or Frost Tolerance. Many plants increase their tolerance to cold if they are allowed to acclimate to it in low, nonfreezing temperatures. For example, rye exposed to a period of low nonfreezing temperatures can withstand temperature as low as -30° C, while the same rye without the acclimation period dies at -5° C [135]. The acclimation mechanisms include the expression of genes that function to protect membranes, the primary site of freezing injury, such as dehydrins that prevent ice nucleation [136]. In addition, CBF/DREB1 proteins, a family of transcription factors in *Arabidopsis*, control a group of genes that confer freezing tolerance [135].

Cold tolerance is the ability to grow and produce a satisfactory yield under relatively cold (but not freezing) conditions. Several sorghum accessions from Ethiopia and Uganda have been shown to be cold tolerant during both the reproductive and the vegetative stages [43]. The mechanism for this tolerance has, however, not been investigated.

Seed yield in chickpea can be increased fourfold by sowing in early winter as opposed to traditional spring sowing. However, this increases the probability of exposure to cold, necessitating cold-tolerant lines. Screening of 3276 chickpea germplasm accessions and breeding lines identified 21 cold-tolerant lines [137]. When exposed to cold stress in the reproductive phase, cold-tolerant chick peas produced viable seeds, while cold susceptible cultivars suffered from abortion due to pollen sterility [137]. The analysis of the bambara groundnut transcriptome obtained from suboptimal low temperature of 18°C showed that transcription factors such as MYB, NAC (NAM, ATAF, and CUC), and WRKY are involved in response to the stress [138].

5. Techniques Enhancing Abiotic Stress Tolerance in Orphan Crops

The application of modern genetic, genomic, and agronomic tools to the improvement of indigenous crops can provide enormous opportunities for ensuring global food security. A recent review indicated that there are large gaps between farmers' average yield and potential yield for several African crops [66] indicating that productivity of these crops could be boosted using improved cultivars and proper crop management practices. Recent reviews have suggested several efficient breeding and genomics techniques with immense potential in the development of crops tolerant or resistant to abiotic stresses [139–142]. Table 4 summarizes some techniques with a high potential for the improvement of African orphan crops.

5.1. Genetic Tools. Diverse types of genetic and breeding techniques and their applications to African orphan crop improvement were earlier reviewed [65]. Only widely implemented techniques are briefly presented below.

5.1.1. Hybridization. Hybridization refers to crossing two plants of the same or related species in order to create genetic variability which is utilized to improve trait(s) of choice. Successes in hybridization resulted in semidwarf cultivars

Technique	Improved trait	Crop	Reference
	Breeding & genomics		
Selection/screening	Soil acidity	Tef	[44]
Hybridization (intra- & interspecific)	Drought tolerance	NERICA rice	[45]
Mutation breeding	Diverse traits	Tef	[46]
Marker-assisted method	Drought tolerance	Pearl millet	[47]
TILLING (Targeting Induced Local Lesions IN Genomes)	Several traits	Chickpea, millets, tef	[48, 49]
	Agronomy & physiology		
Conservation tillage	Drought tolerance	Sorghum and chickpea	[50]
Biochar application	Soil acidity tolerance; improved soil properties	Tef & other crops	[51, 52]
BBF (broad-beds & furrows)	Improved drainage	Faba bean and tef	[53]
SCI (System of Crop Intensification)	Several	Finger millet, tef, mustard	[54]
Plant hormones	Salt & cold tolerance	Several crops	[55, 56]

TABLE 4: Major techniques applied to develop abiotic stress-tolerant African orphan crops.

of wheat and rice, which boosted the productivity of both crops during and after the famous Green Revolution [143]. The major breakthrough in hybridization in Africa was the interspecific crossing between the Asian rice (Oryza sativa L.) and the African rice (Oryza glaberrima Steudel), which is widely cultivated in West Africa. This led to the development of the popular NERICA (New Rice for Africa) varieties. NERICA harbors desirable properties from its two parents: high grain yield and protein content from the Asian rice and tolerance to drought and poor soil fertility as well as early maturity from the African rice [45]. Due to its early maturing property, African rice escapes terminal drought, which normally occurs after the flowering stage. Hence, it is the source of food during food shortage particularly just before other crops are harvested. NERICA is currently cultivated in several African countries [144].

5.1.2. Mutation Breeding. This refers to the application of either physical (such as gamma-ray and X-ray) or chemical (e.g., ethyl methane sulfonate or EMS) mutagens to alter the composition or arrangement of DNA of an organism so that mutants harboring traits of choice are selected and bred to popular cultivar(s). EMS, which mainly creates point mutations, is the most desirable mutagen as it results in a single nucleotide lesion in the gene of interest. Most mutation breeding programs aim at altering traits such as plant height and disease resistance in well-adapted plant varieties of rice, barley, and wheat. Mutation breeding is a cornerstone for releasing over 3000 globally known crop varieties including some varieties from African indigenous crops [145]. Mutation breeding and molecular breeding techniques have been successfully implemented to develop salt-tolerant crops [129].

5.1.3. Plant Cell and Tissue Culture. Tissue culture techniques or *in vitro* regeneration have been widely utilized for several indigenous crops especially in tree crops due to a number of benefits. Some of these benefits are as follows: (i) a large number of planting materials can be generated from small area; (ii) planting materials are maintained free of diseases particularly from viruses; and (iii) the regeneration process is made all year round irrespective of climatic conditions since regenerates are grown in greenhouses under controlled environmental conditions. An efficient regeneration method resulting in plantlets has been developed for wild banana [146].

5.1.4. Marker-Assisted Selection (MAS). This refers to the utilization of molecular markers in breeding for traits of interest. When markers are found that are linked to genes controlling traits, the marker (not the gene) can be used to identify and select breeding stock. Markers are selected that are easily identifiable with common laboratory tests. Commonly utilized markers in crop breeding are SSRs (Simple Sequence Repeats, or microsatellites) and SNPs (Single Nucleotide Polymorphisms). SSR refers to a repeat of two to six nucleotides in the DNA sequences while SNP refers to a single nucleotide polymorphism. Other recently developed SNP-based methods include GBS (genotyping-bysequencing) [147] and GWAS (Genome-Wide Association Studies) [148]. The potential application of GBS was recently investigated in several crops with different genome sizes and breeding systems [149]. In breeding for abiotic stress-tolerant crops, GWAS was efficiently applied in identifying aluminum resistance wheat cultivars [150].

5.1.5. Transgenesis and Genome Editing. The transgenic method is the fastest adopted agricultural technology due to its high acceptance by farmers. Since the release of the first commercial transgenic crops, the annual cultivated area by transgenic crops has increased 106-fold and the production volume has increased from just 1.7 million tons in 1996 to 181 million ha in 2014 [151]. The development of cisgenesis involves a system in which plant-specific promoters are used to drive the gene of interest, instead of foreign promoters

from bacteria or other organisms in case of transgenesis [152]. Genome editing especially the CRISPR/Cas9 (clustered regularly interspaced short palindromic repeats or CRISPR-associated protein 9) is an efficient and specific technique targeting multiple sites simultaneously [153, 154]. The manipulation of signaling pathways or regulatory mechanisms involved in the protection of plants against drought through this transgenic method was effective in developing plants for this particular abiotic stress [155–158].

5.1.6. High-Throughput Techniques. TILLING (Targeting Induced Local Lesions IN Genomes) and EcoTILLING which screen mutagenized and natural populations, respectively, have been instrumental in identifying mutants of interest within a short period of time [159]. Both TILLING and EcoTILLING have been applied to the improvement of indigenous crops such as tef [48, 160].

5.2. Omics Tools. Omics techniques refer to the use of genomic, transcriptomic, proteomic, and related techniques for improving crops. Omics tools provide useful information regarding the response and adaptation of plants to abiotic stresses [161, 162]. Genomic tools have proven to be robust in developing crop varieties with improved traits [163]. A list of genes known to be involved in diverse abiotic stresses and crops has been documented for various crops [164, 165]. The genome sequencing of an orphan crop tef has been utilized in the investigation of drought and nutrition-related genes [166].

Some genes are known to be involved in multiple abiotic stresses tolerance. Among these, NAC transcription factor enhances both drought and salt tolerance [167, 168], while RCI2/PMP3 (which encodes small membrane proteins of the PMP3 family) and bZIP (Basic leucine zipper) transcription regulate several abiotic stresses [169, 170].

RNASeq, quantitatively sequencing mRNA using nextgeneration sequencing technologies, has in recent years become a standard; for example, gene expression profiling of chickpea responses to drought, cold, and high-salinity has been performed [171, 172].

MicroRNAs, a class of small noncoding RNAs that are between 21 and 24 nucleotides long, are associated with responses to diverse abiotic stresses including drought, salinity, extreme temperatures, nutrient deprivation, and heavy metals [173]. miR169 is one of the largest miRNA families that is conserved in all plant species and significantly contributes to proper plant development and in plant response to environmental stress [174]. The study in foxtail millet (Setaria italica) showed that, after exposing the plant to moisture scarcity, 32 dehydration-responsive miRNAs were upregulated in tolerant cultivar and 22 miRNAs were downregulated in sensitive cultivar, suggesting that miRNAmediated molecular regulation might play important roles in providing contrasting characteristics to these cultivars [175]. Successes in developing abiotic stress tolerance in crop plants using either the overexpression or downregulation of the miRNAs and their targets were reviewed [173, 174]. Although reports for African indigenous crops are not yet available,

microRNAs induced in drought have been reported for rice [176].

Proteomic tools have also been implemented to study drought tolerance in rice [177] and salinity and drought tolerance in cereals [178].

5.3. Agronomic Tools. Throughout the world, roughly 1.5 billion people live in risk-prone, marginal environments and are not using modern agricultural technologies. New management systems that can adapt to local, highly variable small farms typical of the developing world are necessary [179]. Agroecology, the study of the ecology of agricultural systems, has brought novel land management approaches to subsistence farmers. Several agronomic techniques have been developed which enhance the tolerance or adaptation of crops to diverse types of abiotic stresses. Four of these techniques are briefly discussed below.

5.3.1. Conservation Tillage. Conservation tillage, also known as minimum tillage or no-till, is widely used in some parts of the world mainly to control erosion and increase soil fertility. Conservation tillage, leaving the previous year's crop residue on the fields, has the effect of reducing soil erosion and runoff. A study in drought-prone areas of northern Ethiopia showed that conservation tillage enhanced the availability of soil moisture to sorghum and chickpea [50]. Similarly, in Zimbabwe's semiarid areas receiving low amounts of rainfall, conservation tillage increased the productivity of maize compared to the conventional tillage [180].

5.3.2. Proper Drainage of Waterlogged Soils. As indicated above, waterlogging is among the major constraints to crop production in some parts of Africa. The broad-bed maker (BBM), a low-cost modification of the traditional Ethiopian plow, has been efficiently utilized to drain excess moisture from fields containing *Vertisols* through the creation of broad-beds and furrows (BBF). Due to the improved drainage obtained using BBM, a grain yield increase of 78% and straw yield increase of 56% were obtained for wheat in the central highlands of Ethiopia [53].

5.3.3. Presowing Seed Treatments to Boost Tolerance to Salts. Since seed germination and the early establishment of seedlings are the stages of plant development most sensitive to salt stress, rapid and uniform germination is required [129]. Enhanced growth can be obtained by immersing seeds in salt solution before sowing. This process is commonly known as "priming." Five types of priming are known: (i) osmopriming in which seeds are treated with solutions of sugars, glycerol, or mannitol that promote germination; (ii) halopriming involving soaking of seeds in solutions of inorganic salts such as MnSO₄; (iii) hydropriming in which seeds are soaked in water before sowing; (iv) thermopriming involving treating seeds with high or low temperatures before sowing; and (v) hormone priming, which involves the use of plant growth regulators such as auxin, gibberellic acid, and ethylene.

5.3.4. The System of Crop Intensification (SCI). SCI was formed in order to transfer the knowledge and experience developed by System Rice Intensification (SRI) to diverse crops. SRI has been tested in 50 countries and coordinated by the International Programs of Cornell's College of Agriculture and Life Sciences. SCI involves improving planting and growing techniques such that more yield can be obtained using fewer seeds and less water through management of the relationship between the plant and soil. Plant spacing and age of rice at transplantation time are two examples of this kind of management. An application of this technique to abiotic stresses is the reduction of toxic levels of inorganic arsenic in rice fields by using the optimum amount of water instead of excess water [181, 182]. In addition to major crops such as rice and wheat, SCI gave positive results for indigenous crops such as finger millet, tef, and mustard [54].

5.4. Plant Growth Regulators or Hormones. Several plant growth regulators (PGRs) play a key role in the plant response to diverse environmental conditions especially to abiotic stresses. Major PGRs including abscisic acid (ABA), cytokinin, ethylene, jasmonate (JA), auxin, brassinosteroids (BRs), and gibberellins (GAs) are known to be involved in the protection of plants against salt stress [55]. Exogenous application of Methyl jasmonate (MeJA) protects plants from both biotic and abiotic stresses [183]. Jasmonates are also involved in the protection of plants from both heat and cold [56]. Protection from the cold is through the regulation of C-repeat binding factor (CBF) by jasmonate during cold stress [56]. A recent study on orphan cereals showed that deficiency in gibberellic acid did not only reduce the height of tef and finger millet but also enhanced the tolerance of these crops to drought [184]. A study on a model plant Arabidopsis showed that strigolactones, a small class of carotenoid-derived compounds, play a positive regulatory role in the plant response to drought and salt stress [185].

6. Initiatives to Mitigate Abiotic Stresses in Africa

A number of projects have been initiated to improve productivity of crops through tackling major constraints. Among these, three initiatives relevant to Africa are briefly discussed below.

6.1. Stress-Tolerant Rice for Africa and South Asia (STRASA). STRASA was formed in 2007 by IRRI (International Rice Research Institute) and Africa Rice to develop and deliver rice varieties tolerant to diverse abiotic stresses in the unfavorable rice-growing environments [57] (Table 5). The initiative focused on developing rice varieties tolerant to drought, flooding (submergence), soil salinity, iron toxicity, and cold. Improved rice varieties tolerant to drought, flooding, and salinity have been extensively investigated in South Asian countries such as Bangladesh, India, and Nepal while those tolerant to iron toxicity have been tested in several African countries where a single variety has been released in Sierra Leone. The cold-tolerant lines have been evaluated in coldprone African countries where a total of seven improved varieties have been released in Burundi, Madagascar, and Mali.

6.2. Drought-Tolerant Maize for Africa (DTMA). DTMA was formed by CIMMYT (International Maize and Wheat Improvement Center) and IITA (International Institute of Tropical Agriculture) in close collaboration with private and public sectors where they developed over 200 maize varieties for 13 African countries, namely, Angola, Benin, Ethiopia, Ghana, Kenya, Malawi, Mali, Mozambique, Nigeria, Tanzania, Uganda, Zambia, and Zimbabwe (Table 5). In these regions where maize significantly suffers from moisture scarcity, about 54,000 metric tons of certified drought-tolerant seed was produced through the program in a single year [58, 186]. Countries which benefited a lot from the program by releasing maximum number of varieties are Nigeria (21), Zambia (18), and Malawi (17).

6.3. Feed the Future Innovation Lab. Feed the Future, the US Government's global hunger and food security initiative, addresses the global agricultural challenges through its 24 Feed the Future Innovation Labs [59] (Table 5). Five programs related to developing climate-resilient crops and focus countries are on beans (Haiti, Honduras, Malawi, Mozambique, Tanzania and Zambia), on chickpea (Ethiopia, India and Turkey), on cowpea (Burkina Faso, Ghana, Nigeria and Senegal), on millet (India, Mali and Nigeria) and on sorghum (Ethiopia, India, Mali and South Africa). From the above 15 countries in the resilient program, about 75% are from Africa from which three countries (Ethiopia, Mali and Nigeria) are represented each in two programs. In general, this shows a high level of commitment by scientists and government of USA to contribute to food security in Africa through tackling major climatic and soil constraints.

7. Conclusions and Prospects

A variety of abiotic stresses cause considerable crop yield loss annually. Compared to major or exotic crops, orphan crops are more tolerant to locally prevailing abiotic stresses. However, even these resilient orphan crops do not produce optimum yields when subjected to abiotic stresses. Although indigenous crops are better protected than major crops of the world against certain abiotic stresses, the effects on orphan crops are substantial, especially under extreme moisture scarcity. African researchers are faced with the challenge of improving the productivity of orphan crops through tackling major yield limiting factors, which include a variety of abiotic stresses. Moreover, researchers and policy makers need to prioritize research directions based on the urgency and extent of the prevailing abiotic stresses. Useful lessons could also be learned from the work made in other parts of the world. For instance, in Japan, a salt-tolerant rice variety called "kaijin" was developed in a very short period through mutation breeding and TILLING in response to the large areas of land devoted to rice cultivation that were affected by the

Name	Crons	Ahintic stress targeted	African country of focus	Achievement/improved	Reference
	o dovo			varieties	
		Drought	Burundi, Guinea Conakry, Madagascar, Mali, Nigeria & Sierra Leone	20 varieties	
	;U	Submergence	Mali	2 varieties	
1 KADA	KICE	Salinity	Gambia & Sierra Leone	6 varieties	[57]
		Cold	Burundi, Madagascar & Mali	7 varieties	
		Iron toxicity	Sierra Leone	1 variety	
			Angola, Benin, Ethiopia, Ghana, Kenya,	Improved	
DTMA	Maize	Drought	Malawi, Mali, Mozambique, Nigeria,	drought-tolerant maize	[58]
			Tanzania, Uganda, Zambia & Zimbabwe	varieties	
	Beans	Drought & heat	Malawi, Mozambique, Tanzania & Zambia		
	Chickpea		Ethiopia		
Feed the Future Innovation Labs	Cowpea	Drought	Burkina Faso, Ghana, Nigeria & Senegal		[20]
	Millet	Drought & heat	Mali & Nigeria		
	Sorghum	Drought & heat	Ethiopia, Kenya, Mali & South Africa		

sia; DTMA: Drought-Tolerant Maize for Africa;	
STRASA: Stress-Tolerant Rice for Africa and South A	
tackling diverse types of abiotic stresses in Africa. (Management in Development.
BLE 5: Selected projects focused on t	ARMD: Forum for Agricultural Risk l

high level of salts from the sea during the 2011 Tsunami [2]. Global warming and climate change call for the development of climate-smart crops. Definition of the optimum ideotype for each crop type is necessary so that breeders focus on developing crop varieties with a desirable architecture and high yield potential. Both conventional and modern breeding techniques as well as omics tools play key roles in tackling the problems of abiotic stresses. Once candidate lines with desirable abiotic stress tolerance have been obtained, increased tolerance to other abiotic stresses or enhanced productivity can then be obtained through hybridization. Some key achievements from the global initiatives are contributing to increased productivity of crops cultivated in Africa. However, further commitments are required to include major staple crops in the continent which are largely neglected from global scientific community.

Conflicts of Interest

The author declares that they have no conflicts of interest.

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References

- FAO, The impact of natural hazards and disasters on agriculture and food security and nutrition a call for action to build resilient livelihoods, FAO (Food and Agriculture Organization), Rome, Italy, 2015.
- [2] H. Takagi, M. Tamiru, A. Abe et al., "MutMap accelerates breeding of a salt-tolerant rice cultivar," *Nature Biotechnology*, vol. 33, no. 5, pp. 445–449, 2015.
- [3] S. Rosell and B. Holmer, "Rainfall change and its implications for Belg harvest in South Wollo, Ethiopia," *Geografiska Annaler: Series A, Physical Geography*, vol. 89, no. 4, pp. 287–299, 2007.
- [4] A. Lemi, "Rainfall probability and agricultural yield in Ethiopia," *Eastern Africa Social Science Research Review*, vol. 21, no. 1, pp. 57–96, 2005.
- [5] WFP, "El Niño: implications and scenarious for 2015. VAM: Food Security Analysis," in *World food Program*, 17, 2015.
- [6] O. Rojas, Y. Li, and R. Cumani, Understanding the drought impact of El Niño on the global agricultural areas: an assessment using FAO's Agricultural Stress Index (ASI), FAO, Rome, Italy, 2015.
- [7] M. V. K. Sivakumar, "Empirical analysis of dry spells for agricultural applications in West Africa," *Journal of Climate*, vol. 5, no. 5, pp. 532–539, 1992.
- [8] "FAO (food and agricultural organization) statistical data," http://faostat3.fao.org/home/E.
- [9] CSA, "Agricultural sample survey for 2013/14," in *Statistical Bulletin 532*, Addis Ababa, Ethiopia, 2014.
- [10] C. G. Campell, Grass pea (Lathyrus sativus L.), IPK, Gartersleben/IPGRI, Rome, Italy, 1997.
- [11] S. A. Brandt, *The 'tree against hunger': enset-based agricultural system in Ethiopia*, American Association for the Advancement of Science, Washington, DC, USA, 1997.

- [12] NAP, Lost Crops of Africa, Volume I: Grains, National Academies Press, Washington, DC, USA, 1996.
- [13] NAP, Ed., *Lost Crops of Africa; Volume II: Vegetables*, National Academies Press, 2006.
- [14] H. Ceballos, C. A. Iglesias, J. C. Pérez, and A. G. O. Dixon, "Cassava breeding: opportunities and challenges," *Plant Molecular Biology*, vol. 56, no. 4, pp. 503–516, 2004.
- [15] J. T. Williams and N. Haq, GlobaL Research on Underutilised Crops: An Assessment of Current Activities and Proposals for Enhanced Cooperation, vol. 50, Southampton, UK.
- [16] A. Getinet, G. Rakow, and R. K. Downey, "Agronomic performance and seed quality of Ethiopian mustard in Saskatchewan," *Canadian Journal of Plant Science*, vol. 76, no. 3, pp. 387–392, 1996.
- [17] S. Ketema, *Tef, Eragrostis tef (Zucc.) Trotter*, vol. 52, Institute of Plant Genetics and Crop Plant Research, Gatersleben/International Plant Genetic Resources Institute, Rome, Italy, 1997.
- [18] "Policy brief: use of saline water in agriculture to improve livelihoods in West Asia and North Africa," http://www.iwmi .cgiar.org/assessment/files_new/research_projects/Policy%20 Brief_ICBA.pdf.
- [19] N. Sanginga, O. Lyasse, and B. B. Singh, "Phosphorus use efficiency and nitrogen balance of cowpea breeding lines in a low P soil of the derived savanna zone in West Africa," *Plant and Soil*, vol. 220, no. 1-2, pp. 119–128, 2000.
- [20] H. Valenzuela and J. Smith, "Cowpea," in Sustainable Agriculture, Green Manure Crops, vol. 3, 2002.
- [21] IPGRI, "Promoting fonio production in West and Central Africa through germplasm management and improvement of post harvest technology," in *Project Number: 200078600-00100*, 18, p. 00100, International Plant Genetic Resources Institute, Benin, Africa, 2004.
- [22] R. Fungo, "Potential of bananas in alleviating micronutrient eficiencies in the great lakes region of East Africa," in *African Crop Science Conference Proceedings*, 9:8, 2009.
- [23] J. S. Heslop-Harrison and T. Schwarzacher, "Domestication, genomics and the future for banana," *Annals of Botany*, vol. 100, no. 5, pp. 1073–1084, 2007.
- [24] A. Getinet and S. M. Sharma, *Guizotia abyssinica (L. f.) Cass*, Institute of Plant Genetics and Crop Plant Research, Gatersleben/International Plant Genetic Resources Institute, Rome, Italy, 1996.
- [25] L. Spaenij-Dekking, Y. Kooy-Winkelaar, and F. Koning, "The Ethiopian cereal Tef in celiac disease," *The New England Journal* of *Medicine*, vol. 353, no. 16, pp. 1748-1749, 2005.
- [26] A. N. Doust, E. A. Kellogg, K. M. Devos, and J. L. Bennetzen, "Foxtail millet: a sequence-driven grass model system," *Plant Physiology*, vol. 149, no. 1, pp. 137–141, 2009.
- [27] A. Chandrashekar, "Finger millet: Eleusine coracana.," *Advances in Food and Nutrition Research*, vol. 59, pp. 215–262, 2010.
- [28] A. Chandrasekara and F. Shahidi, "Antiproliferative potential and DNA scission inhibitory activity of phenolics from whole millet grains," *Journal of Functional Foods*, vol. 3, no. 3, pp. 159– 170, 2011.
- [29] V. Vadez, J. Kholová, R. S. Yadav, and C. T. Hash, "Small temporal differences in water uptake among varieties of pearl millet (Pennisetum glaucum (L.) R. Br.) are critical for grain yield under terminal drought," *Plant and Soil*, vol. 371, no. 1-2, pp. 447–462, 2013.

- [30] F. R. Bidinger, V. Mahalakshmi, and G. D. P. Rao, "Assessment of drought resistance in pearl millet [*Pennisetum americanum* (*L.*) Leeke]. I. Factors affecting yields under stress," *Australian Journal of Agricultural Research*, vol. 38, no. 1, pp. 37–48, 1987.
- [31] I. P. Ajithkumar and R. Panneerselvam, "ROS scavenging system, osmotic maintenance, pigment and growth status of panicum sumatrense roth. under drought stress," *Cell Biochemistry and Biophysics*, vol. 68, no. 3, pp. 587–595, 2014.
- [32] R. A. Balsamo, C. Vander Willigen, A. M. Bauer, and J. Farrant, "Drought tolerance of selected Eragrostis species correlates with leaf tensile properties," *Annals of Botany*, vol. 97, no. 6, pp. 985– 991, 2006.
- [33] M. Kawase and R. E. Whitmoyer, "Aerenchyma development in waterlogged plants," *American Journal of Botany*, vol. 67, no. 1, pp. 18–22, 1980.
- [34] A. Promkhambut, A. Younger, A. Polthanee, and C. Akkasaeng, "Morphological and physiological responses of sorghum (Sorghum bicolor L. Moench) to waterlogging," *Asian Journal* of *Plant Sciences*, vol. 9, no. 4, pp. 183–193, 2010.
- [35] S. S. Kulkarn and P. D. Chavan, "Study of effect of waterlogging on root anatomy of ragi and rice," *American Journal of Plant Physiology*, vol. 9, no. 2, pp. 46–51, 2014.
- [36] D. Kumutha, K. Ezhilmathi, R. K. Sairam, G. C. Srivastava, P. S. Deshmukh, and R. C. Meena, "Waterlogging induced oxidative stress and antioxidant activity in pigeonpea genotypes," *Biologia Plantarum*, vol. 53, no. 1, pp. 75–84, 2009.
- [37] S. S. Kulkarni and P. D. Chavan, "Study of some aspects of anaerobic metabolism in roots of finger millet and rice plants subjected to waterlogging stress," *International Journal of Botany*, vol. 9, no. 2, pp. 80–85, 2013.
- [38] P. Kumar, M. Pal, R. Joshi, and R. K. Sairam, "Yield, growth and physiological responses of mung bean [Vigna radiata (L.) Wilczek] genotypes to waterlogging at vegetative stage," *Physiology and Molecular Biology of Plants*, vol. 19, no. 2, pp. 209–220, 2013.
- [39] M. Thudi, H. D. Upadhyaya, A. Rathore et al., "Genetic dissection of drought and heat tolerance in chickpea through genome-wide and candidate gene-based association mapping approaches," *PLoS ONE*, vol. 9, no. 5, Article ID e96758, 2014.
- [40] M. Ashraf and M. Hafeez, "Thermotolerance of pearl millet and maize at early growth stages: Growth and nutrient relations," *Biologia Plantarum*, vol. 48, no. 1, pp. 81–86, 2004.
- [41] J. D. Ehlers and A. E. Hall, "Heat tolerance of contrasting cowpea lines in short and long days," *Field Crops Research*, vol. 55, no. 1-2, pp. 11–21, 1998.
- [42] E. J. Landry, J. E. Lafferty, C. J. Coyne, W. L. Pan, and J. Hu, "Registration of four winter-hardy faba bean germplasm lines for use in winter pulse and cover crop development," *Journal of Plant Registrations*, vol. 9, no. 3, pp. 367–370, 2015.
- [43] S. P. Singh, "Sources of cold tolerance in grain-sorghum," *Canadian Journal of Plant Science*, vol. 65, no. 2, pp. 251–257, 1985.
- [44] E. Abate, S. Hussein, M. Laing, and F. Mengistu, "Quantitative responses of tef [Eragrostis tef (Zucc.) Trotter] and weeping love grass [Eragrostis curvula (Schrad.) Nees] varieties to acid soil," *Australian Journal of Crop Science*, vol. 7, no. 12, pp. 1854–1860, 2013.
- [45] O. F. Linares, "African rice (Oryza glaberrima): history and future potential," *Proceedings of the National Acadamy of Sciences of the United States of America*, vol. 99, no. 25, pp. 16360– 16365, 2002.

- [46] Z. Tadele, "Tef Improvement Project: harnessing genetic and genomic tools to boost productivity," in Achievements and Prospects of Tef Improvement, K. Assefa, S. Chanyalew, and Z. Tadele, Eds., pp. 333–342, EIAR-University of Bern, Bern, Switzerland, 2013.
- [47] R. S. Yadav, C. T. Hash, F. R. Bidinger, G. P. Cavan, and C. J. Howarth, "Quantitative trait loci associated with traits determining grain and stover yield in pearl millet under terminal drought-stress conditions," *Theoretical and Applied Genetics*, vol. 104, no. 1, pp. 67–83, 2002.
- [48] K. Esfeld, C. Uauy, and Z. Tadele, "Application of TILLING for orphan crop improvement," in *Biotechnology of Neglected and Underutilized Crops*, S. M. Jain and S. D. Gupta, Eds., pp. 83– 113, Springer, 2013.
- [49] C. Bharadwaj, T. Kumar, G. Rajguru, and A. K. Verma, "Standardization of mutagens for developing TILLING populations in chickpea," *Journal of Science & Engineering Research*, vol. 16, no. 1, pp. 23–27, 2011.
- [50] O. V. McHugh, T. S. Steenhuis, Berihun Abebe, and E. C. M. Fernandes, "Performance of in situ rainwater conservation tillage techniques on dry spell mitigation and erosion control in the drought-prone North Wello zone of the Ethiopian highlands," *Soil & Tillage Research*, vol. 97, no. 1, pp. 19–36, 2007.
- [51] A. Abewa, B. Yitaferu, Y. Selassie, and T. Amare, "The role of biochar on acid soil reclamation and yield of teff (Eragrostis tef [Zucc] Trotter) in Northwestern Ethiopia," *Journal of Agricultural Science*, vol. 6, no. 1, pp. 1–12, 2014.
- [52] A. Mukherjee and R. Lal, "Biochar impacts on soil physical properties and greenhouse gas emissions," *Agronomy Journal*, vol. 3, no. 2, pp. 313–339, 2013.
- [53] S. Jutzi, "Deep black clay soils (Vertisols): management options for the Ethiopian highlands," *Mountain Research & Development*, vol. 8, no. 2-3, pp. 153–156, 1988.
- [54] B. Abraham, H. Araya, T. Berhe et al., "The system of crop intensification: reports from the field on improving agricultural production, food security, and resilience to climate change for multiple crops," *Agriculture and Food Security*, vol. 3, article no. 4, 2014.
- [55] H. Ryu and Y.-G. Cho, "Plant hormones in salt stress tolerance," *Journal of Plant Biology*, vol. 58, no. 3, pp. 147–155, 2015.
- [56] M. Sharma and A. Laxmi, "Jasmonates: emerging players in controlling temperature stress tolerance," *Frontiers in Plant Science*, vol. 6, article no. 1129, 2016.
- [57] "Stress-tolerant rice for Africa and Asia," http://strasa.irri .org/home.
- [58] "DTMA (Drought Tolerant Maize)," http://dtma.cimmyt.org/.
- [59] "Advancing food security and nutrition together," https://feedthefuture.gov/.
- [60] R. Hall, Neglected and Underutilized Species (NUS): for a Food-Secure Africa, A. P. Rudebjer and S. Padulosi, Eds., Bioversity International, Rome, Italy, 2013.
- [61] I. Dawson and H. Jaenicke, "Underutilised plant species: the role of biotechnology," in *Position Paper No 1*, vol. 27, 2006.
- [62] "Crops for the future," http://www.cffresearch.org.
- [63] D. D. Kuta, E. Kwon-Ndung, S. Dachi, M. Ukwungwu, and E. D. Imolehin, "Potential role of biotechnology tools for genetic improvement of 'lost crops of Africa': the case of fonio (Digitaria exilis and Digitaria iburua)," *African Journal of Biotechnology*, vol. 2, no. 12, pp. 580–585, 2003.
- [64] R. L. Naylor, W. P. Falcon, R. M. Goodman et al., "Biotechnology in the developing world: a case for increased investments in orphan crops," *Food Policy*, vol. 29, no. 1, pp. 15–44, 2004.

- [65] Z. Tadele and K. Assefa, "Increasing food production in Africa by boosting the productivity of understudied crops," *Agronomy*, vol. 2, no. 4, pp. 240–283, 2012.
- [66] Z. Tadele, "Role of crop research and development in food security of Africa," *International Journal of Plant Biology and Research*, vol. 2, no. 3, article 1019, 2014.
- [67] S. Padulosi, M. Bhag, R. S. Bala et al., "Food security and climate change: role of plant genetic resources of minor millets," *Indian Journal of Plant Genetic Resource*, vol. 22, pp. 1–16, 2009.
- [68] "The svalbard global seed vault," https://www.croptrust.org/ what-we-do/svalbard-global-seed-vault/.
- [69] SPGRC, SADC Plant Genetic Resources Centre: Twenty Fifth Annual Report 2014/2015, 36, SPGRC, Lusaka, Zambia, 2015.
- [70] S. Chanyalew, K. Assefa, and G. Metaferia, "Phenotypic and molecular diversity in tef," in *Achievements and Prospects of Tef Improvement*, K. Assefa, S. Chanyalew, and Z. Tadele, Eds., pp. 21–31, EIAR-University of Bern, Bern, Switzerland, 2013.
- [71] C. Sautter, S. Poletti, P. Zhang, and W. Gruissem, "Biofortification of essential nutritional compounds and trace elements in rice and cassava," *Proceedings of the Nutrition Society*, vol. 65, no. 2, pp. 153–159, 2006.
- [72] Z. Tadele, "Drought Adaptation in Millets," in *Abiotic and Biotic Stress in Plants: Recent Advances and Future Perspectives*, A. Shanker and C. Shanker, Eds., pp. 639–662, InTech, 2016.
- [73] B. I. G. Haussmann, H. F. Rattunde, E. Weltzien-Rattunde, P. S. C. Traoré, K. vom Brocke, and H. K. Parzies, "Breeding strategies for adaptation of pearl millet and sorghum to climate variability and change in West Africa," *Journal of Agronomy and Crop Science*, vol. 198, no. 5, pp. 327–339, 2012.
- [74] K. Assefa, G. Cannarozzi, D. Girma et al., "Genetic diversity in tef [Eragrostis tef (Zucc.) Trotter]," *Frontiers in Plant Science*, vol. 6, no. 177, 2015.
- [75] G. Alemaw and A. T. Wold, "An agronomic and seed-quality evaluation of noug (Guizotia abyssinica Cass.) germplasm in Ethiopia," *Plant Breeding*, vol. 114, no. 4, pp. 375-376, 1995.
- [76] Asiwe J. A. N., Ed., Field Evaluation of Bambara Groundnut, University of Bern, Stampfl, Bern, Switzerland, 2009.
- [77] I. Léder, Ed., Sorghum and Millets, UNESCO, Eolss, Oxford, UK, 2004.
- [78] B. O. De Lumen, S. Thompson, and W. J. Odegard, "Sulfur amino acid-rich proteins in acha (Digitaria exilis), a promising underutilized African cereal," *Journal of Agricultural and Food Chemistry*, vol. 41, no. 7, pp. 1045–1047, 1993.
- [79] B. Písaříková, S. Kráčmar, and I. Herzig, "Amino acid contents and biological value of protein in various amaranth species," *Czech Journal of Animal Science*, vol. 50, no. 4, pp. 169–174, 2005.
- [80] A. Yami, "Tef straw: a valuable feed resource to improve animal production and productivity," in *Achievements and Prospects of Tef Improvement*, K. Assefa, S. Chanyalew, and Z. Tadele, Eds., pp. 233–251, EIAR-University Bern, Bern, Switzerland, 2013.
- [81] E. Hopman, L. Dekking, M.-L. Blokland et al., "Tef in the diet of celiac patients in The Netherlands," *Scandinavian Journal of Gastroenterology*, vol. 43, no. 3, pp. 277–282, 2008.
- [82] P. Pontieri, G. Mamone, S. De Caro et al., "Sorghum, a healthy and gluten-free food for celiac patients as demonstrated by genome, biochemical, and immunochemical analyses," *Journal* of Agricultural and Food Chemistry, vol. 61, no. 10, pp. 2565– 2571, 2013.
- [83] P. Caron, P. Craufurd, A. Martin et al., "Impacts of AKST on development and sustainability goals: global report," in *Agriculture at a Crossroads*, B. D. McIntyre, H. R. Herren, J.

Wakhungu, and R. T. Watson, Eds., pp. 145–253, : International Assessment of Agricultural Knowledge, Science and Technology for Development, IAASTD, Washington, DC, USA, 2009.

- [84] A. Kebede, J. C. McCann, A. E. Kiszewski, and Y. Ye-Ebiyo, "New evidence of the effects of agro-ecologic change on malaria transmission," *The American Journal of Tropical Medicine and Hygiene*, vol. 73, no. 4, pp. 676–680, 2005.
- [85] Y. Ye-Ebiyo, R. J. Pollack, and A. Spielman, "Enhanced development in nature of larval Anopheles arabiensis mosquitoes feeding on maize pollen," *The American Journal of Tropical Medicine and Hygiene*, vol. 63, no. 1-2, pp. 90–93, 2000.
- [86] J. McCann, Maize and Grace: Africa's Encounter with a Neew World Crop, 1500-2000, Harvard University Press, Cambridge, Mass, USA, 2005.
- [87] "Abiotic stress," http://levitycropscience.com/poster/.
- [88] FAO, *Agriculture, Food and Water*, Food and Agriculture Organization, Rome, Italy, 2003.
- [89] FAO, AQUASTAT Main Database, Food and Agriculture Organization of the United Nations (FAO), 2016.
- [90] J. Rockström and M. Falkenmark, "Agriculture: increase water harvesting in Africa," *Nature*, vol. 519, no. 7543, pp. 283–285, 2015.
- [91] A. Matsuura, W. Tsuji, P. An, S. Inanaga, and K. Murata, "Effect of pre- and post-heading water deficit on growth and grain yield of four millets," *Plant Production Science*, vol. 15, no. 4, pp. 323– 331, 2012.
- [92] M. Maqsood and S. N. A. Ali, "Effects of drought on growth, develompent, radiation use efficiency and yield of finger millet (Eleucine coracana)," *Pakistan Journal of Botany*, vol. 39, no. 1, pp. 123–134, 2007.
- [93] T. Winkel, J.-F. Renno, and W. A. Payne, "Effect of the timing of water deficit on growth, phenology and yield of pearl millet (Pennisetum glaucum (L.) R. Br.) grown in Sahelian conditions," *Journal of Experimental Botany*, vol. 48, no. 310, pp. 1001–1009, 1997.
- [94] M. T. Abraha, S. Hussein, M. Laing, and K. Assefa, "Genetic management of drought in tef: current status and future research directions," *Global Journal of Crop, Soil Science and Plant Breeding*, vol. 3, no. 3, pp. 156–161, 2015.
- [95] V. Mahalakshmi, F. R. Bidinger, and D. S. Raju, "Effect of timing of water deficit on pearl millet (Pennisetum americanum)," *Field Crops Research*, vol. 15, no. 3-4, pp. 327–339, 1987.
- [96] V. O. Sadras, P. Grassini, and P. Steduto, Status of Water Use Efficiency of Main Crops: SOLAW Background Thematic Report - TR07, FAO, Rome, Italy.
- [97] J. M. Slingo, A. J. Challinor, B. J. Hoskins, and T. R. Wheeler, "Introduction: food crops in a changing climate," *Philosophical Transactions of the Royal Society B: Biological Sciences*, vol. 360, no. 1463, pp. 1983–1989, 2005.
- [98] J. R. Porter and M. A. Semenov, "Crop responses to climatic variation," *Philosophical Transactions of the Royal Society B: Biological Sciences*, vol. 360, no. 1463, pp. 2021–2035, 2005.
- [99] C. Parent, N. Capelli, A. Berger, M. Crèvecoeur, and J. F. Dat, "An overview of plant responses to soil waterlogging," *Plant Stress*, vol. 2, pp. 20–27, 2008.
- [100] S. M. Virmani, "Agroclimatology of the Vertisols and vertic soil areas of Africa," in *Management of Vertisols in Sub-Saharan Africa*, S. C. Jutzi, I. Haque, J. McIntire, and J. E. S. Stares, Eds., ILCA (International Livestock Center for Africa), Addis Ababa, Ethiopia, 1988.

- [101] A. Matsuura, P. An, K. Murata, and S. Inanaga, "Effect of preand post-heading waterlogging on growth and grain yield of four millets," *Plant Production Science*, vol. 19, no. 3, pp. 348– 359, 2016.
- [102] S. Silva, "Aluminium toxicity targets in plants," *Journal of Botany*, vol. 2012, Article ID 219462, 8 pages, 2012.
- [103] "Effects of soil acidity," https://www.agric.wa.gov.au/soilacidity/effects-soil-acidity.
- [104] E. Abate, S. Hussien, M. Laing, and F. Mengistu, "Aluminium toxicity tolerance in cereals: Mechanisms, genetic control and breeding methods," *African Journal of Agricultural Research*, vol. 8, no. 9, pp. 711–722, 2013.
- [105] "Salt-affected soils," http://www.fao.org/soils-portal/soil-management/management-of-some-problem-soils/salt-affectedsoils/more-information-on-salt-affected-soils/en/.
- [106] L. Postiglione, "Soil salinization in the Mediterranean: soils, processes and implications," in *Mediterranean Desertification: A Mosaic of Processes and Responses*, N. A. Geeson, C. J. Btandt, and B. Thornes, Eds., pp. 163–173, John Wiley & Sons, 2002.
- [107] E. G. Barrett-Lennard, "The interaction between waterlogging and salinity in higher plants: Causes, consequences and implications," *Plant and Soil*, vol. 253, no. 1, pp. 35–54, 2003.
- [108] K. G. Asfaw and F. I. Danno, "ffects of salinity on yield and yield components of tef [Eragrostis tef (Zucc.) Trotter] accessions and varieties," *Current Research Journal of Biological Sciences*, vol. 3, no. 4, pp. 289–299, 2011.
- [109] C. E. Bita and T. Gerats, "Plant tolerance to high temperature in a changing environment: scientific fundamentals and production of heat stress-tolerant crops," *Frontiers in Plant Science*, vol. 4, no. 273, 2013.
- [110] W. Schlenker and M. Roberts, "Estimating the impact of climate change on crop yields: the importance of nonlinear temperature effects," NBER Working Paper No 13799, JEL No C23,Q54, 2008.
- [111] G. C. Nelson, M. W. Rosegrant, J. Koo et al., "Climate change impact on agriculture and costs of adaptation," Food Policy Report, International Food Policy Research Institute, Washington, DC, USA, 2009.
- [112] K. Miura and T. Furumoto, "Cold signaling and cold response in plants," *International Journal of Molecular Sciences*, vol. 14, no. 3, pp. 5312–5337, 2013.
- [113] C. L. Walthall, J. Hatfield, P. Backlund et al. et al., Climate Change and Agriculture in the United States: Effects and Adaptation, United States Department of Agricultulure (USDA), Washington, DC, USA, 2012.
- [114] M. V. Mickelbart, P. M. Hasegawa, and J. Bailey-Serres, "Genetic mechanisms of abiotic stress tolerance that translate to crop yield stability," *Nature Reviews Genetics*, vol. 16, no. 4, pp. 237– 251, 2015.
- [115] A. S. Duque, A. M. de Almeida, A. B. da Silva et al., "Abiotic stress responses in plants: unraveling the complexity of genes and networks to survive, abiotic stress," in *Plant Responses and Applications in Agriculture*, K. Vahdati, Ed., pp. 49–101, Intech, 2013.
- [116] P. Chivenge, T. Mabhaudhi, A. T. Modi, and P. Mafongoya, "The potential role of neglected and underutilised crop species as future crops under water scarce conditions in Sub-Saharan Africa," *International Journal of Environmental Research and Public Health*, vol. 12, no. 6, pp. 5685–5711, 2015.
- [117] M. M. Chaves and M. M. Oliveira, "Mechanisms underlying plant resilience to water deficits: prospects for water-saving agriculture," *Journal of Experimental Botany*, vol. 55, no. 407, pp. 2365–2384, 2004.

- [118] V. Vadez, "Root hydraulics: the forgotten side of roots in drought adaptation," *Field Crops Research*, vol. 165, pp. 15–24, 2014.
- [119] C. G. Lopez, G. M. Banowetz, C. J. Peterson, and W. E. Kronstad, "Dehydrin expression and drought tolerance in seven wheat cultivars," *Crop Science*, vol. 43, no. 2, pp. 577–582, 2003.
- [120] M. A. Hossain and S. N. Uddin, "Mechanisms of waterlogging tolerance in wheat: morphological and metabolic adaptations under hypoxia or anoxia," *Australian Journal of Crop Science*, vol. 5, no. 9, pp. 1094–1101, 2011.
- [121] R. K. Sairam, D. Kumutha, K. Ezhilmathi, P. S. Deshmukh, and G. C. Srivastava, "Physiology and biochemistry of waterlogging tolerance in plants," *Biologia Plantarum*, vol. 52, no. 3, pp. 401– 412, 2008.
- [122] L. V. Kochian, M. A. Piñeros, J. Liu, and J. V. Magalhaes, "Plant adaptation to acid soils: the molecular basis for crop aluminum resistance," *Annual Review of Plant Biology*, vol. 66, pp. 571–598, 2015.
- [123] T. Sasaki, Y. Yamamoto, B. Ezaki et al., "A wheat gene encoding an aluminum-activated malate transporter," *The Plant Journal*, vol. 37, no. 5, pp. 645–653, 2004.
- [124] J. V. Magalhaes, J. Liu, C. T. Guimarães et al., "A gene in the multidrug and toxic compound extrusion (MATE) family confers aluminum tolerance in sorghum," *Nature Genetics*, vol. 39, no. 9, pp. 1156–1161, 2007.
- [125] J. Furukawa, N. Yamaji, H. Wang et al., "An aluminum-activated citrate transporter in barley," *Plant & Cell Physiology (PCP)*, vol. 48, no. 8, pp. 1081–1091, 2007.
- [126] E. Delhaize, P. R. Ryan, D. M. Hebb, Y. Yamamoto, T. Sasaki, and H. Matsumoto, "Engineering high-level aluminum tolerance in barley with the ALMT1 gene," *Proceedings of the National Acadamy of Sciences of the United States of America*, vol. 101, no. 42, pp. 15249–15254, 2004.
- [127] J. F. Pereira, G. Zhou, E. Delhaize, T. Richardson, M. Zhou, and P. R. Ryan, "Engineering greater aluminium resistance in wheat by over-expressing TaALMT1," *Annals of Botany*, vol. 106, no. 1, pp. 205–214, 2010.
- [128] R. Munns and M. Tester, "Mechanisms of salinity tolerance," Annual Review of Plant Biology, vol. 59, pp. 651–681, 2008.
- [129] M. Ashraf, H. R. Athar, P. J. C. Harris, and T. R. Kwon, "Some prospective strategies for improving crop salt tolerance," *Advances in Agronomy*, vol. 97, pp. 45–110, 2008.
- [130] D. Mittal, N. Sharma, V. Sharma, S. K. Sopory, and N. Sanan-Mishra, "Role of microRNAs in rice plant under salt stress," *Annals of Applied Biology*, vol. 168, no. 1, pp. 2–18, 2016.
- [131] B. Asthir, "Mechanisms of heat tolerance in crop plants," *Biologia Plantarum*, vol. 59, no. 4, pp. 620–628, 2015.
- [132] M. Hasanuzzaman, K. Nahar, M. M. Alam, R. Roychowdhury, and M. Fujita, "Physiological, biochemical, and molecular mechanisms of heat stress tolerance in plants," *International Journal of Molecular Sciences*, vol. 14, no. 5, pp. 9643–9684, 2013.
- [133] B. Asthir, "Protective mechanisms of heat tolerance in crop plants," *Journal of Plant Interactions*, vol. 10, no. 1, pp. 202–210, 2015.
- [134] N. K. Gupta, S. Agarwal, V. P. Agarwal, N. S. Nathawat, S. Gupta, and G. Singh, "Effect of short-term heat stress on growth, physiology and antioxidative defence system in wheat seedlings," *Acta Physiologiae Plantarum*, vol. 35, no. 6, pp. 1837–1842, 2013.
- [135] M. F. Thomashow, "Plant cold acclimation: freezing tolerance genes and regulatory mechanisms," *Annual Review of Plant Biology*, vol. 50, pp. 571–599, 1999.

- [136] L. A. Bravo, T. J. Close, L. J. Corcuera, and G. Charles L, "Characterization of an 80-kDa dehydrin-like protein in barley responsive to cold acclimation," *Physiologia Plantarum*, vol. 106, no. 2, pp. 177–183, 1999.
- [137] K. B. Singh, R. S. Malhotra, and M. C. Saxena, "Relationship between cold seventy and yield loss in chickpea (Cicer arietinuw L.)," *Journal of Agronomy and Crop Science*, vol. 170, pp. 121–127, 1993.
- [138] V. S. Bonthala, K. Mayes, J. Moreton et al., "Identification of gene modules associated with low temperatures response in bambara groundnut by network-based analysis," *PLoS ONE*, vol. 11, no. 2, Article ID e0148771, 2016.
- [139] D. Barabaschi, A. Tondelli, F. Desiderio et al., "Next generation breeding," *Journal of Plant Sciences*, vol. 242, pp. 3–13, 2016.
- [140] J. G. Schaart, C. C. M. van de Wiel, L. A. P. Lotz, and M. J. M. Smulders, "Opportunities for products of new plant breeding techniques," *Trends in Plant Science*, vol. 21, no. 5, pp. 438–449, 2016.
- [141] B. A. Akpinar, S. J. Lucas, and H. Budak, "Genomics approaches for crop improvement against abiotic stress," *The Scientific World Journal*, vol. 2013, Article ID 361921, 9 pages, 2013.
- [142] C. Kole, M. Muthamilarasan, R. Henry et al., "Application of genomics-assisted breeding for generation of climate resilient crops: Progress and prospects," *Frontiers in Plant Science*, vol. 6, article no. 563, 2015.
- [143] N. E. Borlaug, "Sixty-two years of fighting hunger: personal recollections," *Euphytica*, vol. 157, no. 3, pp. 287–297, 2007.
- [144] A. Diagne, S. K. G. Midingoyi, M. C. S. Wopereis, and I. Akintayo, "Increasing rice productivity and strengthening food security through new rice for Africa (NERICA)," in *Yes Africa Can: Success Stories from a Dynamic Continent*, P. Chuhan-Pole and M. Angwafo, Eds., pp. 253–267, The World Bank, Washington, DC, USA, 2011.
- [145] "FAO/IAEA programme's database of mutation enhanced technologies for agriculture (META)," http://mvgs.iaea.org/.
- [146] S. Uma, S. Lakshmi, M. S. Saraswathi, A. Akbar, and M. M. Mustaffa, "Embryo rescue and plant regeneration in banana (Musa spp.)," *Plant Cell, Tissue and Organ Culture*, vol. 105, no. 1, pp. 105–111, 2011.
- [147] R. J. Elshire, J. C. Glaubitz, Q. Sun et al., "A robust, simple genotyping-by-sequencing (GBS) approach for high diversity species," *PLoS ONE*, vol. 6, no. 5, Article ID e19379, 2011.
- [148] K. Zhao, C.-W. Tung, G. C. Eizenga et al., "Genome-wide association mapping reveals a rich genetic architecture of complex traits in Oryza sativa," *Nature Communications*, vol. 2, no. 1, article 467, 2011.
- [149] C. Kim, H. Guo, W. Kong, R. Chandnani, L.-S. Shuang, and A. H. Paterson, "Application of genotyping by sequencing technology to a variety of crop breeding programs," *Journal of Plant Sciences*, vol. 242, pp. 14–22, 2016.
- [150] H. Raman, B. Stodart, P. R. Ryan et al., "Genome-wide association analyses of common wheat (Triticum aestivum L.) germplasm identifies multiple loci for aluminium resistance," *Genome*, vol. 53, no. 11, pp. 957–966, 2010.
- [151] C. James, Global Status of Commercialized Biotech/GM Crops, 2014 vol. Brief 49, International Service for the Acquisition of Agri-biotech Applications, 2015.
- [152] E. Jacobsen and H. J. Schouten, "Cisgenesis strongly improves introgression breeding and induced translocation breeding of plants," *Trends in Biotechnology*, vol. 25, no. 5, pp. 219–223, 2007.

- [153] M. Araki and T. Ishii, "Towards social acceptance of plant breeding by genome editing," *Trends in Plant Science*, vol. 20, no. 3, pp. 145–149, 2015.
- [154] L. Bortesi and R. Fischer, "The CRISPR/Cas9 system for plant genome editing and beyond," *Biotechnology Advances*, vol. 33, no. 1, pp. 41–52, 2015.
- [155] A. S. Moffat, "Finding new ways to protect drought-stricken plants," *Science*, vol. 296, no. 5571, pp. 1226–1229, 2002.
- [156] S. Yang, B. Vanderbeld, J. Wan, and Y. Huang, "Narrowing down the targets: towards successful genetic engineering of droughttolerant crops," *Molecular Plant*, vol. 3, no. 3, pp. 469–490, 2010.
- [157] D. Todaka, K. Shinozaki, and K. Yamaguchi-Shinozaki, "Recent advances in the dissection of drought-stress regulatory networks and strategies for development of drought-tolerant transgenic rice plants," *Frontiers in Plant Science*, vol. 6, article no. 84, 2015.
- [158] P. Bhatnagar-Mathur, V. Vadez, and K. K. Sharma, "Transgenic approaches for abiotic stress tolerance in plants: retrospect and prospects," *Plant Cell Reports*, vol. 27, no. 3, pp. 411–424, 2008.
- [159] C. M. McCallum, L. Comai, E. A. Greene, and S. Henikoff, "Targeted screening for induced mutations," *Nature Biotechnology*, vol. 18, no. 4, pp. 455–457, 2000.
- [160] Z. Tadele, C. Mba, and B. J. Till, "TILLING for mutations in model plants and crops," in *Molecular Techniques in Crop Improvement*, S. M. Jain and D. S. Brar, Eds., pp. 307–332, Springer, 2nd edition, 2010.
- [161] K. Urano, Y. Kurihara, M. Seki, and K. Shinozaki, "Omics' analyses of regulatory networks in plant abiotic stress responses," *Current Opinion in Plant Biology*, vol. 13, no. 2, pp. 132–138, 2010.
- [162] S. Vij and A. K. Tyagi, "Emerging trends in the functional genomics of the abiotic stress response in crop plants: review article," *Plant Biotechnology Journal*, vol. 5, no. 3, pp. 361–380, 2007.
- [163] S. Rasool, P. Ahmad, M. U. Rehman, A. Arif, and N. A. Anjum, "Achieving crop stress tolerance and improvement—an overview of genomic techniques," *Applied Biochemistry and Biotechnology*, vol. 177, no. 7, pp. 1395–1408, 2015.
- [164] J. Amudha and G. Balasubramani, "Recent molecular advances to combat abiotic stress tolerance in crop plants," in *Biotechnology and Molecular Biology Reviews*, vol. 6, pp. 31–58, 2 edition, 2011.
- [165] L. F. Ciarmiello, P. Woodrow, A. Fuggi, G. Pontecorvo, and P. Carillo, "Plant genes for abiotic stress," in *Abiotic Stress in Plants: Mechanisms and Adaptations*, A. Shanker, Ed., Intech, 2011.
- [166] G. Cannarozzi, S. Plaza-Wüthrich, K. Esfeld et al., "Genome and transcriptome sequencing identifies breeding targets in the orphan crop tef (Eragrostis tef)," *BMC Genomics*, vol. 15, no. 1, article no. 581, 2014.
- [167] X. Zheng, B. Chen, G. Lu, and B. Han, "Overexpression of a NAC transcription factor enhances rice drought and salt tolerance," *Biochemical and Biophysical Research Communications*, vol. 379, no. 4, pp. 985–989, 2009.
- [168] H. Hu, M. Dai, J. Yao et al., "Overexpressing a NAM, ATAF, and CUC (NAC) transcription factor enhances drought resistance and salt tolerance in rice," *Proceedings of the National Acadamy* of Sciences of the United States of America, vol. 103, no. 35, pp. 12987–12992, 2006.
- [169] P. S. C. F. Rocha, "Plant abiotic stress-related RCI2/PMP3s: multigenes for multiple roles," *Planta*, vol. 243, no. 1, pp. 1–12, 2016.

- [170] P. Sornaraj, S. Luang, S. Lopato, and M. Hrmova, "Basic leucine zipper (bZIP) transcription factors involved in abiotic stresses: a molecular model of a wheat bZIP factor and implications of its structure in function," *Biochimica et Biophysica Acta (BBA)* -*General Subjects*, vol. 1860, no. 1, pp. 46–56, 2016.
- [171] N. L. Mantri, R. Ford, T. E. Coram, and E. C. K. Pang, "Transcriptional profiling of chickpea genes differentially regulated in response to high-salinity, cold and drought," *BMC Genomics*, vol. 8, article no. 303, 2007.
- [172] R. Garg, R. Shankar, B. Thakkar et al., "Transcriptome analyses reveal genotype- and developmental stage-specific molecular responses to drought and salinity stresses in chickpea," *Scientific Reports*, vol. 6, Article ID 19228, 2016.
- [173] V. Shriram, V. Kumar, R. M. Devarumath, T. S. Khare, and S. H. Wani, "MicroRNAs as potential targets for abiotic stress tolerance in plants," *Frontiers in Plant Science*, vol. 7, article 817, 2016.
- [174] B. Zhang, "MicroRNA: a new target for improving plant tolerance to abiotic stress," *Journal of Experimental Botany*, vol. 66, no. 7, pp. 1749–1761, 2015.
- [175] A. Yadav, Y. Khan, and M. Prasad, "Dehydration-responsive miRNAs in foxtail millet: genome-wide identification, characterization and expression profiling," *Planta*, vol. 243, no. 3, pp. 749–766, 2016.
- [176] D.-H. Jeong and P. J. Green, "The role of rice microRNAs in abiotic stress responses," *Journal of Plant Biology*, vol. 56, no. 4, pp. 187–197, 2013.
- [177] G. M. Ali and S. Komatsu, "Proteomic analysis of rice leaf sheath during drought stress," *Journal of Proteome Research*, vol. 5, no. 2, pp. 396–403, 2006.
- [178] R. Ngara and B. K. Ndimba, "Understanding the complex nature of salinity and drought-stress response in cereals using proteomics technologies," *Proteomics*, vol. 14, no. 4-5, pp. 611– 621, 2014.
- [179] M. A. Altieri, "Agroecology: the science of natural resource management for poor farmers in marginal environments," *Agriculture, Ecosystems & Environment*, vol. 93, no. 1-3, pp. 1– 24, 2002.
- [180] I. W. Nyakudya and L. Stroosnijder, "Conservation tillage of rainfed maize in semi-arid Zimbabwe: a review," *Soil & Tillage Research*, vol. 145, pp. 184–197, 2015.
- [181] SRI, The System of Crop Intensification: Agroecological Innovations for Improving Agricultural Production, Food Security, and Resilience to Climate Change, Cornell University, Ithaca, NY, USA; Technical Centre for Agricultural and Rural Cooperation, Wageningen, Netherlands, 2014.
- [182] "SRI international network and resources center," http://sri .cals.cornell.edu/.
- [183] T. A. Dar, M. Uddin, M. M. A. Khan, K. R. Hakeem, and H. Jaleel, "Jasmonates counter plant stress: a review," *Environmen*tal and Experimental Botany, vol. 115, pp. 49–57, 2015.
- [184] S. Plaza-Wüthrich, R. Blösch, A. Rindisbacher, G. Cannarozzi, and Z. Tadele, "Gibberellin deficiency confers both lodging and drought tolerance in small cereals," *Frontiers in Plant Science*, vol. 7, article no. 643, 2016.
- [185] C. V. Ha, M. A. Leyva-Gonzalez, Y. Osakabe et al., "Positive regulatory role of strigolactone in plant responses to drought and salt stress," *Proceedings of the National Acadamy of Sciences* of the United States of America, vol. 111, no. 2, pp. 851–856, 2014.
- [186] M. Fisher, T. Abate, R. W. Lunduka, W. Asnake, Y. Alemayehu, and R. B. Madulu, "Drought tolerant maize for farmer

adaptation to drought in sub-Saharan Africa: determinants of adoption in eastern and southern Africa," *Climatic Change*, vol. 133, no. 2, pp. 283–299, 2015.



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