Clinical-Pathological Conference



Wien Klin Wochenschr (2015) 127:974–980 DOI 10.1007/s00508-015-0882-8

Wiener klinische Wochenschrift The Central European Journal of Medicine

Clinical-Pathological Conference Series from the Medical University of Graz



Case No 156: 82-year-old woman with chronic diarrhea and weight loss of 20 kilograms

Elisabeth Fabian · Dietmar Schiller · Heimo Wenzl · Carolin Lackner · Josef Donnerer · Alexander Ziachehabi · Rene Silye · Rainer Schöfl · Guenter J. Krejs

Received: 17 June 2015 / Accepted: 10 October 2015 / Published online: 26 November 2015 © The Author(s) 2015. This article is published with open access at Springerlink.com

Presentation of case

Keywords Chronic diarrhea · Sprue · Subtotal villous atrophy · Olmesartan · Drug-associated enteropathy · Adverse drug effect · Delayed hypersensitivity reaction · Hypokalemia

G. J. Krejs, MD, AGAF (⊠) · H. Wenzl Division of Gastroenterology and Hepatology, Department of Internal Medicine, Medical University of Graz, Auenbruggerplatz 15, 8036 Graz, Austria e-mail: guenter.krejs@medunigraz.at

E. Fabian Division of Gastroenterology and Hepatology, Department of Internal Medicine III, Medical University of Vienna, Vienna, Austria

D. Schiller · A. Ziachehabi · R. Schöfl Department of Internal Medicine IV, Elisabethinen Hospital, Linz, Austria

C. Lackner Department of Pathology, Medical University of Graz, Graz, Austria

J. Donnerer Department of Experimental and Clinical Pharmacology, Medical University of Graz, Graz, Austria

R. Silye Department of Clinical Pathology, General Hospital, Linz, Austria *Dr. D. Schiller:* After two previous hospitalizations, an 82-yearold woman was readmitted with chronic diarrhea of 8-month duration and weight loss of 20 kg. This had left her so weak that she could no longer look after herself.

The watery diarrhea had started 8 months before and sometimes was worse at night. She had up to 12 bowel movements per day without abdominal pain and had lost her appetite. Duodenal biopsies revealed subtotal villous atrophy with crypt hyperplasia and increased intraepithelial lymphocytes (40 per 100 epithelial cells). Tissue transglutaminase antibodies and serum anti-endomysial antibodies were negative. Serum immunoglobulins (IgG, IgA, and IgM), thyroid stimulating hormone (TSH), differential blood count, anti-HIV antibody, ova and parasites in stool (three times), colonoscopy with multiple biopsies, capsule endoscopy of the small bowel, and abdominal computerized tomography were all negative or normal. Biopsies from ileoscopy showed changes similar to the proximal small bowel. Human leukocyte antigen (HLA)-DO2 (positive) and DO8 (negative) genotyping were compatible with celiac disease. Intestinal lymphocytes showed no T-cell receptor rearrangement. A strict gluten-free diet did not ameliorate the diarrhea but it improved slightly under budesonide medication. Finally, during this third hospitalization, a diagnosis of "refractory sero-negative celiac disease" was established.

Her medical history was positive for paroxysmal atrial fibrillation, hypertension, hypothyroidism following autoimmune thyreoiditis and chronic depression. Her history was negative for previous gastrointestinal complaints, foreign travel, and fever. Her family history was non-revealing.

Her medications included budesonide, amiodarone, olmesartan, amlodipine, thyroid replacement therapy (levothyroxine, liothyronine), trazodone, risperidone, and lorazepam.

On admission the patient was afebrile, body weight was 45 kg, height 162 cm; she was in poor general and nutritional condition (body mass index: 17.1 kg/m²). Blood pressure was 135/90 mmHg, bowel sounds were hyperactive. She appeared slightly dehydrated. Abnormal laboratory results included serum potassium 3.3 mmol/l (normal: 3.6–4.8 mmol/l), C-reactive protein (CRP) 1.5 mg/l (normal: <0.5 mg/l), and hemoglobin 11.1 g/dl (normal: 13.0–17.5 g/dl). All other results were negative or normal including urine dip stick, fecal occult blood, and calprotectin.

A diagnostic insight allowed an explanation for the severe diarrhea.

Dr. C. Lackner: Histological examination of the duodenal biopsies mentioned in the protocol revealed subtotal villous atrophy, chronic inflammation of the lamina propria and increased intraepithelial lymphocytes (approximately 40 per 100 epithelial cells), reactive and degenerative epithelial changes as well as crypt hyperplasia (Fig. 1). These histopathological features are not very specific and can be found with celiac disease, Crohn's disease, enteric infection, collagenous sprue, tropical sprue, bacterial overgrowth, common variable immunodeficiency, autoimmune enteropathy, and hematological malignancies, and have also been reported as side effects of immunosuppressant drugs [1, 2]. In this case, small bowel histology is compatible with a wide spectrum of diseases.

Differential diagnosis

Dr. H. Wenzl: This is a case of chronic diarrhea. When a patient presents with this condition, a detailed medical history is pivotal for establishing the correct diagnosis. The patient under discussion is an elderly woman who had been suffering from chronic watery diarrhea for 8 months. She reported up to 12 bowel movements in 24 h, even at night. This important finding suggests an organic rather than functional cause for her chronic diarrhea [3]. Osmotic diarrhea can also be excluded in cases with noctural diarrhea because osmotic diarrhea always follows shortly after meals, so that it usually subsides during the night. Abdominal pain would be typical for inflammatory bowel diseases such as Crohn's disease, irritable bowel syndrome or ischemic colitis but was not present here. The negative hemoccult and calprotectin speak against inflammatory mucosal diseases, as did the normal colon biopsies. A weight loss of 20 kg is remarkable and suggests severe bowel dysfunction including malabsorption, chronic inflammation and neoplasia, and demands extensive morphological examination. The patient underwent a pan-endoscopy including esoph-

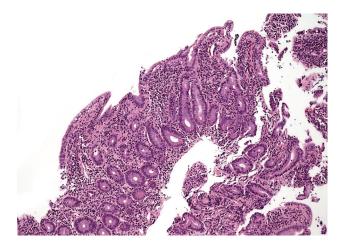


Fig. 1 Duodenal biopsy showing subtotal villous atrophy and crypt hyerplasia (H&E, $\times 100$)

ago-gastro-duodenal endoscopy, ileocolonoscopy with multiple biopsies, and capsule endoscopy of the small bowel. The protocol does not describe the macroscopic appearance of the duodenum and ileum, but histological assessment documented subtotal villous atrophy, chronic inflammation, and crypt hyperplasia suggesting celiac disease. In 20-25% of patients, celiac disease first manifests itself at the age of 60 years or older [4], so based on histology alone this diagnosis could indeed be made for this 82-year-old patient. Tests for celiac disease with antibodies to tissue transglutaminase and endomysium were, however, negative. Both parameters have a high sensitivity and specificity for celiac disease [5]. Serum IgA levels were normal. Her HLA-type was DQ2 positive and DQ8 negative. In Europe, 85-90% of patients with celiac disease are HLA-DQ2 positive and 10-15% are HLA-DQ8 positive [6]; however, up to 30% of the general population is also HLA-DQ2 or DQ8 positive but without the disease [7]. Celiac disease is defined as immune-mediated enteropathy with a strong genetic predisposition, which improves upon dietary exclusion of gluten [8]. Gluten is digested by luminal and brush-border enzymes into amino acids and polypeptides. The gliadin peptides induce immunological changes in the epithelium and in the lamina propria. Gliadin damages epithelial cells, leading to increased expression of interleukin-15, which in turn activates intraepithelial lymphocytes. These lymphocytes are cytotoxic and damage enterocytes expressing the surface protein MIC-A (major-histocompatibilitycomplex class I chain-related A). During infections or as the result of permeability changes, gliadin can enter the lamina propria, where it is deamidated by tissue transglutaminase, allowing interaction with HLA-DQ2 or HLA-DQ8 on the surface of antigen-presenting cells. Presentation of gliadin to gliadin-reactive CD4 + T cells through a T cell receptor thus results in production of cytokines that cause tissue damage, subsequently leading to villous atrophy, crypt hyperplasia, and activation and expansion of B cells that produce antibodies [7].

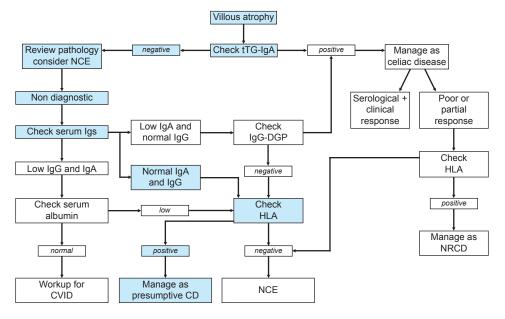


Fig. 2 Diagnostic algorithm for small intestinal villous atrophy [15]. The diagnostic path followed with our patient is marked in *blue*. When the result ("Manage as presumptive CD") did not satisfy the clinicians they returned to square "Review pathology, consider NCE" (*left upper corner*). Only when at this point additional other differential diagnoses were considered

Treatment for celiac disease is a gluten-free diet, which usually results in recovery of the mucosa and return to normal health. Our patient, however, did not respond to this diet, as is the case with 0.3-30% of patients with celiac disease, who are subsequently diagnosed as refactory celiac disease (RCD) [9-12]. There are two types of RCD: type 1 shows a normal intraepithelial lymphocyte population, whereas type 2 displays a predominantly aberrant intraepithelial lymphocyte phenotype [13]. Taking all these findings together, our patient could have RCD type I, but I think this diagnosis has to be questioned because she is an elderly woman who might have had trouble following a strict gluten-free diet. Moreover, we speak of non-responsive sprue if patients do not respond to strict gluten-free diet for 6-12 months and of RCD if patients do not respond to strict gluten-free diet for \geq 12 months and other causes of villous atrophy have been excluded [14]. Based on the information given in the protocol, our patient had, however, not been on a gluten-free diet long enough and as already mentioned, the elderly patient may not have been able to comply with it. Still, the negative celiac serology and the supposed nonresponse to a gluten-free diet urge us to consider other non-celiac enteropathies (NCE) presenting with villous atrophy. The diagnostic algorithm for small intestinal villous atrophy (Fig. 2) indicates that other etiologies of NCE have to be considered here [15]. These include common variable immunodeficiency, food allergies, tropical sprue, post viral enteropathy, eosinophilic gastroenteritis, peptic duodenitis, autoimmune enteropathy, small intestinal bacterial overgrowth, immune-mediated enteropathy, Crohn's disease, and collagenous sprue

(see text), did her physicians arrive at the right diagnosis. *tTG* tissue transglutaminase, *Ig* immunoglobulin, *NCE* non-celiac enteropathy, *DGP* deamidated gliadin peptide, *HLA* human leukocyte antigen, *CVID* common variable immune deficiency, *CD* celiac disease, *NRCD* non-responsive celiac disease

[15]. Other diseases with small intestinal villous atrophy that merit consideration are giardiasis, HIV enteropathy, tuberculosis, radiation enteritis, Zollinger-Ellison syndrome, Whipple's disease, intestinal lymphoma, food intolerances [16], alpha chain disease, graft-versus-host disease, hypogammaglobulinemia, severe malnutrition [5], and certain medications [17]. The medical history shows no evidence for most of these diseases; there are no further specific histopathological findings and there is no other serological abnormality. This leads us to limit the differential diagnosis to non-responsive sprue, postgastroenteritis syndrome, small intestinal bacterial overgrowth, autoimmune enteropathy, and Zollinger-Ellison syndrome. A strict gluten-free diet over a longer period of time would show whether the patient suffers from truly non-responsive sprue. A post-gastroenteritis syndrome would be self-limiting ("watch and wait"); small intestinal bacterial overgrowth could be sought with an H₂-breath test and analysis of duodenal aspirate, and treated with oral antibiotics; an autoimmune enteropathy could be diagnosed by an experienced gastrointestinal pathologist's review of the biopsies and enterocyte antibodies; and in case of Zollinger-Ellison syndrome, serum gastrin would be increased, usually together with a tumor in the pancreas or duodenal wall. Actually, none of these diagnoses really seems to be likely in our patient. According to our list of potential causes for NCE, only a careful review of the patient's medication is left, because some drugs might have adverse gastrointestinal effects (Table 1). Of the eight different medications our patient took, five document diarrhea as an adverse effect. Diarrhea is indeed a ommon drug-associated adverse

 Table 1
 Patient's medication and documented adverse gastrointestinal effects (Summary of Product Characteristics)

Medication	Gastrointestinal side effect	
	Diarrhea	Villous atrophy
Budenoside	-	-
Amiodarone	-	-
Lorazepam	-	-
Amlodipine	+	-
Olmesartan	+	+
Trazodone	+	-
Risperidone	+	-
Levothyroxine/liothyronine	+	-

event, and the underlying mechanisms are often unclear [18]. Diarrhea due to drug-induced enteropathy has been reported for medications such as mycophenolate mofetil [19, 20], methotrexate [21, 22], azathioprine [23] and most recently olmesartan. Among the medications taken by our patient, only one, olmesartan, is associated with villous atrophy resulting in sprue-like enteropathy. Olmesartan is an angiotensin II receptor antagonist that has been commonly prescribed for treatment of hypertension since 2002 [24]. The association between spruelike enteropathy and olmesartan was first reported in 2012 [18]. Clinical features of this enteropathy include gastrointestinal symptoms such as diarrhea, weight loss and steatorrhea, negative IgA tissue transglutaminase antibodies (or endomysial-antibodies), evidence of enteropathy (villous atrophy) with or without collagen deposition or intraepithelial lymphocytes, lack of clinical response to a gluten-free diet, and exclusion of other causes of enteropathy [18]. Since our patient took olmesartan and presented with unexplained diarrhea, I suggest that she suffers from olmesartan-associated sprue-like enteropathy. Symptoms typically occur after olmesartan has been taken for months or years; this enteropathy can affect almost the entire gut [18], and can present a spectrum of histological damage (from normal villi to partial or total villous atrophy with variable degrees of mucosal inflammation) [25]. The mechanisms underlying olmesartan-associated enteropathy are still unclear. Interestingly, approximately 70% of patients with olmesartan-associated enteropathy are HLA-DQ2 or HLA-DQ8 positive, which is a higher prevalence than expected in the general population (25-30%) [26, 27]. This suggests that the presence of HLA-DQ2 or HLA-DQ8 may increase the risk for immune-mediated mucosal damage in these patients, but further studies must clarify this. Withdrawal of olmesartan intake usually results in rapid clinical and histological improvement of the enteropathy. Withdrawal of olmesartan is thus indicated for our patient.

Dr. H. Wenzl's diagnosis

Olmesartan-associated sprue-like enteropathy.

Discussion of diagnosis

Dr. G. J. Krejs: Dr. Schiller was the attending physician who cared for the patient and will describe further details of the patient's course.

Dr. D. Schiller: Arriving at a diagnosis in this case was challenging. Within a short period of time we observed a second patient with the same clinical picture and enteropathy. When we contacted Dr. Nadine Cerf-Bensussan at the Laboratory for Intestinal Immunity in Paris requesting determination of anti-enterocyte antibodies (which subsequently turned out to be negative in both patients), we were asked the decisive question of whether our patients were taking olmesartan. Discontinuation of the medication resolved the diarrhea within several days and both patients began to gain weight. Although there are many potential causes for NCE, gastroenterologists should be aware that olmesartan can cause enteropathy. Olmesartan has now been reported to be associated with sprue-like enteropathy [18, 20, 23, 25]. Our first two cases of olmesartan-associated enteropathy are currently being published as a clinical vignette in the journal "Gut" [28].

Meanwhile, a third patient presented with olmesartan-associated enteropathy at our hospital in Linz. All patients received a primary diagnosis of non-responsive/refractory celiac disease or unexplained sprue. Typical symptoms such as diarrhea and massive weight loss (10-20 kg) always occurred after a longer period of olmesartan intake. In all patients, clinical response was rapid after olmesartan was discontinued and histological recovery was confirmed in all three patients by followup biopsy within 2 months. Since our patients' recoveries were so striking and they had already been suffering for so long, we did not want to rechallenge them with olmesartan to reproduce the chronic diarrhea and villous atrophy.

Dr. G. J. Krejs: Dr. Lackner, our pathologist, will show and explain the follow-up biopsy after the patient had been off olmesartan for 2 months.

Dr. C. Lackner: Follow-up duodenal biopsy showed complete mucosal recovery. The initially described histopathological features including villous atrophy, chronic inflammation, reactive and degenerative epithelial alterations, and crypt hyperplasia were not present in the specimen after discontinuation of olmesartan. Villus:crypt ratio was within normal range (Fig. 3).

Dr. G. J. Krejs: Currently, an association between olmesartan intake and development of sprue-like enteropathy, mainly characterized by diarrhea, weight loss and variable degrees of duodenal mucosal damage is being noted by gastroenterologists and some 90 cases have been documented worldwide. Dr. Donnerer is a clinical pharmacologist and will explain possible mechanisms of olmesartan-associated enteropathy.

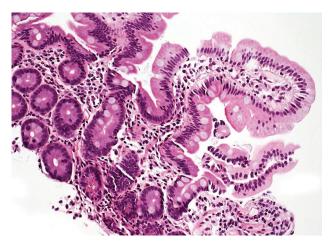


Fig. 3 Normal villus and crypt architecture in duodenal biopsy 3 months after discontinuation of olmesartan (H&E, ×200)

Dr. J. Donnerer: Generally, many drugs can exert adverse effects involving different organ systems and tissues. When medication is administered orally, the gastrointestinal tract is often affected by adverse events for the simple reason of being exposed to high drug concentrations. Olmesartan is an orally administered prodrug (olmesartan medoxomil) that is rapidly metabolized to the active component (olmesartan) by esterases in the gastrointestinal mucosa, portal blood and liver [29]. The mechanisms involved in olmesartan-associated enteropathy are still not fully understood.

A delayed drug hypersensitivity reaction may be the underlying mechanism leading to enteropathy under olmesartan therapy. Some medications can provoke allergic type B reactions if the pharmaceutical substance acts as a hapten and combines with a body's own protein to form an antigen, thus inducing allergization. Generally, there are four different types of hypersensitivity reactions which can result in drug-induced allergization: type I (anaphylaxis), type II (cytotoxic effects), type III (immune-complex hypersensitivity), and type IV (T lymphocyte-mediated reactions, delayed hypersensitivity). Considering the long period of months or years between onset of olmesartan therapy and appearance of villous atrophy, it is suggested that a delayed type IV hypersensitivity reaction may cause the sprue-like enteropathy.

Some angiotensin (AT) receptor blockers are suggested to have inhibitory effects on transforming growth factor-beta 1 [30, 31], which is pivotal for maintaining gut immune homeostasis [32, 33]. Duodenal biopsies from patients under olmesartan therapy show a significant increase in the number of CD8 + cells, assuming that olmesartan-associated sprue-like enteropathy may be due to either an expansion of cytotoxic CD8 + T cells, or a dysfunctional intestinal regulatory mechanism that normally suppresses CD8 + T cell activity [34].

Another possible mechanism underlying villous atrophy in olmesartan-associated enteropathy might be a pro-apoptotic effect on intestinal epithelial cells mediated by angiotensin II. In the human gut, angiotensin II binds to two forms of receptors with different properties. AT1 receptor is expressed throughout the gastrointestinal tract; it activates growth-promoting factors and mediates major effects of angiotensin II. AT2 is mainly found in the duodenum and jejunum, and induces effects opposing those of AT1 [35, 36]. Recent studies have shown that binding of angiotensin II to AT2 promotes apoptosis of enterocytes by up-regulation of pro-apoptotic protein associated with a down-regulation of anti-apoptotic protein [37]. Moreover, in rat smooth muscle cells, druginduced AT1 receptor blocking has been found to cause translocation of AT2 receptors from cytosol to external membrane, which may favor binding of angiotensin II to AT2 [38]. Olmesartan has a high affinity for AT1 receptors; in the case of drug-mediated saturation of these receptors, angiotensin II could increasingly bind to AT2 receptors and thus induce apoptotic effects, leading to villous atrophy without inflammation and increase in intraepithelial lymphocytes [1]. This mechanism is, however, still controversial because such effects have not been described for other angiotensin II receptor antagonists, which would be expected since interaction with the AT1 receptor is the basic therapeutic mechanism underlying this group of antihypertensives.

At present, olmesartan-associated sprue-like enteropathy may be considered an adverse effect. Since the underlying mechanisms are still unclear, more studies involving rechallenge with either olmesartan or placebo are needed. At this time a cause-effect relationship should be classified as only probable [39]. A similar enteropathy has been reported after telmisartan and irbesartan, and it is open to question whether these changes could be a class effect of sartans [40-42]. In relation to the worldwide use of olmesartan for antihypertensive therapy, olmesartan-associated sprue-like enteropathy occurs only rarely. However, doctors should be aware of this association when a patient under therapy with olmesartan presents with unexplained chronic diarrhea. The regulatory authorities have recently ordered that a special warning regarding sprue-like enteropathy be included in the SPC (Summary of Product Characteristics) of olmesartan medoxomil; however, since this side effect is so rare, no suggestion has been made to take olmesatran off the market.

One other comment on the medication of this patient is that she received three drugs (trazodone, risperidone, and lorazepam) that target the central nervous system (CNS). The weakness mentioned at the beginning of the protocol was probably due to her dramatic weight loss and possibly to hypokalemia, but the CNS effects of her medication should also be considered and the necessity for all the medication she is receiving should be reevaluated.

Dr. G. J. Krejs: Dr. Wenzl, as a diarrhea expert, what do you say about the hypokalemia albeit mild in this patient? When does chronic diarrhea lead to hypokalemia? And what is your final comment on this case?

Clinical-Pathological Conference

Dr. H. Wenzl: Hypokalemia is always a sign of chronic largevolume diarrhea. The transport mechanisms in the colon try to conserve sodium but potassium has to go into the lumen in exchange. With a normal stool, 5 meq of potassium are discharged every day; with a liter of watery stool there is a potassium loss of 50 meq/l per day [43-45]. This slowly depletes total body potassium (50 meq/kg) and ultimately results in hypokalemia. Hypokalemia is prominent in pancreatic cholera or Verner-Morrison syndrome, where it is part of the name of the condition (WDHH syndrome or watery diarrhea hypochlorhydria hypokalemia syndrome) [46]. It is also observed with chronic laxative abuse. That the patient had slight hypokalemia indicates long-standing large-volume diarrhea.

This case is very interesting and emphasizes the diagnostic dilemma when patients with villous atrophy have either negative celiac serology or do not respond to a gluten-free diet. Under these circumstances, a broad spectrum of other etiologies for enteropathy has to be considered. A side effect of medication as a cause, especially when olmesartan is involved, should be part of the initial differential diagnosis of such unexplained enteropathy, as the newer guidelines now suggest [18, 47].

Final diagnosis

Olmesartan-associated sprue-like enteropathy.

Acknowledgments

The authors express their sincere gratitude to Eugenia Lamont for language editing of the manuscript.

Conflict of interest

All authors declare that there are no conflicts of interest.

Open Access This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

References

- 1. Ianiro G, Bibbo S, Montalto M, Ricci R, Gasbarrini A, Cammarota G. Systematic review: sprue-like enteropathy associated with olmesartan. Aliment Pharmacol Ther. 2014;40(1):16-23.
- Greenson JK. The biopsy pathology of non-coeliac enteropathy. Histopathology. 2015;66(1):29–36.
- 3. Krejs GJ. Cecil textbook of medicine. 19th ed. Philadelphia: WB Saunders; 1992. pp. 680-7. (Chapter: Diarrhea).
- 4. Beaumont DM, Mian MS. Coeliac disease in old age: "a catch in the rye". Age Ageing. 1998;27(4):535-8.
- 5. Feldmann M, Friedmann LS, Brandt LJ, editors. Sleisenger and Fordtran's gastrointestinal and liver disease. 10th ed. Philadelphia: Elsevier Sounders; 2015.
- Megiorni F, Mora B, Bonamico M, Berbato M, Nenna R, Maiella G, Lulli P, Mazzilli MC. HLA-DQ and risk gradient for celiac disease. Hum Immunol. 2009;70(1):55–9.

- Green PHR, Cellier C. Celiac disease. N Engl J Med. 2007;357:1731-43.
- 8. Walker MM, Murray JA. An update in the diagnosis of coeliac disease. Histopathology. 2011;59(2):166-79.
- 9. Ciclitira PJ, King AL, Fraser JS. AGA technical review on celiac sprue. American Gastroenterological Association. Gastroenterology. 2001;120(6):1526-40.
- Cellier C, Cerf-Bensussan N. Treatment of clonal refractory celiac disease or cryptic intraepithelial lymphoma: a long road from bench to bedside. Clin Gastroenterol Hepatol. 2006;4(11):1320-1.
- 11. Wahab PJ, Meijer JW, Goerres MS, Mulder CJ. Coeliac disease: changing views on gluten-sensitive enteropathy. Scand J Gastroenterol. 2002;236:60–5.
- Ilus T, Kaukinen K, Virta LJ, Huhtala H, Mäki M, Kurppa K, Heikkinen M, Heikura M, Hirsi E, Jantunen K, Moilanen V, Nielsen C, Puhto M, Pölkki H, Vihriälä I, Collin P. Refractory coeliac disease in a country with a high prevalence of clinically-diagnosed coeliac disease. Aliment Pharmacol Ther. 2014;39(4):418–25.
- 13. Daum S, Cellier C, Mulder CJ. Refractory coeliac disease. Best Pract Res Clin Gastroenterol. 2005;19(3):413-24.
- Rubio-Tapia A, Hill ID, Kelly CP, Calderwood AH, Murray JA, American College of Gastroenterology. ACG clinical guidelines: diagnosis and management of celiac disease. Am J Gastroenterol. 2013;108(5):656-76.
- Pallav K, Leffler DA, Tariq S, Kabbani T, Hansen J, Peer A, Bhansali A, Najarian R, Kelly CP. Noncoeliac enteropathy: the differential diagnosis of villous atrophy in contemporary clinical practice. Aliment Pharmacol Ther. 2012;35(3):380–90.
- Yamada T, Alpers DH, Kalloo AN, Kaplowitz N, Owyang C, Powell DW, editors. Textbook of gastroenterology. West Sussex: Blackwell Publishing Ltd; 2009.
- 17. Kelly CP, Lamont JT, Grover S, editors. Diagnosis of celiac disease. Waltham: UpToDate; 2015. (Post TW).
- Rubio-Tapia A, Herman ML, Ludvigsson JF, Kelly DG, Mangan TF, Wu TT, Murray JA. Severe spruelike enteropathy associated with olmesartan. Mayo Clin Proc. 2012;87(8):732–8.
- Kamar N, Faure P, Dupuis E, Cointault O, Joseph-Hein K, Durand D, Moreau J, Rostaing L. Villous atrophy induced by mycophenolate mofetil in renal-transplant patients. Transpl Int. 2004;17(8):463–7.
- Weclawiak H, Ould-Mohamed A, Bournet B, Guilbeau-Frugier C, Fortenfant F, Muscari F, Sallusto F, Dambrin C, Esposito L, Guitard J, Abbal M, Rostaing L, Kamar N. Duodenal villous atrophy: a cause of chronic diarrhea after solid-organ transplantation. Am J Transplant. 2011;11(3):575-82.
- Boscá MM, Añón R, Mayordomo E, Villagrasa R, Balza N, Amorós C, Corts JR, Benages A. Methotrexate induced sprue-like syndrome. World J Gastroenterol. 2008;14(45):7009-11.
- 22. Houtman PM, Hofstra SS, Spoelstra P. Non-coeliac sprue possibly related to methotrexate in a rheumatoid arthritis patient. Neth J Med. 1995;47(3):113–6.
- Ziegler TR, Fernández-Estívariz C, Gu LH, Fried MW, Leader LM. Severe villus atrophy and chronic malabsorption induced by azathioprine. Gastroenterology. 2003;124(7):1950-7.
- 24. The Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation. and Treatment of High Blood Pressure. Bethesda, MD: National Institutes of Health, National Heart, Lung, and Blood Institute, US Department of Health and Human Services, 2004. NIH Publication No. 04-5230.

- 25. Marthey L, Cadiot G, Seksik P, Pouderoux P, Lacroute J, Skinazi F, Mesnard B, Chayvialle JA, Savoye G, Druez A, Parlier D, Abitbol V, Gompel M, Eoche M, Poncin E, Bobichon R, Colardelle P, Wils P, Salloum H, Peschard S, Zerbib F, Méresse B, Cerf-Bensussan N, Malamut G, Carbonnel F. Olmesartan-associated enteropathy: results of a national survey. Aliment Pharmacol Ther. 2014;40(9):1103–9.
- Sollid LM, Markussen G, Ek J, Gjerde H, Vartdal F, Thorsby E. Evidence for a primary association of celiac disease to a particular HLA-DQ alpha/beta heterodimer. J Exp Med. 1989;169(1):345–50.
- 27. Kaukinen K, Partanen J, Mäki M, Collin P. HLA-DQ typing in the diagnosis of celiac disease. Am J Gastroenterol. 2002;97(3):695–9.
- Schiller D, Ziachehabi A, Silye R, Schöfl R. Two coincident cases of easily curable "refractory sprue". Gut. 2015;64(11):1773.
- 29. Scott LJ, McCormack PL. Olmesartan medoxomil: a review of its use in the management of hypertension. Drugs. 2008;68(9):1239-72.
- 30. Matt P, Schoenhoff F, Habashi J, Holm T, Van Erp C, Loch D, Carlson OD, Griswold BF, Fu Q, De Backer J, Loeys B, Huso DL, McDonnell NB, Van Eyk JE, Dietz HC, GenTAC Consortium. Circulating transforming growth factor-beta in Marfan syndrome. Circulation. 2009;120(6):526–32.
- Kagami S, Border WA, Miller DE, Noble NA. Angiotensin II stimulates extracellular matrix protein synthesis through induction of transforming growth factor-beta expression in rat glomerular mesangial cells. J Clin Invest. 1994;93(6):2431-7.
- 32. Macdonald TT, Monteleone G. Immunity, inflammation, and allergy in the gut. Science. 2005;307(5717):1920–5.
- Coombes JL, Robinson NJ, Maloy KJ, Uhlig HH, Powrie F. Regulatory T cells and intestinal homeostasis. Immunol Rev. 2005;204:184–94.
- 34. Rubio-Tapia A. Sprue-like enteropathy associated with olmesartan—broadening the differential diagnosis of enteropathy. Aliment Pharmacol Ther. 2014;40(11-12):1362-3.
- Fändriks L. The angiotensin II type 2 receptor and the gastrointestinal tract. J Renin Angiotensin Aldosterone Syst. 2010;11(1):43-8.
- Mavromoustakos T, Agelis G, Durdagi S. AT1 antagonists: a patent review (2008-2012). Expert Opin Ther Pat. 2013;23(11):1483-94.

- 37. Sun L, Wang W, Xiao W, Liang H, Yang Y, Yang H. Angiotensin II induces apoptosis in intestinal epithelial cells through the AT2 receptor, GATA-6 and the Bax pathway. Biochem Biophys Res Commun. 2012;424(4):663–8.
- De Godoy MA, Rattan S. Translocation of AT1- and AT2receptors by higher concentrations of angiotensin II in the smooth muscle cells of rat internal anal sphincter. J Pharmacol Exp Ther. 2006;319(3):1088-95.
- Naranjo CA, Busto U, Sellers EM, Sandor P, Ruiz I, Roberts EA, Janecek E, Domecq C, Greenblatt DJ. A method for estimating the probability of adverse drug reactions. Clin Pharmacol Ther. 1981;30(2):239-45.
- Cyrano J, Vasatko T, Machac J, Nova M, Szanyi J, Kopacova M. Telmisartan-associated enteropathy—is there any class effect? Aliment Pharmacol Ther. 2014;40(5):569-70.
- 41. Cammarota G, Ianiro G, Bibbo S, Gasbarrini A. Telmisartan associated enteropthy—is there any class effect? Aliment Pharmacol Ther. 2014;40(5):570.
- 42. Marietta EV, Cartee A, Rishi A, Murray JA. Drug-induced enteropathy. Dig Dis. 2015;33(2):215-20.
- Wenzl HH, Fine KD, Santa Ana CA, Porter JL, Fordtran JS. Effect of fludrocortisone and spironolactone on sodium and potassium losses in secretory diarrhea. Dig Dis Sci. 1997;42(1):119–28.
- 44. Agarwal R, Afzalpurkar R, Fordtran JS. Pathophysiology of potassium absorption and secretion by the human intestine. Gastroenterology. 1994;107(2):548–71.
- Giebisch G. Challenges to potassium metabolism: internal distribution and external balance. Wien Klin Wochenschr. 2004;116(11-12):353-66.
- Krejs GJ. VIPoma syndrome. Am J Med. 1987;82(Suppl. 513): 37–48.
- 47. Ludvigsson JF, Bai JC, Biagi F, Card TR, Ciacci C, Ciclitira PJ, Green PH, Hadjivassiliou M, Holdoway A, van Heel DA, Kaukinen K, Leffler DA, Leonard JN, Lundin KE, McGough N, Davidson M, Murray JA, Swift GL, Walker MM, Zingone F, Sanders DS, BSG Coeliac Disease Guide-lines Development Group, British Society of Gastroenterology. Diagnosis and management of adult coeliac disease: guidelines from the British Society of Gastroenterology. Gut. 2014;63(8):1210-1228.