

## Review Article

# Distinct Functions of Neutrophil in Cancer and Its Regulation

Zvi Granot<sup>1</sup> and Jadwiga Jablonska<sup>2</sup>

<sup>1</sup>Department of Developmental Biology and Cancer Research, The Institute for Medical Research Israel-Canada, The Hebrew University-Hadassah Medical School, 91120 Jerusalem, Israel

<sup>2</sup>Translational Oncology, Internal Medicine II, University Hospital, Tuebingen, Otfried-Mueller-Street 10, 72076 Tuebingen, Germany

Correspondence should be addressed to Zvi Granot; [zvikag@ekmd.huji.ac.il](mailto:zvikag@ekmd.huji.ac.il) and Jadwiga Jablonska; [jadwiga.jablonska-koch@med.uni-tuebingen.de](mailto:jadwiga.jablonska-koch@med.uni-tuebingen.de)

Received 24 August 2015; Accepted 27 October 2015

Academic Editor: Ronald Gladue

Copyright © 2015 Z. Granot and J. Jablonska. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Neutrophils are the most abundant of all white blood cells in the human circulation and are usually associated with inflammation and with fighting infections. In recent years the role immune cells play in cancer has been a matter of increasing interest. In this context the function of neutrophils is controversial as neutrophils were shown to possess both tumor promoting and tumor limiting properties. Here we provide an up-to-date review of the pro- and antitumor properties neutrophils possess as well as the environmental cues that regulate these distinct functions.

## 1. Introduction

Neutrophils are the most abundant of all white blood cells and play a key role in host protection against microbial infections and in inflammation. Chronic inflammation has been associated with increased susceptibility for cancer. Hepatitis B [1] and inflammatory bowel disease [2] are common examples for this correlation, leading to hepatocellular carcinoma and colorectal cancer, respectively. Neutrophils, as a key component in inflammation, may play a crucial role in inflammation driven tumorigenesis. This was well exemplified when neutrophils were shown to directly promote carcinogenesis in a mouse model of colitis [3]. Indeed, neutrophils at the primary tumor site were shown to provide a wide range of different tumor promoting functions. Neutrophils were shown to support angiogenesis via secretion of proangiogenic factors as well as the proteolytic activation of proangiogenic factors. Neutrophils were also implicated in promoting tumor growth through the proteolytic release of EGF, TGF $\beta$ , and PDGF from the extracellular matrix (ECM). Neutrophils express high levels of metalloproteinases which can also modify the ECM to allow tumor cell dissemination thereby promoting tumor spread. Furthermore, neutrophils were shown to

recruit other tumor promoting cells to the tumor bed. Finally, immature neutrophils, also termed G-MDSC (granulocytic myeloid derived suppressor cells), were implicated in the establishment of an immunosuppressive tumor microenvironment thereby limiting antitumor immunity. On the other hand, neutrophils were shown to have antitumor properties including the capacity to kill tumor cells either through direct cytotoxicity or via antibody dependent cell cytotoxicity (ADCC) [4]. Similar conflicting reports were made as to the role neutrophils play in the premetastatic niche. Neutrophils accumulate in large numbers in premetastatic organs [5–7]. The fact that bone marrow derived cells were implicated in priming of the premetastatic niche prompted the hypothesis that neutrophils may be the cells that mediate this process. Indeed, neutrophils were shown to have a positive effect on tumor cell seeding in the premetastatic site [6]. In contrast, we and others have shown that neutrophils actively limit metastatic seeding by killing tumor cells [5, 7].

Interestingly, while neutrophils play a role in modulating tumor cell seeding in the metastatic site, it seems like they do not affect the growth rate of the metastatic nodules [5, 7]. This suggested that neutrophil antitumor functions are not always manifested inside the tumor and may depend on

the chemokine landscape in the tumor microenvironment. This notion was further supported by findings showing that upon entering the tumor microenvironment neutrophils acquire a different set of traits. This was referred to as “polarization” of neutrophils toward a tumor promoting or an anti-tumor phenotype which is mediated via cytokines available in the tumor microenvironment (i.e.,  $TGF\beta$  and IFNs, resp.). Furthermore, recent studies suggested that neutrophils are not a homogeneous population of cells and may consist of both pro- and antitumor subpopulations [8]. Together, the observations made thus far suggest that the mere accumulation of neutrophils in the tumor site may not necessarily be indicative of their contribution or of their prognostic value. Along these lines, the ongoing efforts to correlate neutrophil counts, or the ratio between neutrophils and other immune cells, with patient prognosis and ultimate outcome are conflicting and show that neutrophil abundance may correlate with a better prognosis in some studies and with a worse prognosis in others [9].

## 2. Molecular Mechanisms of Neutrophil Polarization in the Tumor Microenvironment

Neutrophils were shown to have diverse functions in the tumor microenvironment including both promoting and inhibiting tumor growth. As neutrophils are quick to respond to environmental cues, the most plausible explanation for the different neutrophil phenotypes was that neutrophil function is dictated by the local chemokine milieu. Advances in our understanding of how neutrophil function is regulated in cancer have led to the realization that neutrophils may be directed towards a specific phenotype, be it tumor promoting or tumor limiting, upon entering the tumor. Here we will discuss how interferons and  $TGF\beta$  polarize neutrophils in the tumor microenvironment.

**2.1. Interferons.** Type I interferons (IFNs) were first characterized in the process of viral interference. However, since then IFNs were found to be involved in a wide range of biological processes. In the context of cancer, IFNs show strong antitumor function as they inhibit tumor cell proliferation and promote apoptosis [10]. However, IFNs were also found to play a key role in mounting an antitumor immune response through the activation of T-cells, NK cells, and macrophages [11]. In recent years it has become apparent that IFNs also affect neutrophil function and promote antitumor processes mediated by neutrophils. Jablonska et al. have shown that  $IFN-\beta$  is critical for suppressing the expression of proangiogenic factors, such as VEGF and MMP9, in tumor infiltrating neutrophils leading to enhanced tumor vascularization and growth in  $IFN-\beta$  deficient animals [12]. Furthermore,  $IFN-\beta$  was found to play a significant role in regulating the recruitment of neutrophils and their longevity in the primary tumor [13, 14]. Finally, type I IFN activity was found to inhibit neutrophil-mediated formation of “fertile” premetastatic niche [15].

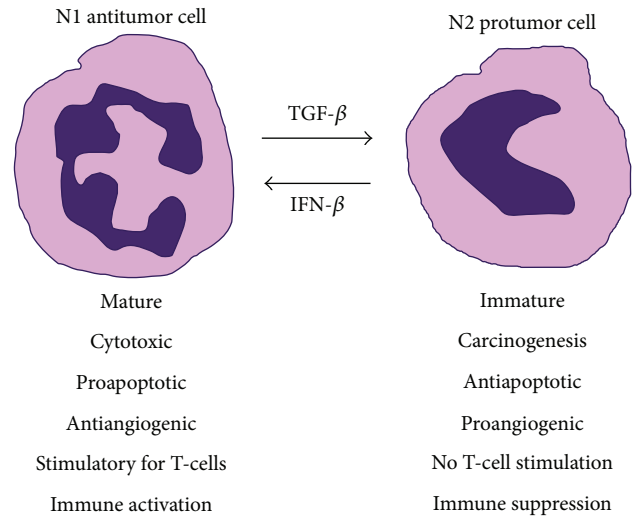


FIGURE 1: Neutrophil function in cancer is dictated by environmental cues. Neutrophils may be divided into N1 antitumor and N2 protumor cells.  $TGF\beta$  is a potent driver of the transition from N1 to N2 phenotype whereas  $IFN-\beta$  is a potent driver of the transition in the opposite direction. This exemplifies the notion that neutrophil function in cancer is determined by the chemokine milieu in the microenvironment.

**2.2.  $TGF\beta$ .**  $TGF\beta$  is a multipotent molecule known to have diverse effects in cancer. One of the most explored functions of  $TGF\beta$  in cancer is its role in generating an immunosuppressive tumor microenvironment. A groundbreaking study by Fridlender and colleagues [16] demonstrated that  $TGF\beta$  plays a critical role in suppression of antitumor neutrophil cytotoxicity. In this study, the authors showed that blocking  $TGF\beta$  signaling leads to a change in the cellular composition of the tumor and allows the influx of large numbers of neutrophils. More importantly, they showed that tumor-associated neutrophils (TANs) recruited in the absence of  $TGF\beta$  signaling have an antitumor N1 phenotype. The authors concluded that  $TGF\beta$  in the tumor microenvironment is involved in polarizing TAN towards N2 protumor phenotype. This concept was supported by other studies showing that  $TGF\beta$  can directly block antitumor neutrophil cytotoxicity [5] and that  $TGF\beta$  receptor deficient myeloid cells, including neutrophils, maintain an antitumor phenotype and limit tumor growth [17].

The conflicting effects of  $TGF\beta$  and IFNs on neutrophil function in the context of cancer are an example of how neutrophils respond to cues in the microenvironment (Figure 1). While understanding the mechanisms that regulate neutrophil function is clearly important from a therapeutic point of view, the realization that neutrophils may play conflicting roles, depending on their context, is an important notion.

### 3. Antitumor N1 Phenotype

Antitumor N1 neutrophils act to limit tumor growth and metastatic progression. This is accomplished via distinct mechanisms including direct and antibody dependent cytotoxicity as well as through the activation of other cell types including T-cells and dendritic cells.

**3.1. Direct Cytotoxicity.** Direct cytotoxicity of neutrophils towards tumor cells is not a novel concept and was first observed in the early 1970s [18]. Neutrophils are highly motile phagocytic cells whose primary function is antimicrobial protection of the host. Accordingly, neutrophils generate a variety of antimicrobial molecules. However, most of these molecules are harmless to eukaryotic cells. Still, the reactive molecules generated by the NADPH oxidase complex, superoxides,  $H_2O_2$ , and HOCl. Indeed, these molecules were found to be directly involved in antitumor neutrophil cytotoxicity [19–21]. Several studies have shown that physical contact is required for neutrophil cytotoxicity. However, stimulating cultured neutrophils with a potent agonist, such as PMA, leads to the generation and secretion of very high levels of  $H_2O_2$  alleviating the need for physical contact [5].

**3.2. ADCC.** Antibody dependent cell-mediated cytotoxicity (ADCC) is another mechanism for neutrophil antitumor cytotoxicity. Tumor cell-specific antibodies may be successfully used as an anticancer therapy. Antibody labeled cells are susceptible to destruction by immune cells expressing Fc receptors (FcR). Neutrophils express several FcRs that can mediate ADCC including FcγRI (CD64), FcγRIIa (CD32), FcγRIIIa (CD16a), and FcγRIIIb (CD16b) [22–24]. Indeed, neutrophils were shown to take part in ADCC in several types of cancer including glioma, squamous cell, and ovarian carcinoma. Neutrophils were also shown to contribute to the antitumor ADCC in Non-Hodgkin's Lymphoma [25], in breast cancer using [26], and in B-cell lymphoma [27].

**3.3. Stimulation of T-Cells and DCs.** Neutrophils, on top of having a role in killing tumor cells directly, can also stimulate adaptive antitumor immune responses. This was well exemplified by experiments showing that neutrophils are required for proper antitumor CD8<sup>+</sup> T-cell immune response [16, 28–30]. Stimulation of adaptive antitumor immunity by neutrophils has two arms, the recruitment of other immune cells and their antigen presenting abilities.

*(a) Recruitment of Immune Cells.* Neutrophils secrete several cytokines including TNFα, Cathepsin G, and neutrophil elastase which have a direct effect on T-cells and promote their proliferation and cytokine production. Neutrophils, under these conditions, act to recruit and activate T-cells and enhance the overall adaptive immune antitumor response. Specifically, TAN were shown to stimulate T-cell proliferation and IFNγ secretion in early stage lung cancer patients [31].

*(b) Neutrophil Extracellular Traps (NETs).* Production of extracellular traps by neutrophils is an interesting feature in neutrophil biology. These NETs are composed of chromatin

fibers decorated with histones and other proteins and are considered as an additional tool in neutrophils' arsenal of antimicrobial properties. However, Tillack and colleagues showed that NETs may also be utilized to prime T-cells [32]. This was also linked to a possible role of NETs in immunoeediting in cancer and the propagation of antitumor immune responses [33].

*(c) Antigen Presentation.* For a long time antigen presentation was thought to be exclusively mediated by macrophages and more so by dendritic cells (DCs). However, in 2007 Beauvillain and colleagues demonstrated that neutrophils can efficiently process and present antigens to directly stimulate T-cell immune responses [34]. While this does not directly link neutrophil presentation of antigens to antitumor cytotoxicity, Fridlender and colleagues showed in 2009 that N1 TANs require T-cells for their antitumor activity in the primary tumor [16], an observation that may be explained by neutrophils' ability to present tumor antigens to stimulate T-cells.

### 4. Protumor N2 Phenotype

Neutrophils have been traditionally considered as guards of the host immune system. However, in the context of tumor, the function of these cells is frequently modified to act against the host and promote tumor growth and metastasis formation. A possible reason for this could be tumor-secreted factors that elicit wound-repair responses by neutrophils that in turn inadvertently stimulate tumor progression [35]. Moreover, wound-infiltrating neutrophils are rapidly diverted from a wound to preneoplastic cells and such interactions lead to increased proliferation of the preneoplastic cells. Prostaglandin E2 (PGE2) seems to be the factor responsible for this process [36]. These results have shown that repeated wounding with subsequent inflammation leads to a greater incidence of local melanoma formation. Along these lines, several studies have shown that infiltration of tumors by neutrophils is associated with poor clinical outcome. Tumor-associated neutrophils (TANs) have been shown to promote tumor growth and progression via a variety of mechanisms, including extracellular matrix remodeling, promotion of tumor cell invasion and metastasis, angiogenesis, lymphangiogenesis, and immune suppression [12, 13, 15].

**4.1. Protumor Cytokines.** One of the mechanisms responsible for neutrophil-mediated tumor angiogenesis, growth, and metastasis is the secretion of protumor cytokines by these cells [37]. Depending on the cytokine milieu, neutrophils are able to secrete multiple growth factors such as EGF, TGFβ, PDGF, HGF, VEGF, and oncostatin M [12, 38–41].

Evidence suggests that EGF and its receptor EGFR are involved in the pathogenesis and progression of different carcinoma types [42]. Amplification of the EGFR gene and mutations of the EGFR tyrosine kinase domain have been recently demonstrated to occur in carcinoma patients. EGFR causes neoangiogenesis but also increased proliferation, decreased apoptosis, and enhanced tumor cell motility [43] since its receptor (EGFR; HER1; erbB1) is highly expressed on

variety of human tumors including non-small cell lung cancer (NSCLC) and breast, head and neck, gastric, colorectal, esophageal, prostate, bladder, renal, pancreatic, and ovarian cancers [42, 44].

TGF $\beta$  is frequently upregulated in human cancers [45] and has been linked to the regulation of tumor initiation, progression, and metastasis [46]. Tumor-secreted TGF $\beta$  is usually sequestered to the extracellular matrix as an inactive complex and becomes activated through enzymes such as neutrophil-derived elastase and MMP9 [46]. Furthermore, reactive oxygen free radicals produced by activated neutrophils can activate latent TGF $\beta$  [47]. Thus, activated neutrophils, through production of elastase, MMP9, and ROS, may contribute to TGF $\beta$ -mediated immunosuppression [9]. Furthermore, TGF $\beta$  has been shown to be a potent chemoattractant for neutrophils facilitating their recruitment to sites of inflammation [48, 49] and to promote their protumor N2 phenotype, as mentioned above [16].

Another important neutrophil-derived growth factor is platelet-derived growth factor (PDGF). Interestingly, this growth factor was shown to be chemotactic for monocytes and neutrophils [50]. It was recently established that PDGF stimulation cooperates with genetic changes caused by retroviral insertions in induction of fully malignant tumor phenotype [51]. Moreover, the autocrine PDGF signaling seems to play a role in the growth and metastasis of epithelial cancers.

VEGF is a very potent proangiogenic factor but also serves as a potent chemoattractant for neutrophils. It has been implicated as the key endothelial cell-specific factor required for pathological angiogenesis, including tumor neovascularization. Inhibition of the VEGF signaling blocks angiogenesis in growing tumors, leading to regression of tumor growth [52]. The function of vascular endothelial growth factor (VEGF) in cancer is not limited to angiogenesis and vascular permeability [53]. VEGF-mediated signaling occurs in tumor cells, and this signaling contributes to key aspects of tumorigenesis, including the function of cancer stem cells and tumor initiation [54]. Autocrine VEGF signaling can promote the growth, survival, migration, and invasion of cancer cells [55–57].

Oncostatin M is another pleiotropic cytokine that is secreted by neutrophils [58]. It has been shown to exert proinflammatory effects by inducing adhesion and chemotaxis of neutrophils and chemokine production by endothelial cells [59]. Although oncostatin M was originally identified as an inhibitor of tumor cell growth *in vitro* [60, 61], it is increasingly apparent that this cytokine plays a role in breast cancer cell detachment [62] and angiogenesis [41].

In addition to growth factors, neutrophils are able to secrete other cytokines that influence tumor development and spreading. For instance, neutrophil delivered TNF $\alpha$ , IL-6, and IL-17 were shown to promote tumor growth by modifying the function of stromal cells surrounding the tumor [63, 64]. TNF $\alpha$  produced by tumor cells or inflammatory cells in the tumor microenvironment can promote tumor cell survival through the induction of NF $\kappa$ B-dependent antiapoptotic molecules [65]. TNF $\alpha$  was also shown to promote angiogenesis [66] and induce the expression of VEGF and

HIF-1 $\alpha$  in tumor cells [67]. IL-6 promotes angiogenesis and the expression of VEGF [68] through JAK2/STAT3 signaling [64] and the tumor promoting effects of IL-17 are in part mediated through upregulation of IL-6 [63, 64].

**4.2. Angiogenesis and Modulation of the ECM.** Angiogenesis is one of the hallmarks of the development of malignant neoplasias. Primary tumors of a certain size require the growth of new blood vessels in order to be supplied with nutrients and oxygen. Accordingly, at a size of 1–2 mm<sup>3</sup>, tumors alter their angiogenic phenotype and support continuous proliferation of endothelial cells. This “angiogenic switch” is activated by disturbed balance between endogenous pro- and antiangiogenic factors. It leads to the uncontrolled growth of blood vessels, mainly via stimulation of VEGF. Importantly, experimental *in vivo* models of angiogenesis have demonstrated that neutrophils affect neovascularization in the tissues [69]. Accordingly, Gr-1-mediated neutrophil depletion was found to significantly reduce tumor angiogenesis [70, 71]. Notably, in patients with myxofibrosarcoma, elevated numbers of neutrophils were observed in high-grade malignant tumors and this correlated positively with increased intratumoral microvessel density [72]. The mechanism by which tumor-associated neutrophils modulate tumor angiogenesis has not been fully elucidated. Activated neutrophils can release a variety of proteases that can degrade and remodel the ECM, a process that is crucial for angiogenesis. These cells have recently been shown to express high amounts of VEGF and MMP9 that is known to be responsible for initiation of the angiogenic switch and to support vessel growth in tumors [12]. MMP9 has been shown to have the most profound effects in mediating tumor angiogenesis [73]. Proteolysis of the ECM by this MMP releases such potent angiogenic factors such as vascular endothelial growth factor (VEGF) and FGF2 that are usually sequestered in an inactivated form to the ECM [74, 75]. MMP9 is also involved in the regulation of leukocytosis, for example, by potentiating proangiogenic and neutrophil attracting IL-8 expression [76] and by the release of hematopoietic progenitor cells from the bone marrow [77]. Huang et al. could show that MMP9-deficient mice display significantly reduced tumor microvessel density, compared with wild-type mice [78]. Neutrophil-derived MMP9 has also been shown to contribute to tumor angiogenesis and progression of squamous cell carcinoma [74]. Finally, Bv8, a potent proangiogenic factor, was shown to be upregulated in neutrophils in the context of cancer and to directly contribute to tumor angiogenesis and progression [79, 80].

**4.3. Tumor Cell Dissemination.** Metastasis is a highly complex process requiring tumor cell detachment from the primary tumor and migration to secondary target organs through the lymphatic or blood circulatory systems [81]. Neutrophils can exhibit both pro- and antimetastatic properties under certain conditions [82–85]. In prometastatic state neutrophils secrete soluble factors, including proteases and cytokines, that activate endothelium and parenchymal cells, leading to improvement of adhesion of circulating tumor cells in distant sites [74, 83, 86] and enhanced metastasis formation. Moreover, contact-dependent mechanisms, whereby

neutrophils act as a bridge, tethering circulating tumor cells (CTCs) to target organ endothelium, have been described [87]. Such interaction is mediated by the binding of  $\beta 2$  integrins on neutrophils to ICAM-1 on tumor cells and was described for lung and liver metastasis model [84, 88]. In studies by Spicer et al. neutrophils promote cancer cell adhesion within liver sinusoids and their depletion before cancer cell inoculation resulted in decreased number of metastases in an intrasplenic model of liver metastasis [84]. Another interesting study showed that neutrophils can support lung metastasis development through physical interaction and anchoring of circulating tumor cells to endothelium [89]. It is not clear if this process supports tumor cell extravasation into target organ or neutrophils hold melanoma cells in the capillaries until they grow into a secondary tumor [89].

In addition to the mechanisms proposed thus far, novel aspects of neutrophil biology recently got attention as possible mechanism that contributes to cancer progression and metastasis. Recent studies suggest that NETs are able to trap tumor cells and depending on neutrophil activation such sequestered tumor cells can be destroyed by ROS that results in inhibition of metastasis formation [82] or be kept in place thus supporting early adhesion of tumor cells to distant organ sites and metastatic processes [90].

In the recent work of Wu et al. an inhibitory role of endogenous type I IFNs on neutrophil-mediated metastasis formation could be shown. The lack of endogenous type I IFNs drives neutrophils to prometastatic phenotype at least in two ways, supporting neutrophil migration and the formation of the premetastatic niche in the lung and inhibiting neutrophil cytotoxicity against tumor cells in circulation.

**4.4. Formation of the Premetastatic Niche.** Tumor induced changes in the microenvironment of distal organs make tissues more receptive to colonization of migrating tumor cells [91, 92]. Consequently, bone marrow derived cells, including neutrophils, are mobilized and accumulate in the future site of metastasis [93] where they participate in the formation of supportive metastatic microenvironment termed “premetastatic niche” [94–96]. These cells are recruited by Bv8, MMP9, S100A8, and S100A9 [6, 97] and this process seems to be strongly dependent on granulocyte colony-stimulating factor (G-CSF) [6].

Recent studies have shown that neutrophils make up the main cell population involved in formation of premetastatic niche [82]. This process seems to be enhanced by the absence of type I interferons that results in upregulation of CXCR2 expression on neutrophils from these mice. Moreover, prometastatic molecules like S100A8, S100A9, Bv8, and MMP9 are upregulated in lungs of *Ifnar1*<sup>-/-</sup> mice. Both S100A8 and S100A9 are known to influence tumor cell proliferation, survival, and migration [97, 98] but also to stimulate migration and proliferation of neutrophils. Bv8, next to induction of tumor cell extravasation [6], increases neutrophil accumulation within premetastatic tissue. MMP9 is responsible for formation of leaky vasculature in the premetastatic lung [99] and support of tumor cells survival in this organ [100].

**4.5. Recruitment of Other Cells and Immune Evasion.** The immune regulatory functions of neutrophils are recently getting growing attention. Interactions between neutrophils and other immune cells obviously are regulating many inflammatory processes, including tumorigenesis. There is evidence that activated neutrophils can interact with T-cells in dichotomous ways. Several studies have shown that neutrophils can present antigens and provide accessory signals for T-cell activation [101–103]. Other studies have suggested that neutrophils can suppress antigen-nonspecific T-cell proliferation [104, 105]. The suppressive function of granulocytic cells in cancer patients has generally been attributed to a circulating low-density granulocytic myeloid derived suppressor cell (G-MDSC) population [60–62]. However, there is some uncertainty about whether G-MDSCs do exist in humans. In mice this heterogeneous group of cells consists mainly of immature neutrophils and monocytes.

Neutrophil-mediated T-cell suppression requires arginase 1 or ROS [105–107]. In humans with metastatic cancer disease, arginase 1-mediated suppression of lymphocytes was reported [108, 109]. Lately, mature blood neutrophil subset was shown to suppress T-cell activation in cancer [8] and during severe inflammation [104]. This suppression requires release of H<sub>2</sub>O<sub>2</sub> into the immunological synapse in a Mac-1 (CD11b/CD18) dependent manner.

Very recent studies show that neutrophils cooperate with  $\gamma\delta$  T-cells in promotion of breast cancer metastasis [110]. Neutrophil depletion in the highly aggressive metastatic breast cancer mouse model KEP results in significant reduction of both spontaneous pulmonary and lymph node metastasis [110]. Moreover, combined depletion of both neutrophils and CD8<sup>+</sup> cells results in inhibition of metastasis formation, implicating cooperation of these cells during this process.

## 5. Recruitment of Neutrophils into Tumor and Premetastatic Sites

Neutrophils make up substantial population of cells infiltrating tumors and premetastatic niche, in mice and human [12, 15, 111]. Many cell subtypes, including tumor cells, produce chemokines that attract neutrophils, for example, CXCL1 or CXCL2.

**5.1. Factors That Mediate Neutrophil Recruitment.** The migration of neutrophils into solid tumors depends on chemotactic factors. There are several chemotactic factors that may stimulate the migration of neutrophils, but the most potent are members of the CXCL chemokine family. Human CXCL8 (IL-8) is one of the best studied neutrophil chemoattractants with respect to tumor biology and is overexpressed in different human carcinomas and tumor cell lines including breast, colon, cervical, lung, brain, prostate, ovarian, and renal cell carcinomas, acute myelogenous and B-cell lymphocytic leukemia, melanoma, and Hodgkin's disease [112]. Importantly, both stromal and tumor cells can produce CXCL8. Other human chemokines such as CCL3 (MIP-1 $\alpha$ ) and CXCL6 (huGCP-2) or murine chemokines CXCL1, CXCL2, and CXCL5 are potent chemoattractants and activators for neutrophils [12] and are produced by many tumors [113–116].

Recent study on hepatocellular carcinoma indicated importance of CXCL16 and its receptor CXCR6 in neutrophil recruitment and tumor progression, due to its ability to stimulate tumor cells to release CXCL8. Another recent study shows that human metastatic melanoma cells entrapped in the lungs secrete IL-8 to attract neutrophils, which promotes tumor cell tethering to the vascular endothelium. Prolonged cell retention in the lungs facilitated transendothelial migration and metastasis development [89]. Experiments have shown that inhibition of neutrophil migration, for example, by blocking of chemokine receptor CXCR2 or CXCR2<sup>-/-</sup> in mice, leads to reduced tumor angiogenesis and growth in B16F10 melanoma [14, 117] and Lewis lung carcinoma model [107]. Inhibited myeloid cell infiltration due to the loss of CXCR2 was also shown to be responsible for significantly suppressed chronic colonic inflammation and colitis-associated tumorigenesis [118].

A number of additional mediators might serve as chemoattractants for neutrophil recruitment to the tumor tissue. It has been shown that bioactive lipids, such as sphingosine-1-phosphate (S1P), could promote neutrophil activation and chemotaxis [119, 120]. Similarly, the hypoxia-inducible factor 1- $\alpha$  and its downstream products like CXCL12, VEGF, or MMP9 are involved in recruitment and retention of neutrophils in angiogenic environments [12, 121]. VEGF, in addition to its proangiogenic role during tumor growth, is also capable of inducing neutrophil adhesion to postcapillary venules followed by homing to tissues of its high expression, for example, tumor or premetastatic niche [122, 123].

Recent studies suggest that the myeloid-related proteins (MRPs) are also involved in neutrophil migration. The MRPs S100A8 and S100A9 are strongly expressed by tumors and in the premetastatic niche and act as strong chemoattractants for neutrophils into these sites [82, 97, 124]. However, the exact mechanism of MRPs mediated neutrophil mobilization is not clear and still needs to be investigated.

**5.2. Survival of Neutrophils in Tumor Microenvironment.** Due to their proinflammatory functions and potential toxicity against host tissue, the neutrophil life span is strictly regulated [125]. In the absence of inflammatory stimuli, neutrophils are removed from circulation shortly after their mobilization from the bone marrow, mainly by apoptosis. Importantly, several proinflammatory cytokines have been shown to influence the longevity of neutrophils [126]. Recent observations of Andzinski et al. [13] show that the life span of tumor-associated neutrophils is remarkably prolonged in tumor-bearing IFN- $\beta$  deficient (Ifnb1<sup>-/-</sup>) mice, compared to wild-type controls. This is apparently due to the fact that IFN- $\beta$  is able to influence both the extrinsic and the intrinsic apoptosis pathways of neutrophilic granulocytes. Lower expression of Fas, reactive oxygen species, active Caspases 3 and 9, as well as a change in expression pattern of proapoptotic and antiapoptotic members of the Bcl-2 family and the major apoptosome constituent Apaf-1, is observed under such conditions. The death receptor Fas on neutrophils has been shown to be involved in spontaneous extrinsic cell death signaling [127]. Fas has been shown to play a role in type I

IFN-induced apoptosis in several types of neoplasias such as melanoma, multiple myeloma, and chronic myeloid leukemia cells [128, 129].

ROS production by neutrophils might also play an important role in regulation of life span of neutrophils. For example, a delayed spontaneous apoptosis was shown in patients deficient for NADPH oxidase [130, 131]. It has also been shown that hypoxia or pharmacological inhibition of NADPH oxidase and hydrogen peroxide scavengers decreases the rate of neutrophil apoptosis [132]. Recent data indicate that spontaneous production of ROS is diminished in the absence of endogenous IFN- $\beta$ , potentially contributing to the delayed apoptosis of tumor infiltrating neutrophils in Ifnb1<sup>-/-</sup> mice [13]. G-CSF is one of the major survival factors of neutrophilic granulocytes and has been reported to reduce Bax expression and redistribution [133] and restore its phosphorylation status thus leading to its inactivation. This mechanism is responsible for G-CSF-mediated repression of Caspase activation [134]. Regulation of G-CSF expression is responsible for altered neutrophils survival in tumors.

## 6. Concluding Remarks

Neutrophil function in cancer has long been a matter of debate as these cells were shown to possess a range of tumor promoting as well as tumor limiting properties. We propose that these conflicting observations stem from the fact that neutrophils are not a homogeneous population of cells. Neutrophil heterogeneity stems from two facts that are not mutually exclusive and have to do with the changes in the chemokine milieu in the context of cancer: The first is the fact that neutrophils are highly responsive to cues in their microenvironment and may adopt a protumor phenotype in certain conditions and an antitumor phenotype in others. The second is the fact that there are distinct neutrophil subsets which differ in their contribution in the context of cancer. Together, these observations support the notion that neutrophil function in cancer may be dictated in a context dependent fashion (Figure 1). These observations also identify potential elements which may be therapeutically targeted to enhance antitumor neutrophil activity while limiting their protumor properties.

## Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

## References

- [1] Y. Nakamoto, L. G. Guidotti, C. V. Kuhlen, P. Fowler, and F. V. Chisari, "Immune pathogenesis of hepatocellular carcinoma," *Journal of Experimental Medicine*, vol. 188, no. 2, pp. 341–350, 1998.
- [2] G. Rogler, "Chronic ulcerative colitis and colorectal cancer," *Cancer Letters*, vol. 345, no. 2, pp. 235–241, 2014.

- [3] K. Shang, Y.-P. Bai, C. Wang et al., "Crucial involvement of tumor-associated neutrophils in the regulation of chronic colitis-associated carcinogenesis in mice," *PLoS ONE*, vol. 7, no. 12, Article ID e51848, 2012.
- [4] M. van Egmond and J. E. Bakema, "Neutrophils as effector cells for antibody-based immunotherapy of cancer," *Seminars in Cancer Biology*, vol. 23, no. 3, pp. 190–199, 2013.
- [5] Z. Granot, E. Henke, E. A. Comen, T. A. King, L. Norton, and R. Benezra, "Tumor entrained neutrophils inhibit seeding in the premetastatic lung," *Cancer Cell*, vol. 20, no. 3, pp. 300–314, 2011.
- [6] M. Kowanzet, X. Wu, J. Lee et al., "Granulocyte-colony stimulating factor promotes lung metastasis through mobilization of Ly6G+Ly6C+ granulocytes," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 50, pp. 21248–21255, 2010.
- [7] M. A. López-Lago, S. Posner, V. J. Thodima, A. M. Molina, R. J. Motzer, and R. S. K. Chaganti, "Neutrophil chemokines secreted by tumor cells mount a lung antimetastatic response during renal cell carcinoma progression," *Oncogene*, vol. 32, no. 14, pp. 1752–1760, 2013.
- [8] J. Y. Sagiv, J. Michaeli, S. Assi et al., "Phenotypic diversity and plasticity in circulating neutrophil subpopulations in cancer," *Cell Reports*, vol. 10, no. 4, pp. 562–573, 2015.
- [9] R. V. Sionov, Z. G. Fridlender, and Z. Granot, "The multifaceted roles neutrophils play in the tumor microenvironment," *Cancer Microenvironment*, pp. 1–34, 2014.
- [10] D. V. Kalvakolanu, "Interferons and cell growth control," *Histology and Histopathology*, vol. 15, no. 2, pp. 523–537, 2000.
- [11] M. M. Brierley and E. N. Fish, "Review: IFN- $\alpha/\beta$  receptor interactions to biologic outcomes: understanding the circuitry," *Journal of Interferon and Cytokine Research*, vol. 22, no. 8, pp. 835–845, 2002.
- [12] J. Jablonska, S. Leschner, K. Westphal, S. Lienenklaus, and S. Weiss, "Neutrophils responsive to endogenous IFN- $\beta$  regulate tumor angiogenesis and growth in a mouse tumor model," *Journal of Clinical Investigation*, vol. 120, no. 4, pp. 1151–1164, 2010.
- [13] L. Andzinski, C.-F. Wu, S. Lienenklaus, A. Kröger, S. Weiss, and J. Jablonska, "Delayed apoptosis of tumor associated neutrophils in the absence of endogenous IFN- $\beta$ ," *International Journal of Cancer*, vol. 136, no. 3, pp. 572–583, 2015.
- [14] J. Jablonska, C.-F. Wu, L. Andzinski, S. Leschner, and S. Weiss, "CXCR2-mediated tumor-associated neutrophil recruitment is regulated by IFN- $\beta$ ," *International Journal of Cancer*, vol. 134, no. 6, pp. 1346–1358, 2014.
- [15] C.-F. Wu, L. Andzinski, N. Kasnitz et al., "The lack of type I interferon induces neutrophil-mediated pre-metastatic niche formation in the mouse lung," *International Journal of Cancer*, vol. 137, no. 4, pp. 837–847, 2015.
- [16] Z. G. Fridlender, J. Sun, S. Kim et al., "Polarization of tumor-associated neutrophil phenotype by TGF- $\beta$ : 'N1' versus 'N2' TAN," *Cancer Cell*, vol. 16, no. 3, pp. 183–194, 2009.
- [17] S. V. Novitskiy, M. W. Pickup, A. Chytil, D. Polosukhina, P. Owens, and H. L. Moses, "Deletion of TGF- $\beta$  signaling in myeloid cells enhances their anti-tumorigenic properties," *Journal of Leukocyte Biology*, vol. 92, no. 3, pp. 641–651, 2012.
- [18] J. Bubeník, P. Perlmann, K. Helmstein, and G. Moberger, "Cellular and humoral immune responses to human urinary bladder carcinomas," *International Journal of Cancer*, vol. 5, no. 3, pp. 310–319, 1970.
- [19] J. Dissemmond, T. K. Weimann, L. A. Schneider et al., "Activated neutrophils exert antitumor activity against human melanoma cells: reactive oxygen species-induced mechanisms and their modulation by granulocyte-macrophage-colony-stimulating factor," *Journal of Investigative Dermatology*, vol. 121, no. 4, pp. 936–938, 2003.
- [20] M. Zivkovic, M. Poljak-Blazi, G. Egger, S. B. Sunjic, R. J. Schaur, and N. Zarkovic, "Oxidative burst and anticancer activities of rat neutrophils," *BioFactors*, vol. 24, no. 1–4, pp. 305–312, 2005.
- [21] R. A. Clark and S. J. Klebanoff, "Neutrophil-mediated tumor cell cytotoxicity: role of the peroxidase system," *Journal of Experimental Medicine*, vol. 141, no. 6, pp. 1442–1447, 1975.
- [22] B. H. Kushner and N.-K. V. Cheung, "Absolute requirement of CD11/CD18 adhesion molecules, FcRII and the phosphatidylinositol-linked FcRIII for monoclonal antibody-mediated neutrophil antihuman tumor cytotoxicity," *Blood*, vol. 79, no. 6, pp. 1484–1490, 1992.
- [23] D. Iliopoulos, C. Ernst, Z. Steplewski et al., "Inhibition of metastases of a human melanoma xenograft by monoclonal antibody to the GD2/GD3 gangliosides," *Journal of the National Cancer Institute*, vol. 81, no. 6, pp. 440–444, 1989.
- [24] T. Valerius, R. Repp, T. P. M. de Wit et al., "Involvement of the high-affinity receptor for IgG (Fc gamma RI; CD64) in enhanced tumor cell cytotoxicity of neutrophils during granulocyte colony-stimulating factor therapy," *Blood*, vol. 82, no. 3, pp. 931–939, 1993.
- [25] F. J. Hernandez-Ilizaliturri, V. Jupudy, J. Ostberg et al., "Neutrophils contribute to the biological antitumor activity of rituximab in a non-Hodgkin's lymphoma severe combined immunodeficiency mouse model," *Clinical Cancer Research*, vol. 9, no. 16, part 1, pp. 5866–5873, 2003.
- [26] P. Hubert, A. Heitzmann, S. Viel et al., "Antibody-dependent cell cytotoxicity synapses form in mice during tumor-specific antibody immunotherapy," *Cancer Research*, vol. 71, no. 15, pp. 5134–5143, 2011.
- [27] Y. Guettinger, K. Barbin, M. Peipp et al., "A recombinant bispecific single-chain fragment variable specific for HLA class II and Fc $\alpha$ RI (CD89) recruits polymorphonuclear neutrophils for efficient lysis of malignant B lymphoid cells," *Journal of Immunology*, vol. 184, no. 3, pp. 1210–1217, 2010.
- [28] F. Fioretti, D. Fradelizi, A. Stoppacciaro et al., "Reduced tumorigenicity and augmented leukocyte infiltration after monocyte chemotactic protein-3 (MCP-3) gene transfer: perivascular accumulation of dendritic cells in peritumoral tissue and neutrophil recruitment within the tumor," *Journal of Immunology*, vol. 161, no. 1, pp. 342–346, 1998.
- [29] A. Stoppacciaro, C. Melani, M. Parenza et al., "Regression of an established tumor genetically modified to release granulocyte colony-stimulating factor requires granulocyte-T cell cooperation and T cell-produced interferon  $\gamma$ ," *Journal of Experimental Medicine*, vol. 178, no. 1, pp. 151–161, 1993.
- [30] F. Cavallo, M. Giovarelli, A. Gulino et al., "Role of neutrophils and CD4+ T lymphocytes in the primary and memory response to nonimmunogenic murine mammary adenocarcinoma made immunogenic by IL-2 gene," *The Journal of Immunology*, vol. 149, no. 11, pp. 3627–3635, 1992.
- [31] E. B. Eruslanov, P. S. Bhojnagarwala, J. G. Quatromoni et al., "Tumor-associated neutrophils stimulate T cell responses in early-stage human lung cancer," *Journal of Clinical Investigation*, vol. 124, no. 12, pp. 5466–5480, 2014.

- [32] K. Tillack, P. Breiden, R. Martin, and M. Sospedra, "T lymphocyte priming by neutrophil extracellular traps links innate and adaptive immune responses," *Journal of Immunology*, vol. 188, no. 7, pp. 3150–3159, 2012.
- [33] S. Berger-Achituv, V. Brinkmann, U. A. Abed et al., "A proposed role for neutrophil extracellular traps in cancer immunoeediting," *Frontiers in Immunology*, vol. 4, article 48, 2013.
- [34] C. Beauvillain, Y. Delneste, M. Scotet et al., "Neutrophils efficiently cross-prime naive T cells in vivo," *Blood*, vol. 110, no. 8, pp. 2965–2973, 2007.
- [35] G. F. Whalen, "Solid tumours and wounds: transformed cells misunderstood as injured tissue?" *The Lancet*, vol. 336, no. 8729, pp. 1489–1492, 1990.
- [36] N. Antonio, M. L. Bonnelykke-Behrntz, L. C. Ward et al., "The wound inflammatory response exacerbates growth of pre-neoplastic cells and progression to cancer," *The EMBO Journal*, vol. 34, no. 17, pp. 2219–2236, 2015.
- [37] C. Tecchio, P. Scapini, G. Pizzolo, and M. A. Cassatella, "On the cytokines produced by human neutrophils in tumors," *Seminars in Cancer Biology*, vol. 23, no. 3, pp. 159–170, 2013.
- [38] M. A. Cassatella, "Neutrophil-derived proteins: selling cytokines by the pound," *Advances in Immunology*, vol. 73, pp. 369–509, 1999.
- [39] G. R. Grotendorst, G. Smale, and D. Pancev, "Production of transforming growth factor beta by human peripheral blood monocytes and neutrophils," *Journal of Cellular Physiology*, vol. 140, no. 2, pp. 396–402, 1989.
- [40] M. McCourt, J. H. Wang, S. Sookhai, and H. P. Redmond, "Activated human neutrophils release hepatocyte growth factor/scatter factor," *European Journal of Surgical Oncology*, vol. 27, no. 4, pp. 396–403, 2001.
- [41] M. M. Queen, R. E. Ryan, R. G. Holzer, C. R. Keller-Peck, and C. L. Jorcyk, "Breast cancer cells stimulate neutrophils to produce oncostatin M: potential implications for tumor progression," *Cancer Research*, vol. 65, no. 19, pp. 8896–8904, 2005.
- [42] N. Normanno, A. De Luca, C. Bianco et al., "Epidermal growth factor receptor (EGFR) signaling in cancer," *Gene*, vol. 366, no. 1, pp. 2–16, 2006.
- [43] C. L. Arteaga, "Epidermal growth factor receptor dependence in human tumors: more than just expression?" *Oncologist*, vol. 7, supplement 4, pp. 31–39, 2002.
- [44] D. S. Salomon, R. Brandt, F. Ciardiello, and N. Normanno, "Epidermal growth factor-related peptides and their receptors in human malignancies," *Critical Reviews in Oncology and Hematology*, vol. 19, no. 3, pp. 183–232, 1995.
- [45] S. Wojtowicz-Praga, "Reversal of tumor-induced immunosuppression by TGF- $\beta$  inhibitors," *Investigational New Drugs*, vol. 21, no. 1, pp. 21–32, 2003.
- [46] B. Bierie and H. L. Moses, "Transforming growth factor beta (TGF- $\beta$ ) and inflammation in cancer," *Cytokine and Growth Factor Reviews*, vol. 21, no. 1, pp. 49–59, 2010.
- [47] M. H. Barcellos-Hoff and T. A. Dix, "Redox-mediated activation of latent transforming growth factor- $\beta$ 1," *Molecular Endocrinology*, vol. 10, no. 9, pp. 1077–1083, 1996.
- [48] T. Parekh, B. Saxena, J. Reibman, B. N. Cronstein, and L. I. Gold, "Neutrophil chemotaxis in response to TGF-beta isoforms (TGF-beta 1, TGF-beta 2, TGF-beta 3) is mediated by fibronectin," *The Journal of Immunology*, vol. 152, no. 5, pp. 2456–2466, 1994.
- [49] J. Reibman, S. Meixler, T. C. Lee et al., "Transforming growth factor beta 1, a potent chemoattractant for human neutrophils, bypasses classic signal-transduction pathways," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 88, no. 15, pp. 6805–6809, 1991.
- [50] T. F. Deuel, R. M. Senior, J. S. Huang, and G. L. Griffin, "Chemotaxis of monocytes and neutrophils to platelet-derived growth factor," *The Journal of Clinical Investigation*, vol. 69, no. 4, pp. 1046–1049, 1982.
- [51] L. Uhrbom, G. Hesselager, M. Nistér, and B. Westermark, "Induction of brain tumors in mice using a recombinant platelet-derived growth factor B-chain retrovirus," *Cancer Research*, vol. 58, no. 23, pp. 5275–5279, 1998.
- [52] G. McMahon, "VEGF receptor signaling in tumor angiogenesis," *Oncologist*, vol. 5, supplement 1, pp. 3–10, 2000.
- [53] D. R. Senger, "Vascular endothelial growth factor: much more than an angiogenesis factor," *Molecular Biology of the Cell*, vol. 21, no. 3, pp. 377–379, 2010.
- [54] H. L. Goel and A. M. Mercurio, "VEGF targets the tumour cell," *Nature Reviews Cancer*, vol. 13, no. 12, pp. 871–882, 2013.
- [55] R. C. Bates, J. D. Goldsmith, R. E. Bachelder et al., "Flt-1-dependent survival characterizes the epithelial-mesenchymal transition of colonic organoids," *Current Biology*, vol. 13, no. 19, pp. 1721–1727, 2003.
- [56] R. E. Bachelder, A. Crago, J. Chung et al., "Vascular endothelial growth factor is an autocrine survival factor for neuropilin-expressing breast carcinoma cells," *Cancer Research*, vol. 61, no. 15, pp. 5736–5740, 2001.
- [57] M. J. Barr, D. J. Bouchier-Hayes, and J. H. Harmey, "Vascular endothelial growth factor is an autocrine survival factor for breast tumour cells under hypoxia," *International Journal of Oncology*, vol. 32, no. 1, pp. 41–48, 2008.
- [58] A. Grenier, M. Dehoux, A. Boutten et al., "Oncostatin M production and regulation by human polymorphonuclear neutrophils," *Blood*, vol. 93, no. 4, pp. 1413–1421, 1999.
- [59] S. M. Kerfoot, E. Raharjo, M. Ho et al., "Exclusive neutrophil recruitment with oncostatin M in a human system," *The American Journal of Pathology*, vol. 159, no. 4, pp. 1531–1539, 2001.
- [60] J. M. Zarling, M. Shoyab, H. Marquardt, M. B. Hanson, M. N. Lioubin, and G. J. Todaro, "Oncostatin M: a growth regulator produced by differentiated histiocytic lymphoma cells," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 83, no. 24, pp. 9739–9743, 1986.
- [61] J. Liu, N. Hadjokas, B. Mosley, Z. Estrov, M. J. Spence, and R. E. Vestal, "Oncostatin M-specific receptor expression and function in regulating cell proliferation of normal and malignant mammary epithelial cells," *Cytokine*, vol. 10, no. 4, pp. 295–302, 1998.
- [62] R. G. Holzer, R. E. Ryan, M. Tommack, E. Schlekeway, and C. L. Jorcyk, "Oncostatin M stimulates the detachment of a reservoir of invasive mammary carcinoma cells: role of cyclooxygenase-2," *Clinical and Experimental Metastasis*, vol. 21, no. 2, pp. 167–176, 2004.
- [63] R. J. Moore, D. M. Owens, G. Stamp et al., "Mice deficient in tumor necrosis factor- $\alpha$  are resistant to skin carcinogenesis," *Nature Medicine*, vol. 5, no. 7, pp. 828–831, 1999.
- [64] F.-M. Gu, Q.-L. Li, Q. Gao et al., "IL-17 induces AKT-dependent IL-6/JAK2/STAT3 activation and tumor progression in hepatocellular carcinoma," *Molecular Cancer*, vol. 10, article 150, 2011.



- [65] J.-L. Luo, S. Maeda, L.-C. Hsu, H. Yagita, and M. Karin, "Inhibition of NF- $\kappa$ B in cancer cells converts inflammation-induced tumor growth mediated by TNF $\alpha$  to TRAIL-mediated tumor regression," *Cancer Cell*, vol. 6, no. 3, pp. 297–305, 2004.
- [66] R. C. A. Sainson, D. A. Johnston, H. C. Chu et al., "TNF primes endothelial cells for angiogenic sprouting by inducing a tip cell phenotype," *Blood*, vol. 111, no. 10, pp. 4997–5007, 2008.
- [67] Y. Jing, N. Ma, T. Fan et al., "Tumor necrosis factor- $\alpha$  promotes tumor growth by inducing vascular endothelial growth factor," *Cancer Investigation*, vol. 29, no. 7, pp. 485–493, 2011.
- [68] H.-E. Tzeng, C.-H. Tsai, Z.-L. Chang et al., "Interleukin-6 induces vascular endothelial growth factor expression and promotes angiogenesis through apoptosis signal-regulating kinase 1 in human osteosarcoma," *Biochemical Pharmacology*, vol. 85, no. 4, pp. 531–540, 2013.
- [69] H. Nozawa, C. Chiu, and D. Hanahan, "Infiltrating neutrophils mediate the initial angiogenic switch in a mouse model of multistage carcinogenesis," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 33, pp. 12493–12498, 2006.
- [70] R. Benelli, M. Morini, F. Carrozzino et al., "Neutrophils as a key cellular target for angiostatin: implications for regulation of angiogenesis and inflammation," *The FASEB Journal*, vol. 16, no. 2, pp. 267–269, 2002.
- [71] J. P. Shaw, N. Chuang, H. Yee, and P. Shamamian, "Polymorphonuclear neutrophils promote rFGF-2-induced angiogenesis *in vivo*," *Journal of Surgical Research*, vol. 109, no. 1, pp. 37–42, 2003.
- [72] T. Mentzel, L. F. Brown, H. F. Dvorak et al., "The association between tumour progression and vascularity in myxofibrosarcoma and myxoid/round cell liposarcoma," *Virchows Archiv*, vol. 438, no. 1, pp. 13–22, 2001.
- [73] P. E. Van Den Steen, B. Dubois, I. Nelissen, P. M. Rudd, R. A. Dwek, and G. Opdenakker, "Biochemistry and molecular biology of gelatinase B or matrix metalloproteinase-9 (MMP-9)," *Critical Reviews in Biochemistry and Molecular Biology*, vol. 37, no. 6, pp. 375–536, 2002.
- [74] L. M. Coussens, C. L. Tinkle, D. Hanahan, and Z. Werb, "MMP-9 supplied by bone marrow-derived cells contributes to skin carcinogenesis," *Cell*, vol. 103, no. 3, pp. 481–490, 2000.
- [75] G. Bergers, R. Brekken, G. McMahon et al., "Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis," *Nature Cell Biology*, vol. 2, no. 10, pp. 737–744, 2000.
- [76] P. E. Van den Steen, P. Proost, A. Wuyts, J. Van Damme, and G. Opdenakker, "Neutrophil gelatinase B potentiates interleukin-8 tenfold by aminoterminal processing, whereas it degrades CTAP-III, PF-4, and GRO- $\alpha$  and leaves RANTES and MCP-2 intact," *Blood*, vol. 96, no. 8, pp. 2673–2681, 2000.
- [77] B. Heissig, K. Hattori, S. Dias et al., "Recruitment of stem and progenitor cells from the bone marrow niche requires MMP-9 mediated release of Kit-ligand," *Cell*, vol. 109, no. 5, pp. 625–637, 2002.
- [78] S. Huang, M. Van Arsdall, S. Tedjarati et al., "Contributions of stromal metalloproteinase-9 to angiogenesis and growth of human ovarian carcinoma in mice," *Journal of the National Cancer Institute*, vol. 94, no. 15, pp. 1134–1142, 2002.
- [79] F. Shojaei, M. Singh, J. D. Thompson, and N. Ferrara, "Role of Bv8 in neutrophil-dependent angiogenesis in a transgenic model of cancer progression," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 105, no. 7, pp. 2640–2645, 2008.
- [80] F. Shojaei, X. Wu, C. Zhong et al., "Bv8 regulates myeloid-cell-dependent tumour angiogenesis," *Nature*, vol. 450, no. 7171, pp. 825–831, 2007.
- [81] G. P. Gupta and J. Massagué, "Cancer metastasis: building a framework," *Cell*, vol. 127, no. 4, pp. 679–695, 2006.
- [82] C.-F. Wu, W.-C. Chiang, C.-F. Lai et al., "Transforming growth factor  $\beta$ -1 stimulates profibrotic epithelial signaling to activate pericyte-myofibroblast transition in obstructive kidney fibrosis," *American Journal of Pathology*, vol. 182, no. 1, pp. 118–131, 2013.
- [83] J. E. De Larco, B. R. K. Wuertz, and L. T. Furcht, "The potential role of neutrophils in promoting the metastatic phenotype of tumors releasing interleukin-8," *Clinical Cancer Research*, vol. 10, no. 15, pp. 4895–4900, 2004.
- [84] J. D. Spicer, B. McDonald, J. J. Cools-Lartigue et al., "Neutrophils promote liver metastasis via Mac-1-mediated interactions with circulating tumor cells," *Cancer Research*, vol. 72, no. 16, pp. 3919–3927, 2012.
- [85] Z. G. Fridlender and S. M. Albelda, "Tumor-associated neutrophils: friend or foe?" *Carcinogenesis*, vol. 33, no. 5, pp. 949–955, 2012.
- [86] A. M. Houghton, D. M. Rzymkiewicz, H. Ji et al., "Neutrophil elastase-mediated degradation of IRS-1 accelerates lung tumor growth," *Nature Medicine*, vol. 16, no. 2, pp. 219–223, 2010.
- [87] P. Auguste, S. Lemiere, F. Larrieu-Lahargue, and A. Bikfalvi, "Molecular mechanisms of tumor vascularization," *Critical Reviews in Oncology/Hematology*, vol. 54, no. 1, pp. 53–61, 2005.
- [88] M. J. Slattery and C. Dong, "Neutrophils influence melanoma adhesion and migration under flow conditions," *International Journal of Cancer*, vol. 106, no. 5, pp. 713–722, 2003.
- [89] S. J. Huh, S. Liang, A. Sharma, C. Dong, and G. P. Robertson, "Transiently entrapped circulating tumor cells interact with neutrophils to facilitate lung metastasis development," *Cancer Research*, vol. 70, no. 14, pp. 6071–6082, 2010.
- [90] J. Cools-Lartigue, J. Spicer, B. McDonald et al., "Neutrophil extracellular traps sequester circulating tumor cells and promote metastasis," *Journal of Clinical Investigation*, vol. 123, no. 8, pp. 3446–3458, 2013.
- [91] J. A. Joyce and J. W. Pollard, "Microenvironmental regulation of metastasis," *Nature Reviews Cancer*, vol. 9, no. 4, pp. 239–252, 2009.
- [92] J. T. Erler, K. L. Bennewith, T. R. Cox et al., "Hypoxia-induced lysyl oxidase is a critical mediator of bone marrow cell recruitment to form the premetastatic niche," *Cancer Cell*, vol. 15, no. 1, pp. 35–44, 2009.
- [93] R. N. Kaplan, R. D. Riba, S. Zacharoulis et al., "VEGFR1-positive haematopoietic bone marrow progenitors initiate the premetastatic niche," *Nature*, vol. 438, no. 7069, pp. 820–827, 2005.
- [94] R. N. Kaplan, S. Rafii, and D. Lyden, "Preparing the 'soil': the premetastatic niche," *Cancer Research*, vol. 66, no. 23, pp. 11089–11093, 2006.
- [95] J. P. Sleeman, "The metastatic niche and stromal progression," *Cancer and Metastasis Reviews*, vol. 31, no. 3-4, pp. 429–440, 2012.
- [96] J. Sceneay, M. J. Smyth, and A. Möller, "The pre-metastatic niche: finding common ground," *Cancer and Metastasis Reviews*, vol. 32, no. 3-4, pp. 449–464, 2013.
- [97] S. Hiratsuka, A. Watanabe, H. Aburatani, and Y. Maru, "Tumour-mediated upregulation of chemoattractants and recruitment of myeloid cells predetermines lung metastasis," *Nature Cell Biology*, vol. 8, no. 12, pp. 1369–1375, 2006.

- [98] S. Hiratsuka, A. Watanabe, Y. Sakurai et al., "The S100A8-serum amyloid A3-TLR4 paracrine cascade establishes a pre-metastatic phase," *Nature Cell Biology*, vol. 10, no. 11, pp. 1349–1355, 2008.
- [99] H. H. Yan, M. Pickup, Y. Pang et al., "Gr-1+CD11b+ myeloid cells tip the balance of immune protection to tumor promotion in the premetastatic lung," *Cancer Research*, vol. 70, no. 15, pp. 6139–6149, 2010.
- [100] H. B. Acuff, K. J. Carter, B. Fingleton, D. L. Gorden, and L. M. Matrisian, "Matrix metalloproteinase-9 from bone marrow-derived cells contributes to survival but not growth of tumor cells in the lung microenvironment," *Cancer Research*, vol. 66, no. 1, pp. 259–266, 2006.
- [101] M. Radsak, C. Iking-Konert, S. Stegmaier, K. Andrassy, and G. M. Hänsch, "Polymorphonuclear neutrophils as accessory cells for T-cell activation: major histocompatibility complex class II restricted antigen-dependent induction of T-cell proliferation," *Immunology*, vol. 101, no. 4, pp. 521–530, 2000.
- [102] N. S. Potter and C. V. Harding, "Neutrophils process exogenous bacteria via an alternate class I MHC processing pathway for presentation of peptides to T lymphocytes," *Journal of Immunology*, vol. 167, no. 5, pp. 2538–2546, 2001.
- [103] E. Reali, R. Guerrini, S. Moretti et al., "Polymorphonuclear neutrophils pulsed with synthetic peptides efficiently activate memory cytotoxic T lymphocytes," *Journal of Leukocyte Biology*, vol. 60, no. 2, pp. 207–213, 1996.
- [104] J. Pillay, V. M. Kamp, E. Van Hoffen et al., "A subset of neutrophils in human systemic inflammation inhibits T cell responses through Mac-1," *Journal of Clinical Investigation*, vol. 122, no. 1, pp. 327–336, 2012.
- [105] M. Munder, F. Mollinedo, J. Calafat et al., "Arginase I is constitutively expressed in human granulocytes and participates in fungicidal activity," *Blood*, vol. 105, no. 6, pp. 2549–2556, 2005.
- [106] J.-I. Youn, S. Nagaraj, M. Collazo, and D. I. Gabrilovich, "Subsets of myeloid-derived suppressor cells in tumor-bearing mice," *Journal of Immunology*, vol. 181, no. 8, pp. 5791–5802, 2008.
- [107] L. C. Jacobsen, K. Theilgaard-Mönch, E. I. Christensen, and N. Borregaard, "Arginase 1 is expressed in myelocytes/metamyelocytes and localized in gelatinase granules of human neutrophils," *Blood*, vol. 109, no. 7, pp. 3084–3087, 2007.
- [108] P. C. Rodriguez, M. S. Ernstoff, C. Hernandez et al., "Arginase I-producing myeloid-derived suppressor cells in renal cell carcinoma are a subpopulation of activated granulocytes," *Cancer Research*, vol. 69, no. 4, pp. 1553–1560, 2009.
- [109] A. H. Zea, P. C. Rodriguez, M. B. Atkins et al., "Arginase-producing myeloid suppressor cells in renal cell carcinoma patients: a mechanism of tumor evasion," *Cancer Research*, vol. 65, no. 8, pp. 3044–3048, 2005.
- [110] S. B. Coffelt, K. Kersten, C. W. Doornebal et al., "IL-17-producing  $\gamma\delta$  T cells and neutrophils conspire to promote breast cancer metastasis," *Nature*, vol. 522, no. 7556, pp. 345–348, 2015.
- [111] H. K. Jensen, F. Donskov, N. Marcussen, M. Nordmark, F. Lundbeck, and H. von der Maase, "Presence of intratumoral neutrophils is an independent prognostic factor in localized renal cell carcinoma," *Journal of Clinical Oncology*, vol. 27, no. 28, pp. 4709–4717, 2009.
- [112] K. Xie, "Interleukin-8 and human cancer biology," *Cytokine and Growth Factor Reviews*, vol. 12, no. 4, pp. 375–391, 2001.
- [113] A. Bellocq, M. Antoine, A. Flahault et al., "Neutrophil alveolitis in bronchioloalveolar carcinoma: induction by tumor-derived interleukin-8 and relation to clinical outcome," *American Journal of Pathology*, vol. 152, no. 1, pp. 83–92, 1998.
- [114] L.-F. Lee, R. P. Hellendall, Y. Wang et al., "IL-8 reduced tumorigenicity of human ovarian cancer in vivo due to neutrophil infiltration," *Journal of Immunology*, vol. 164, no. 5, pp. 2769–2775, 2000.
- [115] H. Schaidler, M. Oka, T. Bogenrieder et al., "Differential response of primary and metastatic melanomas to neutrophils attracted by IL-8," *International Journal of Cancer*, vol. 103, no. 3, pp. 335–343, 2003.
- [116] G. Opendakker and J. Van Damme, "The countercurrent principle in invasion and metastasis of cancer cells. Recent insights on the roles of chemokines," *International Journal of Developmental Biology*, vol. 48, no. 5–6, pp. 519–527, 2004.
- [117] M. P. Keane, J. A. Belperio, Y. Y. Xue, M. D. Burdick, and R. M. Strieter, "Depletion of CXCR2 inhibits tumor growth and angiogenesis in a murine model of lung cancer," *Journal of Immunology*, vol. 172, no. 5, pp. 2853–2860, 2004.
- [118] H. Katoh, D. Wang, T. Daikoku, H. Sun, S. K. Dey, and R. N. DuBois, "CXCR2-expressing myeloid-derived suppressor cells are essential to promote colitis-associated tumorigenesis," *Cancer Cell*, vol. 24, no. 5, pp. 631–644, 2013.
- [119] J. Arnold, E. C. De Boer, M. A. O'Donnell, A. Böhle, and S. Brandau, "Immunotherapy of experimental bladder cancer with recombinant BCG expressing interferon- $\gamma$ ," *Journal of Immunotherapy*, vol. 27, no. 2, pp. 116–123, 2004.
- [120] O. Florey and D. O. Haskard, "Sphingosine 1-phosphate enhances Fc $\gamma$  receptor-mediated neutrophil activation and recruitment under flow conditions," *The Journal of Immunology*, vol. 183, no. 4, pp. 2330–2336, 2009.
- [121] R. H. Wenger, "Cellular adaptation to hypoxia: O<sub>2</sub>-sensing protein hydroxylases, hypoxia-inducible transcription factors, and O<sub>2</sub>-regulated gene expression," *The FASEB Journal*, vol. 16, no. 10, pp. 1151–1162, 2002.
- [122] G. Christoffersson, E. Vågesjö, J. Vandooren et al., "VEGF-A recruits a proangiogenic MMP-9-delivering neutrophil subset that induces angiogenesis in transplanted hypoxic tissue," *Blood*, vol. 120, no. 23, pp. 4653–4662, 2012.
- [123] M. Grunewald, I. Avraham, Y. Dor et al., "VEGF-induced adult neovascularization: recruitment, retention, and role of accessory cells," *Cell*, vol. 124, no. 1, pp. 175–189, 2006.
- [124] J. M. Devery, N. J. C. King, and C. L. Geczy, "Acute inflammatory activity of the S100 protein CP-10. Activation of neutrophils in vivo and in vitro," *Journal of Immunology*, vol. 152, no. 4, pp. 1888–1897, 1994.
- [125] B. Amulic, C. Cazalet, G. L. Hayes, K. D. Metzler, and A. Zychlinsky, "Neutrophil function: from mechanisms to disease," *Annual Review of Immunology*, vol. 30, pp. 459–489, 2012.
- [126] M. L. Gabelloni, A. S. Trevani, J. Sabaté, and J. Geffner, "Mechanisms regulating neutrophil survival and cell death," *Seminars in Immunopathology*, vol. 35, no. 4, pp. 423–437, 2013.
- [127] W. C. Liles, P. A. Kiener, J. A. Ledbetter, A. Aruffo, and S. J. Klebanoff, "Differential expression of Fas (CD95) and Fas ligand on normal human phagocytes: implications for the regulation of apoptosis in neutrophils," *Journal of Experimental Medicine*, vol. 184, no. 2, pp. 429–440, 1996.
- [128] M. Chawla-Sarkar, D. W. Leaman, and E. C. Borden, "Preferential induction of apoptosis by interferon (IFN)-beta compared with IFN-alpha2: correlation with TRAIL/Apo2L induction in melanoma cell lines," *Clinical Cancer Research*, vol. 7, no. 6, pp. 1821–1831, 2001.

- [129] C. Sellaeri, T. Sato, L. Del Vecchio et al., "Involvement of fas-mediated apoptosis in the inhibitory effects of interferon- $\alpha$  in chronic myelogenous leukemia," *Blood*, vol. 89, no. 3, pp. 957–964, 1997.
- [130] T. Kuijpers and R. Lutter, "Inflammation and repeated infections in CGD: two sides of a coin," *Cellular and Molecular Life Sciences*, vol. 69, no. 1, pp. 7–15, 2012.
- [131] Y. Kasahara, K. Iwai, A. Yachie et al., "Involvement of reactive oxygen intermediates in spontaneous and CD95 (Fas/APO-1)-mediated apoptosis of neutrophils," *Blood*, vol. 89, no. 5, pp. 1748–1753, 1997.
- [132] H. Lundqvist-Gustafsson and T. Bengtsson, "Activation of the granule pool of the NADPH oxidase accelerates apoptosis in human neutrophils," *Journal of Leukocyte Biology*, vol. 65, no. 2, pp. 196–204, 1999.
- [133] B. Dibbert, M. Weber, W. H. Nikolaizik et al., "Cytokine-mediated Bax deficiency and consequent delayed neutrophil apoptosis: a general mechanism to accumulate effector cells in inflammation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 96, no. 23, pp. 13330–13335, 1999.
- [134] N. A. Maianski, D. Roos, and T. W. Kuijpers, "Bid truncation, Bid/Bax targeting to the mitochondria, and caspase activation associated with neutrophil apoptosis are inhibited by granulocyte colony-stimulating factor," *Journal of Immunology*, vol. 172, no. 11, pp. 7024–7030, 2004.



**Hindawi**  
Submit your manuscripts at  
<http://www.hindawi.com>

