

Research Article

In Vitro Vitamin K₃ Effect on Conjunctival Fibroblast Migration and Proliferation

I. Pinilla,^{1,2} L. B. Izaguirre,³ F. J. Gonzalvo,⁴ E. Piazuelo,² M. A. Garcia-Gonzalez,² A. I. Sanchez-Cano,^{2,5} and F. Sopeña^{2,6}

¹ Department of Ophthalmology, Lozano Blesa University Hospital, C/San Juan Bosco 15, 50009 Zaragoza, Spain

² Aragon Institute of Health Sciences (IIS Aragon), 50009 Zaragoza, Spain

³ Department of Ophthalmology, Hospital García Orcoyen, Navarra, 31200 Estella, Spain

⁴ Department of Ophthalmology, Complejo Hospitalario de Navarra, Navarra, 31008 Pamplona, Spain

⁵ Department of Applied Physics, Zaragoza University, 50009 Zaragoza, Spain

⁶ Department of Gastroenterology, Lozano Blesa University Hospital, C/San Juan Bosco 15, 50009 Zaragoza, Spain

Correspondence should be addressed to I. Pinilla; ipinilla@unizar.es

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Purpose. To evaluate the dose effect of vitamin K_3 on wound healing mechanisms. *Methods.* Conjunctival fibroblasts were incubated for 24 hours. An artificial wound was made and the cells were incubated with fresh medium plus doses of vitamin K_3 to be tested. Wound repair was monitored at 0, 18, 24, and 48 hours. Proliferation was measured in actively dividing cells by [³H]thymidine uptake. Six different groups were tested: group 1/no drugs added, group 2/ethanol 0.1%, group 3/vitamin K_3 1 mg/L, group 4/vitamin K_3 2 mg/L, group 5/vitamin K_3 4 mg/L, and group 6/vitamin K_3 6 mg/L. Each experiment was carried out in triplicate and 4 times. *Results.* There were no differences among groups at the initial time. *In vitro* wound repair was slower in groups 4, 5, and 6. There were no differences between control and ethanol groups and between control and vitamin K_3 1 mg/L groups. Fibroblast mitogenic activity was statistically decreased in all vitamin K groups; statistical differences were found among vitamin K_3 1 mg/mL and higher doses too. In groups 5 and 6, cellular toxicity was presented. *Conclusions.* Vitamin K_3 is able to inhibit fibroblast proliferation. Vitamin K_3 2 mg/L or higher doses inhibit wound healing repair, exhibiting cellular toxicity at 4 and 6 mg/L.

1. Introduction

Antimetabolites and other fibroblast inhibitor drugs have been shown to enhance the success rate of filtering surgery although, depending on the dose, they can lead to severe complications and may result in the failure of the surgery.

Corticosteroids [1–6] antiproliferative agents (5-fluorouracil and other fluoropyrimidines, taxol, doxorubicin, mycophenolate mofetil..., alone or in combination or with different delivery systems) [3–12], systemic, periocular, intraocular steroidal, and nonsteroidal anti-inflammatory agents [5, 13–16], colchicine [8], daunomycin [8], tissue plasminogen activator [17], heparin [12, 18–20], interferon-gamma [21, 22], calcium channel blockers [23], prolyl and lysyl hydroxylase inhibitors [19, 24–26], retinoic acid [27, 28], alpha-tocopherol [29–31], disintegrins [32], siRNA-PKC α [33]... are some of the useful drugs that have been used in the treatment of conditions such as proliferative vitreoretinopathy, bleb scarring after trabeculectomy, and other disorders with cell proliferation (progressive conjunctival or extraocular cicatrization).

Vitamin K_3 (menadione, 2-methyl-1,4-naphthoquinone) has been used as antihemorrhagic agent. Its ability to inhibit proliferation of tumor cells has already been reported; its activity has been demonstrated in human tumor stem cell and it is used in clinical trial for advanced malignancies acting in different pathways and has also been related to other oxidative stress processes at the eye level as cataract formation... [34–40]. Liu et al. reported that this drug could inhibit proliferation of rabbit conjunctive cells [41].



FIGURE 1: Phase-contrast microphotographs showing the process of wound healing in an ulcer treated with vitamin $K_3 1 \text{ mg/L}$. Lesion time, 0 hour: *in vitro* ulcer after being produced in a confluent monolayer; 24 hours fibroblasts migrations was evident; 48 hours: some fibroblast were filling the ulcer area.

The aim of this study was to evaluate and to compare the antiproliferative properties of vitamin K_3 in cultured human fibroblasts.

2. Methods

2.1. Material. All supplies for cell culture were purchased from Nunc (Roskilde, DK). Dulbecco's Modified Eagles Medium (DMEM), phosphate buffer saline (PBS), fetal calf serum (FCS), and antibiotics-antifungals were purchased from GIBCO (Madison, WI). [methyl-³H]thymidine was purchased from Amershm Iberica (Madrid, Spain). 2-Methyl 1,4-Naphthoquinone (Menadione) (98%) was obtained from Sigma (St. Louis, MO). The drug was initially dissolved in 90% ethanol. This alcohol solution was then diluted into BSS to yield a final ethanol concentration of 0.1%.

2.2. Cell Cultures. Conjunctival fibroblasts were obtained from explants of a healthy adult subject who underwent ophthalmic surgery for retinal detachment. All subjects gave informed consent to participate in the study, which was conducted in accordance with the tenets of the Declaration of Helsinki, and the experimental protocol was approved by the local Ethics Committee of the Aragon Health Science Institute. Cells were cultured in uncoated plastic flasks in DMEM supplemented with antibiotics and antifungals (100 IU/mL penicillin, 100 μ g/mL streptomycin, and 0.25 μ g/mL amphotericin B), and 20% fetal calf serum (FCS) in a humidified atmosphere at 37 degrees Celsius and 5% CO₂. The culture medium was changed every 3 day and the experiments were performed with cells obtained between the 5th and 8th passages.

2.3. Wounding Assays. Wounding assays were performed using the method described by Sato and Watanabe [42]. An artificial wound was made by mechanical cell denudation with a rotating tip, as described in previous papers [26]. The wound repair process was monitored by two independent observers measuring the cell-free area (mm²) in a blind fashion, at different times: 0, 18, 24, and 48 hours (Figure 1). The cell-free area was quantified in the elliptic or circular shape wounds with homogeneous size. Then, the major and the minor axes were measured in a phase-contrast microscopy equipped with a calibrated visor. The area was calculated applying the mathematical formula: area = $A \times B \times \pi/4$, where A and B were the major and the minor axes, respectively.

2.4. Assay of Cell Mitogenic Activity. Freshly trypsinized fibroblasts were seeded in 24-well plates at a density of 15×10^4 cells/well and were incubated for 24 hours in fresh medium and the drugs to be tested. Cells were labeled for the last 3 hours period with 1 μ Ci/mL of [methyl-³H]thymidine.

TABLE 1: Measurement of the wound areas (values are expressed as mean \pm SD).

AREA (mm ²)	0 hours	18 hours	24 hours	48 hours	
Control	0.599 ± 0.118	0.332 ± 0.108	0.267 ± 0.118	0.069 ± 0.066	
K ₃ 1 mg/L	0.569 ± 0.099	0.405 ± 0.056	0.356 ± 0.048	0.125 ± 0.073	
K ₃ 2 mg/L	0.669 ± 0.120	0.672 ± 0.095	0.619 ± 0.082	0.459 ± 0.232	
K ₃ 4 mg/L	0.554 ± 0.162	0.771 ± 0.096	>1 mm ²	>1 mm ²	
K ₃ 6 mg/L	0.582 ± 0.110	0.774 ± 0.039	$>1 \mathrm{mm}^2$	>1 mm ²	

After removing the media, the cells were washed 3 times with ice-cold PBS and then 2 times with 5% ice-cold trichloroacetic acid to precipitate the DNA. The precipitate was dissolved in 500 μ L 0.1 N NaOH and 0.1% sodium dodecyl sulfate. The extract was neutralized with 0.1 N HCl and radioactivity was counted in a liquid scintillation counter (1900 TR, Packard Instrument Company, Meriden, CT).

2.5. Groups of Treatment. There were 6 different treatment groups: group 1/control group: no drugs added, group 2/0.5 μ L of ethanol 10% (final ethanol concentration 0.1%); group 3/vitamin K₃ 1 mg/L; group 4/vitamin K₃ 2 mg/L; group 5/vitamin K₃ 4 mg/L; group 6/vitamin K₃ 6 mg/L.

2.6. Statistical Analysis. Each experiment was carried out in triplicate and at least 4 times. The results were expressed as mean \pm standard deviation ($x \pm$ SD). Statistical significances between mean values were assessed with Mann-Whitney *U*-test. The probability level at which the Null Hypothesis was rejected was set at *P* < 0.05.

3. Results and Discussion

Wound healing in some ocular diseases and surgeries, as mucous-cutaneous diseases or after glaucoma filtering surgery, or complicated retinal detachment, is one of the problems that needs to be solved. We were studying the effect of vitamin K on wound healing and its possible toxicity.

The wound area is presented in Table 1. There were no differences among the groups at the initial time (Table 1). The mean size was $0.586 \text{ mm}^2 \pm 0.082$. There were no differences between control group and ethanol group. Ethanol 0.1 mg/mL did not show effect on fibroblast migration and proliferation. No toxic effect had been related to its use. Vitamin K is a liposoluble drug. This fact can be remarkable in order to use it to assist the effect of the silicone oil on complicated retinal detachments.

Wound healing process can be divided into three phases: inflammation, proliferation, and modulation of the scar. This process begins immediately after the injury. The fibroblast proliferation appears after 24 hours [43]. In this experimental model of wound healing, we evaluate the fibroblast migration 18 hours after the ulcer has been done. In the 24 hours' time migration and proliferation are evident.

Vitamin K_3 at the doses 2, 4, and 6 mg/L significantly decreased the speed of wound repair during the experiment. There were no differences between control group and vitamin $K_3 1 \text{ mg/L}$ (Table 2). Vitamin K_3 at the doses 2, 4, and 6 mg/L

TABLE 2: Differences between the wound areas of the groups at 0 (a), 18 (b), 24 (c), and 48 hours (d). NS means P > 0.05. S means $P \le 0.05$.

(a)

(a)								
0 hours	Control	K ₃ 1 mg/L	K ₃ 2 mg/L	K ₃ 4 mg/L	K ₃ 6 mg/L			
Control		NS	NS	NS	NS			
K ₃ 1mg/L			NS	NS	NS			
K ₃ 2 mg/L				NS	NS			
K ₃ 4 mg/I					NS			
(b)								
18 hours	Control	$K_3 1 mg/L$	$K_3 2 mg/L$	$K_3 4 mg/L$	K ₃ 6 mg/L			
Control		NS	S	S	S			
$K_3 1 mg/L$			S	S	S			
K ₃ 2 mg/L				NS	NS			
K ₃ 4 mg/I					NS			
(c)								
24 hours	Control	K ₃ 1 mg/L	K ₃ 2 mg/L	K ₃ 4 mg/L	K ₃ 6 mg/L			
Control		NS	S	S	S			
$K_3 1 mg/L$			S	S	S			
K ₃ 2 mg/L				S	S			
K ₃ 4 mg/I					NS			
(d)								
48 hours	Control	K ₃ 1mg/L	K ₃ 2 mg/L	K ₃ 4 mg/L	K ₃ 6 mg/L			
Control		NS	S	S	S			
K ₃ 1 mg/L			S	S	S			
K ₃ 2 mg/L	,			S	S			
K ₃ 4 mg/L	,				NS			

inhibited the cell migration and proliferation and showed slower closure of the wounds than the other groups (from 18 hours on). Liu et al. found that most of the cells died at concentrations of 7.5 mg/L; the concentration of 4.0 mg/L inhibited fifty percent of the cellular growth. The cellular border became clearer and some cells started to die at 5 mg/L [41].

Fibroblast mitogenic activity was significantly inhibited by all vitamin K_3 doses. There were differences between vitamin $K_3 \ 1 \ mg/mL$ and all the others vitamin K_3 groups. In this study, vitamin K_3 at $1 \ mg/L$ did not show differences with the control group in the speed of wound repair. Fibroblast mitogenic activity was inhibited by all doses of vitamin K_3 ; differences were found among vitamin $K_3 \ 1 \ mg/mL$ and the greater doses. Vitamin $K_3 \ 1 \ mg/mL$ is able to inhibit fibroblast mitogenic activity with no influence in wound repair; this effect has probably been counteracted by its no migration inhibitory effect.

In our study, doses of 4 mg/L induced great cellular alterations. Vitamin K₃ 4 mg/L and 6 mg/L induced cellular toxicity. Cells presented changes in their morphology, which characterized apoptosis, including nuclear and cytoplasmic condensation with intact plasma membrane cell. They lost their adherence to the plate, showing a growing ulcer throughout the time being the ulcer sizes larger than in the other groups. MMC and 5-FU are also able to induce apoptosis in cultured tenon's fibroblast [44].

The application of experimental data derived from cell cultures to clinical use has limitations. Variables such as bioavailability, diffusional barriers, metabolic inactivation, excretion, drug resistance, and enzyme induction prohibit simple extrapolation of cell culture data to human diseases. Nevertheless, this basic approach to drug selection is invaluable.

The mechanism of cytotoxicity of vitamin K_3 is not well known and it has been the focus of multiple papers. The drug is able to affect the cell by two mechanisms. One is its ability to disturb the intracellular calcium flux and calcium-dependent potassium flux [45]. The other is that its chemical transformation within the cell may generate reactive oxygen species and potentially deplete intracellular glutathione [46]. Effects on different cells have been described such as inhibition of PTP-1B in keratinocytes [47], induction of tumor cell death through hydrogen peroxide generation, and regulation of the expression of G1 phase-related cell cycle molecules [48, 49].

We can conclude that all the studied doses of vitamin K were able to inhibit fibroblast mitogenic activity. Vitamin K_3 , at 2 mg/L or higher doses, interfere the mechanisms of cell repair, delaying the wound healing process in this *in vitro* model. Vitamin K_3 at 4 and 6 mg/L in cell culture showed fibroblast toxicity. The drug could be considered an alternative to the drug treatment and prevention of exaggerated scarring in some ocular diseases.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

References

- S. J. Ryan, "Traction retinal detachment XLIX Edward Jackson Memorial Lecture," *The American Journal of Ophthalmology*, vol. 115, no. 1, pp. 1–20, 1993.
- [2] F. Koerner, A. Merz, B. Gloor, and E. Wagner, "Postoperative retinal fibrosis—a controlled clinical study of systemic steroid therapy," *Graefe's Archive for Clinical and Experimental Ophthalmology*, vol. 219, no. 6, pp. 268–271, 1982.
- [3] A. García-Layana, M. T. Hernando, L. Manzanas, and J. C. Pastor, "Tratamiento profiláctico de la vitreorretinopatía proliferante," *Archivos de la Sociedad Española de Oftalmología*, vol. 60, pp. 315–322, 1991.
- [4] R. Fiscella, G. A. Peyman, J. Elvart, and B. Yue, "In vitro evaluation of cellular inhibitory potential of various antineoplastic

drugs and dexamethasone," *Ophthalmic Surgery*, vol. 16, no. 4, pp. 247–249, 1985.

- [5] M. S. Blumenkranz, A. Claflin, and A. S. Hajek, "Selection of therapeutic agents for intraocluar proliferative disease. Cell culture evaluation," *Archives of Ophthalmology*, vol. 102, no. 4, pp. 598–604, 1984.
- [6] A. S. Berger, C. K. Cheng, P. A. Pearson et al., "Intravitreal sustained release corticosteroid-5-fluoruracil conjugate in the treatment of experimental proliferative vitreoretinopathy," *Investigative Ophthalmology and Visual Science*, vol. 37, no. 11, pp. 2318–2325, 1996.
- [7] M. S. Blumenkranz, M. K. Hartzer, and A. S. Hajek, "Selection of therapeutic agents for intraocular proliferative disease. II. Differing antiproliferative activity of the fluoropyrimidines," *Archives of Ophthalmology*, vol. 105, no. 3, pp. 396–399, 1987.
- [8] C. Verdoorn, V. W. Renardel de Lavalette, J. Dalma-Weizhausz, G. M. Orr, N. Sorgente, and S. J. Ryan, "Cellular migration, proliferation, and contraction. An in vitro approach to a clinical problem—proliferative vitreoretinopathy," *Archives of Ophthalmology*, vol. 104, no. 8, pp. 1216–1219, 1986.
- [9] C. Heinz, T. Hudde, K. Heise, and K. P. Steuhl, "Antiproliferative effect of mycophenolate mofetil on cultured human Tenon fibroblasts," *Graefe's Archive for Clinical and Experimental Ophthalmology*, vol. 240, no. 5, pp. 408–414, 2002.
- [10] C. Heinz, K. Heise, T. Hudde, and K. P. Steuhl, "Mycophenolate mofetil inhibits human Tenon fibroblast proliferation by guanosine depletion," *The British Journal of Ophthalmology*, vol. 87, no. 11, pp. 1397–1398, 2003.
- [11] M. Akimoto, T. Miyahara, J. Arai et al., "A new delivery system for 5-fluorouracil using prodrug and converting enzyme," *The British Journal of Ophthalmology*, vol. 86, no. 5, pp. 581–586, 2002.
- [12] V. Sundaram, A. Barsam, and G. Virgili, "Intravitreal low molecular weight heparin and 5-fluorouracil for the prevention of proliferative vitreoretinopathy following retinal reattachment surgery," *Cochrane Database of Systematic Reviews*, no. 1, Article ID CD006421, 2013.
- [13] M. Reibaldi, A. Russo, A. Longo et al., "Rhegmatogenous retinal detachment with a high risk of proliferative vitreoretinopathy treated with episcleral surgery and an intravitreal dexamethasone 0.7-mg implant," *Case Reports in Ophthalmology*, vol. 4, no. 1, pp. 79–83, 2013.
- [14] P. Yang, B. S. McKay, J. B. Allen, and G. J. Jaffe, "Effect of NF-κ B inhibition on TNF-α-induced apoptosis in human RPE cells," *Investigative Ophthalmology and Visual Science*, vol. 45, no. 7, pp. 2438–2446, 2004.
- [15] R. G. Williams, S. Chang, M. R. Comaratta, and G. Simoni, "Does the presence of heparin and dexamethasone in the vitrectomy infusate reduce reproliferation in proliferative vitreoretinopathy?" *Graefe's Archive for Clinical and Experimental Ophthalmology*, vol. 234, no. 8, pp. 496–503, 1996.
- [16] W. Chen, H. Chen, P. Hou, A. Fok, Y. Hu, and D. S. C. Lam, "Midterm results of low-dose intravitreal triamcinolone as adjunctive treatment for proliferative vitreoretinopathy," *Retina*, vol. 31, no. 6, pp. 1137–1142, 2011.
- [17] G. A. Williams, F. H. Lambrou, G. A. Jaffe et al., "Treatment of postvitrectomy fibrin formation with intraocular tissue activator," *Archives of Ophthalmology*, vol. 106, no. 8, pp. 1055– 1058, 1988.
- [18] R. N. Johnson and G. Blankenship, "A prospective, randomized, clinical trial of heparin therapy for postoperative intraocular fibrin," *Ophthalmology*, vol. 95, no. 3, pp. 312–317, 1988.

- [19] S. Scheer, C. Morel, O. Touzeau, J. A. Sahel, and L. Laroche, "Pharmacological adjuvants for surgical treatment of proliferative vitreoretinopathy," *Journal Francais d'Ophtalmologie*, vol. 27, no. 9, pp. 1051–1059, 2004.
- [20] D. G. Charteris, G. W. Aylward, D. Wong, C. Groenewald, R. H. Y. Asaria, and C. Bunce, "A randomized controlled trial of combined 5-fluorouracil and low-molecular-weight heparin in management of established proliferative vitreoretinopathy," *Ophthalmology*, vol. 111, no. 12, pp. 2240–2245, 2004.
- [21] M. A. Latina, S. J. Belmonte, C. Park, and E. Crean, "Gammainterferon effects on human fibroblasts from Tenon's capsule," *Investigative Ophthalmology and Visual Science*, vol. 32, no. 10, pp. 2806–2815, 1991.
- [22] L. M. Hjelmeland, J. W. Li, C. A. Toth, and M. B. Landers III, "Antifibrotic and uveitogenic properties of gamma interferon in the rabbit eye," *Graefe's Archive for Clinical and Experimental Ophthalmology*, vol. 230, no. 1, pp. 84–90, 1992.
- [23] Y. Kang, D. A. Lee, and E. J. Higginbotham, "In vitro evaluation of antiproliferative potential of calcium channel blockers in human Tenon's fibroblasts," *Experimental Eye Research*, vol. 64, no. 6, pp. 913–925, 1997.
- [24] J. T. Handa, S. Murad, and G. J. Jaffe, "Minoxidil inhibits ocular cell proliferation and lysyl hydroxylase activity," *Investigative Ophthalmology and Visual Science*, vol. 34, no. 3, pp. 567–575, 1993.
- [25] J. Wu and A. Danielsson, "Inhibition of hepatic fibrogenesis: a review of pharmacologic candidates," *Scandinavian Journal of Gastroenterology*, vol. 29, no. 5, pp. 385–391, 1994.
- [26] L. Izaguirre, I. Pinilla, F. Gonzalvo, S. Pérez, and F. M. Honrubia, "Effect of doxorubicin on fibroblast migration and proliferation," *Annals of Ophthalmology*, vol. 35, no. 1, pp. 48–52, 2003.
- [27] J. J. Araiz, M. F. Refojo, M. H. Arroyo, F. L. Leong, D. M. Albert, and F. I. Tolentino, "Antiproliferative effect of retinoic acid in intravitreous silicone oil in an animal model of proliferative vitreoretinopathy," *Investigative Ophthalmology and Visual Science*, vol. 34, no. 3, pp. 522–530, 1993.
- [28] J. W. Doyle, R. K. Dowgiert, and S. M. Buzney, "Factors modulating the effect of retinoids on cultured retinal pigment epithelial cell proliferation," *Current Eye Research*, vol. 11, no. 8, pp. 753–765, 1992.
- [29] J. M. Larrosa, A. A. S. Veloso Jr., F. L. Leong, and M. F. Refojo, "Antiproliferative effect of intravitreal α-tocopherol and α-tocopheryl-acid-succinate in a rabbit model of PVR," *Current Eye Research*, vol. 16, no. 10, pp. 1030–1035, 1997.
- [30] J. M. Larrosa, V. Polo, T. Ramirez, I. Pinilla, L. E. Pablo, and F. M. Honrubia, "Alpha-tocopherol derivatives and wound healing in an experimental model of filtering surgery," *Ophthalmic Surgery* and Lasers, vol. 31, no. 2, pp. 131–135, 2000.
- [31] I. Pinilla, J. M. Larrosa, V. Polo, E. Piazuelo, P. Jimenez, and F. M. Honrubia, "Comparison of fibroblast inhibitory effect of α-tocopherol succinate and 13-cis retinol," *Annals of Ophthalmology*, vol. 34, no. 2, pp. 108–112, 2002.
- [32] C. H. Yang, T. F. Huang, K. R. Liu, M. S. Chen, and P. T. Hung, "Inhibition of retinal pigment epithelial cell-induced tractional retinal detachment by disintegrins, a group of Arg-Gly-Asp-containing peptides from viper venom," *Investigative Ophthalmology and Visual Science*, vol. 37, no. 5, pp. 843–854, 1996.
- [33] Q. Gao, W. Wang, Y. Lan et al., "The inhibitory effect of small interference RNA protein kinase $C-\alpha$ on the experimental proliferative vitreoretinopathy induced by dispase in mice,"

International Journal of Nanomedicine, vol. 8, pp. 1563–1572, 2013.

- [34] R. T. Chlebowski, S. A. Akman, and J. B. Block, "Vitamin K in the treatment of cancer," *Cancer Treatment Reviews*, vol. 12, no. 1, pp. 49–63, 1985.
- [35] D. Lim, R. J. Morgan Jr., S. Akman et al., "Phase I trial of menadiol diphosphate (vitamin K3) in advanced malignancy," *Investigational New Drugs*, vol. 23, no. 3, pp. 235–239, 2005.
- [36] P. Oztopçu, S. Kabadere, A. Mercangöz, and R. Uyar, "Comparison of vitamins K1, K2, and K3 effects on growth of rat glioma and human glioblastoma multiforme cells in vitro," *Acta Neurologica Belgica*, vol. 104, no. 3, pp. 106–110, 2004.
- [37] M. Ishibashi, M. Arai, S. Tanaka, K. Onda, and T. Hirano, "Antiproliferative and apoptosis-inducing effects of lipophilic vitamins on human melanoma A375 cells in vitro," *Biological* and Pharmaceutical Bulletin, vol. 35, no. 1, pp. 10–17, 2012.
- [38] B. R. Acharya, D. Choudhury, A. Das, and G. Chakrabarti, "Vitamin K3 disrupts the microtubule networks by binding to tubulin: a novel mechanism of its antiproliferative activity," *Biochemistry*, vol. 48, no. 29, pp. 6963–6974, 2009.
- [39] A. M. Marchionatti, G. Picotto, C. J. Narvaez, J. Welsh, and N. G. T. de Talamoni, "Antiproliferative action of menadione and 1, 25(OH)2D3 on breast cancer cells," *Journal of Steroid Biochemistry and Molecular Biology*, vol. 113, no. 3-5, pp. 277– 232, 2009.
- [40] K. R. Hegde and S. D. Varma, "Combination of glycemic and oxidative stress in lens: implications in augmentation of cataract formation in diabetes," *Free Radical Research*, vol. 39, no. 5, pp. 513–517, 2005.
- [41] X. H. Liu, X. W. Song, Y. Xu, and C. Zhang, "The inhibition of vitamin K3 on rabbit fibroblast proliferation in vitro," *Ophthalmologica*, vol. 210, no. 3, pp. 180–182, 1996.
- [42] N. Sato and S. Watanabe, "Influence of extracellular matrix in gastric mucosal repair in vitro," *Biochemical and Biophysical Research Communications*, vol. 202, no. 1, pp. 285–292, 1994.
- [43] R. J. Kirsner and H. Eaglestein, "The wound healing process," *Dermatologic Clinics*, vol. 11, no. 4, pp. 629–640, 1993.
- [44] J. G. Crowston, A. N. Akbar, P. H. Constable, N. L. Occleston, J. T. Daniels, and P. T. Khaw, "Antimetabolite-induced apoptosis in Tenon's capsule fibroblasts," *Investigative Ophthalmology and Visual Science*, vol. 39, no. 2, pp. 449–454, 1998.
- [45] F. Kawamura, N. Hirashima, T. Furuno, and M. Nakanishi, "Effects of 2-methyl-1,4-naphtoquinone (Menadione) on cellular signaling in RBL-2H3 cells," *Biological and Pharmaceutical Bulletin*, vol. 29, no. 4, pp. 605–607, 2006.
- [46] H. Morrison, B. Jernstrom, and M. Nordenskjold, "Induction of DNA damage by menadione (2-methyl-1,4-naphthoquinone) in primary cultures of rat hepatocytes," *Biochemical Pharmacology*, vol. 33, no. 11, pp. 1763–1769, 1984.
- [47] J. I. Beier, C. von Montfort, H. Sies, and L. O. Klotz, "Activation of ErbB2 by 2-methyl-1,4-naphthoquinone (menadione) in human keratinocytes: role of EGFR and protein tyrosine phosphatases," *FEBS Letters*, vol. 580, no. 7, pp. 1859–1864, 2006.
- [48] C. Lin, J. Kang, and R. Zheng, "Vitamin K3 triggers human leukemia cell death through hydrogen peroxide generation and histone hyperacetylation," *Pharmazie*, vol. 60, no. 10, pp. 765– 771, 2005.
- [49] S. Kuriyama, M. Hitomi, H. Yoshiji et al., "Vitamins K2, K3 and K5 exert in vivo antitumor effects on hepatocellular carcinoma by regulating the expression of G1 phase-related cell cycle molecules," *International Journal of Oncology*, vol. 27, no. 2, pp. 505–511, 2005.



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