Acetylation controls sterol export

Supplementary information

Supplementary materials and methods

Mass spectrometry

For lipid analysis by mass spectrometry, heme-deficient *say1* Δ (YRS1853) mutant cells were grown in cholesterol containing media. Lipids were extracted from the cell pellet, concentrated and analyzed in the positive ion mode on a Bruker Esquire HCT ion trap mass spectrometer (ESI) with a flow rate of 120 µl/h and a capillary tension of -250 V. Ion fragmentation was induced with argon as collision gas at a pressure of 8 mbar.

Lipid labeling and analysis

To examine ergosterol dependent export into the culture media, cells were grown in SC media containing Tween 80 (0.05 mg/ml) and [¹⁴C]cholesterol (0.025 μ Ci/ml), for 16 h at 24°C. Cells were collected by centrifugation, washed twice with SC media to remove the unincorporated label. Cells were then diluted to OD₆₀₀ of ~1 in media either containing no ergosterol, 2 µg/ml, 20 µg/ml or 200 µg/ml of cold ergosterol and cells were cultivated for 24 h. Cells were collected and lipids were extracted from the cell pellet and culture supernatant. Lipids were separated by thin-layer chromatography on silica gel 60 plates (TLC; Merck, Darmstadt, Germany) with the solvent system petroleum ether/diethylether/acetic acid (70:30:2; per vol.) and radiolabeled lipids were analyzed and quantified by scanning with a Berthold Tracemaster 40 Automatic TLC-Linear Analyzer (Berthold Technologies, Bad Wildbad, Germany).

To examine the substrate specificity of the acetylation and export pathway,

heme-deficient cells were cultivated at 24°C for 16 h in YPD media containing Tween 80 (0.05 mg/ml) and either [³H]7-ketocholesterol, [³H]lanosterol at 1 μ Ci/ml or [¹⁴C]25-hydroxycholesterol, [¹⁴C] β -sitosterol, or [¹⁴C]progesterone (American Radiolabeled Chemicals Inc) at 0.025 μ Ci/ml. Cells were collected by centrifugation and lipids were extracted from the cell pellet and the culture supernatant. Samples were dried and analyzed by TLC.

Oxygen-dependence of Say1 and Atf2 levels

To examine the expression levels of Say1 and Atf2 under aerobic and heme-deficient conditions, hem-deficient cells expressing C-terminally myc- or GFP-tagged versions of Say1 and Atf2, respectively, were cultivated in YPD media containing either Tween 80 (5 mg/ml) and cholesterol (20 μ g/ml) or 10 μ g/ml aminolevulinic acid (ALA) for 16 h. Cells were diluted to OD₆₀₀ of ~0.5 in fresh media and samples were removed after 0, 2, 8 and 16 h of growth at 24°C. Proteins were extracted from equal OD units of cells, precipitated, separated by electrophoresis, blotted and probed with antibodies against myc, GFP or Pgk1.

Supplementary references

- Nakamura, K., Niimi, M., Niimi, K., Holmes, A.R., Yates, J.E., Decottignies, A., Monk, B.C., Goffeau, A. and Cannon, R.D. (2001) Functional expression of Candida albicans drug efflux pump Cdr1p in a Saccharomyces cerevisiae strain deficient in membrane transporters. Antimicrob Agents Chemother, 45, 3366-3374.
- Winzeler, E.A., Shoemaker, D.D., Astromoff, A., Liang, H., Anderson, K., Andre, B.,
 Bangham, R., Benito, R., Boeke, J.D., Bussey, H., Chu, A.M., Connelly, C.,
 Davis, K., Dietrich, F., Dow, S.W., El Bakkoury, M., Foury, F., Friend, S.H.,
 Gentalen, E., Giaever, G., Hegemann, J.H., Jones, T., Laub, M., Liao, H.,

Davis, R.W. and et al. (1999) Functional characterization of the *S. cerevisiae* genome by gene deletion and parallel analysis. *Science*, **285**, 901-906.

Supplementary tables

Table SI: S. cerevisiae strains used in this study

Strain	Relevant genotype	Source or reference
YRS1533	BY4742; MATα his3Δ1 leu2Δ0 ura3Δ0 lys2Δ0	EUROSCARF; Winzeler et al., 1999
YRS1845	MATa ade $2\Delta 0$ his $3\Delta 1$ leu $2\Delta 0$ ura $3\Delta 0$ sec 12^{ts}	This study
YRS1849	MATα his3Δ1 leu2Δ0 ura3Δ0 lys2Δ0 hem1::LEU2	This study
YRS1853	MATα his3∆1 leu2∆0 ura3∆0 lys2∆0 say1::kanMX4 hem1::LEU2	This study
YRS2133	MATα his3Δ1 leu2Δ0 ura3Δ0 lys2Δ0 atf2::kanMX4 say1::HIS3MX6	This study
YRS2135	MATα his3∆1 leu2∆0 ura3∆0 lys2∆ atf2::kanMX4 hem1::LEU2	This study
YRS2136	MATα his3Δ1 leu2Δ0 ura3Δ0 lys2Δ0 atf2::kanMX4 say1::HIS3MX6 hem1::LEU 2	This study
YRS2209	MATa. his3∆1 leu2∆0 ura3∆0 lys2∆0 atf1::kanMX4 say1::HIS3MX6 hem1::LEU2	This study
YRS2211	MATα his3Δ1 leu2Δ0 ura3Δ0 lys2Δ GAL1- GFP-SAY1-HIS3MX6	This study
YRS2212	MATα his3Δ1 leu2Δ0 ura3Δ0 lys2Δ GAL1- GFP-SAY1-HIS3MX6 hem1::LEU2	This study
YRS2483	MATα his3Δ1 leu2Δ0 ura3Δ0 lys2Δ0 ATF2- GFP-HIS3MX6	This study
YRS2486	MATα his3Δ1 leu2Δ0 ura3Δ0 lys2Δ0 are1::kanMX4 are2::kanMX4	This study
YRS2495	MATa his3∆1 leu2∆0 ura3∆0 lys2∆0 say1::HIS3MX6 hem1::LEU2 pdr5::kanMX4 snq2::kanMX4	This study
YRS2528	MATα his3Δ1 leu2Δ0 ura3Δ0 lys2Δ SAY1- MYC-HIS3MX6	This study
YRS2529	MATα his3 Δ 1 leu2 Δ 0 ura3 Δ 0 lys2 Δ SAY1-	This study

This study

- YRS2529MAT α his $3\Delta 1$ leu $2\Delta 0$ ura $3\Delta 0$ lys 2Δ SAY1- This study
MYC-HIS3MX6 hem1::LEU2
- YRS2537MAT α his 3 $\Delta 1$ leu 2 $\Delta 0$ ura 3 $\Delta 0$ lys 2 Δ GAL1- This study
GFP-ATF2-HIS3MX6
- YRS2538MAT α his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ GAL1- This study
GFP-ATF2-HIS3MX6 hem1::LEU2
- YRS2539 $MAT\alpha his3\Delta 1 leu2\Delta 0 ura3\Delta 0 lys2\Delta GAL1$ -GFP-ATF2-HIS3MX6 say1::KanMX4 hem1::LEU 2
- YRS2550MAT α his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ EUROSCARF;
say1::kanMX4Winzeler et al., 1999
- YRS2551MAT α his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ EUROSCARF;
atf2::kanMX4Winzeler et al., 1999
- YRS2655MATa $ade2\Delta 0$ $his3\Delta 1$ $leu2\Delta 0$ $ura3\Delta 0$ $sec12^{ts}$ This study
say1::HIS3MX6 hem1::LEU2 $sec12^{ts}$
- YRS2656 MATα pdr1-3 his1 ura3 yor1::hisG snq2::hisG Nakamura et al., 2001 pdr5::hisG pdr10::hisG pdr11::hisG ycf1::hisG pdr3::hisG pdr15::hisG
- YRS2776 MAT α his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ This study ygr263cT/G757::MYC-HIS3MX6
- YRS2846 $MAT\alpha$ $his3\Delta 1$ $leu2\Delta 0$ $ura3\Delta 0$ $lys2\Delta 0$ This studysay1::HIS3MX6erg4::kanMX4
- YRS2851 $MAT\alpha$ his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ This study atf2::HIS3MX6 erg4::kanMX4
- YRS2857MAT α his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ This study
ygr263cT/G757::MYC-HIS3MX6 hem1::LEU2
- YRS2862 $MAT\alpha$ $his3\Delta 1$ $leu2\Delta 0$ $ura3\Delta 0$ $lys2\Delta 0$ EUROSCARF;erg4::kanMX4Winzeler et al., 1999
- YRS2890 $MAT\alpha$ his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 Δ say1::KanMX4 hem1::LEU2 pYES [URA3]
- YRS2891MAT α his 3 $\Delta 1$ leu 2 $\Delta 0$ ura 3 $\Delta 0$ lys 2 Δ This studysay1::KanMX4 hem1::LEU2
pRecAADACL1::[URA3]This study
- YRS2985 $MAT\alpha$ his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 Δ This studysay1::KanMX4 hem1::LEU2
pRecAADAC::[URA3]This study

- YRS3018 $MAT\alpha$ $his3\Delta 1$ $leu2\Delta 0$ $ura3\Delta 0$ $lys2\Delta 0$ This studyare1::kanMX4are2::kanMX4trp1::URA3
- YRS3019 $MAT\alpha$ his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ This study trp1::URA3
- YRS3052MATahis3 $\Delta 11,15$ leu2 $\Delta 3,112$ trp1 Δ -1This studyHDEL::URA3GFP-SAY1-HIS3MX6
- YRS3100 $MAT\alpha$ $his3\Delta 1$ $leu2\Delta 0$ $ura3\Delta 0$ $lys2\Delta$ This studyare1::kanMX4are2::kanMX4hem1::LEU2
- YRS3101 $MAT\alpha$ $his3\Delta 1$ $leu2\Delta 0$ $ura3\Delta 0$ $lys2\Delta$ This studyare1::kanMX4are2::kanMX4say1::HIS3MX6hem1::LEU2
- YRS3102 $MAT\alpha$ $his3\Delta 1$ $leu2\Delta 0$ $ura3\Delta 0$ $lys2\Delta$ This studyare1::kanMX4are2::kanMX4say1::HIS3MX6
- YRS3275 $MAT\alpha$ his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ This study say1::kanMX4 trp1::URA3
- YRS3276 MAT α his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ This study atf2::kanMX4 trp1::URA3
- YRS3277 MAT α his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 Δ GAL1- This study GFP-SAY1-HIS3MX6 trp1::URA3
- YRS3278 MAT α his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 Δ GAL1- This study GFP-ATF2-HIS3MX6 trp1::URA3
- YRS3279 $MAT\alpha$ his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 Δ This study are1::kanMX4 are2::kanMX4 atf2::HIS3MX6
- YRS3280 MAT α his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 Δ This study are1::kanMX4 are2::kanMX4 atf2::HIS3MX6 hem1::LEU2
- YRS3281 $MAT\alpha$ $his3\Delta 1$ $leu2\Delta 0$ $ura3\Delta 0$ $lys2\Delta 0$ This study are1::kanMX4 are2::kanMX4 atf2::HIS3MX6 trp1::URA3
- YRS3282 $MAT\alpha$ his3 $\Delta 1$ leu2 $\Delta 0$ ura3 $\Delta 0$ lys2 $\Delta 0$ ATF2- This study GFP-HIS3MX6 hem1::LEU2

Supplementary figure legends

Figure S1. Mass spectrometric analysis of cholesterol acetate.

A) say1 Δ mutant cells accumulate an ion with m/z ratio identical to that of cholesterol acetate. Heme-deficient say1 Δ (YRS1853) mutant cells were grown in media containing cholesterol. Lipids were extracted from the cell pellet and analyzed by mass spectrometry. The sodium adduct of cholesterol acetate (428.69 Da) employed as chemical standard gives a major peak at m/z=451.3. say1 Δ mutant cells contain a lipid with identical m/z ratio.

B) The fragmentation profile of the lipid present in $say1\Delta$ mutant cells is identical to that of cholesterol acetate. Fragmentation of the cholesterol acetate (m/z 451.3) reveals a daughter ion of m/z=396.3, which corresponds to a loss of sodium acetate. Fragmentation of the ion at m/z=451.3 in the lipid extract of $say1\Delta$ mutant cells reveals the same daughter ion at m/z=396.3. CA, cholesterol acetate.

Figure S2. Export of cholesterol acetate is stimulated by the presence of ergosterol in the media.

Heme-deficient *say1* Δ (YRS1853) mutant cells were labeled with [¹⁴C]cholesterol for 16 h, diluted into fresh media containing the indicated concentration of cold ergosterol and cultivated for 24 h. Lipids were extracted from the cell pellet and the culture media, analyzed by TLC and quantified. The proportion of cholesterol acetate (CA), free cholesterol (FC), and steryl esters (STE) that is exported into the culture media is plotted as a function of ergosterol concentration in the media.

Figure S3. Formation of cholesterol acetate is independent of Are1 and Are2, but efficient export of cholesterol acetate requires long-chain steryl esters.

A) Formation of cholesterol acetate is independent of Are1 and Are2. Hemedeficient wild-type (YRS1849), $say1\Delta$ (YRS1853), $atf2\Delta$ (YRS2135), $are1\Delta$ $are2\Delta$ (YRS3100), $are1\Delta$ $are2\Delta$ $say1\Delta$ (YRS3101), and $are1\Delta$ $are2\Delta$ $atf2\Delta$ (YRS3280) mutant cells were labeled with [¹⁴C]cholesterol for 16 h. Lipids were extracted, separated by TLC and visualized using a phosphorimager. FC, free cholesterol; CA, cholesteryl acetate; STE, steryl ester.

B) Export of sterol acetate is strongly reduced in cells lacking Are1 and Are2. Heme-deficient *say1* Δ (YRS1853), *are1* Δ *are2* Δ (YRS3100), and *are1* Δ *are2* Δ *say1* Δ (YRS3101) mutant cells were labeled with [¹⁴C]cholesterol for 16 h, diluted into fresh media containing cold ergosterol and cultivated for 6 h. Lipids were extracted from the cell pellet (intracellular) and the culture media (extracellular) and analyzed by TLC. FC, free cholesterol; CA, cholesteryl acetate; STE, steryl ester. The open star indicates an unidentified secreted sterol derivative.

Figure S4. Say1 or Atf2 are not essential in cells that form no long-chain steryl esters.

Deletion of either Say1 or Atf2 in an *are1* Δ *are2* Δ mutant background does not impair growth. Heme-competent and heme-deficient wild-type (YRS1533, YRS1849), *say1* Δ (YRS2550, YRS1853), *atf2* Δ (YRS2551, YRS2135), *are1* Δ *are2* Δ (YRS2486, YRS3100), *are1* Δ *are2* Δ *say1* Δ (YRS3102, YRS3101), and *are1* Δ *are2* Δ *atf2* Δ (YRS3279, YRS3280) mutant cells were serially diluted 10fold and spotted on YPD plates or media supplemented with cholesterol and Tween (Chol/Tween) or with ALA. Plates were incubated at 30°C for 4 d.

Figure S5. Substrate specificity of the sterol acetylation and export pathway.

Heme-deficient wild-type (YRS1849), *say1* Δ (YRS1853), *atf2* Δ (YRS2135), and *are1* Δ *are2* Δ (YRS3100) mutant cells were labeled with either [¹⁴C] β -sitosterol, [¹⁴C]progesterone, [¹⁴C]25-hydroxycholesterol, [³H]7-ketocholesterol, or [³H]lanosterol for 16 h. Lipids were extracted from the cell pellet (intracellular) and the culture media (extracellular), separated by TLC and visualized using a phosphorimager. FSito, free sitosterol; STE, steryl ester; FProg, free progesterone; OH-CA, hydroxycholesteryl acetate; FOH-C, free hydroxycholesterol; FKC, free 7-ketocholesterol; FL, free lanosterol.

Figure S6. Expression of Say1 and Atf2 is independent of oxygen availability.

Heme-deficient cells expressing chromosomally tagged versions of Say1-myc (YRS2529) or Atf2-GFP (YRS3282) were cultivated in YPD media containing either cholesterol/Tween or ALA. Cells were diluted to an OD of 0.5 and samples were removed after growth for the time indicated, proteins were extracted, separated by electrophoresis, and probed with antibodies against myc or GFP. Phosphoglycerate kinase (Pgk1) was used as a loading control.



Figure S1

В

Α



Figure S2



heme-deficient





wι say1Δ atf2Δ are1Δ are2Δ are1Δ are2Δ say1Δ are1Δ are2Δ atf2Δ

Figure S4

Figure S5







Figure S6