

This document is the unedited author's version of a Submitted Work that was subsequently accepted for publication in *Environmental Science & Technology*, copyright © American Chemical Society after peer review. To access the final edited and published work see <http://pubs.acs.org/doi/abs/10.1021/es901612v>.

1 **The Influence of Operating Parameters on the Biodegradation of Steroid Estrogens**
2 **and Alkylphenolic Compounds during Biological Wastewater Treatment Processes**

3 Yoong K.K. Koh¹, Tze Y. Chiu² Alan R. Boobis³, Mark D. Scrimshaw⁴, John P. Bagnall⁵,
4 Ana Soares⁵, Simon Pollard⁵, Elise Cartmell^{5*} and John N. Lester⁵

5

6 ¹Public Utilities Board, Technology and Water Quality Office, 40 Scotts Road #15-01,
7 Environment Building, 228231, Singapore.

8 ²Earth Tech Engineering Limited, Wentworth Business Park, Tankersley, Barnsley, S75
9 3DL, UK.

10 ³Faculty of Medicine, Division of Experimental Medicine and Toxicology, Imperial
11 College London, Hammersmith Campus, London, W12 0NN, UK.

12 ⁴Institute for the Environment, Brunel University, Uxbridge, Middlesex, UB8 3PH, UK

13 ⁵Centre for Water Science, School of Applied Sciences, Cranfield University,
14 Bedfordshire, MK43 0AL, UK.

15

16 *Corresponding author:

17

18 **Abstract.** This study investigated operational factors influencing the removal of steroid
19 estrogens and alkylphenolic compounds in two sewage treatment works, one a
20 nitrifying/denitrifying activated sludge plant and the other a nitrifying/denitrifying
21 activated sludge plant with phosphorus removal. Removal efficiencies of >90% for
22 steroid estrogens and for longer chain nonylphenol ethoxylates (NP₄₋₁₂EO) were observed
23 at both works, which had equal sludge ages of 13 days. However, the biological activity
24 in terms of milligrams of estrogen removed per tonne of biomass was found to be 50-60%
25 more efficient in the nitrifying/denitrifying activated sludge works compared to the works
26 which additionally incorporated phosphorus removal. A temperature reduction of 6°C
27 had no impact on the removal of free estrogens, but removal of the conjugated estrone-3-
28 sulphate was reduced by 20%. The apparent biomass sorption (LogK_p) values were
29 greater in the nitrifying/denitrifying works than those in the nitrifying/denitrifying works
30 with phosphorus removal for both steroid estrogens and alkylphenolic compounds
31 possibly indicating a different cell surface structure and therefore microbial population.
32 The difference in biological activity (mg tonne⁻¹) identified in this study, of up to seven

33 times, suggests that there is the potential for enhancing the removal of estrogens and
34 alkylphenols if more detailed knowledge of the factors responsible for these differences
35 can be identified and maximised, thus potentially improving the quality of receiving
36 waters.

37

38 **Introduction**

39 Natural and synthetic estrogens and non-ionic surfactants such as alkylphenol
40 polyethoxylates (APEOs) are endocrine disrupting chemicals (EDCs) that can cause
41 adverse effects on the sexual and reproductive systems in wildlife and fish (1,2). The
42 effluents discharged from sewage treatment works (STWs) are major sources of these
43 anthropogenic chemicals to the aquatic environment (3). In addition, APEOs biodegrade
44 during wastewater treatment to generate the parent alkylphenols (AP), octylphenol (OP)
45 and nonylphenol (NP), the shorter chain mono to triethoxylates (NP₁EO, NP₂EO and
46 NP₃EO) and a range of carboxylated intermediate by-products (4,5) which are more
47 estrogenic than their parent substances (5-8). In the aquatic environment these
48 compounds are amenable to further biotransformation and bioconcentration (9) and may
49 potentially bioaccumulate (10); as a consequence of this behaviour complex issues for
50 environmental health arise (2). While secondary biological treatment of wastewater
51 significantly reduces the concentration of some of these compounds, as presently
52 configured and operated, these processes cannot afford adequate protection of the aquatic
53 environment (11). Regulatory authorities are seeking to reduce and ultimately eliminate
54 this problem. In the UK, a £40 million (\$75 million) National Demonstration Program
55 (NDP) has been undertaken by the water industry as part of the asset management plan
56 four (AMP4) settlement, initiated by the Environment Agency (EA) of England and
57 Wales to investigate the potential removal of steroid estrogens from final effluents (12).
58 An initial report from the study has concluded that STWs, where treatment involved
59 nitrifying activated sludge, were able to remove steroid estrogens more effectively than
60 those with other biological treatment processes (13).

61

62 The primary objective of conventional wastewater treatment is the removal of carbon and
63 nitrogen and possibly phosphorus (14), hence current configurations are not designed or
64 operated to remove EDCs (15-17). Tertiary treatment technologies such as granular
65 activated carbon (GAC), advanced oxidation processes (AOPs), and membrane filtration
66 have been suggested to remove these micropollutants (18,19). However, the presence of

67 high levels of insoluble and dissolved organic matter may interfere with the adsorption
68 process and could therefore result in lower than anticipated removals when GAC is used
69 (20) The same issue also means that employment of AOPs may require high doses of
70 oxidants, thus resulting in increased cost (21). Undoubtedly, advanced treatment
71 technologies will remove these compounds and ameliorate the impact of EDCs on surface
72 waters, but they will inevitably result in significant financial and environmental costs
73 through increased energy consumption and carbon dioxide emissions (21). Environmental
74 sustainability therefore requires the consideration of alternative strategies such as
75 optimisation or modification of existing STWs by determining the operating parameters
76 that govern the removal of these substances within the STW.

77

78 Several studies have attempted to link certain operating parameters and the removal of
79 EDCs in STWs. In activated sludge systems, solid retention time (SRT) (22-25) and
80 hydraulic retention time (HRT) (3,26) have both been proposed as factors which may
81 regulate EDC removal, however, explicit information on their precise mode of effect is
82 lacking. There is also no conclusive evidence on the significance of other variables, such
83 as temperature, partitioning to solids and dissolved oxygen concentrations that would
84 inform decisions on the optimisation of STWs for the removal of these chemicals (11,27).

85

86 This study was undertaken to determine the role of operating parameters in the removal
87 and biodegradation of selected steroid estrogens, APEOs and their metabolites in two
88 treatment processes: a nitrifying/denitrifying activated sludge plant (N/DN) and a
89 nitrifying/denitrifying activated sludge plant with phosphorus (P) removal (N/DN-P).
90 Such treatment processes are frequently installed at large urban STW where discharges
91 contribute significantly to flows in receiving waters, which are therefore likely to be
92 impacted by discharges of EDCs. The objective of this study was to compare the
93 biological activity of each process, and investigate factors that may influence it, such as
94 organic loading, temperature and dissolved oxygen concentration.

95

96 **Materials and Methods**

97 **Sewage Treatment Works.** Samples were collected at appropriate stages of the
98 biological treatment processes (after primary sedimentation) at two full-scale STWs
99 (Figure 1), the characteristics of which are described in Table 1. Both STWs were
100 required to nitrify in order to comply with effluent ammonia requirements and the N/DN-

101 P works had an additional anaerobic zone for biological P removal. Chemical
102 precipitation (ferrous) was used following secondary treatment to further reduce final
103 effluent phosphorus concentrations in the N/DN-P works.

104

105 **Sampling Regime.** Three separate sampling campaigns were undertaken: at the N/DN
106 works in Summer 2004 and Winter 2006; and finally at the N/DN-P works in Summer
107 2006. Discrete samples were collected in 2.5 l amber borosilicate glass vessels with
108 Teflon lined caps from 08:00 on a Monday morning through to 12:00 on Friday. The
109 maximum interval between samples was 4-6 hours depending on the retention time of the
110 unit processes, for practical reasons such as health and safety and accessibility to the
111 sampling points. The samples were not preserved as they were extracted onto solid phase
112 extraction (SPE) cartridges on site within 15 minutes of collection. Sampling frequency
113 was such, that in conjunction with the average daily flows, representative mass balances
114 could be calculated. The monitoring programme allowed for coverage of diurnal
115 (day/night) variation, seasonality (winter/summer) and process type (N/DN and N/DN-P).
116 Little or no precipitation (rain or snow) was experienced during any sampling period.
117 Samples were taken from the settled sewage leading to the activated sludge units, the
118 returned activated sludge (RAS), the final effluents and at the N/DN-P works only, the
119 liquors from sludge thickening treatment containing volatile fatty acids (VFAs) (Figure
120 1).

121

122 **Analytical Procedure for Steroid Estrogens and Alkylphenolic Compounds.** Steroid
123 estrogens and APEOs were determined in the dissolved and adsorbed phases in all
124 samples. Methodology for the determination of the natural and synthetic steroid estrogens:
125 estrone (E1); 17 β -estradiol (E2); estriol (E3); sulphate conjugate of estrone (E1-3S); and
126 17 α -ethinylestradiol (EE2) in the dissolved phase (28) and on solids (29) has previously
127 been reported. Estrogens are predominantly excreted as either glucuronide or sulphate
128 conjugates, although only estrone 3-sulfate (E1-3S) has been detected in UK sewages
129 (30), probably as a result of the predominance of this conjugate in urine and the rapid
130 deconjugation of the glucuronides (31). Therefore, only this conjugate was analysed. The
131 alkylphenolic compounds: alkylphenols, alkylphenol polyethoxylates (APEO) and
132 alkylphenol ethoxycarboxylates (APEC) in the dissolved phase were determined by the
133 method of Koh et al. (32). Methodology for the determination of these compounds on the
134 solid phase, along with full descriptions of all methods and their performance are

135 provided in the supplementary information. In summary, 1 l sewage samples for both
136 steroid estrogens and APEOs were filtered through glass fibre GF/C filters (VWR
137 International, Leicestershire, UK) prior to solid phase extraction on separate tC18 SPE
138 cartridges. For the dissolved phase, steroid estrogens were extracted from the tC18
139 cartridges followed by two further sample clean-up stages and quantification using
140 LC/ESI(-)/MS/MS. The alkylphenolic compounds were eluted from the tC18 cartridges
141 and without further clean-up, quantified using LC/MS/MS. The adsorbed and sludge
142 samples for steroid estrogens and APEOs were freeze dried then solvent extracted and
143 subjected to clean-up (three stage for estrogens and single stage for APEO) before
144 quantification by LC/MS/MS.

145

146 **Mass Balance and Biomass Activity Calculations.** Mass balances were completed by
147 multiplying average daily steroid estrogen concentrations (E1, E2, E3, E1-3S and EE2) or
148 alkylphenolic concentrations (NPEO, NPEC and NP) by average daily flows and utilising
149 these values to calculate an average daily flux. The removal efficiencies of the biomass
150 were evaluated by activity i.e. milligram steroid estrogen degraded per tonne of biomass
151 for each estrogen individually and for the sum of all steroid estrogens (Σ EST). The
152 degradation was obtained from the flux data, and was the mass of each compound which
153 entered the biological treatment stage and was not accounted for through analysis of
154 effluent or RAS and was assumed to be degraded. For the alkylphenolic compounds
155 activity was calculated for each alkylphenolic compound and for the groups NP₁₋₃EC and
156 NP₄₋₁₂EO. The calculation was based on the mass difference between the settled sewage
157 and the final effluent in milligrams of estrogens or alkylphenolic and dividing it by the
158 mass of the mixed liquor volatile suspended solids (MLVSS) (tonnes).

159

160 **Results and Discussion**

161 It has been postulated that sludge age, also referred to as solid retention time (SRT), and
162 hydraulic retention time (HRT) are both key factors in the removal of EDCs in biological
163 wastewater treatment processes (11,27,33-35). However, Joss et al. (34) alluded to the
164 fact that SRT only explained part of the difference in removal efficiency. Sludge loading
165 was suggested as a key parameter due to the potential for competitive substrate inhibition
166 limiting estrogen biodegradation, although to date, no clear relationship has been
167 established, suggesting that other parameters may also be involved. In this study the

168 works examined had equivalent SRTs and HRTs but varying sludge loadings as measured
169 by the food to microorganism ratio (F:M) ($\text{g BOD} \cdot \text{g}^{-1} \text{MLVSS} \cdot \text{d}^{-1}$) (Table 1).

170

171 **The Impact of Process Type and Operational Parameters on EDC Degradation.**

172 Based on the mass fluxes, degradation of estrogens was 70 - 76% in both the N/DN and
173 N/DN-P works indicating no difference in removal efficiency. The biological degradation
174 efficiencies for NPEOs were lower with 41% and 55% of the flux entering the biological
175 treatment stage degraded in the N/DN and N/DN-P works respectively in 2006 (Figure 2).
176 These NP, NPEC and NPEO data were comparable to that observed by Loyo-Rosales et
177 al. in a nitrifying activated sludge plant (plant 3) (36) with 59% of influent ($\text{NP}_{0-16}\text{EO}$)
178 degraded with production of the NP_{1-4}EC .

179

180 The total steroid estrogen load (dissolved and adsorbed) in both STWs decreased by
181 almost 1 order of magnitude during treatment from 1806-5508 mg d^{-1} in the settled
182 sewage influents to 117-375 mg d^{-1} in the final effluents. Carballa et al. (37) also
183 performed a similar mass balance over the secondary treatment process, albeit for the
184 combination of E1+E2 only, with 497 mg d^{-1} in the settled sewage and 325 mg d^{-1} in the
185 final effluent from a 100,000 population equivalent (PE) STW (37). In this present study,
186 the mass of E1+E2 in the settled sewage ranged from 757-3859 mg d^{-1} and in final
187 effluents from 66 mg d^{-1} to 291 mg d^{-1} , indicating that significantly more biodegradation
188 was occurring in the STWs in this study. The negative removal efficiencies for E1
189 observed by Carballa et al. were suggested to be due to conversion of E2 to E1 which was
190 then more slowly degraded during secondary treatment (38). In this present study, higher
191 removals were observed for E1, with degradation occurring during biological treatment in
192 both the N/DN and N/DN-P works. It can only be hypothesized that the lack of E1
193 degradation in the study by Carballa et al. was potentially due to the low SRTs (38).
194 Kreuzinger et al. proposed that higher SRTs (e.g. in works with nitrification) allowed the
195 enrichment of slowly growing bacteria and consequently the establishment of a more
196 diverse biocoenosis with broader physiological capabilities and greater potential for EDC
197 removal compared to STWs with low SRTs (35). At the works studied by Kreuzinger et
198 al. the E1+E2+E3 mass balance removals varied from: 16% with a SRTs of <1 day; 66%
199 with a SRT of 9.6 days; 98% with a SRT of 24 days. The removal efficiencies in this
200 study support these findings and were between 78 – 80% in both the N/DN and N/DN-P
201 works for E1+E2+E3 which both had equivalent SRTs of up to 13 days (Table 1).

202

203 The total NPEO load (dissolved and absorbed) in both STWs in 2006 also decreased by
204 nearly 1 order of magnitude from 330486 - 646209 mg d⁻¹ in the settled sewage influents
205 to 54910 – 112924 mg d⁻¹ in the final effluents (Figure 3). At the N/DN works in 2004,
206 similar reductions in the total NPEO load was observed from 1787390 mg d⁻¹ to 83630
207 mg d⁻¹ (39) with removal efficiencies of 93 – 96% observed for NP₄₋₁₂EO (Table 2).
208 Formation of NP₂₋₃EO was observed at both the N/DN and N/DN-P works in 2006 which
209 reduced the overall removal efficiencies to 73 – 91 % for NP₁₋₁₂EO, below the 99.1% and
210 93.7% removals reported by Loyo-Rosales et al. during summer and winter periods
211 respectively for NP₀₋₁₆EO (36). The increased biodegradation (95%) observed in 2004
212 (Figure 2) may be a reflection of the higher concentrations of NPEO detected in 2004
213 compared to 2006 or the higher sewage temperature in the summer of 2004 of 18°C
214 compared to 12°C in the winter of 2006. The low concentrations of total NPEO detected
215 in the return activated sludge (RAS) in 2004 were probably because NPEC compounds
216 were not determined or included in the mass balance. In 2006, the total NPEC mass in the
217 RAS was 71141 mg d⁻¹ in the N/DN works which was approximately equivalent to the
218 NP and NPEO concentration of 69082 mg d⁻¹. If the 2006 proportions of NPEC to NP
219 and NPEO were applied across the 2004 mass balance the biodegradation would be
220 reduced to 88% from 95%.

221

222 The NPEC compounds comprised about 50% of the total alkylphenolic compounds in the
223 final effluent compared to <5% in the settled sewage, with NP₁₋₃EC exhibiting an
224 increase in concentration from <1 µg l⁻¹ in the settled sewage to 1.3 and 2 µg l⁻¹ in the
225 N/DN and N/DN-P works respectively..

226

227 **Evaluation of Biomass Activity as a Determining Factor on EDC Removal.** Both the
228 removal efficiencies and final effluent concentrations given in Table 4 at the two STWs
229 were similar. However, the removal efficiencies of the biomass at the STWs were also
230 evaluated by activity i.e. milligram steroid estrogen or NPEO biodegraded per tonne of
231 biomass. The two STWs were equally efficient in removing organics with removal of
232 chemical oxygen demand (COD) being >82% (Table 1), however, the biomass activity
233 was different. The food to micro-organism (F:M) ratio in the N/DN works was twice that
234 of the N/DN-P works with 0.1 g BOD. g⁻¹ MLVSS d⁻¹ and 0.05 g BOD. g⁻¹ MLVSS d⁻¹
235 respectively. The overall biomass activity per tonne of steroid estrogen removed was

236 highest in the N/DN works in 2004 at 116.7 mg tonne⁻¹ and lowest in the N/DN-P works
237 at 39.4 mg tonne⁻¹. This difference in efficiency of the biomass was also observed in
238 relation to the removal of NPEO, with an activity of 11977 mg NP₄₋₁₂EO tonne⁻¹ in the
239 N/DN works and 4221 mg tonne⁻¹ in the N/DN-P works (Table 2). It is apparent that
240 these results do not necessarily support the hypothesis that higher organic loadings, as
241 measured by the F:M ratios, result in the inhibition of steroid estrogen biodegradation
242 (24). This is assuming that the influent inert non-degradable material is consistent
243 between the works. Furthermore, although the MLVSS concentrations on both sampling
244 occasions at the N/DN works were much lower than the N/DN-P works, steroid estrogen
245 biodegradation based on removal in mg tonne⁻¹ of biomass, was higher for the N/DN
246 works.. It is hypothesized that the biomass in the N/DN works was different to that in the
247 N/DN-P works. The apparent LogK_p values also infer possible differences between the
248 biomass at the two works. The generally lower LogK_p values determined for all
249 compounds at the N/DN-P works compared with the N/DN works (Table 3), are
250 indicative of different absorption capacities of the biomass, as it is known that some
251 genera of bacteria are far more hydrophobic than others, and that the proportional
252 abundance hydrophobic genera increases with sludge age (Davenport et al., 2000)

253

254 The greater biological activity observed at the N/DN works does not support the
255 hypothesis that the varied environmental conditions with respect to redox (aerobic,
256 anoxic and anaerobic) present in the N/DN-P works provides a more diverse bacterial
257 community with more complex biochemistry which can potentially enhance the
258 biodegradation of EDCs (34). Therefore, it appears that examining the correlation
259 between SRT and EDC removal efficiencies, although useful in predicting if removals
260 can occur, does not provide a true representation of the biomass activity and propensity
261 for EDC removal.

262

263 It has been postulated that increasing the sludge age increases the diversity of the
264 consortia of bacteria present in a treatment plant allowing the growth of EDC degrading
265 organisms (40) and that the ability to remove EDCs is assumed to be a property of some
266 of the slower growing organisms that can only colonise the treatment plant at long sludge
267 ages. It is unlikely that the presence of EDCs are specifically selecting for these
268 organisms, as the low concentrations of EDC found in wastewaters could only support a
269 small number of cells. It is more likely that EDC degradation occurs fortuitously in

270 organisms scavenging a wide range of carbon sources and there is recent evidence that
271 the primary mechanism for EE2 degradation in STWs is more likely to be due to the
272 activity of heterotrophic bacteria than ammonia oxidising bacteria (41). Heterotrophic
273 organisms that efficiently scavenge low concentrations of a resource are sometimes
274 referred to under the descriptive population term of “K strategists” (42). “K strategists”
275 have a high affinity for resources (i.e. a low Monod half saturation coefficient) and low
276 growth rates (i.e. a low μ_{\max}), a property consistent with the long sludge ages required for
277 degradation and utilisation of low concentrations of EDCs.

278

279 **Influence of Temperature on the Biodegradation of Steroid Estrogens.** The
280 evaluation of removal based on biomass activity established that there were differences
281 between the N/DN and N/DN-P works, postulated to be due to variations in the
282 establishment of a more diverse biocoenosis. Therefore, because removal efficiencies of
283 organic micropollutants are known to be more sensitive to temperature than removal of
284 biochemical oxygen demand (BOD) and suspended solids (SS) (43,44), the effect of
285 temperature was evaluated by undertaking a further study of the N/DN works during the
286 winter for steroid estrogens. Recorded sewage temperatures were 18°C (summer) and
287 12°C (winter) with corresponding air temperatures of 21°C and 6°C. The 6 °C reduction in
288 sewage temperature did not have an effect on the removal of free estrogens, which was
289 consistent with other studies which have observed minimal impact of temperature on the
290 removal of unconjugated estrogens (e.g. E2) (16,45,46).

291

292 However, the 6°C reduction in temperature did effect the removal of the conjugate E1-3S
293 which was ~20% lower in the N/DN works in winter (59%±6) compared to summer
294 (78%±4). Hence it appears that deconjugation was inhibited by the reduction in
295 temperature, rather than the biodegradation of the deconjugated moiety E1. This is
296 consistent with the observation by D’Ascenzo et al. who reported removal of E1-3S at 64%
297 in six activated sludge plants in Rome during the Autumn, although values for the
298 temperature of either sewage or air temperature were reported (47). It could be postulated
299 that the low removal of E1-3S was probably due to the low activity of arylsulphatase
300 enzymes (caused by the low temperature) or the absence of bacteria containing these
301 enzymes during the cold season (48).

302

303 **Impact of Nitrification on the Biodegradation of Steroid Estrogens.** Nitrification
304 activity has been reported to be correlated with estrogen removal (49). Both works in this
305 study fully nitrified and therefore dissolved oxygen (DO) was not thought to directly
306 influence EDC biodegradation in this study. It has been demonstrated that degradation of
307 steroid estrogens is associated with the co-metabolism of the ammonia oxidizing bacteria
308 in nitrifying activated sludge (50) which may dominate nitrifying plants such as those
309 sampled in this study. However, there is strong evidence that cometabolic degradation of
310 EE2 by ammonia oxidising bacteria is not an important removal mechanism in STWs
311 (41). Therefore, the difference in removal of EDCs between the N/DN and N/DN-P
312 works was probably not due to the biochemical activity of ammonia oxidizing bacteria,
313 but may result from the metabolic activity of heterotrophic organisms able to utilise
314 resources present at low concentrations, such as the “K strategists” (42) which remains to
315 be established.

316

317 **Partitioning of EDCs to Particulate Matter.** To determine the significance of sorption
318 in the removal process, the distribution of the estrogens and NPEOs between the solid
319 and liquid phase in the mixed liquor was evaluated by calculating the apparent partition
320 coefficient (K_p). This was undertaken to confirm EDC removal mechanisms (sorption or
321 biodegradation) and to establish if there were any differences between the N/DN and
322 N/DN-P works. The observed $\text{Log}K_p$ values for E1 and E2 in the N/DN works in both
323 summer (2004) and winter (2006) were above those observed in the N/DN-P works in
324 summer (2006) (Table 3). These $\text{Log}K_p$ values were in the same range as $\text{Log}K_d$ values
325 for E1, E2 and EE2 reported by Carbella et al. (37); Joss et al. (34) and Ternes et al. (52).
326 There is an indication in the data presented in Table 3 that overall for steroid estrogens,
327 with the exception of E1-3S in the summer of 2004, that apparent $\text{Log}K_p$ values were
328 greater in the N/DN works than those in the N/DN-P works. This is supported by the
329 NPEO data with adsorption to solids also being more important in the N/DN works (42%)
330 compared to the N/DN-P works (28%) (Figure 2). This is reflected in the apparent $\text{Log}K_p$
331 with higher values observed for each alkylphenol group for the N/DN biomass (1.4-3.2 l
332 kg^{-1}) compared with (0.05-1.4 l kg^{-1}) the N/DN-P biomass (Table 3).

333

334 Results for the two STWs examined in this study demonstrated that in the settled sewage
335 20-30% of E1, E2 and EE2 were associated with suspended solids, however, for the more
336 hydrophilic E3 and E1-3S this decreased to around 10%. This is in agreement with results

337 from studies using radiolabelled E2 to determine the fate of estrogens in STWs (51)
338 which found that at low concentrations, the majority of the radiolabelled E2 remained in
339 the liquid phase and did not adsorb to the solids. Therefore, biodegradation appears to be
340 the dominant removal pathway for steroid estrogens, as demonstrated in Figure 2, where
341 mass balance calculations indicate that $\geq 70\%$ of the total steroid estrogens were
342 biodegraded. In contrast adsorption to solids was a more significant for NPEOs with
343 biodegradation observed at $\geq 41\%$.

344

345 **The Distribution of EDCs in Settled Sewage.** At the N/DN works, which had
346 approximately double the retention time in the sewerage system (13 hours) (based on
347 Water Utility design information), higher concentrations of E3 than E1 were observed in
348 the influent and settled sewage. At the N/DN works, the E3:E1 ratio was 1.33 (2004) and
349 1.69 (2006) compared to 0.42 for the N/DN-P works influent (Figure 4). Deconjugation
350 of the conjugated estrogens in the time taken for the sewage to reach the works was
351 clearly demonstrated by the detection of the unconjugated estrogens E1, E2, EE2 and E3
352 in the settled sewage. This finding was expected as it has already been reported that the
353 deconjugation of glucuronide conjugates may occur in the sewerage system, while
354 cleavage of the sulphonated conjugates, which require arylsulphatase for cleavage, will
355 only occur in the STWs as this demands more specialized micro-organisms (53,54). This
356 observation corroborates that of D'Ascenzo et al. who concluded that unconjugated
357 estrogens and sulphated (not glucuronide) estrogens were the dominant species in the
358 influent of STWs (47). This study further confirms the importance of this conjugated
359 hormone since it inevitably contributes to the overall estrogenic burden leading to the
360 release of E1 as a consequence of the hydrolysis of the sulphate conjugate.

361

362 **The Distribution of EDCs in Final Effluents.** The concentration of E1 in the final
363 effluents in this study ranged from 4.3 to 5.5 ng l⁻¹ (N/DN 2004/06 and N/DN-P 2006)
364 (Table 4) which is in agreement with concentrations (low nanogram per litre) reported in
365 other countries: Italy 9.3 ng l⁻¹ (53); the Netherlands 4.5 ng l⁻¹ (54); and Canada 3 ng l⁻¹
366 (24). Recent work from France (55) has reported E1 concentrations ranging from <1 up
367 to 75 ng l⁻¹ (median 5.3 ng l⁻¹), although concentrations of E2 in the effluent were lower,
368 with a median of 1 ng l⁻¹, in good agreement with values reported in Table 4 (0.4-1.1 ng l⁻¹).
369 The N/DN-P works did not achieve the proposed requirement (EEq<1) for all the

370 compounds whilst the N/DN works (both seasons) was within the Predicted No-Effect
371 Concentration (PNEC) value for E2 and EE2 . The combined PNEC value of <1 EEq was
372 not achieved at either STWs (Table 4) (56).

373

374 In contrast the final effluent concentrations of NP were below the PNEC value of 330 ng
375 l^{-1} (57) with 44 ng l^{-1} and 55 ng l^{-1} in the N/DN and N/DN-P works respectively. This
376 was in agreement with another study where concentrations of 50-300 ng l^{-1} were
377 observed in the final effluent (58) but lower than the median value of 1649 ng l^{-1} reported
378 in 2003 from a number of further STWs final effluents (59).

379

380 The use of tertiary treatment processes (GAC, Ozone, Membrane Filtration) are currently
381 being evaluated to assess their ability to achieve PNEC values (12). However, all of these
382 processes come at a high environmental and economic cost (21). It would therefore, be
383 highly desirable to operate secondary biological treatment processes to achieve an
384 environmental sustainable solution for EDC removal. The difference in specific biomass
385 activity identified in this study, does suggest that there is the potential for enhancement of
386 EDC removal by biological wastewater treatment. If more detailed knowledge of the
387 factors responsible for these differences can be identified it may allow for
388 maximising .removal during the treatment process.

389

390 **Acknowledgements**

391 One of the authors (Y.K.K. Koh) is grateful to the Public Utilities Board of Singapore for
392 the award of a PhD scholarship. The authors would like to thank the following companies:
393 Anglian Water Ltd, Severn Trent Water Ltd, Thames Water Utilities Ltd, United Utilities
394 Plc and Yorkshire Water Services Ltd for providing their support and funding. In addition
395 we would like to acknowledge those who have assisted with fieldwork: Joe Peters from
396 Anglian Water, Angela Barugh from Thames Water, Carlos Constantino, Nafsika Ganidi,
397 Emma Goslan, Pantelis Kampas, Ewan McAdam, Davide Minervini, Dan Murray, Nick
398 Paterakis, Florine Sternenberg, Andrew Thornton and Xenofon Varidakis from Cranfield
399 and Brunel Universities. Finally, Dan McMillan at Waters Ltd. for analytical support.

400

401 **Brief**

402 Biodegradation of steroid estrogens and alkylphenolic compounds was examined
403 with >90% removal and 50-60% greater biological activity observed in a

404 nitrifying/denitrifying activated sludge plant compared to a nitrifying/denitrifying
405 activated sludge plant with phosphorus removal..
406

Table 1. Overview of the operating parameters of the two sewage treatment works

Operating parameters	N/DN		N/DN-P
	Summer 2004	Winter 2006	Summer 2006
Biological process	Nitrifying/ denitrifying	Nitrifying/ denitrifying	Nitrifying/denitrifying /P-removal
Process technology	Anoxic/Aerobic	Anoxic/Aerobic	Anoxic/Anaerobic/Aerobic
PE	150,000	150,000	250,323
Q activated sludge process ($\text{m}^3 \text{d}^{-1}$)	12000	17200	44000
HRT θ_r (h)	13.6 (0.6d)	10.2 (0.4d)	12.1 (0.5d)
SRT θ_c (d)	13	13	9 – 13
DO (g m^{-3})	1.4	3.2	1.8
MLVSS (g m^{-3})	2740	3282	4971
F:M ratio (g BOD. g^{-1} MLVSS.d^{-1})	0.09	0.1	0.05
pH	7-7.5	7-7.6	7.2-7.4
Trade input	<1%	<1%	10%
Ambient ($^{\circ}\text{C}$)	21	6	27
Sewage ($^{\circ}\text{C}$)	18	12	22
Settled Sewage Influent Characteristics			
COD (g m^{-3})	252	286	489
BOD (g m^{-3})	151	141	148
$\text{NH}_4\text{-N}$ (g m^{-3})	34.5	38	37
$\text{NO}_3\text{-N}$ (g m^{-3})	3	3.2	2.5
P (g m^{-3})	n/d	n/d	9
TSS (g m^{-3})	266	122	118
Final Effluent Characteristics			
COD (g m^{-3})	40.1	51	30
BOD (g m^{-3})	4	16	11
$\text{NH}_4\text{-N}$ (g m^{-3})	<1	0.4	<0.2
$\text{NO}_3\text{-N}$ (g m^{-3})	28.2	31.9	15
P (g m^{-3})	n/d	n/d	<0.03
TSS (g m^{-3})	8	36	8

KEY: PE – population equivalent; Q - total flow; HRT – hydraulic retention time; SRT – solids retention time; DO – dissolved oxygen; MLVSS – mixed liquor volatile suspended solids; F:M food to micro-organism ratio; COD – chemical oxygen demand; BOD – biological oxygen demand; $\text{NH}_4\text{-N}$ – ammoniacal nitrogen; $\text{NO}_3\text{-N}$ – nitrate nitrogen; P – orthophosphate; TSS – total suspended solids. The works MLVSS, COD, BOD, $\text{NH}_4\text{-N}$, $\text{NO}_3\text{-N}$, P, TSS values are from daily duplicate samples averaged over the 5 day sampling period. Total flow, DO, temperature, pH were daily averages from on-line continuous recorders. The DO set point was 1.5 mg l^{-1} for both works. The variation was $\pm 1 \text{ mg l}^{-1}$ for the N/DN works and $\pm 3 \text{ mg l}^{-1}$ for the N/DN-P works.
n/d = not determined

Table 2. Biomass activity (mg tonne⁻¹) and removal efficiency (%) of steroid estrogens and alkylphenolic compounds in secondary treatment

Steroid estrogens	Biomass activity (mg tonne ⁻¹) and removal efficiency (%) ^{ab}					
	N/DN			N/DN-P		
	2004		2006	2006		2006
E1	28.8	(89)	34.2	(89)	20.7	(91)
E2	5.4	(94)	11.4	(96)	6.7	(94)
E3	41.7	(99)	62.7	(99)	9.2	(98)
EE2	0.7	(68)	0.8	(65)	0.3	(60)
E1-3S	6.9	(78)	7.6	(59)	2.5	(88)
∑EST	83.5	(93)	116.7	(92)	39.4	(92)
Loading (mg ∑EST m ⁻³ d ⁻¹)	0.25		0.41		0.21	
Alkylphenolic compounds	c	c				
NP	115	(30)	-19	(-12)	11	(11)
NP ₁ EC	n/d	n/d	-167	(-136)	-125	(-241)
NP ₂ EC	n/d	n/d	-611	(-163)	-59	(-52)
NP ₃ EC	n/d	n/d	-13	(-211)	-10	(-594)
NP ₁ EO	23	(80)	263	(77)	71	(41)
NP ₂ EO	497	(68)	-22	(-160)	-4	(-107)
NP ₃ EO	4021	(85)	-28	(-460)	-5	(-190)
NP ₄ EO	7368	(92)	180	(78)	103	(86)
NP ₅ EO	2977	(95)	519	(88)	241	(88)
NP ₆ EO	6284	(96)	1715	(92)	653	(90)
NP ₇ EO	9470	(96)	2120	(95)	736	(92)
NP ₈ EO	14311	(97)	2114	(96)	716	(94)
NP ₉ EO	13991	(97)	2039	(97)	676	(95)
NP ₁₀ EO	12062	(98)	1609	(97)	534	(96)
NP ₁₁ EO	7065	(98)	1112	(98)	366	(96)
NP ₁₂ EO	6069	(98)	569	(98)	196	(97)
NP ₁₋₃ EC	n/d	n/d	-791	(-157)	-195	(-110)
NP ₄₋₁₂ EO	79597	(96)	11977	(93)	4221	(93)

^aBiomass activity was calculated by taking the mass difference of the settled sewage and the final effluent in milligrams of estrogens and dividing it by the MLSS concentration in tonne in the secondary tank;

^bRemoval % was calculated as $\frac{(M_{in} - (M_{WAS} + M_{out}))}{M_{in}} \times 100\%$ - waste activated sludge (WAS) was

estimated to be 2–5% the flow rate of RAS to maintain the SRT of the aeration tank (VFA return is negligible since return flow is circa 1% of main flow).

^cValues obtained from Koh et al. (2005) (39)

Key: ∑EST = sum of steroid estrogens; n/d not determined

Table 3. Apparent biomass sorption coefficient LogKp (l kg^{-1}) for secondary activated sludge

Steroid Estrogen	N/DN 2004 (this study)	N/DN 2006 (this study)	N/DN-P 2006 (this study)	Carbella et al. 2007 <i>LogKd</i> (l kg^{-1})	Joss et al. 2004 <i>LogKd</i> (l kg^{-1})	Ternes et al. 2004 <i>LogKd</i> (l kg^{-1})
E1	2.53	2.40	1.99	2.9	2.95	n/d
E2	2.78	2.67	1.11	4.5	n/d	n/d
E3	2.79	2.35	1.46	n/d	n/d	n/d
EE2	2.93	3.35	2.00	n/d	n/d	2.5
E1-3S	2.05	1.52	1.60	n/d	n/d	n/d
Alkylphenol group						
NP ₃₋₁₂ EO	n/d	1.6	1.2	n/d	n/d	n/d
NP ₁₋₂ EO	n/d	2.6	0.8	n/d	n/d	n/d
NP ₁₋₁₂ EO	n/d	1.8	1.2	n/d	n/d	n/d
NP	n/d	3.2	1.4	n/d	n/d	n/d
NP ₁₋₃ EC	n/d	1.4	0.05	n/d	n/d	n/d

Key: n/d not determined

In this study K_p was calculated using average steroid estrogen or APEO return activated sludge (RAS) concentration data in ng l^{-1} . The adsorbed concentration of steroid estrogens and APEOs in RAS was divided by the mixed liquor concentration - MLVSS (mg l^{-1}) to determine $\text{ng steroid estrogen kg}^{-1}$ biomass or ng APEO kg^{-1} and then divided by the dissolved concentration of steroid estrogens or APEOS after filtration of RAS to determine the partitioning as
$$\text{LogKp} = \frac{(\text{ng.chemical.sorbed} / \text{kg.biomass})}{\text{ng.chemical.dissolved}}$$

Table 4. Concentrations of estrogens in the final effluents of the investigated works (ng l⁻¹) and range in brackets with their EEq values.

Steroid estrogens	Concentration (ng l ⁻¹)			PNEC*
	N/DN		N/DN-P	
	2004	2006	2006	
E1	5.1 (2-7.2)	4.3 (3.2-6.2)	5.5 (1.9-9)	3
E2	0.4 (<MDL-0.6)	0.4 (0.2-0.6)	1.1 (<MDL-2.2)	1
E3	0.5 (<MDL-0.8)	0.4 (0.2-0.9)	0.3 (<MDL-1.1)	-
EE2	0.2 (<MDL-1.3)	0.2 (<MDL-0.4)	0.2 (<MDL-1.1)	0.1
E1-3S	3.1 (0.8-4.8)	7.7 (4-12)	0.8 (0.3-1.6)	-
EEq	4.1	3.8	4.9	<1

*Environment Agency (56).

$$EEq \text{ (ng l}^{-1}\text{)} = \frac{[17\alpha - \text{Ethinylest radiol}]}{PNEC = 0.1} + \frac{[17\beta - \text{Estradiol}]}{PNEC = 1} + \frac{[\text{Estrone}]}{PNEC = 3} < 1$$

Key: MDL = Method detection limit 0.1 ng l⁻¹ for E1 and E1-3S; 0.2 ng l⁻¹ for E2, E3 and EE2

References

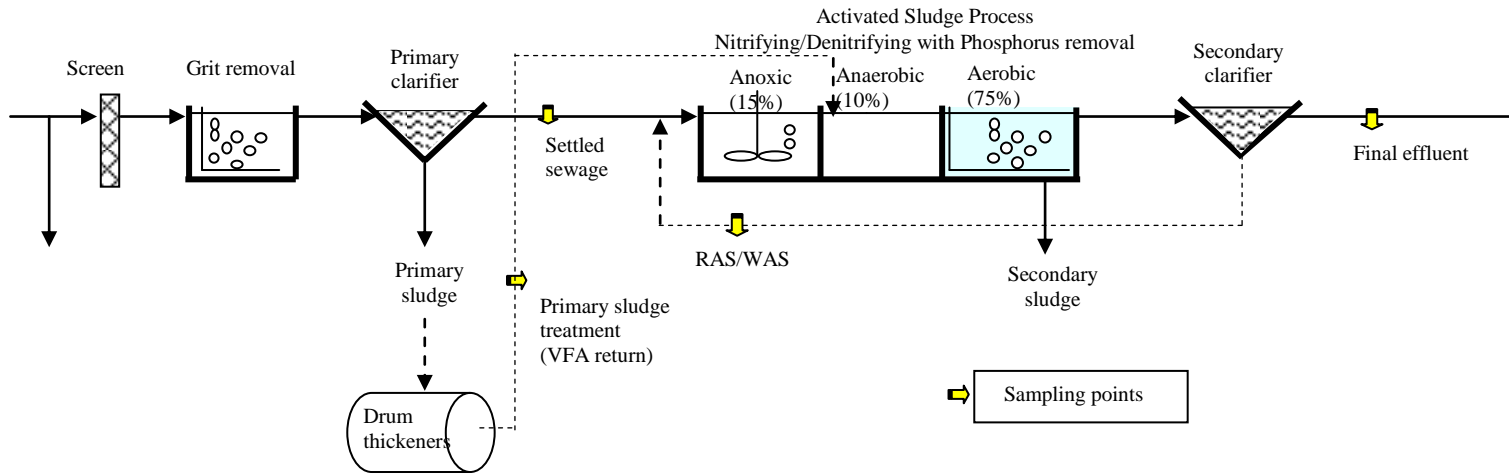
1. Jobling, S.; Nolan, M.; Tyler, C. R.; Brighty, G.; Sumpter, J. P. Widespread sexual disruption in wild fish. *Environ. Sci. Technol.* **1998**, *32*, 2498-2506.
2. Lai, K.M., Scrimshaw, M.D. and Lester, J.N. The effects of natural and synthetic steroid estrogens in relation to their environmental occurrence. *Crit. Revs. Toxicol.* **2002**, *32*, 113-132.
3. Kirk, L. A.; Tyler, C. R.; Lye, C. M.; Sumpter, J. P. Changes in estrogenic and androgenic activities at different stages of treatment in wastewater treatment works. *Environ. Toxicol. Chem.* **2002**, *21*, 972-979.
4. Giger, W.; Brunner, P. H.; Schaffner, C. 4-Nonylphenol in Sewage-Sludge - Accumulation of Toxic Metabolites from Nonionic Surfactants. *Science* **1984**, *225*, 623-625.
5. Chiu, T.Y.; Paterakis, N.; Scrimshaw, M.D.; Cartmell, E.; Lester, J.N. A critical review of the formation of mono and dicarboxylated metabolic intermediates of alkylphenol polyethoxylates during wastewater treatment and their environmental significance. *Crit. Revs. Environ. Sci. Technol.* (in press).
6. Renner, R. European bans on surfactant trigger transatlantic debate. *Environ. Sci. Technol.* **1997**, *31*, 316A-321A.
7. Jobling, S.; Sheahan, D.; Osborne, J. A.; Matthiessen, P.; Sumpter, J. P. Inhibition of testicular growth in rainbow trout (*Oncorhynchus mykiss*) exposed to estrogenic alkylphenolic chemicals. *Environ. Toxicol. Chem.* **1996**, *15*, 194-202.
8. Routledge, E. J.; Sumpter, J. P. Estrogenic activity of surfactants and some of their degradation products assessed using a recombinant yeast screen. *Environ. Toxicol. Chem.* **1996**, *15*, 241-248.
9. Lai, K.M., Scrimshaw, M.D. and Lester, J.N. Biotransformation and bioconcentration of steroid estrogens by *Chlorella vulgaris*. *App. Environ. Microbiol.* **2002**, *68*, 859-864.
10. Gomes, R.L., Deacon, H.E., Lai, K.M., Birkett, J.W., Scrimshaw, M.D. and Lester, J.N. An assessment of the bioaccumulation of estrone in *Daphnia magna*. *Environ. Toxicol. Chem.* **2004**, *23*, 105-108.
11. Langford, K. and Lester, J.N., Chapter 4, Fate and behaviour of endocrine disruptors in wastewater treatment processes. In: *Endocrine Disruptors in Wastewater and Sludge Treatment Processes*. Birkett, J.W. and Lester, J.N (Eds.), CRC Press, Boca Raton, Florida, USA, **2002**; pp103-144.
12. Burke, M. UK to tackle endocrine disrupters in wastewater. *Environ. Sci. Technol.* **2004**, *38*, 362A-363A.
13. UKWIR. Endocrine Disrupting Chemicals National Demonstration Programme: Assessment of the Performance of WwTW in Removing Oestrogenic Substances (09/TX/04/16), UKWIR, London, UK. ISBN 1 84057 525 5, **2009**.
14. Birkett, J. W. Sources of Endocrine Disrupters. In *Endocrine Disruptors in Wastewater and Sludge Treatment Processes*; Birkett, J. W., Lester, J. N., (Eds.) Lewis Pub.: Boca Raton; Florida, USA, **2003**; pp 35-58.
15. Johnson, A. C.; Aerni, H.-R.; Gerritsen, A.; Gibert, M.; Giger, W.; Hylland, K.; Jurgens, M.; Nakari, T.; Pickering, A.; Suter, M. J.-F. Comparing steroid estrogen, and nonylphenol content across a range of European sewage plants with different treatment and management practices. *Water Res.* **2005**, *39*, 47-58.

16. Snyder, S. A.; Westerhoff, P.; Yoon, Y.; Sedlak, D. L. Pharmaceuticals, personal care products, and endocrine disruptors in water: Implications for the water industry. *Environ. Eng. Sci.* **2003**, *20*, 449-469.
17. Servos, M. R.; Bennie, D. T.; Burnison, B. K.; Jurkovic, A.; McInnis, R.; Neheli, T.; Schnell, A.; Seto, P.; Smyth, S. A.; Ternes, T. A. Distribution of estrogens, 17 β -estradiol and estrone, in Canadian municipal wastewater treatment plants. *Sci. Total Environ.* **2005**, *336*, 155-170.
18. Rosenfeldt, E. J.; Linden, K. G. Degradation of endocrine disrupting chemicals bisphenol A, ethinyl estradiol, and estradiol during UV photolysis and advanced oxidation processes. *Environ. Sci. Technol.* **2004**, *38*, 5476-5483.
19. Ifelebuegu, A.O., Lester, J.N., Churchley, J., Cartmell, E. Removal of an endocrine disrupting chemical (17 α -ethinyloestradiol) from wastewater effluent by activated carbon adsorption: Effects of activated carbon type and competitive adsorption. *Environ. Technol.* **2006**, *27*, 1343-1349.
20. Quinlivan, P. A.; Li, L.; Knappe, D. R. U. Effects of activated carbon characteristics on the simultaneous adsorption of aqueous organic micropollutants and natural organic matter. *Water Res.* **2005**, *39*, 1663-1673.
21. Jones, O. A. H.; Green, P.; Voulvoulis, N.; Lester, J. N. Questioning the excessive use of advanced treatment to remove organic micropollutants from wastewater. *Environ. Sci. Technol.* **2007**, *41*, 5085-5089.
22. Andersen, H.; Siegrist, H.; Halling-Sorensen, B.; Ternes, T. A. Fate of estrogens in a municipal sewage treatment plant. *Environ. Sci. Technol.* **2003**, *37*, 4021-4026.
23. Holbrook, R.D.; Novak, J.T.; Grizzard, T.J.; Love, N.G. Estrogen receptor agonist fate during wastewater and biosolids treatment processes: A mass balance analysis. *Environ. Sci. Technol.* **2002**, *36*, 4533-4539.
24. Ternes, T. A.; Stumpf, M.; Mueller, J.; Haberer, K.; Wilken, R. D.; Servos, M. Behavior and occurrence of estrogens in municipal sewage treatment plants - I. Investigations in Germany, Canada and Brazil. *Sci. Total Environ.* **1999**, *225*, 81-90.
25. Clara, M.; Kreuzinger, N.; Strenn, B.; Gans, O.; Kroiss, H. The solids retention time - a suitable design parameter to evaluate the capacity of wastewater treatment plants to remove micropollutants. *Water Res.* **2005**, *39*, 97-106.
26. Svenson, A.; Allard, A. S.; Ek, M. Removal of estrogenicity in Swedish municipal sewage treatment plants. *Water Res.* **2003**, *37*, 4433-4443.
27. Koh, Y. K. K.; Chiu, T. Y.; Boobis, A.; Cartmell, E.; Scrimshaw, M. D.; Lester, J. N. Treatment and removal Strategies of natural Estrogens in the Wastewater. *Environ. Technol.* **2008**, *29*, 245-268.
28. Koh, Y. K. K.; Chiu, T. Y.; Boobis, A.; Cartmell, E.; Lester, J. N.; Scrimshaw, M. D. Determination of steroid estrogens in wastewater by high performance liquid chromatography-tandem mass spectrometry. *J. Chromatogr. A.* **2007**, *1173*, 81-87.
29. Chiu, T.Y.; Koh, Y.K.K.; Paterakis, N.; Boobis, A.R.; Cartmell, E.; Richards, K.H.; Lester, J.N. and Scrimshaw, M.D. The significance of sample mass in the analysis of steroid estrogens in sewage sludges and the derivation of partition coefficients in wastewaters. *J. Chromatog. A.*, **2009**, available on line doi:10.1016/j.chroma.2009.04.019
30. Gomes, R.L.; Birkett, J.W.; Scrimshaw, M.D.; Lester, J.N. Simultaneous determination of natural and synthetic steroid estrogens and their conjugates in aqueous matrices by liquid chromatography / mass spectrometry. *Int. J. Environ. Anal. Chem.* **2005**, *85*, 1-14.

31. Gomes, R.L.; Scrimshaw, M.D.; Lester J.N. Fate of conjugated natural and synthetic steroid estrogens in crude sewage and activated sludge batch studies. *Environ. Sci. Technol.* **2009**, in press.
32. Koh, K.; Chiu, T.Y.; Boobis, A.R.; Cartmell, E.; Pollard, S.; Scrimshaw, M.; Lester, J. A sensitive and robust method for the determination of alkylphenol polyethoxylates and their carboxylic acids and their transformation in a trickling filter wastewater treatment plant. *Chemosphere* **2008**, *73*, 551–556.
33. Ternes, T.A.; Joss, A.; Siegrist, H. Scutinizing pharmaceutical and personal care products in wastewater treatment. *Environ. Sci. Technol.* **2004**, *38*, 393A-399A.
34. Joss, A.; Andersen, H.; Ternes, T.; Richle, P. R.; Siegrist, H. Removal of estrogens in municipal wastewater treatment under aerobic and anaerobic conditions: Consequences for plant optimization. *Environ. Sci. Technol.* **2004**, *38*, 3047-3055.
35. Kreuzinger, N; Clara, M.; Strenn, B.; Kroiss, H. Relevance of the sludge retention time (SRT) as design criteria for wastewater treatment plants for the removal of endocrine disruptors and pharmaceuticals from wastewater. *Water Sci. Technol.* **2004**, *50*, 149-156.
36. Loyo-Rosales, J.E.; Rice, C.P.; Torrents, A. Fate of octyl- and nonylphenol ethoxylates and some carboxylated derivatives in three American wastewater treatment plants. *Environ. Sci. Technol.* **2007**, *41*, 6815-6821.
37. Carballa, M.; Omil, F.; Lema, J.M. Calculation methods to perform mass balances of micropollutants in sewage treatment plants. Application to pharmaceutical and personal care products (PPCPs). *Environ. Sci. Technol.* **2007**, *41*, 884-890.
38. Carballa, M; Omil, F.; Lema, J.M.; Llombart, M.; Garcıa-Jares, C.; Rodrıguez, I.; G3mez, M.; Ternes, T. Behavior pf pharmaceuticals, cosmetics and hormones in a sewage treatment plant. *Water Res.* **2004**, *38*, 2918-2926.
39. Koh, Y.K.K.; Lester, J.N.; Scrimshaw, M.D. Fate and behavior of alkylphenols and their polyethoxylates in an activated sludge plant, *Bull. Environ. Contam. Toxicol.* **2005**, *75*, 1098-1106.
40. Davenport, R.J.; Curtis, T.P.; Goodfellow, M.; Stainsby, F.M.; Bingley, M. Quantitative use of fluorescent in situ hybridization to examine relationships between mycolic acid-containing actinomycetes and foaming in activated sludge plants. *Appl. Environ. Microbiol.* **2000**, *66*, 1158-1156.
41. Langford, K.H.; Scrimshaw, M.D.; Lester, J.N. The impact of process variables on the removal of PBDEs and NPEOs during biological sewage treatment. *Arch. Environ. Contam. Toxicol.* **2007**, *57*, 1-7.
42. Gaulk, L.S.; Strand, S.E.; Kalhorn, T.F.; Stensel, H.D. 17 α -ethinylestradiol transformation via abiotic nitration in the presence of ammonia oxidizing bacteria. *Environ. Sci. Technol.* **2008**, *42*, 7622-7627.
43. Graham, D.W.; Curtis, T.P. Ecological theory and bioremediation. In *Bioremediation: A Critical Review*; Head, I.M., Singleton, I., Milner, M.G., Eds.; Horizon Scientific Press, **2003**; pp 61-80, Norwich, UK
44. Obeng, L.A.; Perry, R.; Lester, J.N. The influence of transient temperature changes on the biodegradation of nitrilotriacetic acid in the activated sludge process. *Environ. Pollut. (Series A)*, **1982**, *28*, 149-161.
45. Stephenson, T.; Lester, J.N.; Perry, R. The influence of transient temperature changes on the biodegradation of nitrilotriacetic acid in the activated sludge process: a pilot plant study. *Environ. Pollut. (Series A)*, **1983**, *32*, 1-10.

46. Johnson, A.C.; Belfroid, A.; Di Corcia, A. Estimating steroid oestrogen inputs into activated sludge treatment works and observations on their removal from the effluent. *Sci. Total Environ.* **2000**, *256*, 163-173.
47. Onda, K.; Nakamura, Y.; Takatoh, C.; Miya, A.; Katsu, Y. The behavior of estrogenic substances in the biological treatment process of sewage. *Water Sci. Technol.* **2003**, *47*, 109-116.
48. D'Ascenzo, G.; Di Corcia, A.; Gentili, A.; Mancini, R.; Mastropasqua, R.; Nazzari, M.; Samperi, R. Fate of natural estrogen conjugates in municipal sewage transport and treatment facilities. *Sci. Total Environ.* **2003**, *302*, 199-209.
49. Nakada, N.; Yasojima, M.; Okayasu, Y.; Komori, K.; Tanaka, H.; Suzuki, Y. Fate of oestrogenic compounds and identification of oestrogenicity in a wastewater treatment process. *Water Sci. Technol.* **2006**, *53*, 51-63.
50. Vader, J.S.; Van Ginkel, C.G.; Sperling, F.M.G.M.; De Jong, J.; De Boer, W.; De Graaf, J.S.; Van Der Most, M.; Stokman, P.G.W. Degradation of ethinyl estradiol by nitrifying activated sludge. *Chemosphere* **2000**, *41*, 1239-1243.
51. Ren, Y.-X.; Nakano, K.; Nomura, M.; Chiba, N.; Nishimura, O. Effects of bacterial activity on estrogen removal in nitrifying activated sludge. *Water Res.* **2007**, *41*, 3089-3096.
52. Fürhacker, M.; Breithofer, A.; Jungbauer, A. 17 beta-estradiol: Behavior during waste water analyses. *Chemosphere* **1999**, *39*, 1903-1909.
53. Ternes, T.A.; Herrmann, N.; Bonerz, M.; Knacker, T.; Siegrist, H.; Joss, A.A rapid method to measure the solid-water distribution coefficient (K_d) for pharmaceuticals and musk fragrances in sewage sludge. *Water Res.* **2004**, *38*, 4075-4084.
54. Baronti, C.; Curini, R.; D'Ascenzo, G.; Di Corcia, A.; Gentili, A.; Samperi, R. Monitoring natural and synthetic estrogens at activated sludge sewage treatment plants and in a receiving river water. *Environ. Sci. Technol.* **2000**, *34*, 5059-5066.
55. Belfroid, A.C.; Van der Horst, A.; Vethaak, A.D.; Schafer, A.J.; Rijs, G.B.J.; Wegener, J.; Cofino, W.P. Analysis and occurrence of estrogenic hormones and their glucuronides in surface water and waste water in The Netherlands. *Sci. Total Environ.* **1999**, *225*, 101-108.
56. Janex-Habibi, M.L.; Huyard, A.; Esperanza, M.; Bruchet, A. Reduction of endocrine disruptor emissions in the environment: The benefit of wastewater treatment. *Water Res.* **2009**, *43*, 1565-1576.
57. Environment Agency *Proposed Predicted–No–Effect Concentrations (PNECs) for Natural and Synthetic Steroid Estrogens in Surface Waters*; R&D Technical Report P2-T04/1., **2002**; Bristol, UK.
58. European Commission, 4-Nonylphenol (branched) and nonylphenol: European Union risk assessment report. **2002**, EUR 20387 EN.
59. Ahel, M.; Molnar, E.; Ibric, S.; Giger, W. Estrogenic metabolites of alkylphenol polyethoxylates in secondary sewage effluents and rivers. *Water Sci. Technol.* **2000**, *42*, 15-22.
60. Lagana, A.; Bacaloni, A.; De Leva, I.; Faberi, A.; Fago, G.; Marino, A. Analytical methodologies for determining the occurrence of endocrine disrupting chemicals in sewage treatment plants and natural waters. *Anal. Chim. Acta* **2004**, *501*, 79-88.

A. Nitrifying/Denitrifying (N/DN) works operating with no internal recycle and a plug flow aerobic zone.



B. Nitrifying/Denitrifying with Phosphorus removal (N/DN-P) works operating with no internal recycle and a plug flow aerobic zone.

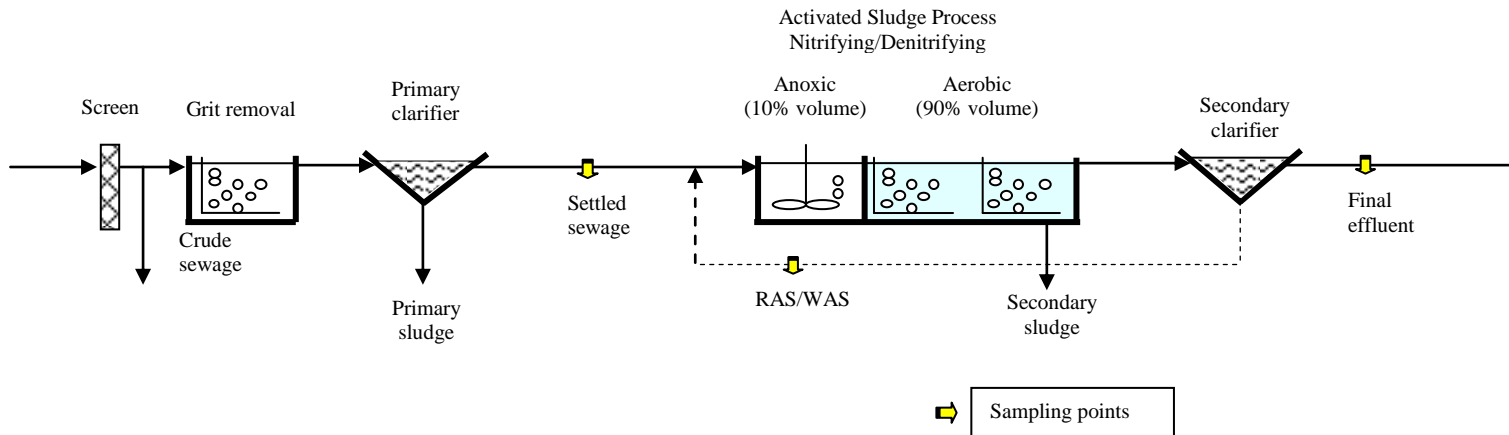


Figure 1. Schematic diagrams of the two activated sludge sewage treatment works sampled – A nitrifying/denitrifying (N/DN) and B nitrifying/denitrifying with phosphorus removal (N/DN-P)

Steroid estrogens

Alkylphenols

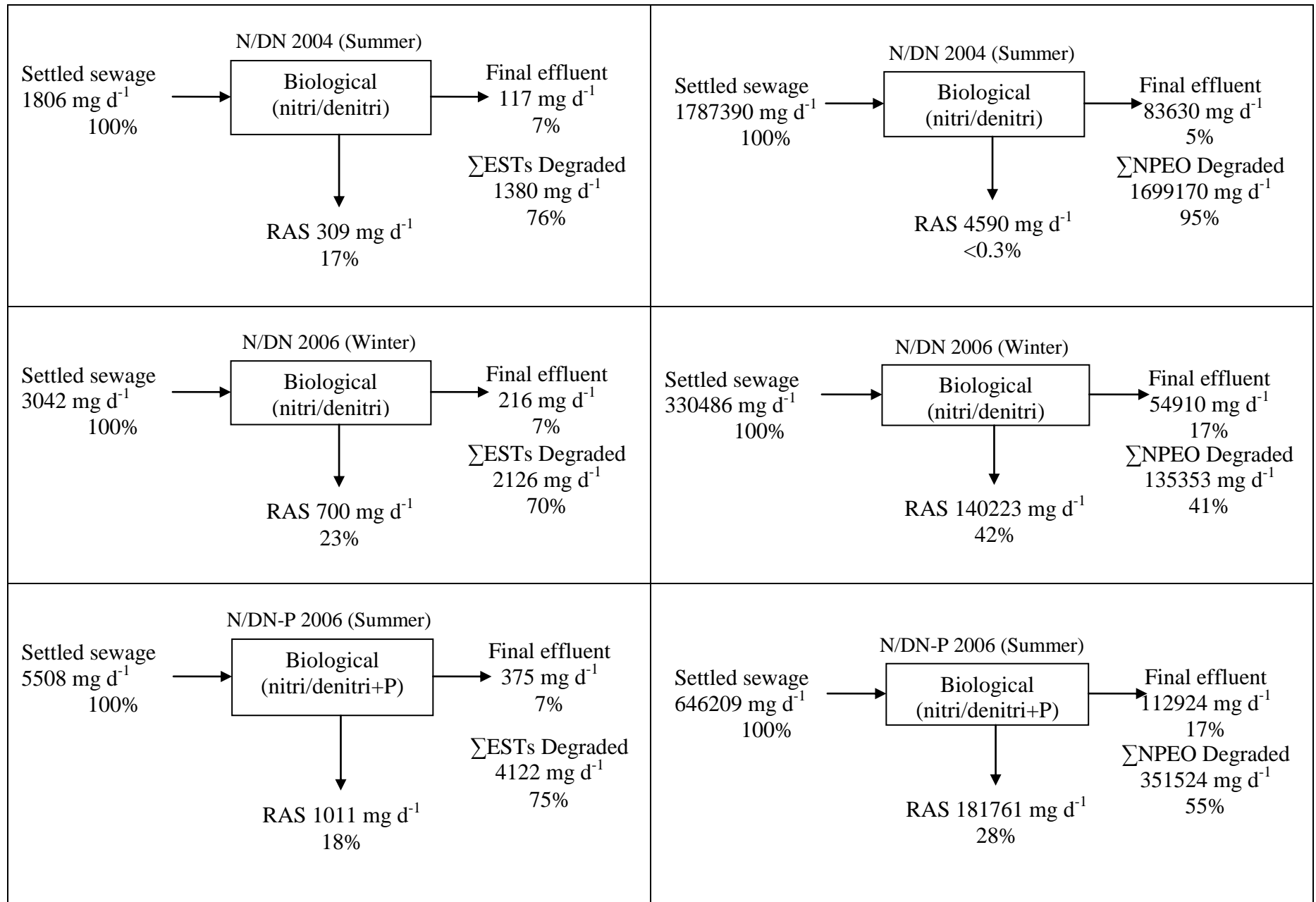


Figure 2. Mass balance of the total steroid estrogens and of alkyl phenolic compounds NPEO, NPEC and NP in the sewage treatment works. The degraded component has been determined from settled sewage – (RAS + final effluent). The N/DN data for alkylphenolic compounds in 2004 are from Koh et al. (50) and do not include NPEC.

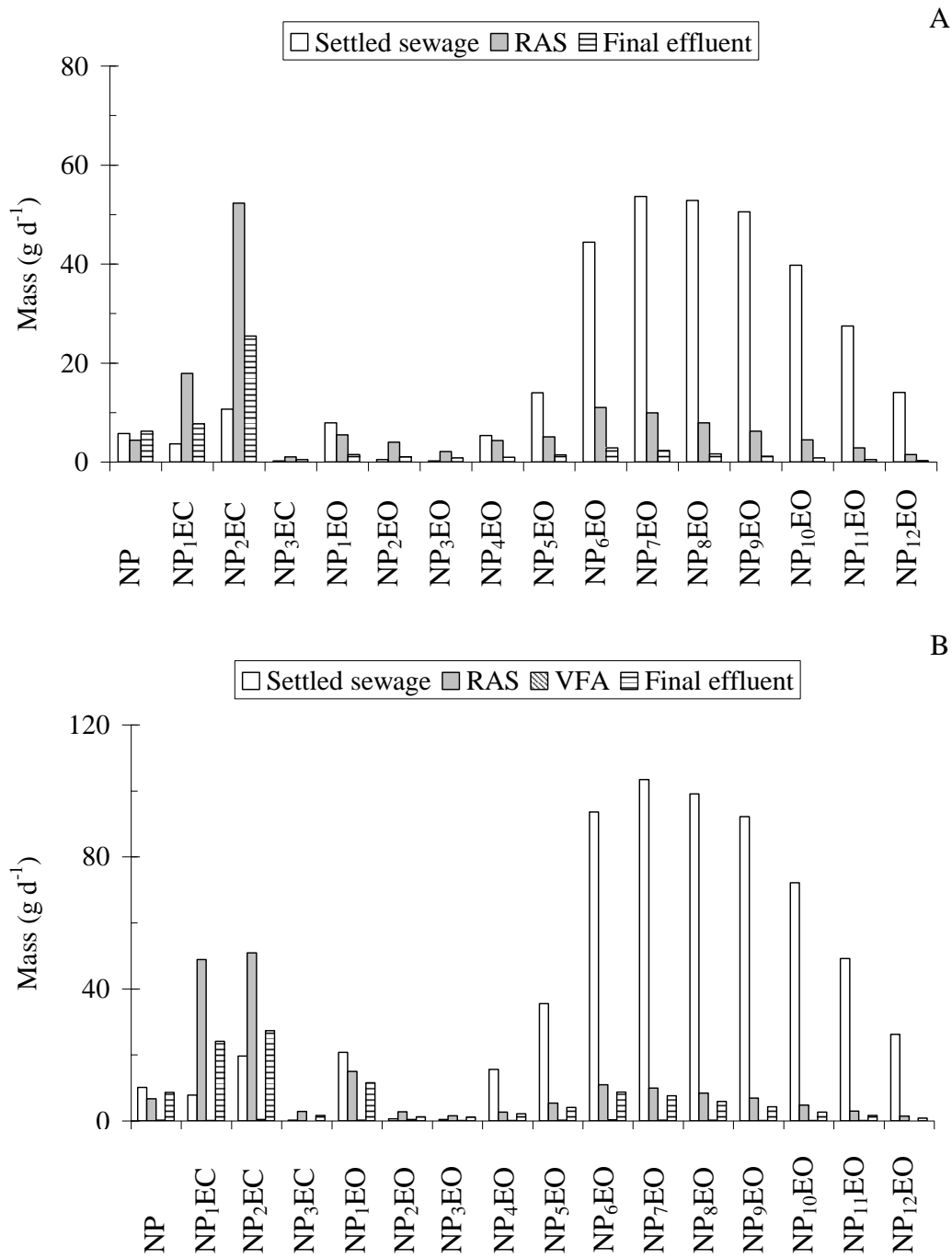


Figure 3. Mass fluxes of NPEO in N/DN (A) (2006) and N/DN-P (B) (2006).

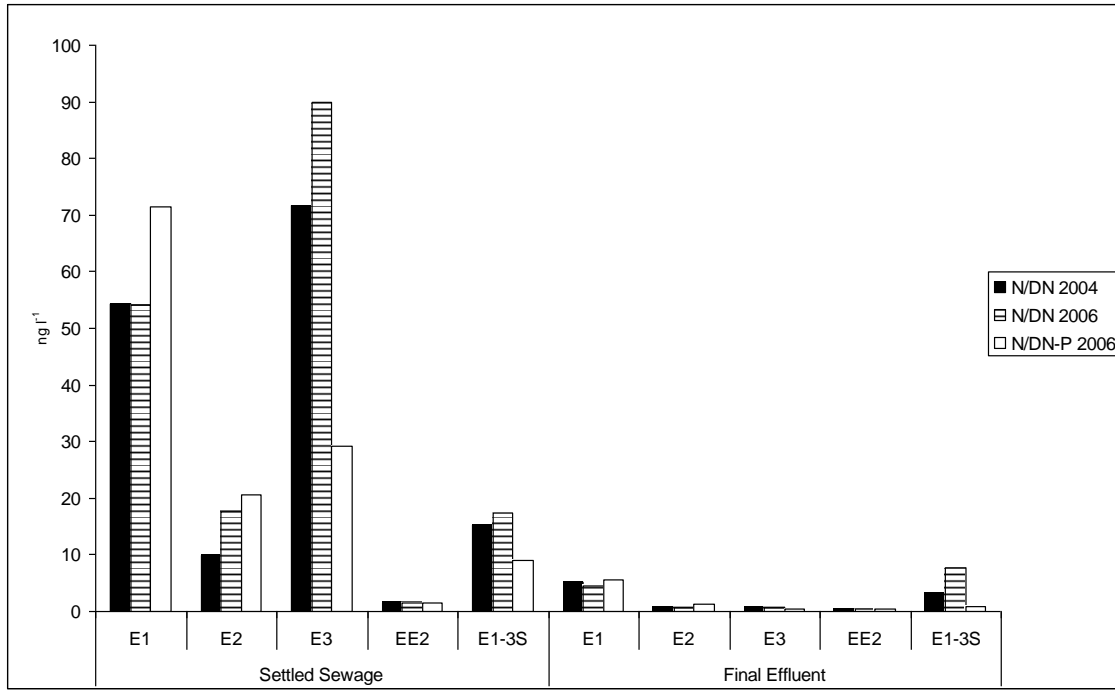


Figure 4. Concentrations of total steroid estrogens (dissolved and adsorbed) in the sewage treatment works