# Atlas of Genetics and Cytogenetics in Oncology and Haematology

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## Gene Section Review

## S100A4 (S100 Calcium Binding Protein A4)

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### Abstract

Review on S100A4, with data on DNA/RNA, on the protein encoded and where the gene is implicated.

## Identity

**Other names:** 18A2, 42A, CAPL, FSP1, MTS1, P9KA, PEL98

HGNC (Hugo): S100A4

Location: 1q21.3

#### Local order

The 1q21 locus harbours the epidermal differentiation complex (EDC) encompassing a 2.05 Mbp of human genomic DNA. The S100 family genes except the S100beta are arranged in the following order: 1 cen-S100A10-S100A11-THH (trychohyalin)-FLG (filaggrin)-IVL (involucrin)-LOR (loricrin)-S100A9-S100A12-S100A8-S100A7A-S100A7P1-S100A7L2-

S100A7P2-S100A7-S100A6-S100A5-S100A4-

S100A3-S100A2-S100A16-S100A14-S100A13-

S100A1-1qtel. S100beta is located on 21q22.3 (Schäfer et al., 1995; Marenholz et al., 1996; Mischke et al., 1996; GeneLoc version 2.41: pseudogenes S100A7P1: HGNC 21654, S100A7P2: HGNC 21656). S100A11 pseudogenes have been listed (Pseudogenes.org); they are not shown here.

The 18A2 cDNA/mRNA (S100A4) was cloned and

its structure, translation product and tissue distribution were described by Jackson-Grusby and colleagues. The sequence of 18A2 was similar to that of the 2A9 clone described by Calabretta and collegues.

### **DNA/RNA**

#### Note

Starts 153516089 bp from pter; ends 153522612 bp from pter; 6524 bases; orientation minus strand.

Homo sapiens chromosome 1, GRCh37 primary reference assembly.

NCBI reference sequence: NC\_000001.10, NT\_004487.19.

#### Description

The human S100A4 gene has four exons. Exon 1 is non-coding and exons 2 and 3 are coding exons. Exon 2 with the start codon and encodes N-terminal EF hand and exon 3 encodes the C-terminal EF-hand. The fourth non-coding exon occurs in the 5'-UTR.

#### Transcription

Two variant RNA transcripts from Source Search. The NM\_019554 is a longer transcript. NM\_002961 possesses alternate 5'-UTR; both encode the same protein isoform.

NCBI reference sequence NM\_019554 ver 01954.2 564 bp mRNA; NM 002961 ver 002961.2; 512 bp mRNA.



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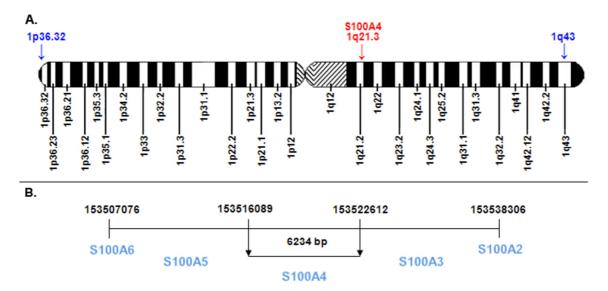


Figure 1. A. Chromosome 1 ideogram showing the location of S100A4 based on National Library of Medicine Handbook. B. The position of S100A4 is based on Entrez Gene ID 6275 (not drawn to scale).

Splice variants have been reported of S100A4. In human osteosarcoma, alternative splicing within the 5'-untranslated region (UTR) generates the two variants. Both variants, hu-mts1 and hu-mts1 (var), contain one open reading frame, differ slightly in translational capacity; possess similar stability. The hu-mts1 and hu-mts1 (var) splice variants with exons 1, 2, 3, and 4 and another with exons 1, 3 and 4, may be differentially expressed. The hu-mts1 (var) is expressed in the colon but not in the liver; it was not found in leukocytes, neutrophils, macrophages and lymphocytes. The hu-mts1 variant predominated in human breast carcinoma (SK-BR-3) and lung carcinoma (A549) is predominant (Ambartsumian et al., 1995). The differential association of the variants has been described in gastric cancers, which seems to relate to disease state possibly relates to progression; however the expression status of the variants in the lymph node metastases is not known. A splice transcript with loss of non-coding exon 1a/1b, but exons 2 and 3 present has been described in infiltrating carcinoma of the breast by Albertazzi et

al.. One would note nonetheless that Alternative Splicing and Transcript Diversity (ASTD) have listed 12 variant transcripts.

#### **Regulation of transcription**

Binding sites for several transcription factors have been identified in the promoter of S100A4. SABiosciences ChIP-qPCR Assay database lists 19 p53 binding sites.

Multiple NFAT (nuclear factor of activated T cells) transcription factor consensus binding sites; NFkappaB related binding site (Tulchinsky et al., 1997). Much evidence is also available regarding activation of NF-kappaB by S100A4. S100A4 can activate NF-kappaB via the classical pathway mediated by MEKK/IKK<sub>β</sub>; S100A6 and S100P also are capable of exerting pro-metastatic effects again pathway. bv activating the NF-kappaB Experimentally induced expression of S100A4 is inhibited by NF-kappaB inhibitors. Aside from these, several other regulatory pathways may be identified, e.g. the Wnt/ $\beta$ -catenin/TCF, HIF/HER among others, as evidenced by the established phenotypic expression induced by the gene.

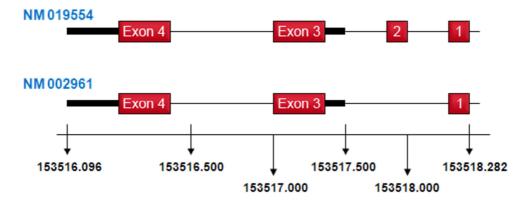


Figure 2. Based on USCS GRCh37/hg19 Feb. 2009. Not drawn to scale.

#### 1 macplekald vmvstfhkys gkegdkfkin kselkelltr elpsfigkrt deaafqkims

#### 61 nldsnrdnev dfgeycvfls ciammcneff egfpdkqprk k

Figure 3. Sequence.

S100A4 has been postulated to signal via RAGE (receptor for advanced glycation end products) which is known to activate NF-kappaB.

A composite enhancer consisting of 6 cis-elements has been identified in the first intron of murine S100A4. This interacts with Sp1 and AP-1 family members and CBF (core binding factor alpha) and KRC (zinc finger transcription factor kappa recognition component) transcription factors.

#### Pseudogene

None reported.

## Protein

#### Description

Human S100A4 (also mouse and rat S100A4) contains 101 aminoacid residues and is approx 12 kDa in size. In common with most S100 family members, S100A4 is an antiparallel homodimer stabilised by noncovalent interactions between two helices from each subunit forming an X-type fourhelix bundle. Each subunit has two calcium-binding EF-hands linked by the intermediate hinge region and a distinctive C-terminal extension. A pseudo-EF hand formed by helices 1 and 2 and the pseudo-EF-hand and a canonical EF-hand that are brought into proximity by a small two-stranded antiparallel beta-sheet. The hinge region and the C-terminal loop of \$100 proteins are involved in target protein binding. Calcium binding produces а conformational change, which leads to the exposure of hydrophobic pocket of residues in helices 3 and 5, the hinge region and the C-terminal loop. This conformational change is required for target protein binding. S100A4 might be post-translationally modified. Charged variants conceivably resulting from post-translational changes have been described in one report, but no confirmation of these findings has been forthcoming. However, calculations from the predicted isoelectric point of S100A4 and separation of the charged variants from the major spot would suggest that two variants may have displayed 17.4 and 26.1% and a third variant with a possible highly extended form and nearly 2.6-fold increase in net negative charge. Alterations in net molecular charge of this magnitude and charge distribution can alter protein

configuration.

NCBI sequence: NM\_002961; NP\_002952; NM\_019554; NP\_062427; UniProtKB/Swiss-Prot: P26447.

#### Features

EF-hand domains:

EF hand 1: length 36; position 12-47,

EF hand 2: length 36; position 50-84.

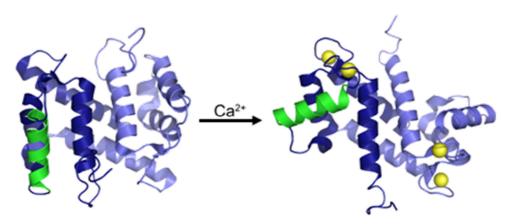
Target protein interaction domains: in the active state S100A4 interacts with many target proteins e.g. p53 family proteins, HDM2, Annexin II, Factin, tropomyosin, and heavy chain of non-muscle myosin IIA, among others. In a closed conformational state S100A4 is inactive, but the protein assumes an open conformation upon calcium binding. In the altered configuration S100A4 can interact with target proteins. These target proteins interact with specific binding domains of S100A4, which are accessible upon conformational change of the apoprotein upon Ca<sup>2+</sup> binding. The Rudland/Barraclough group has shown that specific mutations that inhibit selfassociation of S100A4 markedly reduce its metastasis promoting effects. The mutations reduce self-association and reduce the affinity of S100A4 to two target proteins viz. p53 and non-muscle myosin heavy chain isoform A. The interaction between S100A4 and target proteins can possibly also be disrupted by the packaging of S100A4 in such a way as to sequester S100A4 dimers.

Inhibition of S100A4 polymerisation by suppressing TG2 (tissue transglutaminase 2) function has resulted in the inhibition of cell migration in vitro. This is inspired by the fact that TG2 is a cross-linking protein. Treatment of cells in vitro with EGF seems to up regulate the expression of EGFR and TG2 accompanied by enhanced cell migration. S100A4 over expressing tumours not infrequently tend to be EGFR postive; so tissue transglutaminase could be promoting EGFR dimerisation and facilitate EGF/EGFR signalling.

#### Expression

S100A4 is distributed ubiquitously in normal tissues (Mazzucchelli, 2002).

For expression profile: Human Protein Atlas (HPA): CAB002618 and Human Protein Reference Database HPRD.



**Figure 4.** S100A4 undergoes a calcium-dependent conformational rearrangement that exposes the protein target binding cleft. Ribbon diagrams showing the NMR solution structure of apo-S100A4 (PDB code 1M31) and the X-ray structure of calciumbound S100A4 (PDB code 2Q91). Following the addition of calcium (yellow spheres), helix 3 (green helix in dark blue monomer) moves to expose the target bind cleft. This conformational rearrangement is required for S100A4 binding to protein targets. The author is grateful to Anne R. Bresnick, Ph.D., Department of Biochemistry, Albert Einstein College of Medicine, Bronx, New York, for providing this illustration and the brief legend.

S100A4 occurs in many forms of human cancer, e.g. breast, colorectal, liver, lung, head and neck, ovarian, endometrial, pancreatic, renal, testicular, and prostate cancers, and melanoma; also in many cell lines of myeloid, lymphoid, lung and brain origin and cell lines derived from many forms of leukaemias. The expression of the gene is regulated by methylation. Over expression correlates with hypomethylation and the frequency of hypomethylation relates to tumour progression, e.g. in ovarian cancers. There is no implication at present that the degree of methylation is related to expression. S100A4 has been implicated in other human diseases, e.g. Crohn's disease and rheumatoid arthritis.

#### Localisation

S100A4 occurs extracellulary and also in cytoplasmic and nuclear location. Differential distribution has been reported between stromal components of primary and metastatic tumour. Patterns of distribution could vary between tissues and between species. No firm functional link has been made with the site/s of localisation.

Intracellular distribution is an important factor in determining genetic activity.

It may be noted here that S100A4 is often expressed in component inflammatory cells of tumour stroma. It has been postulated that interactions between the stroma and tumour cells lead to the expression of the protein and modulate its function in either or both. However, both the postulate and its potential influence in tumour progression are yet to be established.

The pattern of intracellular distribution of many genetic determinants has been found to be highly relevant to invasion and metastasis. Nuclear location of S100A4 was shown some while ago to

relate to aggressive tumour behaviour and poor prognosis. Translocation to the nucleus has been associated with EMT induced by TGF- $\beta$ /Smad signalling. IL-induced translocation seems to require sumoylation of specific lysine residues and in this way conceivably regulating target gene expression. Expression patterns need to be explored in more than one tumour system. This might be crucial in the development of strategies of treatment targeting S100A4, especially with the postulated link of S100A4 expression with chemoresistance.

#### Function

S100A4 protein promotes metastasis, functions as a counter point to metastasis suppressor nm23, and is implicated in the regulation of the cell cycle, cell proliferation, motility, invasion, tubulin polymerisation, and angiogenesis. S100A4 might suppress expression of other suppressor genes e.g. PRDM2 and VASH1. PRDM2 (PR domain containing 2, with ZNF domain) is a tumour suppressor gene encoding a zinc finger protein. VASH1 (vasohibin 1) inhibits cell migration, proliferation and tumour growth and angiogenesis. S100A4 promotes metastatic spread of cancer as demonstrated by gene transfer studies. Its expression has shown clear correlation with tumour spread to lymph nodes and with prognosis.

## Cell cycle, cell proliferation, tumour growth and apoptosis.

S100A4 binds to and forms complexes with p53 to regulate cell cycle progression. P53 has been confirmed as a target of S100A4, which stabilises p53. There is conclusive evidence that S100A4 binds to C-terminal regulatory region of p53. S100A4 and certain other members of the S100 family bind to TAD transactivation domain (residues 1-57) of p53.

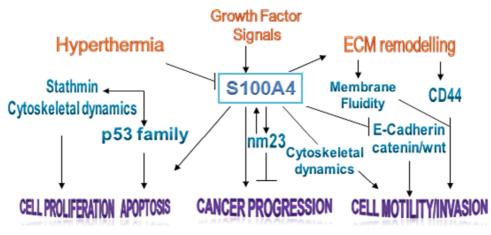


Figure 5.

They may also affect p53 function by binding to the tetramerization domain of p53 (residues 325-355) and interfering with intracellular translocation and subcellular localisation.

This interaction is suggested to be linked with p53 function. Nineteen p53 binding sites have been identified in the promoter of S100A4 (SABiosciences ChIP-qPCR Assay). S100A4 also influences waf1 ID: 139> and mdm2, a regulator of p53 function and the apoptosis family bax gene. It binds to N-terminal domain of mdm2. Signalling pathways include P53-Rb/stathmin/p53 down stream effectors, e.g. p21<sup>waf</sup>, p16 etc. P53/stathmin signalling modulates microtubule dynamics and cell division. Furthermore, p53 and down stream target apoptosis family genes such as BNIP3, caspases; calpain/Fas (?) are postulated as important pathways in S100A4 signalling. Knockdown of S100A4 has been reported to lead to apoptosis. The transcription factor NF-kappaB which involved in anti-apoptosis has been implicated in S100A4 signalling.

S100A4 proliferative signalling seems to involve epidermal growth factor receptors (EGFR). EGFR expression correlates with S100A4 expression. Interactive signalling with HER2 might be postulated with the finding that S100A4 stimulates EGFR/HER2 receptor signalling and on the identification in human S100A4 promoter of an HER2 response element 1099-1487 bp up stream of the transcription start site. The interaction of S100A4 with the TGF-beta system via Smad has also been reported. S100A4 seems able to bind to the N-ter region of Smad3. TGF-beta is an important activator of epithelial mesenchymal transition leading to acquisition of invasive ability. The interaction between S100A4 and Smad thus falls in place with the metastasis-promoting function of the former. Some of these pathways are pictorially represented above (figure 5). S100A4 activates EMT via up regulation of Snail, a negative regulator of E-cadherin. The TGF- $\beta$  family receptor Activin involvement has been implicated.

#### Invasion, motility, and intercellular adhesion.

One of the targets of S100A4 involved in cell motility is myosin filaments. Myosin II consists of two heavy chains (MHC) with globular domains which interact with F-actin. The tail domains of heavy chains form a coiled-coil tail that participates in the assembly of myosin filaments. Wrapped round the neck region of each heavy chain are the essential and the regulatory light chains. Phosphorylation of the regulatory light chain and also of MHC plays an important part in the assesmbly of myosin II monomers into filaments. S100A4 inhibits CK2-mediated phosphorylation of MHC, inhibits the assembly of myosin monomers into filaments. The affinity of S100A4 for the myosin-IIA can be reduced by CK2-mediated phosphorylation. S100A4 destabilises MHCIIA filaments phosphorylated by PKC and inhibits the assembly of monomers. PKC and CK2 can phosphorylate distinct serine residues but yet be additive in their effect. The outcome is that S100A4 promotes dissociation of the filaments and prevents self assembly of monomers resulting in enhanced migration. Thus S100A4 seems to provide a mechanistic link between the actomyosin cytoskeletal and migration.

Signalling systems include modulation of cytoskeletal dynamics; cadherin/catenin complex cytoskeletal linkage and significantly a TCF, a component of the canonical Wnt signalling system, binding site has been identified in the S100A4 promoter and S100A4 directly binds heterodimeric beta-catenin/TCF complexes; CD44/cytoskeletal linkage; ECM associated proteolytic enzyme remodelling. affects system/ECM tubulin polymerisation. S100A4 and tumour suppressor nm23 exert opposite effects on tubulin dynamics. Two C-terminal lysine residues are required for enhanced motility and invasion and interaction with

target proteins. The connective tissue growth factor (CTGF) has been reported to up regulate S100A4 expression and inhibition of S100A4 blocks CTGF-induced cell motility.

S100A4 seems to function via the MMP/TIMP system in promoting invasion as well as induction of angiogenesis. S100A4 is over expressed in invasive glioma cell lines together with down regulation of TIMP-2, indicating a close linkup of S100A4 with the MMP system in the promotion of invasion.

Angiogenesis signalling occurs via activation of MMP/TIMP; activation of angiogenic factors VEGF/endothelial cell proliferation; MetAP2/p53mediated inhibition of endothelial cell proliferation. S100A4 stimulates angiogenic signalling in breast cancer. An indirect link is suggested by the inhibition of S100A4 by Interferon-gamma which might inhibit angiogenesis by down regulating VEGF expression. Hypoxia is a major regulator of angiogenesis. HIF-1 $\alpha$  (hypoxia-inducible factor-1 $\alpha$ ) is a transcription regulator in hypoxia. It can activate VEGF to induce angiogenesis and TGF- $\alpha$ and promotes cell survival. Exposure to hypoxia has been correlated with reduced methylation of the hypoxia response element in S100A4's promoter region and enhanced HIF binding to the promoter and increased transcription of the gene together with increased cell proliferation and invasion. Given that HIF also promotes VEGF expression one can see a potential two pronged approach to control tumour growth with HIF inhibition. Some clinical studies are underway to study the effects of Sorafenib-mediated inhibition of HIF-1 $\alpha$  and VEGF. In laboratory studies Sorafenib has been

found to reduce tumour growth and tumour associated microvessel density.

Osteopontin was identified as a metastasisassociated protein some time ago. Many strands of evidence suggest that osteopontin is an intermediary in S100A4 signalling pathway. In breast cancer expression of osteopontin in the background of S100A4 has generally correlated with poor patient survival.

Osteopontin is associated with several activated NF-kappaB pathways. S100A4 induces the expression and secretion of osteopontin in some osteosarcoma cell lines in an NF-kappaB-dependent fashion. Inhibition of osteopontin inhibits tumour development and angiogenesis; inhibition of both might result in synergistic suppression of tumour progression.

Shown below are the potential pathways of S100A4 signalling in cell motility/invasion and angiogenesis, emphasising the possibility that S100A4 seems able to influence many significant systems leading to angiogenesis.

#### Homology

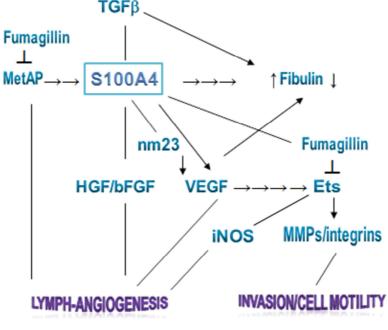
Sequence homology to protein from Pan troglodytes (Chimpanzee) (Gene ID: 457320; Protein NCBI RefSeq: XP\_001138744.1).

Bos taurus (Bovine) (Gene ID: 282343).

Canis lupus familiaris (Gene ID: 403787; NCBI reference sequence: NP\_001003161.1; protein: NP\_777020.1).

Sequence homology 93% to murine S100A4 (Entrez Gene ID 20198; NP\_035441).

Sequence homology 91% to rat protein (Entrez Gene ID 24615; NP\_036750).





### **Mutations**

#### Note

Many SNPs have been identified; 7 shown in NCBI and 26 in Applied Biosystems data source. The NCBI Entrez SNP database lists 19 submissions.

#### Chromosomal rearrangements

The locus 1q21 is a hotspot for chromosomal rearrangements, microdeletions and duplications; significance uncertain and there are no clear implications for metastasis. No translocations leading to hybrid S100A4 have been recorded.

There are 11 common and 1 rare fragile sites on chromosome 1. The common FRA1F occurs in 1q21. Chromosome 1 is prone to sister chromatid recombination (SCR) and >70% SCRs occur at the fragile sites or in the same band as the fragile sites, but no link with S100A4 established.

#### Germinal

None reported.

#### Somatic

No simple mutations, gene fusions, or structural variants detected in breast and colorectal carcinomas and in gliomas (Cosmic: Catalogue Of Somatic Mutations In Cancer, Welcome Trust Sanger Institute).

No mutations have been found coding regions in human, canine and feline S100A4. Mutating phenylalanine 72 to alanine reduces functional effectiveness. Toombak (tobacco rich in tobaccospecific nitrosamine) dipping (placing between the lower lip and gums) has been indirectly linked with S100A4 mutations in oral squamous cell carcinoma, but mutations have been described also in non-dippers. The carcinoma from dippers had 4 mutations (one transition, 3 transversions) and nonsnuff-dippers showed 3 mutations each (one transition, 2 transversions). The suggestion is that S100A4 mutations could be complementing the effects of more frequent mutations of p53 and  $p21^{waf1}$ .

## Implicated in

# General notes on association with human cancer

#### Note

S100A4 has been implicated in the progression and prognosis of several forms of human cancer, e.g. breast, colorectal, gastric, pancreatic and bladder cancer, SCLC and oesophageal squamous cell carcinoma, among others. Poor prognosis associated with high S100A4 expression is accompanied by clear signs of disease progression, e.g. high histological and clinical grades and involvement of lymph nodes. Also indicative of poor prognosis is high S100A4 expression coupled with reduced E-cadherin expression in pancreatic, oral squamous cell carcinoma and in melanoma. S100A4 expression is inversely related with expression of metastasis suppressor nm23 and with prognosis of breast cancer.

#### Cytogenetics

No cytogenetic data are available.

#### Abnormal protein

No fusion proteins or hybrid genes involving S100A4 are known.

#### Breast cancer

#### Note

Both tumour and serum levels are reportedly enhanced in breast cancer patients. S100A4 expression is inversely related to that of the metastasis suppressor nm23 in breast cancers. Tumour levels might correlate with proliferative state and shown to be linked with p53 dysfunction. S100A4 proliferative signalling seems to involve epidermal growth factor receptors (EGFR). Breast cancers that are high S100A4s expressers tend to be oestrogen (ER)/progesterone receptor (PR) negative.

Given that ER/PR expression is inversely related to the expression of epidermal growth factor receptors, ER/PR status together with S100A4/nm23 expression status could provide significant leads to the prediction of prognosis. S100A4 signalling could interact with HER2 function; this is suggested by the finding that S100A4 stimulates EGFR/HER2.

Up regulated expression was associated with increased tumour angiogenesis and this would be expected to contribute to the invasive spread of breast cancer.

Of some interest is the suggestion that phosphosulindac might target and induce apoptosis of breast cancer stem cells.

#### Prognosis

S100A4 may be regarded as an independent predictor of prognosis.

#### **Colorectal cancer**

#### Note

Primary cancers show enhanced S100A4 expression and associated with metastatic disease in the lymph nodes.

Up regulation of its expression has been correlated with enhanced invasion and nodal dissemination. Nuclear expression has been reported to be a prognostic indicator.

As in the case of breast cancer there are indications that S100A4 might interact with and abrogate p53 function. The implied association with aggressive disease is underscored by the emergence of correlated expression of S100A4 with the extracellular matrix metalloproteinase inducer CD147/Basigin/EMMPRIN but a causal link is yet to be established. One should note in this context that S100A9 can also bind CD147.

Efforts are being made to inhibit Wnt/β-catenin mediated targeting of S100A4 using sulindac.

#### Bladder cancer

#### Note

Higher expression S100A4 has been observed and this might be associated with muscle invasion.

#### Prognosis

High expression has been related to decreased survival.

#### Oesophageal squamous cell carcinoma

#### Note

S100A4 expression levels negatively corresponded with E-cadherin expression in ESCCs patients with metastatic disease. In vitro studies of migration of cells with experimentally enhanced S100A4 expression have lent support to the perceived relationship. Transfection of gall bladder carcinoma cell lines E-cadherin has led to the suppression of S100A4.

#### Ovarian cancer

#### Note

The expression of nuclear S100A4 expression is associated with more aggressive disease in primary carcinoma where the level of expression has been reported to be higher in solid tumours than in effusions.

#### Lung cancer

#### Note

Higher expression of S100A4 has been encountered in squamous cell but not adenocarcinoma of the lung.

#### Prognosis

A large study of the expression levels has revealed S100A4 to be significantly predictive of survival in squamous cell but not adenocarcinoma of the lung. S100A4 was significantly associated with patients' poor prognosis in lung squamous cell carcinoma but not lung adenocarcinoma.

#### Pancreatic cancer

#### Note

S100A4 up regulation might be accompanied by reduced E-cadherin expression. This inverse relationship has also been encountered in melanoma and oral squamous cell carcinoma cell lines. This might generate an additive effect on tumour aggression.

Perineural invasion has been associated with enhanced S100A4 expression. Worthy of note is that perineural invasion is a feature linked with tumour spread and poor prognosis and its correlation with S100A4 might have implications for disease management.

#### Prognosis

Some preliminary evidence is available indicating that S100A4 expression levels relate to shorter overall survival of patients with pancreatic cancer.

#### Gastric cancer

#### Note

Higher expression of S100A4 has been noted in gastric cancer and correlated with the presence of the tumour in lymph node and the occurrence of distant metastases, and with poor prognosis. Consistent with the situation in certain other forms of cancer, in gastric cancer S100A4 levels inversely relate to E-cadherin expression. Indeed, down regulation of E-cadherin has been found to occur in parallel with hypomethylation of S100A4.

#### Melanoma

#### Note

A marked inverse relationship has been described between S100A4 and E-cadherin in these tumours.

#### Gliomas

#### Note

S100A4 is over expressed in invasive glioma cell lines together with down regulation of TIMP-2, indicating a close linkup of S100A4 with the MMP system in the promotion of invasion. This ability to induce angiogenesis and metastatic dissemination could be complemented and indeed augmented by its postulated ability to enhance endothelial permeability. Occludin is a transmembrane tightjunction protein essential for maintaining integrity of both epithelia and endothelia. S100A4 is said to reduce occludin expression and could compromise in this way the integrity of the vascular endothelium. This might enhance endothelial permeability. Indeed the ability of tumours to form metastatic deposits in the brain has been attrributed to possible dysfunction of the blood brain barrier. If future work provides substantive evidence for this, this might have potential implication for the management of metastatic disease. But then it ought to be recognised here that gliomas do not normally metastasise to extracranial sites. Glioma cell lines do over express S100A4, but there is little information concerning the tumours. S100A8 and S100A9 have been implicated to impede diapedesis with up regulated expression of the adhesion proteins ICAM-1 and VCAM-1. S100A4 and A9 can form heterodimers in vivo and the heterodimers carry features that resemble S100A9 homodimers in

respect of the ability to bind pro-inflammatory receptors. It is not known if S100A4 and A9 heterodimers and A9 homodimers differ with regard to effects on endothelial permeability.

# Crohn's disease (a form of irritable bowel disease)

#### Note

S100A4 expression is increased in structure fibroblasts of fibrostenosing Crohn's disease promoting intestinal fibroblast migration.

#### Rheumatoid arthritis

#### Note

Increase S100A4 mRNA found in proliferating synovial fibroblasts. Also protein expression up regulated in rheumatoid arthritis synovial tissues and linked with joint invasion. IL-7 and S100A4 occurs in cartilage osteoarthritis and can lead to increased MMP-13 production by chondrocytes. The JAK/STAT/RAGE signalling has been implicated here.

#### Psoriasis

#### Note

Many S100 proteins are found in the dermis. S100A4 is up regulated in the dermis and colossal release of the protein has been reported. Enhanced stabilisation of p53 near cells expressing S100A4 has been noticed. It appears to affect cell proliferation and induce angiogenesis. Suppression of S100A4 using antibodies seems to suppress vascularisation of psoriatic skin xenografts, together with diminution of both number and size of blood vessels. There are no suggestions of cooperative or interactive function of S100A4 with S100A7 (Psoriasin).

## Cardio-vascular, nervous and pulmonary systems

#### Note

The focus in this review is on the role of S100A4 in the disease process. Indeed, S100A4 is expressed in normal as well as in pathological conditions, subserves several physiological functions such as regulating macrophage motility, and participates in fibrosis and tissue remodeling in several diseased and damaged states, e.g. fibrosis of the kidney and loss of renal function, cardiac fibrosis, tissue repair and regeneration and wound healing; central nervous system injury, and pulmonary vascular disease.

S100A4 may be involved in disorders of these systems, but data currently available are somewhat fragmentary. Both S100A4 mRNA and protein are said to be up regulated in the hypertrophic hearts. Up regulation is associated with hypertrophy induced by aortic stenosis or myocardial infarction. In vitro, recombinant S100A4 protein increases the number of viable cardiac myocytes. The ERK1/ERK2 signalling system has been found to be activated in these processes.

Two putative neurotropic motifs have been identified in S100A4 and neuroprotective function has been attributed to the protein.

S100A4 might be associated with PAH (pulmonary arterial hypertension) and putatively linked with 17 $\beta$ -oestradiol modulated S100A4/RAGE signalling. A clinical trial is underway at present to study S100A4 as a marker in PAH treatment (NCT01305252).

## **Breakpoints**

#### Note

A 1q21 breakpoint was described some time ago in renal cell carcinoma (RCC)-associated (X;1)(p11;q21) translocation. This has been mapped to the S100 gene cluster, but its link with S100A4 is uncertain. No translocations involving S100A4 have been recorded. No fusion proteins or hybrid genes involving S100A4 are known.

There are 11 common and 1 rare fragile sites on chromosome 1. The common FRA1F occurs in 1q21. Chromosome 1 is prone to sister chromatid recombination (SCR) and >70% SCRs occur at the fragile sites or in the same band as the fragile sites, but no link with S100A4 has been established.

## To be noted

#### Note

S100A4 expression has been linked with chemoresistance, but the mechanisms involved remain to be elucidated. Whether this occurs via engagement of RAGE by S100A4 and activation of RAGE signalling leading up to chemoresistance is an avenue yet to be explored.

S100A4 activates interacting and multi-functional signalling systems, including EMT signalling systems such as Wnt/ $\beta$ -catenin, NF-kappaB, and E-cadherin among others, highly relevant in the context of cancer. Mediation of its function by osteopontin and potential function of S100 family proteins to act as RAGE ligands are important considerations. These would make S100A4 an eminently valuable chemotherapeutic target. Some downstream effectors of the S100A4 pathways might also lend themselves as targets of interest.

The diversity of biological effects flowing from the inappropriate expression of S100A4 and interactions of S100A4 signalling with several systems which modulate biological response would merit investigations into suppressing S100A4 expression as a possible therapeutic approach. S100A4 function is allied with growth factor and steroid hormone receptors and osteopontin which is itself subject to regulation by Wnt, NF-kappaB

among others, so here is ample provision of options available for therapeutic studies. Also the perceived link up between EMT and S100A4 expression affords fresh avenues of approach.

The antihelminth drug Niclosamide which targets the Wnt/ $\beta$ -catenin pathway can suppress S100A4 expression with parallel inhibition of cell proliferation, migration, promotion of apoptosis, and metastatic spread in vitro and xenograft tumour models.

Xanthohumol is a prenylflavonoid antioxidant, derived from the female flowers of the hops plant (Humulus lupulus). It was identified to possess anticancer properties some while ago. Recent work has shown its ability to suppress cell proliferation, invasion and tumour progression. The flavonoid has been found to inhibit proliferation and invasion in many breast cancer cell lines, including the triple negative breast cancers MDA-MB-231 cells. It induces apoptosis and might be functioning by inhibiting Akt and NF-kappaB activation. The postulated suppression of tumour progression by Xanthohumol is based on assays purportedly resembling intravasation of tumour emboli through defects in the endothelial barrier, so needs much further scrutiny. It would be necessary to perform in vivo studies, ethically undesirable they might be, and ascertain these claims by using stringent criteria to evaluate intravasation of tumour cells and formation of metastases.

It may be noted here that Xanthohumol is said to inhibit cell motility and EMT activation and this is accompanied by inhibition of S100A4 among other determinants. Also of interest is that Xanthohumol induces apoptosis which is caspase-mediated and requires Annexin I. This might be of particular interest since S100A4 as well as other S100A proteins bind to and regulate the function of many target proteins which include annexins. Disruption of Annexin/S100A11 alters the phenotypic behaviour. Annexins display divergent effects on cell proliferation, apoptosis and invasion. The effects may relate to whether the specific S100A is a tumour promoter or suppressor. It would be needless iteration that further investigation is warranted. A clinical study of its pharmacokinetics has been undertaken (NCT01367431).

Phenanthrenes are a class of compounds originally obtained from various members of Orchidaceae and described to possess cytotoxic, antimicrobial and anti-inflammatory activity. They may be therapeutically important in preventing metastasis by inhibiting the interaction of S100A4 with its target molecules (see Bresnick AR patent WO2011146101 A1).

The induction of motility response to S100A4 seems to be mediated by Rho signalling. Rho, Rac and Cdc42 are most prominent players in cytoskeletal reorganization and modulation of cell motility. Of this RhoA has been linked with membrane ruffling and cell motility. Recently S100A4 has been shown to bind to Rhotekin, a RhoA interacting scaffold protein, via the RBD (Rho binding domain), but Rho and S1004 seem to bind to different residues of RBD. The Rho-Rhotekin-S100A4 complex generates the invasive phenotype. At the practical level it is worthy of note and future pursuit that Paclitaxel at low doses well below therapeutic levels has been shown to inhibit S100A4 in the nuclear compartment and in parallel reduce cell migration and invasion in vitro of human cholangiocarcinoma cells in a Rho-GTPase mediated manner.

Membrane bound mucins such as MUC16 (CA125) and MUC2, MUC4, MUC13 have been associated with cancer malignancy. MUC16 is associated with the formation of peritoneal metastases in ovarian cancer. MUCs 2 and 13 are over expressed in pancreatic cancer. MUC4 is overexpressed in oesophageal, lung and colon cancer. Its expression correlates with progression of pancreatic cancer. Experimental suppression of MUC4 in oesophageal cancer cells by shRNA reduces S100A4 expression and reduces cell proliferation and tumorigenic ability as compared with MUC4 expressing parent cells.

The causal linkup is not established; however being integral membrane glycoproteins possibly they anchor S100A4 to the cell membrane and interferes with activation of S100A4 signalling pathways. Growth factors are known to induce phosphoryalation of the cytoplasmic domain of MUC1 and activate nuclear localization of MUC1 and  $\beta$ -catenin and participate in growth factor signalling.

Little is known about the involvement of the immune system in relation to S100A4 function, but there are indications that it may be recruited to NK cell immune synapses and possibly contribute to immune synapse formation. Whether S100A4 participates in restraining NK lytic function or promotes the formation and function of inhibitory synapses is uncertain. The cytoplasmic Src kinases regulate the activating or inhibitory pathways and it has been suggested S100A4 might be capable of influencing the operation of these pathways. CTLs and NK cells induce apoptosis via the pore forming protein perforin and granzyme B pathway. S100A4 may not be involved in this apoptosis pathway, but could be involved in the FasL pathway.

The non-steroidal anti-inflammatory agent sulindac has been found to interfere with Wnt signalling. The  $\beta$ -catenin/TCF transcription complex targets and regulates S100A4. Using an in vivo model involving intrasplenic xenografting of colon cancer cells, sulidac has been shown to down regulate S100A4 promoter activity and expression together with inhibition of Wnt/ $\beta$ -catenin signalling. Phosphosulindac is said to target breast CSCs in vitro and induce apoptosis.

Sulindac is currently being investigated in clinical trial on advanced stage IV colorectal cancer (NCT01856322).

### References

Calabretta B, Battini R, Kaczmarek L, de Riel JK, Baserga R. Molecular cloning of the cDNA for a growth factorinducible gene with strong homology to S-100, a calciumbinding protein. J Biol Chem. 1986 Sep 25;261(27):12628-32

Jackson-Grusby LL, Swiergiel J, Linzer DI. A growthrelated mRNA in cultured mouse cells encodes a placental calcium binding protein. Nucleic Acids Res. 1987 Aug 25;15(16):6677-90

Sherbet GV.. The metastatic spread of cancer. Macmillan Press, Basingstoke London, 1987, ISBN 0-333-44362-4.

Lakshmi MS, Sherbet GV.. Genetic recombination in human melanoma and astrocytoma cell lines involves oncogenes and growth factor genes. Clin Exp Metastasis. 1990 Jan-Feb;8(1):75-87.

Davies BR, Davies MP, Gibbs FE, Barraclough R, Rudland PS.. Induction of the metastatic phenotype by transfection of a benign rat mammary epithelial cell line with the gene for p9Ka, a rat calcium-binding protein, but not with the oncogene EJ-ras-1. Oncogene. 1993 Apr;8(4):999-1008.

Lakshmi MS, Parker C, Sherbet GV.. Metastasis associated MTS1 and NM23 genes affect tubulin polymerisation in B16 melanomas: a possible mechanism of their regulation of metastatic behaviour of tumours. Anticancer Res. 1993 Mar-Apr;13(2):299-303.

Watanabe Y, Usada N, Minami H, Morita T, Tsugane S, Ishikawa R, Kohama K, Tomida Y, Hidaka H.. Calvasculin, as a factor affecting the microfilament assemblies in rat fibroblasts transfected by src gene. FEBS Lett. 1993 Jun 7;324(1):51-5.

Cajone F, Debiasi S, Parker C, Lakshmi MS, Sherbet GV.. Metastasis-associated mts1 gene expression is downregulated by heat shock in variant cell lines of the B16 murine melanoma. Melanoma Res. 1994 Jun;4(3):143-50.

Kriajevska MV, Cardenas MN, Grigorian MS, Ambartsumian NS, Georgiev GP, Lukanidin EM.. Nonmuscle myosin heavy chain as a possible target for protein encoded by metastasis-related mts-1 gene. J Biol Chem. 1994 Aug 5;269(31):19679-82.

Merzak A, Parker C, Koochekpour S, Sherbet GV, Pilkington GJ.. Overexpression of the 18A2/mts1 gene and down-regulation of the TIMP-2 gene in invasive human glioma cell lines in vitro. Neuropathol Appl Neurobiol. 1994 Dec;20(6):614-9.

Parker C, Whittaker PA, Usmani BA, Lakshmi MS, Sherbet GV.. Induction of 18A2/mts1 gene expression and its effects on metastasis and cell cycle control. DNA Cell Biol. 1994 Oct;13(10):1021-8.

Takenaga K, Nakamura Y, Sakiyama S.. Cellular localization of pEL98 protein, an S100-related calcium binding protein, in fibroblasts and its tissue distribution analyzed by monoclonal antibodies. Cell Struct Funct. 1994 Jun;19(3):133-41.

Takenaga K, Nakamura Y, Sakiyama S, Hasegawa Y, Sato K, Endo H.. Binding of pEL98 protein, an S100-

related calcium-binding protein, to nonmuscle tropomyosin. J Cell Biol. 1994 Mar;124(5):757-68.

Ambartsumian N, Tarabykina S, Grigorian M, Tulchinsky E, Hulgaard E, Georgiev G, Lukanidin E.. Characterization of two splice variants of metastasis-associated human mts1 gene. Gene. 1995 Jun 14;159(1):125-30.

Davies M, Harris S, Rudland P, Barraclough R.. Expression of the rat, S-100-related, calcium-binding protein gene, p9Ka, in transgenic mice demonstrates different patterns of expression between these two species. DNA Cell Biol. 1995 Oct;14(10):825-32.

Schafer BW, Wicki R, Engelkamp D, Mattei MG, Heizmann CW.. Isolation of a YAC clone covering a cluster of nine S100 genes on human chromosome 1q21: rationale for a new nomenclature of the S100 calcium-binding protein family. Genomics. 1995 Feb 10;25(3):638-43.

Sherbet GV, Parker C, Usmani BA, Lakshmi MS.. Epidermal growth factor receptor status correlates with cell proliferation-related 18A2/mts1 gene expression in human carcinoma cell lines. Ann N Y Acad Sci. 1995 Sep 30;768:272-6.

Marenholz I, Volz A, Ziegler A, Davies A, Ragoussis I, Korge BP, Mischke D.. Genetic analysis of the epidermal differentiation complex (EDC) on human chromosome 1q21: chromosomal orientation, new markers, and a 6-Mb YAC contig. Genomics. 1996 Nov 1;37(3):295-302.

Mischke D, Korge BP, Marenholz I, Volz A, Ziegler A.. Genes encoding structural proteins of epidermal cornification and S100 calcium-binding proteins form a gene complex ("epidermal differentiation complex") on human chromosome 1q21. J Invest Dermatol. 1996 May;106(5):989-92.

Oates AJ, Barraclough R, Rudland PS.. The role of osteopontin in tumorigenesis and metastasis. Invasion Metastasis. 1997;17(1):1-15. (REVIEW)

Tulchinsky E, Prokhortchouk E, Georgiev G, Lukanidin E.. A kappaB-related binding site is an integral part of the mts1 gene composite enhancer element located in the first intron of the gene. J Biol Chem. 1997 Feb 21;272(8):4828-35.

Albertazzi E, Cajone F, Sherbet GV.. Characterization of a splice variant of metastasis-associated h-mts1 (S100A4) gene expressed in human infiltrating carcinomas of the breast. DNA Cell Biol. 1998a Dec;17(12):1003-8.

Albertazzi E, Cajone F, Leone BE, Naguib RN, Lakshmi MS, Sherbet GV.. Expression of metastasis-associated genes h-mts1 (S100A4) and nm23 in carcinoma of breast is related to disease progression. DNA Cell Biol. 1998b Apr;17(4):335-42.

Lloyd BH, Platt-Higgins A, Rudland PS, Barraclough R.. Human S100A4 (p9Ka) induces the metastatic phenotype upon benign tumour cells. Oncogene. 1998 Jul 30;17(4):465-73.

Sherbet GV, Lakshmi MS.. S100A4 (MTS1) calcium binding protein in cancer growth, invasion and metastasis. Anticancer Res. 1998 Jul-Aug;18(4A):2415-21. (REVIEW)

Cajone F, Sherbet GV.. Stathmin is involved in S100A4mediated regulation of cell cycle progression. Clin Exp Metastasis. 1999;17(10):865-71.

Kriajevska M, Bronstein IB, Scott DJ, Tarabykina S, Fischer-Larsen M, Issinger O, Lukanidin E.. Metastasisassociated protein Mts1 (S100A4) inhibits CK2-mediated phosphorylation and self-assembly of the heavy chain of nonmuscle myosin. Biochim Biophys Acta. 2000 Dec 20;1498(2-3):252-63.

Mayr B, Brem G, Reifinger M.. Absence of S100A4 (mts1) gene mutations in various canine and feline tumours. Detection of a polymorphism in feline S100A4 (mts1). J Vet Med A Physiol Pathol Clin Med. 2000 Mar;47(2):123-8.

Rudland PS, Platt-Higgins A, Renshaw C, West CR, Winstanley JH, Robertson L, Barraclough R.. Prognostic significance of the metastasis-inducing protein S100A4 (p9Ka) in human breast cancer. Cancer Res. 2000 Mar 15;60(6):1595-603.

Ambartsumian N, Klingelhofer J, Grigorian M, Christensen C, Kriajevska M, Tulchinsky E, Georgiev G, Berezin V, Bock E, Rygaard J, Cao R, Cao Y, Lukanidin E.. The metastasis-associated Mts1(S100A4) protein could act as an angiogenic factor. Oncogene. 2001 Aug 2;20(34):4685-95.

Chen H, Fernig DG, Rudland PS, Sparks A, Wilkinson MC, Barraclough R.. Binding to intracellular targets of the metastasis-inducing protein, S100A4 (p9Ka). Biochem Biophys Res Commun. 2001 Sep 7;286(5):1212-7.

Cohn MA, Hjelmso I, Wu LC, Guldberg P, Lukanidin EM, Tulchinsky EM.. Characterization of Sp1, AP-1, CBF and KRC binding sites and minisatellite DNA as functional elements of the metastasis-associated mts1/S100A4 gene intronic enhancer. Nucleic Acids Res. 2001 Aug 15;29(16):3335-46.

Donato R.. S100: a multigenic family of calcium-modulated proteins of the EF-hand type with intracellular and extracellular functional roles. Int J Biochem Cell Biol. 2001 Jul;33(7):637-68. (REVIEW)

Ninomiya I, Ohta T, Fushida S, Endo Y, Hashimoto T, Yagi M, Fujimura T, Nishimura G, Tani T, Shimizu K, Yonemura Y, Heizmann CW, Schafer BW, Sasaki T, Miwa K... Increased expression of S100A4 and its prognostic significance in esophageal squamous cell carcinoma. Int J Oncol. 2001 Apr;18(4):715-20.

Sherbet GV.. Calcium signalling in cancer. CRC Press, Boca Raton, New York Washington. 2001.

Ibrahim SO, Lillehaug JR, Dolphine O, Johnson NW, Warnakulasuriya KA, Vasstrand EN.. Mutations of the cell cycle arrest gene p21WAF1, but not the metastasisinducing gene S100A4, are frequent in oral squamous cell carcinomas from Sudanese toombak dippers and nonsnuff-dippers from the Sudan, Scandinavia, USA and UK. Anticancer Res. 2002 May-Jun;22(3):1445-51.

Mazzucchelli L.. Protein S100A4: too long overlooked by pathologists? Am J Pathol. 2002 Jan;160(1):7-13.

Vallely KM, Rustandi RR, Ellis KC, Varlamova O, Bresnick AR, Weber DJ.. Solution structure of human Mts1 (S100A4) as determined by NMR spectroscopy. Biochemistry. 2002 Oct 22;41(42):12670-80.

Andersen K, Smith-Sorensen B, Pedersen KB, Hovig E, Myklebost O, Fodstad O, Maelandsmo GM.. Interferongamma suppresses S100A4 transcription independently of apoptosis or cell cycle arrest. Br J Cancer. 2003 Jun 16;88(12):1995-2001.

Grigorian M, Lukanidin E.. [Activator of metastasis in cancer cells, Mst1/S100A4 protein binds to tumor suppressor protein p53]. Genetika. 2003 Jul;39(7):900-8.

Hernan R, Fasheh R, Calabrese C, Frank AJ, Maclean KH, Allard D, Barraclough R, Gilbertson RJ.. ERBB2 upregulates S100A4 and several other prometastatic genes in medulloblastoma. Cancer Res. 2003 Jan 1;63(1):140-8. Li ZH, Spektor A, Varlamova O, Bresnick AR.. Mts1 regulates the assembly of nonmuscle myosin-IIA. Biochemistry. 2003 Dec 9;42(48):14258-66.

Andersen K, Nesland JM, Holm R, Florenes VA, Fodstad O, Maelandsmo GM.. Expression of S100A4 combined with reduced E-cadherin expression predicts patient outcome in malignant melanoma. Mod Pathol. 2004 Aug;17(8):990-7.

Lee WY, Su WC, Lin PW, Guo HR, Chang TW, Chen HH.. Expression of S100A4 and Met: potential predictors for metastasis and survival in early-stage breast cancer. Oncology. 2004;66(6):429-38.

Schmidt-Hansen B, Klingelhofer J, Grum-Schwensen B, Christensen A, Andresen S, Kruse C, Hansen T, Ambartsumian N, Lukanidin E, Grigorian M.. Functional significance of metastasis-inducing S100A4(Mts1) in tumor-stroma interplay. J Biol Chem. 2004 Jun 4;279(23):24498-504. Epub 2004 Mar 26.

Cho YG, Kim CJ, Nam SW, Yoon SH, Lee SH, Yoo NJ, Lee JY, Park WS.. Overexpression of S100A4 is closely associated with progression of colorectal cancer. World J Gastroenterol. 2005 Aug 21;11(31):4852-6.

Dulyaninova NG, Malashkevich VN, Almo SC, Bresnick AR.. Regulation of myosin-IIA assembly and Mts1 binding by heavy chain phosphorylation. Biochemistry. 2005 May 10;44(18):6867-76.

Fernandez-Fernandez MR, Veprintsev DB, Fersht AR.. Proteins of the S100 family regulate the oligomerization of p53 tumor suppressor. Proc Natl Acad Sci U S A. 2005 Mar 29;102(13):4735-40. Epub 2005 Mar 21.

Ikoma N, Yamazaki H, Abe Y, Oida Y, Ohnishi Y, Suemizu H, Matsumoto H, Matsuyama T, Ohta Y, Ozawa A, Ueyama Y, Nakamura M.. S100A4 expression with reduced E-cadherin expression predicts distant metastasis of human malignant melanoma cell lines in the NOD/SCID/gammaCnull (NOG) mouse model. Oncol Rep. 2005 Sep;14(3):633-7.

Moriyama-Kita M, Endo Y, Yonemura Y, Heizmann CW, Miyamori H, Sato H, Yamamoto E, Sasaki T.. S100A4 regulates E-cadherin expression in oral squamous cell carcinoma. Cancer Lett. 2005 Dec 18;230(2):211-8.

Uchikoga N, Takahashi SY, Ke R, Sonoyama M, Mitaku S.. Electric charge balance mechanism of extended soluble proteins. Protein Sci. 2005 Jan;14(1):74-80. Epub 2004 Dec 2.

Viemann D, Strey A, Janning A, Jurk K, Klimmek K, Vogl T, Hirono K, Ichida F, Foell D, Kehrel B, Gerke V, Sorg C, Roth J.. Myeloid-related proteins 8 and 14 induce a specific inflammatory response in human microvascular endothelial cells. Blood. 2005 Apr 1;105(7):2955-62. Epub 2004 Dec 14.

Zhang S, Wang G, Liu D, Bao Z, Fernig DG, Rudland PS, Barraclough R.. The C-terminal region of S100A4 is important for its metastasis-inducing properties. Oncogene. 2005 Jun 23;24(27):4401-11.

de Silva Rudland S, Martin L, Roshanlall C, Winstanley J, Leinster S, Platt-Higgins A, Carroll J, West C, Barraclough R, Rudland P.. Association of S100A4 and osteopontin with specific prognostic factors and survival of patients with minimally invasive breast cancer. Clin Cancer Res. 2006 Feb 15;12(4):1192-200.

Garrett SC, Varney KM, Weber DJ, Bresnick AR.. S100A4, a mediator of metastasis. J Biol Chem. 2006 Jan 13;281(2):677-80. Epub 2005 Oct 21. (REVIEW) Hemandas AK, Salto-Tellez M, Maricar SH, Leong AF, Leow CK.. Metastasis-associated protein S100A4--a potential prognostic marker for colorectal cancer. J Surg Oncol. 2006 May 1;93(6):498-503.

Kikuchi N, Horiuchi A, Osada R, Imai T, Wang C, Chen X, Konishi I.. Nuclear expression of S100A4 is associated with aggressive behavior of epithelial ovarian carcinoma: an important autocrine/paracrine factor in tumor progression. Cancer Sci. 2006 Oct;97(10):1061-9.

Oida Y, Yamazaki H, Tobita K, Mukai M, Ohtani Y, Miyazaki N, Abe Y, Imaizumi T, Makuuchi H, Ueyama Y, Nakamura M.. Increased S100A4 expression combined with decreased E-cadherin expression predicts a poor outcome of patients with pancreatic cancer. Oncol Rep. 2006 Sep;16(3):457-63.

Ren J, Raina D, Chen W, Li G, Huang L, Kufe D.. MUC1 oncoprotein functions in activation of fibroblast growth factor receptor signaling. Mol Cancer Res. 2006 Nov;4(11):873-83.

Sherbet GV, Lakshmi MS.. The modus operandi of S100A4 signalling in cancer growth, progression and prognosis. The molecular and cellular pathology of cancer progression and prognosis. Sherbet GV (ed.), 2006; pp. 359-377.

Sjoblom T, Jones S, Wood LD, Parsons DW, Lin J, Barber TD, Mandelker D, Leary RJ, Ptak J, Silliman N, Szabo S, Buckhaults P, Farrell C, Meeh P, Markowitz SD, Willis J, Dawson D, Willson JK, Gazdar AF, Hartigan J, Wu L, Liu C, Parmigiani G, Park BH, Bachman KE, Papadopoulos N, Vogelstein B, Kinzler KW, Velculescu VE.. The consensus coding sequences of human breast and colorectal cancers. Science. 2006 Oct 13;314(5797):268-74. Epub 2006 Sep 7.

Stein U, Arlt F, Walther W, Smith J, Waldman T, Harris ED, Mertins SD, Heizmann CW, Allard D, Birchmeier W, Schlag PM, Shoemaker RH.. The metastasis-associated gene S100A4 is a novel target of beta-catenin/T-cell factor signaling in colon cancer. Gastroenterology. 2006 Nov;131(5):1486-500. Epub 2006 Aug 22.

Boye K, Andersen K, Tveito S, Oyjord T, Maelandsmo GM.. Interferon-gamma-induced suppression of S100A4 transcription is mediated by the class II transactivator. Tumour Biol. 2007;28(1):27-35. Epub 2006 Dec 12.

Chen PS, Wang MY, Wu SN, Su JL, Hong CC, Chuang SE, Chen MW, Hua KT, Wu YL, Cha ST, Babu MS, Chen CN, Lee PH, Chang KJ, Kuo ML.. CTGF enhances the motility of breast cancer cells via an integrin-alphavbeta3-ERK1/2-dependent S100A4-upregulated pathway. J Cell Sci. 2007 Jun 15;120(Pt 12):2053-65.

Debacker K, Kooy RF.. Fragile sites and human disease. Hum Mol Genet. 2007 Oct 15;16 Spec No. 2:R150-8. Epub 2007 Jun 13. (REVIEW)

Mahon PC, Baril P, Bhakta V, Chelala C, Caulee K, Harada T, Lemoine NR.. S100A4 contributes to the suppression of BNIP3 expression, chemoresistance, and inhibition of apoptosis in pancreatic cancer. Cancer Res. 2007 Jul 15;67(14):6786-95.

Matsumoto K, Irie A, Satoh T, Ishii J, Iwabuchi K, Iwamura M, Egawa S, Baba S.. Expression of S100A2 and S100A4 predicts for disease progression and patient survival in bladder cancer. Urology. 2007 Sep;70(3):602-7. Epub 2007 Aug 3.

Pleger ST, Most P, Katus HA.. S100 proteins: a missing piece in the puzzle of heart failure? Cardiovasc Res. 2007 Jul 1;75(1):1-2. Epub 2007 May 10.

Moniaux N, Chaturvedi P, Van Seuningen I, Porchet N, Singh AP, Batra SK.. MUC4 (mucin 4, cell surface associated) Atlas Genet Cytogenet Oncol Haematol. 2007; 11(3):201-206. URL: http://AtlasGeneticsOncology.org/Genes/MUC4ID41459ch 3q29.html

Schneider M, Kostin S, Strom CC, Aplin M, Lyngbaek S, Theilade J, Grigorian M, Andersen CB, Lukanidin E, Lerche Hansen J, Sheikh SP.. S100A4 is upregulated in injured myocardium and promotes growth and survival of cardiac myocytes. Cardiovasc Res. 2007 Jul 1;75(1):40-50. Epub 2007 Apr 6.

Boye K, Grotterod I, Aasheim HC, Hovig E, Maelandsmo GM.. Activation of NF-kappaB by extracellular S100A4: analysis of signal transduction mechanisms and identification of target genes. Int J Cancer. 2008 Sep 15;123(6):1301-10.

El-Abd E, El-Tahan R, Fahmy L, Zaki S, Faid W, Sobhi A, Kandil K, El-Kwisky F.. Serum metastasin mRNA is an important survival predictor in breast cancer. Br J Biomed Sci. 2008;65(2):90-4.

Grigorian M, Ambartsumian N, Lukanidin E.. Metastasisinducing S100A4 protein: implication in non-malignant human pathologies. Curr Mol Med. 2008 Sep;8(6):492-6. (REVIEW)

Haugen MH, Flatmark K, Mikalsen SO, Malandsmo GM.. The metastasis-associated protein S100A4 exists in several charged variants suggesting the presence of posttranslational modifications. BMC Cancer. 2008 Jun 13;8:172.

Ismail TM, Fernig DG, Rudland PS, Terry CJ, Wang G, Barraclough R.. The basic C-terminal amino acids of calcium-binding protein S100A4 promote metastasis. Carcinogenesis. 2008 Dec;29(12):2259-66. Epub 2008 Sep 10.

Li Y, Zhang KL, Sun Y, Yang Y, Chen XY, Kong QY, Wu ML, Liu J, Li H.. Frequent S100A4 Expression with Unique Splicing Pattern in Gastric Cancers: A Hypomethylation Event Paralleled with E-cadherin Reduction and Wnt Activation. Transl Oncol. 2008 Dec;1(4):165-76.

Oslejskova L, Grigorian M, Gay S, Neidhart M, Senolt L.. The metastasis associated protein S100A4: a potential novel link to inflammation and consequent aggressive behaviour of rheumatoid arthritis synovial fibroblasts. Ann Rheum Dis. 2008 Nov;67(11):1499-504. Epub 2007 Dec 4. (REVIEW)

Parsons DW, Jones S, Zhang X, Lin JC, Leary RJ, Angenendt P, Mankoo P, Carter H, Siu IM, Gallia GL, Olivi A, McLendon R, Rasheed BA, Keir S, Nikolskaya T, Nikolsky Y, Busam DA, Tekleab H, Diaz LA Jr, Hartigan J, Smith DR, Strausberg RL, Marie SK, Shinjo SM, Yan H, Riggins GJ, Bigner DD, Karchin R, Papadopoulos N, Parmigiani G, Vogelstein B, Velculescu VE, Kinzler KW.. An integrated genomic analysis of human glioblastoma multiforme. Science. 2008 Sep 26;321(5897):1807-12. Epub 2008 Sep 4.

Pathuri P, Vogeley L, Luecke H.. Crystal structure of metastasis-associated protein S100A4 in the active calcium-bound form. J Mol Biol. 2008 Oct 31;383(1):62-77. doi: 10.1016/j.jmb.2008.04.076. Epub 2008 May 7.

Schneider M, Hansen JL, Sheikh SP.. S100A4: a common mediator of epithelial-mesenchymal transition, fibrosis and regeneration in diseases? J Mol Med (Berl). 2008 May;86(5):507-22. doi: 10.1007/s00109-007-0301-3. Epub 2008 Mar 6. (REVIEW)

Antonyak MA, Li B, Regan AD, Feng Q, Dusaban SS, Cerione RA.. Tissue transglutaminase is an essential participant in the epidermal growth factor-stimulated signaling pathway leading to cancer cell migration and invasion. J Biol Chem. 2009 Jul 3;284(27):17914-25. doi: 10.1074/jbc.M109.013037. Epub 2009 Apr 29.

Chen M, Sinha M, Luxon BA, Bresnick AR, O'Connor KL.. Integrin alpha6beta4 controls the expression of genes associated with cell motility, invasion, and metastasis, including S100A4/metastasin. J Biol Chem. 2009 Jan 16;284(3):1484-94. Epub 2008 Nov 14.

Ikenaga N, Ohuchida K, Mizumoto K, Yu J, Fujita H, Nakata K, Ueda J, Sato N, Nagai E, Tanaka M.. S100A4 mRNA is a diagnostic and prognostic marker in pancreatic carcinoma. J Gastrointest Surg. 2009 Oct;13(10):1852-8. Epub 2009 Aug 4.

Kim JH, Kim CN, Kim SY, Lee JS, Cho D, Kim JW, Yoon SY.. Enhanced S100A4 protein expression is clinicopathologically significant to metastatic potential and p53 dysfunction in colorectal cancer. Oncol Rep. 2009 Jul;22(1):41-7.

Klingelhofer J, Moller HD, Sumer EU, Berg CH, Poulsen M, Kiryushko D, Soroka V, Ambartsumian N, Grigorian M, Lukanidin EM.. Epidermal growth factor receptor ligands as new extracellular targets for the metastasis-promoting S100A4 protein. FEBS J. 2009 Oct;276(20):5936-48. Epub 2009 Sep 9.

Maelandsmo GM, Florenes VA, Nguyen MT, Flatmark K, Davidson B.. Different expression and clinical role of S100A4 in serous ovarian carcinoma at different anatomic sites. Tumour Biol. 2009;30(1):15-25. Epub 2009 Feb 5.

Maruko A, Ohtake Y, Katoh S, Ohkubo Y... Transglutaminase down-regulates the dimerization of epidermal growth factor receptor in rat perivenous and periportal hepatocytes. Cell Prolif. 2009 Oct;42(5):647-56. doi: 10.1111/j.1365-2184.2009.00622.x. Epub 2009 Jul 9.

Sherbet GV.. Metastasis promoter S100A4 is a potentially valuable molecular target for cancer therapy. Cancer Lett. 2009 Jul 18;280(1):15-30. Epub 2008 Dec 6. (REVIEW)

Tsuna M, Kageyama S, Fukuoka J, Kitano H, Doki Y, Tezuka H, Yasuda H.. Significance of S100A4 as a prognostic marker of lung squamous cell carcinoma. Anticancer Res. 2009 Jul;29(7):2547-54.

van Dieck J, Teufel DP, Jaulent AM, Fernandez-Fernandez MR, Rutherford TJ, Wyslouch-Cieszynska A, Fersht AR.. Posttranslational modifications affect the interaction of S100 proteins with tumor suppressor p53. J Mol Biol. 2009 Dec 18;394(5):922-30. Epub 2009 Oct 9.

Yammani RR, Long D, Loeser RF.. Interleukin-7 stimulates secretion of S100A4 by activating the JAK/STAT signaling pathway in human articular chondrocytes. Arthritis Rheum. 2009 Mar;60(3):792-800.

Cunningham MF, Docherty NG, Burke JP, O'Connell PR.. S100A4 expression is increased in stricture fibroblasts from patients with fibrostenosing Crohn's disease and promotes intestinal fibroblast migration. Am J Physiol Gastrointest Liver Physiol. 2010 Aug;299(2):G457-66. Epub 2010 May 20.

Grotterod I, Maelandsmo GM, Boye K.. Signal transduction mechanisms involved in S100A4-induced activation of the transcription factor NF-kappaB. BMC Cancer. 2010 May 28;10:241. doi: 10.1186/1471-2407-10-241.

Hua J, Chen D, Fu H, Zhang R, Shen W, Liu S, Sun K, Sun X.. Short hairpin RNA-mediated inhibition of S100A4  $\,$ 

promotes apoptosis and suppresses proliferation of BGC823 gastric cancer cells in vitro and in vivo. Cancer Lett. 2010 Jun 1;292(1):41-7. doi: 10.1016/j.canlet.2009.11.007. Epub 2009 Nov 28.

Li ZH, Dulyaninova NG, House RP, Almo SC, Bresnick AR.. S100A4 regulates macrophage chemotaxis. Mol Biol Cell. 2010 Aug 1;21(15):2598-610. doi: 10.1091/mbc.E09-07-0609. Epub 2010 Jun 2.

Miranda KJ, Loeser RF, Yammani RR.. Sumoylation and nuclear translocation of S100A4 regulate IL-1betamediated production of matrix metalloproteinase-13. J Biol Chem. 2010 Oct 8;285(41):31517-24. doi: 10.1074/jbc.M110.125898. Epub 2010 Aug 4.

Ismail TM, Zhang S, Fernig DG, Gross S, Martin-Fernandez ML, See V, Tozawa K, Tynan CJ, Wang G, Wilkinson MC, Rudland PS, Barraclough R.. Selfassociation of calcium-binding protein S100A4 and metastasis. J Biol Chem. 2010 Jan 8;285(2):914-22. Epub 2009 Nov 16.

Malashkevich VN, Dulyaninova NG, Ramagopal UA, Liriano MA, Varney KM, Knight D, Brenowitz M, Weber DJ, Almo SC, Bresnick AR.. Phenothiazines inhibit S100A4 function by inducing protein oligomerization. Proc Natl Acad Sci U S A. 2010 May 11;107(19):8605-10. doi: 10.1073/pnas.0913660107. Epub 2010 Apr 26.

Matsuura I, Lai CY, Chiang KN.. Functional interaction between Smad3 and S100A4 (metastatin-1) for TGF-betamediated cancer cell invasiveness. Biochem J. 2010 Feb 24;426(3):327-35.

Mencia N, Selga E, Rico I, de Almagro MC, Villalobos X, Ramirez S, Adan J, Hernandez JL, Noe V, Ciudad CJ.. Overexpression of S100A4 in human cancer cell lines resistant to methotrexate. BMC Cancer. 2010 Jun 1;10:250.

van Dieck J, Lum JK, Teufel DP, Fersht AR.. S100 proteins interact with the N-terminal domain of MDM2. FEBS Lett. 2010 Aug 4;584(15):3269-74. Epub 2010 Jun 19.

Wang HY, Zhang JY, Cui JT, Tan XH, Li WM, Gu J, Lu YY.. Expression status of S100A14 and S100A4 correlates with metastatic potential and clinical outcome in colorectal cancer after surgery. Oncol Rep. 2010 Jan;23(1):45-52.

Wang YY, Ye ZY, Zhao ZS, Tao HQ, Chu YQ.. High-level expression of S100A4 correlates with lymph node metastasis and poor prognosis in patients with gastric cancer. Ann Surg Oncol. 2010 Jan;17(1):89-97. Epub 2009 Oct 10.

Zibert JR, Skov L, Thyssen JP, Jacobsen GK, Grigorian M.. Significance of the S100A4 protein in psoriasis. J Invest Dermatol. 2010 Jan;130(1):150-60.

Berge G, Pettersen S, Grotterod I, Bettum IJ, Boye K, Maelandsmo GM.. Osteopontin--an important downstream effector of S100A4-mediated invasion and metastasis. Int J Cancer. 2011 Aug 15;129(4):780-90. doi: 10.1002/ijc.25735. Epub 2011 Mar 11.

Bruyere E, Jonckheere N, Frenois F, Mariette C, Van Seuningen I.. The MUC4 membrane-bound mucin regulates esophageal cancer cell proliferation and migration properties: Implication for S100A4 protein. Biochem Biophys Res Commun. 2011 Sep 23;413(2):325-9. doi: 10.1016/j.bbrc.2011.08.095. Epub 2011 Aug 26.

Cerezo LA, Kuncova K, Mann H, Tomcik M, Zamecnik J, Lukanidin E, Neidhart M, Gay S, Grigorian M, Vencovsky J, Senolt L.. The metastasis promoting protein S100A4 is increased in idiopathic inflammatory myopathies. Rheumatology (Oxford). 2011 Oct;50(10):1766-72. doi: 10.1093/rheumatology/ker218. Epub 2011 Jun 28.

Dempsie Y, Nilsen M, White K, Mair KM, Loughlin L, Ambartsumian N, Rabinovitch M, Maclean MR.. Development of pulmonary arterial hypertension in mice over-expressing S100A4/Mts1 is specific to females. Respir Res. 2011 Dec 20;12:159. doi: 10.1186/1465-9921-12-159.

Lu W, Lin C, Roberts MJ, Waud WR, Piazza GA, Li Y.. Niclosamide suppresses cancer cell growth by inducing Wnt co-receptor LRP6 degradation and inhibiting the Wnt/b-catenin pathway. PLoS One. 2011;6(12):e29290. doi: 10.1371/journal.pone.0029290. Epub 2011 Dec 16.

Mace EM, Orange JS.. Multiple distinct NK-cell synapses. Blood. 2011 Dec 15;118(25):6475-6. doi: 10.1182/blood-2011-10-381392.

Nishioku T, Furusho K, Tomita A, Ohishi H, Dohgu S, Shuto H, Yamauchi A, Kataoka Y.. Potential role for S100A4 in the disruption of the blood-brain barrier in collagen-induced arthritic mice, an animal model of rheumatoid arthritis. Neuroscience. 2011 Aug 25;189:286-92. doi: 10.1016/j.neuroscience.2011.05.044. Epub 2011 May 26.

Osada T, Chen M, Yang XY, Spasojevic I, Vandeusen JB, Hsu D, Clary BM, Clay TM, Chen W, Morse MA, Lyerly HK.. Antihelminth compound niclosamide downregulates Wnt signaling and elicits antitumor responses in tumors with activating APC mutations. Cancer Res. 2011 Jun 15;71(12):4172-82. doi: 10.1158/0008-5472.CAN-10-3978. Epub 2011 Apr 29.

Sack U, Walther W, Scudiero D, Selby M, Kobelt D, Lemm M, Fichtner I, Schlag PM, Shoemaker RH, Stein U.. Novel effect of antihelminthic Niclosamide on S100A4-mediated metastatic progression in colon cancer. J Natl Cancer Inst. 2011 Jul 6;103(13):1018-36. doi: 10.1093/jnci/djr190. Epub 2011 Jun 17.

Sherbet GV.. Growth factors and their receptors in cell differentiation, cancer and cancer therapy. E-book; 2011 July 12; Elsevier

Stein U, Arlt F, Smith J, Sack U, Herrmann P, Walther W, Lemm M, Fichtner I, Shoemaker RH, Schlag PM.. Intervening in beta-catenin signaling by sulindac inhibits S100A4-dependent colon cancer metastasis. Neoplasia. 2011 Feb;13(2):131-44.

Boye K, Nesland JM, Sandstad B, Haugland Haugen M, Maelandsmo GM, Flatmark K.. EMMPRIN is associated with S100A4 and predicts patient outcome in colorectal cancer. Br J Cancer. 2012 Aug 7;107(4):667-74. doi: 10.1038/bjc.2012.293. Epub 2012 Jul 10.

Hibino T, Sakaguchi M, Miyamoto S, Yamamoto M, Motoyama A, Hosoi J, Shimokata T, Ito T, Tsuboi R, Huh NH.. S100A9 is a novel ligand of EMMPRIN that promotes melanoma metastasis. Cancer Res. 2013 Jan 1;73(1):172-83. doi: 10.1158/0008-5472.CAN-11-3843. Epub 2012 Nov 7.

Chen D, Zheng XF, Yang ZY, Liu DX, Zhang GY, Jiao XL, Zhao H.. S100A4 silencing blocks invasive ability of esophageal squamous cell carcinoma cells. World J Gastroenterol. 2012 Mar 7;18(9):915-22. doi: 10.3748/wjg.v18.i9.915.

Dmytriyeva O, Pankratova S, Owczarek S, Sonn K, Soroka V, Ridley CM, Marsolais A, Lopez-Hoyos M, Ambartsumian N, Lukanidin E, Bock E, Berezin V, Kiryushko D.. The metastasis-promoting S100A4 protein

confers neuroprotection in brain injury. Nat Commun. 2012;3:1197. doi: 10.1038/ncomms2202.

Du M, Wang G, Ismail TM, Gross S, Fernig DG, Barraclough R, Rudland PS.. S100P dissociates myosin IIA filaments and focal adhesion sites to reduce cell adhesion and enhance cell migration. J Biol Chem. 2012 May 4;287(19):15330-44. doi: 10.1074/jbc.M112.349787. Epub 2012 Mar 6.

Fabris L, Cadamuro M, Sambado L, Beretta I, Spirli C, IndraccoloS, Strazzabosco M.. Selective reduction in S100A4 nuclear expression by low dose paclitaxel halts invasiveness of human cholangiocarcinoma cells through a Rho-A/Cdc42 dependent mechanism. J Hepatol 2012 April; 56, Suppl 2, S112.

Horiuchi A, Hayashi T, Kikuchi N, Hayashi A, Fuseya C, Shiozawa T, Konishi I.. Hypoxia upregulates ovarian cancer invasiveness via the binding of HIF-1? to a hypoxia-induced, methylation-free hypoxia response element of S100A4 gene. Int J Cancer. 2012 Oct 15;131(8):1755-67. doi: 10.1002/ijc.27448. Epub 2012 Mar 14.

Li X, Baskin JG, Mangan EK, Su K, Gibson AW, Ji C, Edberg JC, Kimberly RP.. The unique cytoplasmic domain of human Fc?RIIIA regulates receptor-mediated function. J Immunol. 2012 Nov 1;189(9):4284-94. doi: 10.4049/jimmunol.1200704. Epub 2012 Sep 28.

Liu LP, Ho RL, Chen GG, Lai PB.. Sorafenib inhibits hypoxia-inducible factor-1? synthesis: implications for antiangiogenic activity in hepatocellular carcinoma. Clin Cancer Res. 2012 Oct 15;18(20):5662-71. doi: 10.1158/1078-0432.CCR-12-0552. Epub 2012 Aug 28.

Zhu C, Cheng KW, Ouyang N, Huang L, Sun Y, Constantinides P, Rigas B.. Phosphosulindac (OXT-328) selectively targets breast cancer stem cells in vitro and in human breast cancer xenografts. Stem Cells. 2012 Oct;30(10):2065-75. doi: 10.1002/stem.1139.

Bresnick AR.. Preventing or inhibiting tumor metastasis in subject involves administering aromatic compound including 9,10-dihydro-phenanthrene derivative, (1,4) naphthoquinone derivative, quinoline-5,8-dione derivative, to the subject. US Patent WO2011146101-A1; US2013090355-A1, 2013 Apr 11.

Chen M, Bresnick AR, O'Connor KL.. Coupling S100A4 to Rhotekin alters Rho signaling output in breast cancer cells. Oncogene. 2013 Aug 8;32(32):3754-64. doi: 10.1038/onc.2012.383. Epub 2012 Sep 10.

Kawakita T, Espana EM, Higa K, Kato N, Li W, Tseng SC.. Activation of Smad-mediated TGF-? signaling triggers epithelial-mesenchymal transitions in murine cloned corneal progenitor cells. J Cell Physiol. 2013 Jan;228(1):225-34. doi: 10.1002/jcp.24126.

Nomura M, Tanaka K, Wang L, Goto Y, Mukasa C, Ashida K, Takayanagi R.. Activin type IB receptor signaling in prostate cancer cells promotes lymph node metastasis in a xenograft model. Biochem Biophys Res Commun. 2013 Jan 4;430(1):340-6. doi: 10.1016/j.bbrc.2012.11.011. Epub 2012 Nov 15.

Scheiter M, Lau U, van Ham M, Bulitta B, Grobe L, Garritsen H, Klawonn F, Konig S, Jansch L.. Proteome analysis of distinct developmental stages of human natural killer (NK) cells. Mol Cell Proteomics. 2013 May;12(5):1099-114. doi: 10.1074/mcp.M112.024596. Epub 2013 Jan 13.

Sherbet GV.. Therapeutic strategies in cancer biology and pathology. E-book 2013 Aug; Elsevier.

Tamaki Y, Iwanaga Y, Niizuma S, Kawashima T, Kato T, Inuzuka Y, Horie T, Morooka H, Takase T, Akahashi Y, Kobuke K, Ono K, Shioi T, Sheikh SP, Ambartsumian N, Lukanidin E, Koshimizu TA, Miyazaki S, Kimura T.. Metastasis-associated protein, S100A4 mediates cardiac fibrosis potentially through the modulation of p53 in cardiac fibroblasts. J Mol Cell Cardiol. 2013 Apr;57:72-81. doi: 10.1016/j.yjmcc.2013.01.007. Epub 2013 Jan 23.

Tsukamoto N, Egawa S, Akada M, Abe K, Saiki Y, Kaneko N, Yokoyama S, Shima K, Yamamura A, Motoi F, Abe H, Hayashi H, Ishida K, Moriya T, Tabata T, Kondo E, Kanai N, Gu Z, Sunamura M, Unno M, Horii A.. The expression of S100A4 in human pancreatic cancer is associated with invasion. Pancreas. 2013 Aug;42(6):1027-33. doi: 10.1097/MPA.0b013e31828804e7.

Viola K, Kopf S, Rarova L, Jarukamjorn K, Kretschy N, Teichmann M, Vonach C, Atanasov AG, Giessrigl B, Huttary N, Raab I, Krieger S, Strnad M, de Martin R, Saiko P, Szekeres T, Knasmuller S, Dirsch VM, Jager W, Grusch M, Dolznig H, Mikulits W, Krupitza G.. Xanthohumol attenuates tumour cell-mediated breaching of the lymphendothelial barrier and prevents intravasation and metastasis. Arch Toxicol. 2013 Jul;87(7):1301-12. doi: 10.1007/s00204-013-1028-2. Epub 2013 Mar 17. Wang Z, Griffin M.. The role of TG2 in regulating S100A4mediated mammary tumour cell migration. PLoS One. 2013;8(3):e57017. doi: 10.1371/journal.pone.0057017. Epub 2013 Mar 1.

Yu CC, Tsai CH, Hsu HI, Chang YC.. Elevation of S100A4 expression in buccal mucosal fibroblasts by arecoline: involvement in the pathogenesis of oral submucous fibrosis. PLoS One. 2013;8(1):e55122. doi: 10.1371/journal.pone.0055122. Epub 2013 Jan 31.

Zhang J, Antonyak MA, Singh G, Cerione RA.. A mechanism for the upregulation of EGF receptor levels in glioblastomas. Cell Rep. 2013 Jun 27;3(6):2008-20. doi: 10.1016/j.celrep.2013.05.021. Epub 2013 Jun 13.

Zheng X, Gai X, Wu Z, Liu Q, Yao Y.. Metastasin leads to poor prognosis of hepatocellular carcinoma through partly inducing EMT. Oncol Rep. 2013 May;29(5):1811-8. doi: 10.3892/or.2013.2341. Epub 2013 Mar 12.

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