Keywords:

T-SPOT TB

Interferon γ -based assays

QuantiFERON-TB Gold In-Tube

Enferm Infecc Microbiol Clin. 2011;29(Supl 3):25-32



Diagnosis of tuberculosis infection using interferon-γ-based assays

Miguel Santín Cerezales^{a,*} and José Domínguez Benítez^b

^aServicio de Enfermedades Infecciosas, Hospital Universitario de Bellvitge, IDIBELL, Departamento de Ciencias Clínicas, Universidad de Barcelona, Red Española de Investigación en Patología Infecciosa (REIPI), Instituto de Salud Carlos III, Barcelona, Spain

^bServicio de Microbiología, Instituto de Investigación en Ciencias de la Salud Germans Trias i Pujol, Universidad Autónoma de Barcelona, Ciber de Enfermedades Respiratorias, Instituto de Salud Carlos III, Badalona, Barcelona, Spain

ABSTRACT

Interferon- γ -based assays, collectively known as IFN- γ release assays (IGRAs), have emerged as a reliable alternative to the old tuberculin skin test (TST) for the immunodiagnosis of tuberculosis (TB) infection. The 2 commercially available tests, the enzyme-linked immunosorbent assay (ELISA), QuantiFERON-TB Gold Intube (QFI-IT), and the enzyme-linked immunospot assay (ELISPOT), T-SPOT.TB, are more accurate than TST for the diagnosis of TB, since they are highly specific and correlate better with the existence of risk factors for the infection. According to the available data, T-SPOT.TB obtains a higher number of positive results than QFT-IT, while its specificity is lower. Although the sensitivity of the IFN- γ -based assays may be impaired to some extent by cellular immunosuppression and extreme ages of life, they perform better than TST in these situations. Data from longitudinal studies suggest that IFN- γ -based tests are better predictors of subsequent development of active TB than TST; however this prognostic value has not been consistently demonstrated. This review focuses on the clinical use of the IFN- γ -based tests in different risk TB groups, and notes the main limitations and areas for future development.

Diagnóstico de la tuberculosis mediante las técnicas basadas en la detección de interferón- γ

RESUMEN

Las técnicas de detección de la liberación de interferón- γ conocidas como IFN- γ *release assays* (IGRA), constituyen una alternativa fiable a la clásica prueba de la tuberculina (PT) para el inmunodiagnóstico de la infección tuberculosa. Las 2 pruebas comerciales disponibles, QuantiFERON-TB Gold In-tube (QFT-IT) y T-SPOT.TB, son más precisas que la PT para el diagnóstico de tuberculosis (TB), ya que son muy específicas y presentan una mejor correlación con la existencia de factores de riesgo para la infección tuberculosa. Según los datos disponibles, T-SPOT.TB detecta mayor número de positivos que QFT-IT, pero es menos específica. Aunque en determinadas situaciones, como en pacientes con inmunosupresión celular y en las edades extremas de la vida, estas técnicas siguen siendo superiores a la PT. Estudios longitudinales sugieren que las pruebas de liberación de IFN- γ son mejores predictores de la progresión a enfermedad tuberculosa; sin embargo, este hecho no ha sido demostrado completamente. Esta revisión trata el uso de los test de liberación de IFN- γ en diferentes grupos de riesgo de TB. Asimismo, remarca sus principales limitaciones y las áreas de desarrollo futuro.

Palabras clave: Determinación de liberación de interferón-γ QuantiFERON-TB Gold In-Tube T-SPOT TB

*Corresponding author

E-mail: msantin@bellvitgehospital.cat (M. Santín Cerezales).

0213-005X/\$ - see front matter © 2010 Elsevier España, S.L. Todos los derechos reservados.

From the tuberculin skin test to the interferon-γ-based assays

The tuberculin skin test (TST), that recalls the delayed-type hypersensitivity response to the intradermal inoculation of purified protein derivate (PPD),¹ has been used to diagnose TB infection for the last hundred years. The PPD contains a mixture of more than 200 antigens that are widely shared by mycobacteria other than *Mycobacterium tuberculosis*, including the vaccinal strain of *Mycobacterium bovis* bacilli Calmette-Guérin (BCG) and many non-tuberculous mycobacteria (NTM). As a result, individuals sensitized by previous exposure to NTM or BCG vaccine may respond immunologically to PPD. The other main limitation of the TST is its low sensitivity in certain groups of individuals, such as immunosuppressed patients and young children.²

Immunodiagnostic methods have been developed based on the in vitro quantification of the cellular immune response, by detecting interferon-gamma (IFN- γ) released by sensitized T-cells stimulated with specific M. tuberculosis antigens. The two main antigens used are the 6-kD M. tuberculosis early-secreted antigenic target protein (ESAT-6) and the 10-kD culture filtrate protein (CFP-10), encoded in the region of difference 1 (RD1), which is present in M. tuberculosis but not in BCG or in most NTM.3 This new in vitro technology has been rapidly adapted from initial in-house methods to the two commercially available techniques: QuantiFERON-TB Gold assays (QFT-G) (Cellestis Limited, Carnegie, Victoria, Australia) and T-SPOT. TB assay (Oxford Immunotec, Oxford, UK). Both tests, collectively known as IGRAs (Interferon-Gamma Release Assays), have been approved for sale in Europe and have received final approval from the U.S. Food and Drug Administration (FDA) as an aid for diagnosing *M. tuberculosis* infection. T-SPOT.TB detects the number of IFN- γ producing T-cells after stimulating a definite number of isolated peripheral blood mononuclear cells (PBMCs) with ESAT-6 and CFP-10 separately by means of enzyme-linked immunospot assay (ELISPOT). QFT-G tests are whole blood assays that use an enzyme-linked immunosorbent assay (ELISA) to detect IFN-y produced in supernatants by stimulated T-cells. The QFT-G In Tube version (QFT-IT), includes a third antigen, TB7.7. This new antigen is encoded in RD11 and is missing from the BCG strains as well as most common environmental mycobacteria.⁴ In the QFT-IT assay, the three specific M. tuberculosis antigens are already incorporated into the same tube (Fig. 1). Both in vitro tests include a positive control that detects the capacity of T cells to produce IFN- γ upon stimulation with a mitogen (phytohemagglutinin), in order to distinguish false-negatives from indeterminate results.

Interferon- γ -based assays for detecting latent infection in high-risk populations

This section will discuss the potential value of the IFN- γ -based tests in diagnosing latent TB infection (LTBI) in people at high risk of progression to active disease.

Contact tracing study

Between 5% and 10% of recently infected contacts will develop active TB within 2-5 years after exposure. The identification and treatment of these individuals constitutes an essential component of the TB control strategy in low-prevalence countries. Numerous studies have explored the utility of the IFN- γ -based tests in contact investigations.⁵⁻⁹ In the absence of a gold standard test for the diagnosis of LTBI, the best approach to compare IFN- γ -based tests and TST consists of correlating their results with the degree of exposure to an infectious case. Positive results of IFN- γ -based tests were found to be more strongly associated with greater recent exposure than TST;^{5,8} however, this association could not be demonstrated by others.⁹ Besides, IFN- γ -based tests offer the advantage of high specificity, since they are not affected by prior BCG vaccination or by infection with most NTM.

Health care workers

Due to the risk of infection with M. tuberculosis through occupational exposure, periodical testing is recommended for all health care workers (HCWs). Serial TST testing may induce a boosting phenomenon, compromising its interpretation.¹⁰ In a study performed in Barcelona (Spain),¹¹ prevalence of LTBI in HCWs without a previous positive TST was higher according to T-SPOT.TB (23.1%), and QFT-IT (17.3%) than according to TST (15.4%). Positive IFN- γ tests were associated with age and degree of occupational exposure, but not with BCG vaccination, a finding consistent with previous studies with QFT-G tests.^{12,13} Although IFN- γ -based tests have became a good alternative to TST for serial testing of HCWs, factors such as reversions and conversions should be taken into account. Choi et al14 described conversions of QFT-IT in HCWs 2-4 weeks after performing a TST test among TST reactors, but not among non-reactive individuals. Similarly, van Zyl-Smit et al¹⁵ reported conversions a week after TST administration. However, when using a two-step screening strategy, IFN-y test results were not influenced if TST was performed within three days.

Immunocompromised patients

The performance of the IFN- γ tests in immunocompromised patients and the effect of immunosuppression on these tests remains unclear.¹⁶ Previous studies including different groups of immunocompromised patients found impaired performance of IFN- γ -based tests related to malfunction of cellular immune system, but they performed better than TST nonetheless.^{17,18} In a prospective study including 369 immunosuppressed participants, Richeldi et al¹⁹ found that IFN- γ tests detected more patients as being infected with *M. tuberculosis* than did TST.

HIV infected patients

Patients co-infected with HIV and *M. tuberculosis* are particularly prone to a reactivation of LTBI and development of disseminated disease. In studies evaluating T-SPOT.TB and its ELISPOT precommercial version^{20,21} or the QFT-G tests,^{22,23} *in vitro* tests obtained higher rates of positive results than TST in diagnosing LTBI,^{23,24} and a better association between positive results and presence of risk factors for LTBI.^{22,23} In recent years, some studies in HIV-infected populations reported similar sensitivities for both IFN- γ tests.²¹⁻²⁵ As regards indeterminate results, a correlation between low CD4⁺ cell counts and a low control positive response was found with QFT-IT,²² while T-SPOT.TB and *in-house* ELISPOT appeared to be relatively unimpaired by low CD4⁺ cell counts.^{24,26,27} However, higher rates of indeterminate results with T-SPOT.TB and *in-house* ELISPOT have also been reported.^{24,28}

Chronic immune-mediated inflammatory disease

Tumor necrosis factor (TNF)- α antagonists provide reliable treatment in patients with immune-mediated inflammatory diseases (IMID).²⁹ TNF- α is one of the key molecules involved in granuloma formation and containment of TB infection. Due to the increased risk of TB in patients receiving anti-TNF- α agents,³⁰ exclusion of active TB and screening for latent infection is mandatory before starting anti-TNF- α therapy.³¹ However, cellular-mediated response to PPD is compromised by the corticosteroids and/or immunosuppressive drugs that most patients with IMID are already taking.^{29,31} Experience with the IFN- γ -based tests in this population, although promising, is still limited.^{32,33} Overall, agreement with the TST seems to be poor.³⁴⁻³⁶ The discordant positive TST and negative IFN- γ -based test results have been attributed to false-positive TST results,^{37,38} whereas the

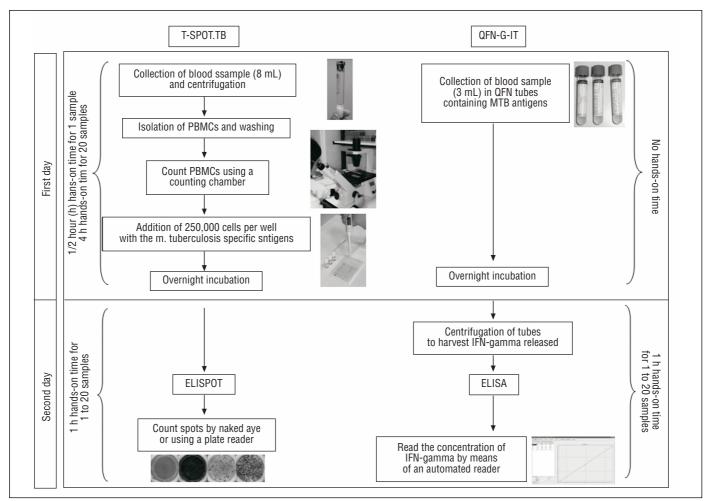


Figure 1. Comparison of T-SPOT.TB and QuantiFERON-TB Gold In Tube (QFT-IT) methodology. ELISA: enzyme-linked immunosorbent assay; ELISPOT: enzyme-linked immunospot assay; IFN: interferon; PBMCs: peripheral blood mononuclear cells.

discordant negative TST and positive IFN- γ -based test results have been considered false-negative TST results due to the immunosuppressive therapy being taken by these patients.^{35,36,39} The available data show that the IFN- γ -based tests detect more cases of LTBI than TST does,^{36,40} and a closer association with the presence of risk factors for TB infection.^{34,35} The effect of DMID-associated immunosuppression on the performance of the IFN- γ -based tests has not been completely clarified. In a study involving 398 consecutive subjects, Bartalesi et al⁴¹ did not find an association between results of the TST or QFT-IT and the use of conventional disease-modifying antirheumatic drugs, but reported an association of steroids with a lower likelihood of a positive result. In view of the high risk of TB in IMID patients receiving anti-TNF- α therapy, a strategy based on a simultaneous TST and one of the IFN- γ tests might maximize diagnostic sensitivity for the detection of LTBI.

Paediatric population

Children have a high risk of progression to active TB, especially infants under the age of two.⁴² Early, specific diagnosis of LTBI is therefore crucial to prevent active disease. The sensitivity of TST in young children is unknown, but the existence of immaturity certainly induces a lower cutaneous response. In addition, BCG-vaccination, especially in TB endemic areas, reduces the test's specificity. Overall, T-SPOT.TB provides higher rates of positive results for LTBI than TST or the QFT-G tests. Recently, Davies et al⁴³ found that, in contrast to TST, ELISPOT results were not affected by young age or severe

immunosuppression. Furthermore, a high correlation with the degree of exposure to *M. tuberculosis* with both IFN- γ -based tests has been demonstrated.⁴⁴ Results of both IFN- γ -based tests are unrelated to the BCG-vaccination status, which contributes to their high specificity.4,7 Discordant results between these tests and TST are frequently found.7,45-48 In a study conducted in Barcelona (Spain),7 among BCG-unvaccinated children, 60% and 57% of those with positive TST had negative QF-IT and T-SPOT.TB respectively. Latorre et al⁴⁹ reported that 48% of children with TST positive and negative T-SPOT.TB had sensitized T cells against Mycobacterium avium sensitins. As regards the indeterminate results, a significantly lower IFN- γ release in response to mitogen (positive control) has been described in young children tested with QFT-IT, suggesting an agedependent response.^{47,50} As for T-SPOT.TB, it does not seem to be related to age,¹⁷ except in the first weeks of life.⁵¹ However, Nicol et al48 reported a decline in positive T-SPOT.TB results in children less than 1 year of age, whereas TST results were unaffected.

Interferon- γ -based assays for diagnosing active tuberculosis

Although the IFN- γ -based tests are widely used together with or in place of TST, their role in the diagnosis of active disease is still undefined. According to the results of two recent meta-analyses,^{52,53} both commercial IFN- γ -based tests, performed in blood, have better sensitivity than TST for active TB. As expected, specificity for active disease was low, ranging from 59% for T-SPOT.TB to 79% for QFT-IT and 75% for TST.⁵³

27

While a positive result of an IFN- γ -based assay does not distinguish between active and latent infection, in combination with the TST result it may help to exclude active TB.⁵⁴⁻⁵⁶ A recent multicentre study⁵⁵ showed a very low likelihood of TB with a negative result on both TST and IFN- γ -based tests. Unfortunately, only 4% of patients were immunosuppressed, which precludes its generalization to the whole risk population. The usefulness of levels of IFN- γ , measured by QFT-IT, to predict clinical outcome was evaluated in two studies.^{56,57} Although active TB was associated with higher IFN- γ levels, the benefit was presumed to be marginal in highly experienced centres.

Cellular immunosuppression and age, among other factors, may impair performance of IFN- γ -based tests.⁵⁸ In 4 studies that compared performance of QFT-IT in HIV-infected and non-infected adults with active TB, mean sensitivity was 64% in HIV-infected patients and 79% in non-HIV-infected patients.⁵⁹⁻⁶² Although the overall sensitivity of T-SPOT.TB is higher than that of QFT-IT in otherwise healthy people, data from head-to-head comparisons in HIV-infected patients are scarce.^{60,63,64} In three studies, covering a total of 39 patients with culture-confirmed TB, QFT-IT and T-SPOT.TB detected 74% and 82% of cases respectively.^{60,64} Despite impaired sensitivity, IFN- γ -based tests are clearly superior to TST for the diagnosis of active TB in these patients.^{61,65-67}

IFN- γ -based tests may be of value in diagnosing TB in childhood, due to the absence of microbiological confirmation in a high proportion of cases. In a large prospective study, the sensitivity of T-SPOT.TB was 83%, and was not affected by HIV status.²⁰ A hospitalbased study reported sensitivity rates of 100% for TST and 73% for T-SPOT.TB and QFT.IT, and specificity rates of 58%, 98% and 100% for TST, T-SPOT.TB and QFT-IT respectively.⁶⁸ A multicentre study comparing both IFN- γ -based tests with TST in 333 children aged 2 months to 16 years found that in 49 TB-confirmed cases, sensitivity was 82% for TST, 78% for QFT-IT and 66% for T-SPOT.TB, increasing to 96% and 91% when TST was combined with T-SPOT and QFT-IT respectively.⁶⁹

As regards aged patients, the data available show that QFT-G is more sensitive than TST in patients older than 80 years with active TB.^{70,71} Although the sensitivity of QFT-G decreased with age, it remained better than that of TST.^{71,72}

Interferon- γ -based assays in fluids other than blood

IFN-gamma is predominantly produced by effector T-cells. The recruitment of specific T cells during active TB and the process via which antigen-specific cells clonally expand and migrate to the site of infection have been described.⁷³ Therefore, during active TB, it makes sense to apply IFN-based assays in samples collected directly from the site of infection. Data from a recent metanalysis⁵³ indicate that the T-SPOT.TB assay in extrasanguineous fluids is a promising tool for the diagnosis of active TB.

Pulmonary tuberculosis

Rapid diagnosis of pulmonary TB relies on the detection of acidfast bacilli (AFB). However, this can be difficult due to the low sensitivity of the sputum smear. In addition, a significant proportion of cases cannot be confirmed by culture. In a prospective study by Jafari et al,⁷⁴ all 12 patients with smear-negative pulmonary TB, but none of the 25 controls, had positive T-SPOT.TB test from the bronchoalveolar lavage (BAL). TB-specific T cells were more concentrated in BAL than in peripheral blood, indicating a highly selective compartmentalization at the site of infection.⁷⁵ A recent large study carried out by the TBNET confirmed the high sensitivity, specificity and predictive values of T-SPOT.TB from the BAL.⁷⁶

Pleural tuberculosis

The diagnosis of pleural TB is often difficult due to the limitations of conventional tests.⁷⁷ Wilkinson et al⁷³ found a 15-fold greater

concentration of ESAT-6-specific spot-forming T cells in pleural fluid than of PBMCs in 10 patients with pleural TB. These cells were not found in the pleural fluid of 8 patients with nontuberculous pleuritis.73 In a TBNET study,78 T-SPOT.TB was performed on mononuclear cells from blood and pleural fluid in 20 patients with pleural TB and in 21 with pleural effusion of other causes. T-SPOT.TB was positive in 90% of cases on blood samples and in 95% of cases pleural fluid. Specificity was 67% for blood and 76% for pleural fluid. In another study of 28 patients with pleural TB, results in pleural fluid were inconclusive in 52% of cases, due to high background IFN-γ production.⁷⁹ Commercial IFN- γ tests, T-SPOT.TB and QFT-IT in pleural fluid were compared to unstimulated IFN-γ for the diagnosis of pleural TB in 74 patients.⁸⁰ In 11 (15%) cases, the cell counts were not large enough to perform the tests. In the 63 remaining patients, sensitivity, specificity, positive predictive value and negative predictive value were: for T-SPOT.TB, 86, 60, 84 and 64% respectively; for QFT-IT, 57, 80, 87 and 44% respectively, and for unstimulated IFN- γ , 97, 100, 100 and 94% respectively. The authors concluded that the IFN- γ -based assays had suboptimal accuracy for the diagnosis of pleural TB.

Tuberculous meningitis

Tuberculous meningitis (TBM) is a challenge for clinicians because of the frequent absence of microbiological confirmation and high mortality if not promptly treated. In one study including 10 patients with a diagnosis of TBM, T-SPOT.TB detected M. tuberculosis antigenspecific IFN- γ in CSF from nine patients (90%), but in none of the seven controls (specificity 100%).⁸¹ In a study with 12 patients with TB of the central nervous system and 25 without TB, T-SPOT's sensitivity and specificity in CSF were 75%.82 Recently, in a prospective observational study of 31 patients with confirmed or probable TBM, the same group of investigators⁸³ reported a sensitivity of 59% and a specificity of 89% for T-SPOT.TB in CSF mononuclear cells. However, since the diagnosis was not confirmed microbiologically in 21 of these patients, the sensitivity may have been underestimated. Similarly, in a study of 140 patients with meningitis (81% HIV-infected), using ≥46 spot-forming cells as cut-off point and after excluding bacterial and cryptococcal meningitis, the positive and negative predictive values of T-SPOT.TB in CSF were 100% and 68% respectively.84

Interferon- γ -based assays for predicting subsequent active tuberculosis

The ability to predict subsequent active TB among latently infected people is essential in order to select those who would benefit from chemoprophylaxis and to avoid unnecessary treatment for low-risk persons. Doherty et al⁸⁵ demonstrated a strong association of reactivity to ESAT-6 and progression to active TB in twenty-four household contacts of smear-positive TB patients. In a study involving 601 close contacts of sputum smear-positive TB, Diel et al⁸⁶ found that while 14.6% of contacts with positive QFT-IT who declined treatment developed TB within the 2-year follow-up, only 2.3% of those with positive TST did. This difference between TST and QFT-IT disappeared when only unvaccinated contacts were considered.⁸⁷ More recently, the same group of investigators⁸⁸ extended the original study and reported progression to active TB for up to four years for a cohort of 954 close contacts of smear-positive index cases. Of 147 untreated contacts with a positive QFT-IT test 19 (12.9%) developed active TB, whereas only 17 of 155 (3.1%) with TST >5 mm did. The progression rate was higher among children (28.6%). In addition, none of 824 untreated contacts with negative QFT-IT developed active TB, confirming the high negative predictive value of the test.⁸⁸ In the study by Kik et al⁸⁹ of 339 close contacts of sputum smear-positive TB, TST, QFT-IT and T-SPOT.TB were comparable in predicting development of TB during a two-year period. Positive predictive values were 3.1% for TST ≥10 mm, 3.8% for TST ≥15 mm, 2.8% for positive QFT-IT and 3.3% for T-SPOT.TB. In a cohort of 308

Table 1

Study (reference)	Country	Study population	Period of follow-up (years)	Test	Positive test n/N (%)	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)
Doherty et al ⁸⁵	Ethiopia	Household contact adults	2	ELISA (PPD)	21/24 (88)	100	18	33	100
				ELISA (ESAT-6)	9/24 (38)	86	83	67	93
Leung et al ⁸⁷	Hong Kong	Silicosis patients	2.5ª	TST (10 mm)	203/308 (66)	77	35	6.4	96
				T-SPOT.TB	204/308 (66)	88	36	7.4	98
Diel et al ⁸⁸	Germany	Close contact adults	4	TST	555/903 (60)	89	61	3.1	99
				QFT-IT	147/903 (16)	100	86	12.9	100
Kik et al ⁸⁹	Netherlands	Contact immigrant adults	2	TST	339 ^b	100	16	3.1	100
				QFT-IT	178/324 (55) ^b	63	45	2.8	98
				T-SPOT.TB	181/299 (61) ^b	75	40	3.3	98
Hill et al ⁹⁰	Gambia	Household contact childre & adults	2	TST	843/2230 (38)	54	62	1.7	99
		(HIV +ve and -ve)		T-SPOT.TB	649/1736 (37)	42	63	1.7	99
Aichelburg et al ⁹¹	Austria	HIV (+) adults	1.6	QFT-IT	44/783 (5.6)	100	95	8.1	100
Santin et al ⁹²	Spain	HIV (+) adults	1.6ª	TST	7/120 (6)	-	94 ^c	-	100 ^c
				QFT-IT	13/120(11)	-	89°	-	100 ^c
Bakir et al93	Turkey	Contact children	1.3	TST	550/908 (61)	80	40	2	99
				T-SPOT.TB	381/908 (42)	73	59	3	99
Higuchi et al ⁹⁴	Japan	Contact adolescents	3.5	TST	95	-	73	-	100
				QFT-G	4/88 ^d	-	96 ^e	-	100 ^e

NPV: negative predictive value; PPV: positive predictive value.

^aMean follow-up.

^bOnly subjects with positive TST were tested with QFT-IT and T-SPOT.TB.

Calculated with 120 non-treated patients (those with negative TST and negative/indeterminate QFT-IT).

^dOnly subjects with positive TST were tested with QFT-G.

^eCalculated with 84 non-treated subjects (those with negative QFT-G).

silicosis patients, Leung et al⁸⁷ found that a positive T-SPOT.TB significantly predicted development of active TB during a follow-up of more than 2 years (RR 7.80; 95%CI 1.02-59.6). Unexpectedly, TST was not predictive of TB, regardless of the cut-off point used. In a study with 2348 household contacts in Gambia, Hill et al⁹⁰ found that neither the TST nor the T-SPOT.TB predicted development of TB. The lack of predictive value was attributed to the high-burden of TB and recent transmission.

Two studies assessed QFT-IT and progression to active disease in HIV-seropositive individuals.^{91,92} In a large study in a low-prevalence country, 8.1% of HIV-seropositive patients with a positive QFT-IT result at baseline, and left untreated, developed TB during a median follow-up of 19 months. None of the 738 patients with negative results had TB.⁹¹ In a study in Spain of 135 HIV-infected individuals without active disease, none of the 103 patients who had a negative or indeterminate QFT-IT result at baseline had TB after a median follow-up of 20 months.⁹²

Development of TB was also assessed in child and adolescent contacts. Bakir et al⁹³ studied 908 children with recent household exposure to TB, most of whom received preventive therapy. During a follow-up of 1.3 years, children with positive T-SPOT.TB had a 3- to 4-fold higher risk of developing active TB than those with negative T-SPOT.TB. However, rates of progression were similar in children with positive T-SPOT.TB and TST reactors. Since a high proportion of children were treated, the true incidence rates may have been underestimated. In the study by Higuchi et al,⁹⁴ 349 students underwent QFT-G and TST simultaneously, but only those with

positive QFT-G were given chemoprophylaxis. Follow-up of the 91 students with positive TST but negative QFT-G showed no cases of active TB.

Although IFN- γ -based tests seem to predict subsequent active TB better than TST, the majority of high-risk people with positive tests will not develop active TB. Conversely, subsequent active TB in the next two to three years seems to be extremely low among people with a negative result.^{85-87,89-94} Table 1 summarizes the nine studies assessing development of active TB with IFN- γ -based tests.

Final remarks and areas of future development

IFN- γ -based assays have become a reliable alternative to the old TST for the diagnosis of TB infection. Both commercial tests, QFT-IT and T-SPOT.TB, have a higher specificity than TST, and a better correlation with risk factors for TB and the degree of contact with an infectious case. Although their sensitivity may be affected to some extent by immunosuppression and extreme ages of life, they perform better than TST in these situations. Besides, IFN- γ -based tests do not induce boosting, and no additional visits are required for reading.

A great deal is now known about IFN- γ -based assays, and their use has expanded considerably. However, the prognostic value of a positive/negative result for the development of active TB, the significance of discordant results, the cut-off points to use in immunosuppressed people, the conversion/reversion phenomenon, and their role in the diagnosis of paucibacillary forms of TB, are some of the important questions that remain unresolved.

The actual prognostic value of a positive IFN-γ result needs to be clarified. Although the available data suggest that IFN- γ tests predict progression to active disease better than TST, most people with a positive result will not develop TB. Large prospective studies are urgently needed. Furthermore, the question of whether quantification of IFN- γ release may be of help in this situation, as has been previously suggested,^{85,86} should also be addressed. Because of the discordance between IFN- γ tests and TST results, practitioners are reluctant to use them in everyday clinical practice. Trials focusing specifically on understanding the discordant results between IFN- γ tests and the TST, and between IFN- γ tests themselves, are required. This issue is especially relevant in childhood, where the effect of NTM infection may play an important role.49 Since in vitro assays rely on the secretion of IFN- γ , which is largely produced by CD4⁺ T cells, determining the CD4 threshold at which the performance of these assays declines is of particular importance. In addition, studies exploring the effect of the different immunosuppressor drugs on the response, as well as the accuracy of new cut-offs for diagnosing LTBI in immunosuppressed patients, are needed. Detection of *M. tuberculosis* specific T cells in samples other than blood with ELISPOT is a promising tool for the diagnosis of smear-negative pulmonary TB⁷⁶ and other paucibacillary forms of TB.78,81 The methodological procedures and appropriate cut-offs should be established.

While awaiting answers to these questions, the use of the IFN- γ -based tests in clinical practice should be guided by clinical judgement and evidence-based guidelines for different groups of patients must be developed.

Finally, technical modifications of IFN- γ -based tests are being explored.⁹⁵ The attempts to improve IFN- γ -based tests include the study of alternative readouts to measure IFN- γ release.^{95,96} the use of alternative *M. tuberculosis* specific antigens,^{4,97} and the simultaneous measurement of chemokines⁹⁸ and interleukins.⁹⁹ The next generation of IFN- γ -based tests will significantly enhance diagnostic sensitivity without diminishing specificity, and will also reduce the rate of indeterminate results, especially in immunosuppressed patients and children.

Conflict of interest

The authors have no relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript. This includes employment, consultancies, honoraria, stock ownership or options, expert testimony, grants or patents received or pending, or royalties. No writing assistance was utilized in the production of this manuscript.

M. Santín received a fee from Inverness Medical Ibérica, S.A.U. (Distributor of QFT-IT in Spain) for giving lectures on IGRA. J. Domínguez is a researcher funded from the *Miguel Servet* programme of the *Instituto de Salud Carlos III* (Spain).

References

- Jasmer RM, Nahid P, Hopewell PC. Clinical practice. Latent tuberculosis infection. N Engl J Med. 2002;347:1860-6.
- Horsburgh CR Jr. Priorities for the treatment of latent tuberculosis infection in the United States. N Engl J Med. 2004;350:2060-7.
- Andersen P, Munk ME, Pollock JM, Doherty TM. Specific immune-based diagnosis of tuberculosis. Lancet. 2000;356:1099-104.
- Brock I, Weldingh K, Leyten EM, Arend SM, Ravn P, Andersen P. Specific T-cell epitopes for immunoassay-based diagnosis of *Mycobacterium tuberculosis* infection. J Clin Microbiol. 2004;42:2379-87.
- Arend SM, Thijsen SF, Leyten EM, Bouwman JJ, Franken WP, Koster BF, et al. Comparison of two interferon-gamma assays and tuberculin skin test for tracing TB contacts. Am J Respir Crit Care Med. 2007;175:618-27.
- Diel R, Loddenkemper R, Meywald-Walter K, Gottschalk R, Nienhaus A. Comparative performance of tuberculin skin test, QuantiFERON-TB-Gold In Tube assay, and T-Spot.TB test in contact investigations for tuberculosis. Chest. 2009;135:1010-8.

- Domínguez J, Ruiz-Manzano J, De Souza-Galvao M, Latorre I, Milà C, Blanco S, et al. Comparison of two commercially available gamma interferon blood tests for immunodiagnosis of tuberculosis. Clin Vaccine Immunol. 2008;15:168-71.
- Lalvani A, Pathan AA, McShane H, Wilkinson RJ, Latif M, Conlon CP, et al. Rapid detection of *Mycobacterium tuberculosis* infection by enumeration of antigenspecific T cells. Am J Respir Crit Care Med. 2001;163:824-8.
 Janssens JP, Roux-Lombard P, Perneger T, Metzger M, Vivien R, Rochat T.
- Janssens JP, Roux-Lombard P, Perneger T, Metzger M, Vivien R, Rochat T. Contribution of a IFN-gamma assay in contact tracing for tuberculosis in a lowincidence, high immigration area. Swiss Med Wkly. 2008;138:585-93.
- Menzies R, Vissandjee B, Rocher I, St Germain Y. The booster effect in two-step tuberculin testing among young adults in Montreal. Ann Intern Med. 1994;120:190-8.
- Casas I, Latorre I, Esteve M, Ruiz-Manzano J, Rodríguez D, Prat C, et al. Evaluation of interferon-gamma release assays in the diagnosis of recent tuberculosis infection in health care workers. PLoS One. 2009;4:e6686.
- 12. Nienhaus A, Schablon A, Bacle CL, Siano B, Diel R, et al. Evaluation of the interferongamma release assay in healthcare workers. Int Arch Occup Environ Health. 2008;81:295-300.
- Pai M, Gokhale K, Joshi R, Dogra S, Kalantri S, Mendiratta DK, et al. Mycobacterium tuberculosis infection in health care workers in rural India: comparison of a wholeblood interferon gamma assay with tuberculin skin testing. JAMA. 2005;293:2746-55.
- Choi JC, Shin JW, Kim JY, Park IW, Choi BW, Lee MK. The effect of previous tuberculin skin test on the follow-up examination on whole-blood interferon-gamma assay in the screening for latent tuberculosis infection. Chest. 2008;133:1415-20.
- Van Zyl-Smit RN, Pai M, Peprah K, Meldau R, Kieck J, Juritz J, et al. Within-subject variability and boosting of T-cell interferon-gamma responses after tuberculin skin testing. Am J Respir Crit Care Med. 2009;180:49-58.
 Menzies D, Pai M, Comstock G. Meta-analysis: new tests for the diagnosis of latent
- Menzies D, Pai M, Comstock G. Meta-analysis: new tests for the diagnosis of latent tuberculosis infection: areas of uncertainty and recommendations for research. Ann Intern Med. 2007;146:340-54.
- 17. Ferrara G, Losi M, D'Amico R, Roversi P, Piro R, Meacci M, et al. Use in routine clinical practice of two commercial blood tests for diagnosis of infection with *Mycobacterium tuberculosis*: a prospective study. Lancet. 2006;367:1328-34.
- Kobashi Y, Mouri K, Obase Y, Fukuda M, Miyashita N, Oka M. Clinical evaluation of QuantiFERON TB-2G test for immunocompromised patients. Eur Respir J. 2007;30:945-50.
- Richeldi L, Losi M, D'Amico R, Luppi M, Ferrari A, Mussini C, et al. Performance of tests for latent tuberculosis in different groups of immunocompromised patients. Chest. 2009;136:198-204.
- Liebeschuetz S, Bamber S, Ewer K, Deeks J, Pathan AA, Lalvani A. Diagnosis of tuberculosis in South African children with a T-cell-based assay: a prospective cohort study. Lancet. 2004;364:2196-203.
- Mandalakas AM, Hesseling AC, Chegou NN, Kirchner HL, Zhu X, Marais BJ, et al. High level of discordant IGRA results in HIV-infected adults and children. Int J Tuberc Lung Dis. 2008;12:417-23.
- Brock I, Ruhwald M, Lundgren B, Westh H, Mathiesen LR, Ravn P. Latent tuberculosis in HIV positive, diagnosed by the *M. tuberculosis* specific interferon-gamma test. Respir Res. 2006;7:56.
- Jones S, De Gijsel D, Wallach FR, Gurtman AC, Shi Q, Sacks H. Utility of QuantiFERON-TB Gold in-tube testing for latent TB infection in HIV-infected individuals. Int J Tuberc Lung Dis. 2007;11:1190-5.
- Stephan C, Wolf T, Goetsch U, Bellinger O, Nisius G, Oremek G, et al. Comparing QuantiFERON-tuberculosis gold, T-SPOT tuberculosis and tuberculin skin test in HIV-infected individuals from a low prevalence tuberculosis country. AIDS. 2008;22:2471-9.
- Rangaka MX, Wilkinson KA, Seldon R, Van Cutsem G, Meintjes GA, Morroni C, et al. Effect of HIV-1 infection on T-Cell-based and skin test detection of tuberculosis infection. Am J Respir Crit Care Med. 2007;175:514-20.
- 26. Clark SA, Martin SL, Pozniak A, Steel A, Ward B, Dunning J, et al. Tuberculosis antigen-specific immune responses can be detected using enzyme-linked immunospot technology in human immunodeficiency virus (HIV)-1 patients with advanced disease. Clin Exp Immunol. 2007;150:238-44.
- Rivas I, Latorre I, Sanvisens A, Domínguez J, Tor J, Prat C, et al. Prospective evaluation of latent tuberculosis with interferon-gamma release assays in drug and alcohol abusers. Epidemiol Infect. 2009;137:1342-7.
- Karam F, Mbow F, Fletcher H, Senghor CS, Coulibaly KD, LeFevre AM, et al. Sensitivity of IFN-gamma release assay to detect latent tuberculosis infection is retained in HIV-infected patients but dependent on HIV/AIDS progression. PLoS ONE. 2008;3:e1441.
- Panes J, Gomollón F, Taxonera C, Hinojosa J, Clofent J, Nos P. Crohn's disease: a review of current treatment with a focus on biologics. Drugs. 2007;67:2511-37.
- Gardam MA, Keystone EC, Menzies R, Manners S, Skamene E, Long R, et al. Antitumour necrosis factor agents and tuberculosis risk: mechanisms of action and clinical management. Lancet Infect Dis. 2003;3:148-55.
- Carmona L, Gomez-Reino JJ, Rodríguez-Valverde V, Montero D, Pascual-Gómez E, Mola EM, et al. Effectiveness of recommendations to prevent reactivation of latent tuberculosis infection in patients treated with tumor necrosis factor antagonists. Arthritis Rheum. 2005;52:1766-72.
- 32. Domínguez J, Latorre I. Role of the T-cell interferon-gamma release assays in preventing reactivation of latent tuberculosis infection in immunosuppressed patients in treatment with anti-TNF agents. J Crohn's & Colitis. 2008;2:250-4.
- Lalvani A, Millington KA. Screening for tuberculosis infection prior to initiation of anti-TNF therapy. Autoimmun Rev. 2008;8:147-52.
- 34. Bocchino M, Matarese A, Bellofiore B, Giacomelli P, Santoro G, Balato N, et al. Performance of two commercial blood IFN-gamma release assays for the detection

of *Mycobacterium tuberculosis* infection in patient candidates for anti-TNF-alpha treatment. Eur J Clin Microbiol Infect Dis. 2008;27:907-13.

- Matulis G, Juni P, Villiger PM, Gadola SD. Detection of latent tuberculosis in immunosuppressed patients with autoimmune diseases: performance of a *Mycobacterium tuberculosis* antigen-specific interferon gamma assay. Ann Rheum Dis. 2008;67:84-90.
- 36. Ponce de León D, Acevedo-Vásquez E, Alvizuri S, Gutiérrez C, Cucho M, Alfaro J, et al. Comparison of an interferon-gamma assay with tuberculin skin testing for detection of tuberculosis (TB) infection in patients with rheumatoid arthritis in a TB-endemic population. J Rheumatol. 2008;35:776-81.
- Cobanoglu N, Ozcelik U, Kalyoncu U, Ozen S, Kiraz S, Gurcan N, et al. Interferongamma assays for the diagnosis of tuberculosis infection before using tumour necrosis factor-alpha blockers. Int J Tuberc Lung Dis. 2007;11:1177-82.
- Martin J, Walsh C, Gibbs A, McDonnell T, Fearon U, Keane J, et al. Comparison of interferon-{gamma}-release assays and conventional screening tests before tumour necrosis factor-{alpha} blockade in patients with inflammatory arthritis. Ann Rheum Dis. 2010;69:181-5.
- 39. Vassilopoulos D, Stamoulis N, Hadziyannis E, Archimandritis AJ. Usefulness of enzyme-linked immunosorbent assay (Elispot) compared to tuberculin skin testing for latent tuberculosis screening in rheumatic patients scheduled for antitumor necrosis factor treatment. Addendum. J Rheumatol. 2008;35:1464.
- 40. Takahashi H, Shigehara K, Yamamoto M, Suzuki C, Naishiro Y, Tamura Y, et al. Interferon gamma assay for detecting latent tuberculosis infection in rheumatoid arthritis patients during infliximab administration. Rheumatol Int. 2007;27:1143-8.
- arthritis patients during infliximab administration. Rheumatol Int. 2007;27:1143-8.
 41. Bartalesi F, Vicidomini S, Goletti D, Fiorelli C, Fiori G, Melchiorri D, et al. QuantiFERON-TB Gold and the TST are both useful for latent tuberculosis infection screening in autoimmune diseases. Eur Respir J. 2009;33:586-93.
- Lalvani A, Millington KA. T cell-based diagnosis of childhood tuberculosis infection. Curr Opin Infect Dis. 2007;20:264-71.
- Davies MA, Connell T, Johannisen C, Wood K, Pienaar S, Wilkinson KA, et al. Detection of tuberculosis in HIV-infected children using an enzyme-linked immunospot assay. AIDS. 2009;23:961-9.
- 44. Altet N, De Souza-Galvao M, Latorre I, et al. Diagnosing TB infection in children: analysis of discordances using in vitro tests and tuberculin skin test. Eur Respir J. 2010;doi:10.1183/09031936.00022710.
- Connell TG, Ritz N, Paxton GA, Buttery JP, Curtis N, Ranganathan SC. A three-way comparison of tuberculin skin testing, QuantiFERON-TB gold and T-SPOT.TB in children. PLoS ONE. 2008;3:e2624.
- 46. Kampmann B, Whittaker E, Williams A, Walters S, Gordon A, Martínez-Alier N, et al. Interferon-gamma release assays do not identify more children with active TB than TST. Eur Respir J. 2009;33:1374-82.
- Lighter J, Rigaud M, Eduardo R, Peng CH, Pollack H. Latent tuberculosis diagnosis in children by using the QuantiFERON-TB Gold In-Tube test. Pediatrics. 2009;123:30-7.
- 48. Nicol MP, Davies MA, Wood K, Hatherill M, Workman L, Hawkridge A, et al. Comparison of T-SPOT.TB assay and tuberculin skin test for the evaluation of young children at high risk for tuberculosis in a community setting. Pediatrics. 2009;123:38-43.
- Latorre I, De Souza-Galvao M, Ruiz-Manzano J, Lacoma A, Prat C, Altet N, et al. Evaluating the non-tuberculous mycobacteria effect in the tuberculosis infection diagnosis. Eur Respir J. 2010;35:338-42.
- Connell TG, Curtis N, Ranganathan SC, Buttery JP. Performance of a whole blood interferon gamma assay for detecting latent infection with Mycobacterium tuberculosis in children. Thorax. 2006;61:616-20.
- Richeldi L, Ewer K, Losi M, Bergamini BM, Roversi P, Deeks J, et al. T cell-based tracking of multidrug resistant tuberculosis infection after brief exposure. Am J Respir Crit Care Med. 2004;170:288-95.
- Diel R, Loddenkemper R, Nienhaus A. Evidence-based comparison of commercial interferon-gamma release assays for detecting active TB: a metaanalysis. Chest. 2010;137:952-68.
- Sester M, Sotgiu G, Lange C, Giehl C, Girardi E, Migliori GB, et al. Interferon-(gamma) release assays for the diagnosis of active tuberculosis: a systematic review and meta-analysis. Eur Respir J. 2011;37:100-11.
- 54. Domínguez J, De Souza-Galvao M, Ruiz-Manzano J, Latorre I, Prat C, Lacoma A, et al. T-cell responses to the *Mycobacterium tuberculosis*-specific antigens in active tuberculosis patients at the beginning, during, and after antituberculosis treatment. Diagn Microbiol Infect Dis. 2009;63:43-51.
- Goletti D, Stefania C, Butera O, Amicosante M, Ernst M, Sauzullo I, et al. Accuracy of immunodiagnostic tests for active tuberculosis using single and combined results: a multicenter TBNET-Study. PLoS ONE. 2008;3:e3417.
- 56. Metcalfe JZ, Cattamanchi A, Vittinghoff E, Ho C, Grinsdale J, Hopewell C, et al. Evaluation of quantitative IFN-gamma response for risk stratification of active tuberculosis suspects. Am J Respir Crit Care Med. 2010;181:87-93.
- Latorre I, De Souza-Galvao M, Ruiz-Manzano J, Lacoma A, Prat C, Fuenzalida L, et al. Quantitative evaluation of T-cell response after specific antigen stimulation in active and latent tuberculosis infection in adults and children. Diagn Microbiol Infect Dis. 2009;65:236-46.
- 58. Pai M, Menzies D. Interferon-gamma release assays: what is their role in the diagnosis of active tuberculosis? Clin Infect Dis. 2007;44:74-7.
- 59. Aabye MG, Ravn P, PrayGod G, Jeremiah K, Mugomela A, Jepsen M, et al. The impact of HIV infection and CD4 cell count on the performance of an interferon gamma release assay in patients with pulmonary tuberculosis. PLoS ONE. 2009;4:e4220.
- Chee CB, Gan SH, Khinmar KW, Barkham TM, Koh CK, Liang S, et al. Comparison of sensitivities of two commercial gamma interferon release assays for pulmonary tuberculosis. J Clin Microbiol. 2008;46:1935-40.

- 61. Raby E, Moyo M, Devendra A, Banda J, De Haas P, Ayles H, et al. The effects of HIV on the sensitivity of a whole blood IFN-gamma release assay in Zambian adults with active tuberculosis. PLoS ONE. 2008;3:e2489.
- 62. Tsiouris SJ, Coetzee D, Toro PL, Austin J, Stein Z, El-Sadr W. Sensitivity analysis and potential uses of a novel gamma interferon release assay for diagnosis of tuberculosis. J Clin Microbiol. 2006;44:2844-50.
- Markova R, Todorova Y, Drenska R, et al. Usefulness of interferon-gamma release assays in the diagnosis of tuberculosis infection in HIV-infected patients in Bulgaria. Biotechnol & Biotechnol Eq. 2009;23:1103-8.
 Leidl L, Mayanja-Kizza H, Sotgiu G, Baseke J, Ernst M, Hirsch C, et al. Relationship
- 64. Leidl L, Mayanja-Kizza H, Sotgiu G, Baseke J, Ernst M, Hirsch C, et al. Relationship of immunodiagnostic assays for tuberculosis and numbers of circulating CD4+ T-cells in HIV infection. Eur Respir J. 2010;35:619-26.
- 65. Vincenti D, Carrara S, Butera O, Bizzoni F, Casetti R, Girardi E, et al. Response to region of difference 1 (RD1) epitopes in human immunodeficiency virus (HIV)infected individuals enrolled with suspected active tuberculosis: a pilot study. Clin Exp Immunol. 2007;150:91-8.
- 66. García-Gasalla M, Fernández-Baca V, Mir-Viladrich I, Cifuentes-Luna C, Campins-Rosselló A, Payeras-Cifre A, et al. Quantiferon-TB-Gold In Tube test in the diagnosis of pulmonary and extra-pulmonary tuberculosis. Enferm Infecc Microbiol Clin. 2010;28:685-9.
- Syed Ahamed Kabeer B, Sikhamani R, Swaminathan S, et al. Role of interferon gamma release assay in active TB diagnosis among HIV infected individuals. PLoS One. 2009;4:e5718.
- 68. Detjen AK, Keil T, Roll S, Hauer B, Mauch H, Wahn U, et al. Interferon-gamma release assays improve the diagnosis of tuberculosis and nontuberculous mycobacterial disease in children in a country with a low incidence of tuberculosis. Clin Infect Dis. 2007;45:322-8.
- 69. Bamford AR, Crook AM, Clark JE, Nademi Z, Dixon G, Paton JY, et al. Comparison of interferon-{gamma} release assays and tuberculin skin test in predicting active tuberculosis (TB) in children in the UK: a paediatric TB network study. Arch Dis Child. 2010;95:180-6.
- Kobashi Y, Mouri K, Yagi S, Obase Y, Miyashita N, Okimoto N, et al. Clinical utility of the QuantiFERON TB-2G test for elderly patients with active tuberculosis. Chest. 2008;133:1196-202.
- Mori T, Sakatani M, Yamagishi F, Takashima T, Kawabe Y, Nagao K, et al. Specific detection of tuberculosis infection: an interferon-gamma-based assay using new antigens. Am J Respir Crit Care Med. 2004;170:59-64.
- Kobashi Y, Mouri K, Miyashita N, Okimoto N, Matsushima T, Kageoka T, et al. QuantiFERON TB-2G test for patients with active tuberculosis stratified by age groups. Scand J Infect Dis. 2009;41:841-6.
- 73. Wilkinson KA, Wilkinson RJ, Pathan A, Ewer K, Prakash M, Klenerman P, et al. Ex vivo characterization of early secretory antigenic target 6-specific T cells at sites of active disease in pleural tuberculosis. Clin Infect Dis. 2005;40:184-7.
- 74. Jafari C, Ernst M, Kalsdorf B, Greinert U, Diel R, Kirsten D, et al. Rapid diagnosis of smear-negative tuberculosis by bronchoalveolar lavage enzyme-linked immunospot. Am J Respir Crit Care Med. 2006;174:1048-54.
- Jafari C, Ernst M, Strassburg A, Greinert U, Kalsdorf B, Kirsten D, et al. Local immunodiagnosis of pulmonary tuberculosis by enzyme-linked immunospot. Eur Respir J. 2008;31:261-5.
- 76. Jafari C, Thijsen S, Sotgiu G, Goletti D, Domínguez-Benítez JA, Losi M, et al. Bronchoalveolar lavage enzyme-linked immunospot for a rapid diagnosis of tuberculosis: a Tuberculosis Network European Trialsgroup study. Am J Respir Crit Care Med. 2009;180:666-73.
- Trajman A, Pai M, Dheda K, Van Zyl Smit R, Zwerling AA, Joshi R, et al. Novel tests for diagnosing tuberculous pleural effusion: what works and what does not? Eur Respir J. 2008;31:1098-106.
- Losi M, Bossink A, Codecasa L, Jafari C, Ernst M, Thijsen S, et al. Use of a T-cell interferon-{gamma} release assay for the diagnosis of tuberculous pleurisy. Eur Respir J. 2007;30:1173-9.
- Baba K, Sornes S, Hoosen AA, Lekabe JM, Mpe MJ, Langeland M, et al. Evaluation of immune responses in HIV infected patients with pleural tuberculosis by the QuantiFERON TB-Gold interferon-gamma assay. BMC Infect Dis. 2008;8:35.
- Dheda K, Van Zyl-Smit RN, Sechi LA, Badri M, Meldau R, Meldau S, et al. Utility of quantitative T-cell responses versus unstimulated interferon-{gamma} for the diagnosis of pleural tuberculosis. Eur Respir J. 2009;34:1118-26.
- Thomas MM, Hinks TS, Raghuraman S, Ramalingam N, Ernst M, Nau R, et al. Rapid diagnosis of *Mycobacterium tuberculosis* meningitis by enumeration of cerebrospinal fluid antigen-specific T-cells. Int J Tuberc Lung Dis. 2008;12:651-7.
- Kim SH, Chu K, Choi SJ, Song KH, Kim HB, Kim NJ, et al. Diagnosis of Central Nervous System Tuberculosis by T-cell-based Assays on Peripheral Blood and Cerebrospinal Fluid Mononuclear cells. Clin Vaccine Immunol. 2008;15:1356-62.
- 83. Kim SH, Cho OH, Park SJ, Lee EM, Kim MN, Lee SO, et al. Rapid diagnosis of tuberculous meningitis by T cell-based assays on peripheral blood and cerebrospinal fluid mononuclear cells. Clin Infect Dis. 2010;50:1349-58.
- 84. Patel VB, Singh R, Connolly C, Coovadia Y, Peer AK, Parag P, et al. Cerebrospinal T-cell responses aid in the diagnosis of tuberculous meningitis in a human immunodeficiency virus- and tuberculosis-endemic population. Am J Respir Crit Care Med. 2010;182:569-77.
- Doherty TM, Demissie A, Olobo J, Wolday D, Britton S, Eguale T, et al. Immune responses to the *Mycobacterium tuberculosis*-specific antigen ESAT-6 signal subclinical infection among contacts of tuberculosis patients. J Clin Microbiol. 2002;40:704-6.
- Diel R, Loddenkemper R, Meywald-Walter K, et al. Predictive value of a wholeblood IFN-gamma assay for the development of active TB disease. Am J Respir Crit Care Med. 2008;177:1164-70.

- 87. Leung CC, Chang KC, Chau CH. Is the whole-blood gamma interferon assay better than the tuberculin skin test in predicting active tuberculosis? Am J Respir Crit Care Med. 2008;178:210-11; author reply 211.
- Diel R, Loddenkemper R, Niemann S, Meywald-Walter K, Nienhaus A. Negative and Positive Predictive Value of a Whole-Blood IGRA for Developing Active TB - An Update. Am J Respir Crit Care Med. 2011;183:88-95.
 Kik SV, Franken WP, Mensen M, Cobelens FG, Kamphorst M, Arend SM, et al.
- Kik SV, Franken WP, Mensen M, Cobelens FG, Kamphorst M, Arend SM, et al. Predictive value for progression to tuberculosis by IGRA and TST in immigrant contacts. Eur Respir J. 2010;35:1346-53.
 Hill PC, Jackson-Sillah D, Fox A, Brookes RH, de Jong BC, Lugos MD, et al. Incidence
- Hill PC, Jackson-Sillan D, Fox A, Brookes RH, de Jong BC, Lugos MD, et al. Incidence of tuberculosis and the predictive value of ELISPOT and Mantoux tests in Gambian case contacts. PLoS ONE. 2008;3:e.1379.
- Aichelburg MC, Rieger A, Breitenecker F, Pfistershammer K, Tittes J, Eltz S, et al. Detection and prediction of active tuberculosis disease by a whole-blood interferongamma release assay in HIV-1-infected individuals. Clin Infect Dis. 2009;48:954-62.
- 92. Santín M, Casas S, Saumoy M, Andreu A, Moure R, Alcaide F, et al. Detection of latent tuberculosis by the tuberculin skin test and a whole-blood interferongamma release assay, and the development of active tuberculosis in HIVseropositive persons. Diagn Microbiol Infect Dis. 2011;69:59-65.

- Bakir M, Millington KA, Soysal A, Deeks JJ, Efee S, Aslan Y, et al. Prognostic value of a T-cell-based, interferon-gamma biomarker in children with tuberculosis contact. Ann Intern Med. 2008;149:777-87.
- 94. Higuchi K, Harada N, Mori T, Sekiya Y. Use of QuantiFERON-TB Gold to investigate tuberculosis contacts in a high school. Respirology. 2007;12:88-92.
- Lalvani A, Millington KA. T-cell interferon-gamma release assays: can we do better? Eur Respir J. 2008;32:1428-30.
 Lalvani A, Pareek M. Interferon gamma release assays: principles and practice.
- Enferm Infecc Microbiol Clin. 2010;28:245-2.
 Liu XQ, Dosanjh D, Varia H, Ewer K, Cockle P, Pasvol G, et al. Evaluation of T-cell
- Du XQ, Dosanji D, Vala H, Ever K, Cocke F, Pasvo G, et al. Evidation of 1-cen responses to novel RD1- and RD2-encoded Mycobacterium tuberculosis gene products for specific detection of human tuberculosis infection. Infect Immun. 2004;72:2574-81.
- Ruhwald M, Bodmer T, Maier C, Jepsen M, Haaland MB, Eugen-Olsen J, et al. Evaluating the potential of IP-10 and MCP-2 as biomarkers for the diagnosis of tuberculosis. Eur Respir J. 2008;32:1607-15.
- Millington KA, Innes JA, Hackforth S, Hinks TS, Deeks JJ, Dosanjh DP, et al. Dynamic relationship between IFN-gamma and IL-2 profile of *Mycobacterium tuberculosis*specific T cells and antigen load. J Immunol. 2007;178:5217-26.