

Serveur Académique Lausannois SERVAL serval.unil.ch

Author Manuscript

Faculty of Biology and Medicine Publication

This paper has been peer-reviewed but does not include the final publisher proof-corrections or journal pagination.

Published in final edited form as:

Title: Bacteremia caused by *Comamonas kerstersii* in a patient with diverticulosis.

Authors: Opota O, Ney B, Zanetti G, Jatton K, Greub G, Prod'homme G

Journal: Journal of clinical microbiology

Year: 2014 Mar

Volume: 52

Issue: 3

Pages: 1009-12

DOI: 10.1128/JCM.02942-13

In the absence of a copyright statement, users should assume that standard copyright protection applies, unless the article contains an explicit statement to the contrary. In case of doubt, contact the journal publisher to verify the copyright status of an article.

1 Title: *Comamonas kerstersii* bacteremia in a patient with diverticulosis

2

3 Onya Opota¹, Barbara Ney², Giorgio Zanetti^{2,3}, Katia Jaton¹, Gilbert Greub^{1,2} and Guy Prod'homme¹

4 Institute of Microbiology¹, Infectious Diseases Service², Hospital Preventive Medicine and Infectious
5 Diseases Service³, University Hospital of Lausanne, Lausanne, Switzerland

6

7 Corresponding author:

8 Gilbert Greub,

9 Institute of Microbiology,

10 University Hospital of Lausanne,

11 Bugnon 46, 1010 Lausanne,

12 Switzerland.

13 Phone +41 (0)21 314 4979

14 Fax +41 (0)21 314 4060

15 E-mail : Gilbert.Greub@chuv.ch

16

17 Keywords: *Comamonas kerstersii*, bacteremia, intra-abdominal infection, *Delftia acidovorans*, MALDI-
18 TOF.

19 Abstract: 40 words

20 Text: 1232 words

21

22 **ABSTRACT**

23 We report for the first time a case of bacteremia caused by *Comamonas kerstersii* in a 65-year-old patient
24 with sign of diverticulosis. In addition, we review the isolation of *Comamonas* sp. and related organisms
25 in our hospital over 25 years.

26

27 **CASE REPORT**

28 *Comamonas kerstersii* is a non-fermenting β -Proteobacteria described in 2003 that has long been
29 considered as non-pathogenic (19). This organism has been recently associated with intra-abdominal
30 infection consecutive to perforation of the digestive tract (1). Herein, we describe a case of polymicrobial
31 bacteremia due to *Comamonas kerstersii* and *Bacteroides fragilis* in a 65-year-old diabetic man that was
32 admitted to the emergency department of the hospital due to sudden onset of fever and chills. The patient
33 reported episodes of vomiting and diarrhea and mentioned that he drank water from a small river. Stool
34 cultures did not disclose *Salmonella*, *Shigella*, *Aeromonas* or *Campylobacter* species. The detection of
35 *Clostridium difficile* toxin A and B and glutamate dehydrogenase antigen was also negative. Blood
36 cultures (two pairs of bottles) were drawn from a peripheral vein and the patient was discharged under
37 treatment with oral ciprofloxacin for a gastroenteritis of unknown origin. The Blood cultures were
38 processed into a BACTEC FX automated blood culture system (Becton Dickinson, Sparks, MD). A first
39 aerobic blood culture bottle became positive and the Gram staining revealed the presence of long
40 filamentous Gram-negative bacilli (Figure 1). The bacterial identification by MALDI-TOF (Bruker
41 Daltonics GmbH, Leipzig, Germany) analysis was performed the same day using a protocol that we
42 recently developed based on the analyses of a bacterial pellet preparation from the blood bottles (5, 16,
43 17). The strain was identified as *Comamonas kerstersii*, a Gram-negative non-fermentative bacterium, and
44 prompted the hospitalization of the patient. The patient was afebrile at that time, but palpation of the left
45 lower abdominal quadrant was painful. An abdominal CT scan revealed diverticulosis without evidence
46 of diverticulitis. Consecutively, the anaerobic blood bottles from the same pair became positive for

47 *Bacteroides fragilis*. We monitored the following MIC ($\mu\text{g.ml}$) for the *Comamonas kerstersii* strain:
48 ceftazidime, 0.75; meropenem, 0.004; minocycline, 0.38; levofloxacin, 4; co-trimoxazole, >32;
49 ciprofloxacin, 32. For the *Bacteroides fragilis* strain, the MIC were: amoxicillin-clavulanate, 1.5;
50 piperacillin-tazobac, 6; imipenem, 0.12; meropenem, 0.025; metronidazole, 0.25; clindamycin, 16;
51 ciprofloxacin, 32. A treatment of imipenem-cilastatine was given for 10 days and the patient recovered.
52 The final diagnosis was a mixed bacteremia with *Comamonas kerstersii* and *Bacteroides fragilis* in the
53 setting of a diverticulosis.

54

55 Comamonads are Gram negative, non-fermentative bacteria, oxydase- and catalase-positive, largely
56 motile due to the presence of polar flagella. The *Comamonas* genus originally contained *Comamonas*
57 *terrigena*, *Comamonas testosteroni* (previously *Pseudomonas testosteroni*) and *Comamonas acidovorans*
58 (previously *Pseudomonas acidovorans*) (6). It now contains seventeen species while *Comamonas*
59 *acidovorans* has been separated from the *Comamonas* genus on the basis of 16S rRNA and is now known
60 as *Delftia acidovorans* (20). Although ubiquitously distributed in the environment (soil and water),
61 *Comamonas* and *Delftia* sp. are rarely associated with infections in humans. However, several
62 publications have incriminated *Comamonas testosteroni* and *Delftia acidovorans* in particular in human
63 diseases, including severe invasive infections such as bacteremia and meningitis (2-4, 7, 10, 11, 20).

64 *Comamonas kerstersii*, described in 2003 (19), has recently been reported as an agent of intra-abdominal
65 infection by Almuzara and colleagues (1). The present case is the first report of *Comamonas kerstersii*
66 bacteremia. We initially performed the identification of the strain at the species level directly from the
67 positive blood bottle using MALDI-TOF, with a spectral score of 2.176 (5, 16, 17). Subsequently, it was
68 recovered both from the blood agar plate (with a spectral score of 2.26), on which the growth was
69 maximal and from the “chocolate” agar plate that is supplemented with NAD (factor V) and hemin (factor
70 X). We also proceeded to the amplification and sequencing of the 16S rRNA ribosomal gene in order to
71 confirm the MALDI-TOF identification of this strain (8). The analysis of the sequences using the BLAST

72 V2.0 software (<http://www.ncbi.nlm.nih.gov/BLAST/>) showed 100% of identity with the sequences
73 corresponding to the 16S RNA ribosomal gene of *Comamonas kerstersii* strain LMG 5323 (19). From
74 both the blood bottle and the agar plates, the strain appeared as an extremely long Gram-negative
75 filamentous bacillus which is a very unusual phenotype for bacteria of this genus (Figure 1). The
76 *Comamonas* and *Delftia* strains previously isolated in our hospital are Gram-negative short bacilli or rods
77 (Figure 1), which is the morphology described for these organisms (19, 20).

78 Translocation from the digestive tract seems to be a predominant cause of infections by *Delftia* and
79 *Comamonas* species. Recently, Hagiya and colleagues reported a *Delftia acidovorans* bacteremia in a 46-
80 year-old woman caused by translocation of the bacteria consecutive to pesticide poisoning (10). A
81 bacteremia caused by *Comamonas testosteroni* was previously reported, in a 22-year-old man with
82 perforated appendix (9). In the four cases reported by Almuzara and colleagues, the *Comamonas*
83 *kerstersii* strains were isolated from intra-abdominal collections (1). We previously identified another
84 *Comamonas kerstersii* strain in an intra-peritoneal collection of an 11-year-old child with a perforated
85 appendix (table 1 and figure 1). Herein, the digestive origin of the *Comamonas kerstersii* strain is
86 supported by the fact that: i) the patient reported abdominal pain, vomiting and diarrhea, ii) the CT scan
87 revealed evidence of diverticulosis, and iii) the enteric bacteria *Bacteroides fragilis* was isolated from the
88 blood culture in this setting. The infection could originate from the water that the patient drank in the
89 countryside.

90 *Comamonads* have been rarely associated with infection in humans despite their ubiquitous distribution in
91 the environment possibly due to the difficulty to accurately distinguish *Comamonas* species from
92 *Pseudomonas* species in the pre-MALDI-TOF area (1). Alternatively, *Comamonads* could have been
93 under-recognized due to their common occurrence in the setting of a polymicrobial infection. In our 1027-
94 bed tertiary care university hospital, thirty-three *Comamonas* sp. strains and thirty-eight *Delftia*
95 *acidovorans* strains were isolated from 1997 to 2013. They were primarily isolated from respiratory
96 samples (33%), urogenital samples (23%) and digestive samples (21%); bacteremia represented 5% (three

97 patients) of all cases (Table 1). All 3 cases were poly-microbial bacteremia. The first bacteremia case
98 was due to *Comamonas testosteroni* in association with *Streptococcus parasanguis* and *Ralstonia pickettii*
99 in a 33-year old man. A second case involved *Delftia acidovorans* in association with *Streptococcus*
100 *agalactiae* in blood cultures from a 61-year-old man. The last case is the present *Comamonas kerstersii*
101 and *Bacteroides fragilis* co-infection.

102 Like *Delftia acidovorans*, *Comamonas testosteroni* is the *Comamonas* species predominantly associated
103 with bacteremia (table 1) (7, 9, 10, 20). Translocation from the digestive tract and catheters are the
104 predominant source of infection (9, 10, 13-15). Children or patients with compromised immune systems
105 (AIDS or patients treated with chemotherapies) appear to be particularly at risk to develop *Comamonas*
106 sp. or *Delftia acidovorans* bacteremia (12, 13). Interestingly, Khan and colleagues reported a fatal
107 outcome in an 4-year old immuno-competent child presenting a *Delftia acidovorans* bacteremia (12). The
108 patient presented herein did not display any sign of immunodeficiency suggesting that such bacteremia
109 may also occur in the absence of immunosuppression. The likely high inoculum in the water that was
110 drunk and the diabetic status of the patient are two significant co-factors that may explain the occurrence
111 of a bacteremia in the setting of a gastro-intestinal infection. Similarly, a *Comamonas* species bacteremia
112 has been associated with exposure to possibly contaminated water of a fish tank (18).

113 This report reveals that *Comamonas kerstersii* and other non-fermenting related bacteria may be
114 involved in severe diseases independently of perforation of the digestive tract. Moreover, this report
115 highlights the usefulness of MALDI-TOF in redefining the epidemiology and clinical syndromes due to
116 some non-fermentative Gram negative bacteria that were difficult to identify in the pre-MALDI-TOF
117 area.

118 **ACKNOWLEDGEMENTS**

119 We are grateful to all the technicians of the Diagnostic Microbiology Laboratory of the Lausanne
120 Hospital for their technical contribution. In particular, we thank Maria Senra-Ortiz and Christian
121 Durussel.

- 123 1. **Almuzara, M. N., R. Cittadini, C. Vera Ocampo, R. Bakai, G. Traglia, M. S. Ramirez, M.**
124 **del Castillo, and C. A. Vay.** 2013. Intra-abdominal infections due to *Comamonas kerstersii*. *J*
125 *Clin Microbiol* **51**:1998-2000.
- 126 2. **Arda, B., S. Aydemir, T. Yamazhan, A. Hassan, A. Tunger, and D. Serter.** 2003. *Comamonas*
127 *testosteroni* meningitis in a patient with recurrent cholesteatoma. *APMIS* **111**:474-6.
- 128 3. **Chotikanatis, K., M. Backer, G. Rosas-Garcia, and M. R. Hammerschlag.** 2011. Recurrent
129 intravascular-catheter-related bacteremia caused by *Delftia acidovorans* in a hemodialysis patient.
130 *J Clin Microbiol* **49**:3418-21.
- 131 4. **Cooper, G. R., E. D. Staples, K. A. Iczkowski, and C. J. Clancy.** 2005. *Comamonas*
132 (*Pseudomonas*) *testosteroni* endocarditis. *Cardiovasc Pathol* **14**:145-9.
- 133 5. **Croxatto, A., G. Prod'hom, and G. Greub.** Applications of MALDI-TOF mass spectrometry in
134 clinical diagnostic microbiology. *FEMS Microbiol Rev* **36**:380-407.
- 135 6. **De Vos, P., K. Kersters, E. Falsen, B. Pot, M. Gillis, P. Segers, and J. De Ley.** 1985.
136 *Comamonas* Davis and Park 1962 gen. nov., nom. rev. emend., and *Comamonas terrigena* Hugh
137 1962 sp. nov., nom. rev. *International Journal of Systematic Bacteriology* **35**:443– 453.
- 138 7. **Ender, P. T., D. P. Dooley, and R. H. Moore.** 1996. Vascular catheter-related *Comamonas*
139 *acidovorans* bacteremia managed with preservation of the catheter. *Pediatr Infect Dis J* **15**:918-
140 20.
- 141 8. **Goldenberger, D., T. Schmidheini, and M. Altwegg.** 1997. Detection of *Bartonella henselae*
142 and *Bartonella quintana* by a simple and rapid procedure using broad-range PCR amplification
143 and direct single-strand sequencing of part of the 16S rRNA gene. *Clin Microbiol Infect* **3**:240-
144 245.
- 145 9. **Gul, M., P. Ciragil, E. Bulbuloglu, M. Aral, S. Alkis, and F. Ezberci.** 2007. *Comamonas*
146 *testosteroni* bacteremia in a patient with perforated acute appendicitis. Short communication.
147 *Acta Microbiol Immunol Hung* **54**:317-21.
- 148 10. **Hagiya, H., T. Murase, J. Sugiyama, Y. Kuroe, H. Nojima, H. Naito, S. Hagioka, and N.**
149 **Morimoto.** 2013. *Delftia acidovorans* bacteremia caused by bacterial translocation after
150 organophosphorus poisoning in an immunocompetent adult patient. *J Infect Chemother* **19**:338-
151 41.
- 152 11. **Khan, S., S. Sistla, R. Dhodapkar, and S. C. Parija.** 2012. Fatal *Delftia acidovorans* infection
153 in an immunocompetent patient with empyema. *Asian Pac J Trop Biomed* **2**:923-4.
- 154 12. **Khan, S., S. Sistla, R. Dhodapkar, and S. C. Parija.** Fatal *Delftia acidovorans* infection in an
155 immunocompetent patient with empyema. *Asian Pac J Trop Biomed* **2**:923-4.
- 156 13. **Le Moal, G., M. Paccalin, J. P. Breux, F. Roblot, P. Roblot, and B. Becq-Giraudon.** 2001.
157 Central venous catheter-related infection due to *Comamonas testosteroni* in a woman with breast
158 cancer. *Scand J Infect Dis* **33**:627-8.
- 159 14. **Nseir, W., J. Khateeb, M. Awawdeh, and M. Ghali.** Catheter-related bacteremia caused by
160 *Comamonas testosteroni* in a hemodialysis patient. *Hemodial Int* **15**:293-6.
- 161 15. **Nseir, W., J. Khateeb, M. Awawdeh, and M. Ghali.** 2011. Catheter-related bacteremia caused
162 by *Comamonas testosteroni* in a hemodialysis patient. *Hemodial Int* **15**:293-6.
- 163 16. **Prod'hom, G., A. Bizzini, C. Durussel, J. Bille, and G. Greub.** 2010. Matrix-assisted laser
164 desorption ionization-time of flight mass spectrometry for direct bacterial identification from
165 positive blood culture pellets. *J Clin Microbiol* **48**:1481-3.
- 166 17. **Prod'hom, G., C. Durussel, and G. Greub.** 2013. A simple blood-culture bacterial pellet
167 preparation for faster accurate direct bacterial identification and antibiotic susceptibility testing
168 with the VITEK 2 system. *J Med Microbiol* **62**:773-7.
- 169 18. **Smith, M. D., and J. D. Gradon.** 2003. Bacteremia due to *Comamonas* species possibly
170 associated with exposure to tropical fish. *South Med J* **96**:815-7.

- 171 19. **Wauters, G., T. De Baere, A. Willems, E. Falsen, and M. Vaneechoutte.** 2003. Description of
172 *Comamonas aquatica* comb. nov. and *Comamonas kerstersii* sp. nov. for two subgroups of
173 *Comamonas terrigena* and emended description of *Comamonas terrigena*. Int J Syst Evol
174 Microbiol **53**:859-62.
- 175 20. **Wen, A., M. Fegan, C. Hayward, S. Chakraborty, and L. I. Sly.** 1999. Phylogenetic
176 relationships among members of the Comamonadaceae, and description of *Delftia acidovorans*
177 (den Dooren de Jong 1926 and Tamaoka et al. 1987) gen. nov., comb. nov. Int J Syst Bacteriol
178 **49**;2:567-76.

179

180

181

182

183 **Figure legend:**

184 **Figure 1: Gram staining of *Comamonas kerstersii*, *Comamonas testosteroni* and *Delftia acidovorans***
185 **strains isolated in clinical samples from Lausanne University Hospital.** (A) The *Comamonas*
186 *kerstersii* strain of the present case report directly from the blood-cultures bottle or B after culture on
187 blood agar medium. The strain displays long filaments when compared to the other strains that appears as
188 Gram-negative short bacilli or rods. C) A *Comamonas kerstersii* strain identified in an intra-peritoneal
189 collection of an 11-year-old child with a perforated appendix. D) *Comamonas testosteroni* involved in a
190 bacteremia in a 33-year- old man. E) *Delftia acidovorans* identified in blood cultures from a 61-year-old
191 man.

Table 1: *Comamonas* sp. and *Delfia acidovorans* isolated from clinical samples in the Lausanne University Hospital from 1997 to 2013.

	Respiratory and ENT ^a	Urogenital ^b	Intra-abdominal ^c	Skin	Blood culture	Surgical wound	Others ^d	Total patient (samples)	% of patient
<i>Delfia acidovorans</i>	19 (20)	12 (13)	-	5	1 (13)	-	1 (4)	38 (55)	54.3
<i>Comamonas testosteroni</i>	2 (4)	3	8	1 (4)	1	4 (5)	1	20 (26)	28.57
<i>Comamonas kerstersii</i>	-	-	1	-	1	-	-	2	2.86
<i>Comamonas aquatica</i>	-	1 (4)	-	-	-	-	-	1 (4)	1.43
<i>Comamonas specis</i> ^e	2	- (2)	6 (8)	1	-	-	- (1)	9 (14)	12.86
Number of patients (number of samples)	23 (26)	16 (22)	15 (17)	7 (10)	3 (15)	4 (5)	2 (6)	70 (101)	100
Percentage of patients (percentage of samples)	32.86 (25.74)	22.86 (21.78)	21.43 (16.83)	10 (9.9)	4.29 (14.85)	5.71 (4.95)	2.85 (5.94)	100	

^aEar nose and throat

^bUrine, vaginal swab and placenta

^cascitic fluid, peritoneal fluid, penrose liquid, kehr drain

^dstools (1), bone fragment (1), orifice smear (1)

^eno identification at the species level

Number represent patients, brackets represent the number of samples

