

Spatial and temporal shifts in the diet of the barnacle *Amphibalanus eburneus* within a subtropical estuary

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ABSTRACT

The success of many sessile invertebrates in marine benthic communities is linked to their ability to efficiently remove suspended organic matter from the surrounding water column. To investigate the diet of the barnacle Amphibalanus eburneus, a dominant suspension feeder within the Indian River Lagoon (IRL) of central Florida, we compared the stable isotopes ratios (δ^{13} C and δ^{15} N) of barnacle tissue to those of particulate organic matter (POM). Collections were carried out quarterly for a year from 29 permanent sites and at sites impacted by an Aureoumbra lagunensis bloom. δ^{13} C and δ^{15} N values of Amphibalanus eburneus varied across sites, but $\delta^{15}N$ was more stable over time. There was a range of $\delta^{15}N$ values of Amphibalanus eburneus tissue from 6.0% to 10.5% across sites. Because land-based sources such as sewage are generally enriched in ¹⁵N, this suggests a continuum of anthropogenic influence across sites in the IRL. Over 70% of the variation in δ^{15} N values of *Amphibalanus eburneus* across sites was driven by the δ^{15} N values of POM, supporting a generalist feeding strategy on available sources of suspended organic matter. The dominance of this generalist consumer in the IRL may be linked to its ability to consume spatially and temporally variable food resources derived from natural and anthropogenic sources, as well as Aureoumbra lagunensis cells. Generalist consumers such as Amphibalanus eburneus serve an important ecological role in this ecosystem and act as a sentinel species and recorder of local, site-specific isotopic baselines.

Subjects Biodiversity, Ecology, Marine BiologyKeywords Stable isotopes, Estuary, Indian River Lagoon, Suspension feeders, Barnacles, Generalist, Algal bloom

INTRODUCTION

Organisms that can exploit suspended organic matter fill a unique niche in aquatic ecosystems (*Ricciardi & Bourget*, 1999; *Riisgård & Larsen*, 2010). By efficiently consuming both living cells and detritus, suspension feeders play critical roles in the regulation of primary production and benthic-pelagic coupling of nutrients and organic matter (*Gili & Coma*, 1998). This feeding mechanism is widely successful in estuarine and coastal ecosystems, and coincides with highly diverse communities in benthic habitats (*Karlson & Osman*, 2012; *Cresson*, *Ruitton & Harmelin-Vivien*, 2016). Suspension

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feeders encounter various forms of organic matter that change over space and time (*Richoux, Vermeulen & Froneman, 2014*). These sources of nutrition vary in size and quality and can include bacteria [from $<2~\mu$ m], phytoplankton [2–200 μ m], zooplankton, and detritus from terrestrial plants, phytoplankton, and submerged aquatic vegetation (*Deegan & Garritt, 1997; Hsieh et al., 2000; Cresson, Ruitton & Harmelin-Vivien, 2016*).

Although suspension feeders appear to be ecologically similar and can occupy overlapping niches within a habitat, there is substantial variation in the feeding structures and mechanisms of coexisting species (Stuart & Klumpp, 1984; Lesser et al., 1992; Riisgård & Manríquez, 1997; Karlson, Gorokhova & Elmgren, 2015). This variation impacts an organism's ability to acquire, sort, and select food particles and may allow individual species to specialize in a subset of available suspended particulate matter (Dubois et al., 2007a; Riisgård & Larsen, 2010; Dubois & Colombo, 2014; Richoux, Vermeulen & Froneman, 2014; Cresson, Ruitton & Harmelin-Vivien, 2016; Whalen & Stachowicz, 2017). For example, bivalves are able to sort and selectively feed on particles, releasing inorganic matter via pseudofeces and efficiently retaining high quality particles greater than 5 µm (Jørgensen, 1974; Møhlenberg & Riisgård, 1978; Riisgård, 1988; Galimany et al., 2017a, 2017b). Tunicates, barnacles, and gastropod mollusks, in contrast, largely lack structures that facilitate particle selection and are considered generalist or indiscriminate suspension feeders (Lesser et al., 1992; Petersen, 2007; Dubois et al., 2007a; Cresson, Ruitton & Harmelin-Vivien, 2016). These generalists are, however, capable of retaining a broader range of particles that may not be consumed by selective suspension feeders or even adopting an omnivorous nutritional strategy that allows them to feed at higher trophic levels (Lesser et al., 1992; Bone, Carre & Chang, 2003; Petersen, 2007, Decottignies et al., 2007, Beninger et al., 2007, Kach & Ward, 2008, Riisgård & Larsen, 2010; Richoux, Vermeulen & Froneman, 2014; Cresson, Ruitton & Harmelin-Vivien, 2016).

Coastal ecosystems are subject to seasonal shifts in environmental conditions and strong resource gradients from a combination of inputs from marine and terrestrial sources (Deegan & Garritt, 1997). In addition, food webs in these systems are increasingly impacted by anthropogenically-derived pollution and harmful algal blooms (Carlton, Newman & Pitombo, 2011; Lapointe et al., 2015; Galimany et al., 2017b). Divergence in feeding mechanisms among coexisting suspension feeders may lead to differential responses of species to these changes and shape community composition across sites (Dubois et al., 2007a; Cresson, Ruitton & Harmelin-Vivien, 2016). Trophic plasticity may allow generalist species to exploit a broader range of ecological niches and adapt to both natural and anthropogenic changes in food availability. Barnacles, for instance, are a dominant component of many intertidal and estuarine ecosystems and are one of the most prolific invaders into coastal ecosystems worldwide (Carlton, Newman & Pitombo, 2011). Their success is likely linked to their ability to consume a wide range of particle sizes from zooplankton to phytoplankton (down to 2-5 μm), detritus, and organic matter from both natural and anthropogenic sources (Barnes, 1959; Crisp & Southward, 1961; Lesser et al., 1992; Riisgård & Larsen, 2010; Dolenec et al., 2006). The diet of barnacles has also been shown to vary over space and time (Dolenec et al., 2006; Dubois et al., 2007a;

Dubois & Colombo, 2014; Richoux, Vermeulen & Froneman, 2014) resulting from shifts in organic matter composition.

The Indian River Lagoon (IRL) of central Florida is a subtropical, shallow (mean depth of <1 m) estuary that spans 250 km of the east coast of central Florida. The IRL supports a high diversity of marine species due to a variety of habitats (mangrove, seagrass, oyster reefs, and artificial substrates) and its location in a tropical/temperate transition zone in close proximity to the Gulf Stream (Gilmore, 1995; Swain et al., 1995). Like many estuaries, both acute and chronic stressors are increasingly impacting communities within the IRL, leading to cascading effects throughout local food webs. For example, nutrient loading (Lapointe et al., 2015) and the loss of planktonic grazers have led to an increased frequency and severity of algal blooms (Phlips et al., 2014). The "superbloom" of a Picocyanobacteria and a Pedinophyceae (both 1-2 μm) sp. in 2011 and blooms of the brown tide Aureoumbra lagunensis (4-5 µm) in 2012, 2013, and the winter of 2016 (SJRWMD, 2013; Phlips et al., 2014; Kamerosky, Cho & Morris, 2015; Lapointe et al., 2015) were particularly devastating within the sublagoons of the northern IRL (NIRL). Although these algal species are all non-toxic, high concentrations of algal cells blocked sunlight from reaching seagrass beds and, ultimately, resulted in hypoxic events that led to fish kills within the NIRL (Phlips et al., 2014).

Surveys of epifauna communities at 90 sites in the NIRL have found high species diversity (175 taxa in 11 phyla) and a dominance of the barnacle *Amphibalanus eburneus*, with a mean percent cover of 30% to 40% across sites (D.S. Janiak, 2016, unpublished data; Fig. S1). The success of *Amphibalanus eburneus* across sites in the NIRL may be linked to its generalist feeding on available sources of suspended organic matter, but little is known about how the diet of *Amphibalanus eburneus* changes over time and space and whether these changes mirror general shifts in particulate organic matter (POM). To investigate this, we compared the stable isotope ratios (δ^{13} C and δ^{15} N) of *Amphibalanus eburneus* tissue to that of POM from the water column. Collections were carried out quarterly for a year and also during an *Aureoumbra lagunensis* bloom. We tested the following hypotheses: (1) the δ^{13} C and δ^{15} N values of *Amphibalanus eburneus* vary over space and time and will be closely tied to the δ^{13} C and δ^{15} N values of POM, and (2) *Amphibalanus eburneus* will demonstrate shifts in δ^{13} C and/or δ^{15} N values that indicate consumption of the brown tide *Aureoumbra lagunensis*.

MATERIALS AND METHODS

Sample collection

As part of a project monitoring epifaunal community composition and diversity over space and time, we established 29 permanent monitoring sites spanning 150 km within the three sub-lagoons (Indian River and Mosquito Lagoons, and the Banana River) of the northern region of the greater IRL (NIRL; Fig. 1; Table S1). Collections of *Amphibalanus eburneus* (N = 5-10 individuals) were carried out at each of these monitoring sites on a quarterly basis (in January, April, July, and October of 2015). Permits for species collections were provided by the Florida Fish and Wildlife Conservation Commission (SAL-14-1567-SR). To test whether a bloom of *Aureoumbra lagunensis* influenced the diet

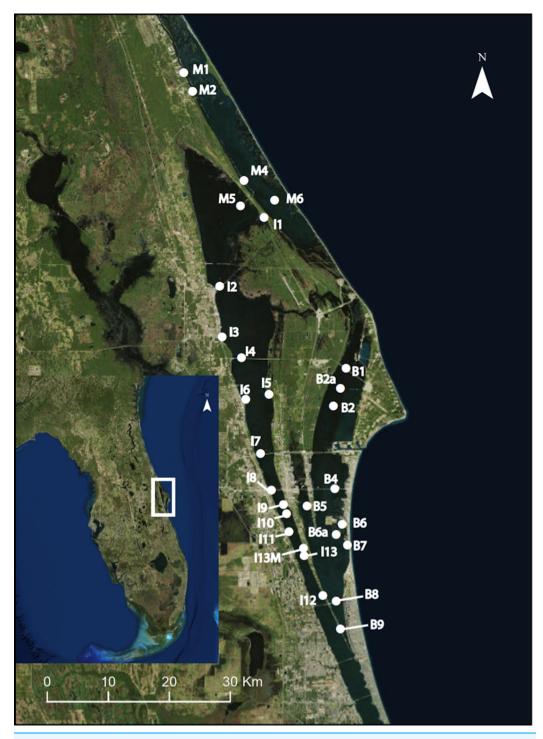


Figure 1 Map of 29 permanent monitoring sites established in the three sublagoons (Banana River [B], Indian River Lagoon [I], and Mosquito Lagoon [M]) of the Northern IRL of central Florida (inset map). Map data: Google, SIO, NOAA, U.S. Navy, NGA, GEBCO, and Landsat/Copernicus.

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of *Amphibalanus eburneus*, samples were also collected opportunistically at sites within the Banana River and IRL that were influenced by *Aureoumbra lagunensis* from December of 2015 to March of 2016 (Fig. S2; *Galimany et al., 2017a*). Barnacles were removed from mangrove prop roots or artificial substrates (dock and bridge pilings or seawalls) using a paint scraper and placed into a 4 L plastic bag containing seawater for transit back to the Smithsonian Marine Station. There were missing data for some sites on particular dates resulting from sample loss or inaccessibility.

During seasonal and *Aureoumbra lagunensis* collections, a sample of seawater (20 L) for POM was also taken at each site to monitor the $\delta^{13}C$ and $\delta^{15}N$ values of general sources of particulate carbon and nitrogen available to *Amphibalanus eburneus*. In the laboratory, this seawater was prefiltered through 105 μ m mesh and then filtered through a Millepore quartz fiber filter (2 μ m porosity) stacked on top of a Whatman glass fiber (GF) filter (0.7 μ m porosity) to obtain a single POM sample for each time point at each site. The organic matter content on GF filters was below detection limits, so POM $\delta^{13}C$ and $\delta^{15}N$ values are based on the results from organic matter on quartz fiber filters (2 μ m porosity) that were stacked on top of the GF/F.

Sample preparation and stable isotope (δ^{13} C and δ^{15} N) analysis

Samples of *Amphibalanus eburneus* were kept separate and held overnight in flowing, sand-filtered seawater to allow for gut evacuation and then frozen at –20 °C. Once thawed, the shell diameter was measured for each individual (*n* = 10 per site for each sampling period) and, using forceps, all tissue within the shell of a barnacle was placed into an individual glass vial (*Richoux, Vermeulen & Froneman, 2014*). Tissue was dried at 60 °C for 24 h and homogenized using a mortar and pestle. Homogenized samples were acidified to remove carbonate by exposure to 12 N HCl fumes for 12 h, after which samples were returned to the oven at 60 °C for 24 h (*Freeman & Thacker, 2011*). Quartz and GF filters containing POM were also dried and acidified prior to analysis. POM was scraped from each filter, and POM from each filter and barnacle samples were separately weighed to the nearest 0.001 mg into tared tin capsules. Isotope analysis was carried out at the Stable Isotope Facility at UC Davis using a PDZ Europa ANCA-GSL (for barnacle tissue samples) or Micro Cube (for POM samples) elemental analyzer interfaced to a PDZ Europa 20–20 isotope ratio mass spectrometer (Sercon Ltd, Cheshire, UK). Isotope values are reported in δ notation in units of permille (‰).

Data analysis

Isotope values provide a time-integrated record of an organism's diet, with δ^{13} C values providing information about the primary sources of carbon fueling local food webs and δ^{15} N values acting as a proxy for trophic level and the sources of nitrogen assimilated (*Michener & Kaufman*, 2007). To test the effect of season and site on the placement of *Amphibalanus eburneus* samples within bivariate (δ^{13} C and δ^{15} N) isotopic space, we calculated isotopic dissimilarity (measured as Euclidean distance) among samples and analyzed dissimilarity using a permutational multivariate analysis of variance (PERMANOVA) (Primer 6 with PERMANOVA+ add-on). Seasonal variation in

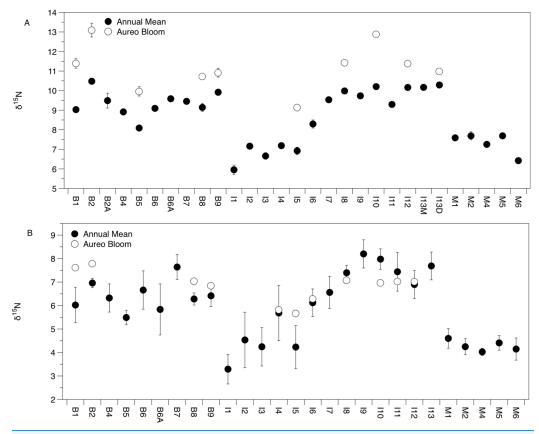


Figure 2 Mean (\pm SE) δ^{15} N values of *Amphibalanus eburneus* tissue (A) and particulate organic matter (POM; (B)) at individual sites within the three sublagoons of the northern Indian River Lagoon. Sublagoons include the Banana River [B], Indian River Lagoon [I], and Mosquito Lagoon [M]. Data are shown as annual means (from each season from January to October of 2015) and mean (*Amphibalanus eburneus* tissue) and single POM values from an *Aureoumbra lagunensis* bloom in January 2016.

individual isotope values was assessed using a Kruskal–Wallis One-Way Analysis of Variance (ANOVA). Linear regressions were used to investigate the relationship between the δ^{13} C and δ^{15} N values of *Amphibalanus eburneus* tissue and the δ^{13} C and δ^{15} N values of POM at each site. These analyses were carried out using Systat.

RESULTS

There was significant variation in the $\delta^{15}N$ and $\delta^{13}C$ values of *Amphibalanus eburneus* tissue over space and time (PERMANOVA testing the effect of site [PseudoF29,1046 = 23.89; $p_{(perm)} = 0.001$] and season [PseudoF3,1072 = 140.6; $p_{(perm)} = 0.001$]) (Figs. 2A and 3A; Fig. S3 in supplemental information) during annual collections. Within each season, $\delta^{15}N$ and $\delta^{13}C$ values also varied among sites (PERMANOVA testing the effect of individual site nested within collection period: PseudoF96,976 = 56.58; $p_{(perm)} = 0.001$) (Fig. S3). Annual mean $\delta^{15}N$ values from each site ranged from 6.0% to 10.5%, with depleted (6.0% to 7.7%) $\delta^{15}N$ values in the northern region of the IRL (site #s I1–5) and in the Mosquito Lagoon (site #s M1–6) compared to sites in the Banana River and the southern IRL (site #s B

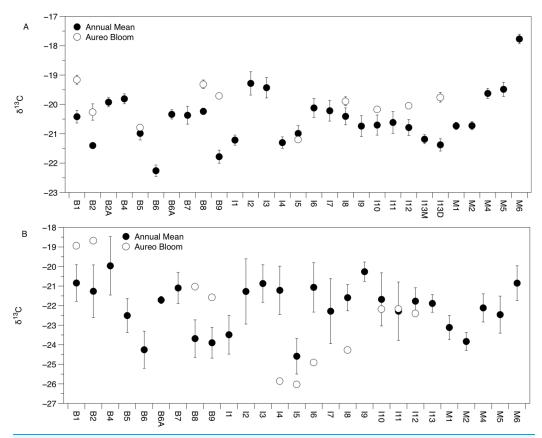


Figure 3 Mean (\pm SE) δ^{13} C values of Amphibalanus eburneus tissue (A) and particulate organic matter (POM; (B)) at sites within the sublagoons of the northern Indian River Lagoon. Sublagoons include the Banana River [B], Indian River Lagoon [I], and Mosquito Lagoon [M]. Data are shown as annual mean (from each season from January to October of 2015) and mean (Amphibalanus eburneus tissue) and single POM values from an Aureoumbra lagunensis bloom in January 2016.

and I6–13) that had δ^{15} N values between 8.3‰ and 10.5‰ (Fig. 2A). Annual mean δ^{13} C values ranged from –22.3‰ to –17.8‰, with variation at finer spatial scales (within lagoons and even between geographically adjacent sites) than δ^{15} N values (Fig. 3A; Fig. S3). In addition, δ^{13} C values varied more across seasons than δ^{15} N values (Kruskal–Wallis: $H: 506_3$; p < 0.001 and $H: 10.2_3$, p < 0.05 for δ^{13} C and δ^{15} N, respectively) (Figs. 2A and 3A; Fig. S3).

The δ^{13} C and δ^{15} N values of POM varied across space and time (Figs. 2B, 3B and 4; Fig. S4), with a range of annual means from -24.6% to -20% and 3.3% to 8.2% for δ^{13} C and δ^{15} N, respectively. The average δ^{13} C and δ^{15} N values of POM at each site explained 22% and 71% of the variation in the mean δ^{13} C and δ^{15} N values of *Amphibalanus eburneus* at the same site (linear regression: $r^2 = 0.22$; p < 0.01; N = 35 and $r^2 = 0.71$; p < 0.001; N = 35 for δ^{13} C and δ^{15} N values, respectively) (Fig. 4; Fig. S4 for individual site values for each season). Tissue values of *Amphibalanus eburneus* were, on average, enriched in both δ^{15} N and δ^{13} C compared to POM (+2.78 \pm 0.15 SE and +1.76 \pm 0.17 SE for δ^{15} N and δ^{13} C, respectively) (Fig. 4; Fig. S4).

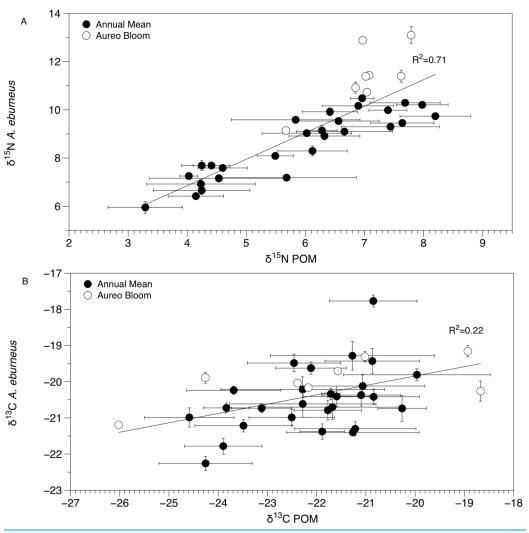


Figure 4 Mean (\pm SE) $\delta^{15}N$ (A) and $\delta^{13}C$ (B) values of Amphibalanus eburneus tissue at a site as a function of mean (\pm SE) $\delta^{15}N$ and $\delta^{13}C$ values of particulate organic matter (POM) at the same site. Data include 29 sites within the sublagoons of the northern Indian River Lagoon. Tissue and POM samples were taken each season from January to October of 2015 (shaded circles) and also during an Aureoumbra lagunensis bloom in January of 2016 (open circles).

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 δ^{15} N of *Amphibalanus eburneus* tissue was between 9.1‰ and 13.1‰ during the *Aureoumbra lagunensis* bloom, with an average enrichment of 1.8‰ (range of 0.7 to 2.7‰) compared to annual mean values (Figs. 2A, 4A and 5; Fig. S3). Likewise, δ^{13} C of *Amphibalanus eburneus* tissue during the *Aureoumbra* bloom ranged from –21‰ to –19‰, with a mean enrichment of 0.9‰ (range of –0.2‰ to 2.1‰) compared to the annual mean at a site (Figs. 3A, 4B and 5; Fig. S3). The δ^{15} N and δ^{13} C values of POM were variable during the *Aureoumbra lagunensis* bloom, with enrichment at some, but not all, sites (mean enrichment of 0.3‰ [range of –1‰ to 1.6‰] and –0.4‰ [range of –4.6‰ to 2.7‰] for δ^{15} N and δ^{13} C, respectively) relative to the annual mean (Figs. 2B, 3B and 4; Fig. S4). Tissue values of *Amphibalanus eburneus* were enriched in

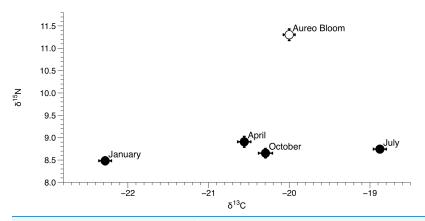


Figure 5 Mean (\pm SE) δ^{15} N and δ^{13} C values of Amphibalanus eburneus for each season from January to October of 2015 and during an Aureoumbra lagunensis bloom in January 2016. Values were calculated from δ^{15} N and δ^{13} C values of Amphibalanus eburneus from all sites within the three sublagoons of the Northern Indian River Lagoon. Full-size DOI: 10.7717/peerj.5485/fig-5

both $\delta^{15}N$ and $\delta^{13}C$ relative to POM under bloom conditions (+4.37 \pm 0.30 SE (range +3.47 to +5.9) and +1.91 \pm 0.75 SE (range –1.59 to +4.83) for $\delta^{15}N$ and $\delta^{13}C$, respectively (Fig. 4; Fig. S4).

DISCUSSION

Trophic ecology of Amphibalanus eburneus

The δ^{13} C and δ^{15} N values of POM suggest that particulate sources of organic matter vary across seasons and small spatial scales in the IRL. Variation in δ^{13} C values is likely due to inputs of organic carbon from a combination of marine phytoplankton (δ^{13} C values of -18‰ to -24‰) and detritus from terrestrial C₃ plants such as mangroves (-35‰ to -25‰) and seagrasses (-13.5‰ and -5.2‰) (Deegan & Garritt, 1997; Michener & *Kaufman*, 2007). Likewise, the δ^{15} N values of POM provide information about the sources of nitrogen fueling local food webs. For instance, while depleted $\delta^{15}N$ values suggest natural N-fixation, elevated δ¹⁵N values (>3‰) are suggestive of nitrogen derived from ¹⁵N-enriched sources such as wastewater (*Lapointe et al.*, 2015). POM δ^{15} N values in our study that range from ~3 to 8‰ therefore suggest a continuum of impact from anthropogenically-derived nutrients across sites in the IRL. High levels of dissolved inorganic nitrogen and total dissolved nitrogen have been reported previously in regions of the northern IRL resulting from long water residence times and inputs of anthropogenically-derived nitrogen via surface water runoff and groundwater from septic tanks (Smith, 1993; Lapointe et al., 2015). Our data suggest that sites close to human development in the Banana River and southern sites in the Indian River (δ^{15} N values of ~6 to 8‰) are more impacted by these ¹⁵N-enriched sources than those in the more sparsely populated Mosquito Lagoon and northern IRL sites; our POM values are in general agreement with the magnitude of ¹⁵N enrichment found in macroalgae from this region of the IRL (Lapointe et al., 2015).

The isotopic composition of *Amphibalanus eburneus* tissue was coupled to the temporal and spatial dynamics of POM δ^{15} N and, to a lesser extent, δ^{13} C values, supporting the

contention that barnacles are generalist suspension feeders utilizing predominant components of the organic matter pool in the water column (Cresson, Ruitton & Harmelin-Vivien, 2016). In contrast, if barnacles had a broader capacity to sort and select particles based on size or nutritional quality, we would expect less variation in the δ^{13} C and $\delta^{15}N$ of barnacle tissue over time and space and a decoupling of POM and Amphibalanus eburneus isotope values (Decottignies et al., 2007; Dubois et al., 2007a; Dubois & Colombo, 2014). For example, in the oyster Crassostrea gigas, the δ^{13} C and δ^{15} N values of POM explained less than 5% of the variation in isotope values of oyster muscle tissue (Marchais et al., 2013). Although this relationship was significant in our study (explaining 71% and 22% of the variance for mean $\delta^{15}N$ and $\delta^{13}C$ values at each site, respectively), the trend may have been even stronger if we had finer temporal resolution instead of a single POM isotope "snapshot" for each season from a site. The tissue of Amphibalanus eburneus was generally enriched in ¹³C and ¹⁵N (higher δ¹³C and δ¹⁵N values) relative to POM (by +1.76‰ and +2.78‰, respectively). Because consumers are generally enriched in both ¹³C and ¹⁵N due to the process of trophic enrichment, this implies the presence of a "trophic step" between particulate matter and Amphibalanus eburneus (Dubois et al., 2007b). The magnitude of trophic enrichment varies across species, but is generally hypothesized to range from <1.0% to 2.0% for δ^{13} C and 3.0% to 3.6% for $\delta^{15}N$ (DeNiro & Epstein, 1981; Zanden & Rasmussen, 2001; McCutchan et al., 2003; Dubois et al., 2007b). Our values are therefore well within the estimated range for trophic enrichment, supporting generalist feeding on bulk POM by Amphibalanus eburneus (Hsieh et al., 2000).

Unlike many suspension feeders, barnacles are able to consume higher trophic level prey such as zooplankton (*Richoux, Vermeulen & Froneman, 2014*). Enriched δ¹⁵N values of Amphibalanus eburneus from the IRL may therefore reflect feeding on zooplankton (Dix & Hanisak, 2015). We posit, however, that if zooplankton had been a dominant component of the diet of Amphibalanus eburneus in the IRL, we would have observed an additional trophic step between POM (predominately phytoplankton) and Amphibalanus eburneus tissue isotope values. Enriched $\delta^{15}N$ values are thus likely the result of the passage of anthropogenically-derived nitrogen assimilated by phytoplankton into the epifaunal food web. Enriched $\delta^{15}N$ values in POM and barnacle tissue at sites impacted by anthropogenically-derived nutrients has been reported before in other systems, with up to a 5% enrichment in barnacles from impacted sites and strong linear correlations between POM and barnacle δ^{15} N values (*Dolenec et al.*, 2006, 2007). Elevated δ^{15} N values of POM and Amphibalanus eburneus tissue (>8%) in the more urbanized regions of the NIRL (Banana River and southern sites in the Indian River) and relatively stable $\delta^{15}N$ values across seasons therefore suggest chronic exposure to nitrogen from anthropogenic sources at some sites. In contrast, δ^{13} C values within a site were more variable over both space and time, implying shifts in carbon sources over small spatial scales and the potential for seasonal fluctuations in phytoplankton productivity or growth rates (Cifuentes, Sharp & Fogel, 1988). It is possible that higher variability in δ^{13} C values of *Amphibalanus eburneus* tissue over time may be due to

differences in the turnover rate of this isotope relative to $\delta^{15}N$ (*Dubois et al.*, 2007b), but little is currently known about the turnover rate of carbon and nitrogen isotopes in barnacles.

Amphibalanus eburneus and algal blooms

The IRL has been exposed to multiple, acute algal blooms over the past decade, with a particularly detrimental superbloom in 2011 and recurring Aureoumbra lagunensis blooms in 2012, 2013, and 2016 (Phlips et al., 2014; Galimany et al., 2017a). The Aureoumbra bloom in 2016 occurred after we had gathered a year of baseline quarterly sampling on Amphibalanus eburneus feeding in the NIRL, providing an opportunity to investigate how the trophic ecology of Amphibalanus eburneus changes during an algal bloom. A single "snapshot" assessment of the $\delta^{15}N$ and $\delta^{13}C$ values of POM during the Aureoumbra bloom demonstrated $\delta^{15}N$ values that were at or above annual means and δ¹³C values that were highly variable, with evidence of both enrichment and depletion in δ^{13} C relative to annual means. In contrast, a more time integrated assessment (via δ^{15} N and δ^{13} C values of Amphibalanus eburneus tissue) revealed that δ^{15} N and, to a lesser extent, δ¹³C values of Amphibalanus eburneus were enriched during the Aureoumbra bloom compared to annual means. For δ^{13} C, this shift likely reflects a combination of source homogenization (predominately Aureoumbra) and isotopic fractionation associated with algal growth and high productivity that alters the δ^{13} C signal at the base of the food web (Cifuentes, Sharp & Fogel, 1988). Although the average enrichment of δ^{13} C in Amphibalanus eburneus tissue relative to POM was still within the range of a trophic step (mean 1.91% ± 0.75 SE) during the bloom, substantial variation in this enrichment (range -1.59% to 4.83%) suggests that direct reliance of Amphibalanus eburneus on phytoplankton may be reduced during blooms.

Enrichment of $\delta^{15}N$ (up to 13%) may reflect increased Amphibalanus eburneus consumption of phytoplankton that are relying on enriched sources of anthropogenicallyderived N. Although blooms of Aureoumbra in the IRL have previously been shown to elicit lower POM^{15N} values than under non-bloom conditions (Kang, Koch & Gobler, 2015), Aureoumbra is also known to rapidly assimilate NH₄, a common component of nitrogen derived from anthropogenic sources such as septic tanks (Lapointe et al., 2015; Kang, Koch & Gobler, 2015). We therefore propose that Amphibalanus eburneus may be consuming ¹⁵N-enriched *Aureoumbra* cells at bloom sites. Alternatively, because barnacles have been shown to reduce feeding efficiency at lower particle sizes (3 to 5 µm; Lesser et al., 1992), enriched δ^{15} N values under bloom conditions may also reflect an additional trophic step (δ^{15} N values of Amphibalanus eburneus tissue was on average enriched by 4.37‰ (range of 3.47% to 5.9%) compared to POM during the bloom) as Amphibalanus eburneus is feeding more heavily on zooplankton than phytoplankton during an algal bloom. These data provide initial evidence of nutritional shifts in Amphibalanus eburneus under bloom conditions, but additional work in the laboratory is needed to verify the role of this epifaunal species in bloom mitigation (Galimany et al., 2017b).

CONCLUSIONS

The generalist feeding strategy of Amphibalanus eburneus appears to allow it to exploit spatially and temporally variable sources of organic matter and may contribute to the successful dominance of this species across sites in the NIRL (Carlton, Newman & Pitombo, 2011; Karlson & Osman, 2012). The abundance of Amphibalanus eburneus on diverse substrates and across both impacted and pristine sites within the NIRL is in direct contrast to other epifaunal organisms that are more constrained in their distribution or are currently present at only a fraction of their historical abundance (e.g., Crassostrea virginica and Mercenaria mercenaria; MacKenzie, Taylor & Arnold, 2001; Wilson et al., 2005; Garvis, Sacks & Walters, 2015; D.S. Janiak, 2016, unpublished data). As a dominant and stable faunal component of the NIRL, Amphibalanus eburneus is likely playing an important role in nutrient and organic matter cycling in this system (*Dubois et al.*, 2007a). In addition, with a wide distribution and an integration of diverse sources of carbon and nitrogen into its tissues, Amphibalanus eburneus acts as an important sentinel species and recorder of isotopic baselines (Post, 2002; Dolenec et al., 2006; Fukumori et al., 2008). Finally, its nutritional plasticity may also allow Amphibalanus eburneus to capture and consume bloom particles such as Aureoumbra, providing a potential means for bioremediation and the prevention of algal blooms via top-down control.

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ADDITIONAL INFORMATION AND DECLARATIONS

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Competing Interests

The authors declare that they have no competing interests.

Author Contributions

• Christopher J. Freeman conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.

- Dean S. Janiak conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Malcolm Mossop performed the experiments, analyzed the data, authored or reviewed drafts of the paper, approved the final draft.
- Richard Osman conceived and designed the experiments, performed the experiments, authored or reviewed drafts of the paper, approved the final draft.
- Valerie J. Paul conceived and designed the experiments, performed the experiments, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft.

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Data Availability

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Supplemental Information

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REFERENCES

- **Barnes H. 1959.** Stomach contents and microfeeding of some common cirripedes. *Canadian Journal of Zoology* **37(3)**:231–236 DOI 10.1139/z59-027.
- Beninger PG, Decottignies P, Guiheneuf F, Barillé L, Rincé Y. 2007. Comparison of particle processing by two introduced suspension feeders: selection in *Crepidula fornicata* and *Crassostrea gigas*. *Marine Ecology Progress Series* 334:165–177 DOI 10.3354/meps334165.
- Bone Q, Carre C, Chang P. 2003. Tunicate feeding filters. *Journal of the Marine Biological Association of the United Kingdom* 83(5):907–919 DOI 10.1017/s002531540300804xh.
- **Carlton JT, Newman WA, Pitombo FB. 2011.** Barnacle invasions: introduced, cryptogenic, and range expanding Cirripedia of North and South America. In: Galil BS, Clark PF, Carlton JT, eds. *In the Wrong Place-Alien Marine Crustaceans: Distribution, Biology and Impacts.* London: Springer, 159–213.
- **Cifuentes LA, Sharp JH, Fogel ML. 1988.** Stable carbon and nitrogen isotope biogeochemistry in the Delaware estuary. *Limnology and Oceanography* **33(5)**:1102–1115 DOI 10.4319/lo.1988.33.5.1102.
- Cresson P, Ruitton S, Harmelin-Vivien M. 2016. Feeding strategies of co-occurring suspension feeders in an oligotrophic environment. *Food Webs* 6:19–28 DOI 10.1016/j.fooweb.2015.12.002.
- Crisp DJ, Southward AJ. 1961. Different types of cirral activity of barnacles. *Philosophical Transactions of the Royal Society B: Biological Sciences* 243(705):271–307 DOI 10.1098/rstb.1961.0003.

- **Decottignies P, Beninger PG, Rincé Y, Riera P. 2007.** Trophic interactions between two introduced suspension-feeders, *Crepidula fornicata* and *Crassostrea gigas*, are influenced by seasonal effects and qualitative selection capacity. *Journal of Experimental Marine Biology and Ecology* **342(2)**:231–241 DOI 10.1016/j.jembe.2006.10.005.
- **Deegan LA, Garritt RH. 1997.** Evidence for spatial variability in estuarine food webs. *Marine Ecology Progress Series* **147**:31–47 DOI 10.3354/meps147031.
- **DeNiro MJ, Epstein S. 1981.** Influence of diet on the distribution of nitrogen isotopes in animals. *Geochimica et Cosmochimica Acta* **45(3)**:341–351 DOI 10.1016/0016-7037(81)90244-1.
- **Dix N, Hanisak MD. 2015.** Microzooplankton grazing experiments in the subtropical Indian River Lagoon, Florida challenge assumptions of the dilution technique. *Journal of Experimental Marine Biology and Ecology* **465**:1–10 DOI 10.1016/j.jembe.2014.12.010.
- **Dolenec T, Lojen S, Dolenec M, Lambaša Z, Dobnikar M, Rogan N. 2006.** ¹⁵N and ¹³C Enrichment in *Balanus perforatus*: tracers of municipal particulate waste in the Murter sea (Central Adriatic, Croatia). *Acta Chimica Slovenica* **53**:469–476.
- **Dolenec T, Lojen S, Kniewald G, Dolenec M, Rogan N. 2007.** Nitrogen stable isotope composition as a tracer of fish farming in invertebrates *Aplysina aerophoba*, *Balanus perforatus*, and *Anemonia sulcata* in central Adriatic. *Aquaculture* **262(1–2)**:237–249 DOI 10.1016/j.aquaculture.2006.11.029.
- **Dubois SF, Colombo F. 2014.** How picky can you be? Temporal variations in trophic niches of co-occurring suspension-feeding species. *Food Webs* **1(1–4)**:1–9 DOI 10.1016/j.fooweb.2014.07.001.
- **Dubois S, Jean-Louis B, Bertrand B, Lefebvre S. 2007b.** Isotope trophic-step fractionation of suspension-feeding species: implications for food partitioning in coastal ecosystems. *Journal of Experimental Marine Biology and Ecology* **351(1–2)**:121–128 DOI 10.1016/j.jembe.2007.06.020.
- **Dubois S, Orvain F, Marin-Léal JC, Ropert M, Lefebvre S. 2007a.** Small-scale spatial variability of food partitioning between cultivated oysters and associated suspension-feeding species, as revealed by stable isotopes. *Marine Ecology Progress Series* **336**:151–160 DOI 10.3354/meps336151.
- **Freeman CJ, Thacker RW. 2011.** Complex interactions between marine sponges and their symbiotic microbial communities. *Limnology and Oceanography* **56**(**5**):1577–1586 DOI 10.4319/lo.2011.56.5.1577.
- Fukumori K, Oi M, Doi H, Takahashi D, Okuda N, Miller TW, Kuwae M, Miyasaka H, Genkai-Kato M, Koizumi Y, Omori K. 2008. Bivalve tissue as a carbon and nitrogen isotope baseline indicator in coastal ecosystems. *Estuarine*, *Coastal*, *and Shelf Science* 79(1):45–50 DOI 10.1016/j.ecss.2008.03.004.
- Galimany E, Freeman CJ, Lunt J, Domingos A, Sacks P, Walters L. 2017b. Feeding competition between the native oyster *Crassostrea virginica* and the invasive mussel *Mytella charruana*. *Marine Ecology Progress Series* 564:57–66 DOI 10.3354/meps11976.
- Galimany E, Lunt J, Freeman CJ, Reed S, Segura-García I, Paul VJ. 2017a. Feeding behavior of eastern oysters *Crassostrea virginica* and hard clams *Mercenaria mercenaria* in shallow estuaries. *Marine Ecology Progress Series* 567:125–137 DOI 10.3354/meps12050.
- Garvis SK, Sacks PE, Walters LJ. 2015. Formation, movement, and restoration of dead intertidal oyster reefs in Canaveral National Seashore and Mosquito Lagoon, Florida. *Journal of Shellfish Research* 34(2):251–258 DOI 10.2983/035.034.0206.
- Gili JM, Coma R. 1998. Benthic suspension feeders: their paramount role in littoral marine food webs. *Trends in Ecology and Evolution* 13(8):316–321 DOI 10.1016/s0169-5347(98)01365-2.

- **Gilmore GR. 1995.** Environmental and biogeographic factors influencing ichthyofaunal diversity: Indian River Lagoon. *Bulletin of Marine Science* **57(1)**:153–170.
- Hsieh HL, Kao WY, Chen CP, Liu PJ. 2000. Detrital flows through the feeding pathway of the oyster (*Crassostrea gigas*) in a tropical shallow lagoon: δ^{13} C signals. *Marine Biology* **136(4)**:677–684 DOI 10.1007/s002270050727.
- Jørgensen CB. 1974. On gill function in the mussel *Mytilus edulis* L. *Ophelia* 13(1-2):187-232 DOI 10.1080/00785326.1974.10430595.
- **Kach DJ, Ward JE. 2008.** The role of marine aggregates in the ingestion of picoplankton-size particles by suspension feeding molluscs. *Marine Biology* **153(5)**:797–805 DOI 10.1007/s00227-007-0852-4.
- Kamerosky A, Cho HJ, Morris L. 2015. Monitoring of the 2011 super algal bloom in Indian River Lagoon, FL, USA, using MERIS. *Remote Sensing* 7(2):1441–1460 DOI 10.3390/rs70201441.
- Kang Y, Koch F, Gobler CJ. 2015. The interactive roles of nutrient loading and zooplankton grazing in facilitating the expansion of harmful algal blooms caused by the pelagophyte, *Aureoumbra lagunensis*, to the Indian River Lagoon, FL, USA. *Harmful Algae* 49:162–173 DOI 10.1016/j.hal.2015.09.005.
- **Karlson AM, Gorokhova E, Elmgren R. 2015.** Do deposit-feeders compete? Isotopic niche analysis of an invasion in a species-poor system. *Scientific Reports* **5(1)**:1–8 DOI 10.1038/srep09715.
- **Karlson RH, Osman RW. 2012.** Species composition and geographic distribution of invertebrates in fouling communities along the east coast of the USA: a regional perspective. *Marine Ecology Progress Series* **458**:255–268 DOI 10.3354/meps09767.
- Lapointe BE, Herren LW, Debortoli DD, Vogel MA. 2015. Evidence of sewage-driven eutrophication and harmful algal blooms in Florida's Indian River Lagoon. *Harmful Algae* 43:82–102 DOI 10.1016/j.hal.2015.01.004.
- **Lesser MP, Shumway SE, Cucci T, Smith J. 1992.** Impact of fouling organisms on mussel rope culture: interspecific competition for food among suspension-feeding invertebrates. *Journal of Experimental Marine Biology and Ecology* **165(1)**:91–102 DOI 10.1016/0022-0981(92)90291-h.
- **MacKenzie CL Jr, Taylor DL, Arnold WS. 2001.** A history of hard clamming. In: Kraeurer JN, Castagna M, eds. *Biology of the Hard Clam*. Amsterdam: Elsevier, 651–671.
- Marchais V, Schaal G, Grall J, Lorrain A, Nerot C, Richard P, Chauvaud L. 2013. Spatial variability of stable isotope ratios in oysters (*Crassostrea gigas*) and primary producers along an estuarine gradient (Bay of Brest, France). *Estuaries and Coasts* 36(4):808–819 DOI 10.1007/s12237-012-9584-x.
- McCutchan JH, Lewis WM, Kendall C, McGrath CC. 2003. Variation in trophic shift for stable isotope ratios of carbon, nitrogen, and sulfur. *Oikos* 102(2):378–390 DOI 10.1034/j.1600-0706.2003.12098.x.
- **Michener RH, Kaufman L. 2007.** Stable isotope ratios as tracers in marine aquatic food webs: an update. In: Michener RH, Lajtha K, eds. *Stable Isotopes in Ecology and Environmental Science*. Second Edition. Oxford: Blackwell Publishing Ltd, 238–282.
- **Møhlenberg F, Riisgård HU. 1978.** Efficiency of particle retention in 13 species of suspension feeding bivalves. *Ophelia* **17(2)**:239–246 DOI 10.1080/00785326.1978.10425487.
- **Petersen JK. 2007.** Ascidian suspension feeding. *Journal of Experimental Marine Biology and Ecology* **342(1)**:127–137 DOI 10.1016/j.jembe.2006.10.023.
- Phlips EJ, Badylak S, Lasi MA, Chamberlain R, Green WC, Hall LM, Hart JA, Lockwood JC, Miller JD, Morris LJ, Steward JS. 2014. From red tides to green and brown tides: bloom dynamics in a restricted subtropical lagoon under shifting climatic conditions. *Estuaries and Coasts* 38(3):886–904 DOI 10.1007/s12237-014-9874-6.

- **Post DM. 2002.** Using stable isotopes to estimate trophic position: models, methods, and assumptions. *Ecology* **83(3)**:703–718 DOI 10.1890/0012-9658(2002)083[0703:USITET] 2.0.CO;2.
- **Ricciardi A, Bourget E. 1999.** Global patterns of macroinvertebrate biomass in marine intertidal communities. *Marine Ecology Progress Series* **185**:21–35 DOI 10.3354/meps185021.
- **Richoux NB, Vermeulen I, Froneman PW. 2014.** Stable isotope ratios indicate differential omnivory among syntopic rocky shore suspension-feeders. *Marine Biology* **161(5)**:971–984 DOI 10.1007/s00227-013-2358-6.
- **Riisgård HU. 1988.** Efficiency of particle retention and filtration rate in 6 species of Northeast American bivalves. *Marine Ecology Progress Series* **45(3)**:217–223 DOI 10.3354/meps045217.
- **Riisgård HU, Larsen PS. 2010.** Particle capture mechanisms in suspension-feeding invertebrates. *Marine Ecology Progress Series* **418**:255–293 DOI 10.3354/meps08755.
- **Riisgård HU, Manríquez P. 1997.** Filter-feeding in fifteen marine ectoprocts (Bryozoa): particle capture and water pumping. *Marine Ecology Progress Series* **154**:223–239 DOI 10.3354/meps154223.
- **SJRWMD** (St. Johns River Water Management District). 2013. The Indian River Lagoon: an estuary of national significance. *Available at https://www.sjrwmd.com/waterways/indian-river-lagoon/*.
- Smith NP. 1993. Tidal and nontidal flushing of Florida's Indian River Lagoon. *Estuaries* 16(4):739–746 DOI 10.2307/1352432.
- **Stuart V, Klumpp DW. 1984.** Evidence for food-resource partitioning by kelp-bed filter feeders. *Marine Ecology Progress Series* **16**:27–37 DOI 10.3354/meps016027.
- Swain HM, Breininger DR, Busby DS, Clark KB, Cook SB, Day RA, De Freese DE, Gilmore RG, Hart AW, Hinkle CR, McArdle DA, Mikkelsen PM, Nelson WG, Zahorcak AJ. 1995. Indian River Lagoon conference—introduction. *Bulletin of Marine Science* 57(1):1–7.
- Whalen MA, Stachowicz JJ. 2017. Suspension feeder diversity enhances community filtration rates in different flow environments. *Marine Ecology Progress Series* 570:1–13 DOI 10.3354/meps12133.
- Wilson C, Scotto L, Scarpa J, Volety A, Laramore S, Haunert D. 2005. Survey of water quality, oyster reproduction and oyster health status in the St. Lucie Estuary. *Journal of Shellfish Research* 24(1):157–165 DOI 10.2983/0730-8000(2005)24[157:sowqor]2.0.co;2.
- **Zanden MJV, Rasmussen JB. 2001.** Variation in δ^{15} N and δ^{13} C trophic fractionation: implications for aquatic food web studies. *Limnology and Oceanography* **46(8)**:2061–2066 DOI 10.4319/lo.2001.46.8.2061.