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Original Paper

Identification and Validation of Two Novel **Prognostic IncRNAs in Kidney Renal Clear Cell Carcinoma**

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Key Words

Long non-coding RNAs • Kidney Renal Clear • Cell Carcinoma • Competitive endogenous RNA

Abstract

Background/Aims: Kidney renal clear cell carcinoma (KIRC) is one of the most fatal malignancies due to late diagnosis and poor treatment. To improve its prognosis, a screening for molecular biomarkers of KIRC is urgently needed. Long non-coding RNAs (IncRNAs) play important roles in tumorigenesis and prognosis of cancers. However, it is not clear whether lncRNAs can be used as molecular biomarkers in predicting the survival of KIRC patients. Methods: In this study, our aim was to identify IncRNAs/mRNAs signatures and their prognostic values in KIRC. The aberrant expression profile of mRNAs and lncRNAs in 529 KIRC tissues and 72 adjacent non-tumor pancreatic tissues were obtained from the Cancer Genome Atlas (TCGA). A weighted gene co-expression network analysis (WGCNA) of two key IncRNAs was constructed. We constructed an aberrant IncRNA-mRNA-miRNA ceRNA network in CESC. In addition, Gene Ontology (GO) and KEGG pathway analysis were performed. *Results:* Using IncRNA/mRNA expression profiling data, the overall analysis revealed that two novel IncRNA signatures (DNM1P35 and MIR155HG) and several mRNAs were found to be significantly correlated with KIRC patient's overall analysis. Based on the target gene of the two IncRNA in co-expression network, the GO and KEGG analysis were also performed. A dysregulated IncRNA-related ceRNA network was also observed. **Conclusion:** These results suggested that the two novel lncRNAs signatures may act as independent prognostic biomarkers for predicting the survival of KIRC patient.

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Song et al.: Screen of Two Prognostic IncRNAs in Kidney Renal Clear Cell Carcinoma

Introduction

Kidney cancer (KC), a genitourinary type of cancer, accounts for 2%-3% of all adult malignancies worldwide [1, 2]. It has one of the highest incidence rates among the urinary system tumors. The morbidity and mortality of KC are increasingly and the most common subtype of KC is kidney renal clear cell carcinoma (KIRC)[3]. In 2017, approximately 63, 990 new cases and 14, 440 deaths associated with kidney cancer were reported in the United States [4]. The low survival rates are due to late diagnosis of KIRC and ineffective therapeutic methods currently available. To improve prognosis and decrease mortality and morbidity due to KIRC, diagnostic biomarkers are required to facilitate early detection and risk stratification of KIRC and to guide decisions related to the choice of proper treatments. However, numerous studies indicated that KIRC is heterogeneous in various aspects including clinicopathological, molecular, and cellular heterogeneity. Therefore, identification of molecular biomarkers or therapeutic targets to conquer KIRC is imperative.

Due to rapid technological advancements in genome-wide sequencing, research of clinical disease and pathological mechanisms in various cancers has made tremendous achievement [5]. The sequencing of the human genome produced remarkable findings that protein-coding genes comprise <3% of human DNA. Yet over 80% of our genome is actively transcribed to a set of RNA transcripts without protein-coding functions [6, 7]. Long noncoding RNAs (lncRNAs) are broadly defined as RNAs over 200 nucleotides (nt) in length, which have many structural features of the mRNAs [8-11]. Accumulating evidence has demonstrated that lncRNAs play important molecular roles in regulating gene expression at the level of transcription and post-transcriptional, and chromatin modification [12-14]. Although small ncRNAs, in particular miRNAs, have been extensively studied for over 20 years and many aspects of their biology have been unraveled, still very little is known about the functions of lncRNAs. In this study, aberrant expression profiles of lncRNAs and mRNAs in 529 KIRC patients and 72 non-tumor samples were acquired from the TCGA database and a panel of lncRNAs and mRNAs were detected. Additionally, aberrant lncRNA related ceRNA network was constructed via WGCNA in KIRC. This study can help clinicians to understand the function of lncRNAs through lncRNA-associated ceRNA network in KIRC and provide new lncRNAs as novel diagnostic biomarkers.

Materials and Methods

Raw data

The mRNA expression profiles and corresponding clinical information of KIRC patients were obtained from TCGA data portal (https://tcga-data.nci.nih.gov/tcga/), which was imputed on IlluminaHiSeq RNA-Seq platform, containing 529 KIRC tissues and 72 adjacent non-tumor pancreatic tissues. The lncRNA expression profiles were identified based on the annotation from the GENCODE project (http://www.gencodegenes. org) [15]. Both mRNA profiles data and clinical characteristics of KIRC are publicly available and in open-access platforms. Therefore, approval by local ethics committee was not needed.

Screening of differentially expressed IncRNAs and mRNAs

The differential expression of lncRNAs (DElncRNAs) and mRNAs (DEmRNAs) between CESC and adjacent tissue were calculated using R/Bioconductor package of edgeR[16]. The differentially expressed genes (DEGs) of the data set with $|\log 2$ fold change| ≥ 1.0 and P-value less than 0.05 were selected for subsequent analysis.

Functional enrichment analysis of DEmRNAs

To better understand the biological effects and pathways of the aberrantly expression DEmRNA, Gene Ontology (GO) Biological Process, Kyoto Encyclopedia of Genes, and Genomes (KEGG) pathway analyses were conducted using the R/Bioconductor package of Clusteprofiler [17]. Functional enrichment analysis was based on the threshold of P-value <0.05.



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Prognostic analysis

A univariate Cox model was employed to impute the relationship between the expression level of each lncRNAs/mRNAs and patient's overall survival (OS). The lncRNAs with P-values less than 0.05 were considered to be statistically significant in univariate Cox analysis. Thereafter, multivariate Cox analysis was employed to evaluate the contribution of lncRNAs/mRNAs as independent prognostic factors of patient survival. These analyses were conducted using the R package of survival and KMsurv.

Weighted co-expression network construction with WGCNA and target prediction

We analyzed the incorporated network using weighted gene co-expression network analysis (WGCNA), which enables the description of the correlation patterns and gene expression profiles [18, 19]. The WGCNA R package was used to evaluate the significance of the two lncRNAs and their module membership. We assessed the weighted co-expression relationship among all dataset subjects in an adjacency matrix using the pairwise Pearson correlation. The appropriate soft threshold power was automatically calculated and generated as described for the standard scale-free network. In the study, the soft threshold was set at β =11 (scale-free R2= 0.85). Following the identification of weighted correlation, characteristics of the network were presented by Cytoscape 3.4.0[20]. We also predicted the target genes of the two lncRNAs via co-expression network.

Functional enrichment analysis of IncRNAs target genes

The target genes of the candidate lncRNAs were predicted via co-expression network using WGCNA. The enrichment analysis of these target genes was determined using the R/Bioconductor package of Clusteprofiler. Functional enrichment analysis was conducted at the threshold of P-value <0.05.

Validation of the differentially expressed IncRNAs with GEO data

To verify the DElncRNAs/DEmRNAs obtained from TCGA database, we screened the mRNA datasets of KIRC on the GEO database. To identify eligible studies, we employed the following search strategies: "Kidney renal clear cell carcinoma" or "KIRC". The lncRNAs expression level was also extracted for further analysis. The differentially expressed genes were also imputed using the R package of Limma.

ONCOMINE and CCLE analysis

The two DElncRNAs in different types of cancers and cell lines were determined through analysis on the ONCOMINE database (www.oncomine.org) and CCLE database (https://portals.broadinstitute.org/ccle/home), which is a publicly accessible online microarray database to facilitate discovery and identification of genome-wide expression analyses.

Prediction of IncRNA-related ceRNA

The target miRNA of lncRNAs were predicted and minimum free energy (MFE) of miRNA-lncRNA duplexes was imputed using the RNAhybrid program. The data of lncRNA-miRNA interactions were downloaded from highly reliable online miRNA reference database of miRcode (http://www.mircode.org/) [21]. The integrated lncRNA-miRNA pairs were predicted using miRcode, in combination with the selected miRNA. The filter threshold of selected miRNA was max energy <= - 20 and score> 160. Then, the prediction of targeted mRNAs of miRNAs was retrieved from databases of starBase [22]. The lncRNA-associated ceRNA was constructed and visualized using Cytoscape 3.3.2[20]

Results

Pre-treated data

The mRNA expression and corresponding clinical information of KIRC patients were obtained from TCGA data portal (https://tcga-data.nci.nih.gov/tcga/), which was based on the IlluminaHiSeq RNA-Seq platform; containing 529 KIRC tissues and 72 adjacent non-tumor pancreatic tissues. The raw counts of KIRC mRNA expression profiles (level 3 data) were downloaded from the TCGA databases. After initial screening, 16, 315 genes were obtained.



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The mRNA expression profiles were transformed lncRNA expression profiles according to the annotation of GENCODE project. Both mRNA profiles data and clinical characteristics of PDAC are publicly available and in open-access platforms. Therefore, approval by a local ethics committee was not needed.

Screening of differentially expressed lncRNAs/mRNAs in KIRC

Analysis of mRNAs/lncRNAs expression profiles in KIRC patients tissues (n=529) and in normal tissues (n=72) identified 4, 544 differentially expressed genes, including differentially expressed mRNAs (n=4497) and lncRNAs (n=47) (Fig. 1). Of these, 3032 genes were over-expressed and 1512 genes were down-expressed. Among the differentially expressed lncRNA, 43 were over-expressed while 4 were down-expressed.

Functional enrichment analysis of DEmRNAs

To explore the functional implication of DEmRNAs, we performed the functional enrichment analysis of GO and KEGG category. The results of GO analysis revealed that GO-enriched categories in the biological process (n=1031), cellular component (n=80), and molecular function (n=117) were among the up-regulated genes. The top five GO-enriched categories are shown in Fig. 2A. A total of 52 KEGG pathways related to biological pathways were enriched among the upregulated genes, including hsa04940; type I diabetes mellitus, hsa05330; allograft rejection, hsa05332: graft-versus-host disease. hsa05150; Staphylococcus aureus infection, and hsa04060; cytokine-cytokine receptor

KARGFR



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Fig. 1. Volcano plot of the differentially expressed genes. Blue color indicates differentially expressed genes with $|\log 2$ fold change| ≥ 1.0 and P-value < 0.05.



Fig. 2. The top five of GO terms and pathways. Above and below showing upregulated and downregulated DEmRNAs, respectively.

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interaction. The results analysis revealed of GO GO-enriched categories in the biological process (n=347), cellular component (n=48), and molecular function (n=76) among the downregulated genes. The top five GO categories are shonw in Fig. 2B. A total of 36 KEGG pathways related to the biological pathways were enriched among the downregulated genes, including hsa00280; valine, leucine, and isoleucine degradation, hsa01200; carbon metabolism, hsa00640; metabolism, propanoate hsa00260; glycine, serine, and threonine metabolism, and hsa00071; fatty acid degradation.

> Prognostic assessment of DElncRNAs/DEmRNAs profiles and clinical characteristics

The univariate Cox regression was performed between DEmRNAs/ **DElncRNAs** profiles and KIRC patients, and the results showed that 932 DEmRNAs and **DElncRNAs** were significantly associated with overall survival (OS) (P<0.05). In addition, multivariate Cox proportional regression was performed, and we found that 2 DElncRNAs and 78 DEmRNAs are independent prognostic indicators of KIRC (Table 1). The high expression of two DElncRNAs (DNM1P35 and MIR155HG) was significantly associated

Table 1. multivariate Cox regression analysis for DEmRNAs/DElncRNAs

| GENE | Z | Pr(> z) | HR | lower .95 CI | upper .95 CI |
|------------------------|---------------|-------------|--------------|---|--------------|
| NTN5.126147 | 12.19952683 | 0 | 3.08197E+12 | 3.24E-13 | 30365823159 |
| DAND5.199699 | 2.305107202 | 0.02116057 | 14290711362 | 7.00E-11 | 33.15233195 |
| CCDC154.645811 | 2.5422388 | 0.011014489 | 3967357315 | 2.52E-10 | 157.921916 |
| SPC24.147841 | 2.005602176 | 0.04489871 | 217454121.8 | 4.60E-09 | 1.547817557 |
| GSG2.83903 | 4.619587573 | 3.85E-06 | 29199840.07 | 3.42E-08 | 19862.08382 |
| KIAA1751.85452 | 2.702769962 | 0.006876432 | 3538398.747 | 2.83E-07 | 63.06923325 |
| KIR3DL2.3812 | 2.654365794 | 0.007945764 | 122971.9579 | 8.13E-06 | 21.45512661 |
| KIF14.9928 | 3.31932974 | 0.000902338 | 14416.54726 | 6.94E-05 | 50.48696484 |
| C19orf35.374872 | 7.194473204 | 6.27E-13 | 11387.39047 | 8.78E-05 | 894.0203852 |
| ACRC.93953 | 2.018081802 | 0.043582743 | 393.4687612 | 0.002541498 | 1.187762575 |
| RFX8.731220 | 4.462430509 | 8.10E-06 | 211.6676026 | 0.004724389 | 20.14621038 |
| DNM1P35.100128285 | 6.88901596 | 5.62E-12 | 184.7264857 | 0.005413409 | 41.84935869 |
| SLC46A2.57864 | 2.416639361 | 0.015664529 | 134.4850386 | 0.007435771 | 2.524981468 |
| KIF19.124602 | 2.360379384 | 0.018256254 | 123.3742125 | 0.008105422 | 2.263371662 |
| NEK2.4751 | 2.916382649 | 0.003541159 | 89.79681353 | 0.011136253 | 4.370849735 |
| BIRC5.332 | 2.450085553 | 0.014282228 | 76.26165933 | 0.013112749 | 2.379805703 |
| C11orf35.256329 | 2.287471633 | 0.022168309 | 75.6248348 | 0.013223169 | 1.857705537 |
| ATAD5.79915 | 7.665621903 | 1.78E-14 | 71.17200222 | 0.014050469 | 23.91698747 |
| CENPI.2491 | 2.112468878 | 0.034646253 | 59.08051924 | 0.016926053 | 1.34241068 |
| AGER.177 | 9.743421932 | 0 | 58.85309657 | 0.016991459 | 25.92785913 |
| BRIP1.83990 | 8.413027803 | 0 | 45.60954225 | 0.021925236 | 18.7304957 |
| LILRA2.11027 | 6.661521776 | 2.71E-11 | 43.3130434 | 0.023087733 | 14.29190209 |
| PRSS36.146547 | 5.997315922 | 2.01E-09 | 39.91612632 | 0.025052531 | 11.96415722 |
| CTLA4.1493 | 2.164790164 | 0.030403758 | 37.12412616 | 0.026936661 | 1.407720159 |
| C17orf56.146705 | 2.053378029 | 0.040035928 | 34.9844756 | 0.028584107 | 1.175534312 |
| PBK.55872 | 2.552003603 | 0.010710541 | 32.34339146 | 0.030918217 | 2.240037714 |
| C7orf53.286006 | 5.204250003 | 1.95E-07 | 28.81653836 | 0.034702294 | 8.126967483 |
| YPEL4.219539 | 5.240353704 | 1.60E-07 | 25.40633521 | 0.039360262 | 7.576602174 |
| DNHD1.144132 | 4.950855672 | 7.39E-07 | 24.67361368 | 0.040529126 | 6.935379726 |
| CDC6.990 | 5.844255669 | 5.09E-09 | 16.04076471 | 0.062341168 | 6.324609189 |
| TRIM46.80128 | 2.877116629 | 0.004013272 | 12.59502614 | 0.079396421 | 2.242414431 |
| KIFC1.3833 | 5.422624466 | 5.87E-08 | 12.39903469 | 0.08065144 | 4.991046599 |
| CHEK2.11200 | 2.420179979 | 0.015512827 | 11.75166498 | 0.085094325 | 1.597672283 |
| KLRA1.10748 | 3.170680295 | 0.001520824 | 11.21686761 | 0.089151449 | 2.517044144 |
| FAM72B.653820 | 2.275818345 | 0.022856884 | 9.933711631 | 0.100667307 | 1.375264644 |
| E2F1.1869 | 3.818919341 | 0.000134038 | 8.855395077 | 0.112925509 | 2.89119489 |
| MIR155HG.114614 | 1.966408188 | 0.049251478 | 5.158421696 | 0.193857745 | 1.005391063 |
| PRC1.9055 | 5.157125624 | 2.51E-07 | 4.754986139 | 0.210305555 | 2.629050281 |
| TBC1D3B.414059 | 6.07734797 | 1.22E-09 | 4.271648978 | 0.234101633 | 2.674400786 |
| CSAD.51380 | 7.014674294 | 2.30E-12 | 3.755325539 | 0.266288499 | 2.594691996 |
| LOC100129637.100129637 | 3.379853768 | 0.000725244 | 3.018593475 | 0.331280117 | 1.590624988 |
| CACNB1.782 | 3.3862544 | 0.000708537 | 3.000687683 | 0.333256942 | 1.588566529 |
| TAZ.6901 | 7.407599 | 1.29E-13 | 2.978921007 | 0.335692017 | 2.231662564 |
| NBPF9.400818 | 9.273790424 | 0 | 2.713152825 | 0.368574889 | 2.197161021 |
| SPHK1.8877 | 2.136610756 | 0.032629664 | 2.603337852 | 0.384122253 | 1.082316972 |
| SLC17A9.63910 | 6.859553794 | 6.91E-12 | 2.303251316 | 0.434168861 | 1.814723246 |
| LUC150776.150776 | 3.1449/480/ | 0.001661011 | 2.245102622 | 0.445413938 | 1.356263417 |
| CN1NAP1.8506 | 4.037768108 | 5.40E-05 | 2.218508274 | 0.450753334 | 1.506885769 |
| L190FI48.84/98 | 2.588200482 | 1 205 00 | 1.900039563 | 0.52030483 | 1.10859584 |
| AT 102.0900 | 2 142676420 | 1.37E-08 | 1.771206472 | 0.302481023 | 1.43/42/840 |
| ZNE602 55657 | 2 167070205 | 0.001074108 | 1.721200472 | 0.30090/022 | 1.220/41902 |
| CDM 6722 | 0.005690119 | 0.030229495 | 1 60065700 | 0.20024014/ | 1.032310056 |
| OTRT1 81890 | 2 062210245 | 0 039197724 | 1 6404 77605 | 0.0000000000000000000000000000000000000 | 1.024945560 |
| PPEIA4 8497 | 4 701272022 | 2 595 06 | 1.618972261 | 0.617712542 | 1 324316794 |
| IIRXN11 91544 | 3 21 37 28910 | 0.001310232 | 1.618168052 | 0.617982785 | 1.324310704 |
| PLAUR 5329 | 9 283725821 | 0.001310233 | 1 614429847 | 0.619413722 | 1 459158712 |
| ASAP1.50807 | 3.759319685 | 0.000170376 | 1.602477364 | 0.624033776 | 1.253202696 |

with overall survival (P<0.05) and exhibited positive effects (See Fig. 3, Table 2). To further examine the association between clinical features and prognosis of KIRC patients, we conducted univariate and multivariate Cox proportional regression. The results are shown in Table 3. In univariate Cox analysis, STAGE, Primary Tumor-Lymph Nodes-Metastasis (TNM), Metastasis (M), and TNM, Tumor (T) were significantly associated with OS of KIRC patients. However, only TNM and M were significantly associated with OS of KIRC patients in the multivariate Cox analysis.





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Fig. 3. Kaplan-Meier survival curves for overall survival outcomes according to the risk cutoff point for two lncRNAs. The p value of the log-rank test was less than 0.01.

Table 2. Two lncRNAs were significantly correlated with overall survival in multivariate Cox regression analysis

| GENE | Z | Pr(> z) | HR | lower .95 CI | upper .95 CI |
|----------|-------------|-------------|-------------|--------------|--------------|
| DNM1P35 | 6.88901596 | 5.62E-12 | 184.7264857 | 41.84935869 | 815.3977887 |
| MIR155HG | 1.966408188 | 0.049251478 | 5.158421696 | 1.005391063 | 26.46663112 |

| Clinical factures | Univariate ana | lysis | Multivariate analysis | | |
|-------------------|-----------------------|-------------|-----------------------|-----------------------|--|
| Cillical leatures | P-value | HR(95%CI) | P-value | HR(95%CI) | |
| GENDER | 1.0578(0.8827-1.2676) | 0.541811087 | | | |
| AGE | 0.9999(0.9927-1.0071) | 0.976078039 | | | |
| STAGE | 1.1269(1.0457-1.2144) | 0.015231324 | 0.172389844 | 0.8770(0.7264-1.0589) | |
| TNM_M | 1.6316(1.3782-1.9315) | 1.04E-07 | 2.56E-07 | 1.6871(1.3828-2.0584) | |
| TNM_N | 1.0160(0.9300-1.1100) | 0.118369952 | | | |
| TNM_T | 1.0461(1.0143-1.0788) | 0.007735265 | 0.084015258 | 1.0674(0.9913-1.1493) | |
| GRADE | 1.1142(0.9890-1.2553) | 0.078317061 | | | |

Target prediction and functional enrichment of the two lncRNAs

The biological functions of lncRNAs were still unknown. Therefore, to explore the target genes of the two key lncRNAs, we employed the Weighted Co-expression network construction to examine. A total of 11 modules were detected in the co-expression network. The lncRNA DNM1P35 and MIR155HG were included in the turquoise module and yellow module, respectively. The number of genes in the turquoise module was 466, of which 113 genes were identified to be potentially regulated by the lncRNA DNM1P35 in the co-expression network (see Fig. 4). Among these target genes in co-expression network, ACRC, AGER, AP1G2, C17orf56, CCDC14, CSAD, DNHD1, ENGASE, GOLGA8B, LOC100129637, LOC150776, LOC339047, LPIN3, UBXN11, ZNF692, and DNM1P35 were up-regulated in KIRC and hence may act as independent prognostic factors for the OS of KIRC patients. The number of genes in the yellow module was 221, of which 107 genes were identified to be potentially regulated by the lncRNA MIR155HG in the co-expression network (see Fig. 4). Among these target genes were identified to be potentially regulated by the lncRNA MIR155HG in the co-expression network (see Fig. 4). Among these target genes in the yellow module was 221, of which 107 genes were identified to be potentially regulated by the lncRNA MIR155HG in the co-expression network (see Fig. 4). Among these target genes in the co-expression network (see Fig. 4). Among these target genes in the co-expression network (see Fig. 4). Among these target genes in the co-expression network (see Fig. 4). Among these target genes in the co-expression network (see Fig. 4).



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Fig. 4. The target genes of two key lncRNAs in the coexpression network (the crimson in the Fig. is the prognostic risk gene, the diamond represents lncRNAs, and the red represents the up-regulation).

AP1G2, C17orf56, CCDC14, CSAD, DNHD1, ENGASE, GOLGA8B, LOC100129637, LOC150776, LOC339047, LPIN3, UBXN11, and ZNF692 were up-regulated in KIRC and hence may act as independent prognostic factors for the OS of KIRC patients.

The enrichment analysis was conducted to describe the biological function of the target genes



Fig. 5. Comparison of GO and KEGG enrichment results for target genes of two lncRNAs.

of two key lncRNAs. We found enrichment of 334 gene ontology categories. There are 238 GO terms with the biological process (BP), 26 GO terms with cellular component, and 70 GO categories with molecular function. A total of 14 KEGG pathways were enriched by target genes of two key lncRNAs. Comparison of the enrichment analysis of the two lncRNAs demonstrated that their target gene MIR155HG was enriched in T cell receptor signaling pathway (See Fig. 5).

Validation of DEmRNAs/DElncRNAs with GEO data

One study (GSE53757) was considered eligible for GEO. Analysis of mRNA expression profiles identified a total of 3135 differentially expressed mRNA. Among these genes, 1628 DEmRNAs were over-expressed and 1507 DEmRNAs were down-expressed. In both TCGA KIRC and GSE53757 studies, 1127 genes were over-expressed while 736 genes were down-expressed. Among significant prognostic risk of 78 mRNAs and 2 lncRNAs, 51 genes which FDR was less than 0.01 were up-regulated. As for expression profiles of the two lncRNAs, only lncRNA DNM1P35 was detected in the GSE53757. However, the differential expression profiles of lncRNA DNM1P35 were not statistically significant in tumor and normal group. lncRNA MIR155HG was not detected in GSE53757.

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Expression level of two IncRNAs in Oncomine and CCLE database Two IncRNAs

(DNM1P35 and MIR155HG) were found in human solid tumors (Fig. 6). Only two studies reported the of expression IncRNA **DNM1P35** breast in cancer [23] and kidney cancer of TCGA cohort. The study on TCGA kidney cancer suggested that the expression of lncRNA DNM1P35 was significantly higher in clear cell renal cell carcinoma than other types of renal cell carcinoma as shown in Fig. 7. In Oncomine database, several studies reported the expression level of lncRNAs MIR155HG in clear cell renal cell carcinoma samples [24, 25]. The pooled results revealed that lncRNAs MIR155HG was significantly higher in KIRC compared to other cancer samples (cancer vs. cancer, P<0.05) (Fig. 8). In addition, the CCLE database analysis demonstrated that lncRNA expression the level of DNM1P35 and MIR155HG was high in many cancer cell-lines (Fig. 9).

Fig. 8. The combined result of lncRNA MIR155HG expression across three studies in Oncomine database.







Fig. 7. The expression level of lncRNA DNM1P35 was significantly higher in Clear Cell Renal Cell Carcinoma compared to other types of Renal Cell Carcinoma.



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Fig. 9. The two lncRNAs (DNM1P35 and MIR155HG) were highly expressed in kidney cancer cell lines as revealed from CCLE analysis.

Construction and analysis of IncRNA-related ceRNA network

A total of 295 microRNAs that bind to DNM1P35 were identified at the set threshold, and 115 microRNAs which are associated with MIR155HG were screened. The miRNA-lncRNA network is illustrated in Fig. 10. The starBase database was used to explore the downstream target genes of microRNAs that bind to lncRNA, and those which are prognostic genes for KIRC. A ceRNA network was constructed based on a combination of microRNAs, lncRNAs, and target genes. The network contains 62 nodes and 135 edges, in which 32 miRNAs, 2 lncRNAs, and 28 target genes were detected (Fig. 11). The combination of MIR135HG with hsa-miR-155-5p and hsa-miR-223-3p is also mentioned in the starBase database.

Discussion

Kidney Renal Clear Cell Carcinoma (KIRC) is an aggressive and malignant type of kidney cancer which has poor prognosis and is characterized by a complex molecular and cellular heterogeneity [26]. Although KIRC is curable at early-stage by surgery resection, the prognosis of metastatic KIRC is poor. Researchers have demonstrated that the 5-year survival rate of KIRC patients is less than 10%[27]. In the past, great efforts have been made to provide insights into the molecular mechanisms underlying KIRC, but the focus has been on protein-coding genes or miRNA[28, 29]. Therefore, understanding the molecular mechanism of KIRC may provide clinicians with new treatment strategies for this disease. To date, there are no conventional molecular biomarkers for KIRC prognosis. In biological processes, lncRNAs acts through miscellaneous mechanisms [30, 31]. It has been documented that lncRNAs are a novel class of non-protein-coding transcripts found in cancer biogenesis and prognosis. Aberrant lncRNAs expression has been reported in many cancers, suggesting that they may function as oncogenes or tumor suppressors [32-34]. Integration genomic studies have demonstrated that lncRNAs are thought to improve the clinical outcome of







Fig. 10. The lncRNA-miRNA based ceRNA network (triangle represents miRNAs, and diamond represents lncRNAs).

Fig. 11. The integrated ceRNA network (squares are target genes, triangles are miRNAs, diamonds are lncRNAs, blue lines indicates that microRNAs regulate target genes, dark yellow lines represent the reaction between miRNAs and lncRNAs).





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KIRC patients [35, 36]. However, no specific biomarker has been found to display therapeutic efficiency for KIRC, and therefore there is need for specific prognostic factors to improve the treatment of KIRC patients.

In the present work, we compared gene expression profiles between KIRC samples and adjacent non-tumor tissues to identify potential lncRNA molecular biomarkers using a database of TCGA. The differentially expressed mRNAs/lncRNAs were identified after which the univariate and multivariate Cox analysis was performed to determine whether these mRNAs/lncRNAs can be used for prediction of KIRC prognosis. We found two key lncRNAs and several mRNAs as key predictors of KIRC prognosis. Among the lncRNAs, overexpression of lncRNAs (DNM1P35 and MIR155HG) was associated with poor prognosis of patients with KIRC. We further used the GEO data to confirm these findings. Indeed, the results revealed that DEmRNAs/lncRNAs can be independent prognostic predictors for patients with KIRC. Through a Weighted Co-expression network using WGCNA, we found that two lncRNAs were correlated with 230 protein-coding genes. In addition, a lncRNA-miRNA-mRNA ceRNA network was constructed based on the starBase and miRanda database. Using the enrichment and functional analysis using the R/Bioconductor package of Clusteprofiler, we found that MIR155HG and it's target gene Cytotoxic T lymphocyte antigen 4 (CTLA-4) were up-regulated in KIRC and both were significantly associated with OS in patients with KIRC. CTLA-4, a transmembrane protein encoded by the CTLA-4 gene, is expressed in activated CD4 + and CD8 + T cells. CTLA-4 binds to its ligand B7 molecule to produce inhibitory signals that prevent activation of T cell and protect tumor cells from T lymphocytes. Thus, blocking the immune effect of CTLA-4 can stimulate cell proliferation of many immunity cells, thereby inducing or enhancing the anti-tumor immune response [37-39]. The target gene of DNM1P35 did not show enrichment function. However, the expression of DNM1P35 and its target genes: ACRC, AGER, AP1G2, C17orf56, CCDC14, CSAD, DNHD1, ENGASE, GOLGA8B, LOC100129637, LOC150776, LOC339047, LPIN3, UBXN11, and ZNF692 were significantly increased in KIRC and are prognostic risk genes for this disease.

Several studies have reported that lncRNA MIR155HG plays key roles in the development of myeloproliferative disorders, leukemia, lymphoma, and glioma [40-42]. lncRNA MIR155HG can suppress lymphoma by regulating the activity of NF- κ B and mesenchymal genes [42]. However, the contribution of lncRNA DNM1P35 to the development of KIRC is still not known, and further studies are required to address these aspects.

The expression of two incRNAs (MIR155HG and DNM1P35) and target genes of CTLA-4, ACRC, AGER, AP1G2, C17orf56, CCDC14, CSAD, DNHD1, ENGASE, GOLGA8B, LOC100129637, LOC150776, LOC339047, LPIN3, UBXN11, and ZNF692 was significantly increased in KIRC and thus they are prognostic risk genes for this disease. Among these key mRNAs, diseases associated with AGER include Diabetic Angiopathy and Thymic Hyperplasia. Among its related pathways are activated TLR4 signaling and a-beta signaling pathways. GO annotations related to this gene include identical protein binding and transmembrane signaling receptor activity [43, 44].

AP1G2 has been reported to be involved in cardiac arrest and long Qt syndrome 1. Among its related pathways are CTLA-4 signaling and clathrin derived vesicle budding. GO annotations related to this gene include binding and protein transporter activity [45, 46].

CSAD was associated with Disuse Amblyopia. Among its related pathways are β -alanine metabolism (TR) and sulfur amino acid metabolism. GO annotations related to this gene include pyridoxal phosphate binding and carboxy-lyase activity [47].

Diseases associated with ENGASE include congenital isorder of deglycosylation. Among its related pathways is transport to the golgi and subsequent modification and metabolism of proteins. GO annotations related to this gene include mannosyl-glycoprotein endo-beta-N-acetylglucosaminidase activity [48].

LPIN3 (Lipin 3) is a protein-coding gene. Among its related pathways are mitotic prophase and synthesis of PC. GO annotations related to this gene include transcription coactivator activity and phosphatidate phosphatase activity [49].

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Several limitations of the study should be considered. Firstly, the two lncRNAs signature were analyzed and validated only in the TCGA and GEO data set, and no other KIRC-related lncRNAs expression profile was used for further validation. Secondly, the TCGA data was obtained from a single central source, and the ethnic composition of the population in TCGA database was mainly white and black, hence the findings in the work cannot be extrapolated to other ethnicities. Thirdly, no experimental data on the mechanisms of lncRNA have been reported, and further experimental studies are needed to improve our understanding of the functional role of the lncRNA in KIRC.

Findings in this study demonstrated that two key lncRNAs and several mRNAs can be used as molecular biomarkers and prognostic factors to predict the survival rate in patients with KIRC. However, the biological function of the two lncRNAs needs to be further validated through experiments.

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Disclosure Statement

The authors declare to have no competing financial interests.

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