

Journal of Chemical Health Risks

www.jchr.org



### **ORIGINAL ARTICLE**

## Fabrication of Mesoporous Silica-coated Graphene Oxide by New Method and Application in Solid Phase Extraction for **Preconcentration of Cu<sup>2+</sup>**

Ali Moghimi

Silica;

SPE;

FAAS:

Copper

Department of Chemistry, Varamin (Pishva) Branch, Islamic Azad University, Varamin, Iran

(Received: 25 February 2018 Accepted: 11 May 2018)

ABSTRACT: Graphene oxide is a derivates of graphene that has a ultrahigh specific surface space with variety of chemically reactive functionalities, such as epoxy and hydroxyl groups on the basal **KEYWORDS** plane and carboxylic acid groups along the area edge, which can be use different groups for Graphene oxide; functionalization, consequently it has a great promise for use as sorbent materials. On the other hand silica is well-known one of the best sorbent for adsorption that we use it as substance for coated on the graphene oxide for produce an ultra-sensitive sorbent. A new technique using a solid phase extraction Preconcentration; (SPE) cartridge with graphene oxide functionalized by silica as sorbent was developed for the preconcentration of trace amounts of copper and was determined by flame atomic absorption spectrometry (FAAS). Some of the important parameters were selected and optimized. Under the optimized conditions the limit of detection (LOD) and limit of quantification (LOQ) were 0.175, 0.585 and the proposed method has a good reproducibility 0.85% (RSD %). The enrichment factor was 200 and the percentage of recovery was in the range of 97-100%. The method was successfully applied to the recovery of  $Cu^{2+}$  in different type of water samples. Graphene oxide and its derivates such as GO@SiO2 in this study are full of potential to use as an excellent adsorbent in the extraction method like solid phase extraction (SPE).

#### **INTRODUCTION**

Though copper (II) ions are really important for several biological processes, its levels in some states are representation of a problem or illness in many living systems or environments. although trace copper ions promote rancidity and off-flavors in nutrients, copper accumulation in the liver is a feature of Wilson's illness, producing neurological and psychiatric defects .Hence copper is both vital and toxic for many biological systems [1]. Consequently, determination of trace amounts of copper has received considerable attention in the battle against environmental pollution [2].

FAAS (flame atomic absorption spectroscopy) with its relative low cost and decent analytical performance is the significant instrument in the research laboratories for determination of a range of heavy metals. Accurate determination of trace heavy metals by FAAS is usually one of the important problems for the analytical chemist because of their low concentrations. Also other important

problem in FAAS determinations of heavy metals are the effects of the matrix of the analyzed samples. In order hand, these problem analytical chemists mostly use separation and preconcentration methods such as coprecipitation, liquid–liquid extraction (LLE), cloud point extraction (CPE), electrodeposition, solid phase extraction (SPE), solid phase micro extraction (SPME), etc. [3, 4 and 5].

In recent years, SPE method have been well used for determination of copper ions in numerous environmental samples [6,7] because of its simplicity, rapidity, negligible cost, low consumption of reagents and its ability to combine with different detection techniques either in online or off-line mode [8]. The main part of the SPE is the sorbent material that determines the selectivity and sensitivity of the method. Though, the commonly used SPE sorbents, such as C<sub>18</sub>, silica and graphitic carbon, are often the only suitable option for a limited number of analytes. Reusability of the SPE column is also a problem. Thus, developing new SPE sorbent material is important [9]. Carbon derivates are well-known for their high adsorption capacity. They have been established to possess excessive potential as adsorbents for removing many types of environmental pollutants such as heavy metals [10, 11], Carbon nanotubes (CNTs)[12,13] ,Fullerenes can be used as chromatographic stationary phases to offer high selectivity for specific compounds[14,15] or as sorbent materials for on-line clear up and preconcentration[16,17]. Recently, CNTs have been shown to be excellent classes of sorbent materials for SPE [18]. Since the first application of CNTs in SPE by Cai et al that he use multi-walled carbon nanotubes (MWCNTs) [18], In recent years many reports have been distributed focusing on progress of use CNTsbased SPE methods for a great variety of analytes, including phenolic compounds [19], insecticides [20], pharmaceuticals [21, 22], inorganic ions [23].

organometallic compounds [24].

Graphene oxide is a novel two-dimensional nanomaterial or in fact can be mention graphene oxide is graphene but with more chemically reactive functionalities [25, 26] which can be more functionalized [27, 28]. Graphene oxide surface has a major amount of the sp<sup>2</sup>-hybridized carbon

backbone structure remains intact [29], allowing the nano plate to retain a high degree of planarity, as proven in thinfilm deposition and fabrication experiments [30, 31], and a consistently high surface to volume ratio [32]. The extraordinary properties of graphene and its derivates like graphene oxide make it a greater candidate as an adsorbent for SPE technique. Firstly, graphene has a large surface area (theoretical value 2630 m<sup>2</sup> g<sup>-1</sup> [33]), suggesting a high sorption capacity. Precisely, both sides of the planar sheets of graphene are accessible for molecule adsorption, while for CNTs and fullerenes, steric hindrance may exist when molecules access their inner walls. Secondly, graphene can be simply modified with functional groups, particularly via graphene oxide which has many reactive groups [34]. Functionalization may more enhance the selectivity and sensitivity of SPE. Thirdly, CNTs usually contain trace amounts of metallic impurities that originate their synthesis and using the metal catalysts. These impurities could have negative effects on the applications of CNTs [35, 36]. Nonetheless, graphene and graphene oxide instead, can be synthesized from graphite without the using metal catalysts. In fact, graphene is a layer of graphite, which does not use of any metal catalysts in its synthesis and exfoliation process. Accordingly, the pure material would be obtained [37].

Actually, the SPE technique using graphene and derivates of graphene such as functionalization graphene oxide as a novel and great adsorbent [38]. Chemical modification of graphene oxide has been a promising way to achieve mass production of chemically modified graphene (CMG) platelets. Graphene oxide contains a range of reactive oxygen functional groups, which renders it a good candidate to use in the above-mentioned applications.

In this study the aim was to develop fast and efficient method for selective extraction, concentration and determination of copper with a novel SPE method using graphene oxide functionalized and reduced by mesoporous silica via direct reaction as adsorbent in a SPE column combined with FAAS .The silica used for SPE extraction columns is essentially porous and noncrystal-line, with the general formula  $SiO_2 \times H_2O$ . The silanol groups are the polar groups through which other organic moieties are bonded which we used it on the surface of graphene oxide for the higher ability of graphene oxide in adsorption Cu ( $\Pi$ ) ions.

#### MATERIALS AND METHODS

#### Apparatus

The concentration of the Cu (II) ions solutions was determined by using the Varian model spectra AA-240 (Mulgrara, Victoria, Australia). The pH-measurements of the metal ion and buffer solutions were carried out by an Orion 420. The size and morphology of graphene oxide was observed by scanning electron microscopy (SEM model LEO 440i), before taken image with SEM the surface of sample was coated with gold. X-ray diffraction (XRD) measurements were carried out with Cu K $\alpha$ radiation (40 kV, 60 mA, 2 $\Theta$  from 5 -115). Fourier transform-infrared (FT-IR) spectra were taken in KBr pressed pellets on a FT-IR Thermo Nicolet (USA).

#### Materials and reagents

All the materials and reagents were of analytical grade and were purchased Merck, Aldrich and Sigma Company. All the dilutions were prepared by deionized water.

#### Preparation of Graphene Oxide Nano sheets

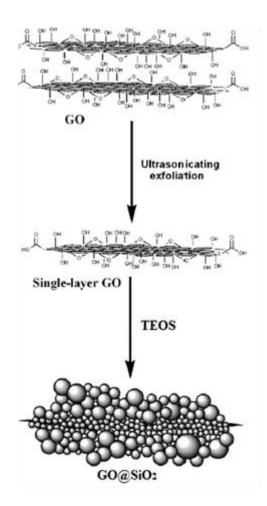
Graphene oxide was prepared by acid oxidation of graphite powder according to the modified Hummers's method. Concisely, 2 g of 300 mesh graphite powder and 5 g of sodium nitrate were added to 50 mL of sulfuric acid (98%). The mixture was then cooled down to 0°C. Afterward, 7 g of KMnO<sub>4</sub> was carefully added to the reaction bottle with stirring at a temperature below 20°C; the temperature was then increased up to 35°C and kept stirring for 1 h. After that, 150 mL of water was added, and the mixture was stirred at 90°C for 15 min.

Then, an additional 100 mL of water was added into the reaction container, followed by the addition of 7 mL of 30%  $H_2O_2$ . The reaction mixture was filtered through a PVDF filter membrane with a pore size of 0.22 µm, washed one time with 5% HCl for remove all of metals on the surface of graphene oxide and washed repeatedly with DI water until the pH of mixture reached pH=7. Finally, the resultant dispersion in water was ultrasonicated for 1 h, followed by centrifugation at 7000 rpm for 30 min and dry in oven 60 °C to produce the graphene oxide.

#### Synthesis of mesoporous SiO<sub>2</sub> on graphene oxide

A simple and feasible synthesis method for silica coating of GO was modified according to the literature [1]. As shown in Scheme 1, in a typical synthesis, 100 mg GO and (0.1–3g) CTAB were dispersed into 30 mL deionized water, and the mixture was sonicated for one hour. Then, the above mixture was added to 80 mL

anhydrous ethanol and further sonicated for 0.5 h to form a stable dispersion. Instantly, 2 mL NH<sub>3</sub>.H<sub>2</sub>O was added into the as-prepared GO dispersion.



Scheme 1. Schematic illustration of synthesis for preparing GO@SiO<sub>2</sub>

Subsequently, a TEOS solution (1 mL TEOS in 40 mL ethanol) was dropped in under mechanical stirring, and the reaction mixture was stirred for another 12 h without sonication. Finally, the GO solution was centrifuged and washed with ethanol. The process results in the formation of a uniform and thick layer of silica on every individual GO. The surfactants were removed by a fast and efficient ion exchange method (1.2 g of  $NH_4NO_3$  in 60 mL of ethanol) to obtain  $SiO_2$ -coated GO with mesoporous structure.

#### Characterization

Figure 1a XRD was used to investigate the phase and structure of the synthesized graphene oxide, As expected, GO shows a broad band over low diffraction angles ( $2\theta = 11.2^{\circ}$ ) and suggested that the GO layers were decorated with oxygen-containing groups of carboxyl, hydroxyl and epoxide and that the strong  $\pi$ - $\pi$  interaction of graphite planes .Figure 1b affirmed that SiO<sub>2</sub> was amorphous, Figure 1c showed similar pattern with amorphous in Figure 1b and did not exhibited any peaks of graphite or GO, which confirmed that SiO<sub>2</sub> particles prevented the restacking of the most graphene sheets [2].

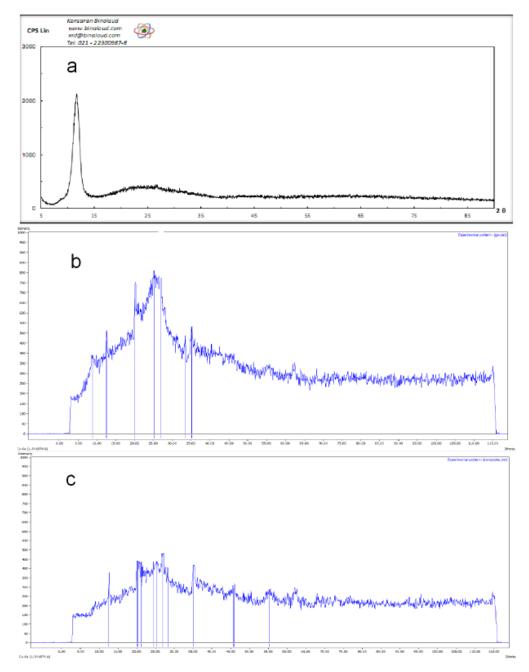


Figure 1. XRD patterns of (a) GO, (b) SiO<sub>2</sub> and (c) GO@SiO<sub>2</sub>.

The FESEM image of Go shown in Figure 2(a) that in image we can see diameter of layers are 54.39 nm and 4

6.88nm. In Figure 2(b) we see SEM of  $GO@SiO_2$  that its clearly displayed nanoparticle of  $SiO_2$  coated on GO.

A. Moghimi / Journal of Chemical Health Risks 8(3) (2018) 223-238

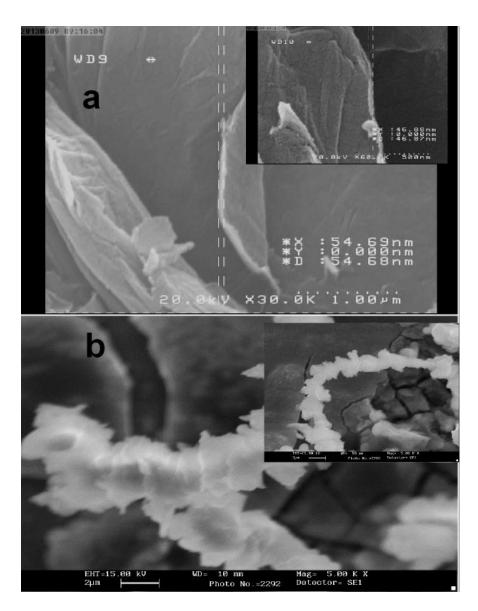


Figure 2. (a) FESEM of GO, (b) SEM of GO@SiO<sub>2</sub>

#### Column preparation and suggested procedures

The adsorption of copper by the adsorbents was studied by the SPE technique. The SPE mode adsorption was selected because of its simplicity, fast and economic for the preconcentration and determination of trace amounts of cooper in diverse samples. GO@SiO<sub>2</sub> (0.1 g) was placed in a 2mL SPE column using an upper frit and a lower frit to avoid adsorbent loss. Prior to extraction, the column was conditioned with 10mL of MeOH and 10mL of deionized water, respectively. The sample solution (50mL) was passed through the column at a flow rate of 2mL/min. Then, the column was washed with 5mL of 10% (v/v) MeOH aqueous solution to remove the co-adsorbed matrix materials from the column. The analytes retained on the column were eluted with 5mL of 4M HNO<sub>3</sub> aqueous solution that flow rate of eluent was 2ml/min.

Finally, the analyte ions in the eluent were determined by FAAS at 324.7 nm for  $Cu^{2+}$ . The effects of several parameters, such effect of pH, eluent type and its volume, effect of flow rates of sample and eluent solution and amount of adsorbent, effect of interfering ions and breakthrough volume were also studied. The results of these studies were used to obtain the optimum conditions

for adsorption capacity measurements. Using the procedure described above, the percent of recovery was calculated from the following equation) Eq.(1)):

$$R\% = \frac{(Cs.Vs) \times (Vw / Vs)}{Vw.Cw}$$
(1)

Where Cs is analyte concentration in eluent (round by FAAS) (mg/L), and Cw is analyte concentration in sample solution (known) (mg/L), Vs is volume of eluent (mL) and Vw is volume of sample solution (mL).

#### Sampling

The first Tap water sample were collected from a water tap in our lab (Tehran shargh (Ghiamdasht), Islamic Azad University, Tehran, Iran), and the second tab water sample were collected from a water tap in our location live (Pakdasht, Tehran, Iran) and the bottle of mineral water purchase from grocery store that the company name was Damavand.

Tap water samples used for development of the method were collected in glasses containers. Before the analysis, the organic content of the water samples was oxidized in the presence of 7%  $H_2O_2$  and then concentrated nitric acid was added. These water samples were then filtered through a 0.45 $\mu$ m Millipore cellulose membrane to remove suspended particulate matter and stored in a refrigerator at 4°C in the dark before analysis.

#### **RESULTS AND DISCUSSION**

# Characterization of GO and $GO@SiO_2$ packed SPE cartridges

#### IR spectrum of GO and GO@SiO<sub>2</sub>

The FT-IR spectra of the GO and GO@SiO2 shown in Figure 3. In Figure 3a shows the FT-IR patterns of graphene oxide. The characteristic features were the absorption bands corresponding to O-H stretching at 3434.33cm<sup>-1</sup>, the peaks at 2922.01 and 2850.93cm<sup>-1</sup> were designated as the asymmetric stretching and symmetric vibrations of CH<sub>2</sub>, C=O stretching vibration of carboxyl appearance of a strong band at 1721.99cm<sup>-1</sup>, the C-OH stretching at 1216cm<sup>-1</sup>, O-H deformation at 1411cm<sup>-1</sup> and C-O vibration at 1025.79cm<sup>-1</sup>, the peak observed at 916cm<sup>-1</sup> might be assigned to the C-H out of plane vibration. In Figure 3b we have pattern of GO@SiO<sub>2</sub> that shows us a new weak band at 1637 cm<sup>-1</sup> which may be contributed from the skeletal vibrations of the reduced graphene lattice [7, 8] or water. The band at  $1077.23 \text{ cm}^{-1}$ was assigned to the stretching vibrations of Si-O-Si bond [9]. In the meantime, it was clear to see that there was no characteristic bands for oxygen containing groups between 1300 and 1600 cm<sup>-1</sup> and that the band at 1637.47 cm<sup>-1</sup> (C=O) disappeared, which conformed that the oxygen containing groups were removed to a high extent based on this proposed method.

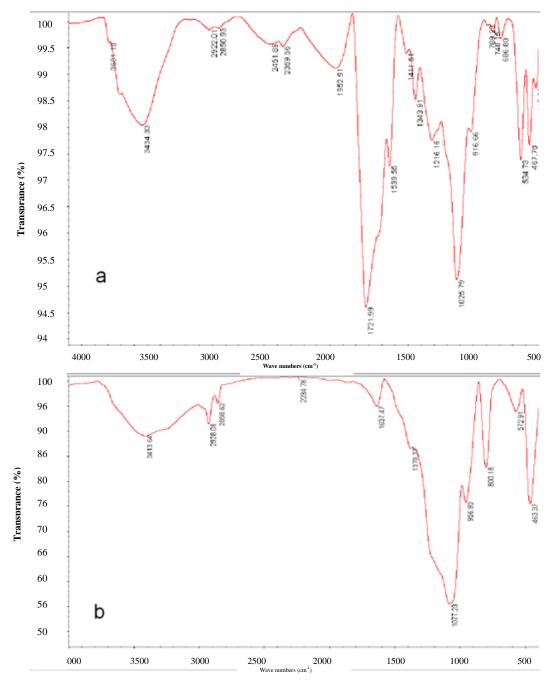


Figure 3. FTIR spectra of (a) GO, (b) GO@SiO<sub>2</sub>.

#### Raman spectrum of GO and GO@SiO<sub>2</sub>

Raman spectroscopy is a great technique non-destructive to study graphene and its derivates GO, especially for investigating the ordered or disordered crystal structures and assessing defects of graphene or GO. The usual features of carbon materials in Raman spectra are the G band at 1580 cm<sup>-1</sup> deriving from the  $E_{2g}$  phonon of C sp<sup>2</sup> atoms and D band at 1350 cm<sup>-1</sup> considered as a breathing mode of  $\kappa$ -point photos of A<sub>1g</sub> symmetry which is assigned to local defects and disorder mostly at the edges of graphene and GO sheets [3, 4 and 5]. As shown in Figure 4a, the Raman spectrum for GO presents a broadened G band at 1584 cm<sup>-1</sup> and D band at 1304 cm<sup>-1</sup>, Figure 2b, the Raman spectrum for GO@SiO<sub>2</sub> presents G band at 1590 cm<sup>-1</sup> and D band at 1360 cm<sup>-1</sup>. The G line was related to the first-order scattering of the  $E_{2g}$  phonons of sp<sup>2</sup> carbon atoms and the D line as a breathing mode of  $\kappa$ -point

phonons of  $A_{1g}$  symmetry is assigned to structural imperfections induced by the attachment of hydroxyl and/or epoxide groups on the carbon surface [6].

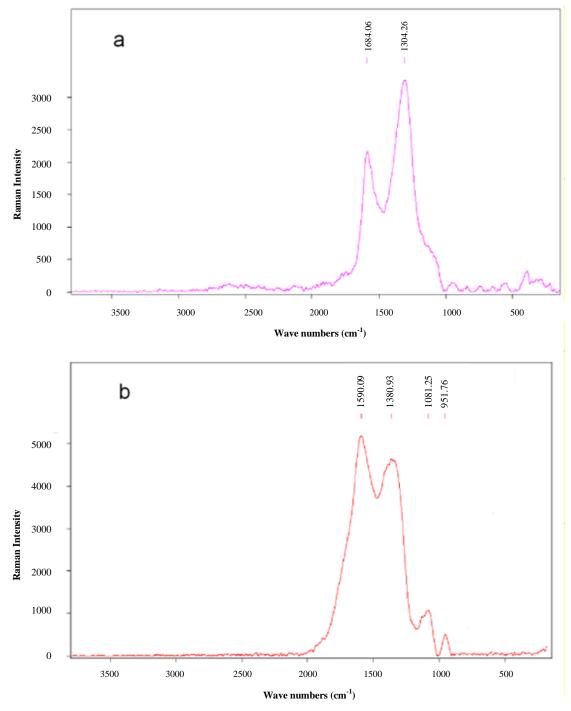


Figure 4. Raman spectrum of (a) GO, b GO@SiO2.

#### **Optimization of SPE procedures**

#### Effect of pH

The metal chelate stability constant and its chemical stability considerably influence the SPE recovery. The pH shows a very important role on metal chelate formation and following extraction. Therefore, pH was the first optimized parameter. pH of the analyte solutions was adjusted to desired values with diluted hydrochloric acid (0.1 mol  $L^{-1}$ ) and/or ammonia solution (0.1 mol  $L^{-1}$ ) The variation in recovery of Cu(II) with pH is shown in Figure 5. According to the results shown in Figure 4 up to pH 4.5-5.0, complete

recoveries are obtained. However, at higher pH values, percentage recovery decreases. This is due to fact that in an acidic solution the protonation of  $GO@SiO_2$  occurs and there is a weak tendency for retention between Cu(II) and  $GO@SiO_2$ , whereas at higher values (pH>5), Cu(II) reacts with hydroxide ions to produce Cu(OH)<sub>2</sub>. Therefore, pH 4.5 was selected for subsequent work and real sample analysis.

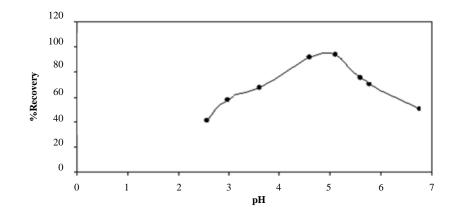


Figure 5. Influence of sample pH on the percentage recovery of Cu (II), conditions: 100 mg adsorbent, 50 ml of 5 mg/L of copper ions.

#### Effect of flow rates of sample and eluent solution

The effects of flow rates of sample solution and eluent solution on the recovery of Cu (II) was also examined between under the optimum conditions in the range of 1–10.0 mL min<sup>-1</sup> by controlling the flow rate with peristaltic pump. The recovery of the ions were independent of flow rate in the range of 0.5–2.0 mL min<sup>-1</sup> for eluent solution and range of 1–5.0 mL min<sup>-1</sup> for sample solution.

#### Effect of sample solution volume

Another parameter studied to find the best experimental conditions is the volume of sample solution and/or analyte

concentration. For this determination, 50.0–1100.0 mL of sample solutions containing 5ppm Cu(II) was processed according to the suggested procedure. The recovery of Cu(II) was quantitative (>98%) obtained up to a sample volume of 1000.0 mL and the adsorbed Cu(II) can be eluted with 5 mL eluent. Therefore, an enrichment factor of 200 was achieved by this method. Finally In our suggested procedure, a sample volume of 50.0 mL was chosen for preconcentration method.

#### Effect of amount of sorbent

To reach a high extraction recovery, different amounts of  $GO@SiO_2$  ranging from 50 to 300 mg were applied to extract the target compounds from the sample solutions.

The results are shown in Figure 5 from which it can be seen that the extraction recovery achieved by 100 mg, but almost the same as obtained with 300 mg or more than of the adsorbent. Based on the above results, 100 mg of  $GO@SiO_2$  was selected for the following experiments.

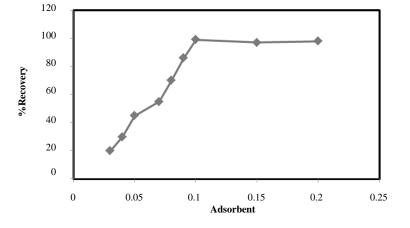


Figure 6. Removal percentages of cooper ions at different amounts of adsorbent, conditions: 50 ml of 5 mg/L of copper ions, pH= 4.5.

#### Eluent type and its volume

Another important factors which affect the percent of recovery are the type, volume, and concentration of the eluent solution used for the removal of metal ions from the sorbent. For this determination, various types of eluents were examined according to the suggested procedure. The results for this study are listed in Table 1. 4.0 mol  $L^{-1}$ 

HNO<sub>3</sub> was found to be adequate for quantitative elution ( $\geq$ 95%).The effect of eluent volume on the recovery of Cu (II) was also studied and listed in table.2. the results show the best quantitative recovery is 5.0 mL of 4.0 molL<sup>-1</sup> HNO<sub>3</sub>. As a result 5.0 mL of 4.0 molL<sup>-1</sup> HNO<sub>3</sub> was nominated in the subsequent preconcentration method.

Table 1. Effect of type and	concentration of various eluen	t on the recovery of Cu.

%Recovery				
Eluent concentration (M)	1	2	3	4
NaOH	9	11	12	22
NH <sub>3</sub>	12.5	12.5	18	19
HCI	12.5	16.5	30	35
H <sub>2</sub> SO <sub>4</sub>	25	28	28.5	36
HClO <sub>4</sub>	21	33	38	39
HNO <sub>3</sub>	65	77	98	98.5

A. Moghimi / Journal of Chemical Health Risks 8(3) (2018) 223-238

Table 2. Effect of eluent volume on the recovery of Cu	a.
--	----

Volume of eluent	%Recovery
HNO <sub>3</sub> (4M) 2 ml	44
HNO <sub>3</sub> (4M) 4 ml	79.5
HNO <sub>3</sub> (4M) 5 ml	98.5
HNO <sub>3</sub> (4M) 8 ml	98.5
HNO3 (4M)10ml	99

#### **Reusability of Column**

The stability and potential regeneration of the column were studied. After each time extraction, the column was washed with 10 mL of MeOH and 10mL of deionized water. Therefore, the column was available for a next extraction immediately at least 50 adsorption elution cycles without significant decrease in the recovery of Cu(II) ions.

#### Effect of foreign ions

The influence of common foreign ions on the adsorption of Cu(II) on  $GO@SiO_2$  were studied. In this work, 50.0 ml

solutions containing 5ppm of Cu and various amounts of interfering ions were treated according to the suggested procedure. The tolerance level was defined as the maximum concentration of the foreign ion causing a change in the analytical signal no higher than 5%, when compared with the signal of 5ppm copper alone. The results, listed in Table 3, demonstration that the presence of major cations and anions in natural water hasno important influence on the adsorption of Cu (II) ions under the designated conditions.

Table 3. Effect of foreign	ions on the percent recove	ery of 5 ppm Cu in w	ater samples.

Diverse ion	Amounts taken(mg)	% Found	%Recovery of Cu <sup>2+</sup> ion
Na <sup>+</sup>	92.4	1.15(2.4) <sup>b</sup>	98.3(1.8)
$\mathbf{K}^{+}$	92.6	1.32(2.3)	98.6(2.9)
$Mg^{2+}$	14.6	0.3(1.2)	98.9(1.8)
Ca <sup>2+</sup>	26.3	2.26(3.0)	98.6(1.6)
<b>Sr</b> <sup>2+</sup>	2.46	2.86(2.6)	98.4(2.0)
Ba <sup>2+</sup>	2.66	3.16(2.1)	98.3(2.3)
Mn <sup>2+</sup>	2.66	1.36(2.2)	93.3(2.8)
Co <sup>2+</sup>	2.16	1.4(2.3)	99.1(2.9)
Ni <sup>2+</sup>	1.66	2.0(2.4)	98.6(2.6)
Zn <sup>2+</sup>	2.38	1.93(2.1)	98.4(2.2)
$\mathbf{Cd}^{2+}$	2.66	1.92.0)	98.2(2.8)
Pb <sup>2+</sup>	0.64	2.3(1.9)	93(2.3)
$Hg^{2+}$	0.44	2.81(2.1)	93.3(2.8)
$\mathbf{Ag}^{+}$	2.63	3.46(2.9)	96.6(2.9)
Cr <sup>3+</sup>	1.33	2.92(2.30	93.3(2.4)
<b>UO</b> <sup>2+</sup>	2.84	2.8(2.1)	98.3(2.3)

<sup>a</sup> Initial samples contained  $10\mu g \operatorname{Cu}^{2+}$  and different amounts of various ions in 100 mL water(0.1 M acetate ion).<sup>b</sup> Values in parentheses are RSDs based on five individual replicate analysis.

#### Analytical figures of merits

Under optimized conditions, a calibration curve for Cu (II) was found by preconcentrating a series of Cu (II) standards according to the procedure mentioned. The curve was linear from 1.0 mg/l to 7.0 mg/l for Cu. As analytical figures of merit, limit of detection (LOD), limit of

quantification (LOQ) defined as  $3S_B/m$  and  $10S_B/m$ , relative standard deviation (RSD) for the proposed preconcentration and speciation method have been determined and listed in Table 4.

Table 4. Analytical figures of merits for Cu.

0.85%
0.175
0.585
200
5
Y=0.0854X+0.0109
0.9966

#### Determination of copper in real water samples

Three type of water samples (information described in section 2.7sampling) were used for the determination of copper. The analytical results are given in Table 5. The percent of recoveries for the addition of different concentrations of Cu (II) to water samples were 97.5 and 99%. These satisfactory percent of recoveries indicate no significant effects from the matrix composition of the real water samples.

Table 5. Determination of copper in real water samples.
---

Sample	Taken amount (µg)	Flame atomic absorption (µg)	%Recovery
Tap water (Tehran,	0	$3.0(2.2)^{a}$	-
Iran)	30	34.5(1.0)	98
Tap water	0	4.2(1.9)	-
(Pakdasht, Tehran, Iran)	30	36.3(1.0)	97.5
Mineral water	0	$N.D^b$	-
(Damavand Company)	30	30.5(0.8)	99

a) Values in parentheses are RSD based on four replicated analyses.

b) No adsorption, passes through column

#### CONCLUSIONS

In conclusion, the projected SPE possesses the following advantages: the technique is rapid when compared with the previously reported procedures for the separation and determination of copper; the time taken for the separation and determination of copper in a 500 mL sample is at the

most 30 min. Furthermore, it is a simple, very sensitive, selective and reproducible method for the separation of  $Cu^{2+}$  and in this work the recovery yields obtained with  $GO@SiO_2$  were about 95-100% then it show graphene oxide and its derivation e.g. in this work  $GO@SiO_2$  is full

of potential for use as a adsorbent in the extraction method like SPE and SPME. This composite may find applications in the development of sensor, super capacitor device, or as support for catalysts, and others.

#### ACKNOWLEDGEMENTS

The authors are so thankful to the Laboratory Complex of I.A.U. for valuable technical assistance and support.

#### REFERENCES

1. Bulut V.N., Tufekci M., Duran C., Soylak M., Kantekin H., 2010. 3D Visualization of iron oxide nanoparticles in MRI of inflammatory. Clean–Soil Air Water. 38, 678-684.

2. Graf D., Molitor F., Ensslin K., Stampfer C., Jungen A., Hierold C., 2007. Spatially Resolved Raman Spectroscopy Nano Lett. 7, 238–242.

3. Ansari I.A., Dewani V.K., Khuhawar M.Y., 1999. Flavonoid constituents of Chorizanthe diffusa with potential cancer chemopreventive activity. J Chem Soc Pak. 21, 359-365.

4. Margel S., Gura S., Bamnolker H., Nitzan B., Tennenbaum T., Bar-Toov B., Hinz M., Seliger H., in: U. Hafeli, W.schutt, J.Teller,M.Zborowski (Eds.),1997, Scientific and Clinical Applications of Magnetic Carriers, Plenum Press, New York. 481–494.

5. Afkhami A., Moosavi R., 2010. Adsorptive removal of Congored, acarcinogenic textiledye, fromaqueous solutions by maghemite nano particles. J Hazard Mater. 174, 398–403.

6. White B.R., Stackhouse B.T., Holocombe J.A., 2009. Magnetic  $\gamma$ -Fe2O3 nanoparticles coated withpoly-l-cysteine for chelation of As(III), Cu(II), Cd(II), Ni(II), Pb(II) and Zn(II). J Hazard Mater. 161, 848–853.

7. Tuutijarvi T., Lu J., Sillanpaa M., Chen G., 2009.As(V)adsorption on maghemitena- noparticles. J Hazard Mater. 166, 1415–1420.

8. Batterham G.J., Munksgaard N.C., Parry D.L., 1997.Tthe effects of poverty on child health and development Sociology. J Anal At Spectrom. 12, 1277-1282.

9. Hummers W.S., Offeman R.E., 1958. Functionalized Graphene and Graphene Oxide: Materials Synthesis. J Am Chem Soc. 80, 1339-1344.

10. Camel V., 2003. Solid phase extraction of trace elements - CNSTN, Spectrochim. Acta Part B. 58, 1177-1182.

 Castillo M., Pina-Luis G., D'ıaz-Garc'ıa M.E., Rivero I.A., Redalyc, 2005. Solid-Phase Organic Synthesis of Sensing Sorbent Materials. J Braz Chem. Soc. 16, 412-417.
Eaton A.D., Clesceri L.S., Greenberg A.E., 1995. Standard Methods for the examination of water and waste

water, 19th ed ,American Public Health Association, Washington, DC.

13. Welcher F.J., Boschmann E., 1979. Organic Reagents for Copper, Krieger Huntington, New York.

 Marczenko Z., 1986. Separation and Spectrophotometric Determination of Elements, Ellis Horwood, London.

15. Carasek E., Tonjes J.W., Scharf M., 2002. Solvent Microextraction: Theory and Practice. Quim Nova. 25, 748-755.

16. Ceccarini A., Cecchini I., Fuoco R., 2005. Flow injection micelle-mediated methodology for determination of lead. Microchem J. 79, 21-29.

17. Tuzen M., Soylak M., Citak D., Ferreira H.S., Korn M.G.A., Bezerra M.A., 2009. A preconcentration system for determination of copper and nickel in water and food samples employing flame atomic absorption spectrometry. J Hazard Mater.162, 1041-1047.

18. Choi Y.S., Choi H.S., 2003. Separation of Trace Amount Zn (II) - Journal of Chemical Health Risks, Bull. Korean Chem Soc. 24, 222-229.

19. Tuzen M., Narin I., Soylak M., Elci L., 2004. XAD-4/PAN Solid Phase Extraction System for Atomic Absorption Spectrometric Determinations of Some Trace Metals in Environmental Samples. Anal Lett. 37,473-480.

20. Karousis N., Sandanayaka A.S.D., Hasobe T., Economopoulos S.P., Sarantopouloua E., Tagmatarchis N., 2011. The Role of Oxygen during Thermal Reduction of Graphene Oxide. J Mater Chem. 21, 109-118. 21. Smith M.B., March J., 2001. March's advanced organic chemistry: reactions, mechanisms, and structure. New York: John Wiley & Sons Inc, 1182–3.

22. Mermoux M., Chabre Y., Rousseau A., 1991. FTIR and carbon-13 NMR study of graphite oxide. Carbon, 29(3),469–74.

23. Cataldo F., 2003. Structural analogies and differences between graphite oxide and C60 and C70 polymeric oxides (fullerene ozopolymers). Fuller Nanotub Car N. 11(1), 1–13.

24. Tohidifar H., Moghimi A., Ayvazzadeh O., Eskandari S., 2013. Determination of lead (II) in milk by flame atomic absorption spectrometry after solid phase extraction. Asian Journal of Chemistry. 25 (11), 5981-5987.

25. Thistlethwaite P.J., Hook M.S., 2000. Diffuse reflectance Fourier transform infrared study of the adsorption of oleate / oleic acid onto titania, Langmuir. 16, 4993-5007.

26.Taguchi, H., Paal B., Armarego W.L.F., 1997. Glycerylether monooxygenase [EC 1.14. 16.5], Part 9. Stereospecificity of the oxygenase reaction. J. Chem. Soc. Perkin Trans, 1(3), 303–7.

27. Moghimi A., Yari M., 2014. Preconcentration of trace Ni (II) using C18 disks nano graphene with amino propyltriethoxysilane (APTES), Merit Research Journal of Environmental Science and Toxicology. 2(5), 110-119.

28. Moghimi A., Akbarieh S.P., 2014. Evaluation of Solidphase Extraction Sorbent with Octadecane-functionalized Nano Graphene (ODG) for the Preconcentration of Chromium Species in Water, International Journal of Scientific Research in Knowledge. 2(1), 8-21.

29. Moghimi A., 2013. Detection of trace amounts of Pb(II) by schiff base-chitosan-grafted multi-walled carbon nanotubes. Russian J Physic Chem A. 87(7), 1203-1209.

30. Su X.G., Wang M.J., Zhang Y.H., Zhang J.H., Zhang H.Q., Jin Q.H., 2003. Separation and preconcentration procedures for the determination of lead using spectrometric techniques: A review , Talanta. 59, 989-995.

31. Soylak M., Karatepe A.U., Elci L., Dogan M., Column preconcentration/separation and atomic absorption spectrometric determinations of some heavy metals in table

salt samples using amberlite XAD-1180,2003.Turkish Journal of Chemistry. 27(2), 235-242.

32. Szabo T., Berkesi O., Dekany I., 2005. DRIFT study of deuterium-exchanged graphite oxide. Carbon. 43, 3186–9.

33. Moghimi A., 2014. Extraction of Ni (II) on micro crystalline naphthalene modified with organic-solution-processable functionalized nano graphene. Russian Journal of Physical Chemistry A. 88(7), 1177-1183.

34. Moghimi A., Abdouss M., 2013. Extraction of Co (II) by Isocyanate Treated Graphite Oxides (iGOs) Adsorbed on Surfactant Coated C18 Before Determination by FAAS. Int J Bio-Inorg Hybd Nanomat. 2(1), 319-327.

35. Moghimi A., Sabertehrani M., Waquif-Husain S., 2007. Preconcentration and determination of chromium species using octadecyl silica membrane disks and flame atomic absorption spectrometry. Chinese Journal of Chemistry. 25 (12), 1859-1865.

36. Tarigh. G.D., Shemirani F., 2013. Magneticmulti-wall carbon nano tubenano composite as an adsorbent for preconcentration and determination of lead(II) and manganese (II) in variousmatrices . Talanta. 115, 744–750.

 Moghimi A., Poursharifi M.J., 2012. Perconcentration of Ni (II) from Sample Water by Modified Nano Fiber, Oriental Journal of Chemistry. Orient J Chem. 28(1), 353-356.

38. Moghimi A., 2014. Separation and extraction of Co (II) using magnetic chitosan nanoparticles grafted with  $\beta$ -cyclodextrin and determination by FAAS. Russ J Phys Chem A. 88(12), 2157-2164.

39. Moghimi A., Siahkalrodi S.Y., 2013.Extraction and Determination of Pb (II) by Organic Functionalisation of Graphenes Adsorbed on Surfactant Coated C18 in Environmental Sample. Journal of Chemical Health Risks. 3 (3), 01-12.

40. Liu Q., Shi J.B., Wang T., Zeng L.X., Jiang G.B., 2011.Graphene and graphene oxide sheets supported on silica as versatile and high-performance adsorbents forsolid-phase.

41. Akhavan O., 2010. The effect of heat treatment on formation of graphene thin films from graphene oxide nan sheets, Carbon. 48, 509–519.

42. Kudin K.N., Ozbas B., Schniepp H.C., Prud'homme R.K., Aksay I.A., Car B., 2008. Raman spectra of graphite oxide and functionalized graphene sheets. Nano Letters. 8, 36–41.

43. Ferrari A.C., Meyer J.C., Scardaci V., Casiraghi C., Lazzeri M., Mauri F., Piscanec S., Jiang D., Novoselov K.S., Roth S., Geim A.K., 2006. Raman spectrum of graphene and graphene layers. Physical Review Letters. 97, 187401–1874014.

44. Zhu Y., Murali S., Cai W., Li X., Suk J.W., Potts J.R., Ruoff R.S., 2010. Graphene and graphene oxide: synthesis, properties, and applications, Adv. Mater. 22, 3906–3924. 45. Xu Y.X., Bai H., Lu G.W., Li C., Shi G.Q., 2008.Flexible graphene films via the filtration of watersoluble no covalent functionalized graphene sheets. Journal of the American Chemical Society. 130, 5856–5857.

46. Guo H.L., Wang X.F., Qian Q.Y., Wang F.B., Xia X.H., 2009. A green approach to the synthesis of graphene Nano sheets. ACS Nano. 3, 2653–2659.

47. Leng W.G., Chen M., Zhou S.X., Wu L.M., 2010. Capillary force induced formation of monodisperse polystyrene/silica organic–inorganic hybrid hollow spheres. Langmuir. 26, 14271–14275.