

Effects of *Cyperus rotundus* extract

Eur J Transl Myol 28 (4): 362-369, 2018

Cell Toxicity and inhibitory effects of *Cyperus rotundus* extract on *Streptococcus mutans*, *Aggregatibacter actinomycetemcomitans* and *Candida albicans*

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Abstract

Periodontitis and tooth decay are common dental diseases. There are some bacterial risk factors such as *Streptococcus mutans*, *Aggregatibacter actinomycetemcomitans* and *Candida albicans* complications, tubers of *Cyperus rotundus* was used to determine the antimicrobial effect of fungi. Considering the increasing resistance of bacteria and fungi to antibiotics and their alcoholic and aqueous extracts and essential oil on *Streptococcus mutans*, *Aggregatibacter actinomycetemcomitans* and *Candida albicans*, as well as an examination cytotoxicity on gingival fibroblast cells. Alcoholic and aqueous extracts and essential oil prepared from tubers of *Cyperus rotundus*. After bacterial and fungal cultures, the inhibition zone, Minimum inhibitory concentrations (MIC), and Minimum Bactericidal Concentration (MBC), were studied. Cytotoxicity was also evaluated using the Methyl tetrazolium test (MTT). The results showed that *Cyperus rotundus* alcoholic extract had the greatest effect on inhibition of growth and death of *Streptococcus mutans*, *Aggregatibacter actinomycetemcomitans*. According to the present study, *Cyperus rotundus* extract can be considered as a suitable candidate for the treatment and prevention of periodontitis and tooth decay.

Key Words: *Cyperus rotundus*, Periodontitis, *Streptococcus mutans*, *Candida albicans*, *Aggregatibacter actinomycetemcomitans*

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Periodontitis is one of the most common oral diseases and one of the main causes of tooth loss worldwide.¹ In Periodontitis, periodontal ligament and other tissues that hold the teeth, are invaded by microorganisms. The clinical symptoms includes gingivitis with bleeding, tooth loosening, gum dwindle, unpleasant odor of mouth, alveolar bone destruction and finally tooth loss. *Aggregatibacter actinomycetemcomitans* is a gram-negative anaerobic bacterium that plays an important role in the development of periodontitis, especially aggressive periodontitis.² Periodontal pockets can easily be saturated with saliva, and are suitable sites for growth of *Candida albicans* species.³ *Candida albicans* accumulate as biofilm and its proliferation occurs in biofilm at periodontal pockets. Indeed, deep periodontal pockets can alter the balance of subfamily microflora, and then

this imbalance leads to colonization of destructive microorganisms in periodontal tissue.⁴ *Candida albicans* colonizes various parts of the mouth especially caries lesion and dental plaque. Dental caries is a common infectious disease of oral cavity⁵ Several factors are effective in predicting the risk of tooth decay such as oral microbial flora especially *Streptococcus mutans*, the major microbial cause of tooth decay. Twenty-dive strains of *Streptococcus mutans* have been discovered, which can destroy the enamel of the tooth by producing lactic acid and sucrose fermentation.^{5,6} One of routine ways to reduce bacterial damages is administration of antibiotics that is accompanied with challenges such as the long course of treatment, side effects and bacterial resistance. Therefore, finding an appropriate replacement

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Table 1. Mean of inhibition zone

Microorganism	Mean of inhibition zone			
	alcoholic extract	aqueous extract	essential oil	chlorhexidine
<i>S. mutans</i>	19.6 mm	9.3 mm	13.3 mm	22.3 mm
<i>A. actinomycetemcomitans</i>	33.3 mm	No inhibition zone	22.3 mm	42.3 mm
<i>C. albicans</i>	No inhibition zone	No inhibition zone	No inhibition zone	25.3 mm

for antibiotic in treatment of periodontitis is particularly important. *L. Cyperus rotundus* from the Cyperaceae family, is a perennial herb.^{7,8} Tubers of *Cyperus rotundus* has terpenoid endoperoxide called 10,12-calamenene, a steroid glycoside called sitosterol- β -D-galactopyranoside and herbal components such as khellin, visnagin, salicylic acid, and p-coumaric acid.⁹ To date, several therapeutic properties have been investigated for Tubers of *Cyperus rotundus* but the antimicrobial properties and effects of Iranian species of this plant have not been studied yet. Therefore, due to the increased drug resistance of bacteria and side effects of antibiotics and advantages of using herbal medicine, the aim of the present study was to evaluate the inhibitory effect of Essential Oil, alcoholic and aqueous extracts of the tubers of *Cyperus rotundus* and its comparison with Chlorhexidine on *Streptococcus mutans*, *Aggregatibacter actinomycetemcomitans* and *Candida albicans*.

Materials and Methods

Preparation and Extraction

Tubers of *Cyperus rotundus* plant was provided from Tehran Agricultural Research Center (Iran) and dried for 2 weeks. 1000 grams of Tubers of *Cyperus rotundus* were thoroughly crushed. Then 400 ml water was added to 400 g of crushed plant and the standard hydrodistillation method was performed using Clevenger type apparatus to prepare the essential oil. To prepare aqueous and alcoholic extracts, separately, 75 g of the crushed plant was mixed 750 ml of distilled water and pure ethanol and stirred slowly for 72 hours and then separated using filters to obtain initial extracts. The initial extract was introduced into a vacuum distillation apparatus at 80°C,

the solvent evaporated slowly for one hour and the concentrated extract was obtained.

Preparation and culture of microorganisms

Three microorganisms including *C. albicans* (ATCC 10231), *S. mutans* (ATCC 35668), and *A. actinomycetemcomitans* (A.a. Jp2 Nov996) were used in this study. Sabouraud dextrose broth (SDA) (Sigma, USA) and BHI agar (Sigma, USA) was used for culture of *C. albicans* and *S. Mutans* respectively. Cultures were incubated in aerobic conditions at 37 °C and after 24 hours, they were cultured on a BHI agar and SDA culture medium to create a single colony and again incubated in aerobic conditions for 24 hours. *A. actinomycetemcomitans* was first cultured for enrichment on the BHI medium (Sigma, USA) under anaerobic conditions (with type A gas pack) for 48 hours at 37 °C and then was cultured on BHI agar and incubated in anaerobic conditions for 72 hours to create a single colony

Inhibition zone analysis

In order to evaluate the antibacterial effect of the plant extracts, well diffusion agar method was used. Standard dilutions of microorganism were provided according to MacFarland.¹¹ Sterilized swabs were used to prepare suspensions of each microorganism and then to culture them on BHI agar medium. After cultivation, tiny wells were created on BHI agar medium and finally, 50 μ l of aqueous and alcoholic extracts were separately inoculated into the wells. Incubation conditions for *C. albicans* and *S. mutans* were aerobic at 37 °C for 18-24 hours, while for *A. actinomycetemcomitans* were anaerobic at 37 °C, for 72 hours. The wells containing distilled water and chlorhexidine were considered as negative and positive controls, respectively. After incubation, the diameter of

Table 2. MIC and MBC test results

Test	<i>S. mutans</i>		
	alcoholic extract	aqueous extract	essential oil
MIC	1.56 %	50%	6.25%
MBC	25%	100%	-
Test	<i>A. actinomycetemcomitans</i>		
	alcoholic extract	aqueous extract	essential oil
MIC	3.12%	-	6.25%
MBC	12.5%	-	12.5%

Table 3. Mean cell viability in study groups

Group	N	Mean	Std. Deviation
Negative Control	8	100.000	22.07
Positive Control	8	15.248	14.00
chlorohexidine	8	20.026	4.73
Essential oil	9	109.424	40.71
Alcoholic extraction	9	150.320	51.14
Total	42	81.426	61.85

the inhibition zone was measured. The tests were repeated three times to obtain more reliable data.

Determining Minimum Inhibitory Concentrations (MIC) and Minimum Bactericidal Concentration MBC

Minimum inhibitory concentrations (MIC) test was performed using microdilution and colorimetric methods. Sterilized BHI broth medium (100 µl) of was added to each well of the plate. Tubers of *Cyperus rotundus* extracts (100 µl) were added to the first well and then serial dilutions were added to others until the 12th well. Finally, 10 µl of diluted McFarland suspension was added into all wells except for negative control, so that the final concentration of the microorganism was 5.55 cfu/ml. Negative control included extract and BHI broth medium without inoculation of microorganisms and positive control including BHI broth and microorganisms without *Cyperus rotundus* extracts. This test was repeated three times and performed for each microorganism separately. *S. mutans* plates were incubated for 18 to 24 hours, and incubation for *A. actinomycetemcomitans* was in anaerobic conditions for 48 hours. After incubation, 20 µl of sterilized Resazurin dye (with 0.01% concentration) (Sigma, USA) was added to each well and then incubated for 2 hours at 37 °C. After incubation, the lowest concentration (blue to pink) was considered as MIC. In order to conduct the Minimum Bactericidal Concentration (MBC) Test, MIC well and a one lower concentration and other higher concentrations were considered. Microorganisms of above wells were cultured on BHI agar, and the lowest concentration, which killed 99% of the microorganisms, was considered as MBC for *S. mutans* and *A. actinomycetemcomitans*.

Cytotoxicity test using Methyl tetrazolium assay (MTT)

Cytotoxicity measurement was performed using Methyl tetrazolium assay (MTT). In order to evaluate the effect of cytotoxicity, MTT test was performed at a higher and a lower concentration than the minimum inhibitory concentration for Tubers of *Cyperus rotundus* extracts. Samples were divided into 9 groups: 1: negative control

containing culture medium. 2: positive control containing distilled water. 3: chlorhexidine (with 0.2 % concentration). 4: Alcoholic extracts at one concentration higher than MIC. 5: Alcoholic extracts at MIC concentration. 6: Alcoholic extracts at one concentration lower than MIC. 7: Essential oil at one concentration higher than MIC. 8: Essential oil at MIC concentration. 9: Essential oil at one concentration lower than MIC. In case of multiple MIC values, the higher value was used in cytotoxicity test.

Cell Culture and treatment

Human gingival fibroblast cells (HGF) from the Pasteur Institute of Iran were purchased in a 75 cm² flask. Cell culture performed on a complete culture medium containing Dulbecco's modified eagle medium (DMEM), (Gibco, USA) supplemented with 10% heat-inactivated fetal bovine serum (FBS), 100 U / mL penicillin G (Sigma, USA) and 100 mg / mL streptomycin (sigma, USA) and then incubated in CO₂ incubator. After reaching the 80% confluency, the cells were collected by trypsin and EDTA and centrifuged at 800g for 5 min. The supernatant was discarded and 1.5×10³ cells / ml of HGF cells in a volume of 200 µL were seeded in each well of plate. The plate was stored at 37 °C for 24 hours. The supernatant was then discarded and washed with PBS. 100 µl of MTT was added to all wells in dark place. The cells then returned to the CO₂ incubator to view the Formazan crystals by inverted microscope. The supernatant was discarded and 100 µl DMSO (Sigma, USA) was added to each well. Then the absorbance of each well was read at 570 and 620 nm using ELISA reader. The percentage of subtraction of the absorbed cells affected by the drug was divided to absorption of the control cell as the percentage of cells that remained stable.

Statistical analysis

Data was first analyzed by SPSS v.20 program. Analysis of variance and Tukey's range test were used to

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Table 4 – A. Comparison of cytotoxicity between study groups

Group	Group	Mean Difference	Significance
Negative Control	Positive Control	84.75131*	.001
	CLX	79.97382*	.001
	EO High	-3.83944	1.000
	EO Medium	-12.91449	1.000
	EO Low	-11.51832	1.000
	Alcohol High	-63.52531	.167
	Alcohol Medium	-44.67714	.593
	Alcohol Low	-42.75742	.646
Positive Control	Negative Control	-84.75131*	.001
	CLX	-4.77749	1.000
	EO High	-88.59075*	.013
	EO Medium	-97.66579*	.005
	EO Low	-96.26963*	.006
	Alcohol High	-148.27661*	.000
	Alcohol Medium	-129.42845*	.000
	Alcohol Low	-127.50873*	.000
CLX	Negative Control	-79.97382*	.001
	Positive Control	4.77749	1.000
	EO High	-83.81326*	.023
	EO Medium	-92.88831*	.008
	EO Low	-91.49215*	.010
	Alcohol High	-143.49913*	.000
	Alcohol Medium	-124.65096*	.000
	Alcohol Low	-122.73124*	.000
EO High	Negative Control	3.83944	1.000
	Positive Control	88.59075*	.013
	CLX	83.81326*	.023
	EO Medium	-9.07504	1.000
	EO Low	-7.67888	1.000
	Alcohol High	-59.68586	.460
	Alcohol Medium	-40.83770	.861
	Alcohol Low	-38.91798	.889

demonstrate differences between observed variables. P value of <0.05 used to indicate statistical significance.

Results

Antibacterial properties of Tubers of Cyperus rotundus extracts

Inhibition zone for *S. mutans* and *A. actinomycetemcomitans* was the highest in the 0.2% chlorhexidine group 22mm and 42mm diameters respectively. In the case of *S. mutans*, inhibition zone in presence of the alcoholic extract, essential oil and aqueous extract was 19mm, 14mm and 10mm

respectively. Aqueous extract had no inhibitory effect on growth of *A. actinomycetemcomitans*, while a diameter of 42 mm for chlorhexidine, 35mm for alcoholic extract and 23mm for essential oil observed. Also, none of alcoholic extract, aqueous extracts and essential oil, had no effect on *C. albicans*. Therefore, MIC and MFC tests for *C. albicans* were not performed. Results of inhibition zone have been demonstrated in table 1. For microbial tests, the highest concentrations available in the aqueous extract and alcoholic extract of 500 mg / ml were used and the subsequent dilutions were prepared using this concentration. In the study of minimum inhibitory and

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Table 4 – B. Comparison of cytotoxicity between study groups

EO Medium	Negative Control	12.91449	1.000
	Positive Control	97.66579*	.005
	CLX	92.88831*	.008
	EO High	9.07504	1.000
	EO Low	1.39616	1.000
	Alcohol High	-50.61082	.668
	Alcohol Medium	-31.76265	.963
	Alcohol Low	-29.84293	.974
EO Low	Negative Control	11.51832	1.000
	Positive Control	96.26963*	.006
	CLX	91.49215*	.010
	EO High	7.67888	1.000
	EO Medium	-1.39616	1.000
	Alcohol High	-52.00698	.636
	Alcohol Medium	-33.15881	.952
	Alcohol Low	-31.23909	.966
Alcohol High	Negative Control	63.52531	.167
	Positive Control	148.27661*	.000
	CLX	143.49913*	.000
	EO High	59.68586	.460
	EO Medium	50.61082	.668
	EO Low	52.00698	.636
	Alcohol Medium	18.84817	.999
	Alcohol Low	20.76789	.998
Alcohol Medium	Negative Control	44.67714	.593
	Positive Control	129.42845*	.000
	CLX	124.65096*	.000
	EO High	40.83770	.861
	EO Medium	31.76265	.963
	EO Low	33.15881	.952
	Alcohol High	-18.84817	.999
	Alcohol Low	1.91972	1.000
Alcohol Low	Negative Control	42.75742	.646
	Positive Control	127.50873*	.000
	CLX	122.73124*	.000
	EO High	38.91798	.889
	EO Medium	29.84293	.974
	EO Low	31.23909	.966
	Alcohol High	-20.76789	.998
	Alcohol Medium	-1.91972	1.000

bactericidal concentration, the lowest concentration with the ability to inhibit growth in *S. mutans* was related to 1.56% alcoholic extract, which is 6.25% in essential oil and 50% in aqueous extracts. The minimum concentration of MBC was also observed in alcoholic extract at 25% concentration followed by aqueous extract at a concentration of 100% (Table 2). In *A. actinomycetemcomitans*, the lowest concentration of MIC was 3.12% for alcoholic extract and 6.25% for essential oil. The MBC content of alcoholic extract and

essential oil was obtained at 12.5% concentration (Table 2).

Cell cytotoxicity

After observing the more suitable antimicrobial effect of the alcoholic extract and essential oil, MTT assay were evaluated for HGF cells to investigate the possible cytotoxic properties. For this purpose, alcoholic extracts and essential oil were obtained at higher concentrations and lower than the MIC, and their cytotoxic effects were compared with 0.2% chlorhexidine over a 24 hour period.

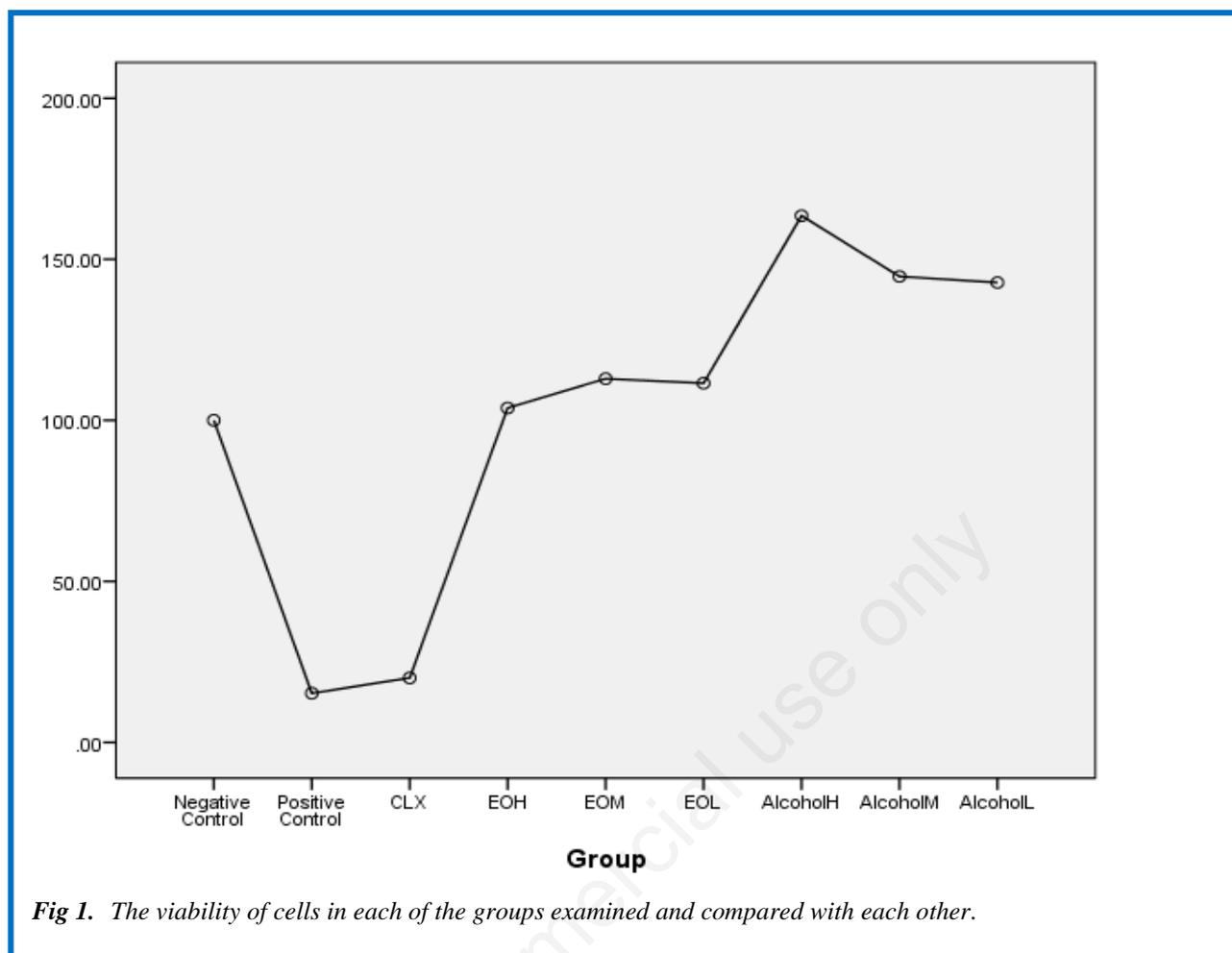


Fig 1. The viability of cells in each of the groups examined and compared with each other.

Based on the toxic effects on the cells, there was a significant difference between the groups of alcoholic extract, essential oil and chlorhexidine and positive control (p value = 0.000). chlorhexidine had the highest toxic effect on the cells with mean 20.02 ± 4.7 . Results of MTT showed that there was a significant difference in these groups with positive control ($P < 0.05$). (Table 4, A and B). However, there was no significant difference between chlorhexidine and positive control (p value = 1.000). Also, there was a significant difference between the treated cells using essential oil and alcoholic extracts with chlorhexidine ($p < 0.05$) for both groups at all concentrations. However, there was no significant difference between the essential oil and alcoholic extract (p value > 0.05) and both treatments showed low toxicity on the cells after 24 hours (Table 4, A and B). Figure 1 shows the viability of the cells in each of the groups examined and compared with each other.

Discussion

A. actinomycetemcomitans is one of the most important causes of periodontal disease.^{12,13} The main treatment of this disease is scaling and root planing and ultrasonic debridement to remove bacterial deposits. *S. mutans* is the most important bacteria causing dental caries.

Therefore, the use of mechanisms to reduce these microorganisms is essential.¹⁴ The aim of the present study was to investigate the effects of ethalonic and aqueous extracts and essential oil of Tubers of *Cyperus rotundus* on *S. mutans*, *A. actinomycetemcomitans* and *C. albicans*. The results of inhibition zone in this study, using statistical analyzes, showed a significant difference in growth of *S. mutans*, *A. actinomycetemcomitans* under the influence of ethalonic and aqueous extracts and essential oil of Tubers of *Cyperus rotundus*. In this study, alcoholic extract of Tubers of *Cyperus rotundus* in very low concentrations (7.18 mg / ml and 15.62 mg / ml) has inhibitory effects on *S. mutans* and *A. actinomycetemcomitans*. In terms of toxicity effect, alcoholic extracts of Tubers of *Cyperus rotundus* in 31.25 mg / ml concentration and essential oil in 6.25% volume concentration showed the lowest cytotoxicity. Chlorhexidine 0.2% showed the highest cytotoxicity after 24 hours compared to the control group (p value = 0.01). This comparison suggests that different concentrations of alcoholic extract and essential oil have no cytotoxic, while chlorhexidine has a 100% toxicity for cells. Antibacterial effects of extracts of Tubers of *Cyperus rotundus* have been observed in previous studies. However this effect on *A.*

actinomycetemcomitans as one of the most important factors involved in periodontal disease, has not been studied yet. Somayeh Kilani et al., Studied the antibacterial effects of essential oil of Tubers of *Cyperus rotundus*, on *Staphylococcus aureus*, *Enterococcus faecalis*, *Escherichia coli*, *Salmonella enteritidis* and *Salmonella typhimurium*, and concluded that the oil extracted from this plant has a strong inhibitory effect on *S. aureus* and *E. faecalis* and a moderate inhibitory effect on *S. typhimurium*.¹⁵ In another study, Yu et al., investigated the effects of Tubers of *Cyperus rotundus* extracts on *S. mutans*. The results showed that the extract was effective in inhibiting the growth of this bacteria and its acid production.¹⁶ Sharma and colleagues evaluated the effects of aqueous, alcoholic, chloroform and ether extracts of Tubers of *Cyperus rotundus* on *E. coli*, *Staphylococcus epidermidis*, *Bacillus cereus*, *Pseudomonas aeruginosa*, *Aspergillus niger* and *C. albicans*. They concluded that alcoholic extract had the highest inhibitory effect on *B. cereus*, *S. epidermium* and *P. aeruginosa* respectively. None of the extracts had inhibitory effect on *E. coli* and *C. albicans*.¹⁷ These results are in concordance with our study, as we found that none of the aqueous, alcoholic extracts and essential oil, had no inhibitory effect on *C. albicans* and the best antibacterial effect, and was due to alcoholic extract. Also, unlike other studies, in the present study, we used essential oil of Tubers of *Cyperus rotundus* to evaluate and compare its inhibitory and antibacterial effects with aqueous and alcoholic extracts. Our results demonstrated its antibacterial effect, although in comparison with alcoholic extract, this compound has less inhibitory effect. In the present study, the effects of aqueous extract of Tubers of *Cyperus rotundus* on *S. mutans* was observed only at a high concentration of 500 mg / ml, but did not have any inhibitory effects on *A. actinomycetemcomitans*. Results of cytotoxicity test suggested that alcoholic extract and essential oil not only have no toxic effects on HGF cells, but also could have proliferative effects. According to this study, it seems that the alcoholic extract of Tubers of *Cyperus rotundus* can be a potential candidate for mouthwash in the future. However, more laboratory and clinical studies are needed to confirm the results of this study. In conclusion, the results of this study show that extracts of Tubers of *Cyperus rotundus* have a good antibacterial effect and can be used as a treatment option in dental infections.

List of acronyms

HGF - Human gingival fibroblast cells
MBC - Minimum Bactericidal Concentration
MIC -Minimum inhibitory concentrations
MTT – Methyl tetrazolium test
SDA - Sabouraud dextrose broth

Author's contributions

MY, ET, MSH, BH, RSH, and MKH equally participated in experimental design, data collection, writing and revision of the manuscript.

Acknowledgments

None.

Funding This research was supported by Emam Khomeyni Dental Clinic and Shahid Shokri Dental Hospital of Baqiyatallah University of Medical Sciences.

Conflict of Interest

The authors report no conflicts of interests.

Ethical Publication Statement

We confirm that we have read the Journal's position on issues involved in ethical publication and affirm that this report is consistent with those guidelines.

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Received for publication: 06/11/2018

Accepted for publication: 12/11/2018