




REVIEW

Transcripts and tumors: regulatory and metabolic programming during biotrophic phytopathogenesis [version 1; peer review: 3 approved]

Lara Schmitz^{1*}, Sean McCotter^{2*}, Matthias Kretschmer^{2*}, James W. Kronstad ², Kai Heime¹

¹Institute for Microbiology and Genetics, Department of Molecular Microbiology and Genetics, Georg-August-University-Göttingen, Göttingen, Lower Saxony, D-37077, Germany

²Michael Smith Laboratories, Department of Microbiology and Immunology, University of British Columbia, Vancouver, BC, V6T 1Z4, Canada

* Equal contributors

v1 First published: 19 Nov 2018, 7(F1000 Faculty Rev):1812 (<https://doi.org/10.12688/f1000research.16404.1>)

Latest published: 19 Nov 2018, 7(F1000 Faculty Rev):1812 (<https://doi.org/10.12688/f1000research.16404.1>)

Abstract




Biotrophic fungal pathogens of plants must sense and adapt to the host environment to complete their life cycles. Recent transcriptome studies of the infection of maize by the biotrophic pathogen *Ustilago maydis* are providing molecular insights into an ordered program of changes in gene expression and the deployment of effectors as well as key features of nutrient acquisition. In particular, the transcriptome data provide a deeper appreciation of the complexity of the transcription factor network that controls the biotrophic program of invasion, proliferation, and sporulation. Additionally, transcriptome analysis during tumor formation, a key late stage in the life cycle, revealed features of the remodeling of host and pathogen metabolism that may support the formation of tremendous numbers of spores. Transcriptome studies are also appearing for other smut species during interactions with their hosts, thereby providing opportunities for comparative approaches to understand biotrophic adaptation.

Keywords

Ustilago maydis, transcriptome, RNAseq, pathogenicity, regulators, effectors, nutrients, metabolism

Open Peer Review

Reviewer Status 

	Invited Reviewers		
	1	2	3
version 1 published 19 Nov 2018			

F1000 Faculty Reviews are written by members of the prestigious F1000 Faculty. They are commissioned and are peer reviewed before publication to ensure that the final, published version is comprehensive and accessible. The reviewers who approved the final version are listed with their names and affiliations.

- 1 **Armin Djamei**, Austrian Academy of Sciences (OEA), Vienna, Austria
- 2 **Barry Saville**, Trent University, Peterborough, Canada
- 3 **José Ruiz-Herrera**, Centro de Investigación y de Estudios Avanzados del Instituto Politécnico Nacional, Irapuato, Mexico

Any comments on the article can be found at the end of the article.

Corresponding authors: James W. Kronstad (kronstad@msl.ubc.ca), Kai Heibel (kheibel@gwdg.de)

Author roles: **Schmitz L:** Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **McCotter S:** Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Kretschmer M:** Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; **Kronstad JW:** Conceptualization, Funding Acquisition, Supervision, Writing – Original Draft Preparation, Writing – Review & Editing; **Heibel K:** Visualization, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: Our research on *Ustilago maydis* is supported by a Discovery grant (to J.W.K.) and a CREATE training award (to the PRoTECT group) from the Natural Sciences and Engineering Research Council of Canada and by grants from the Deutsche Forschungsgemeinschaft (DFG) (to K.H) and the DFG-funded International Research Training Group (PRoTECT).

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Copyright: © 2018 Schmitz L *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution Licence](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Schmitz L, McCotter S, Kretschmer M *et al.* **Transcripts and tumors: regulatory and metabolic programming during biotrophic phytopathogenesis [version 1; peer review: 3 approved]** F1000Research 2018, 7(F1000 Faculty Rev):1812 (<https://doi.org/10.12688/f1000research.16404.1>)

First published: 19 Nov 2018, 7(F1000 Faculty Rev):1812 (<https://doi.org/10.12688/f1000research.16404.1>)

Introduction

Fungal phytopathogens in the order Ustilaginales generally attack cereal and grass plants to cause smut diseases, so named because of the tremendous masses of sooty spores produced in infected tissue (Figure 1). Biotrophic pathogens such as the smut fungi are dependent on living hosts to complete their life cycles. Effective adaptation to the host environment is therefore critical for overcoming the plant immune response and successfully exploiting host nutrients through remodeling of metabolism and effective competition. The mechanisms by which biotrophic pathogens manipulate host metabolism to divert carbon, nitrogen, or micronutrients such as iron for their own use are starting to be identified^{1–4}. Among the Ustilaginales, the *Zea mays* (maize) pathogen *Ustilago maydis* has emerged as an experimentally tractable model for studying the adaptation of biotrophic fungal pathogens to the host environment. As an example of the intricate interaction with the plant host, the life cycle of *U. maydis* involves germination of diploid teliospores on host tissue with subsequent mating between haploid meiotic progeny to form an invasive filamentous cell type^{5,6}. The filaments then form appressoria (invasion structures) to penetrate host tissue with subsequent extensive proliferation, induction of tumors (galls), and eventual formation of melanized teliospores^{5,7} (Figure 1). A large number of effectors are predicted for *U. maydis*, and these proteins are thought to play key roles in managing the infection process. To date, characterized effectors include Cmu1, Pit2, Pep1, and Tin2, which influence host defense, and See1 and Rsp3, which influence tumor progression and the fungal response to host defense, respectively^{3,8}.

In this review, we focus on recent genome-wide transcriptome studies of the infection process that have provided insights into the transcriptional regulation associated with disease, the deployment of effectors by *U. maydis* to manage the infection process, and the remodeling of host metabolism during fungal proliferation. Although not covered here, earlier studies



Figure 1. Tumor formation on maize by *Ustilago maydis*. An infected maize cob found in a cornfield close to Göttingen, Germany, in September 2018. At the end of the growing season, infected kernels give rise to greatly enlarged and bulbous plant tumors filled with black teliospores. Tumors can develop on all aerial parts of the plant but are most prominent in infected cobs.

also examined the transcriptome of *U. maydis* in culture and during infection^{6,7,9–23}. We also refer readers to a wealth of primary literature and recent reviews on the role of transcription factors, regulators, and effectors in the disease process for *U. maydis*^{13,14,20,24–27}.

Transcriptome analysis throughout the infection process

A recent time-resolved and genome-wide study of transcriptional changes occurring during the *U. maydis*–maize interaction provides detailed insights into the processes underlying infection, tumor formation, and sporogenesis²⁸. Among 14 regulatory modules identified, three gene sets/developmental programs were specifically associated with virulence and were upregulated during discrete developmental stages (Figure 2). These gene sets are expressed during 1) growth on the plant surface (early), 2) biotrophic development *in planta* (middle), and 3) tumor formation (late/sporogenesis). In all three sets, genes encoding secreted proteins (effectors) are significantly over-represented, emphasizing their critical role during biotrophic development. Metabolic changes are also predicted and, importantly, several transcription factors with a strong connection to each respective module were identified as potential key regulators (Figure 2).

For the early stage of infection, it is known that gene regulation by the zinc finger transcription factor Rbf1 is required for the initial steps of pathogenic development, including filament formation, induction of G2 cell cycle arrest, and appressoria-mediated plant penetration^{6,25}. Recent work by Lanver *et al.*²⁸ revealed that Rbf1 shows a strong regulatory connectivity to genes expressed at the early stage (0.5–1 days post inoculation [DPI]). Among the 398 genes that are upregulated, 49 encode secreted proteins that are enriched for hydrolytic enzymes with a predicted function in appressoria-mediated plant penetration. Several genes induced on the leaf surface encode sugar- or nitrogen-related transporters, indicating priming of *U. maydis* for altered nutrient availability during plant colonization.

The subsequent middle phase of biotrophic development (2–4 DPI) involves the re-initiation of fungal proliferation, establishment of a compatible biotrophic interaction, and the initiation of tumor formation. Enriched functional categories in the transcriptome include nitrogen and carbon source-related processes and components of the secretory pathway including the unfolded protein response (UPR)²⁸. Transcriptional regulation during the biotrophic stage is modulated by direct interaction between UPR components and key regulators of fungal development and pathogenicity²⁷. Potential transcriptional regulators of this stage include the Rbf1-regulated homeodomain transcription factor Hdp2^{6,7} and the C2H2-zinc finger transcription factor Biz1²⁴.

Among the 228 genes encoding putative effectors that were upregulated in the middle stage, 153 lack functional signatures. However, the characterized genes encoding the effectors Pep1, Pit2, Cmu1, See1, Tin2, and Rsp3 are highly induced at this

	Plant surface (0.5-1 dpi)	Proliferation <i>in planta</i> (2-4 dpi)	Tumor and spore formation (4-8 dpi)
Potential key regulators	bE/bW Rbf1	Hdp2 Biz1	Nlt1 Fox1 Ros1
Effectors	49 potential effectors Pep1	228 potential effectors Pep1 Pit2 Cmu1 Tin2 Rsp3 See1	38 potential effectors See1
Nutrient assimilation	• Hydrolysis and uptake of plant cell wall components	• Ammonium transp.: Ump1, Ump2 • Oligopeptide transp.: Opt2, Opt3, Opt4 • Sucrose transporter Srt1 • Glucose transporter Hxt1 • Aspartic proteases	• Protein catabolism • Autophagy • lipid transport and metabolism • nitrogen and carbon catabolite derepressors

Figure 2. Regulators, effectors, and metabolism during *Ustilago maydis* infection of maize. Overview of transcriptional regulation, the timing of effector function, and metabolic changes and requirements during early, middle, and late stages of the *U. maydis*–maize interaction. DPI, days post inoculation.

stage, and these are required for full virulence of *U. maydis* and/or the suppression of plant defense reactions and fungal defense, respectively (reviewed in 3). Organ- and cell type-specific transcriptome analysis revealed that genes encoding the core effectors Pep1, Pit2, Cmu1, and Tin2 are expressed independent of the plant organ or cell type in leaves and tassels, whereas See1 is exclusively expressed in leaf tissue^{29,30}. Tumor formation occurs by distinct mechanisms in hyperplastic bundle sheath-derived tumor cells (HPT) or hypertrophic mesophyll-derived tumor cells (HTT)³¹. See1 mediates only bundle sheath cell-derived tumor formation, whereas mesophyll-derived tumor formation is independent of See1³¹.

During tumor formation in the middle stage, the ammonium transporters Ump1 and Ump2 and the oligopeptide transporters Opt2, Opt3, and Opt4 appear to mediate nitrogen acquisition. Ump1, Ump2, and Opt2 are induced already on the leaf surface and further upregulated *in planta*^{28,32}. The fact that two secreted aspartic proteases are co-expressed with the OPTs suggests that the breakdown of extracellular proteins and peptide uptake may be part of the biotrophic developmental program of *U. maydis*²⁸. Carbon assimilation during biotrophic development is known to occur mainly via the high-affinity sucrose and glucose transporters Srt1 and Hxt1, respectively. While Srt1 is strongly induced during plant colonization, Hxt1 expression is independent of the fungus–plant interaction^{33,34}.

At later stages of disease (4–8 DPI), a third wave of effectors supports tumor maturation and the production of melanized teliospores. One important regulator after establishment of the biotrophic interaction is the APSES transcription factor Nlt1. Nlt1 appears to guide tumor formation on leaf tissue but not

anthocyanin and tumor formation on the base of the stem²⁸. Another factor contributing to the regulation of effector gene expression at the later stage is the forkhead transcription factor Fox1. Fox1 is required for the attenuation of plant defense responses and for full expression of 141 genes, of which 38 encode potential effectors, as well as several proteins involved in sugar processing and transport and secondary metabolism¹⁴. Genes encoding effectors that are specifically expressed at late stages of biotrophic development are largely unexplored functionally. Potentially, they function to sustain tumor development and guide spore formation as well as the formation of a mucilaginous matrix in which spores are embedded. A key regulator of these late-stage events is the DNA-binding WOPR protein Ros1. Ros1 function is essential for teliosporogenesis, and the majority of Ros1 target genes are involved in metabolism and cellular transport. The Ros1-dependent induction of late effector genes and repression of early effector-encoding genes indicates that Ros1-mediated regulation occurs by direct and indirect mechanisms, involving additional regulators²⁰. As described below, further insights into late-stage changes in both host and pathogen gene regulation were obtained by examining transcriptomes specifically in tumors.

Transcriptome changes reveal extensive metabolic remodeling in tumors

Recent transcriptome profiling experiments of the late stage of infection, when tumors were evident, identified host genes that were upregulated (4,086) or downregulated (5,237)^{4,35}. Down-regulated functional categories included metabolic pathways for amino acids, organic acids, lipids, and photosynthesis. By contrast, upregulated functional categories were related to plant defense and lipid and carbohydrate metabolism. As expected

for proliferating maize cells during tumor formation, meristem maintenance functions were upregulated during infection. Additionally, maize transcription factors that are normally expressed in leaf tissue during plant development were downregulated in infected plants. In contrast, transcription factors normally expressed in flowers were upregulated in tumor tissue. Thus, *U. maydis* appears to inhibit the transition to adult tissue development and instead induces a flowering program. Chloroplast functions related to vegetative tissue, such as photosynthesis, lipid production, amino acid formation, and secondary metabolism, were downregulated while reproductive functions such as sucrose and starch metabolism were upregulated during infection. Maize mutants in host components related to the vegetative to flowering transition, the biogenesis of the chloroplast, or starch formation all showed altered susceptibility to *U. maydis*. For example, impaired chloroplast formation in the *why1* mutant led to increased susceptibility, while the inactive/delayed transition to a flowering state (*id1*) and the reduced starch formation leading to lower sink capacity (*su1*) led to increased plant resistance^{4,35}.

Consistent with earlier work, transcriptome profiling of tumor tissue supported an important role of sugar transporters for metabolic exploitation of the host^{4,13,33}. Hexoses and other sugars represent major carbon sources for *U. maydis*, and the fungus possesses 19 proteins with similarities to sugar/hexose-like transporters. As mentioned above, transporters such as *Hxt1* and *Srt1* play important roles during infection. *Hxt1* is constitutively expressed and has transport activity mainly for glucose but also fructose and mannose. As a high-affinity glucose transporter, *Hxt1* also possesses glucose-sensing activity and competes with the host for glucose molecules freed from sucrose by invertase. Previous work involving the deletion of the gene for this transporter or expression of a constitutive active (sensing but not transport activity) version resulted in strains with reduced or fully abolished virulence, respectively³³. As indicated, the transporter *Srt1* is induced only *in planta* and transports sucrose. Based on its extremely high affinity to sucrose (up to 200-fold versus a typical plant transporter), it outcompetes the sucrose transport functions of the host. The virulence of deletion strains for *srt1* was strongly reduced³³. Deletion of both *hxt1* and *srt1* further reduced fungal virulence, thus indicating a simultaneous uptake and use of sucrose and glucose by the pathogen during infection. Other carbon sources such as organic acids (fatty acids or carboxylic acids) may also contribute to the carbon supply of the pathogen at early time points of infection on the cuticle and at a late stage during spore formation³⁶. Monocarboxylic acids are a carbon source for *U. maydis in vitro* but can have detrimental effects on fungal longevity and reactive oxygen species resistance³⁷. *Ustilago* is known to inhibit the transition to C4 metabolism that is dependent on the dicarboxylate malate for transport between cells. However, it is still unknown how and if different host-derived carboxylic acids affect fungal virulence².

In general, the transcriptome analyses of infected tissue support the idea that control of chloroplast functions is a critical aspect of the infection of maize by *U. maydis*. Moreover, primary

and secondary metabolic functions and plant defense contributions of the chloroplast appear to be suppressed or altered during infection. In line with these observations, chloroplasts were observed to accumulate high amounts of starch, consistent with an overall increased starch content in tumor tissue^{1,4,31}. However, the mechanisms by which *U. maydis* impacts the chloroplast are still unknown.

Beyond *Ustilago maydis*: transcriptome studies with other smut fungi

A number of transcriptome studies have also been carried out for host–pathogen interactions involving other species in the Ustilaginales. For example, Que *et al.*³⁸ examined the interaction between the sugarcane smut *Sporisorium scitamineum* and its host *Saccharum officinarum*, comparing the transcriptional responses of resistant (Yacheng05-179) and susceptible (“ROC”22) plant lines. This study focused on the transcriptional responses of the host at 24, 48, and 120 hours after inoculation (HAI) in the resistant and susceptible cultivars. Both lines showed an overall increase in the number of differentially expressed genes (DEGs) as infection progressed, but the resistant cultivar exhibited an earlier response to pathogen infection. Consistently, most DEGs were associated with stress and defense responses to pathogen attack. The resistant cultivar revealed a rapid peak in DEGs at 48 hours, and more DEGs were associated with disease resistance when compared to the susceptible cultivar. The susceptible cultivar showed a similar accumulation of disease resistance-associated DEGs at the 120-hour time point, highlighting the difference in the timing of the response to infection by the resistant and susceptible lines. Categories of genes showing differential expression included metabolic functions associated with resistance, hormone signaling, flavonoid biosynthesis, cell wall fortification, and defense-related transcription. Interestingly, a total of 26 host chitinases were differentially expressed in the *S. scitamineum*-infected plants. Expression of two of the chitinases in tobacco led to increased disease resistance against *Fusarium solani* and *Botrytis cinerea*.

In a more recent transcriptome study, Wang *et al.*³⁹ investigated the interaction between the edible culm gall-forming fungus *Ustilago esculenta* and the host *Zizania latifolia*, an aquatic grass. The transcriptome study compared infections for two strains of the fungus: the teliospore-forming strain Huijiao and the non-sporulating strain Jiaobai. In general, DEGs were more frequently observed in plants infected with the Jiaobai strain versus the Huijiao strain, although the proportion of fungal transcripts was lower in the former infection. Huijiao-infected plants exhibited a peak of 57% of total transcripts of fungal origin at 10 days after swelling (DAS) compared to only ~5% for Jiaobai.

With regard to host gene expression, clear differences were observed over the course of infection with sampling every 4 days starting at 7 days prior to swelling and ending at 13 DAS. That is, for plants infected with the Jiaobai and Huijiao strains, host genes were identified that were upregulated at all three stages as well as host genes that were upregulated only at specific

time points in comparison to uninfected controls. Additionally, 170 and 205 host DEGs with potential roles in culm gall formation were identified at 1 and 10 DAS, respectively. These genes encoded proteins with predicted roles in primary and secondary metabolism as well as in plant–pathogen interactions and plant hormone signal transduction.

An analysis of pathogen genes that were upregulated was also carried out to attempt to identify functions involved in culm gall formation. Applying an analysis that paralleled that used for host genes, 78 and 109 pathogen genes were identified at 1 and 10 DAS, respectively. A significant fraction of these genes lacked annotation, while genes coding for proteins had predicted functions in nucleotide binding, transcriptional regulation, hydrolase activity, transferase activity, and transport.

In addition to *U. maydis*, other members of the Ustilaginales have been investigated as models for studying biotrophic phytopathology. For example, Rabe *et al.*⁴⁰ carried out a comprehensive study of the genome, transcriptome, and usefulness as a model system of the grass pathogen *Ustilago bromivora* and one of its hosts, *Brachypodium* spp. Initially, a susceptible line, Bd28, was identified after screening a number of *Brachypodium* spp. accessions for susceptibility to *U. bromivora*. In addition to extensive characterization of *U. bromivora* and its interaction with the host, a transcriptome analysis compared fungal gene expression of *U. bromivora* in axenic culture with infected stems of Bd28. It was found that 7.3% of the identified fungal transcripts were upregulated while 16.8% were downregulated. It is notable that these percentages are far lower than those reported by Lanver *et al.*²⁸ for differential expression of *U. maydis* genes during plant infection. For *U. bromivora*, transcripts encoding potential effectors were enriched among upregulated transcripts during infection (30.8%). In comparison, only 5.7% of transcripts (genes) overall were predicted to encode secreted proteins. Among the genes for predicted secreted proteins, 84.1% encoded proteins of unknown function compared with a frequency of 46.4% for all of the proteins encoded by *U. bromivora*. This finding is consistent with other studies indicating that most secreted effectors of plant pathogenic fungi are uncharacterized and poorly conserved between species²⁸. The observation that all of the functionally characterized effectors identified in *U. maydis* also have homologs in *U. bromivora* highlights the importance of fungal model systems to investigate molecular principles of effector function.

Conclusions and future perspectives

Overall, recent transcriptome profiling has revealed dynamic changes in the transcription of pathogen genes encoding

metabolic functions (e.g. for nitrogen and carbon metabolism) that likely reflect shifting environmental conditions throughout the disease process as *U. maydis* colonizes tissue, induces tumors, and sporulates. It is fascinating that the changes in pathogen gene expression are integrated with and driven by effector-mediated manipulation of host cell metabolism and plant immunity. Time-resolved analyses greatly increase our understanding of individual stages of pathogenic development and how transitions between those are coordinated. Future work will exploit the accumulated wealth of data to experimentally address predictions about pathogen effectors and their regulators and the importance of specific metabolic changes for both the pathogen and the host. For example, it is of keen interest to understand how different host environments are sensed and translated into adapted transcriptional programs. Also, future work will further examine the details of tumor formation. It is important to mention that these studies will benefit from technical improvements that increase the detection of less-expressed genes and reproducibility in cell type-specific transcriptome studies. Moreover, comparisons of non-tumor-forming smuts, such as *Sporisorium reilianum* and *Ustilago hordei*, with *U. maydis* may provide clues about the timing of tumor induction and the corresponding specific pathogen and host functions. The sophisticated molecular tools established for *U. maydis*, and the emerging tools for *U. bromivora*, will provide novel insights that are relevant specifically for tumor induction and sporulation by other smut fungi as well as generally for the mechanisms of biotrophy.

Abbreviations

DAS, days after swelling; DEGs, differentially expressed genes; DPI, days post inoculation; UPR, unfolded protein response

Grant information

Our research on *Ustilago maydis* is supported by a Discovery grant (to J.W.K.) and a CREATE training award (to the PRoTECT group) from the Natural Sciences and Engineering Research Council of Canada and by grants from the Deutsche Forschungsgemeinschaft (DFG) (to K.H) and the DFG-funded International Research Training Group (PRoTECT).

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Acknowledgements

We apologize to those colleagues whose work could not be cited owing to space limitations. The picture showing a maize cob infected with *U. maydis* was generously provided by Melina A. Schwier.

References



1. Doehlemann G, Wahl R, Horst RJ, *et al.*: **Reprogramming a maize plant: transcriptional and metabolic changes induced by the fungal biotroph *Ustilago maydis*.** *Plant J.* 2008; **56**(2): 181–95.
[PubMed Abstract](#) | [Publisher Full Text](#)
2. **F** Goulet KM, Saville BJ: **Carbon acquisition and metabolism changes during fungal biotrophic plant pathogenesis: insights from *Ustilago maydis*.** *Can J Plant Pathol.* 2017; **39**(3): 247–266.
[Publisher Full Text](#) | [F1000 Recommendation](#)
3. **F** Lanver D, Tollot M, Schweizer G, *et al.*: ***Ustilago maydis* effectors and their impact on virulence.** *Nat Rev Microbiol.* 2017; **15**(7): 409–21.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
4. Kretschmer M, Croll D, Kronstad JW: **Maize susceptibility to *Ustilago maydis* is influenced by genetic and chemical perturbation of carbohydrate allocation.** *Mol Plant Pathol.* 2017; **18**(9): 1222–37.
[PubMed Abstract](#) | [Publisher Full Text](#)
5. Brefort T, Doehlemann G, Mendoza-Mendoza A, *et al.*: ***Ustilago maydis* as a pathogen.** *Annu Rev Phytopathol.* 2009; **47**: 423–45.
[PubMed Abstract](#) | [Publisher Full Text](#)
6. Heibel K, Scherer M, Vranes M, *et al.*: **The transcription factor Rbf1 is the master regulator for *b*-mating type controlled pathogenic development in *Ustilago maydis*.** *PLoS Pathog.* 2010; **6**(8): e1001035.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
7. Lanver D, Berndt P, Tollot M, *et al.*: **Plant surface cues prime *Ustilago maydis* for biotrophic development.** *PLoS Pathog.* 2014; **10**(7): e1004272.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
8. **F** Ma LS, Wang L, Trippel C, *et al.*: **The *Ustilago maydis* repetitive effector Rsp3 blocks the antifungal activity of mannose-binding maize proteins.** *Nat Commun.* 2018; **9**(1): 1711.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
9. Eichhorn H, Lessing F, Winterberg B, *et al.*: **A ferroxidation/permeation iron uptake system is required for virulence in *Ustilago maydis*.** *Plant Cell.* 2006; **18**(11): 3332–45.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
10. Ho EC, Cahill MJ, Saville BJ: **Gene discovery and transcript analyses in the corn smut pathogen *Ustilago maydis*: expressed sequence tag and genome sequence comparison.** *BMC Genomics.* 2007; **8**: 334.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
11. Horst RJ, Doehlemann G, Wahl R, *et al.*: ***Ustilago maydis* infection strongly alters organic nitrogen allocation in maize and stimulates productivity of systemic source leaves.** *Plant Physiol.* 2010; **152**(1): 293–308.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
12. **F** Skibbe DS, Doehlemann G, Fernandes J, *et al.*: **Maize tumors caused by *Ustilago maydis* require organ-specific genes in host and pathogen.** *Science.* 2010; **328**(5974): 89–92.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
13. **F** Wahl R, Zehri A, Kämper J: **The *Ustilago maydis* *b* mating type locus controls hyphal proliferation and expression of secreted virulence factors in planta.** *Mol Microbiol.* 2010; **75**(1): 208–20.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
14. Zehri A, Heibel K, Wahl R, *et al.*: **The *Ustilago maydis* forkhead transcription factor Fox1 is involved in the regulation of genes required for the attenuation of plant defenses during pathogenic development.** *Mol Plant Microbe Interact.* 2010; **23**(9): 1118–29.
[PubMed Abstract](#) | [Publisher Full Text](#)
15. **F** Jonkers W, Rodriguez Estrada AE, Lee K, *et al.*: **Metabolome and transcriptome of the interaction between *Ustilago maydis* and *Fusarium verticillioides* in vitro.** *Appl Environ Microbiol.* 2012; **78**(10): 3656–67.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
16. Morrison EN, Donaldson ME, Saville BJ: **Identification and analysis of genes expressed in the *Ustilago maydis* dikaryon: uncovering a novel class of pathogenesis genes.** *Can J Plant Pathol.* 2012; **34**(3): 417–35.
[Publisher Full Text](#)
17. Martínez-Soto D, Ruiz-Herrera J: **Transcriptomic analysis of the dimorphic transition of *Ustilago maydis* induced in vitro by a change in pH.** *Fungal Genet Biol.* 2013; **58–59**: 116–25.
[PubMed Abstract](#) | [Publisher Full Text](#)
18. Franco-Frías E, Ruiz-Herrera J, Aréchiga-Carvajal ET: **Transcriptomic analysis of the role of Rim101/PacC in the adaptation of *Ustilago maydis* to an alkaline environment.** *Microbiology.* 2014; **160**(Pt 9): 1985–98.
[PubMed Abstract](#) | [Publisher Full Text](#)
19. Islamovic E, García-Pedrajas MD, Chacko N, *et al.*: **Transcriptome Analysis of a *Ustilago maydis* *ust1* Deletion Mutant Uncovers Involvement of Laccase and Polyketide Synthase Genes in Spore Development.** *Mol Plant Microbe Interact.* 2015; **28**(1): 42–54.
[PubMed Abstract](#) | [Publisher Full Text](#)
20. **F** Tollot M, Assmann D, Becker C, *et al.*: **The WOPR Protein Ros1 is a Master Regulator of Sporogenesis and Late Effector Gene Expression in the Maize Pathogen *Ustilago maydis*.** *PLoS Pathog.* 2016; **12**(6): e1005697.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
21. León-Ramírez CG, Cabrera-Ponce JL, Martínez-Soto D, *et al.*: **Transcriptomic analysis of basidiocarp development in *Ustilago maydis* (DC) Cda.** *Fungal Genet Biol.* 2017; **101**: 34–45.
[PubMed Abstract](#) | [Publisher Full Text](#)
22. **F** Donaldson ME, Ostrowski LA, Goulet KM, *et al.*: **Transcriptome analysis of smut fungi reveals widespread intergenic transcription and conserved antisense transcript expression.** *BMC Genomics.* 2017; **18**(1): 340.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
23. Sánchez-Arreguín JA, Hernández-Oñate MA, León-Ramírez CG, *et al.*: **Transcriptional analysis of the adaptation of *Ustilago maydis* during growth under nitrogen fixation conditions.** *J Basic Microbiol.* 2017; **57**(7): 597–604.
[PubMed Abstract](#) | [Publisher Full Text](#)
24. Flor-Parra I, Vranes M, Kämper J, *et al.*: **Biz1, a zinc finger protein required for plant invasion by *Ustilago maydis*, regulates the levels of a mitotic cyclin.** *Plant Cell.* 2006; **18**(9): 2369–87.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
25. Heibel K, Scherer M, Schuler D, *et al.*: **The *Ustilago maydis* Clp1 protein orchestrates pheromone and *b*-dependent signaling pathways to coordinate the cell cycle and pathogenic development.** *Plant Cell.* 2010; **22**(8): 2908–22.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
26. Lanver D, Mendoza-Mendoza A, Brachmann A, *et al.*: **Sho1 and Msb2-related proteins regulate appressorium development in the smut fungus *Ustilago maydis*.** *Plant Cell.* 2010; **22**(6): 2085–101.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
27. Heibel K, Freitag J, Hampel M, *et al.*: **Crosstalk between the unfolded protein response and pathways that regulate pathogenic development in *Ustilago maydis*.** *Plant Cell.* 2013; **25**(10): 4262–77.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
28. **F** Lanver D, Müller AN, Happel P, *et al.*: **The Biotrophic Development of *Ustilago maydis* Studied by RNA-Seq Analysis.** *Plant Cell.* 2018; **30**(2): 300–23.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
29. Schilling L, Matei A, Redkar A, *et al.*: **Virulence of the maize smut *Ustilago maydis* is shaped by organ-specific effectors.** *Mol Plant Pathol.* 2014; **15**(8): 780–9.
[PubMed Abstract](#) | [Publisher Full Text](#)
30. Redkar A, Hoser R, Schilling L, *et al.*: **A Secreted Effector Protein of *Ustilago maydis* Guides Maize Leaf Cells to Form Tumors.** *Plant Cell.* 2015; **27**(4): 1332–51.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
31. **F** Matei A, Ernst C, Günl M, *et al.*: **How to make a tumour: Cell type specific dissection of *Ustilago maydis*-induced tumour development in maize leaves.** *New Phytol.* 2018; **217**(4): 1681–95.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
32. **F** Paul JA, Wallen RM, Zhao C, *et al.*: **Coordinate regulation of *Ustilago maydis* ammonium transporters and genes involved in mating and pathogenicity.** *Fungal Biol.* 2018; **122**(7): 639–50.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
33. Schuler D, Wahl R, Wippel K, *et al.*: **Hxt1, a monosaccharide transporter and sensor required for virulence of the maize pathogen *Ustilago maydis*.** *New Phytol.* 2015; **206**(3): 1086–100.
[PubMed Abstract](#) | [Publisher Full Text](#)
34. **F** Wahl R, Wippel K, Goos S, *et al.*: **A novel high-affinity sucrose transporter is required for virulence of the plant pathogen *Ustilago maydis*.** *PLoS Biol.* 2010; **8**(2): e1000303.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)
35. Kretschmer M, Croll D, Kronstad JW: **Chloroplast-associated metabolic functions influence the susceptibility of maize to *Ustilago maydis*.** *Mol Plant Pathol.* 2017; **18**(9): 1210–21.
[PubMed Abstract](#) | [Publisher Full Text](#)
36. Kretschmer M, Klose J, Kronstad JW: **Defects in mitochondrial and peroxisomal β -oxidation influence virulence in the maize pathogen *Ustilago maydis*.** *Eukaryotic Cell.* 2012; **11**(8): 1055–66.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
37. Kretschmer M, Lambie S, Croll D, *et al.*: **Acetate provokes mitochondrial stress and cell death in *Ustilago maydis*.** *Mol Microbiol.* 2018; **107**(4): 488–507.
[PubMed Abstract](#) | [Publisher Full Text](#)
38. Que Y, Su Y, Guo J, *et al.*: **A global view of transcriptome dynamics during *Sporisorium scitamineum* challenge in sugarcane by RNA-Seq.** *PLoS One.* 2014; **9**(8): e106476.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
39. **F** Wang ZD, Yan N, Wang ZH, *et al.*: **RNA-seq analysis provides insight into reprogramming of culm development in *Zizania latifolia* induced by *Ustilago esculenta*.** *Plant Mol Biol.* 2017; **95**(6): 533–47.
[PubMed Abstract](#) | [Publisher Full Text](#) | [F1000 Recommendation](#)
40. **F** Rabe F, Bosch J, Stirnberg A, *et al.*: **A complete toolset for the study of *Ustilago bromivora* and *Brachypodium* sp. as a fungal-temperate grass pathosystem.** *eLife.* 2016; **5**: pii: e20522.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#) | [F1000 Recommendation](#)

Open Peer Review

Current Peer Review Status: 

Editorial Note on the Review Process

F1000 Faculty Reviews are written by members of the prestigious F1000 Faculty. They are commissioned and are peer reviewed before publication to ensure that the final, published version is comprehensive and accessible. The reviewers who approved the final version are listed with their names and affiliations.

The reviewers who approved this article are:

Version 1

- José Ruiz-Herrera**
Departamento de Ingeniería Genética, Centro de Investigación y de Estudios Avanzados del Instituto Politécnico Nacional, Irapuato, Mexico
Competing Interests: No competing interests were disclosed.
- Barry Saville**
Forensic Science Program, Trent University, Peterborough, ON, Canada
Competing Interests: No competing interests were disclosed.
- Armin Djamei**
Vienna Biocenter (VBC), Gregor Mendel Institute (GMI), Austrian Academy of Sciences (OEAW), Vienna, Austria
Competing Interests: No competing interests were disclosed.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com

F1000Research