International Journal of Travel Medicine & Global Health

Original Article

The Susceptibility Evaluation of Multiresistant Gram-Negative Bacilli to Meropenem and Imipenem

Nematollah Jonidi Jafari¹, Morteza Izadi¹, Massoud Hajia², Mahdi Qorbanalizadgan³, Amin Saburi^{*4}

Abstract

Introduction: Nosocomial infections are responsible for the much of the morbidity and mortality found in hospitals. The present study was conducted on 70 bacterial strains isolated from hospitalized patients in various medical units of Baqiyatallah Hospital in Tehran, Iran during a period of 12 months from; March to February 2009.

Methods: The bacterial sensitivity for meropenem and imipenem was evaluated using the E-test and explanations of the MIC values. All patients were included in this study that had been hospitalized with no signs and symptoms of infection within the first 48 hours of hospitalization and began presenting signs and symptoms of infection after 48 hours of hospitalization.

Results: Resistance to meropenem and imipenem was confirmed with E-test (AB Biodisk, Sweden) and disc diffusion methods. Meropenem and imipenem were active against 61 (64.2%) and 62(65.2%) strains, respectively, of the 95 ESBL positive strains.

Conclusion: The activity of meropenem or imipenem against gram negative ESBL-positive bacilli is decreasing rapidly but even so these antibiotics are effective against nosocomial multiresistant organisms.

Keywords:	Gram-Negative	Rods, Imipenem	Meropenem.	Susceptibility
	Orann riegaurie	recous, imperior	,	Sabeeptionity

Introduction

Secondary infections due to Gram-negative bacilli continue to be one of the leading causes of mortality and morbidity. Resistance to beta-lactam antibiotics has increased remarkably in the last two decades and has been documented in both community and hospital settings [1-3]. During the past decade, Gram-negative bacteria have extended resistance too many antibiotics, including quinolones, aminoglycosides and β -lactams. Some gram negative rods, such as: some strains of Proteus mirabilis, Escherichia coli and Klebsiella spp. are known to make extended spectrum β -lactamases (ESBLs) or stably derepressed AmpC Beta -lactamases (AmpC) resulting in their wide resistance to the monobactams and third generation cephalosporins [4,5].

Meropenem and Imipenem are widely used against Betalactamase positive bacteria. They are active against most clinically important gram-positive and gram-negative bacteria, including anaerobic and aerobic forms. Nevertheless, reckless use of these antimicrobials has increased Carbapenem resistance among nosocomial pathogens [6]. Antimicrobial therapy of strains producing extendedspectrum β -lactamases and inducible β -lactamases (IBL) has become restricted and the progression of resistance to carbapenems makes the problem more acute [7,8]. The aim of this investigation was to assess the in vitro activity of meropenem and imipenem against Gram-negative bacilli isolated from hospitalized patients in various medical units of Baqiyatallah hospital in Tehran, Iran over 1 year, and Health Research center, Baqiyatallah University of Medical Sciences, Tehran, Iran.
Research Center of references laboratories of Iran, Tehran, Iran.
Research Center of Molecular Biology, Baqiyatallah University of Medical Sciences,

.

Tehran, Iran. 4. Chemical Injuries Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran.

* Corresponding Author

Amin Saburi, Chemical Injuries Research center, Baqiyatallah University of Medical Sciences, Teharn, Iran.

Email: Aminsaburi@yahoo.com

Received: 8 December 2013 Accepted: 3 February 2014

resistance to meropenem and imipenem was confirmed with disc diffusion and E-test (AB Biodisk, Sweden) methods.

Methods

1. Strain collection

The study was performed during a period of 12 months from March to February 2009 Baqiyatallah hospital in Tehran, Iran. All patients with no symptoms and signs of infection before the first 48 hours of hospitalization and presenting signs and symptoms of infection after 48 hours of hospitalization (nosocomial infection) were included. Finally, 350 specimens were collected via blood samples from peripheral veins.

2. The determination of ESBL

The testing protocols were ratified in accordance to the guidelines of the National Committee for Clinical Laboratory Standards.[9] The ESBL study used two different E test strips on Mueller Hinton Agar (MHA): cefotaxime/cefotaxime with clavulanic acid or ceftazidime/ ceftazidime with clavulanate.[10] The recommendations of the January 2003 NCCLS guide was used as the criterion for ESBL-positivity [9]. The specimens were concluded to be ESBL-positive if the addition of clavulanic acid diminished the MIC of either of the β -lactam agents by three-fold or more. E. coli ATCC 25922, P. aeruginosa ATCC 27853 and K. pneumoniae BCC 1395 were used as control strains.

International Journal of Travel Medicine and Global Health. Winter 2014; Volume 2, Issue 1:1-3



Table 1. Imipenem and meropenem susceptibility of ESBL								
	Total			ESBL positive				
		Imipenem			Meropenem			
Susceptibility		S	R	S	R			
Strains		n	n	n	n			
A. baumannii	45	26	19	28	17			
P. aeruginosa	28	16	12	11	17			
Klebsiella spp	13	11	2	13	0			
E. coli	4	4	0	4	0			
Enterobacter spp.	5	5	0	5	0			
Total	95	62	33	61	34			

S, susceptible_4 mg/l; R, Resistant_16 mg

3. In vitro assessment of ESBL strains for meropenem and imipenem

The susceptibilities to meropenem and imipenem of the 95 ESBL positive specimens were definite using the E-test and explanations of the MIC values in mg/l were made from the NCCLS document [9]. The limit values of MIC were regarded as: more than 16 mg/l; resistant, 8 mg/l; intermediate and less than 4 mg/l; unsusceptible. Resistance to meropenem and imipenem was confirmed with disc diffusion and E-test (AB Biodisk, Sweden) methods.

Results

Ninety five of 350 strains were confirmed to be resistant to β -lactam antibiotics. The frequency of microbial agents in this investigation was stated as; Escherichia coli 4(4.2%), Klebsiella pneumoniae 13(13.6%), Enterobacter spp. 5(5.2%), Pseudomonas aeruginosa 28 (29%), Acinetobacter baumannii 45 (47%).

In our study, meropenem and imipenem were active against 61 (64.2%) and 62(65.2%) strains, respectively, of the 95 ESBL positive strains. In Acinetobacter baumannii, where there was the highest incidence of ESBL production, imipenem and meropenem were effective against 84.6% and 100% of strains, respectively. Activity of imipenem against P. aeruginosa and Klebsiella spp. strains was 57.1% and 84.6% but meropenem activity against these strains was 39.2 and 100% respectively. Imipenem and meropenem were completely active against E. coli and Enterobacter spp. Other results are shown in Table 1.

Discussion

In this study, meropenem and imipenem were active against 61 (64.2%) and 62(65.2%) strains of the ESBL positive strains, respectively. Imipenem was minutely more effective against all ESBL positive strains than meropenem. In the investigations conducted by Garau [7] and Colardyn [8], imipenem and meropenem were effective against 74.3, 74 and 81.7, 75%, respectively [7,8].

The effectiveness of both carbapenems in intra-abdominal surgical patients in three studies on ICU strains was more than 94% but this is in disagreement with our findings [11-13]. The rate of resistance to imipenem was 8% in a Polish study and 13% in a Belgian study, and this is in disagreement with our results [14-15]. In a similar study, Zanetti et al., found meropenem and imipenem to be 87.1% and 92% effective [16]. However, in a study in Turkey, resistance to imipenem (8.4–33.4%) was lower than that of our investigation [17]. Two studies results showed car-

bapenems (imipenem and meropenem) to be more potent in vitro than any other drug against the Enterobacteriaceae [18-19]. Similar studies have explained the efficacy of meropenem and imipenem to be 100 and 95.4 % for ESBL producing strains and 94.9 and 96.9 % for strains producing undefined-lactamases [20-21]. In Iaconis study, 13% of strains were resistant to imipenem and 4% to meropenem [22]. Imipenem resistance was found in two strains of K. pneumoniae, but meropenem resistance was found in none due to ESBLs. Meropenem and Imipenem are absolutely resistant to β -lactamase enzymes of Gram-negative bacteria and acquired resistances due to carbapenemases are scarce in these bacteria [23-25]. The difference of our study with other similar studies is that we screen tested to select ESBL strains of specimens and then a susceptibility test was performed.

Durmaz et al. In a lesser but similar study found no meropenem resistance in 22 Gram-negative strains with ESBL positivity and imipenem resistance existed only in one K. pneumoniae strain [20]. In Iran, Hadadi et al described the resistance pattern of Gram-negative bacteria (but no ESBL) to imipenem. Except E.Coli, imipenem resistance was increased in our study versus the Hadadi study [26]. Hawser's study demonstrated that the most active agents against ESBL-positive K. pneumoniae were imipenem, with susceptibility percentages of 89.5% .[27] In Iran, as a developing country, multidrug resistance isolates are commonly reportedly due to antibiotics abuse [28]. The results explain carbapenems (imipenem and meropenem) being more effective in vitro than any other drug against Gram-negative bacilli.

Conclusion

The prevalence of antibiotic resistance in Gram-negative rods is high in Asia. The activity of meropenem or imipenem against gram negative ESBL-positive bacilli is decreasing rapidly but these antibiotics are continuing to show effectiveness against nosocomial multiresistant organisms. This study's results can be used in preparing evidence-based guidelines for antibiotic therapy, especially empirical treatment of nosocomial bacterial infections.

References

1. Nijssen S, Florijn A, Bonten MJ, Schmitz FJ, Verhoef J, Fluit AC. Beta-lactam susceptibilities and prevalence of ESBLproducing isolates among more than 5000 European Enterobacteriaceae isolates. Int J Antimicrob Agents. 2004;24(6):585–91. PubMed PMID: 15555882 2. Bush K. New h-lactamases in gram-negative bacteria: diversity and impact on the selection of antimicrobial therapy. Clin Infect Dis. 2001;32(7):1085–9. PubMed PMID: 11264037

3. Weber DJ, Raasch R, Rutala WA. Nosocomial infections in the ICU. The growing importance of antibiotic-resistance pathogens. Chest. 1999;115(3 Suppl):34S–41S. PubMed PMID: 10084458

4. Patzer JA, Dzier zanowska D, Alicja Pawi nska A, Turner PJ. High activity of meropenem against Gram-negative bacteria from a paediatric Intensive Care Unit, 2001–2005. Int J Antimicrob Agents. 2007;29(3):285–8. PubMed PMID: 17257814

5. Jones RN, Kehrberg EN, Erwin ME, Anderson SC. Prevalence of important pathogens and antimicrobial activity of parenteral drugs at numerous medical centers in the United States, I. Study on the threat of emerging resistances: real or perceived? Fluoroquinolone resistance surveillance group. Diagn Microbiol Infect Dis.1994;19(4):203–15. PubMed PMID: 7851083

6. Bekir S. Kocazeybek a, et al. Short communication Use of Etests with carbapenems for Gram-negative rods producing β -lactamases, International Journal of Antimicrobial Agents 19 (2002) 159–162.

7. Garau J, Blanquer J, Cobo L, Corcia S, Daquerre M, de Latorre FJ, et al. Prospective, randomised, multicentre study of meropenem versus imipenem/cilastatin as ampiric monotherapy in severe nosocomial infections. Eur J Clin Microbiol Infect Dis 1997;16(11):789–96. PubMed PMID: 9447899

8. Colardyn F, Faulkner KL. Intravenous meropenem versus imipenem/cilastatin in the treatment of serious bacterial infections in hospitalized patients. Meropenem serious infection study group. J Antimicrob Chemother. 1996;38(3):523–37. PubMed PMID: 8889726

9. National Committee for Clinical Laboratory Standard (2003) Performance Standard for Antimicrobial Susceptibility Testing. 12th Information Supplement, M100-S14. Wayne (PA): National Committee for Clinical Laboratory Standard; 2003.

10. Bradford PA. Extended-spectrum beta-lactamases in the 21st century: characterization, epidemiology, and detection of this important resistance threat. Clin Microbiol Rev. 2001;14(4): 933–51. PubMed PMID: 11585791

11. Kanellakopoulou K, Giamarellou H, Papadothomakos P, Tsipras H, Chloroyiannis J, Theakou R, et al. Meropenem versus imipenem/cilastatin in the treatment of intraabdominal infections requiring surgery. Eur J Clin Microbiol Infect Dis. 1993;12(6):449–53. PubMed PMID: 8859165

12. Brismar B, Malmborg AS, Tunevall G, Lindgren V, Bregman L, Mentzing LO, et al. Meropenem versus imipenem/cilastatin in the treatment of intra-abdominal infections. J Antimicrob Chemother 1995;35(1):139–48. PubMed PMID: 7768761

13. Geroulanos SJ. Meropenem versus imipenem/cilastatin in intra-abdominal infections requiring surgery. Meropenem study group. J Antimicrob Chemother. 1995;36(Suppl A):191–205. PubMed PMID: 5843495

14. Glupczynski Y, Delmee M, Goossens H, Struelens M, Belgian Multicenter ICU Study Group. Distribution and prevalence of antimicrobial resistance among gram-negative isolates in intensive care units (ICU) in Belgian hospitals between 1996 and 1999. Acta Clin Belg. 2001;56(5):297–306. PubMed PMID: 11770225

15. Patzer J, Dzierzanowska D, Turner P. Susceptibility patterns of Gramnegative bacteria from a Polish intensive care unit, 1997–2000. Int J Antimicrob Agents. 2002;19(5):431–4. PubMed PMID: 12007852

16. Zanetti G, Harbarth SJ, Trampuz A, Ganeo M, Mosimann F, Chautemps R, et al. Meropenem (1.5 g/day) is as effective as imipenem/cilastatin (2 g/day) for the treatment of moderately severe intra-abdominal infections. Int J Antimicrob Agents. 1999;11(2):107–13. PubMed PMID: 10221413

17. Kucukates E. Antimicrobial resistance among Gram-negative bacteria isolated from intensive care units in a Cardiology Institute in Istanbul Turkey. Jpn J Infect Dis. 2005;58(4):228–31. PubMed PMID: 16116256

18. Koseoglu O, Kocagoz S, Gur D, Akova M. Nosocomial bloodstream infections in a Turkish university hospital: study of Gram-negative bacilli and their sensitivity patterns. Int J Antimicrob Agents. 2006;17(6):477–81. PubMed PMID: 11397618

19.Tan TY, Hos LY, Koh TH, Ng LS, Tee NW, Krishnan P, et al.Antibiotic Resistance in Gram-negative Bacilli: a Singapore perspective. Ann Acad Med Singapore. 2008; 37(10):819-25. PubMed PMID: 19037514

20. Durmaz G, Aydinli A, Yildiz U, Akgu'n Y. The effect of meropenem and imipenem against Gram negative bacteria which are resistant to aminoglycoside and are positive for wide spectrum beta lactamases. Infection. 1997;11:19–22.

21. Koc, NA, Evrensel N, Koc, RK. The effects of meropenem and imipenem on Gram negative bacillus isolated from the intensive care units. Infection. 1997;11:119–21.

22. Iaconis JP, Pitkin DH, Sheikh W, Nadler HL. Comparison of antibacterial activities of meropenem and six other antimicrobials against Pseudomonas aeruginosa isolates from North American studies and clinical trials. Clin Infect Dis. 1997;24(Suppl 2):S191–6. PubMed PMID: 9126693

23. Livermore DM. Acquired carbapenemases. J Antimicrob Chemother 1997; 39(6):673-6. PubMed PMID: 9222034

24. Chen HY, Yuan M, Livermore DM. Mechanisms of resistance to beta-lactam antibiotics amongst Pseudomonas aeruginosa isolates collected in UK in 1993. J Med Microbiol. 1995;43(4):300–9. PubMed PMID: 7562993

25. Livermore DM. Bacterial resistance to carbapenems. In: Jungkind DL, editor. Antimicrobial resistance: a crisis in health care. New York: Plenum Press, 1995:25.

26.Hadadi A1, Rasoulinejad M, Maleki Z, Yonesian M, Shirani A, Kourorian Z. Antimicrobial resistance pattern of Gramnegative bacilli of nosocomial origin at 2 university hospitals in Iran.Diagn Microbiol Infect Dis. 2008;60(3):301–5. PubMed PMID: 18036759

27. Hawser S, Hoban D, Bouchillon S, Badal R. Antimicrobial susceptibility of intra-abdominal Gram-negative bacilli from Europe: SMART Europe 2008. Eur J Clin Microbiol Infect Dis, published online 16 oct 2010.

28.Owlia P, Azimi L, Gholami A, Asghari B, Lari AR. ESBLand MBL-mediated resistance in Acinetobacter baumannii: a global threat to burn patients. Infez Med. 2012;20(3):182-7. PubMed PMID: 22992558