



# High Genetic Similarity of MRSA ST88 Isolated From Pigs and Humans in Kogi State, Nigeria

Otalu Jnr Otalu<sup>1</sup>, Jacob K. P. Kwaga<sup>2</sup>, Emmanuel Chukuwdi Okolocha<sup>2</sup>, Md Zohorul Islam<sup>3</sup> and Arshnee Moodley<sup>3\*</sup>

<sup>1</sup> Department of Microbiology, Faculty of Natural Sciences, Kogi State University, Anyigba, Nigeria, <sup>2</sup> Department of Public Health and Preventive Medicine, Faculty of Veterinary Medicine, Ahmadu Bello University, Zaria, Nigeria, <sup>3</sup> Department of Veterinary and Animal Sciences, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark

## OPEN ACCESS

### Edited by:

Adebayo Osagje Shittu,  
Obafemi Awolowo University, Nigeria

### Reviewed by:

Samuel Sunday Taiwo,  
Ladoke Akintola University of  
Technology, Nigeria  
Alaa Abouelfetouh,  
Alexandria University, Egypt

### \*Correspondence:

Arshnee Moodley  
asm@sund.ku.dk

### Specialty section:

This article was submitted to  
Infectious Diseases,  
a section of the journal  
Frontiers in Microbiology

**Received:** 11 June 2018

**Accepted:** 30 November 2018

**Published:** 17 December 2018

### Citation:

Otalu OJ, Kwaga JKP, Okolocha EC,  
Islam MZ and Moodley A (2018) High  
Genetic Similarity of MRSA ST88  
Isolated From Pigs and Humans in  
Kogi State, Nigeria.  
Front. Microbiol. 9:3098.  
doi: 10.3389/fmicb.2018.03098

We determined the prevalence and genetic characteristics of methicillin-resistant *Staphylococcus aureus* (MRSA) isolated from pigs and humans between September 2013 and February 2015 in Kogi State, a central region in Nigeria. A total of 680 nasal swabs were collected and analyzed from pigs ( $n = 425$ ) and “pig-contact” humans ( $n = 55$ ) on 35 farms, and “non-pig-contact” humans ( $n = 200$ ). MRSA was recovered from 20 (4.7%) pigs on 12 farms and 18 (7.0%) humans. Six (2.4%) of the human isolates were recovered from “pig-contact” humans, of which only three work on farms also harboring MRSA positive pigs. All 38 MRSA were resistant to  $\beta$ -lactams only, belonged to *spa* type t1603, sequence type (ST) 88, and *mecA* was associated with a SCC*mec* IVa element. Four isolates from a pig, a pig-contact human from the same farm, a pig-contact human from a pig farm in a different district, and a non-pig-contact human were subjected to whole genome sequencing (WGS). Core genome SNP analysis revealed high genetic similarity between strains (3–11 SNP differences), despite the temporal (2 year gap) and geographic (165 km) differences between isolates. Furthermore, these Nigerian isolates form a distinct clade when compared to other African MRSA ST88 isolates. All but one porcine strain was positive for *scn* suggesting a possible human origin and that pigs were either transiently contaminated by humans or result of a very recent human-to-pig transmission event. To our knowledge, this is the first report of genetically confirmed MRSA in pigs in Nigeria, which appear to be a typical CA-MRSA clone present in the human population.

**Keywords:** CA-MRSA, porcine, ST88, staphylococci, zoonotic

## INTRODUCTION

Methicillin-resistant *Staphylococcus aureus* (MRSA) is of great concern in both human and veterinary medicine (Vanderhaeghen et al., 2010). Such concerns are related to difficulty in treating infections, prolonged hospitalization and increased health care costs. Pigs can be asymptomatic carriers of *S. aureus* including methicillin-resistant strains, although infections with *S. aureus* are infrequent (van der Wolf et al., 2012). The central concern with MRSA in pigs is the risk of spread to humans (Unnerstad et al., 2017).

MRSA sequence type (ST) 398 is the predominant clone observed in pigs from Europe and North America, whereas MRSA ST9 is predominant in Asia. In Africa, there are only four reports of MRSA in pigs; two in South Africa (Adegoke and Okoh, 2014; Van Lochem et al., 2018), one in Senegal (Fall et al., 2012), and one in Nigeria (Okunlola and Ayandele, 2015). Unfortunately, only two of the four studies used molecular methods to confirm the *S. aureus* species identification and the presence of *mecA* in oxacillin or ceftiofur resistant strains, and only one study further typed the MRSA strains. Fall et al. (2012) reported that their porcine MRSA belonged mainly to ST5 and a singleton ST88, which are both major MRSA lineages responsible for human infections in Senegal (Breurec et al., 2011). Pigs have been shown to harbor human-associated MRSA clones e.g., USA300 (Arriola et al., 2011; Baez et al., 2017). People in close contact with pigs (farmers, veterinarians, transporters, and slaughterhouse workers) have been shown to be at a greater risk of being colonized with livestock associated MRSA (LA-MRSA) (Denis et al., 2009; Liu et al., 2015). However, LA-MRSA has been reported in people with no known contact to pigs or pig farms (Larsen et al., 2017). In Nigeria, there are no published data regarding colonization of pigs with MRSA and possible transmission of strains between pigs and humans. Our objective was to evaluate the occurrence of MRSA in pigs, and compare the strains to those isolated from pig-contact and non-contact humans.

## MATERIALS AND METHODS

### Ethical Statement

The study and protocol were approved by the institutional review board of Kogi State University Teaching Hospital. Informed oral consent was obtained from each participant and pig farm owners consented to the sampling of their animals.

### Sample Collection

The study was conducted between September 2013 and February 2015 and covered 16/21 local government areas of Kogi State (Figure 1). Samples were collected from 35 pig farms housing between 28 and 275 pigs per farm. Between 10 and 15 pigs were randomly sampled per farm. When pigs were housed in pens (i.e., on 5/35 farms), two pigs per pen were sampled. On those farms not housing pigs in pens, pigs were randomly selected. Human samples were collected from 55 pig farms workers (pig contact humans). The number of farm workers per farm ranged from 1 to 6 individuals (mode = 1). Two hundred students attending Kogi State University in Anyigba that had no pig contact were also sampled (non-pig-contact humans). All human participants were provided with a standardized questionnaire to collect demographic data, medical history over the previous 12 months e.g., hospitalization and antimicrobial therapy, and information about contact with pigs.

### Isolation and Characterization of MRSA

Nasal swabs were collected with a sterile cotton-tip swab inserting into both anterior nares of each pig and human and were placed into 3 ml Mueller Hinton broth (Oxoid, UK) supplemented

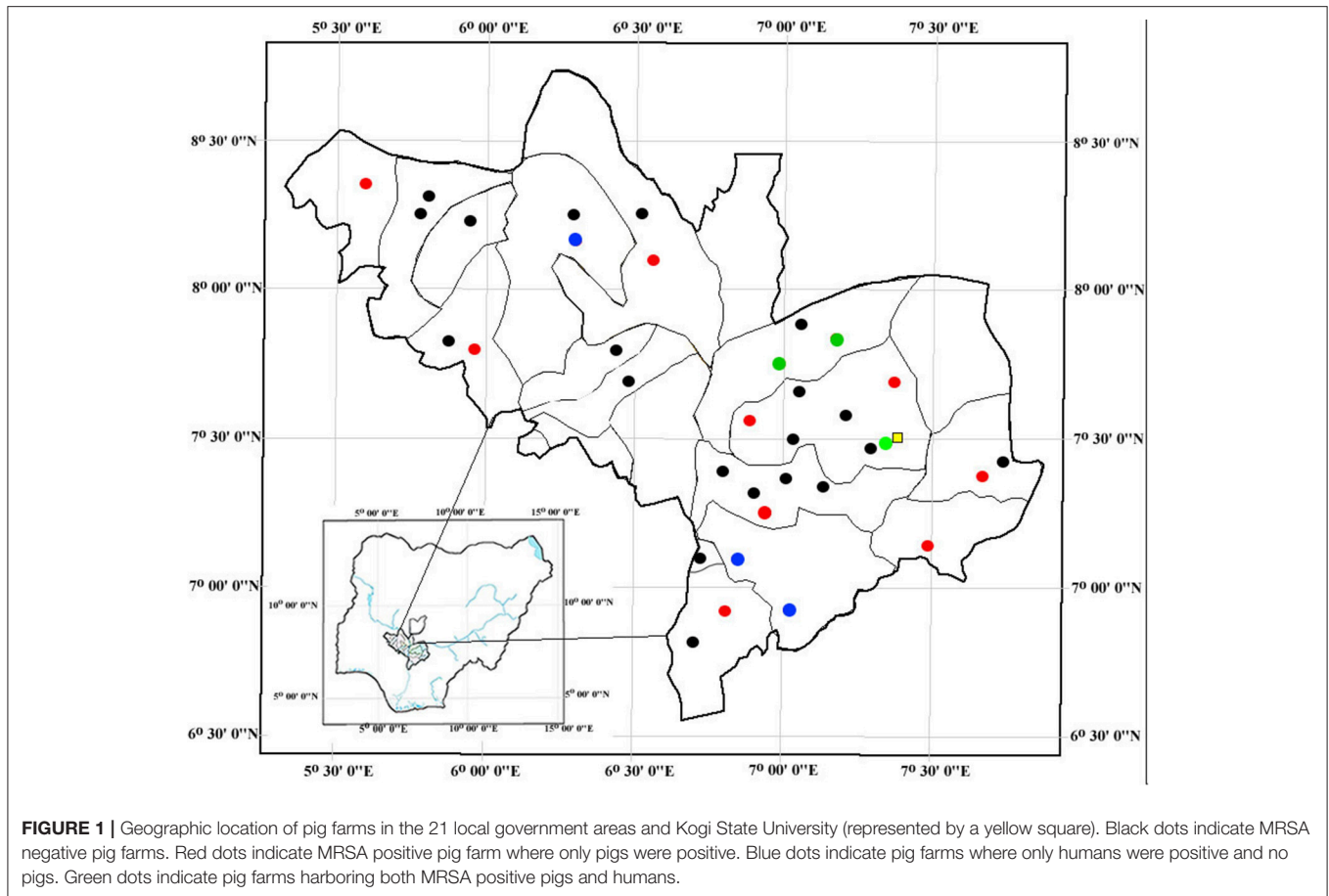
with 6.5% sodium chloride and incubated at 37°C for 24h. A loopful of the inoculum from the enrichment broth was streaked on Brilliance agar (Oxoid, UK). After incubation at 37°C for 24h, suspected MRSA colonies (denim blue colonies) were sub-cultured on 5% sheep blood agar and incubated at 37°C for 24h. Isolates were speciated by matrix-assisted laser desorption/ionization time-of-flight mass spectrophotometry (MALDI-TOF MS) (BioMérieux, France). Confirmed *S. aureus* isolates were stored at -20°C for further analysis. Antimicrobial susceptibility testing was done using the Sensititre system and COMPANIF MIC plate (ThermoFisher Scientific, USA), and susceptibility was interpreted according to CLSI (Clinical and Laboratory Standards Institute (CLSI), 2014). MRSA were characterized by a multiplex PCR to detect *spa*, *mecA*, *lukF-PV* (a marker of the Panton-Valentine leucocidin, PVL), *scn* and a CC398 specific band (Islam et al., 2017). Direct sequencing of the purified PCR reaction was used for *spa* typing (Harmsen et al., 2003). Four isolates (one pig, one pig-contact human from the same pig farm, one non-pig-contact human from the same district, and one pig-contact human from a different district) were selected for further SCC*mec* (Kondo et al., 2007), multi locus sequencing typing (Enright et al., 2000), and whole genome sequencing (WGS).

### Whole-Genome Sequencing

Overnight cultures were grown in tryptic soy broth at 37°C with 200-rpm shaking. Genomic DNA was extracted from cultures by using the automated Maxwell DNA instrument using the Maxwell® RSC Cultured Cells DNA Kit (Promega, United Kingdom). Library preparation was carried out using the Nextera XT kit and paired end 2 × 250 bp sequencing on the MiSeq, all following standard Illumina protocols (Illumina, Inc., United States). All raw reads have been deposited in the Sequence Read Archive database in the European Nucleotide Archive under the study accession number PRJEB26533.

### Phylogenetics and Comparative Genomics

All analyses were performed in CLC Genomics Workbench v. 11.0 using the tools within the Microbial Genomics Module. First, Kmer analysis was performed against the NCBI RefSeq database and including only complete staphylococcal genomes. This analysis was performed to determine if there was any contamination of the DNA prior to sequencing and as a secondary confirmation of the *S. aureus* species identification. Secondly, Adapter trimmed, high quality reads were mapped against the reference genome AUS0325, a closed ST88 methicillin-susceptible, penicillin-resistant genome (NCBI accession no. LT615218) to identify single nucleotide polymorphisms (SNPs). A maximum likelihood tree was generated from core genome SNPs. To further investigate the clonal relationship of MRSA ST88 from the African continent, our strains were compared to the 17 publically available ST88 MRSA isolated in Ghana (Kpeli et al., 2017). The Resfinder v.3.0 and Virulence finder v.1.5 web-based pipelines were used to search for the presence of known virulence and antibiotic resistance genes (Zankari et al., 2012; Joensen et al., 2014).



## RESULT

MRSA was recovered from 20/425 (4.7%) pigs, 6/55 (10.9%) pig farm workers, and 12/200 (6.0%) non-pig contact humans (Table 1). The MRSA positive pigs came from 12 farms (Figure 1). Despite sampling multiple pigs per farm, only 4/12 farms had >1 MRSA positive pigs. Six of the human MRSA isolates were recovered from “pig contact” humans on six pig farms. However, only 3/6 individuals came from farms, which also harbored MRSA positive pigs. Moreover, despite sampling more than one pig farm worker on these six farms, only one individual per farm was positive. The remaining 12 MRSA were isolated from University students not having any pig contact. None of the MRSA positive individuals had reported taking antibiotics and only one pig farm worker had reported being hospitalized in the last 12 months prior to sampling.

All 38 MRSA isolates were resistant to penicillin and cefoxitin only and there was agreement between phenotypic and inferred genotypic resistance. All strains were positive for the CC398 specific band. However, *spa* sequencing demonstrated that all strains had the same *spa* type, t1603, which is unrelated to *spa* types typically associated with CC398. Furthermore, MLST demonstrated that strains belong to ST88. *In silico* PCR using the CC398 specific primers, which target the C398-specific *sau1-hsdS1* variant, (FP2sau1:

5’GAGAATGATTTTGTTTATAACCCTAG3’ and CC398r1: 5’-CAGTATAAAGAGGTGACATGACCCCT-3’) and our *de novo* assembled contigs, showed that the primers bind 100% and produce the expected 106 bp amplicon. The primers bind to an open reading frame (ORF) with 99% nucleotide identity to *hsdS*, which is the sequence specificity gene of the *Sau1* type I restriction-modification system (data not shown). *mecA* was harbored on an SCC*mec* type VIa cassette.

Based on WGS, the core genome consisted of 2407 coding DNA sequences (CDS) with *S. aureus* AUS0325 used as reference. A core genome phylogeny was inferred by mapping reads from the four sequenced isolates to AUS0325, and a phylogenetic tree was constructed using the core genome SNPs and maximum likelihood estimation. Relative to AUS0325, 1,248 SNPs were identified, but only 11 SNPs were found between the four isolates in this study despite the temporal and geographic differences (Table 2; Figure 2). Isolates from the same region i.e., Angyiba and despite a 2-year gap only differed by 3–8 SNPs. A phylogenetic comparison with the 17 ST88 from Ghana indicates that the Nigerian isolates formed a separate, distinct clade with 700 SNPs differences (Figure 3).

All MRSA strains were *pvl* negative and 37/38 strains contained the human immune evasion gene *scn*. The only strain that was *scn*-negative was of porcine origin. Furthermore, based on analysis of the *de novo* contigs in Virulence Finder, all four

**TABLE 1** | Characterization of MRSA isolated from pigs, pig farm workers and non-pig contact humans.

	Number of sampled pigs/humans	Number of pig farms	Number of MRSA positive farms	Number of MRSA positive samples	Location of MRSA positive farms/students	Sampling period	Presence of <i>scn</i> amongst MRSA isolates
Pigs	425	35	12	20	Ankpa, Anyigba, Echeno, Egbe, Ejule, Itama, Lokoja, Ogidi, Oguma, Okpo, Okura-Ofifante, Sheria	Nov 2013–Jan 2015	19
Pig farm workers	55	35	6*	6	Ajaka, Anyigba, Elechi, Kabba, Oguma, Sheria	Nov 2013–Jan 2015	6
Kogi State University students	200	–	–	12	Anyigba	May–Nov 2014	12

\*3/6 farms harbored MRSA positive pigs. Underlined location of the farms.

**TABLE 2** | Single nucleotide polymorphisms in AM492, AM496, AM497, and AM526 relative to the ST88 reference genome, AUS0325.

Nucleotide position	AM526	AM497	AM492	AM496
853428	T	T	C	C
853440	C	Gap	Gap	Gap
853452	C	Gap	Gap	Gap
1349949	T	G	T	T
2673340	G	G	G	A
2673364	A	A	A	G
2673388	G	G	Gap	Gap
2673400	G	G	Gap	Gap
2673412	Gap	Gap	Gap	G
2673424	Gap	Gap	G	G
2673436	Gap	Gap	G	G

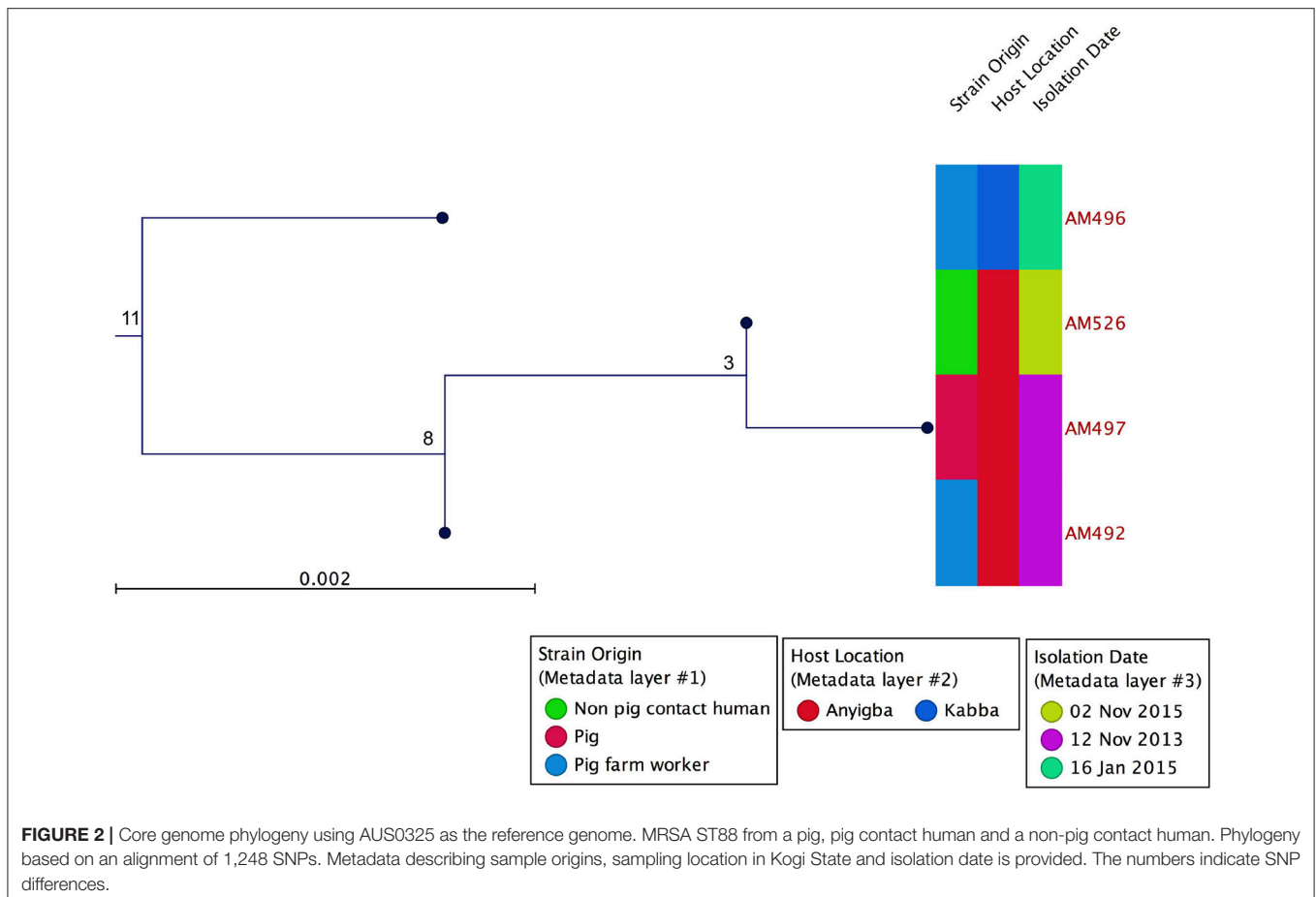
strains contained the following putative virulence associated genes *scn*, *sak*, *lukE*, *lukD*, gamma hemolysin, *aur*, *splA*, and *splB*.

## DISCUSSION

We describe for the first time the isolation of genetically confirmed MRSA in pigs and pig farm workers in Nigeria. This is only the fifth description of MRSA in healthy pigs on the African continent and appears not to be the typical livestock associated clones found in pigs worldwide namely CC398 and CC9.

All MRSA isolates found in this study belonged to ST88, which has been isolated from both hospital- and community-acquired infections in humans in several Sub-Saharan African countries, including Nigeria (Ghebremedhin et al., 2009; Shittu et al., 2012; Kolawole et al., 2013; Raji et al., 2013; Abdulgader et al., 2015). ST88 appears to be primarily associated with the African continent, but has also been reported in Asia and sporadically in other parts of the world (Schaumburg et al., 2014). The association of ST88 and livestock has only been described in Africa. Specifically, MRSA ST88 has been identified in one pig isolate from Senegal and healthy sheep in the Ivory Coast (Fall et al., 2012; Schaumburg et al., 2015). In China, MRSA ST88 has been found in food of animal origin and recently in livestock workers (Wang et al., 2014; Ye et al., 2016).

MRSA in pigs and pig farmers in Africa was first reported in Senegal (Fall et al., 2012) however, no MRSA was isolated from the farm workers. In Nigeria, there has been only one report of phenotypic oxacillin-resistant *S. aureus* in pigs from the Oyo State in South Western, Nigeria (Okunlola and Ayandele, 2015). Only 18/200 (9%) pigs were positive for presumptive MRSA on 7 of the 11 farms sampled. MRSA has been isolated from other animals in Nigeria namely; camels, sheep, cattle, goats and recently in poultry (Mai-siyama et al., 2014; Nworie et al., 2017). Similar to (Okunlola and Ayandele, 2015), we observed few MRSA positive animals on each farm. Moreover, all our MRSA isolates in this study belong to the same *spa* type (t1603) irrespective of the source (pig, pig-contact human or non-pig-contact human) and based on WGS the high genetic similarity,

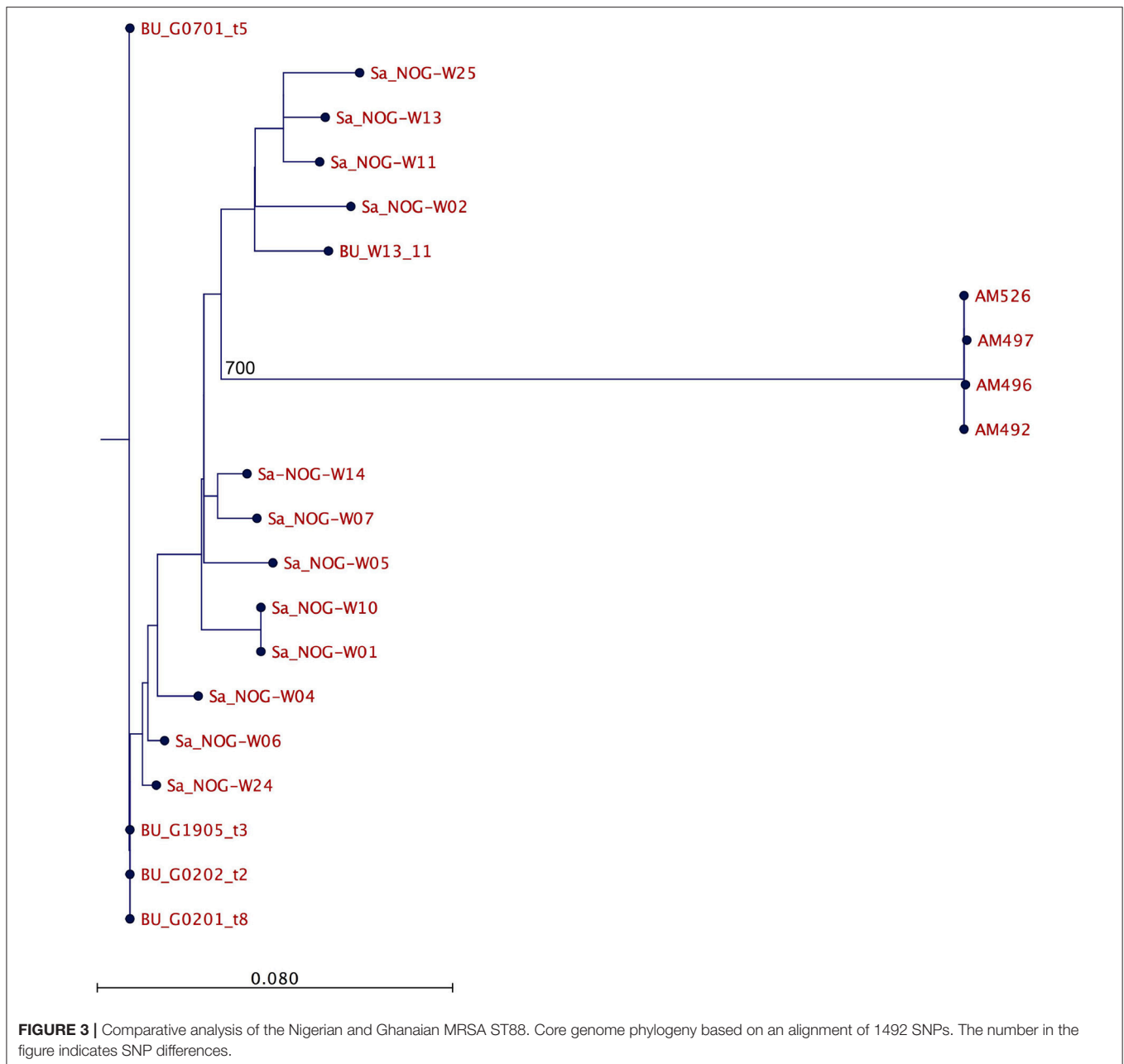


raises questions about the origin of these strains. The immune evasion cluster (IEC), which is carried on a bacteriophage and encodes the secreted proteins staphylococcal complement inhibitor (*scn*), staphylokinase (*sak*), and chemotaxis inhibitory protein (*chp*), are thought to contribute to the immune evasion in humans (van Wamel et al., 2006). Moreover, these genes are less prevalent in livestock adapted *S. aureus* lineages and are hence considered good genetic markers for identification of human associated *S. aureus* clones (McCarthy et al., 2011, 2012; Uhlemann et al., 2012). We found a high incidence of *scn* amongst our porcine isolates (19/20 isolates) and the additional occurrence of *sak* based on WGS data. The presence of these IEC related genes suggest a possible human origin and that pigs were either transiently contaminated by farm workers or the result of a very recent human-to-pig transmission event. Human associated *S. aureus* lineages have been previously described in animals including pigs (Arriola et al., 2011; Schaumburg et al., 2012; Nagel et al., 2013; Baez et al., 2017).

The MRSA ST88 in our study was only resistant to  $\beta$ -lactams. Typically, antibiotic resistance amongst animal *S. aureus* isolates tend to be higher compared to human isolates. A study of methicillin resistant coagulase-negative staphylococci (MRCoNS) of porcine origin in Nigeria, found that isolates were multi-drug resistant (MDR), 85% of strains were resistant

to fusidic acid, and harbored a number of different antibiotic resistance genes (Ugwu et al., 2015). CoNS are a reservoir of antibiotic resistance genes with the potential to transfer to *S. aureus*, as has been demonstrated with the plasmid-mediated transfer of *mupA*, conferring mupirocin resistance, from *S. haemolyticus* to *S. aureus* (Rossi et al., 2016). It is possible that these low-level resistant MRSA ST88 strains if allowed to adapt to pigs and occupy the same niche as MDR MRCoNS, provide an opportunity for the transfer of antibiotic resistance genes to potentially create MDR CA-MRSA.

All our strains were presumptively identified as belonging to CC398. However, upon additional strain typing, this was shown to be incorrect. Typing of MRSA is key for surveillance, epidemiological studies and infection control. Rapid, reliable identification methods such as PCR based methods are important tools to identify MRSA and type to the clonal complex level. The CC398 specific PCR is an excellent tool to rapidly identify *S. aureus* belonging to CC398, which was shown to be 100% specific for CC398 only (Stegger et al., 2011). This PCR is based on *sau1-hsdS* since it was previously shown that variations in this gene is associated with specific *S. aureus* clonal lineages but maintained a high sequence homology within the lineage (Waldron and Lindsay, 2006). This approach has also been used for rapid identification of hospital-acquired MRSA lineages



(Cockfield et al., 2007). Unfortunately, at the time of the study by Stegger et al. (2011) there were no publically available ST88 genomes and strains belonging to ST88 were not included in the study since ST88 is not the typical LA-MRSA and it is not a globally disseminated human associated clone. In light of our findings, we recommend that with the current PCR primers the CC398 specific PCR result should be complemented with *spa* typing or another method to verify the clonal lineage association.

Our study has some limitations. Ideally, all 38 MRSA strains should be whole genome sequenced to better understand ST88 strain diversity in this region and study possible transmission events. However, the latter would be difficult to assess as we do

not have information about the movement of pigs e.g., via trade, and the contact and movement of people. Additionally, we could not sequence all strains because of financial limitations.

Livestock are known reservoirs of new pathogenic bacteria with the ability to cross the host species barrier, and evolve in a host-specific manner to become established in the human population. This has been demonstrated for LA-MRSA CC398 (Price et al., 2013). Further research is required in Nigeria to determine whether MRSA carriage amongst pigs was transient or persistent, establish the exact transmission routes on pig farms and explore biosecurity measures for preventing the spread of MRSA in the farm

environment. While there are limitations to our study; cross-sectional study design conducted at one time point only, nonetheless we provide valuable information on a continent where there are limited studies describing the occurrence of MRSA in animals and even fewer providing molecular typing data.

## AUTHOR CONTRIBUTIONS

OO, JK, and EO designed of the work. OO, JK, EO, MI, and AM performed the acquisition, the analysis, and interpretation of data. OO, JK, EO, MI, and AM drafted and revised the manuscript. All authors approved the final version of this manuscript.

## REFERENCES

- Abdulgader, S. M., Shittu, A. O., Nicol, M. P., and Kaba, M. (2015). Molecular epidemiology of methicillin-resistant *Staphylococcus aureus* in Africa: a systematic review. *Front. Microbiol.* 30:348. doi: 10.3389/fmicb.2015.00348
- Adegoke, A. A., and Okoh, I. A. (2014). Species diversity and antibiotic resistance properties of *Staphylococcus* of farm animal origin in Nkonkobe Municipality, South Africa. *Folia. Microbiol.* 59, 133–140. doi: 10.1007/s12223-013-0275-1
- Arriola, C. S., Güere, M. E., Larsen, J., Skov, R. L., Gilman, R. H., Gonzalez, A. E., et al. (2011). Presence of methicillin-resistant *Staphylococcus aureus* in pigs in Peru. *PLoS ONE.* 6:e28529. doi: 10.1371/journal.pone.0028529
- Baez, M., Collaud, A., Espinosa, I., and Perreten, V. (2017). MRSA USA300, USA300-LV and ST5-IV in pigs, Cuba. *Int. J. Antimicrob. Agents.* 49, 259–261. doi: 10.1016/j.ijantimicag.2016.12.001
- Breurec, S., Zriouil, S. B., Fall, C., Boisier, P., Brisse, S., Djibo, S., et al. (2011). Epidemiology of methicillin-resistant *Staphylococcus aureus* lineages in five major African towns: emergence and spread of atypical clones. *Clin. Microbiol. Infect.* 17, 160–165. doi: 10.1111/j.1469-0691.2010.03219.x
- Clinical and Laboratory Standards Institute (CLSI) (2014). *Performance Standards for Antimicrobial Susceptibility Testing; Twenty-Fourth Information Supplement, Vol. 34.* CLSI document M100-24S. Wayne: Clinical and Laboratory Standards Institute.
- Cockfield, J. D., Pathak, S., Edgeworth, J. D., and Lindsay, J. A. (2007). Rapid determination of hospital-acquired methicillin-resistant *Staphylococcus aureus* lineages. *J. Med. Microbiol.* 56, 614–619. doi: 10.1099/jmm.0.47074-0
- Denis, O., Suetens, C., Hallin, M., Catry, B., Ramboer, I., Dispas, M., et al. (2009). Methicillin-resistant *Staphylococcus aureus* ST398 in swine farm personnel, Belgium. *Emerg. Infect. Dis.* 15, 1098–1101. doi: 10.3201/eid1507.080652
- Enright, M. C., Day, N. P., Davies, C. E., Peacock, S. J., and Spratt, B. G. (2000). Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J. Clin. Microbiol.* 38, 1008–1015.
- Fall, C., Seck, A., Richard, V., Ndour, M., Sembene, M., Laurent, F., et al. (2012). Epidemiology of *Staphylococcus aureus* in pigs and farmers in the largest farm in Dakar, Senegal. *Foodborne Pathog. Dis.* 9, 962–965. doi: 10.1089/fpd.2012.1197
- Ghebremedhin, B., Olugbosi, M. O., and Raji, A. M. (2009). Emergence of a community associated methicillin resistant *Staphylococcus aureus* strain with a unique resistant profile in southwest Nigeria. *J. Clin. Microbiol.* 47, 2975–2980. doi: 10.1128/JCM.00648-09
- Harmsen, D., Claus, H., Witte, W., Rothgänger, J., Claus, H., Turnwald, D., et al. (2003). Typing of methicillin-resistant *Staphylococcus aureus* in a university hospital setting by using novel software for *spa* repeat determination and database management. *J. Clin. Microbiol.* 41, 5442–5448. doi: 10.1128/JCM.41.12.5442-5448.2003
- Islam, M. Z., Espinosa-Gongora, C., Damborg, P., Sieber, R. N., Munk, R., Husted, L., et al. (2017). Horses in Denmark are a reservoir of diverse clones of methicillin-resistant and -susceptible *Staphylococcus aureus*. *Front. Microbiol.* 8:543. doi: 10.3389/fmicb.2017.00543

## FUNDING

Sampling, isolation, and preliminary analysis of MRSA was self financed by OO as part of this PhD project. OO received a travel grant from Kogi State University for a short research stay to perform molecular characterization of MRSA at the University of Copenhagen, Denmark. The molecular characterization was funded by the Danish Council for Independent Research-Technology and Production Sciences (DFF-FTP).

## ACKNOWLEDGMENTS

We would like to thank Tatjana P. Kristensen and Manja Hanegård for technical assistance.

- Joensen, K. G., Scheutz, F., Lund, O., Hasman, H., Kaas, R. S., Nielsen, E. M., et al. (2014). Real-time whole-genome sequencing for routine typing, surveillance, and outbreak detection of verotoxigenic *Escherichia coli*. *J. Clin. Microbiol.* 52, 1501–1510. doi: 10.1128/JCM.03617-13
- Kolawole, O. D., Adeyanju, A., Schaumburg, F., Akinyoola, L. A., Lawal, O. O., Amusa, Y. B., et al. (2013). Characterization of colonizing *Staphylococcus aureus* isolated from surgical wards' patients in a Nigerian University Hospital. *PLoS ONE.* 8:e68721. doi: 10.1371/journal.pone.0068721
- Kondo, Y., Ito, T., Ma, X. X., Watanabe, S., Kreiswirth, B. N., Etienne, J., et al. (2007). Combination of multiplex PCRs for staphylococcal cassette chromosome *mec* type assignment: rapid identification system for *mec*, *ccr*, and major differences in junkyard regions. *Antimicrob. Agents Chemother.* 51, 264–274. doi: 10.1128/AAC.00165-06
- Kpeli, G., Buultjens, A. H., Giulieri, S., Owusu-Mireku, E., Aboagye, S. Y., and Baines, S. L. (2017). Genomic analysis of ST88 community-acquired methicillin resistant *Staphylococcus aureus* in Ghana. *PeerJ.* 28:e3047. doi: 10.7717/peerj.3047
- Larsen, J., Petersen, A., Larsen, A. R., Sieber, R. N., Stegger, M., Koch, A., et al. (2017). Emergence of livestock-associated methicillin-resistant *Staphylococcus aureus* bloodstream infections in Denmark. *Clin. Infect. Dis.* 67, 1072–1076. doi: 10.1093/cid/cix504
- Liu, W., Liu, Z., Yao, Z., Fan, Y., Ye, X., and Chen, S. (2015). The prevalence and influencing factors of methicillin-resistant *Staphylococcus aureus* carriage in people in contact with livestock: a systematic review. *Am. J. Infect. Cont.* 43: 469–475. doi: 10.1016/j.ajic.2014.12.009
- Mai-siyama, I. B., Okon, K. O., Adamu, N. B., Askira, U. M., Isyaka, T. M., Adamu, S. G., et al. (2014). Methicillin-resistant *Staphylococcus aureus* (MRSA) colonization rate among ruminant animals slaughtered for human consumption and contact persons in Maiduguri, Nigeria. *Afr. J. Microbiol. Res.* 8, 2643–2649. doi: 10.5897/AJMR2014.6855
- McCarthy, A. J., van Wamel, W., Vandendriessche, S., Larsen, J., Denis, O., and Garcia-Graells, C., et al (2012). *Staphylococcus aureus* CC398 clade associated with human-to-human transmission. *Appl. Environ. Microbiol.* 78, 8845–8848. doi: 10.1128/AEM.02398-12
- McCarthy, A. J., Witney, A. A., Gould, K. A., Moodley, A., Guardabassi, L., Voss, A., et al. (2011). The distribution of mobile genetic elements (MGEs) in MRSA CC398 is associated with both host and country. *Genome. Biol. Evol.* 3, 1164–1174. doi: 10.1093/gbe/evr092
- Nagel, M., Dischinger, J., Türck, M., Verrier, D., and Oedenkoven, M., Ngoubangoye, B. et al (2013). Human-associated *Staphylococcus aureus* strains within great ape populations in Central Africa (Gabon). *Clin. Microbiol. Infect.* 19, 1072–1077. doi: 10.1111/1469-0691.12119
- Nworie, A., Onyema, A. S., Okeka, S. I., Elom, M. O., Umoh, N. O., Usanga, V. U., et al. (2017). A novel methicillin-resistant *Staphylococcus aureus* t11469 and a poultry endemic strain t002 (ST5) are present in chicken in Ebonyi State, Nigeria. *Biomed. Res. Int.* 2936461. doi: 10.1155/2017/2936461
- Ogunlola, I. O., and Ayandele, A. A. (2015). Prevalence and antimicrobial susceptibility of Methicillin-resistant *Staphylococcus aureus* (MRSA) among

- pigs in selected farms in Ilora, South Western Nigeria. *Euro. J. Exp. Bio.* 5, 50–56.
- Price, L. B., Stegger, M., Hasman, H., Aziz, M., Larsen, J., and Andersen, P. S., et al (2013). *Staphylococcus aureus* CC398: host adaptation and emergence of methicillin resistance in livestock. *MBio.* 3:e00305–e00311. doi: 10.1128/mBio.00305-11
- Raji, A., Ojemhen, O., Umejiburu, U., Ogunleye, A., Blanc, D. S., and Basset, P. (2013). High genetic diversity of *Staphylococcus aureus* in a tertiary care hospital in Southwest Nigeria. *Diagn. Microbiol. Infect. Dis.* 77, 367–369. doi: 10.1016/j.diagmicrobio.2013.08.030
- Rossi, C. C., Ferreira, N. C., Coelho, M. L., Schuenck, R. P., Bastos Mdo, C., and Giambiagi-deMarval, M. (2016). Transfer of mupirocin resistance from *Staphylococcus haemolyticus* clinical strains to *Staphylococcus aureus* through conjugative and mobilizable plasmids. *FEMS Microbiol Lett.* 363:fnw121. doi: 10.1093/femsle/fnw121
- Schaumburg, F., Alabi, A. S., Peters, G., and Becker, K. (2014). New epidemiology of *Staphylococcus aureus* infection in Africa. *Clin. Microbiol. Infect.* 20, 589–596. doi: 10.1111/1469-0691.12690
- Schaumburg, F., Mugisha, L., Peck, B., Becker, K., Gillespie, T. R., and Peters, G., et al (2012). Drug-resistant human *Staphylococcus aureus* in sanctuary apes pose a threat to endangered wild ape populations. *Am. J. Primatol.* 74, 1071–1075. doi: 10.1002/ajp.22067
- Schaumburg, F., Pauly, M., Anoh, E., Mossoun, A., Wiersma, L., Schubert, G., et al. (2015). *Staphylococcus aureus* complex from animals and humans in three remote African regions. *Clin. Microbiol. Infect.* 21, 345.e1-8. doi: 10.1016/j.cmi.2014.12.001
- Shittu, A., Oyedara, O., Abegunrin, F., Okon, K., Raji, A., Taiwo, S., et al. (2012). Characterization of methicillin-susceptible and -resistant staphylococci in the clinical setting: a multicentre study in Nigeria. *BMC Infect Dis.* 12:286. doi: 10.1186/1471-2334-12-286
- Stegger, M., Lindsay, J. A., Moodley, A., Skov, R., Broens, E. M., and Guardabassi, L. (2011). Rapid PCR detection of *Staphylococcus aureus* clonal complex 398 by targeting the restriction-modification system carrying sauI-hsdS1. *J. Clin. Microbiol.* 49, 732–734. doi: 10.1128/JCM.01970-10
- Ugwu, C. C., Gomez-Sanz, E., Agbo, I. C., Torres, C., and Chah, K. F. (2015). Characterization of mannitol-fermenting methicillin-resistant staphylococci isolated from pigs in Nigeria. *Braz. J. Microbiol.* 46, 885–892. doi: 10.1590/S1517-838246320140644
- Uhlemann, A. C., Porcella, S. F., Trivedi, S., Sullivan, S. B., Hafer, C., and Kennedy, A. D., et al (2012). Identification of a highly transmissible animal-independent *Staphylococcus aureus* ST398 clone with distinct genomic and cell adhesion properties. *MBio.* 3:e00027–e00012. doi: 10.1128/mBio.00027-12
- Unnerstad, E. H., Wahlstrom, H., Molander, B., and Bengtsson, B. (2017). Methicillin-resistant *Staphylococcus aureus* not detected in Swedish nucleus and multiplying pig herds. *Infect. Eco Epidemiol.* 7:1313068. doi: 10.1080/2008686.2017.1313068
- van der Wolf, P. J., Rothkamp, A., Junker, K., and de Neeling, A. J. (2012). *Staphylococcus aureus* (MSSA) and MRSA (CC398) isolated from post-mortem samples from pigs. *Vet. Microbiol.* 158, 136–141. doi: 10.1016/j.vetmic.2012.01.025
- Van Lochem, S., Thompson, P. N., and Annandale, C. H. (2018). Prevalence of methicillin-resistant *Staphylococcus aureus* among large commercial pig herds in South Africa. *Onderstepoort. J. Vet. Res.* 85:e1–e4. doi: 10.4102/ojvr.v85i1.1561
- van Wamel, W. J., Rooijackers, S. H., Ruyken, M., van Kessel, K. P., and van Strijp, J. A. (2006). The innate immune modulators staphylococcal complement inhibitor and chemotaxis inhibitory protein of *Staphylococcus aureus* are located on beta-hemolysin-converting bacteriophages. *J. Bacteriol.* 188:1310–1315. doi: 10.1128/JB.188.4.1310-1315.2006
- Vanderhaeghen, W., Hermans, K., Haesebrouck, F., and Butaye, P. (2010). Methicillin-resistant *Staphylococcus aureus* (MRSA) in food production animals. *Epidemiol. Infect.* 138, 606–625. doi: 10.1017/S0950268809991567
- Waldron, D. E., and Lindsay, J. A. (2006). SauI: a novel lineage-specific type I restriction-modification system that blocks horizontal gene transfer into *Staphylococcus aureus* and between *S. aureus* isolates of different lineages. *J. Bacteriol.* 188, 5578–5585. doi: 10.1128/JB.00418-06
- Wang, X., Li, G., Xia, X., Yang, B., Xi, M., and Meng, J. (2014). Antimicrobial susceptibility and molecular typing of methicillin-resistant staphylococcus aureus in retail foods in Shaanxi, China. *Foodborne Pathog. Dis.* 11, 281–286. doi: 10.1089/fpd.2013.1643
- Ye, X., Wang, X., Fan, Y., Peng, Y., Li, L., Li, S., et al. (2016). Genotypic and phenotypic markers of livestock-associated methicillin-resistant *Staphylococcus aureus* CC9 in Humans. *Appl. Environ. Microbiol.* 82, 3892–3899. doi: 10.1128/AEM.00091-16
- Zankari, E., Hasman, H., Cosentino, S., Vestergaard, M., Rasmussen, S., Lund, O., et al. (2012). Identification of acquired antimicrobial resistance genes. *J. Antimicrob. Chemother.* 67, 2640–2644. doi: 10.1093/jac/dks261

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Otalalu, Kwaga, Okolocha, Islam and Moodley. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.