



Serum Insulin-Like Growth Factor-1 in Parkinson's Disease; Study of Cerebrospinal Fluid Biomarkers and White Matter Microstructure

Farzaneh Ghazi Sherbaf^{1†}, Bahram Mohajer^{2†}, Amir Ashraf-Ganjouei¹, Mahtab Mojtaheh Zadeh¹, Ali Javinani¹, Hossein Sanjari Moghaddam¹, Mehdi Shirin Shandiz^{3*} and Mohammad Hadi Aarabi^{1*}

¹ Faculty of Medicine, Tehran University of Medical Sciences, Tehran, Iran, ² Non-communicable Diseases Research Center, Endocrinology and Metabolism Population Sciences Institute, Tehran University of Medical Sciences, Tehran, Iran, ³ Department of Medical Physics, Zahedan University of Medical Sciences, Zahedan, Iran

OPEN ACCESS

Edited by:

Stuart Maudsley,
University of Antwerp, Belgium

Reviewed by:

Jeff S. Davies,
Swansea University, United Kingdom
Yan Ma,
Edwards Lifesciences, United States

*Correspondence:

Mehdi Shirin Shandiz
shandiz1359@gmail.com
Mohammad Hadi Aarabi
mohammadhadiarabi@gmail.com

[†]These authors have contributed
equally to this work

Specialty section:

This article was submitted to
Molecular and Structural
Endocrinology,
a section of the journal
Frontiers in Endocrinology

Received: 21 March 2018

Accepted: 24 September 2018

Published: 02 November 2018

Citation:

Ghazi Sherbaf F, Mohajer B, Ashraf-Ganjouei A, Mojtaheh Zadeh M, Javinani A, Sanjari Moghaddam H, Shirin Shandiz M and Aarabi MH (2018) Serum Insulin-Like Growth Factor-1 in Parkinson's Disease; Study of Cerebrospinal Fluid Biomarkers and White Matter Microstructure. *Front. Endocrinol.* 9:608. doi: 10.3389/fendo.2018.00608

Background: Growing evidence shows that impaired signaling of Insulin-like Growth Factor-1 (IGF-1) is associated with neurodegenerative disorders, such as Parkinson's disease (PD). However, there is still controversy regarding its proinflammatory or neuroprotective function. In an attempt to elucidate the contribution of IGF-1 in PD, we aimed to discover the relation between serum IGF-1 levels in drug-naïve early PD patients and cerebrospinal fluid (CSF) biomarkers as well as microstructural changes in brain white matter.

Methods: The association between quartiles of serum IGF-1 levels and CSF biomarkers (α -synuclein, dopamine, amyloid- β_{1-42} , total tau, and phosphorylated tau) was investigated using adjusted regression models in 404 drug-naïve early PD patients with only mild motor manifestations and 188 age- and sex-matched healthy controls (HC) enrolled in the Parkinson's Progression Markers Initiative (PPMI). By using region of interest analysis and connectometry approach, we tracked the white matter microstructural integrity and diffusivity patterns in a subgroup of study participants with available diffusion MRI data to investigate the association between subcomponents of neural pathways with serum IGF-1 levels.

Results: PD patients had higher levels of IGF-1 compared to HC, although not statistically significant (mean difference: 3.60, $P = 0.44$). However, after adjustment for possible confounders and correction for False Discovery Rate (FDR), IGF-1 was negatively correlated with CSF α -synuclein, total and phosphorylated tau levels only in PD subjects. The imaging analysis proved a significant negative correlation (FDR corrected P -value = 0.013) between continuous levels of serum IGF-1 in patients with PD and the connectivity, but not integrity, in following fibers while controlling for age, sex, body mass index, depressive symptoms, education years, cognitive status and disease duration: middle cerebellar peduncle, cingulum, genu and splenium of the corpus callosum. No significant association was found between brain white matter microstructural measures or CSF markers of healthy controls and levels of IGF-1.

Conclusion: Altered connectivity in specific white matter structures, mainly involved in cognitive and motor deterioration, in association with higher serum IGF-1 levels might propose IGF-1 as a potential associate of worse outcome in response to higher burden of α -synucleinopathy and tauopathy in PD.

Keywords: Parkinson's disease, insulin-like growth factor-1, connectometry, diffusion tensor imaging, synuclein, amyloid, tau, dopamine

INTRODUCTION

Parkinson's disease (PD) is the second most common neurodegenerative disorder after Alzheimer's disease (AD). It is proposed that progressive damage of α -synuclein inclusions to the neural cells by causing neuroinflammation and oxidative imbalance plays the central role in PD pathology (1–4). In addition, the existence of extracellular plaques of amyloid-beta and intracellular aggregation of abnormal tau proteins, the main pathological insults in AD, are as well documented in PD with a great impact on the development of cognitive decline and dementia (5, 6). Presentation of motor and non-motor symptoms of PD may be delayed after a long preclinical phase of neuronal loss (7). Discovering biomarkers of this silent neurodegeneration has recently captured attention in early diagnosis of PD, before the emergence of the debilitating motor and cognitive symptoms, when neuroprotective measures may be promising (8). Among postulated markers, Insulin-like growth factor-1 (IGF-1) has drawn attention as a possible target of modification in the disease course (9, 10).

IGF-1 is a potent neurotrophic factor, involved in neural plasticity, differentiation, myelination, and survival (11), and is increased in the neuroinflammatory responses (12). In spite of some evidence of inhibitory effect of IGF-1 on the inflammatory cascade in a reactive cell (13), there is still controversy regarding the proinflammatory role of this factor or its protective function *in vivo*. Altered IGF-1 signaling is suggested to be associated with neurodegenerative disorders, such as PD (14, 15). Dysregulated signaling of IGF-1 involves brain tissues of post-mortem PD in several regions beyond substantia nigra, in concordance with heterogeneity of symptoms in PD (16). Increased levels of IGF-1 in serum or CSF of early PD patients have been reported in some (17–20), but not all previous studies (21, 22). However, it is not clear whether this is a chronic increase, if there is at all, with imposed susceptibility to neurodegeneration, a compensatory response against neural damage, or merely a consequence of ongoing neurodegenerative processes. However, studies on preclinical models of PD and *in vitro* cell cultures have implicated the neuroprotective effects of IGF-1 in terms of reduced apoptosis, prevention of loss of dopaminergic neurons, blocking formation of α -synuclein aggregates, rescuing neurons from amyloid plaques and neurofibrillary tangles, as well as improvement of motor deficits in the clinical aspect (23–29). These findings imply a possible therapeutic potential of the IGF-1 signaling pathway (9), beyond the proposed role of this factor as a PD biomarker.

Recently, advanced neuroimaging techniques have provided valuable data to detect early PD (8). Diffusion MRI (dMRI) is a powerful tool to identify white matter microstructural changes and has revolutionized our understanding of the neuropathology of PD. Studies have shown that Lewy bodies and coexistent AD-type pathology (amyloid plaques and neurofibrillary tangles) are associated with white matter microstructural damage in PD (30, 31). The most frequently investigated diffusivity metrics, fractional anisotropy (FA), mean diffusivity (MD), radial diffusivity (RD), and axial diffusivity (AD) are indirect indicators of almost every pathology affecting the white matter (32) and have yielded valuable information about the microstructural changes in PD and its clinical relevance (33). dMRI connectometry is a novel analytical method with higher sensitivity and specificity than conventional diffusion metrics, as it only probes subcomponents of the neural tracts, which are significantly related to the study variable and has bypassed the limitations of end-to-end fiber tracking methods (34). In addition, Connectometry is depended on the Spin Distribution Function (SDF), which measures the density of water diffusion in any direction and reports peak SDF for each direction as quantitative anisotropy (QA). Thus, it is highly efficient to measure the connectivity between adjacent voxels of a neural tract and results in what is called "local connectome fingerprint," as it is highly specific to each individual (35).

To date, few studies have explored IGF-1 associations in PD, and no study has yet investigated the association between IGF-1 levels and microstructural white matter disruptions in PD patients. In order to understand the role of IGF-1 in PD pathophysiology, we investigated its association with CSF biomarkers and white matter microstructural changes in early drug-naïve patients with PD. We probed the integrity and connectivity of fiber tracts in relation to serum IGF-1 levels by performing atlas-based region of interest and connectometry analyses of diffusion MRI data.

PATIENTS AND METHODS

Participants

Data used in this study were obtained from the Parkinson's Progression Markers Initiative (PPMI) database (36) (www.ppmi-info.org/data). The study was approved by the institutional review board of all participating sites. Written informed consent was obtained from all participants before study enrolment. The study was performed in accordance with relevant guidelines and regulations according to the PPMI protocol. The participants were tested and confirmed negative

for any neurological disorders apart from PD. Inclusion of PD patients in PPMI meets the following criteria: (1) patients should have at least two of the following signs and symptoms: resting tremor, bradykinesia, and rigidity or either asymmetric resting tremor or asymmetric bradykinesia; (2) maximum of 2 years passed from PD diagnosis in the screening stage; (3) baseline H & Y stage of I or II; (4) Loss of dopaminergic neurons confirmed on DAT scans; (5) no use of PD medications within at least 6 months prior to the baseline visit; and (6) aged ≥ 30 years at the time of diagnosis. Only PD patients and age- and sex-matched healthy control subjects (HC), recruited from 11 different centers, for whom baseline visit values of covariates of interest, i.e., CSF biomarkers, and serum IGF-1 levels were available were enrolled in this study. Among these participants, subjects who had baseline dMRI data with acceptable quality (imaging subgroup) were included in the subsequent imaging analyses.

Baseline assessment of global cognitive status was evaluated using Montreal Cognitive Assessment (MoCA). Data of the 15-item geriatric depression scale (GDS) were used to evaluate depressive symptoms in the study groups. Motor symptom severity was scored based on the Movement Disorders Society Unified Parkinson's Disease Rating Scale (MDS-UPDRS) part III. A detailed method of each can be found on the PPMI website (<http://www.ppmi-info.org/study-design/research-documents-and-sops/>).

CSF Biomarkers Measurement

At the baseline visit, CSF samples were obtained from PPMI participants using lumbar puncture at each participating site. Concentrations of α -synuclein, amyloid-beta ($A\beta_{1-42}$), total tau (T -tau), phosphorylated tau (P -tau), and Dopamine were measured by a commercially available enzyme-linked immunosorbent assay kit (Covance). The detailed method can be found on the PPMI website (<http://www.ppmi-info.org/study-design/research-documents-and-sops/>).

IGF-1 Measurement

As outlined in the PPMI laboratory manual, venous blood samples of participants were collected at baseline visit in the morning after an overnight fasting period. Serum IGF-1 measurement was performed using the Quantikine Human IGF-1 Immunoassay, which employs the quantitative sandwich enzyme immunoassay technique. This assay recognizes natural and recombinant human IGF1 and is calibrated against a highly purified *E. coli*-expressed recombinant human IGF1, produced at R&D Systems. Pretreatment of serum with acid-ethanol extraction was done to release IGF-1 from IGF-binding proteins. Standards and pretreated samples were pipetted into the wells on a microplate, previously coated with a monoclonal antibody specific for IGF1. After washing away unbound substances with the antibody, an enzyme-linked polyclonal antibody specific for IGF1 was added to the wells. Following a second wash, a substrate solution was added to the wells and color developed in proportion to the amount of IGF1 bound in the initial step. A microplate reader (Victor X4-Perkin Elmer) set to 450 nm was used to measure the intensity of the color. A standard curve was

generated for each set of assayed samples. Intra and inter-assay precision were confirmed with multiple tests of three samples of known concentrations. Results of natural human IGF1 showed linear curves that were parallel to the standard curves obtained, using the Quantikine kit standards. The NIBSC/WHO IGF1 International Reference Reagent 02/254 was evaluated in this kit. The dose-response curve of the International Reference Reagent parallels the Quantikine standard curve. Sample values obtained with the Quantikine Human IGF1 kit were finally converted to NIBSC/WHO 02/254 values.

MRI Data Acquisition

This dataset was acquired on 3 Tesla Siemens scanners, producing 64 directions of dMRI (repetition time = 7,748 MS, echo time = 86 ms; voxel size: $2.0 \times 2.0 \times 2.0$ mm³; field of view = 224×224 mm) at $b = 1,000$ s/mm² and one b0 image. Details of the MRI acquisition can be found in the MRI technical operations manual at (<http://www.ppmi-info.org/study-design/research-documents-and-sops/>).

Diffusion MRI Data Analysis

Diffusion MRI data were corrected for subject motion, eddy current distortions, and susceptible artifacts due to the magnetic field inhomogeneity using ExploreDTI toolbox (37). To define the regions of interest (ROI), we performed automated atlas-based analysis with the Johns Hopkins University's Mori white matter atlas using affine and elastic registration based on "elastix". After these preprocessing steps, FA, MD, RD, and AD values were calculated in the 48 brain regions that are provided by the Mori atlas.

Connectometry analyses (34) were performed on subgroups of PD and HC subjects separately. The diffusion data were reconstructed in the MNI space using q-space diffeomorphic reconstruction to obtain the spin distribution function (SDF) (38). Next, dMRI connectometry was used to study the association of white matter connectivity with levels of serum IGF-1. SDF was normalized. A T -score threshold of 2 was assigned to select local connectomes, and the local connectomes were tracked using a deterministic fiber tracking algorithm. A length threshold of 40 mm was used to select tracks. The seeding density was 50 seeds per mm³. Multiple regression models were used to investigate the correlation of IGF-1 with white matter quantitative anisotropy (QA), separately in PD and HC subjects, adjusted for age, sex, Body Mass Index (BMI), disease duration, and education years as covariates in the model. To estimate the False Discovery Rate (FDR), a total of 2,000 randomized permutation was applied to the group label to obtain the null distribution of the track length. The analysis was conducted using publicly available software DSI Studio (<http://dsi-studio.labsolver.org/>) released on 5th April 2018.

Statistical Analysis

Demographic, clinical and CSF biomarker data were compared between HC and PD groups using independent sample t -test and chi-square tests, and in case of non-normal distribution assessed by Shapiro-Wilk test, Kruskal-Wallis was used in a substitute. IGF-1 serum level was further compared between groups using

a linear regression model, considering age, gender, and BMI as possible confounders. Statistical power was calculated for effect size = 0.1 and significance threshold = 0.05. Because of non-linear association of IGF-1 levels with dependent variables, IGF-1 values were categorized in four quartiles as follows: first quartile ≤ 97.9 ng/ml; second quartile = 97.9001–124 ng/ml; third quartile = 124.0001–162.200 ng/ml; and fourth quartile ≥ 162.2001 ng/ml. Adjusted multiple regression models were used to evaluate the association of serum IGF-1 levels with CSF biomarkers and brain white matter microstructural integrity. Models were fitted separately for PD and HC subjects considering age, gender, body mass index (BMI), education years, Hoehn & Yahr (H&Y) stage, and duration of disease in months as covariates (two latter were only fitted in the model of PD subjects). Models regarding CSF biomarkers were additionally adjusted for MOCA score while excluding education years. FDR approach with the significant level of 0.05 was used to correct results for family wise error. In order to clarify the possible effect of gender, motor stage, and global cognitive status (intact cognition and mild cognitive impairment), potential effect modifications by gender, H&Y stage, and MoCA scores were examined by adding interaction terms of (IGF-1 \times gender) or (IGF-1 \times H&Y) or (IGF-1 \times MoCA) separately to the fully adjusted models. Since *P*-values were not < 0.10 , no further stratification was applied based on gender, H&Y or MoCA. All the statistical analyses were done in the R statistical package (v3.4.3).

RESULTS

The study group consisted of 404 PD patients (263 females, mean age: 61.6 ± 9.7 years) and 188 HC (120 females, mean age: 60.8 ± 11.3 years). **Table 1** shows group-specific demographics and baseline level characteristics of total and imaging subgroup participants. All PD subjects had disease duration < 36 months with a mean duration of 6.6 ± 6.6 months. 180 patients were at H&Y stage 1 compatible with unilateral motor involvement and mild to no disability. Two patients were in stage 3, and the remaining patients were in stage 2 of PD progression with bilateral involvement but no impairment in their balance. HC subjects were all cognitively intact according to the MoCA scale (mean MoCA score: 28.23 ± 1.11). 88 PD subjects had the score between 19 and 25 compatible with mild cognitive impairment, and 2 had the score below 19 categorized as severe impairment. 256 PD and 131 HC participants scored 5 to 8 in geriatric depression score indicative of mild depression while the remaining subjects scored below 4. Mean BMI of the PD participants was 27.12 ± 4.52 . As the effect of BMI on the IGF-1 level was controlled in multiple regression analysis, we did not exclude obese participants. Of note, there were few PD patients with comorbid disorders, such as diabetes mellitus (17), thyroid dysfunction (59), and cancer (11). Four patients were taking corticosteroids at the baseline visit. However, excluding these participants did not change the results.

There was no significant difference between PD and HC group regarding age, sex ratio, BMI, and years of education. However, PD subjects had significantly lower scores in MoCA ($p < 0.001$)

and marginally higher scores of GDS ($P: 0.052$). All PD patients had significantly lower $A\beta_{1-42}$, *T*-tau, *P*-tau, α -synuclein, and dopamine compared to HC.

In the subgroup of participants included in the imaging analysis, there were no differences in age, gender ratio, BMI, education, as well as clinical and CSF biomarkers, except for UPDRS III, between 82 PD patients and 57 HC. All PD subjects of the imaging population had an H&Y score of 1 or 2 and none of them scored below 19 on the MoCA.

CSF concentration of IGF-1 was not statistically different between PD and HC subjects before and after adjustment (PD: 136.79 ± 54.42 , HC: 134.55 ± 56.33 ng/ml). Distribution of serum IGF-1 level quartiles were different between diagnoses, but the result did not remain statistically significant after adjustment for age, gender, and BMI (mean difference: 3.60, $P: 0.4$, estimated power ≈ 1).

Association of Serum IGF-1 Level Quartiles With CSF Biomarkers

In PD subjects, a significant negative association was found between serum IGF-1 quartiles and CSF α -synuclein, *T*-tau, and *P*-tau concentrations. Results remained significant after adjustment and FDR correction of *p*-values. The remaining associations between IGF-1 quartiles and CSF biomarkers of PD and HC subjects were not statistically meaningful (**Table 2**). Scaled (0–100) levels of CSF biomarker through IGF-1 quartiles are illustrated in the **Figure 1**.

Association of Serum IGF-1 Level With White Matter Microstructural Integrity and Connectivity

No association was found between IGF-1 levels, neither continuous nor quartile, with FA, MD, RD, or AD values of whole brain white matter tracts in PD or HC (**Supplementary Table**). There was also no interaction between IGF-1 levels and diagnosis with regional FA and MD values.

Results from multiple regression models in dMRI connectometry of PD patients revealed areas where white matter QA was negatively correlated with IGF-1 level. More specifically, the connectometry analysis identified lower connectivity in middle cerebellar peduncle (MCP), left and right cingulum, and genu and splenium of the corpus callosum (CC) in correlation with serum IGF-1 continuous level after adjustment for possible confounders and FDR correction of *p*-values (FDR = 0.013). Dividing patients based on IGF-1 quartiles, connectometry analysis revealed a negative association between IGF-1 continuous levels in the first quartile and QA of aforementioned tracts (FDR: 0.04). Bilateral cingulum and bilateral corticospinal tracts (CST) were the only areas with a trend to significantly lower connectivity in terms of lower QA in association with higher IGF-1 continuous levels in the fourth quartile (FDR: 0.05). No association was observed between IGF-1 continuous levels in the second or third quartiles and white matter QA in PD. When considering IGF-1 quartiles as ordinal variable, MCP, bilateral fornix, cingulum, genu and body of the CC showed significantly negative association in PD,

TABLE 1 | Demographic and baseline clinical data of participants.

Groups	Total population			Imaging population		
	HC (188)	PD (404)	<i>P</i> -value	HC (57)	PD (82)	<i>P</i> -value
Age at baseline visit in years, mean (SD)	60.87 (11.30)	61.66 (9.74)	0.61	59.57 (10.76)	57.70 (8.89)	0.173
Sex, Female, no. (%)	120 (63.8)	263 (65.1)	0.835	38 (66.7)	51 (62.2)	0.718
BMI (Kg/m ²), mean (SD)	26.97 (4.41)	27.12 (4.52)	0.756	26.66 (4.42)	26.84 (4.45)	0.984
Education years, mean (SD)	16.08 (2.89)	15.57 (2.99)	0.061	15.40 (2.84)	15.21 (2.81)	0.795
IGF-1 (ng/ml), mean (SD)	134.55 (56.33)	136.79 (54.42)	0.444	132.09 (55.64)	136.26 (46.06)	0.281
Adjusted IGF-1 difference*	Mean difference: 3.60		0.444	Mean difference: 3.16		0.714
	Power = 1			Power = 0.96		
IGF-1 quartiles (%)			0.013			0.518
1	39 (20.7)	109 (27.0)		16 (28.1)	20 (24.4)	
2	62 (33.0)	87 (21.5)		14 (24.6)	14 (17.1)	
3	48 (25.5)	99 (24.5)		15 (26.3)	23 (28.0)	
4	39 (20.7)	109 (27.0)		12 (21.1)	25 (30.5)	
Adjusted IGF-1 quartiles difference*	Estimate: 0.11,		0.487	Estimate: 0.34		0.280
	Power = 1			Power = 0.97		
Duration of disease in months, mean (SD)		6.66 (6.66)			6.76 (6.99)	
H & Y stage no. (%)						
0	188 (100.0)	0 (0.0)		57 (100.0)	0 (0.0)	
1	–	180 (44.6)		–	37 (45.1)	
2	–	222 (55.0)		–	45 (54.9)	
3	–	2 (0.5)		–	0 (0.0)	
UPDRS III, mean (SD)	1.16 (2.15)	20.72 (8.80)	<0.001	0.72 (1.45)	20.52 (8.76)	<0.001
GDS score, mean (SD)	4.74 (1.00)	4.54 (1.11)	0.052	4.54 (1.02)	4.38 (1.28)	0.647
MOCA score, mean (SD)	28.23 (1.11)	27.10 (2.34)	<0.001	28.19 (1.16)	27.77 (1.90)	0.46
Aβ _{1–42} , mean (SD)	1,024.18 (506.01)	914.94 (414.00)	0.028	973.55 (485.41)	906.41 (355.65)	0.728
Phosphorylated Tau, mean (SD)	16.86 (8.57)	14.36 (5.46)	0.001	15.28 (6.79)	13.90 (4.43)	0.561
Total Tau, mean (SD)	190.29 (81.35)	167.73 (58.63)	0.004	175.15 (72.17)	163.68 (48.81)	0.717
α-synuclein, mean (SD)	1,707.07 (760.51)	1,520.66 (673.34)	0.003	1,484.31 (598.53)	1,519.03 (637.42)	0.689
Dopamine, mean (SD)	0.01 (0.01)	0.01 (0.01)	0.021	0.01 (0.00)	0.01 (0.01)	0.102

H & Y stage, Hoehn and Yahr stage; BMI, body mass index; IGF, insulin-like growth factor; UPDRS III, unified Parkinson's disease rating scale part III; GDS, geriatric depression scale; MoCA, Montreal Cognitive Assessment. Shapiro–Wilk test used to assess normality of distribution. For the normally distributed continuous variables, t-test was implemented and in the case of non-normality in distribution Kruskal–Wallis test was used. Categorical variables were assessed using Pearson Chi-square and Fisher exact test. *Adjusted difference in IGF-1 serum levels were assessed using multiple regression model considering age, gender, and BMI as covariates. Power calculated estimating effect size = 0.1 and significance level = 0.05.

while splenium and bilateral corticospinal tracts were positively correlated with IGF-1 quartiles with *P*-values just below the significant threshold (FDR: 0.033 and 0.049, respectively).

No significant association was found between IGF-1 levels and white matter QA in all HC or divided based on IGF-1 quartiles.

DISCUSSION

In the present study, we have investigated the association of serum IGF-1 levels with CSF biomarkers, as well as whole brain white matter microstructural connectivity and integrity to elucidate the possible role of serum IGF-1 in the central pathophysiology of PD. PD type and AD-type CSF biomarkers in our subjects were lower compared to HC, in agreement with previous studies (39–42). It is speculated that precipitation of α-synuclein, amyloid-β, or tau proteins lowers

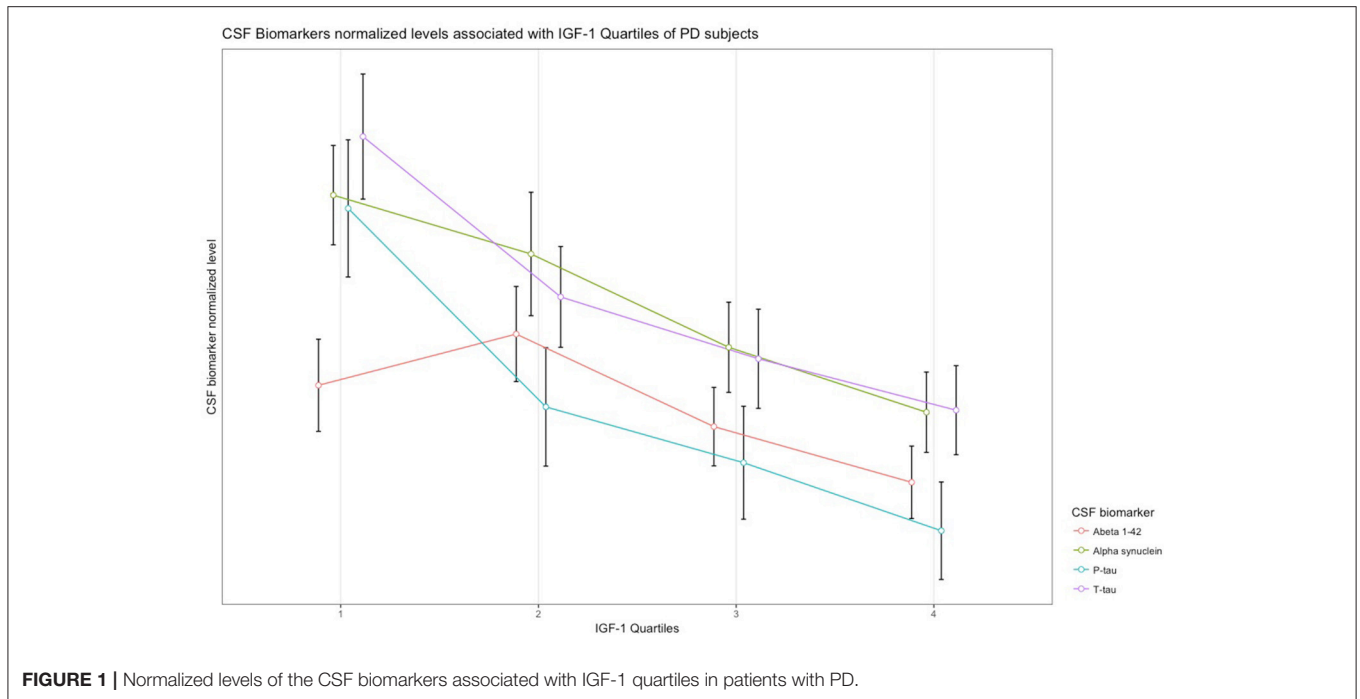
their free concentration in the CSF. Despite non-significant differences between serum IGF-1 levels of early drug-naïve PD patients and HC, we could reveal that decreased concentration of CSF α-synuclein and tau proteins correlate with higher serum IGF-1 quartiles in PD patients, a finding that did not replicate in HC. In addition, we found that structural brain dysconnectivity, but not disintegrity in cingulum, genu, and MCP is associated with higher serum IGF-1 continuous levels only in PD patients, and more particularly in the subgroup of patients with serum IGF-1 levels in the first quartile. On the other hand, higher IGF-1 quartiles correlated with higher connectivity in the splenium and CST, and lower connectivity in the cingulum, fornix, and genu and body of the CC.

IGF-1 receptors are distributed throughout the brain, with high densities in the hippocampus, frontal cortex, and cerebellum

TABLE 2 | Association of CSF biomarkers with IGF-1 quartiles in PD and HC subjects.

IGF-1	1st quartile	2nd quartile	3rd quartile	4th quartile	Estimate ± SE, P-value
PD	109 subjects	87 subjects	99 subjects	109 subjects	
Dopamine	0.01 (0.01)	0.01 (0.01)	0.01 (0.01)	0.01 (0.01)	0 ± 0.001, P: 0.788
α-synuclein	1,669.76 (703.08)	1,588.89 (782.35)	1,460.56 (607.11)	1,371.28 (568.47)	-179.64 ± 66.78, P: 0.007*
Total Tau (t-Tau)	185.72 (71.02)	168.03 (51.26)	161.21 (53.56)	155.54 (50.80)	-15.8 ± 5.705, P: 0.006*
Phosphorylated Tau (p-Tau)	16.12 (6.46)	14.30 (5.00)	13.79 (5.07)	13.17 (4.62)	-1.58 ± 0.53, P: 0.003*
Aβ ₁₋₄₂	940.37 (466.87)	991.03 (428.14)	899.54 (375.93)	844.48 (370.57)	-82.41 ± 41.90, P: 0.05
t-Tau Aβ ₁₋₄₂ ratio	0.22 (0.11)	0.18 (0.06)	0.20 (0.09)	0.21 (0.11)	0.003 ± 0.01, P: 0.718
p-Tau Aβ ₁₋₄₂ ratio	0.02 (0.01)	0.02 (0.01)	0.02 (0.01)	0.02 (0.01)	0 ± 0.001, P: 0.899
p-Tau t-Tau ratio	0.09 (0.01)	0.08 (0.01)	0.09 (0.01)	0.08 (0.01)	-0.002 ± 0.001, P: 0.043
HC	39 subjects	62 subjects	48 subjects	39 subjects	
Dopamine	0.01 (0.00)	0.01 (0.01)	0.01 (0.00)	0.01 (0.00)	-0.002 ± 0.002, P: 0.329
α-synuclein	1,895.98 (926.65)	1,773.07 (690.23)	1,689.51 (737.16)	1,451.42 (675.67)	-224.88 ± 125.87, P: 0.076
Total Tau (t-Tau)	211.65 (113.03)	198.03 (73.56)	181.43 (66.66)	168.86 (69.07)	-17.68 ± 13.12, P: 0.18
Phosphorylated Tau (p-Tau)	19.46 (13.83)	17.55 (7.16)	15.80 (6.05)	14.62 (5.88)	-2.17 ± 1.39, P: 0.12
Aβ ₁₋₄₂	947.08 (372.86)	1,080.15 (549.11)	1,070.43 (445.11)	958.47 (602.31)	17.83 ± 86.48, P: 0.837
t-Tau Aβ ₁₋₄₂ ratio	0.26 (0.25)	0.21 (0.12)	0.18 (0.07)	0.21 (0.13)	-0.02 ± 0.025, P: 0.411
p-Tau Aβ ₁₋₄₂ ratio	0.03 (0.03)	0.02 (0.01)	0.02 (0.01)	0.02 (0.01)	-0.003 ± 0.003, P: 0.382
p-Tau t-Tau ratio	0.09 (0.01)	0.09 (0.01)	0.09 (0.01)	0.09 (0.01)	0 ± 0.001, P: 0.814

Values are reported as mean (SD). *Significant after FDR (threshold < 0.05).



(43, 44). The cingulum bundle and CC have pivotal roles in cognitive function. The most anterior parts of these structures contribute to goal-directed behavior and attention modulation through the network of prefrontal projections, while the most posterior parts are involved in regulating memory with their

harbored temporal and parahippocampal connections (45–47). Remarkably, the cingulum and CC are the fibers most affected in the mild cognitive impairment, AD, and PD with dementia; The microstructural compromise in the parahippocampal and posterior cingulum, and the splenium of the CC is replicated

in several studies of amnesic mild cognitive impairment and AD (48–52). On the other hand, normal aging is shown to be more confined to the frontal circuit with the coexistence of disruptions in the genu and anterior cingulum (53). There are also several reports of the centrality of lower integrity of CC and cingulum in relation with poorer cognition and transition to demented state in patients with PD (54–58). Of note, the prefrontal white matter is the most consistently described region in PD (33), considering the executive dysfunction as the main cognitive disturbance in this neurodegenerative disorder (59). However, the involvement of posterior cognitive circuits confers an increased risk of dementia in PD (60). Moreover, The spread of α -synuclein aggregations to the limbic and neocortical structures can be used to differentiate between PD with and without dementia (61). In addition, AD-type pathology has synergistic pathology with a higher burden of α -synucleinopathy in the limbic and cortical structures of demented patients with PD (62).

Previously, Picillo et al. investigated the association between the serum IGF-1 quartiles with performance in several aspects of cognition in a similar group of subjects enrolled in PPMI database (22). They demonstrated that the lowest serum IGF-1 level is associated with cognitive decline in the domains of executive function, attention and verbal memory, which are mainly mediated by the prefrontal and temporal regions. The association of lower serum IGF-1 levels with poor cognitive ability is replicated in other studies as well (22, 63, 64). Furthermore, it is demonstrated that IGF-1 level rises non-significantly early in the course of PD, peaks at H&Y stage 2 and then decreases when PD ends in stages 3–5 (21, 65). Also, Godau et al. found that higher serum IGF-1 levels are correlated with shorter disease duration (17). A dome-shaped pattern through disease course on the side of *in vitro* and *in vivo* neuroprotective roles suggested for IGF-1 (18, 66, 67), along with our imaging findings of lower white matter QA and CSF biomarkers associated with higher IGF-1 in early PD, may propose a primary compensatory rise in IGF-1 signaling against neural damage of Lewy bodies and neurofibrillary tangles, with consequent depletion of neuroprotective capacity later in the course of the disease (17, 65). However, the exact pathophysiology still needs to be elucidated.

Studies have demonstrated conflicting results trying to link the level of serum IGF-1 to motor severity in PD. Picillo et al. have surveyed the relationship between serum IGF-1 level and progression of motor dysfunction in a 2-years follow-up of 37 early PD patients. They found that higher IGF-1 at baseline predicts more severe motor impairment with possibly better response to dopaminergic administration (20). A negative association was also replicated in a cohort of 79 Japanese PD patients. However, subjects were mostly in H&Y stages 3 and more, and the association was not confirmed in the subgroup of early drug naïve PD patients (65). Godau et al. showed a correlation between decreased serum IGF-1 levels and severity of motor symptoms among 15 newly diagnosed PD patients, but a positive correlation in 11 persons with suspected dopaminergic deficit and putatively at risk for PD (19). Considering limited data on CSF dopamine assessment in PPMI database (total of 90 PD subjects in our sample), we did not find any association

between IGF-1 quartiles and dopamine in early drug-naïve PD. However, there was a negative correlation between IGF-1 level quartiles and the CSF concentrations of α -synuclein and tau proteins. Although dopaminergic depletion is considered the mainstay of extrapyramidal motor manifestations in PD, Kang et al. have shown that CSF α -synuclein and T-tau are decreased in early PD patients, and this reduction is associated with severity of motor symptoms (41, 42). They also revealed that patients with lower P-tau are more likely to have the non-tremor dominant phenotype, who more often experience cognitive deterioration (68). Postural instability, bradykinesia or freezing of gait are shown to be related to structural incoherence in the genu, body and splenium of the CC, and cingulum (69–71). Impaired integrity in MCP is also demonstrated in PD patients with freezing of gait (72–74). Besides associations with motor manifestations, structural disturbance of the cerebellum is also linked to cognitive and executive dysfunction (75–77). Considering observed disconnectivity in the cingulum, CC and MCP in relation to higher IGF-1 continuous levels, increased IGF-1 in early PD may alarm the development of severe motor symptoms in the course of the disease. To further complicate this scenario, we observed a positive association between IGF-1 quartiles and the connectivity in the splenium and CST. There is multitude evidence concerning the reputation of incoherence in these structures regarding motor and cognitive deterioration in PD (33). The positive association of IGF-1 quartile specifically with these structures may reinforce the debate on the compensatory role of IGF-1 in early phases of PD. Although it seems that this compensation does not completely overcome the ongoing neural damage, as noted by its negative association with other major white matter pathways. To support this notion, a comprehensive *in vivo* and *in vitro* preclinical study has demonstrated that IGF-1 downstream signaling pathways enhance the axonal outgrowth in the CST (78). Another study on the mouse model of CC demyelination has revealed that IGF-1 expression is upregulated in the process of neural repair (79). Still, studies on human subjects with longitudinal design are needed to confirm this hypothesis.

Although the major white matter tracts with a central role in cognitive impairment and dementia, and with fundamental associations with motor severity showed altered connectivity in correlation with serum IGF-1 levels, there was no such association with white matter FA, MD, RD, or AD values. In fact, tensor measures rely on the diffusivity of each voxel and mainly detect the pathology in fiber tracts, while connectivity metrics, like QA, are based on density for each fiber orientation and count for individual differences (35). Therefore, QA has resolved the shortage of FA in the areas of crossing fibers. Notably, we were able to detect the microstructural changes in short association fibers, which are located in the region with high crossing fibers, in patients with PD by using the connectometry approach (80). Furthermore, several pathological features, such as axonal loss, demyelination, inflammation, or edema can alter FA. Edema without demyelination may increase FA, and alterations of FA may take a long time to appear in the setting of gradual neuronal loss. However, QA is more related to the neural changes and less affected by the inflammation and edema, and less susceptible

to partial volume effect (81). Therefore, connectivity is more sensitive and specific than tensor analyses. Longer duration of the disease might probably reveal FA changes in specific white matter pathways in relation to IGF-1, a hypothesis that needs to be investigated in well-designed cohort studies. Studying PD patients in advanced stages with dementia may further clarify the possible role of IGF-1 signaling in specific white matter disruptions.

The large sample size, both in total included population and imaging subgroup, has devoted a great statistical power to the observed results. Considering the previously described higher levels of IGF-1 in early phases of neurodegenerative disorders, studying early stages of PD without the conferring effects of treatment appear the best point in the disease course to evaluate the possible role of IGF-1 in the central pathophysiology of PD. Nevertheless, despite growing evidence of IGF-1 protection against amyloid accumulation or dopamine depletion, we could not find any significant association between IGF-1 levels and CSF concentrations of dopamine or $A\beta_{1-42}$. In addition, follow-up studies are warranted to clarify the generalizability of these results to more advanced stages and to determine the casual relationship between this promising neurotrophic factor and progression of PD. Even though we have considered major possible confounders and corrected our results for multiple comparisons, high prevalence of mild depression in the study population, lower number of subjects provided with dopamine levels and measurement of IGF-1 levels in the serum and not CSF should be kept in mind while interpreting our results.

CONCLUSION

In this study, we showed that despite no difference between serum IGF-1 level in PD compared to HC, it is negatively associated with CSF *T*-tau, *P*-tau, and α -synuclein, as well as altered microstructural connectivity in the several brain white matter regions of early drug naïve PD patients with mild motor symptoms. We demonstrated that lower connectivity measures in MCP, cingulum, and CC are correlated with higher continuous levels of IGF-1 exclusively in PD subjects. Disruption of these pathways is previously described in PD and in association with debilitating motor and cognitive disturbances. It is not clear whether the correlation of IGF-1 levels with alteration of CSF biomarkers and neural damage is a modulation in its signaling pathway as a compensatory mechanism, or is only a parallel finding to disease progression. In addition, we found higher IGF-1 quartiles in association with higher connectivity in the splenium and bilateral CST. This specific result may reinforce the debate on the compensatory role of IGF-1 against the ongoing neural damage in PD at least in some specific brain pathways. More studies are needed to assess these findings

in follow-up visits and inspect the development of motor or cognitive symptoms related to the changes in the CSF biomarkers and microstructural alterations in the specified pathways.

ETHICS STATEMENT

All procedures performed here, including human participants were in accordance with the ethical standards of the institutional research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

AUTHOR CONTRIBUTIONS

MA conceived of the presented idea. FG, BM, and MA contributed to the design and implementation of the research. MA and BM performed the analytic calculations. AJ and AA-G drafted the manuscript. FG wrote the manuscript with input from all authors. HS, MM, and MS contributed to data collection, aided in interpreting the results, and worked on the manuscript. All authors provided critical feedback and helped shape the research, analysis, and manuscript.

ACKNOWLEDGMENTS

The dataset of this study was funded by grants from the Michael J. Fox Foundation for Parkinson's Research, the W. Garfield Weston Foundation, and the Alzheimer's Association, the Canadian Institutes for Health Research, and the Natural Sciences and Engineering Research Council of Canada. We thank Christian Beckmann and Simon Eickhoff for their advice on data analysis. Data used in this article were obtained from the Parkinsons Progression Markers Initiative (PPMI) database (www.ppmi-info.org/data). For up-to-date information on the study, visit www.ppmi-info.org. PPMI is sponsored and partially funded by the Michael J. Fox Foundation for Parkinsons Research and funding partners, including AbbVie, Avid Radiopharmaceuticals, Biogen, Bristol-Myers Squibb, Covance, GE Healthcare, Genentech, GlaxoSmithKline (GSK), Eli Lilly and Company, Lundbeck, Merck, Meso Scale Discovery (MSD), Pfizer, Piramal Imaging, Roche, Servier, and UCB (www.ppmi-info.org/fundingpartners). This study was approved by the Ethical Committee of Zahedan University of Medical Sciences (license # Ir.zaums.rec.1397.28).

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2018.00608/full#supplementary-material>

REFERENCES

- Sanjari Moghaddam H, Ghazi Sherbaf F, Mojtahed Zadeh M, Ashraf-Ganjouei A, Aarabi MH. Association between peripheral inflammation and DATSCAN data of the striatal nuclei in different motor subtypes of Parkinson disease. *Front Neurol.* (2018) 9:234. doi: 10.3389/fneur.2018.00234
- Sanjari Moghaddam H, Valitabar Z, Ashraf-Ganjouei A, Mojtahed Zadeh M, Ghazi Sherbaf F, Aarabi MH. Cerebrospinal fluid C-reactive protein

- in Parkinson's disease: associations with motor and non-motor symptoms. *Neuromolecular Med.* (2018) 20:376–85. doi: 10.1007/s12017-018-8499-5
3. Goedert M. Alpha-synuclein and neurodegenerative diseases. *Nat Rev Neurosci.* (2001) 2:492–501. doi: 10.1038/35081564
 4. Haghshomar M, Rahmani F, Hadi Aarabi M, Shahjouei S, Sobhani S, Rahmani M. White matter changes correlates of peripheral neuroinflammation in patients with Parkinson's disease. *Neuroscience* (2017). doi: 10.1016/j.neuroscience.2017.10.050. [Epub ahead of print].
 5. Jellinger KA, Seppi K, Wenning G, Poewe W. Impact of coexistent Alzheimer pathology on the natural history of Parkinson's disease. *J Neural Transm.* (2002) 109:329–39. doi: 10.1007/s007020200027
 6. Dolatshahi M, Pourmirbabaei S, Kamalian A, Ashraf-Ganjouei A, Yaseri M, Aarabi MH. Longitudinal alterations of alpha-synuclein, amyloid beta, total, and phosphorylated tau in cerebrospinal fluid and correlations between their changes in Parkinson's disease. *Front Neurol.* (2018) 9:560. doi: 10.3389/fneur.2018.00560
 7. Berg D, Postuma RB, Bloem B, Chan P, Dubois B, Gasser T, et al. Time to redefine PD? Introductory statement of the MDS task force on the definition of Parkinson's disease. *Mov Disord.* (2014) 29:454–62. doi: 10.1002/mds.25844
 8. Postuma RB, Berg D. Advances in markers of prodromal Parkinson disease. *Nat Rev Neurol.* (2016) 12:622–34. doi: 10.1038/nrneurol.2016.152
 9. Bassil F, Fernagut P-O, Bezard E, Meissner WG. Insulin, IGF-1 and GLP-1 signaling in neurodegenerative disorders: targets for disease modification? *Prog Neurobiol.* (2014) 118:1–18. doi: 10.1016/j.pneurobio.2014.02.005
 10. Athauda D, Foltynie T. Insulin resistance and Parkinson's disease: a new target for disease modification? *Prog Neurobiol.* (2016) 145:98–120. doi: 10.1016/j.pneurobio.2016.10.001
 11. Fernandez AM, Torres-Alemán I. The many faces of insulin-like peptide signalling in the brain. *Nat Rev Neurosci.* (2012) 13:225–39. doi: 10.1038/nrn3209
 12. Labandeira-Garcia JL, Costa-Besada MA, Labandeira CM, Villar-Cheda B, Rodríguez-Perez AI. Insulin-like growth factor-1 and neuroinflammation. *Front Aging Neurosci.* (2017) 9:365. doi: 10.3389/fnagi.2017.00365
 13. Fernandez A, Jimenez S, Mecha M, Davila D, Guaza C, Vitorica J, et al. Regulation of the phosphatase calcineurin by insulin-like growth factor I unveils a key role of astrocytes in Alzheimer's pathology. *Mol Psychiatry* (2012) 17:705–18. doi: 10.1038/mp.2011.128
 14. Gasperi M, Castellano A. Growth hormone/insulin-like growth factor I axis in neurodegenerative diseases. *J Endocrinol Invest.* (2010) 33:587–91. doi: 10.1007/BF03346653
 15. Procaccini C, Santopaolo M, Faicchia D, Colamattéo A, Formisano L, de Candia P, et al. Role of metabolism in neurodegenerative disorders. *Metab Clin Exp.* (2016) 65:1376–90. doi: 10.1016/j.metabol.2016.05.018
 16. Sutherland GT, Matigian NA, Chalk AM, Anderson MJ, Silburn PA, Mackay-Sim A, et al. A cross-study transcriptional analysis of Parkinson's disease. *PLoS ONE* (2009) 4:e4955. doi: 10.1371/journal.pone.0004955
 17. Godau J, Herfurth M, Kattner B, Gasser T, Berg D. Increased serum insulin-like growth factor 1 in early idiopathic Parkinson's disease. *J Neurol Neurosurg Psychiatry* (2010) 81:536–8. doi: 10.1136/jnnp.2009.175752
 18. Mashayekhi F, Mirzajani E, Najji M, Azari M. Expression of insulin-like growth factor-1 and insulin-like growth factor binding proteins in the serum and cerebrospinal fluid of patients with Parkinson's disease. *J Clin Neurosci.* (2010) 17:623–7. doi: 10.1016/j.jocn.2009.08.013
 19. Godau J, Knäuel K, Weber K, Brockmann K, Maetzler W, Binder G, et al. Serum insulinlike growth factor 1 as possible marker for risk and early diagnosis of Parkinson disease. *Arch Neurol.* (2011) 68:925–31. doi: 10.1001/archneurol.2011.129
 20. Picillo M, Erro R, Santangelo G, Pivonello R, Longo K, Pivonello C, et al. Insulin-like growth factor-1 and progression of motor symptoms in early, drug-naive Parkinson's disease. *J Neurol.* (2013) 260:1724–30. doi: 10.1007/s00415-013-6851-0
 21. Bernhard FP, Heinzel S, Binder G, Weber K, Apel A, Roeben B, et al. Insulin-like growth factor 1 (IGF-1) in Parkinson's disease: Potential as trait-, progression- and prediction marker and confounding factors. *PLoS ONE* (2016) 11:e0150552. doi: 10.1371/journal.pone.0150552
 22. Picillo M, Pivonello R, Santangelo G, Pivonello C, Savastano R, Auriemma R, et al. Serum IGF-1 is associated with cognitive functions in early, drug-naive Parkinson's disease. *PLoS ONE* (2017) 12:e0186508. doi: 10.1371/journal.pone.0186508
 23. Hong M, Lee VM-Y. Insulin and insulin-like growth factor-1 regulate tau phosphorylation in cultured human neurons. *J Biol Chem.* (1997) 272:19547–53. doi: 10.1074/jbc.272.31.19547
 24. Guan J, Krishnamurthi R, Waldvogel HJ, Faull RL, Clark R, Gluckman P. N-terminal tripeptide of IGF-1 (GPE) prevents the loss of TH positive neurons after 6-OHDA induced nigral lesion in rats. *Brain Res.* (2000) 859:286–92. doi: 10.1016/S0006-8993(00)01988-0
 25. Offen D, Shtatif B, Hadad D, Weizman A, Melamed E, Gil-Ad I. Protective effect of insulin-like-growth-factor-1 against dopamine-induced neurotoxicity in human and rodent neuronal cultures: possible implications for Parkinson's disease. *Neurosci Lett.* (2001) 316:129–32. doi: 10.1016/S0304-3940(01)02344-8
 26. Krishnamurthi R, Stott S, Maingay M, Faull RL, McCarthy D, Gluckman P, et al. N-terminal tripeptide of IGF-1 improves functional deficits after 6-OHDA lesion in rats. *Neuroreport* (2004) 15:1601–4. doi: 10.1097/01.wnr.0000127461.15985.07
 27. Quesada A, Lee BY, Micevych PE. PI3 kinase/Akt activation mediates estrogen and IGF-1 nigral DA neuronal neuroprotection against a unilateral rat model of Parkinson's disease. *Dev Neurobiol.* (2008) 68:632–44. doi: 10.1002/dneu.20609
 28. Kao S-Y. Rescue of α -synuclein cytotoxicity by insulin-like growth factors. *Biochem Biophys Res Commun.* (2009) 385:434–8. doi: 10.1016/j.bbrc.2009.05.089
 29. Jabir NR, Firoz CK, Baeesa SS, Ashraf GM, Akhtar S, Kamal W, et al. Synopsis on the linkage of Alzheimer's and Parkinson's disease with chronic diseases. *CNS Neurosci Ther.* (2015) 21:1–7. doi: 10.1111/cns.12344
 30. Compta Y, Parkkinen L, O'sullivan SS, Vandrovcova J, Holton JL, Collins C, et al. Lewy-and Alzheimer-type pathologies in Parkinson's disease dementia: which is more important? *Brain* (2011) 134:1493–505. doi: 10.1093/brain/awr031
 31. Abbasi N, Mohajer B, Abbasi S, Hasanabadi P, Abdolizadeh A, Rajimehr R. Relationship between cerebrospinal fluid biomarkers and structural brain network properties in Parkinson's disease. *Mov Disord.* (2018) 33:431–9. doi: 10.1002/mds.27284
 32. Aung WY, Mar S, Benzinger TL. Diffusion tensor MRI as a biomarker in axonal and myelin damage. *Imaging Med.* (2013) 5:427–40. doi: 10.2217/iim.13.49
 33. Hall JM, Ehgoetz Martens KA, Walton CC, O'Callaghan C, Keller PE, Lewis SJ, et al. Diffusion alterations associated with Parkinson's disease symptomatology: a review of the literature. *Parkinsonism Relat Disord.* (2016) 33:12–26. doi: 10.1016/j.parkreldis.2016.09.026
 34. Yeh F-C, Badre D, Verstynen T. Connectometry: a statistical approach harnessing the analytical potential of the local connectome. *Neuroimage* (2016) 125:162–71. doi: 10.1016/j.neuroimage.2015.10.053
 35. Yeh F-C, Vettel JM, Singh A, Poczos B, Grafton ST, Erickson KI, et al. Quantifying differences and similarities in whole-brain white matter architecture using local connectome fingerprints. *PLoS Comput Biol.* (2016) 12:e1005203. doi: 10.1371/journal.pcbi.1005203
 36. Wozniak JR, Muetzel RL. What does diffusion tensor imaging reveal about the brain and cognition in fetal alcohol spectrum disorders? *Neuropsychol Rev.* (2011) 21:133–47. doi: 10.1007/s11065-011-9162-1
 37. Leemans A, Jeurissen B, Sijbers J, Jones DK. ExploreDTI: a graphical toolbox for processing, analyzing and visualizing diffusion MR data. In: *Proceedings of the International Society for Magnetic Resonance in Medicine 17th Annual Meeting*. Honolulu, HI (2009). p. 3536.
 38. Yeh FC, Tseng WY. NTU-90: a high angular resolution brain atlas constructed by q-space diffeomorphic reconstruction. *Neuroimage* (2011) 58:91–9. doi: 10.1016/j.neuroimage.2011.06.021
 39. Montine TJ, Shi M, Quinn JF, Peskind ER, Craft S, Ghingina C, et al. CSF A β 42 and tau in Parkinson's disease with cognitive impairment. *Mov Disord.* (2010) 25:2682–5. doi: 10.1002/mds.23287
 40. Shi M, Bradner J, Hancock AM, Chung KA, Quinn JF, Peskind ER, et al. Cerebrospinal fluid biomarkers for Parkinson disease diagnosis and progression. *Ann Neurol.* (2011) 69:570–80. doi: 10.1002/ana.22311
 41. Kang JH, Irwin DJ, Chen-Plotkin AS, Siderowf A, Caspell C, Coffey CS, et al. Association of cerebrospinal fluid beta-amyloid 1-42, T-tau, P-tau181, and alpha-synuclein levels with clinical features of drug-naive

- patients with early Parkinson disease. *JAMA Neurol.* (2013) 70:1277–87. doi: 10.1001/jamaneurol.2013.3861
42. Kang, J-H, Mollenhauer B, Coffey CS, Toledo JB, Weintraub D, Galasko DR, et al. CSF biomarkers associated with disease heterogeneity in early Parkinson's disease: the Parkinson's Progression Markers Initiative study. *Acta Neuropathol.* (2016) 131:935–49. doi: 10.1007/s00401-016-1552-2
 43. Adem A, Jossan S, d'Argy R, Gillberg P, Nordberg A, Winblad B, et al. Insulin-like growth factor 1 (IGF-1) receptors in the human brain: quantitative autoradiographic localization. *Brain Res.* (1989) 503:299–303. doi: 10.1016/0006-8993(89)91678-8
 44. De Keyser J, Wilczak N, Goossens A. Insulin-like growth factor-I receptor densities in human frontal cortex and white matter during aging, in Alzheimer's disease, and in Huntington's disease. *Neurosci Lett.* (1994) 172:93–6. doi: 10.1016/0304-3940(94)90670-X
 45. Hinkley LB, Marco EJ, Findlay AM, Honma S, Jeremy RJ, Strominger Z, et al. The role of corpus callosum development in functional connectivity and cognitive processing. *PLoS ONE* (2012) 7:e39804. doi: 10.1371/journal.pone.0039804
 46. Fabri M, Pierpaoli C, Barbaresi P, Polonara G. Functional topography of the corpus callosum investigated by DTI and fMRI. *World J Radiol.* (2014) 6:895. doi: 10.4329/wjr.v6.i12.895
 47. Bubb EJ, Metzler-Baddeley C, Aggleton JP. The cingulum bundle: anatomy, function, and dysfunction. *Neurosci Biobehav Rev.* (2018) 92:104–27. doi: 10.1016/j.neubiorev.2018.05.008
 48. Bozzali M, Giulietti G, Basile B, Serra L, Spanò B, Perri R, et al. Damage to the cingulum contributes to Alzheimer's disease pathophysiology by deafferentation mechanism. *Hum Brain Mapp.* (2012) 33:1295–308. doi: 10.1002/hbm.21287
 49. Demey I, Ventrice F, Rojas G, Somale V, Zubiri V. Alzheimer's disease dementia involves the corpus callosum and the cingulum: a diffusion tensor imaging study. *Alzheimers Dementia* (2015) 11:P409–10. doi: 10.1016/j.jalz.2015.06.368
 50. Rémy F, Vayssière N, Saint-Aubert L, Barbeau E, Pariente J. White matter disruption at the prodromal stage of Alzheimer's disease: relationships with hippocampal atrophy and episodic memory performance. *Neuroimage* (2015) 7:482–92. doi: 10.1016/j.nicl.2015.01.014
 51. Wang Z, Dai Z, Shu H, Liu D, Guo Q, He Y, et al. Cortical thickness and microstructural white matter changes detect amnesic mild cognitive impairment. *J Alzheimers Dis.* (2017) 56:415–28. doi: 10.3233/JAD-16 0724
 52. Yu J, Lam CL, Lee TM. White matter microstructural abnormalities in amnesic mild cognitive impairment: a meta-analysis of whole-brain and ROI-based studies. *Neurosci Biobehav Rev.* (2017)83:405–16. doi: 10.1016/j.neubiorev.2017.10.026
 53. Chua TC, Wen W, Slavin MJ, Sachdev PS. Diffusion tensor imaging in mild cognitive impairment and Alzheimer's disease: a review. *Curr Opin Neurol.* (2008) 21:83–92. doi: 10.1097/WCO.0b013e3282f4594b
 54. Hattori T, Orimo S, Aoki S, Ito K, Abe O, Amano A, et al. Cognitive status correlates with white matter alteration in Parkinson's disease. *Hum Brain Mapp.* (2012) 33:727–39. doi: 10.1002/hbm.21245
 55. Kamagata K, Motoi Y, Abe O, Shimoji K, Hori M, Nakanishi A, et al. White matter alteration of the cingulum in Parkinson disease with and without dementia: evaluation by diffusion tensor tract-specific analysis. *AJNR Am J Neuroradiol.* (2012) 33:890–5. doi: 10.3174/ajnr.A2860
 56. Deng B, Zhang Y, Wang L, Peng K, Han L, Nie K, et al. Diffusion tensor imaging reveals white matter changes associated with cognitive status in patients with Parkinson's disease. *Am J Alzheimers Dis Other Dement.* (2013) 28:154–64. doi: 10.1177/1533317512470207
 57. Perea RD, Rada RC, Wilson J, Vidoni ED, Morris JK, Lyons KE, et al. A comparative white matter study with Parkinson's disease, Parkinson's disease with dementia and Alzheimer's disease. *J Alzheimers Dis Parkinsonism* (2013) 3:123. doi: 10.4172/2161-0460.1000123
 58. Chondrogiorgi M, Astrakas LG, Zikou AK, Weis L, Xydis VG, Antonini A, et al. Multifocal alterations of white matter accompany the transition from normal cognition to dementia in Parkinson's disease patients. *Brain Imaging Behav.* (2018) 1–9. doi: 10.1007/s11682-018-9863-7. [Epub ahead of print].
 59. Muslimović D, Post B, Speelman JD, Schmand B. Cognitive profile of patients with newly diagnosed Parkinson disease. *Neurology* (2005) 65:1239–45. doi: 10.1212/01.wnl.0000180516.69442.95
 60. Williams-Gray CH, Evans JR, Goris A, Foltynie T, Ban M, Robbins TW, et al. The distinct cognitive syndromes of Parkinson's disease: 5 year follow-up of the CamPaIGN cohort. *Brain* (2009) 132:2958–69. doi: 10.1093/brain/awp245
 61. Irwin DJ, Lee VM-Y, Trojanowski JQ. Parkinson's disease dementia: convergence of α -synuclein, tau and amyloid- β pathologies. *Nat Rev Neurosci.* (2013) 14:626. doi: 10.1038/nrn3549
 62. Irwin DJ, White MT, Toledo JB, Xie SX, Robinson JL, Van Deerlin V, et al. Neuropathologic substrates of Parkinson disease dementia. *Ann Neurol.* (2012) 72:587–98. doi: 10.1002/ana.23659
 63. Pellecchia MT, Santangelo G, Picillo M, Pivonello R, Longo K, Pivonello C, et al. Insulin-like growth factor-1 predicts cognitive functions at 2-year follow-up in early, drug-naïve Parkinson's disease. *Eur J Neurol.* (2014) 21:802–7. doi: 10.1111/ene.12137
 64. Ma J, Jiang Q, Xu J, Sun Q, Qiao Y, Chen W, et al. Plasma insulin-like growth factor 1 is associated with cognitive impairment in Parkinson's disease. *Dementia Geriatr Cogn Disord.* (2015) 39:251–6. doi: 10.1159/000371510
 65. Numao A, Suzuki K, Miyamoto M, Miyamoto T, Hirata K. Clinical correlates of serum insulin-like growth factor-1 in patients with Parkinson's disease, multiple system atrophy and progressive supranuclear palsy. *Parkinsonism Relat Disord.* (2014) 20:212–6. doi: 10.1016/j.parkreldis.2013.11.005
 66. Ebert AD, Beres AJ, Barber AE, Svendsen CN. Human neural progenitor cells over-expressing IGF-1 protect dopamine neurons and restore function in a rat model of Parkinson's disease. *Exp Neurol.* (2008) 209:213–23. doi: 10.1016/j.expneurol.2007.09.022
 67. Bianchi VE, Locatelli V, Rizzi L. Neurotrophic and neuroregenerative effects of GH/IGF1. *Int J Mol Sci.* (2017) 18:1–25. doi: 10.3390/ijms18112441
 68. Greenland J, Williams-Gray C, Barker R. The clinical heterogeneity of Parkinson's disease and its therapeutic implications. *Eur J Neurosci.* (2018). doi: 10.1111/ejn.14094. [Epub ahead of print].
 69. Chan L-L, Ng K-M, Rumpel H, Fook-Chong S, Li H-H, Tan E-K. Transcallosal diffusion tensor abnormalities in predominant gait disorder parkinsonism. *Parkinsonism Relat Disord.* (2014) 20:53–9. doi: 10.1016/j.parkreldis.2013.09.017
 70. Lee E, Lee JE, Yoo K, Hong JY, Oh J, Sunwoo MK, et al. Neural correlates of progressive reduction of bradykinesia in *de novo* Parkinson's disease. *Parkinsonism Relat Disord.* (2014) 20:1376–81. doi: 10.1016/j.parkreldis.2014.09.027
 71. Canu E, Agosta F, Sarasso E, Volontè MA, Basaia S, Stojkovic T, et al. Brain structural and functional connectivity in Parkinson's disease with freezing of gait. *Hum Brain Mapp.* (2015) 36:5064–78. doi: 10.1002/hbm.22994
 72. Schweder PM, Hansen PC, Green AL, Quaghebeur G, Stein J, Aziz TZ. Connectivity of the pedunculopontine nucleus in parkinsonian freezing of gait. *Neuroreport* (2010) 21:914–6. doi: 10.1097/WNR.0b013e32833ce5f1
 73. Fling BW, Cohen RG, Mancini M, Nutt JG, Fair DA, Horak FB. Asymmetric pedunculopontine network connectivity in parkinsonian patients with freezing of gait. *Brain* (2013) 136:2405–18. doi: 10.1093/brain/awt172
 74. Vercruyse S, Leunissen I, Vervoort G, Vandenbergh W, Swinnen S, Nieuwboer A. Microstructural changes in white matter associated with freezing of gait in Parkinson's disease. *Mov Disord.* (2015) 30:567–76. doi: 10.1002/mds.26130
 75. Camicioli R, Gee M, Bouchard TP, Fisher NJ, Hanstock CC, Emery DJ, et al. Voxel-based morphometry reveals extra-nigral atrophy patterns associated with dopamine refractory cognitive and motor impairment in parkinsonism. *Parkinsonism Relat Disord.* (2009) 15:187–95. doi: 10.1016/j.parkreldis.2008.05.002
 76. Pereira JB, Junqué C, Martí MJ, Ramirez-Ruiz B, Bartrés-Faz D, Tolosa E. Structural brain correlates of verbal fluency in Parkinson's disease. *Neuroreport* (2009) 20:741–4. doi: 10.1097/WNR.0b013e328329370b
 77. Nishio Y, Hirayama K, Takeda A, Hosokai Y, Ishioka T, Suzuki K, et al. Corticolimbic gray matter loss in Parkinson's disease without dementia. *Eur J Neurol.* (2010) 17:1090–7. doi: 10.1111/j.1468-1331.2010.02980.x
 78. Özdinler PH, Macklis JD. IGF-I specifically enhances axon outgrowth of corticospinal motor neurons. *Nat Neurosci.* (2006) 9:1371–81. doi: 10.1038/nn1789
 79. Mason J, Jones J, Taniike M, Morell P, Suzuki K, Matsushima G. Mature oligodendrocyte apoptosis precedes IGF-1 production and oligodendrocyte progenitor accumulation and differentiation during

- demyelination/remyelination. *J Neurosci Res.* (2000) 61:251–62. doi: 10.1002/1097-4547(20000801)61:3<251::AID-JNR3>3.0.CO;2-W
80. Ghazi Sherbaf F, Rostam Abadi Y, Mojtahed Zadeh M, Ashraf-Ganjouei A, Sanjari Moghaddam H, Aarabi MH. Microstructural changes in patients with Parkinson's disease comorbid with REM sleep behaviour disorder and depressive symptoms. *Front Neurol.* (2018) 9:444. doi: 10.3389/fneur.2018.00441
81. Zhang H, Wang Y, Lu T, Qiu B, Tang Y, Ou S, et al. Differences between generalized q-sampling imaging and diffusion tensor imaging in the preoperative visualization of the nerve fiber tracts within peritumoral edema in brain. *Neurosurgery* (2013) 73:1044–53. doi: 10.1227/NEU.0000000000000146

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Ghazi Sherbaf, Mohajer, Ashraf-Ganjouei, Mojtahed Zadeh, Javinani, Sanjari Moghaddam, Shirin Shandiz and Aarabi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.