



Inhibitors of Human ABCG2: From Technical Background to Recent Updates With Clinical Implications

Yu Toyoda, Tappei Takada* and Hiroshi Suzuki

Department of Pharmacy, The University of Tokyo Hospital, Faculty of Medicine, The University of Tokyo, Tokyo, Japan

OPEN ACCESS

Edited by:

Yunkai Zhang,
Vanderbilt University Medical Center,
United States

Reviewed by:

Tomohiro Terada,
Shiga University of Medical Science,
Japan

Ning Ji,
Tianjin Medical University, China
Shinobu Ohnuma,
Tohoku University, Japan

*Correspondence:

Tappei Takada
tappei-ky@umin.ac.jp

Specialty section:

This article was submitted to
Experimental Pharmacology
and Drug Discovery,
a section of the journal
Frontiers in Pharmacology

Received: 25 December 2018

Accepted: 19 February 2019

Published: 05 March 2019

Citation:

Toyoda Y, Takada T and Suzuki H
(2019) Inhibitors of Human ABCG2:
From Technical Background
to Recent Updates With Clinical
Implications.
Front. Pharmacol. 10:208.
doi: 10.3389/fphar.2019.00208

The ATP-binding cassette transporter G2 (ABCG2; also known as breast cancer resistance protein, BCRP) has been suggested to be involved in clinical multidrug resistance (MDR) in cancer like other ABC transporters such as ABCB1 (*P*-glycoprotein). As an efflux pump exhibiting a broad substrate specificity localized on cellular plasma membrane, ABCG2 excretes a variety of endogenous and exogenous substrates including chemotherapeutic agents, such as mitoxantrone and several tyrosine kinase inhibitors. Moreover, in the normal tissues, ABCG2 is expressed on the apical membranes and plays a pivotal role in tissue protection against various xenobiotics. For this reason, ABCG2 is recognized to be an important determinant of the pharmacokinetic characteristics of its substrate drugs. Although the clinical relevance of reversing the ABCG2-mediated MDR has been inconclusive, an appropriate modulation of ABCG2 function during chemotherapy should logically enhance the efficacy of anti-cancer agents by overcoming the MDR phenotype and/or improving their pharmacokinetics. To confirm this possibility, considerable efforts have been devoted to developing ABCG2 inhibitors, although there is no clinically available substance for this purpose. As a clue for addressing this issue, this mini-review provides integrated information covering the technical backgrounds necessary to evaluate the ABCG2 inhibitory effects on the target compounds and a current update on the ABCG2 inhibitors. This essentially includes our recent findings, as we serendipitously identified febuxostat, a well-used agent for hyperuricemia as a strong ABCG2 inhibitor, that possesses some promising potentials. We hope that an overview described here will add value to further studies involving in the multidrug transporters.

Keywords: BCRP, cancer chemotherapy, drug repurposing, febuxostat, Ko143, multidrug resistance, tumor lysis syndrome, vesicle transport

INTRODUCTION

Two decades ago, the ABC transporter G2 (ABCG2) was discovered in drug-resistant cancer cells and human placenta (Allikmets et al., 1998; Doyle et al., 1998; Miyake et al., 1999). Thereafter, many studies were conducted to determine the role of ABCG2 in developing MDR in cancer. Moreover, in the first decade, *in vivo* studies using *Abcg2* knockout mice (Jonker et al., 2002)

Abbreviations: ABC, ATP-binding cassette; BBB, blood–brain barrier; CML, chronic myeloid leukemia; EC₅₀, half-maximal effective concentration; FTC, fumitremorgin C; IC₅₀, half-maximal inhibitory concentration; MDR, multidrug resistance; TKI, tyrosine kinase inhibitor, TLS, tumor lysis syndrome.

coupled with biochemical characterizations revealed the importance of ABCG2 in the biological defense mechanisms against xenobiotics (Vlaming et al., 2009). Indeed, ABCG2—a 655-amino acid protein working as a homodimer on cellular plasma membranes (Robey et al., 2009)—is expressed not only in cancer cells but also in several normal tissues, such as brush border membranes of epithelium in the intestine and of proximal tubules in the kidney, bile canalicular membranes of the hepatocytes in the liver, luminal membranes of the mammary gland epithelium, and blood-facing membranes of the endothelial cells forming the BBB. In these tissues ABCG2 plays a pivotal role in the extrusion of various endogenous and exogenous substrates including drugs (Mizuno et al., 2004, 2007; Adachi et al., 2005; Hirano et al., 2005; Jonker et al., 2005; Ando et al., 2007). Hence, this transporter is recognized as an important determinant of the pharmacokinetic characteristics profiles of various drugs (Giacomini et al., 2010).

In the next decade, after identifying ABCG2 as a physiologically important urate transporter, a positive relationship between ABCG2 dysfunction and increased risk of human diseases, such as gout and hyperuricemia was revealed (Matsuo et al., 2009; Woodward et al., 2009; Ichida et al., 2012; Higashino et al., 2017). In addition to the sulfate conjugates of endogenous steroids (Suzuki et al., 2003) and porphyrins (Zhou et al., 2005; Robey et al., 2009), phytoestrogen sulfate conjugates (van de Wetering and Saphth, 2012) and a uremic toxin indoxyl sulfate (Takada et al., 2018) were added in the growing list of ABCG2 substrates. Contrary to these advances in understanding the pathophysiological importance of ABCG2, the clinical relevance of reversing ABCG2-mediated MDR has been inconclusive (Robey et al., 2018).

ABCG2 overexpression can render the cancer cells resistant to the ABCG2 substrate chemotherapy agents, such as mitoxantrone, doxorubicin, SN-38, and several TKIs. To the best of our knowledge, no published clinical trial has ever succeeded in reversing the ABCG2-mediated MDR. This is because, despite a lot of efforts in ABCG2 inhibitor development, chemical knock-out/down of ABCG2 in clinical situations has not been achieved yet due to the lack of an appropriate candidate molecule. We herein describe some well-used experimental systems to evaluate the ABCG2 inhibitory activity, followed by a recent update on the ABCG2 inhibitors that includes a potent substance, febuxostat.

TECHNICAL BACKGROUND FOR FUNCTIONAL VALIDATION

Various experimental models are available to examine the functions of the ABC transporters. Mainly focusing on ABCG2, with a current update this section introduces some *in vitro* and *in vivo* models that have been used in ABC transporter field. Broadly, the *in vitro* models are classified into two types, namely membrane-based systems and cell-based systems (Figure 1).

Plasma Membrane Vesicle-Based Methods

Preparation of Plasma Membrane Vesicles

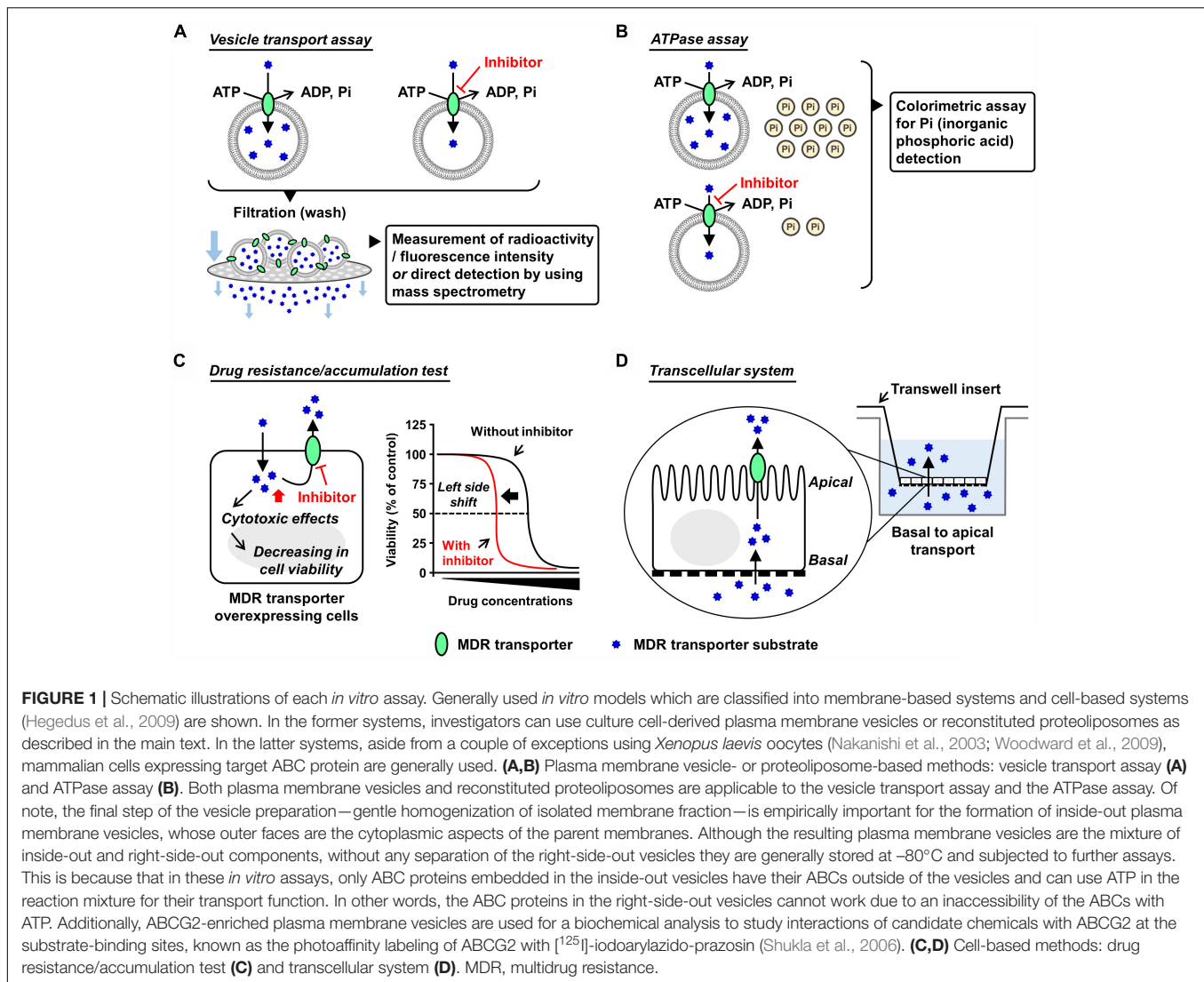
In mammals, most of the ABC transporters are membrane proteins and work as an efflux pump involved in the transport of its substrates from the cytosol, either to the extracellular space or into organelles by an ATP-dependent manner. Therefore, isolation of the target ABC protein-enriched cell membrane is the first step for biochemical analyses. For ABCG2, sucrose density gradient ultracentrifugation for the isolation of plasma membrane fraction is generally employed to prepare plasma membrane vesicles from ABCG2-expressing cells (related notes are inscribed in the legend of Figure 1). For this purpose, not only mammalian cells but also insect cells [e.g., baculovirus-infected Sf9 cells (Saito et al., 2006)] could be used as host cells. Nonetheless, for easy and convenient preparation of ABCG2-expressing cells, we here recommend plasmid-based overexpression in non-polarized cells exhibiting high transfection efficiency, such as HEK293 cells (Miyata et al., 2016).

Vesicle Transport Assay

Vesicle transport assay is a well-established *in vitro* method employed to quantitatively evaluate ABC transporter function. The presence of ATP-dependent active transport across the cell membrane was directly proved using this method (Ishikawa, 1989). In this assay, ATP-regeneration components—enough amount of creatine phosphate and creatine kinase—are employed to maintain ATP levels in the reaction mixtures during prolonged incubation, and AMP is used as an alternative of ATP for the ATP-deficient controls. After incubation for the transport reaction, the plasma membrane vesicles are washed by filtration and then the intravesicularly accumulated substances are detected. To make this process more convenient and sensitive, radiolabeled or fluorescent substrates are usually used; alternatively, mass spectrometry is used (Toyoda et al., 2016; Takada et al., 2018). With ABCG2, [¹⁴C]-urate (Miyata et al., 2016; Stiburkova et al., 2016; Higashino et al., 2017) and [³H]-estrone sulfate (Suzuki et al., 2003) are well-used radiolabeled substrates that exhibit comparatively lower background signal for the quantitative detection due to their relatively hydrophilic properties. Additionally, non-radiolabeled experiments are conducted by the combined use of hematoporphyrin (a fluorescent ABCG2 substrate) and gel filtration techniques (Tamura et al., 2006).

ATPase Assay

Since ABC protein is driven by the free energy of ATP hydrolysis, ATPase activity is recognized as an indicator of the substrate transport. In this assay, the release of inorganic phosphate from ATP coupled with the transport of substrates is estimated using a colorimetric method, such as malachite green procedure (Baykov et al., 1988). This catalytic assay is relatively convenient for estimating the activity of some ABC proteins that prefer lipophilic compounds as their substrates because the non-specifically adsorbed substrates on the vesicles interfere with the measurement of direct transport. Considering that ABCB1 activity has been well



studied based on the ATPase assay, this method will be appropriate when investigators would need to compare the inhibitory effects of target compounds on ABCB1 and ABCG2 (Zhang et al., 2016; Guo et al., 2018). Nonetheless, since the ATPase assay does not evaluate the direct transport, regarding ABCG2, we recommend the vesicle transport assay for more precise evaluation.

Proteoliposome-Based Methods

An artificial lipid membrane system characterized by the reconstitution of purified ABC protein into proteoliposomes (Ambudkar et al., 1998; Jackson et al., 2018) is also a powerful technique. The ABC protein-contained proteoliposomes can be used as an alternative to the plasma membrane vesicles, and their detailed preparation methods are described previously (Geertsma et al., 2008). Additionally, this approach serves as the first choice in the functional studies involving the ABC proteins localized on the organelle membrane (Okamoto et al., 2018).

Cell-Based Methods

Drug Resistance/Accumulation Test

The cells overexpressing MDR machinery show lower sensitivity against its substrate drugs exhibiting cytotoxic or anti-proliferative effects compared to their parent cells, indicating that the ABC protein-expressing cells have higher half maximal effective concentration (EC_{50}) values for cytotoxic transporter substrates. In such situations, co-treatment of the transporter inhibitors with the substrate decreases the EC_{50} values, which is depicted as a left-side shift of cell viability curve in the cytotoxic assay. Additionally, if fluorescence transporter substrates are available, flow cytometry analyses addressing their intracellular accumulation will be useful for inhibitor screening (Murakami et al., 2017; Wu et al., 2017).

Transcellular System

To investigate the transcellular transport of substances, mono-layer culture of polarized cells expressing target transporter(s) in Transwell® inserts system has been used (Hegedus et al.,

2009). For instance, using double-transfected Madin-Darby canine kidney II cells overexpressing both organic anion-transporting polypeptide 1B1 (OATP1B1, a basal uptake transporter) and ABCG2 (an apical efflux transporter), an earlier study has observed an enhanced vectorial transport of [³H]estrone sulfate—an ABCG2 substrate that cannot passively penetrate the plasma membrane—from the basal to the apical side (Matsushima et al., 2005). A similar strategy employing ABCG2-expressing polarized cells was used for *in vitro* studies investigating the active secretion of drugs and toxins into milk via ABCG2 (Wassermann et al., 2013; Ito et al., 2015). Of note, in this transcellular system, endogenous transporters and metabolic reactions in the cells may affect apparent transport activities of target transporters.

In vivo Evaluation Methods

Xenograft Models

Athymic nude mice models with MDR-xenografts have been used to test whether co-administration of a potential MDR inhibitor with an anti-cancer agent can reverse the MDR phenotype (Tiwari et al., 2013; Zhang et al., 2017, 2018). In such models, the obtained results could be affected by the difference in the origin of the transplanted cancer cells. This concern might be important while xenograft models are studied.

Focusing on Pharmacokinetic Characteristics

To examine the *in vivo* effects of different chemicals on the target ABC transporter, its pharmacokinetic role has been focused (Robey et al., 2018). With ABCG2, the pharmacokinetic parameters related to the intestinal absorption or the brain distribution of ABCG2 substrate drugs will be good indicators for ABCG2 activity *in vivo*. For example, pre-dosing of enough quantity of ABCG2 inhibitors increased the bioavailability of sulphasalazine—an ABCG2 substrate—in wild-type mice. This was, however, not observed in *Abcg2*-knockout mice, suggesting the *in vivo* inhibition of *Abcg2* (Kusuhara et al., 2012; Miyata et al., 2016). Interestingly, the utility of a combination of brain-specific firefly luciferase transgenic mice and D-luciferin, a chemiluminescent luciferase substrate transported by ABCG2 (Zhang et al., 2007) to investigate the *in vivo* inhibitory effects of test compounds on ABCG2 in the BBB was reported (Bakhsheshian et al., 2016). Further methodological progress will aid evaluation of *in vivo* ABCG2 function.

Structure-Based *in silico* Approaches

A main approach to abolish MDR is to discover specific inhibitors of the drug-efflux pump. For this purpose, quantitative structure-activity relationship (QSAR) analysis among the series of compounds can serve for the design of lead inhibitors (Nicolle et al., 2009; Ishikawa et al., 2012; Marighetti et al., 2013; Shukla et al., 2014). With ABCG2, since three-dimensional structures of this protein determined by cryo-electron microscopy (EM) were very recently presented (Taylor et al., 2017; Jackson et al., 2018), a deeper understanding of the chemicals–ABCG2 interactions will be achieved as described below.

HISTORY AND RECENT UPDATE OF THE ABCG2 INHIBITORS

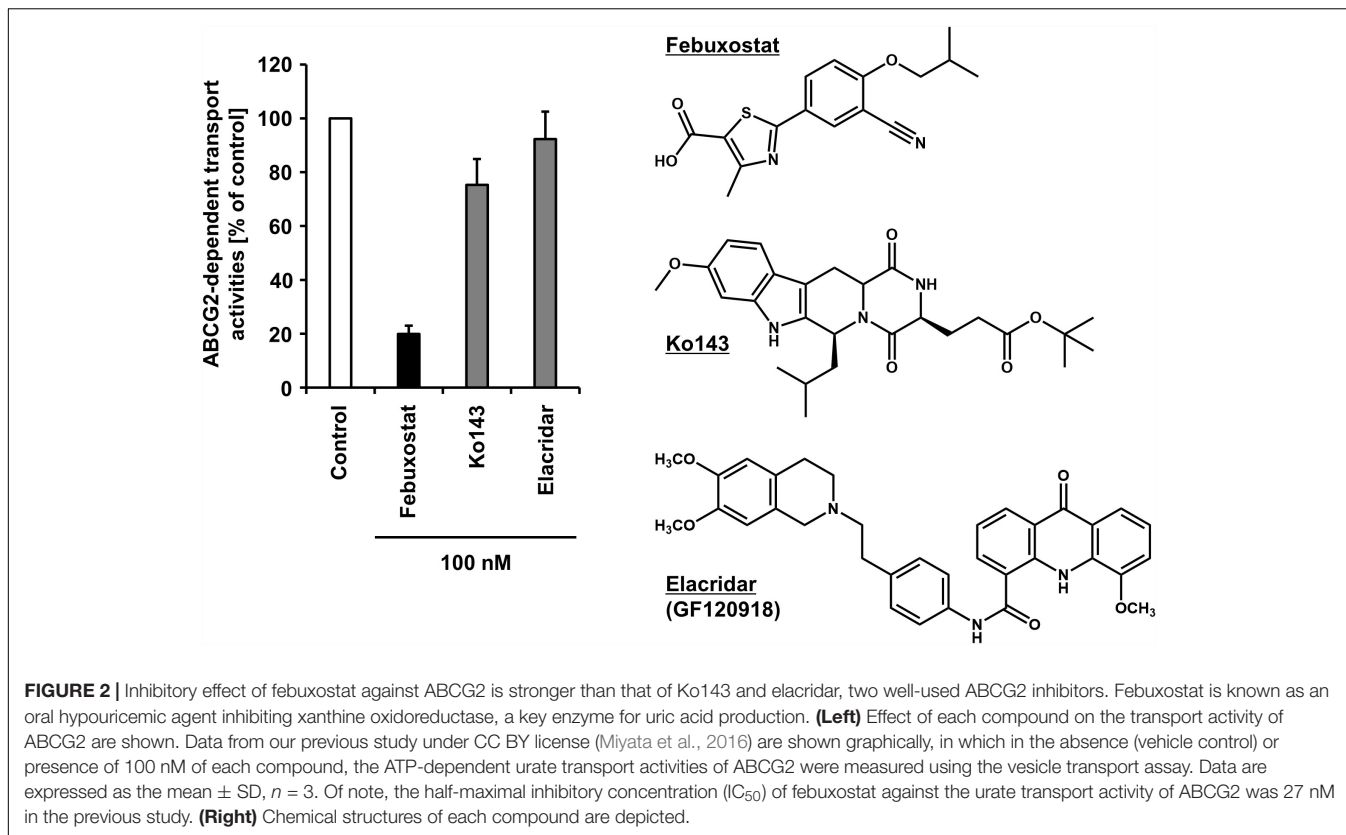
As a MDR machinery in cancer cells and an important drug gatekeeper in tissues like the intestine and the brain, ABCG2 is involved in the efficacy of cancer chemotherapy in patients treated with ABCG2 substrate anti-cancer drugs. To achieve appropriate modulation of ABCG2 by small molecules, the inhibitory potency of various compounds against ABCG2 activity has been extensively evaluated. In this section, we highlight the history and recent update of ABCG2 inhibitors.

Overview of the History of ABCG2 Inhibitors

The first ABCG2 inhibitor reported was FTC, a mycotoxin produced by *Aspergillus fumigatus* (Rabindran et al., 1998, 2000). The *in vivo* use of FTC was unfortunately precluded due to its neurotoxicity. Among the FTC derivatives, Ko143 was identified as a highly potent ABCG2 inhibitor *in vivo* as it was less neurotoxic than the native FTC and was not overtly toxic to mice (Allen et al., 2002). Cell-based assays showed that the EC₉₀ concentrations of Ko143 were 23 nM (*Abcg2*-mediated mitoxantrone resistance), 5.5 μM (ABCB1-mediated paclitaxel resistance) and >8 μM (ABCC1-mediated etoposide resistance), respectively; these results indicated that Ko143 inhibits ABCG2 stronger than ABCB1 and ABCC1, but is not selective to ABCG2 (Allen et al., 2002). Furthermore, from a series of ABCB1 inhibitors, some ABCG2 inhibitors, such as elacridar (GF120918) (Allen et al., 1999; Kruijtz et al., 2002) and tariquidar (XR9576) (Robey et al., 2004) are frequently used in basic research as well as Ko143.

To date, the molecular bases relating to the chemical inhibition of ABCG2 are not fully understood. The cryo-EM structures of ABCG2 (Taylor et al., 2017) and ABCG2 bound to Ko143 derivatives or tariquidar (Jackson et al., 2018) will be an important to address this issue. Besides, another group of researchers has revealed the structural characteristics of ABCG2 protein critical for its function based on a molecular modeling approach combined with biochemical characterizations of ABCG2 mutants (Khunweeraphong et al., 2017). Previous studies employing the QSAR approaches predicted some structural requirements of compounds for interacting with ABCG2 as an inhibitor (Ishikawa et al., 2012; Mao and Unadkat, 2015); not being true for all ABCG2 inhibitors, the representative features are lipophilicity, planar structure, and amine bonded to one carbon of a heterocyclic ring. Furthermore, a virtual screening strategy employing a ligand-based *in silico* classification model to predict the inhibitory potential of drugs toward ABCG2 presented some favorable outcomes (Montanari et al., 2017). Integration of these findings will contribute to providing a basis for the design of new ABCG2 inhibitors.

Hitherto, many studies focusing on the chemicals–ABCG2 interactions identified a large number of ABCG2 inhibitors with diverse chemical structures (Mao and Unadkat, 2015; Wiese, 2015; Pena-Solorzano et al., 2017; Silbermann et al., 2019). An expanding list of the ABCG2 inhibitors, which include ABCG2



substrates (competitive inhibitors), contains such drugs on the market as TKIs and anti-HIV drugs [some of them are often ABCG2 substrates (Polgar et al., 2008; Mao and Unadkat, 2015)] and such dietary phytochemicals as flavonoids and rotenoids. Nevertheless, to the best of our knowledge, the clinical use of such chemicals for ABCG2 inhibition has not yet been achieved probably due to concerns on safety and/or *in vivo* efficacy of them, which may have been common reasons responsible for the failure of the clinical development of ABCG2 inhibitors. Recent studies importantly showed that two potential anti-cancer compounds under clinical development could competitively inhibit both ABCG2 and ABCB1 (Ji et al., 2018a,b).

Febuxostat, a Highly Potent ABCG2 Inhibitor Applicable in Clinical Situations

Regarding the difficulty in the clinical applications of existing ABCG2 inhibitors, our recent study may open up further avenues, in which febuxostat—an approved agent for hyperuricemia globally used in clinical situations—was serendipitously identified as a strong ABCG2 inhibitor both *in vitro* and *in vivo* (Miyata et al., 2016). Using the vesicle transport assay, we revealed that febuxostat inhibits ABCG2 more strongly than Ko143 and elacridar (Figure 2). This indicates that febuxostat has a superior safety profile and better inhibitory ability against ABCG2 compared to these two compounds. Moreover, the study demonstrated that the IC_{50} of febuxostat against urate transport activity of ABCG2 (0.027 μ M)

was lower than its maximum plasma unbound concentrations reported in humans (0.09 μ M), suggesting that febuxostat might inhibit human ABCG2 at a clinically used dose. Thus, febuxostat can be a promising candidate as a potential ABCG2 inhibitor in humans. The structural characteristics and molecular mechanisms of febuxostat as an ABCG2 inhibitor remain to be elucidated, as well as the effects of febuxostat on the function of other ABC transporters, including ABCB1.

Febuxostat will be used in cancer chemotherapy more frequently because recently this drug has been approved in Europe and Japan for the prophylaxis of TLS. TLS is a potentially life-threatening condition caused by an abrupt release of intracellular metabolites after tumor cell lysis in cancer patients on chemotherapy (Alakel et al., 2017). It is the most common treatment-related emergency in patients with hematologic malignancies and characterized by metabolic abnormalities including hyperuricemia that triggers several mechanisms resulting in acute kidney injury. Appropriate control of serum uric acid is therefore important in the prevention of TLS. Recent studies demonstrated that febuxostat—an oral hypouricemic agent—can successfully prevent TLS in cancer patients (Spina et al., 2015; Tamura et al., 2016). In such situations, since the patients will be treated with febuxostat before and during chemotherapy, there would be drug-drug interactions between febuxostat and ABCG2 substrate anti-cancer agents.

Importantly, ABCG2 is reportedly expressed on the malignant hematopoietic and the lymphoid cells frequently; its expression in several types of human leukemia has been investigated

(Jordanides et al., 2006; Eechoute et al., 2011; Stacy et al., 2013). Considering that initiating cells in CML arises from a multipotent hematopoietic stem cell (HSC), together with high expression of ABCG2 in human HSCs (Zhou et al., 2001; Scharenberg et al., 2002), overexpression of ABCG2 has been considered to confer drug resistance ability to the CML stem cell population (Brendel et al., 2007). However, the clinical relevance of ABCG2 inhibition in CML patients from the view point of reversing MDR remains to be clarified. Since several TKIs used for CML are ABCG2 substrates (Hira and Terada, 2018), the combination use of febuxostat may be a future research topic.

Additionally, ABCG2 expressed in tumor cells is reportedly involved in the efflux of photosensitizers in 5-aminolevulinic acid-based photodynamic therapy (Ishikawa et al., 2015). Hence, febuxostat might enhance the efficacy of this minimally invasive modality for treating solid cancers by accumulating photosensitizers in the target cells. The details are discussed in our previous report (Miyata et al., 2016).

CONCLUSION AND PERSPECTIVE

Here, we summarized some key experimental systems that will continuously contribute to generating novel ABCG2 inhibitors and described an overview of the current update of ABCG2 inhibitors. Among them, febuxostat will be one of the most promising candidates for clinical use. Considering that dysfunctional ABCG2 genotypes, which are summarized in a recent review (Heyes et al., 2018), alter the pharmacokinetic characteristics of ABCG2 substrate drugs such as several TKIs (Hira and Terada, 2018) and rosuvastatin (Keskitalo et al., 2009), ABCG2 inhibitors will also exert similar effects in humans. As a beneficial application of this clinical possibility, we have proposed a novel concept named febuxostat-boosted therapy (Miyata et al., 2016), in which febuxostat is expected to enhance the bioavailability of ABCG2 substrate drugs. For a similar purpose, ritonavir and cobicistat are used as pharmacokinetic boosters inhibiting cytochrome P450 3A4, a major pathway of drug metabolism, to increase the plasma concentrations of certain drugs (Shah et al., 2013). No pharmacokinetic enhancer targeting transporter proteins has been, however, successfully

evaluated in clinical trials. In this context, the potential benefits of the febuxostat-boosted therapy should be validated in the near future. Furthermore, this concept could also be applied to enhance the BBB penetration of ABCG2 substrate drugs for brain cancer chemotherapy. Despite the potential risks of adverse events in the combination therapy, further clinical studies to elucidate whether febuxostat is beneficial in enhancing the efficacy of pharmacotherapy via ABCG2 inhibition are warranted.

AUTHOR CONTRIBUTIONS

YT researched the data for the manuscript, provided substantial contributions to discussion of its content, and wrote the manuscript. TT contributed to the discussion and the writing of the manuscript. HS critiqued the manuscript and provided intellectual inputs. All the authors reviewed and edited the manuscript before submission and have made final approval of the manuscript.

FUNDING

This work and relating our findings were supported by the JSPS KAKENHI Grant Numbers 15H05610 to YT, 16H1808 and 18KK0247 to TT, 22136015 to HS; TT has received research grants from Gout Research Foundation, The Uehara Memorial Foundation, Mochida Memorial Foundation for Medical and Pharmaceutical Research, The Takeda Medical Foundation, and MSD Life Science Foundation, Public Interest Incorporated Foundation.

ACKNOWLEDGMENTS

The authors would like to acknowledge Drs. Hiroshi Miyata, Hirotaka Matsuo, and Kimiyoshi Ichida for their contribution to our studies highlighted in this mini-review article as well as Drs. Toshihisa Ishikawa and Yuichi Sugiyama for their previous mentorships and encouragements.

REFERENCES

- Adachi, Y., Suzuki, H., Schinkel, A. H., and Sugiyama, Y. (2005). Role of breast cancer resistance protein (Bcrp1/Abcg2) in the extrusion of glucuronide and sulfate conjugates from enterocytes to intestinal lumen. *Mol. Pharmacol.* 67, 923–928. doi: 10.1124/mol.104.007393
- Alakel, N., Middeke, J. M., Schetelig, J., and Bornhauser, M. (2017). Prevention and treatment of tumor lysis syndrome, and the efficacy and role of rasburicase. *Onco Targets Ther.* 10, 597–605. doi: 10.2147/OTT.S103864
- Allen, J. D., Brinkhuis, R. F., Wijnholds, J., and Schinkel, A. H. (1999). The mouse Bcrp1/Mxr/Abcp gene: amplification and overexpression in cell lines selected for resistance to topotecan, mitoxantrone, or doxorubicin. *Cancer Res.* 59, 4237–4241.
- Allen, J. D., Van Loevezijn, A., Lakhai, J. M., Van Der Valk, M., Van Tellingen, O., Reid, G., et al. (2002). Potent and specific inhibition of the breast cancer resistance protein multidrug transporter in vitro and in mouse intestine by a novel analogue of fumitremorgin C. *Mol. Cancer Ther.* 1, 417–425.
- Allikmets, R., Schriml, L. M., Hutchinson, A., Romano-Spica, V., and Dean, M. (1998). A human placenta-specific ATP-binding cassette gene (ABCP) on chromosome 4q22 that is involved in multidrug resistance. *Cancer Res.* 58, 5337–5339.
- Ambudkar, S. V., Lelong, I. H., Zhang, J., and Cardarelli, C. (1998). Purification and reconstitution of human P-glycoprotein. *Methods Enzymol.* 292, 492–504. doi: 10.1016/S0076-6879(98)92038-9
- Ando, T., Kusuhara, H., Merino, G., Alvarez, A. I., Schinkel, A. H., and Sugiyama, Y. (2007). Involvement of breast cancer resistance protein (ABCG2) in the biliary excretion mechanism of fluoroquinolones. *Drug Metab. Dispos.* 35, 1873–1879. doi: 10.1124/dmd.107.014969
- Bakhsheshian, J., Wei, B. R., Hall, M. D., Simpson, R. M., and Gottesman, M. M. (2016). In vivo bioluminescent imaging of ATP-binding cassette transporter-mediated efflux at the blood-brain barrier. *Methods Mol. Biol.* 1461, 227–239. doi: 10.1007/978-1-4939-3813-1_19
- Baykov, A. A., Evtushenko, O. A., and Avaeva, S. M. (1988). A malachite green procedure for orthophosphate determination and its use in alkaline

- phosphatase-based enzyme immunoassay. *Anal. Biochem.* 171, 266–270. doi: 10.1016/0003-2697(88)90484-8
- Brendel, C., Scharenberg, C., Dohse, M., Robey, R. W., Bates, S. E., Shukla, S., et al. (2007). Imatinib mesylate and nilotinib (AMN107) exhibit high-affinity interaction with ABCG2 on primitive hematopoietic stem cells. *Leukemia* 21, 1267–1275. doi: 10.1038/sj.leu.2404638
- Doyle, L. A., Yang, W., Abruzzo, L. V., Krogmann, T., Gao, Y., Rishi, A. K., et al. (1998). A multidrug resistance transporter from human MCF-7 breast cancer cells. *Proc. Natl. Acad. Sci. U.S.A.* 95, 15665–15670. doi: 10.1073/pnas.95.26.15665
- Eechoute, K., Sparreboom, A., Burger, H., Franke, R. M., Schiavon, G., Verweij, J., et al. (2011). Drug transporters and imatinib treatment: implications for clinical practice. *Clin. Cancer Res.* 17, 406–415. doi: 10.1158/1078-0432.CCR-10-2250
- Geertsma, E. R., Nik Mahmood, N. A., Schuurman-Wolters, G. K., and Poolman, B. (2008). Membrane reconstitution of ABC transporters and assays of translocator function. *Nat. Protoc.* 3, 256–266. doi: 10.1038/nprot.2007.519
- Giacomini, K. M., Huang, S. M., Tweedie, D. J., Benet, L. Z., Brouwer, K. L., Chu, X., et al. (2010). Membrane transporters in drug development. *Nat. Rev. Drug Discov.* 9, 215–236. doi: 10.1038/nrd3028
- Guo, C., Liu, F., Qi, J., Ma, J., Lin, S., Zhang, C., et al. (2018). A novel synthetic dihydroindeno[1,2-b] indole derivative (LS-2-3j) reverses ABCB1- and ABCG2-mediated multidrug resistance in cancer cells. *Molecules* 23:E3264. doi: 10.3390/molecules23123264
- Hegedus, C., Szakacs, G., Homolya, L., Orban, T. I., Telbisz, A., Jani, M., et al. (2009). Ins and outs of the ABCG2 multidrug transporter: an update on in vitro functional assays. *Adv. Drug Deliv. Rev.* 61, 47–56. doi: 10.1016/j.addr.2008.09.007
- Heyes, N., Kapoor, P., and Kerr, I. D. (2018). Polymorphisms of the multidrug Pump ABCG2: a systematic review of their effect on protein expression, function, and drug pharmacokinetics. *Drug Metab. Dispos.* 46, 1886–1899. doi: 10.1124/dmd.118.083030
- Higashino, T., Takada, T., Nakaoka, H., Toyoda, Y., Stiburkova, B., Miyata, H., et al. (2017). Multiple common and rare variants of ABCG2 cause gout. *RMD Open* 3:e000464. doi: 10.1136/rmdopen-2017-000464
- Hira, D., and Terada, T. (2018). BCRP/ABCG2 and high-alert medications: biochemical, pharmacokinetic, pharmacogenetic, and clinical implications. *Biochem. Pharmacol.* 147, 201–210. doi: 10.1016/j.bcp.2017.10.004
- Hirano, M., Maeda, K., Matsushima, S., Nozaki, Y., Kusahara, H., and Sugiyama, Y. (2005). Involvement of BCRP (ABCG2) in the biliary excretion of pitavastatin. *Mol. Pharmacol.* 68, 800–807. doi: 10.1124/mol.105.014019
- Ichida, K., Matsuo, H., Takada, T., Nakayama, A., Murakami, K., Shimizu, T., et al. (2012). Decreased extra-renal urate excretion is a common cause of hyperuricemia. *Nat. Commun.* 3:764. doi: 10.1038/ncomms1756
- Ishikawa, T. (1989). ATP/Mg²⁺-dependent cardiac transport system for glutathione S-conjugates. A study using rat heart sarcolemma vesicles. *J. Biol. Chem.* 264, 17343–17348.
- Ishikawa, T., Kajimoto, Y., Inoue, Y., Ikegami, Y., and Kuroiwa, T. (2015). Critical role of ABCG2 in ALA-photodynamic diagnosis and therapy of human brain tumor. *Adv. Cancer Res.* 125, 197–216. doi: 10.1016/bs.acr.2014.11.008
- Ishikawa, T., Saito, H., Hirano, H., Inoue, Y., and Ikegami, Y. (2012). Human ABC transporter ABCG2 in cancer chemotherapy: drug molecular design to circumvent multidrug resistance. *Methods Mol. Biol.* 910, 267–278. doi: 10.1007/978-1-61779-965-5_11
- Ito, N., Ito, K., Ikebuchi, Y., Toyoda, Y., Takada, T., Hisaka, A., et al. (2015). Prediction of drug transfer into milk considering breast cancer resistance protein (BCRP)-mediated transport. *Pharm. Res.* 32, 2527–2537. doi: 10.1007/s11095-015-1641-2
- Jackson, S. M., Manolaridis, I., Kowal, J., Zechner, M., Taylor, N. M. I., Bause, M., et al. (2018). Structural basis of small-molecule inhibition of human multidrug transporter ABCG2. *Nat. Struct. Mol. Biol.* 25, 333–340. doi: 10.1038/s41594-018-0049-1
- Ji, N., Yang, Y., Cai, C. Y., Lei, Z. N., Wang, J. Q., Gupta, P., et al. (2018a). VS-4718 antagonizes multidrug resistance in ABCB1- and ABCG2-overexpressing cancer cells by inhibiting the efflux function of ABC transporters. *Front. Pharmacol.* 9:1236. doi: 10.3389/fphar.2018.01236
- Ji, N., Yang, Y., Lei, Z. N., Cai, C. Y., Wang, J. Q., Gupta, P., et al. (2018b). Ulixertinib (BVD-523) antagonizes ABCB1- and ABCG2-mediated chemotherapeutic drug resistance. *Biochem. Pharmacol.* 158, 274–285. doi: 10.1016/j.bcp.2018.10.028
- Jonker, J. W., Buitelaar, M., Wagenaar, E., Van Der Valk, M. A., Scheffer, G. L., Scheper, R. J., et al. (2002). The breast cancer resistance protein protects against a major chlorophyll-derived dietary phototoxin and protoporphyria. *Proc. Natl. Acad. Sci. U.S.A.* 99, 15649–15654. doi: 10.1073/pnas.202607599
- Jonker, J. W., Merino, G., Musters, S., Van Herwaarden, A. E., Bolscher, E., Wagenaar, E., et al. (2005). The breast cancer resistance protein BCRP (ABCG2) concentrates drugs and carcinogenic xenotoxins into milk. *Nat. Med.* 11, 127–129. doi: 10.1038/nm1186
- Jordanides, N. E., Jorgensen, H. G., Holyoake, T. L., and Mountford, J. C. (2006). Functional ABCG2 is overexpressed on primary CML CD34+ cells and is inhibited by imatinib mesylate. *Blood* 108, 1370–1373. doi: 10.1182/blood-2006-02-003145
- Keskitalo, J. E., Zolk, O., Fromm, M. F., Kurkinen, K. J., Neuvonen, P. J., and Niemi, M. (2009). ABCG2 polymorphism markedly affects the pharmacokinetics of atorvastatin and rosuvastatin. *Clin. Pharmacol. Ther.* 86, 197–203. doi: 10.1038/clpt.2009.79
- Khunweeraphong, N., Stockner, T., and Kuchler, K. (2017). The structure of the human ABC transporter ABCG2 reveals a novel mechanism for drug extrusion. *Sci. Rep.* 7:13767. doi: 10.1038/s41598-017-11794-w
- Kruitjzer, C. M., Beijnen, J. H., Rosing, H., ten Bokkel Huinink, W. W., Schot, M., Jewell, R. C., et al. (2002). Increased oral bioavailability of topotecan in combination with the breast cancer resistance protein and P-glycoprotein inhibitor GF120918. *J. Clin. Oncol.* 20, 2943–2950. doi: 10.1200/JCO.2002.12.116
- Kusahara, H., Furuie, H., Inano, A., Sunagawa, A., Yamada, S., Wu, C., et al. (2012). Pharmacokinetic interaction study of sulphasalazine in healthy subjects and the impact of curcumin as an in vivo inhibitor of BCRP. *Br. J. Pharmacol.* 166, 1793–1803. doi: 10.1111/j.1476-5381.2012.01887.x
- Mao, Q., and Unadkat, J. D. (2015). Role of the breast cancer resistance protein (BCRP/ABCG2) in drug transport—an update. *AAPS J.* 17, 65–82. doi: 10.1208/s12248-014-9668-6
- Marighetti, F., Steggemann, K., Hanl, M., and Wiese, M. (2013). Synthesis and quantitative structure-activity relationships of selective BCRP inhibitors. *ChemMedChem* 8, 125–135. doi: 10.1002/cmdc.201200377
- Matsuo, H., Takada, T., Ichida, K., Nakamura, T., Nakayama, A., Ikebuchi, Y., et al. (2009). Common defects of ABCG2, a high-capacity urate exporter, cause gout: a function-based genetic analysis in a Japanese population. *Sci. Transl. Med.* 1:5ra11. doi: 10.1126/scitranslmed.3000237
- Matsushima, S., Maeda, K., Kondo, C., Hirano, M., Sasaki, M., Suzuki, H., et al. (2005). Identification of the hepatic efflux transporters of organic anions using double-transfected Madin-Darby canine kidney II cells expressing human organic anion-transporting polypeptide 1B1 (OATP1B1)/multidrug resistance-associated protein 2, OATP1B1/multidrug resistance 1, and OATP1B1/breast cancer resistance protein. *J. Pharmacol. Exp. Ther.* 314, 1059–1067. doi: 10.1124/jpet.105.085589
- Miyake, K., Mickley, L., Litman, T., Zhan, Z., Robey, R., Cristensen, B., et al. (1999). Molecular cloning of cDNAs which are highly overexpressed in mitoxantrone-resistant cells: demonstration of homology to ABC transport genes. *Cancer Res.* 59, 8–13.
- Miyata, H., Takada, T., Toyoda, Y., Matsuo, H., Ichida, K., and Suzuki, H. (2016). Identification of febuxostat as a new strong ABCG2 inhibitor: potential applications and risks in clinical situations. *Front. Pharmacol.* 7:518. doi: 10.3389/fphar.2016.00518
- Mizuno, N., Suzuki, M., Kusahara, H., Suzuki, H., Takeuchi, K., Niwa, T., et al. (2004). Impaired renal excretion of 6-hydroxy-5,7-dimethyl-2-methylamino-4-(3-pyridylmethyl) benzothiazole (E3040) sulfate in breast cancer resistance protein (BCRP1/ABCG2) knockout mice. *Drug Metab. Dispos.* 32, 898–901.
- Mizuno, N., Takahashi, T., Kusahara, H., Schuetz, J. D., Niwa, T., and Sugiyama, Y. (2007). Evaluation of the role of breast cancer resistance protein (BCRP/ABCG2) and multidrug resistance-associated protein 4 (MRP4/ABCC4) in the urinary excretion of sulfate and glucuronide metabolites of edaravone (MCI-186; 3-methyl-1-phenyl-2-pyrazolin-5-one). *Drug Metab. Dispos.* 35, 2045–2052. doi: 10.1124/dmd.107.016352

- Montanari, F., Cseke, A., Wlcek, K., and Ecker, G. F. (2017). Virtual screening of DrugBank reveals two drugs as new BCRP inhibitors. *SLAS Discov.* 22, 86–93. doi: 10.1177/1087057116657513
- Murakami, M., Ohnuma, S., Fukuda, M., Chufan, E. E., Kudoh, K., Kanehara, K., et al. (2017). Synthetic analogs of curcumin modulate the function of multidrug resistance-linked ATP-binding cassette transporter ABCG2. *Drug Metab. Dispos.* 45, 1166–1177. doi: 10.1124/dmd.117.076000
- Nakanishi, T., Doyle, L. A., Hassel, B., Wei, Y., Bauer, K. S., Wu, S., et al. (2003). Functional characterization of human breast cancer resistance protein (BCRP, ABCG2) expressed in the oocytes of *Xenopus laevis*. *Mol. Pharmacol.* 64, 1452–1462. doi: 10.1124/mol.64.6.1452
- Nicolle, E., Boumendjel, A., Macalou, S., Genoux, E., Ahmed-Belkacem, A., Carrupt, P. A., et al. (2009). QSAR analysis and molecular modeling of ABCG2-specific inhibitors. *Adv. Drug Deliv. Rev.* 61, 34–46. doi: 10.1016/j.addr.2008.10.004
- Okamoto, T., Kawaguchi, K., Watanabe, S., Agustina, R., Ikejima, T., Ikeda, K., et al. (2018). Characterization of human ATP-binding cassette protein subfamily D reconstituted into proteoliposomes. *Biochem. Biophys. Res. Commun.* 496, 1122–1127. doi: 10.1016/j.bbrc.2018.01.153
- Pena-Solorzano, D., Stark, S. A., Konig, B., Sierra, C. A., and Ochoa-Puentes, C. (2017). ABCG2/BCRP: specific and nonspecific modulators. *Med. Res. Rev.* 37, 987–1050. doi: 10.1002/med.21428
- Polgar, O., Robey, R. W., and Bates, S. E. (2008). ABCG2: structure, function and role in drug response. *Expert Opin. Drug Metab. Toxicol.* 4, 1–15. doi: 10.1517/17425255.4.1.1
- Rabindran, S. K., He, H., Singh, M., Brown, E., Collins, K. I., Annable, T., et al. (1998). Reversal of a novel multidrug resistance mechanism in human colon carcinoma cells by fumitremorgin C. *Cancer Res.* 58, 5850–5858.
- Rabindran, S. K., Ross, D. D., Doyle, L. A., Yang, W., and Greenberger, L. M. (2000). Fumitremorgin C reverses multidrug resistance in cells transfected with the breast cancer resistance protein. *Cancer Res.* 60, 47–50.
- Robey, R. W., Pluchino, K. M., Hall, M. D., Fojo, A. T., Bates, S. E., and Gottesman, M. M. (2018). Revisiting the role of ABC transporters in multidrug-resistant cancer. *Nat. Rev. Cancer* 18, 452–464. doi: 10.1038/s41568-018-0005-8
- Robey, R. W., Steadman, K., Polgar, O., Morisaki, K., Blayney, M., Mistry, P., et al. (2004). Pheophorbide a is a specific probe for ABCG2 function and inhibition. *Cancer Res.* 64, 1242–1246. doi: 10.1158/0008-5472.CAN-03-3298
- Robey, R. W., To, K. K., Polgar, O., Dohse, M., Fetsch, P., Dean, M., et al. (2009). ABCG2: a perspective. *Adv. Drug Deliv. Rev.* 61, 3–13. doi: 10.1016/j.addr.2008.11.003
- Saito, H., Hirano, H., Nakagawa, H., Fukami, T., Oosumi, K., Murakami, K., et al. (2006). A new strategy of high-speed screening and quantitative structure-activity relationship analysis to evaluate human ATP-binding cassette transporter ABCG2-drug interactions. *J. Pharmacol. Exp. Ther.* 317, 1114–1124. doi: 10.1124/jpet.105.099036
- Scharenberg, C. W., Harkey, M. A., and Torok-Storb, B. (2002). The ABCG2 transporter is an efficient Hoechst 33342 efflux pump and is preferentially expressed by immature human hematopoietic progenitors. *Blood* 99, 507–512. doi: 10.1182/blood.V99.2.507
- Shah, B. M., Schafer, J. J., Priano, J., and Squires, K. E. (2013). Cobicistat: a new boost for the treatment of human immunodeficiency virus infection. *Pharmacotherapy* 33, 1107–1116. doi: 10.1002/phar.1237
- Shukla, S., Kouanda, A., Silvertown, L., Talele, T. T., and Ambudkar, S. V. (2014). Pharmacophore modeling of nilotinib as an inhibitor of ATP-binding cassette drug transporters and BCR-ABL kinase using a three-dimensional quantitative structure-activity relationship approach. *Mol. Pharm.* 11, 2313–2322. doi: 10.1021/mp400762h
- Shukla, S., Robey, R. W., Bates, S. E., and Ambudkar, S. V. (2006). The calcium channel blockers, 1,4-dihydropyridines, are substrates of the multidrug resistance-linked ABC drug transporter, ABCG2. *Biochemistry* 45, 8940–8951. doi: 10.1021/bi060552f
- Silbermann, K., Shah, C. P., Sahu, N. U., Juveale, K., Stefan, S. M., Kharkar, P. S., et al. (2019). Novel chalcone and flavone derivatives as selective and dual inhibitors of the transport proteins ABCB1 and ABCG2. *Eur. J. Med. Chem.* 164, 193–213. doi: 10.1016/j.ejmech.2018.12.019
- Spina, M., Nagy, Z., Ribera, J. M., Federico, M., Aurer, I., Jordan, K., et al. (2015). FLORENCE: a randomized, double-blind, phase III pivotal study of febusostat versus allopurinol for the prevention of tumor lysis syndrome (TLS) in patients with hematologic malignancies at intermediate to high TLS risk. *Ann. Oncol.* 26, 2155–2161. doi: 10.1093/annonc/mdv317
- Stacy, A. E., Jansson, P. J., and Richardson, D. R. (2013). Molecular pharmacology of ABCG2 and its role in chemoresistance. *Mol. Pharmacol.* 84, 655–669. doi: 10.1124/mol.113.088609
- Stiburkova, B., Miyata, H., Zavada, J., Tomcik, M., Pavelka, K., Storkanova, G., et al. (2016). Novel dysfunctional variant in ABCG2 as a cause of severe tophaceous gout: biochemical, molecular genetics and functional analysis. *Rheumatology* 55, 191–194. doi: 10.1093/rheumatology/kev350
- Suzuki, M., Suzuki, H., Sugimoto, Y., and Sugiyama, Y. (2003). ABCG2 transports sulfated conjugates of steroids and xenobiotics. *J. Biol. Chem.* 278, 22644–22649. doi: 10.1074/jbc.M212399200
- Takada, T., Yamamoto, T., Matsuo, H., Tan, J. K., Ooyama, K., Sakiyama, M., et al. (2018). Identification of ABCG2 as an exporter of uremic toxin indoxyl sulfate in mice and as a crucial factor influencing CKD progression. *Sci. Rep.* 8:11147. doi: 10.1038/s41598-018-29208-w
- Tamura, A., Watanabe, M., Saito, H., Nakagawa, H., Kamachi, T., Okura, I., et al. (2006). Functional validation of the genetic polymorphisms of human ATP-binding cassette (ABC) transporter ABCG2: identification of alleles that are defective in porphyrin transport. *Mol. Pharmacol.* 70, 287–296.
- Tamura, K., Kawai, Y., Kiguchi, T., Okamoto, M., Kaneko, M., Maemondo, M., et al. (2016). Efficacy and safety of febusostat for prevention of tumor lysis syndrome in patients with malignant tumors receiving chemotherapy: a phase III, randomized, multi-center trial comparing febusostat and allopurinol. *Int. J. Clin. Oncol.* 21, 996–1003. doi: 10.1007/s10147-016-0971-3
- Taylor, N. M. I., Manolaridis, I., Jackson, S. M., Kowal, J., Stahlberg, H., and Locher, K. P. (2017). Structure of the human multidrug transporter ABCG2. *Nature* 546, 504–509. doi: 10.1038/nature22345
- Tiwari, A. K., Sodani, K., Dai, C. L., Abuznait, A. H., Singh, S., Xiao, Z. J., et al. (2013). Nilotinib potentiates anticancer drug sensitivity in murine ABCB1-, ABCG2-, and ABCC10-multidrug resistance xenograft models. *Cancer Lett.* 328, 307–317. doi: 10.1016/j.canlet.2012.10.001
- Toyoda, Y., Takada, T., and Suzuki, H. (2016). Halogenated hydrocarbon solvent-related cholangiocarcinoma risk: biliary excretion of glutathione conjugates of 1,2-dichloropropane evidenced by untargeted metabolomics analysis. *Sci. Rep.* 6:24586. doi: 10.1038/srep24586
- van de Wetering, K., and Saptho, S. (2012). ABCG2 functions as a general phytoestrogen sulfate transporter in vivo. *FASEB J.* 26, 4014–4024. doi: 10.1096/fj.12-210039
- Vlaming, M. L., Lagas, J. S., and Schinkel, A. H. (2009). Physiological and pharmacological roles of ABCG2 (BCRP): recent findings in Abcg2 knockout mice. *Adv. Drug Deliv. Rev.* 61, 14–25. doi: 10.1016/j.addr.2008.08.007
- Wassermann, L., Halwachs, S., Baumann, D., Schaefer, I., Seibel, P., and Honscha, W. (2013). Assessment of ABCG2-mediated transport of xenobiotics across the blood-milk barrier of dairy animals using a new MDCKII in vitro model. *Arch. Toxicol.* 87, 1671–1682. doi: 10.1007/s00204-013-1066-9
- Wiese, M. (2015). BCRP/ABCG2 inhibitors: a patent review (2009-present). *Expert Opin. Ther. Pat.* 25, 1229–1237. doi: 10.1517/13543776.2015.1076796
- Woodward, O. M., Kottgen, A., Coresh, J., Boerwinkle, E., Guggino, W. B., and Kottgen, M. (2009). Identification of a urate transporter, ABCG2, with a common functional polymorphism causing gout. *Proc. Natl. Acad. Sci. U.S.A.* 106, 10338–10342. doi: 10.1073/pnas.0901249106
- Wu, C. P., Hsiao, S. H., Murakami, M., Lu, Y. J., Li, Y. Q., Huang, Y. H., et al. (2017). Alpha-mangostin reverses multidrug resistance by attenuating the function of the multidrug resistance-linked ABCG2 transporter. *Mol. Pharm.* 14, 2805–2814. doi: 10.1021/acs.molpharmaceut.7b00334
- Zhang, G. N., Zhang, Y. K., Wang, Y. J., Gupta, P., Ashby, C. R. Jr., Alqahtani, S., et al. (2018). Epidermal growth factor receptor (EGFR) inhibitor PD153035 reverses ABCG2-mediated multidrug resistance in non-small cell lung cancer: in vitro and in vivo. *Cancer Lett.* 424, 19–29. doi: 10.1016/j.canlet.2018.02.040
- Zhang, W., Chen, Z., Chen, L., Wang, F., Li, F., Wang, X., et al. (2017). ABCG2-overexpressing H460/MX20 cell xenografts in athymic nude mice maintained original biochemical and cytological characteristics. *Sci. Rep.* 7:40064. doi: 10.1038/srep40064
- Zhang, Y., Bressler, J. P., Neal, J., Lal, B., Bhang, H. E., Lartera, J., et al. (2007). ABCG2/BCRP expression modulates D-Luciferin based bioluminescence imaging. *Cancer Res.* 67, 9389–9397. doi: 10.1158/0008-5472.CAN-07-0944

- Zhang, Y. K., Zhang, G. N., Wang, Y. J., Patel, B. A., Talele, T. T., Yang, D. H., et al. (2016). Bafetinib (INNO-406) reverses multidrug resistance by inhibiting the efflux function of ABCB1 and ABCG2 transporters. *Sci. Rep.* 6:25694. doi: 10.1038/srep25694
- Zhou, S., Schuetz, J. D., Bunting, K. D., Colapietro, A. M., Sampath, J., Morris, J. J., et al. (2001). The ABC transporter Bcrp1/ABCG2 is expressed in a wide variety of stem cells and is a molecular determinant of the side-population phenotype. *Nat. Med.* 7, 1028–1034. doi: 10.1038/nm0901-1028
- Zhou, S., Zong, Y., Ney, P. A., Nair, G., Stewart, C. F., and Sorrentino, B. P. (2005). Increased expression of the Abcg2 transporter during erythroid maturation plays a role in decreasing cellular protoporphyrin IX levels. *Blood* 105, 2571–2576. doi: 10.1182/blood-2004-04-1566

Conflict of Interest Statement: TT and HS have a patent pending.

The remaining author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Toyoda, Takada and Suzuki. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.