

# Regulation of erythrocyte function: multiple evolutionary solutions for respiratory gas transport and its regulation in fish

Journal:	Acta Physiologica
Manuscript ID	APH-2019-04-0263.R1
Manuscript Type:	Review Article
Date Submitted by the Author:	03-May-2019
Complete List of Authors:	Nikinmaa, Mikko; University of Turku, Department of Biology Berenbrink, Michael; University of Liverpool, School of Biological Sciences Brauner, Colin; University of British Columbia, Department of Zoology
Key Words:	Oxygen equilibrium curve (OEC), Bohr effect, Root effect, anion exchange, adrenergically stimulated sodium/proton exchange, carbonic anhydrase, red blood cell



Regulation of erythrocyte function: multiple evolutionary solutions for respiratory gas transport and its regulation in fish Mikko Nikinmaa<sup>1</sup>, Michael Berenbrink<sup>2</sup>, Colin J. Brauner<sup>3</sup> 1. Department of Biology, University of Turku, FI-20014 Turku, Finland 2. Institute of Integrative Biology, Department of Evolution, Ecology and Behaviour, Biosciences Building, Crown Street, University of Liverpool, Liverpool, L69 7ZB, U.K. 3. Department of Zoology, 6270 University Blvd., University of British Columbia, Vancouver, Canada, V6T 1Z4. Short title: Gas transport in fish erythrocytes Corresponding author: Mikko Nikinmaa, Department of Biology, University of Turku, FI-20014 Turku, Finland; tel: +358408361073; e-mail: miknik@utu.fi 

# Abstract

Gas transport concepts in vertebrates have naturally been formulated based on human blood. However, the first vertebrates were aquatic, and fish and tetrapods diverged hundreds of millions years ago. Water-breathing vertebrates live in an environment with low and variable O<sub>2</sub> levels, making environmental  $O_2$  an important evolutionary selection pressure in fishes, and various features of their gas transport differ from humans. Erythrocyte function in fish is of current interest, because current environmental changes affect gas transport, and because especially zebrafish is used as a model in biomedical studies, making it important to understand the differences in gas transport between fish and mammals to be able to carry out meaningful studies. Of the close to thirty thousand fish species, teleosts are the most species-numerous group. However, two additional radiations are discussed: agnathans and elasmobranchs. The gas transport by elasmobranchs may be closest to the ancestors of tetrapods. The major difference in their haemoglobin (Hb) function to humans is their high urea tolerance. Agnathans differ from other vertebrates by having Hbs, where cooperativity is achieved by monomer-oligomer equilibria. Their erythrocytes also lack the anion exchange pathway with profound effects on  $CO_2$  transport. Teleosts are characterized by highly pH sensitive Hbs, which can fail to become fully O<sub>2</sub>-saturated at low pH. An adrenergically stimulated Na<sup>+</sup>/H<sup>+</sup> exchanger has evolved in their erythrocyte membrane, and plasma-accessible carbonic anhydrase can be differentially distributed among their tissues. Together, and differing from other vertebrates, these features can maximize  $O_2$  unloading in muscle while ensuring  $O_2$  loading in gills. key words: Oxygen equilibrium curve (OEC), Bohr effect, Root effect, anion exchange, adrenergically

stimulated Na<sup>+</sup>/H<sup>+</sup> exchange, carbonic anhydrase, erythrocyte

Page 3 of 38

## Acta Physiologica

# 

# 1. Introduction: All vertebrates originated in an aquatic environment

Vertebrates originated in an aquatic environment. Consequently, they evolved in an environment that is characterized by low  $O_2$  solubility, where  $O_2$  content is only ca. 1/20 to 1/30 of that in an equal volume of surrounding air (depending on temperature) and varies markedly daily and seasonally <sup>1</sup>. The low O<sub>2</sub> availability of water is thought to have been an important selective force in the evolution of respiratory systems in fish. As a result, the O<sub>2</sub> transport system of fish is the most versatile among vertebrates <sup>2-4</sup>, with the most extreme adaptation in fishes being the ability to breathe air which occurs in approximately 500 species <sup>5</sup>. Three major alternative solutions in the gas transport by erythrocytes have evolved in fish: that employed by agnathans <sup>6</sup>, that used by elasmobranchs <sup>7,8</sup>, and that found in teleosts <sup>9</sup>. The pattern employed by elasmobranchs may be the most representative of fish ancestors of the tetrapods and hence mammals <sup>10</sup>. Apart from mammals, all vertebrates have erythrocytes that are nucleated and contain other cell organelles, which may partly explain why many features of the regulation of erythrocyte function, important for the different environmental adaptations in fish, are not utilized in air-breathing mammals with their organelle-free erythrocytes. In mammals these features have either been lost, become vestigial or alternatively, some of these features may have uniquely evolved in specific groups of fishes and thus may never have been present in any mammalian ancestor during the evolution of their O<sub>2</sub> transport system <sup>10-12</sup>. This is an important point to consider when using teleost fishes such as zebrafish (Danio *rerio*) to study  $O_2$ -dependent phenomena with a mammalian biomedical application: the zebrafish is a very hypoxia-tolerant tropical cyprinid <sup>13-15</sup>. Although direct link to gas transport cannot be yet made, it is of interest that different from mammals all zebrafish cells exhibit circadian rhythm <sup>16</sup>. In addition to light, oxygen has marked circadian fluctuations <sup>17</sup>, and the major oxygen-regulated transcription factor, HIF, shows (circadian) rhythmic behaviour <sup>18-20</sup>. Many of the discoveries on the regulation of erythrocyte function in fish have been made long after the formulation of the general blood gas transport principles in vertebrates, which were mainly centred on mammals (for a thorough old account, see, e.g. Bishop and Surgenor, 1964)<sup>21</sup>. Consequently, more contemporary

findings on erythrocyte function have been interpreted in light of an earlier gas transport framework that may not be appropriate. In this commentary, we consider erythrocyte function in fish in the context of their unique gas transport characteristics.

A couple of examples indicate the marked differences between mammals and fish (and also other vertebrates). First, as a result of retaining mitochondria, fish erythrocytes are capable of aerobic metabolism, and consume significant amounts of O<sub>2</sub> – producing more than 90 % of their ATP aerobically <sup>22-25</sup>. Second, connected with the presence of a nucleus, fish erythrocytes are capable of active gene expression throughout their life span, although gene transcription and translation decrease with age <sup>26,27</sup>. In view of this, after giving a very short background on the basic vertebrate models of gas transport, we consider the different fish solutions in detail with emphasis on their unique features.

## 2. The basic model of vertebrate oxygen and carbon dioxide transport

Oxygen-carrying pigments are needed to increase the amount of  $O_2$  carried by blood, because without respiratory pigment the amount of  $O_2$  carried would be limited by its low solubility in aqueous media such as plasma. Thus, in the absence of respiratory pigments, the  $O_2$  consumption rate of multicellular animals cannot be high. This is exemplified by icefish, which have secondarily lost Hb entirely from their circulatory system. They are able to live in the absence of Hb largely due to their low metabolic rate in their low-temperature habitat <sup>28,29</sup>.

The simplest solution to increase the  $O_2$ -carrying capacity of a solution is to have monomeric globins or other monomeric respiratory pigments. However, to be the most effective,  $O_2$  binding at the gas exchanger should occur at as low an  $O_2$  tension as possible and  $O_2$  release at the tissues at an  $O_2$ tension as high as possible. This is difficult to accomplish for monomeric globins with hyperbolic oxygen equilibrium curves (OEC); by having tetrameric globins, where subunit interactions result in a sigmoidally shaped OEC, both the  $O_2$  loading and unloading can be more easily fine-tuned to respiratory requirements.

#### Acta Physiologica

A standard OEC is presented in Figure 1. As shown over a century ago, increasing the partial pressure of carbon dioxide ( $PCO_2$ ) decreases Hb-O<sub>2</sub> affinity <sup>30</sup>. When the pH concept was later formalized, the pH-dependent change of O<sub>2</sub> affinity was named the Bohr effect (as an increase in  $PCO_2$  causes a decrease in pH). A single numerical value,  $P_{50}$  is often used to describe Hb-O<sub>2</sub> affinity. The value gives the partial pressure of O<sub>2</sub> ( $PO_2$ ) where Hb is half-saturated, which generally increases as pH decreases, indicating a reduction in Hb-O<sub>2</sub> affinity. A second major discovery in studies of Hb function is now more than 50 years old; in the late 1960's organic phosphates within the erythrocyte were shown to regulate Hb-O<sub>2</sub> affinity <sup>31,32</sup>. Since then, Hb function has been reviewed several times in journals <sup>4,33-35</sup>, and books <sup>36-38</sup> to which the reader is referred. The present contribution briefly reviews the basics to provide the background for a discussion on the unique solutions in fish presented here.

In the simplest models, tetrameric Hb molecules exist in two major conformations: the high affinity R(relaxed)-state and the low affinity T(tense)-state. The existence of high- and low-affinity conformations is the prerequisite of cooperative  $O_2$  binding (sigmoidal OEC). Perutz (1990) <sup>6</sup> has discussed cooperativity in detail. Two main factors influence the T-R-state equilibrium. First, the binding of the first  $O_2$  molecules to deoxygenated T-state Hb facilitates the binding of consecutive  $O_2$  molecules (i.e. tends to shift the Hb conformation from the T-state towards the R-state). Second, the binding of protons (H<sup>+</sup>s), organic phosphates and other anions (the most important of which appear to be Cl<sup>-39</sup> and HCO<sub>3</sub><sup>-40</sup>) stabilize the T-state (these so-called allosteric effectors bind to sites other than the  $O_2$  binding site). The overall OEC of Hb molecules is determined by the probabilities of T- or R-state occurrence; the equilibrium between T- and R-states is virtually instantaneous.

Gas transport in fish is optimized in part through pH effects on Hb function. In this context it is important to emphasize the linked function concept <sup>41</sup>: the effects of H<sup>+</sup>s on Hb-O<sub>2</sub> affinity (Bohr effect) and the effects of O<sub>2</sub> on the binding of H<sup>+</sup>s (Haldane effect) describe the same phenomenon. Thus, a large numerical value of a Bohr coefficient ( $\Delta \log P_{50}/\Delta pH$ ) is necessarily associated with a

large numerical value of Haldane coefficient, i.e. difference in the acid dissociation constants ( $pK_{a}$ values) of oxy- and deoxyHb with the overall  $pK_a$  of deoxyHb being much higher than that of oxyHb. Often, the effect of pH on Hb-O<sub>2</sub> affinity (usually given as  $\Delta \log P_{50}/\Delta pH$ ) is given in terms of plasma pH values. However, the ratio of  $\Delta p H_e / \Delta p H_i$  is less than unity, which means that the Bohr coefficient obtained using plasma pH is smaller than if the value is estimated either based on erythrocyte pH or for Hb solutions i.e. the true value experienced by Hb molecule. The error caused by using plasma pH instead of erythrocyte pH remains constant in mammals regardless of the experimental conditions, since erythrocyte pH cannot be regulated. The same is true for undisturbed fish <sup>42</sup>. However, in many groups of fishes, erythrocyte pH can vary independently from plasma pH (see below), which makes the values for the Bohr coefficient calculated on the basis of plasma pH <sup>43</sup> less meaningful under various stresses and thus such values should be interpreted critically. It is further of note that the effect of pH on the Hb-O<sub>2</sub> affinity may depend on the pH range studied. In the physiologically relevant pH range, a decrease in pH causes a decrease of Hb-O<sub>2</sub> affinity (alkaline Bohr effect). However, in acid conditions (e.g. below pH 6 for human <sup>37</sup>), a decrease in pH may increase Hb- $O_2$  affinity (acid Bohr effect). Since the alkaline Bohr effect is the physiologically relevant one, we simply refer to that from this point forward as the Bohr effect.

In typical tetrameric Hbs, an increase in temperature decreases the overall  $O_2$  affinity, because Hb- $O_2$  binding is exothermic. However, the heat of oxygenation of each successive  $O_2$  molecule that binds to the tetramer varies markedly <sup>44</sup>. In addition to the exothermic  $O_2$  binding by haem, the overall temperature effect on Hb- $O_2$  affinity is affected by the endothermic reactions between Hb and allosteric effectors such as H<sup>+</sup>s and organic phosphates <sup>45</sup>. Consequently, the higher the: i) concentration of allosteric effectors, ii) number of allosteric effector binding sites per Hb tetramer and iii) binding affinities of these allosteric effectors, the smaller the temperature-induced decrease in Hb- $O_2$  affinity. At an extreme, the final result can be a reversed, endothermic overall Hb oxygenation reaction, whereby Hb- $O_2$  affinity increases with increasing temperature <sup>46,47</sup>.

## Acta Physiologica

With regard to the transport of CO<sub>2</sub>, the end product of aerobic respiration, two properties of most vertebrate erythrocytes are considered to be of primary importance. First, erythrocytes possess high carbonic anhydrase (CA) activity <sup>48,49</sup>. Carbonic anhydrase catalyzes the hydration/dehydration reactions between CO<sub>2</sub> and carbonic acid, which are the rate-limiting steps in establishing the equilibrium between CO<sub>2</sub> and HCO<sub>3</sub><sup>-</sup>. Second, the transport of HCO<sub>3</sub><sup>-</sup> across the erythrocyte membrane is accelerated by the anion exchange protein, allowing rapid Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchange <sup>50,51</sup>. The rapid Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchange enables the function of the Jacobs-Stewart cycle, utilized in CO<sub>2</sub> transport <sup>52</sup>. In most vertebrates, more than 50% of CO<sub>2</sub> excreted originates from plasma HCO<sub>3</sub><sup>-</sup> that must enter the erythrocyte and be converted to CO<sub>2</sub> by the CA-catalysed reaction before it can be excreted across the respiratory epithelium. The usual pattern of CO<sub>2</sub> excretion in vertebrates (Figure 2c) has been described in a number of reviews <sup>9,53-55</sup>.

# 3. The agnathan solution

The jawless (agnathan) vertebrates, comprising modern hagfishes and lampreys as the sole survivors of a once more species-rich group, diverged from the jawed vertebrate lineage some 500 million years ago. Its living descendants differ in important aspects from the general pattern of vertebrate erythrocyte function, including the possession of structurally and functionally different globins and the absence of a rapid anion exchange protein in their erythrocyte membrane.

# 3.1. Cooperativity of oxygen binding in haemoglobin is generated via monomer-oligomer equilibria

In addition to the mechanism of cooperativity utilized by tetrameric Hbs, co-operative  $O_2$  binding can occur if the aggregation of globins is oxygen-dependent and the  $O_2$  affinities of aggregation states differ. In such a case, aggregated Hbs have a low  $O_2$  affinity and monomeric Hbs have a high  $O_2$ affinity <sup>6</sup>. Aggregation-dependent generation of cooperativity is utilized by the jawless vertebrates, hagfish and lampreys. Early studies on agnathan Hbs failed to see aggregation (and cooperativity) <sup>56</sup>,

because dilution favours monomerization <sup>57</sup>. Cooperativity stems from the fact that in the oxygenated state, Hb is monomeric but aggregates upon deoxygenation <sup>57-59</sup>. A schematic depiction of aggregation-dependent  $O_2$  affinity regulation is given in Figure 3. In hagfish it is notable that the different Hb types have markedly different capabilities for aggregation and that the aggregation of some components is influenced by anions like  $HCO_3^{-60}$ . Two significant properties of lamprey Hb within erythrocytes are the marked Bohr effect (with a Bohr coefficient of around -1 <sup>56</sup>, which contrasts with a much lower mammalian value of around -0.5 <sup>61</sup>), and the reduction in the  $O_2$  carrying capacity of the blood at air saturated  $O_2$  tensions at low pH <sup>56</sup>, a phenomenon that is more commonly associated with the blood of teleost fishes <sup>62</sup>, where it is referred to as the 'Root effect' (for review see: <sup>10,63</sup>). In contrast, the Bohr coefficient of hagfish Hbs is small – maximally -0.5 <sup>57,60</sup>, and depends on the proportions of different subunits with different aggregation properties <sup>60</sup>.

On the basis of molecular phylogenetic evidence, it appears that agnathan Hbs with monomeroligomer equilibria are only distantly related to the tetramer-forming Hbs of other Vertebrata <sup>64</sup>, and have actually evolved from cytoglobin-like ancestors <sup>65</sup>. As a consequence, although lampreys have Hb characterized by a pronounced sensitivity to H<sup>+</sup>s, resembling Bohr and Root effects of teleost fish, this H<sup>+</sup> sensitivity has probably evolved independently <sup>66</sup>. Another major difference between agnathan and most tetrameric Hbs is that in the former, organic phosphates do not affect Hb-O<sub>2</sub> affinity <sup>67</sup>, whereas in the latter, the effect is usually pronounced, and crucial in fine-tuning Hb-O<sub>2</sub> affinity <sup>4,32,68</sup>, although some Hbs even in mammals are organic-phosphate-insensitive <sup>69</sup>.

# **3.2.** Agnathan erythrocytes do not have rapid anion transport: implications for carbon dioxide transport

In contrast to erythrocytes of other vertebrates, lamprey erythrocytes were shown to regulate intracellular pH actively <sup>70,71</sup>. This was later shown to be associated with the lack of erythrocyte anion exchange activity <sup>72</sup>. A similar absence of rapid erythrocyte anion exchange was reported at the same time for a member of the other agnathan group, the hagfish *Eptatretus stouti* <sup>73</sup>. The lack of an

anion exchanger has a marked influence on CO<sub>2</sub> transport, as plasma HCO<sub>3</sub><sup>-</sup> is consequently not available to erythrocyte CA for CO<sub>2</sub> excretion, as is a common characteristic of other vertebrates <sup>55,74-</sup> <sup>76</sup>. Because of the short transit time of erythrocytes in the gills, and the lack of anion exchange, virtually all excreted CO<sub>2</sub> stems from erythrocyte HCO<sub>3</sub><sup>-</sup>. The total amount of CO<sub>2</sub> and HCO<sub>3</sub><sup>-</sup> that can be transported within erythrocytes, and thus the efficiency of CO<sub>2</sub> excretion (Figure 2), is very dependent on a high erythrocyte pH, which is much higher in lampreys than hagfishes <sup>76</sup> [about 1 pH unit higher in *Lampetra fluviatilis* than in *Myxine glutinosa* at an extracellular pH of 7.6 <sup>70,77</sup>]. Consequently, lampreys can transport CO<sub>2</sub> much more efficiently than hagfish. This is in keeping with the lifestyles of the groups: whereas hagfish are sluggish and incapable of strenuous activity <sup>78</sup>, lampreys undertake long and vigorous spawning migrations <sup>79</sup>.

# 4. The elasmobranchs

A detailed recent review on O<sub>2</sub> and CO<sub>2</sub> transport in elasmobranchs is available <sup>80</sup>. Although elasmobranch Hbs are mostly tetrameric, there is evidence for oxygenation dependent formation of higher polymers and dissociation to dimers <sup>7</sup>. The histidine content of Hb, and thereby the specific hydrogen ion buffer value in the elasmobranchs studied to date is similar to that in mammals <sup>9,80</sup>, and the value of their Bohr coefficient is small, usually below -0.5 <sup>80,81</sup>. The organic phosphate binding site appears to resemble that of mammals <sup>82</sup>, and their erythrocytes have robust anion exchange <sup>83,84</sup>. Consequently, cartilaginous fishes differ the least among fish from the prototype model of jawed vertebrate gas transport <sup>10-12</sup>. However, there are three specific points which warrant attention with regard to differences in O<sub>2</sub> and CO<sub>2</sub> transport in elasmobranchs as compared to the basic model. First, the plasma osmolality of marine elasmobranchs is about three- to four-fold higher than almost all other vertebrates. While their plasma Na<sup>+</sup> and Cl<sup>-</sup> concentrations are much lower than seawater values, their plasma is isosmotic with their seawater environment because of the high plasma urea concentrations, typically up to 0.5 M <sup>85</sup>. Urea at these levels usually denatures proteins, but urea concentrations even up to 5 M appear to have minimal effects on Hb-O<sub>2</sub> transport

function <sup>86</sup>. Although it appears that in most cartilaginous fishes urea has a minimal effect on Hb function <sup>80,86</sup>, there are exceptions <sup>87</sup>. Urea tends to antagonize the effect of organic phosphates on Hb function in many but not all cartilaginous fish <sup>80</sup>. Second, some shark species are heterothermic, maintaining the temperature of some tissues, such as the red swimming muscle, up to 10-15°C higher than that of the environment <sup>88,89</sup>. Although many of the associated adaptations are circulatory with a multitude of heat-exchanging *retia mirabilia*, Hb function also appears responsive <sup>80</sup>. In heterothermic fish, the effect of temperature on Hb-O<sub>2</sub> affinity may be reduced or even reversed (as discussed in more detail below <sup>80,81</sup>), and in elasmobranchs specifically the allosteric effector ATP may reduce or reverse the temperature-effect <sup>90</sup>. Third, there appear to be unique features in the CO<sub>2</sub> transport of elasmobranchs. Both erythrocytic and plasma-accessible CA in gills appear to play a role in CO<sub>2</sub> excretion <sup>80,91</sup>, whereby some plasma HCO<sub>3</sub><sup>-</sup> can be dehydrated to CO<sub>2</sub> directly from the plasma compartment, without entering the erythrocyte, probably by membranebound extracellular gill cell CA <sup>92</sup>. The model of CO<sub>2</sub> excretion in elasmobranchs is schematically represented as Figure 2c.

# 5. The teleosts

# 5.1. The Root effect: extreme pH sensitivity of the oxygen equilibrium curve.

Although the effect of pH on Hb-O<sub>2</sub> affinity is a property of virtually all Hbs, many teleost Hbs are characterized by extreme pH dependence so that Hbs fail to become fully O<sub>2</sub>-saturated at low pH values regardless of the O<sub>2</sub> tension <sup>12,62,63,93-96</sup>; Fig. 4), a phenomenon known as the Root effect. When the pH becomes so low that the Root effect is induced, the cooperativity of O<sub>2</sub> binding eventually disappears, and Hill's n can even become less than 1 <sup>97</sup>, which is suggestive of large subunit heterogeneity in the O<sub>2</sub> affinity of the T-state ( $\alpha$  and  $\beta$ ) globins <sup>63</sup>.

The physiological role of the Root effect in teleost Hbs can be considered as an acid-triggered mechanism by which oxygenated Hb can release substantial amounts of its bound  $O_2$  even in the presence of a high  $PO_2$  <sup>96</sup>. According to the classical viewpoint, the effect is utilised to generate

Page 11 of 38

#### Acta Physiologica

supra-atmospheric *P*O<sub>2</sub> during one or both of the following processes: (1) the secretion of O<sub>2</sub> into the teleost swim bladder for buoyancy regulation and/or (2) the improved O<sub>2</sub> supply to the often avascular and relatively thick teleost retina. In both locations, the Root effect is thought to be elicited by local tissue acidification and co-occurs with vascular counter-current gas exchangers, the swim bladder *rete mirabile* and the choroid *rete mirabile*, respectively. These *retia mirabilia* allow for any O<sub>2</sub> released via the Root effect to diffuse back from the venous to the arterial side of the *rete mirabile* and thereby multiply the initial increase in its partial pressure. Similarly, the increase in CO<sub>2</sub> that is associated with the initial blood acidification, can be kept localised by its back-diffusion from the venous to the arterial side of the *rete mirabile*. While the roles of the Root effect in O<sub>2</sub> delivery to the swim bladder and eye are generally well accepted in the literature, a large scale evolutionary reconstruction of the origins of the Root effect and the swim bladder and choroid *retia mirabilia* has suggested that the Root effect evolved before the first occurrences of either the retinal or the swim bladder O<sub>2</sub> secretion mechanism <sup>12</sup>, raising the question about the initial roles of the Root effect that may confer selective advantages.

The high pH dependency of teleost fish Hbs is also reflected in the magnitude of the Bohr coefficient, which often exceeds -1, even when calculated relative to plasma pH <sup>97-99</sup>. The structural basis of the marked pH dependency of teleost Hbs has been reviewed earlier <sup>96,97</sup>, and here we focus on how the erythrocyte properties of teleost fish, in light of the highly pH sensitive Hbs, affect gas transport.

The initial evolutionarily adaptive role of the Root effect may have been to facilitate  $O_2$  unloading generally in working muscle of teleost fish <sup>100,101</sup>, making it later possible to utilize the property to give greatly enhanced  $O_2$  secretion to the eye and swim bladder. Oxygen uptake and delivery is maximized by a high Hb- $O_2$  affinity at the gills and a low Hb- $O_2$  affinity in the tissues. In simple terms this means that when pH decreases in the muscle to induce the Root effect, the  $PO_2$  in unloading is markedly increased. Since the rate of diffusion depends directly on the partial pressure gradient, this speeds up the diffusion of  $O_2$  to working muscle markedly.

To set the stage, one needs first to consider how the large Haldane effect of teleost fish Hb, i.e. the markedly higher  $pK_a$  of deoxy- than oxyHb, can theoretically influence arterial and venous erythrocyte pH in the absence of acid loads. Since the  $CO_2/HCO_3$  buffer system plays a reduced role in water breathers and Hb is the major non-bicarbonate buffer in their blood, erythrocyte pH in arterial blood would be close to the overall  $pK_a$  value of oxyHb and that in venous blood close to the higher overall  $pK_a$  value of deoxyHb in the absence of any external acid loads from the tissues, i.e. at constant  $CO_2$  tension. The marked increase of erythrocyte pH appears to stop at 50% Hb- $O_2$  saturation  $^{102}$ . In resting conditions the deoxygenation-induced increase in erythrocyte pH is also transmitted to plasma pH. This means that theoretically venous blood can have a pH value that is higher than that of arterial blood even in the presence of an overall acid load. The gradient between erythrocyte and plasma pH in selected conditions is illustrated in Figure 5.

Induction of the Root effect requires a significant, species-dependent, decrease of erythrocyte pH. Assuming that metabolism causes a tissue acid load, which is adequate to cause a 30 % decrease of the maximal  $O_2$  saturation, that the Hb concentration is 100 g/L and that arterial blood is virtually fully  $O_2$ -saturated, then the release of  $O_2$  to the capillary network can be adequate to increase the  $O_2$  tension to 500 mmHg – to a value more than ten times greater than what is found in the absence of significant acid load in rainbow trout (*Oncorhynchus mykiss*)<sup>103</sup>. Consequently, the diffusion gradient for  $O_2$  transfer could be increased dramatically in fish with Root effect Hbs. Such an increase will drastically increase the  $O_2$  available to swimming muscle, whereby the Root effect would markedly favour  $O_2$  delivery to the muscle of strenuously exercising teleost fish. Notably, the above is only a theoretical calculation and assumes that blood is in a closed system. While that is true only for blood in the arterial and venous system, the capillary circulation is at best a semi-open system, since the volume (of fish) which is in contact with tissue capillaries is limited. The only truly open part of the circulation is the gill circulation, where blood is in close contact with the near infinite volume of water in the ambient environment.

#### Acta Physiologica

In line with the above prediction, we have observed that the O<sub>2</sub> tension of blood in the caudal vessel of striped bass (*Morone saxatilis*) was significantly increased (from 118.6 to 173.3 mmHg) and supraatmospheric after 5-min chasing <sup>104</sup>. The pH of sampled blood after exercise was markedly reduced (by 0.3 pH units) during exercise, and may be sufficiently low to induce the Root effect in this species.

In conclusion, the drastically increased  $O_2$  unloading potential due to the extreme pH dependence of Root effect Hbs in teleost fish may represent the basis for a highly efficient  $O_2$  transport system relative to that found in air-breathers, which is also coupled with a more efficient respiratory gas exchange system (counter current exchange <sup>2</sup>). However, the use of Root effect Hbs for acidtriggered, sustained and drastic increases in muscle PO<sub>2</sub> values is of benefit only, if the pH effects in the erythrocyte can be reversed in the time it takes such tissue capillary blood to reach the  $O_2$ uptake site in the gills. Recent work on salmonids in particular, referred to in the following section, suggests that this is made possible by the  $\beta$ -adrenergically stimulated sodium/proton (Na<sup>+</sup>/H<sup>+</sup>) exchange in their erythrocytes <sup>105</sup>.

# 5. 2. The adrenergically stimulated, oxygen-dependent sodium-proton exchanger of erythrocytes

Erythrocyte pH can be rapidly increased in teleost fish through an adrenergically stimulated Na<sup>+</sup>/H<sup>+</sup> exchange <sup>12,42,106</sup>. Adrenergically stimulated Na<sup>+</sup>/H<sup>+</sup> exchange has also been found in erythrocytes of amphibians <sup>107-109</sup>; however, activation in these cells is not rapid <sup>109</sup> precluding a role in the short term regulation of O<sub>2</sub> transport. The adrenergic Na<sup>+</sup>/H<sup>+</sup> exchange activity of teleost erythrocytes is strongly oxygenation dependent <sup>110-112</sup>, and results in rapid net H<sup>+</sup> extrusion following stimulation <sup>113,114</sup>. The increase in erythrocyte pH<sub>i</sub> is dependent upon the absence of CA in the plasma which would otherwise short-circuit this response by rapidly catalysing the conversion of plasma HCO<sub>3</sub><sup>-</sup> and extruded H<sup>+</sup>s to CO<sub>2</sub> <sup>113,114</sup>. Also, a marked effect of the Na<sup>+</sup>/H<sup>+</sup> exchange on erythrocyte pH requires that the buffering capacity of Hb is low. Indeed, the buffering capacity of teleost Hbs is much lower than that of other vertebrate groups <sup>115</sup>. The extremely rapid H<sup>+</sup> extrusion and its O<sub>2</sub> dependency

appear unique for teleost erythrocytes, although  $O_2$  dependency in the adrenergically stimulated Na<sup>+</sup>/H<sup>+</sup> exchange of the amphibian *Bufo marinus*<sup>109</sup> and in other ion transport pathways of vertebrate erythrocytes <sup>116-119</sup> has been described. Common to those ion transport pathways is that they seem to be involved in volume regulation, which does not appear to require as rapid a response as that in the  $O_2$  transport cascade. The  $O_2$  dependency of Na<sup>+</sup>/H<sup>+</sup> exchange appears to be generated not by molecular oxygen but by hydroxyl radicals <sup>120</sup>, suggesting that oxyradicals are not only toxicants but also important signalling molecules in animals <sup>121</sup>.

Characterization of the  $\beta$ -adrenergic receptors of teleost erythrocytes yielded a surprise: the receptors were clearly of the  $\beta_3$ -subtype <sup>122</sup>. Since this receptor subtype, characterized both genetically and pharmacologically, is associated with thermoregulation and fat metabolism in homeothermic vertebrates <sup>123</sup> but in teleost fish with regulation of O<sub>2</sub> transport in stress, it represents an intriguing example of how the same receptor type can evolve to regulate completely different functions <sup>124</sup>.

# 5.3. The simultaneous presence of Root effect and adrenergic Na<sup>+</sup>/H<sup>+</sup> exchange maximizes oxygen unloading from erythrocytes but enables effective oxygen uptake in gills

Although the Root effect and marked pH dependency of Hb-O<sub>2</sub> affinity may increase O<sub>2</sub> unloading during a generalized blood acidosis (such as occurs in hypoxia or exhaustive exercise), O<sub>2</sub> uptake at the gills would decrease if erythrocytic pH<sub>i</sub> did not recover prior to gill entry. However, recent studies on rainbow trout have shown that the erythrocytic pH recovery occurs well within the venous transit time of the erythrocyte from the tissues back to the gills <sup>105</sup> as a result of the adrenergic stimulation of erythrocyte Na<sup>+</sup>/H<sup>+</sup> exchange, whereby pH<sub>i</sub> is largely independent of pH<sub>e</sub> and consequently O<sub>2</sub> loading in gills can be maintained. While the activation of erythrocyte Na<sup>+</sup>/H<sup>+</sup> exchange ensures O<sub>2</sub> loading in the gills, it could prevent any pH decrease upon further acid load in tissues due to tight regulation of pH<sub>i</sub>. However, the effect of Na<sup>+</sup>/H<sup>+</sup> exchange on erythrocytic pH<sub>i</sub> can be reduced by speeding up the extracellular hydration/dehydration reactions between HCO<sub>3</sub><sup>-</sup> and

Page 15 of 38

## Acta Physiologica

CO<sub>2</sub>, as their rate in comparison to the rate of H<sup>+</sup> extrusion determines the extent of the change in pH<sub>i</sub><sup>114,125</sup>. Thus, a decrease in erythrocyte pH<sub>i</sub> at the tissues depends mainly on the activity of extracellular (i.e. plasma-accessible) CA: the greater the activity, the greater the reduction in pH<sub>i</sub> and consequently the decrease in Hb-O<sub>2</sub> affinity and increase in  $O_2$  unloading. In keeping with conditions that could maximize O<sub>2</sub> delivery in the presence of erythrocyte Na<sup>+</sup>/H<sup>+</sup> exchange activation, plasma accessible CA is absent in the gills of most teleosts that have been studied <sup>126,127</sup>, but present in some tissues such as the muscle and the heart <sup>101,128,129</sup> of some species. The differential distribution of plasma accessible CA (absence in the gills and presence in the tissues) creates conditions for greatly enhanced  $O_2$  unloading that could more than double  $O_2$  unloading with no change in tissue perfusion <sup>100</sup>. The potential of this system has been demonstrated in rainbow trout blood both *in vitro* <sup>130</sup> and in vivo <sup>101</sup>, as well as in other salmonids <sup>131</sup>. Most recently the potential has been demonstrated in a more derived teleost, cobia (Rachycentron canadum; Shu and Brauner, unpublished). Functional evidence for the role of this system in enhancing O<sub>2</sub> unloading has been demonstrated in Atlantic salmon (Salmo salar), where injection of a plasma accessible CA inhibitor (C18) in fish swimming at a moderate speed induced a 30% increase in cardiac output to compensate, and at higher swimming speeds fish collapsed (Harter, T.S., Zanuzzo, F.S., Supuran, C.T., Gamperl, A.K. and Brauner, C.J., unpublished). Clearly more studies are required to determine just how widespread this system is among teleosts and its functional importance given that there are more than 25000 teleost fish species with marked differences in the presence and activity of the erythrocyte Na<sup>+</sup>/H<sup>+</sup> exchange of those species where it has been investigated <sup>12,112,132</sup>. Furthermore, the differential tissue distribution of CA has mainly been studied in salmonids.

In conclusion, maximal efficiency of  $O_2$  transport in teleosts requires that the high pH dependency of Hb- $O_2$  affinity, adrenergic activation of erythrocytic Na<sup>+</sup>/H<sup>+</sup> exchange and differential distribution of plasma-accessible CA function in concert (Fig. 6). Alone, each of the phenomena could cause problems for gas transport, but together, they form a highly efficient system that differs markedly

from other vertebrates. How the different components have evolved has been discussed by Randall et al., 2014 <sup>127</sup> and Harter and Brauner, 2017 <sup>133</sup>, but remains a fruitful area for further investigation.

#### 5.4. Temperature effects on teleost haemoglobins: ectothermy and heterothermy

It has been known for over a century that some species of tuna can maintain parts of their body at a temperature much higher than that of the ambient environment <sup>134-136</sup>, a feature shared by some sharks <sup>88</sup>, as discussed above and known as regional heterothermy. The swimming muscle in the body core has a high temperature, which permits improved muscle performance, but the temperature at the body surface and the gills is close to ambient. The marked difference in temperature between the body surface and the core can be maintained by the organization of blood vessels in retia mirabilia in which arteries with warm blood from the core flow counter-current, and in close proximity to the veins with cold blood from the body surface <sup>137</sup>. This system allows for effective heat conservation in those tissues. If the effect of temperature on Hb-O<sub>2</sub> affinity were similar to that of other vertebrates, this could lead to a pronounced loss of O<sub>2</sub> from arteries to veins within the *retia mirabilia* so that the associated peripheral tissues could suffer from  $O_2$  lack. As early as 1960 it was observed that at 50% saturation, the O<sub>2</sub> affinity of tuna Hb was not affected by temperature <sup>138</sup>. It was later observed that in bluefin tuna blood, a reverse temperature effect exists, where Hb-O<sub>2</sub> affinity actually increased with increasing temperature at saturations above 50% and decreased at low saturations <sup>139,140</sup>. While the reversed temperature effect appears counterproductive for  $O_2$  release and  $O_2$  consumption in core muscle, this trait is thought to minimize the loss of  $O_2$  from arteries to veins in *retia mirabilia*. Interestingly, the blood of some ectothermic marine fishes lack the effect of temperature on Hb- O<sub>2</sub> affinity <sup>47,99</sup>, suggesting that a pre-existing trait was utilized in the evolution of Hb function of regionally heterothermic fish.

Regional heterothermy in fishes may involve the whole core of the fish being regulated above ambient temperature, opah (*Lampris guttatus*) as the most extreme case <sup>141</sup> or only a specific tissue such as the eye/brain area as in billfish <sup>46,142</sup>. It is usually thought that in regional heterothermy the

# Acta Physiologica

effects of temperature on Hb-O<sub>2</sub> affinity are reduced or reversed <sup>45,139</sup>. At least two factors influence the overall heat of oxygenation: exothermic O<sub>2</sub> binding to the haem groups of Hb and the endothermic dissociation of allosteric effectors (mainly H<sup>+</sup>s and organic phosphates) from the Hb <sup>45,46</sup>. Because both enthalpies differ depending on the oxygenation state of Hb, the apparent heat of oxygenation also depends on Hb-O<sub>2</sub> saturation <sup>140</sup>.

Apart from the evolution of Hb function in response to temperature in regional heterothermy, the ambient temperature to which fish are adapted appears to have had little influence on functional characteristics of Hb. This is important to note, when considering the possibilities of fish to adapt to increased temperatures, brought about by climate change. It was initially thought that fish from extremely cold, Arctic and Antarctic, habitats, where the temperature is stable and can be close to - 2°C, would have reduced temperature dependency of Hb-O<sub>2</sub> affinity <sup>143,144</sup>. However, more recently it has been shown that temperature sensitivity of Hb-O<sub>2</sub> binding in Antarctic fish appears similar to temperate fish <sup>145</sup>. A complicating factor in evaluating the effect of temperature on Hb-O<sub>2</sub> binding is that, e.g., the enthalpy of the binding of ATP to Hb is influenced by pH <sup>146</sup>, and pH is affected acutely by a change in temperature and following temperature acclimation, both of which can influence the apparent relationship between temperature and Hb-O<sub>2</sub> affinity.

# 6. Evolutionary convergence of responses to achieve similar organismic traits

Similar to adrenergically-stimulated erythrocytes in teleost fishes, erythrocyte pH is also largely independent from plasma pH in the agnathan lampreys. In the latter this feature is mainly due to the presence of constitutively active O<sub>2</sub>-dependent Na<sup>+</sup>/H<sup>+</sup> exchange <sup>147</sup> in the absence of rapid Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchange <sup>72</sup> which results in a high erythrocyte pH <sup>70,71,148</sup>. This is in contrast with the other agnathan group, hagfish, which also lack the anion exchanger <sup>73</sup>, but have low erythrocyte pH because of minimal Na<sup>+</sup>/H<sup>+</sup> exchange or other H<sup>+</sup> extrusion mechanisms <sup>149</sup>. Thus, in lampreys and teleosts, although the mechanism through which erythrocyte pH is to some degree independent of plasma pH is different, the final outcome is similar (convergent evolution). The evolutionary reasons for this

convergent evolution are necessarily speculative. However, at least two possibilities are plausible. First, in both groups of water-breathers, intensive short-term bouts of exercise occur. Since the erythrocyte pH<sub>i</sub> (and thereby Hb-O<sub>2</sub> affinity) can be maintained high in gills, the exercise-induced metabolic acidosis does not appreciably reduce O<sub>2</sub> binding in gills. Second, both teleost fish and lampreys are characterized by highly pH-sensitive Hbs with low buffer values <sup>9</sup>. It is plausible that because of this they cannot afford to lose the ability to regulate erythrocytic pH<sub>i</sub> via Na<sup>+</sup>/H<sup>+</sup> exchange. Regardless of the reason, although the lamprey and teleost fish lineages diverged about 500 million years ago <sup>150</sup>, O<sub>2</sub> transport by erythrocytes has converged to enable similar organismal function.

# 7. Erythrocytes – not only for gas transport: roles in buffering and cell signalling

The above has concentrated completely on respiratory gas transport. However, while this is undoubtedly the primary function of erythrocytes, they possess many other important properties. First, as Hb is a very abundant protein with many histidine residues that can donate or accept H\*s, Hb is a very important extracellular buffer as long as the erythrocytic Hb is accessible to extracellular acid loads. The buffering capacity of Hb is proportional to the histidine content, which is markedly reduced in teleost fish, but high in lungfish, elasmobranchs and tetrapods <sup>11,12,115</sup>. Among the agnathans, the histidine content is distinctly higher in hagfish than in lampreys <sup>9,57</sup>, which is also reflected in their respective H\* buffering capacity in both lampreys and teleost fish may be to benefit O<sub>2</sub> transport characteristics (see above). Haemoglobin has a pronounced role in rapid extracellular buffering as long as erythrocytes have rapid anion exchange <sup>76</sup>, as is the case for all vertebrates except agnathans <sup>72,73</sup>. Because HCO<sub>3</sub><sup>-</sup> transport across the agnathan erythrocyte membrane is exceedingly slow <sup>74,75</sup>, erythrocytic Hb cannot rapidly buffer extracellular acid loads. Although a number of explanations are possible, perhaps this limited ability to buffer an extracellular acid load is associated with a more temperate distribution where metabolic rate and thus metabolic acid loads

#### Acta Physiologica

are reduced. This antitropical distribution is clearly seen for lampreys, which tend to have freshwater ammocoetes (usually river-resident lamprey larvae) with a low thermal tolerance <sup>152</sup>.

Another role for Hb is its influence on redox balance and nitrite-nitrate-nitric oxide equilibrium. This may have an influence on the regulation of erythrocyte adrenergic Na<sup>+</sup>/H<sup>+</sup> exchangers and other oxygenation-dependent ion transporters. For example, potassium transport in crucian carp erythrocytes appears to have two different oxygenation-dependent sensors <sup>119</sup>. One appears to be a hydroxyl radical sensor <sup>121,153</sup>, where Hb may be the primary regulator of hydroxyl radical level. Based on mammalian studies, Hb appears to be a biological Fenton reagent with deoxyhaemoglobin as the form responsible for hydroxyl radical generation <sup>154</sup>. So far, the sensing mechanism of the other O<sub>2</sub> sensor described has not been clarified.

Mammalian studies also suggest that Hb is an oxido-reductase, largely depending on its level of oxygenation <sup>155</sup>. It plays an important role in regulating vasodilatation and vasoconstriction by influencing nitric oxide (and nitrite) level. When Hb is oxygenated, it scavenges NO and the vessels are constricted. When Hb is deoxygenated, it reduces nitrite to form NO, whereby vessels dependent on NO signalling are dilated <sup>155</sup>. Marked formation of NO from nitrite occurs in the erythrocytes of a teleost fish, carp <sup>156</sup>. Further, the nitrite reductase activity correlates with the Hb-O<sub>2</sub> affinity, and changes when the erythrocyte NTP level changes <sup>157</sup>, the latter being modulated in response to hypoxia in fish <sup>68</sup>. In view of these findings Hb appears to play a part in NO-dependent signalling. This is appropriate, as NO-signalling plays a role in vascular tone, i.e. regulating O<sub>2</sub> transport to capillaries. Another signalling molecule, hydrogen sulphide, has also been demonstrated to be especially important in regulating the vasculature in response to hypoxia <sup>158,159</sup>. Not surprisingly, Hb regulates the sulphide turnover both in fish and humans <sup>160,161</sup>. There are significant species-dependent differences both in the formation of sulphaemoglobin <sup>162</sup> and in the effects of sulphide on membrane ion transport <sup>163</sup> in the erythrocytes of teleost fish, but the physiological significance of these differences has not been evaluated. The involvement of Hb in sulphide

signalling may be a reason for the continuous presence of ferric methaemoglobin, which does not carry O<sub>2</sub>, but enables reversible sulphide reactions <sup>160,161</sup>. Also another gaseous signalling molecule, CO, interacts and is regulated by Hb at least in mammals <sup>164</sup>. The overall picture that emerges is that gaseous signalling plays an important role in erythrocyte function and that Hb is important in its regulation. However, so far this remains a little studied area, which would clearly be worthy of further investigation.

# 8. The functions of "new" globins

Tetrameric Hb is at very high concentration within erythrocytes with most fish expressing multiple isoforms <sup>165</sup>. However, in addition, fish erythrocytes transcribe at least cytoglobin, globin x and neuroglobin <sup>166</sup>. In particular, a high mRNA level of the gene encoding neuroglobin was found. In three-spined stickleback, neuroglobin transcription in erythrocytes was the most active of any tissue, and even more active than that of Hb <sup>166</sup>. The function of neuroglobin is far from clear, but it has been suggested to play a role in redox or NO regulation <sup>167</sup> and in regulating free sulphide levels <sup>168</sup>. As discussed above, all of these signalling systems may be active in erythrocytes and can involve Hb. Thus, neuroglobin formation in teleost erythrocytes may be related to redox, NO or sulphide regulation. Globin x is a membrane-bound globin <sup>169</sup>, which can also take part in either redox <sup>170</sup> or NO <sup>171</sup> regulation. Another plausible function for globin x would be in the regulation of adrenergic Na<sup>+</sup>/H<sup>+</sup> exchanger. It was proposed that Hb may be a regulator of the erythrocyte adrenergic Na<sup>+</sup>/H<sup>+</sup> exchanger <sup>172</sup>. However, properties of bulk Hb did not fit the requirements for the O<sub>2</sub> sensor <sup>118</sup>. A minor, membrane-bound globin with O<sub>2</sub> affinity different from bulk Hb, such as globin x, on the other hand, could be involved.

# 9. Conclusions

From the preceding sections it is clear that the erythrocyte functions of fishes are underpinned by various unique mechanisms, which are vastly different in the different fish groups. These erythrocyte functions often deviate from the prototype textbook dogma. Thus when using fish in environmental

# Acta Physiologica

 research, their phylogenetic relationships need to be taken into account. When fish are used with biomedical questions in mind, it must be remembered that their O<sub>2</sub> transport system can be far more efficient than that in mammals because of the need for efficient O<sub>2</sub> extraction from their low-O<sub>2</sub> environment. It is not well understood how the O<sub>2</sub>-dependent phenomena beyond gas transport are different in zebrafish and humans. Perhaps the major difference between human and fish erythrocytes is that the former do not possess nuclei and other cellular organelles. Consequently, while mammalian erythrocytes are devoid of gene expression and produce energy anaerobically, fish erythrocytes are aerobic, and many of their adaptations can involve active protein production. Notably, however, the effectiveness of gene expression decreases with the age of the erythrocyte <sup>27</sup>, adding age-dependent selective removal of erythrocytes to possible regulatory mechanisms behind gas transport. Such selective removal of erythrocytes would affect, e.g., the adrenergic responsiveness of the erythrocytes <sup>173</sup>, and affect the seasonality of the responses of erythrocytes <sup>174</sup>.

# **Conflict of interest**

There is no conflict of interest.

## Acknowledgements

CJB was supported by a Natural Sciences and Engineering Research Council of Canada (NSERC) Discovery Grant (2018-04172). We thank Jacelyn Shu for creating Figure 2 and Ari Karhilahti for drawing the rainbow trout for Figure 6.

# 10. References

- 1. Nikinmaa M. Respiratory adjustments of rainbow trout (Salmo gairdneri Richardson) to changes in environmental temperature and oxygen availability, University of Helsinki, Finland; 1981.
- 2. Piiper J. Respiratory gas exchange at lungs, gills and tissues: mechanisms and adjustments. *J exp Biol.* 1982;100:3-22.

- 3. Piiper J, Dejours P, Haab P, Rahn H. Concepts and basic quantities in gas exchange physiology. *Respir Physiol.* 1971;13:292-304.
  - 4. Weber RE, Jensen FB. Functional adaptations in hemoglobins from ectothermic vertebrates. *Annu Rev Physiol.* 1988;50:161-179.
- 5. Graham JB. *Air-Breathing Fishes*. San Diego: Academic Press; 1997.
- 6. Perutz M. *Mechanisms of cooperativity and allosteric regulation in proteins.* Cambridge: Cambridge University Press; 1990.
- 7. Fyhn UEH, Sullivan B. Elasmobranch hemoglobins: dimerization and polymerization in various species. *Comp Biochem Physiol*. 1975;50B:119-129.
- 8. Wells RMG. Haemoglobin function in aquatic animals: molecular adaptations to environmental challenge. *Marine And Freshwater Research*. 1999;50(8):933-939.
- 9. Nikinmaa M. *Vertebrate red blood cells.* Vol 28. Berlin-Heidelberg-New York: Springer-verlag; 1990.
- 10. Brauner CJ, Berenbrink M. Gas Transport and Exchange. In: David JM, ed. Fish Physiology

Primitive Fishes. Volume 26 ed.: Academic Press; 2007:213-282.

- 11. Berenbrink M. Evolution of vertebrate haemoglobins: Histidine side chains, specific buffer value and Bohr effect. *Respiratory Physiology & Neurobiology.* 2006;154(1-2):165-184.
- 12. Berenbrink M, Koldkjaer P, Kepp O, Cossins AR. Evolution of oxygen secretion in fishes and the emergence of a complex physiological system. *Science*. 2005;307(5716):1752-1757.
- 13. Engeszer RE, Patterson LB, Rao AA, Parichy DM. Zebrafish in the wild: a review of natural history and new notes from the field. *Zebrafish*. 2007;4(1):21-40.
- 14. Barrionuevo WR, Fernandes MN, Rocha O. Aerobic and anaerobic metabolism for the zebrafish, Danio rerio, reared under normoxic and hypoxic conditions and exposed to acute hypoxia during development. *Brazilian Journal of Biology.* 2010;70(2):425-434.
- 15. Abdallah SJ, Thomas BS, Jonz MG. Aquatic surface respiration and swimming behaviour in adult and developing zebrafish exposed to hypoxia. *J Exp Biol.* 2015;218(Pt 11):1777-1786.
- 16. Steindal IAF, Whitmore D. Circadian Clocks in Fish-What Have We Learned so far? *Biology-Basel.* 2019;8(1).
- 17. Nikinmaa M, Rees BB. Oxygen-dependent gene expression in fishes. *American Journal Of Physiology-Regulatory Integrative And Comparative Physiology*. 2005;288(5):R1079-R1090.
- 18. Egg M, Koblitz L, Hirayama J, et al. Linking Oxygen to Time: The Bidirectional Interaction Between the Hypoxic Signaling Pathway and the Circadian Clock. *Chronobiology International.* 2013;30(4):510-529.
- 19. Pelster B, Egg M. Hypoxia-inducible transcription factors in fish: expression, function and interconnection with the circadian clock. *Journal of Experimental Biology*. 2018;221(13).
- 20. Sandbichler AM, Jansen B, Peer BA, Paulitsch M, Pelster B, Egg M. Metabolic Plasticity Enables Circadian Adaptation to Acute Hypoxia in Zebrafish Cells. *Cellular Physiology and Biochemistry*. 2018;46(3):1159-1174.
- 21. Bishop C, Surgenor DM. *The red blood cell: a comprehensive treatise*. New York: Academic Press; 1964.
- 22. Eddy FB. Oxygen uptake by rainbow trout blood, *Salmo gairdneri*. *J Fish Biol*. 1977;10:87-90.
- 23. Ferguson RA, Boutilier RG. Metabolic-membrane coupling in red blood cells of trout:the effects of anoxia and adrenergic stimulation. *J exp Biol.* 1989;143:149-164.
- 24. Sephton DH, Macphee WL, Driedzic WR. Metabolic enzyme activities, oxygen consumption and glucose utilization in sea raven (*Hemitripterus americanus*) erythrocytes. *J exp Biol.* 1991;159:407-418.
- 25. Walsh PJ, Wood CM, Thomas S, Perry SF. Characterization of red blood cell metabolism in rainbow trout. *J exp Biol.* 1990;154:475-489.
- Lund SG, Phillips MCL, Moyes CD, Tufts BL. The effects of cell ageing on protein synthesis in rainbow trout (*Oncorhynchus mykiss*) red blood cells. *Journal of Experimental Biology*. 2000;203(14):2219-2228.

1		
2 3	27.	Gotting M, Nikinmaa MJ. Transcriptomic Analysis of Young and Old Erythrocytes of Fish.
4	۷٦.	Frontiers in Physiology. 2017;8.
5	28.	Cheng CHC, Detrich HW. Molecular ecophysiology of Antarctic notothenioid fishes.
6	20.	Philosophical Transactions of the Royal Society B-Biological Sciences. 2007;362(1488):2215-
7		2232.
8 9	29.	Portner HO, Peck L, Somero G. Thermal limits and adaptation in marine Antarctic
9 10	25.	ectotherms: an integrative view. <i>Philosophical Transactions of the Royal Society B-Biological</i>
10		Sciences. 2007;362(1488):2233-2258.
12	30.	Bohr C, Hasselbalch KA, Krogh A. Uber einen in biologischer Beziehung wichtigen Einfluss,
13	50.	den die Kohlensaurespaunung des Blutes auf dessen Sauerstoffbindung ubt. Skand Arch
14		Physiol. 1904;16:402-412.
15	31.	Benesch R, Benesch RE. The effect of organic phosphates from the human erythrocyte on
16	51.	the allosteric properties of hemoglobin. <i>Biochem Biophys Res Comm.</i> 1967;26:162-167.
17	32.	Chanutin A, Curnish RR. Effect of organic and inorganic phosphates on the oxygen
18 19	52.	equilibrium of human erythrocytes. Arch Biochem Biophys. 1967;121:96-102.
20	33.	Bauer C. On the respiratory function of haemoglobin. <i>Rev Physiol Biochem Pharmacol.</i>
20	55.	1974;70:1-31.
22	34.	Mairbaurl H, Weber RE. Oxygen Transport by Hemoglobin. <i>Comprehensive Physiology.</i>
23	54.	2012;2:1463-1489.
24	25	Bunn HF. Regulation of hemoglobin function in mammals. <i>Am Zool.</i> 1980;20:199-211.
25	35. 36.	
26	50.	Dickerson RE, Geis I. <i>Hemoglobin: Structure, function, evolution, and pathology.</i> Menlo Park:
27	27	Benjamin/Cummings; 1983.
28 29	37.	Bunn HF, Forget BG. <i>Hemoglobin: molecular, genetic and clinical aspects.</i> 690 pp ed.
30	38.	Philadelphia: W.B.Saunders; 1986.
31	50.	Storz JF. <i>Hemoglobin. Insights into Protein Structure, Function, and Evolution.</i> Oxford: Oxford University Press; 2018.
32	39.	Giardina B, Condo SG, Sherbini SE, et al. Arctic life adaptation-I. The function of reindeer
33	59.	hemoglobin. Comp Biochem Physiol. 1989;94B:129-133.
34	40.	Bauer C, Forster M, Gros G, et al. Analysis of bicarbonate binding to crocodilian hemoglobin.
35	40.	J Biol Chem. 1981;256:8429-8435.
36	41.	Wyman J, Jr. Linked functions and reciprocal effects in hemoglobin: a second look. Adv
37 38	41.	Protein Chem. 1964;19:223-286.
30 39	42.	Nikinmaa M. Membrane transport and the control of haemoglobin-oxygen affinity in
40	42.	nucleated erythrocytes. <i>Physiol Rev.</i> 1992;72:301-321.
41	43.	Nikinmaa M, Weber RE. Hypoxic acclimation in the lamprey, <i>Lampetra fluviatilis</i> : Organismic
42	45.	and erythrocytic responses. J exp Biol. 1984;109:109-119.
43	44.	Mayo KH, Chien JCW. Effect of temperature on functional properties of carp hemoglobin. J
44	44.	Mol Biol. 1980;142:63-73.
45	45.	Weber RE, Campbell KL. Temperature dependence of haemoglobin-oxygen affinity in
46 47	45.	heterothermic vertebrates: mechanisms and biological significance. Acta Physiol (Oxf).
47 48		2011;202(3):549-562.
49	46.	Weber RE, Campbell KL, Fago A, Malte H, Jensen FB. ATP-induced temperature
50	40.	independence of hemoglobin-O2 affinity in heterothermic billfish. <i>J exp Biol.</i> 2010;213(Pt
51		9):1579-1585.
52	47	•
53	47.	Clark TD, Rummer JL, Sepulveda CA, Farrell AP, Brauner CJ. Reduced and reversed
54		temperature dependence of blood oxygenation in an ectothermic scombrid fish:
55		implications for the evolution of regional heterothermy? <i>Journal Of Comparative Physiology</i>
56 57	40	B-Biochemical Systemic And Environmental Physiology. 2010;180(1):73-82.
57 58	48.	Carter MJ. Carbonic anhydrase: isozymes, properties, distribution and functional
58 59		significance. Biol Rev (Cambridge). 1972;47:465-513.
60		

49.	Maren TH. Carbonic anhydrase: chemistry, physiology, and inhibition. <i>Physiol Rev.</i> 1967;47:595-781.
50.	Tanner MJA. Molecular and cellular biology of the erythrocyte anion exchanger (AE1). <i>Semin Hematol.</i> 1993;30(1):34-57.
51.	Hamasaki N, Okubo K. Band 3 protein: Physiology, function and structure. <i>Cell Mol Biol.</i> 1996;42:1025-1039.
52.	Jacobs MH, Stewart DR. The role of carbonic anhydrase in certain ionic exchanges involving the erythrocyte. <i>J Gen Physiol.</i> 1942;25:539-552.
53.	Klocke RA. Carbon dioxide transport. In: Farhi LE, Tenney SM, eds. <i>Handbook of Physiology.The respiratory system.Vol IV.Gas exchange.</i> Bethesda Maryland: American
54.	Physiological Society; 1987:173-197. Roughton FJW. Transport of oxygen and carbon dioxide. In: Fenn WO, Rahn H, eds. Handbook of physiology. Respiration Vol I. Washington D.C.: American Physiological Society;
55.	1964:767-825. Tufts BL, Perry SF. Carbon dioxide transport and excretion. In: Perry SF, Tufts BL, eds. <i>Fish</i> <i>Respiration.</i> Vol 17. San Diego: Academic Press; 1998:229-281.
56.	Nikinmaa M. Haemoglobin function in intact <i>Lampetra fluviatilis</i> erythrocytes. <i>Respir Physiol.</i> 1993;91:283-293.
57.	Fago A, Weber RE. Hagfish haemoglobins. In: Jorgensen JM, Lomholt JP, Weber RE, Malte H, eds. <i>The Biology of Hagfishes</i> . London: Chapman & Hall; 1998:321-333.
58.	Briehl RW. The relation between the oxygen equilibrium and aggregation of subunits in lamprey hemoglobin. <i>J Biol Chem.</i> 1963;238:2361-2366.
59.	Qiu Y, Maillett DH, Knapp J, Olson JS, Riggs AF. Lamprey hemoglobin - Structural basis of the Bohr effect. <i>Journal of Biological Chemistry</i> . 2000;275(18):13517-13528.
60.	Fago A, Giangiacomo L, D'Avino R, et al. Hagfish hemoglobins - Structure, function, and oxygen-linked association. <i>Journal of Biological Chemistry</i> . 2001;276(29):27415-27423.
61.	Lapennas GN. The magnitude of the Bohr coefficient: optimal for oxygen delivery. <i>Respir Physiol.</i> 1983;54:161-172.
62.	Root RW. The respiratory function of the blood of marine fishes. <i>Biol Bull mar biol Lab</i> <i>Woods Hole.</i> 1931;61:427-457.
63.	Berenbrink M. TRANSPORT AND EXCHANGE OF RESPIRATORY GASES IN THE BLOOD   Root Effect: Molecular Basis, Evolution of the Root Effect and Rete Systems. In: Farrell AP, ed. Encyclopedia of Fish Physiology. San Diego: Academic Press; 2011:935-943.
64.	Hoffmann FG, Opazo JC, Storz JF. Gene cooption and convergent evolution of oxygen transport hemoglobins in jawed and jawless vertebrates. <i>Proc Natl Acad Sci U S A</i> . 2010;107(32):14274-14279.
65.	Burmester T, Hankeln T. Function and evolution of vertebrate globins. <i>Acta Physiologica</i> . 2014;211(3):501-514.
66.	Berenbrink M. TRANSPORT AND EXCHANGE OF RESPIRATORY GASES IN THE BLOOD   Evolution of the Bohr Effect. In: Farrell AP, ed. <i>Encyclopedia of Fish Physiology</i> . San Diego: Academic Press; 2011:921-928.
67.	Nikinmaa M, Weber RE. Gas transport in lamprey erythrocytes. In: Bicudo JEPW, ed. <i>The vertebrate gas transfer cascade. Adaptations to environment and mode of life</i> . Boca Raton, Florida: CRC Press; 1993:179-187.
68.	Wood SC, Johansen K. Adaptation to hypoxia by increased HbO <sub>2</sub> affinity and decreased red cell ATP concentration. <i>Nature New Biol.</i> 1972;237:278-279.
69.	Bunn HF. Differences in the interaction of 2,3-diphosphoglycerate with certain mammalian hemoglobins. <i>Science</i> . 1971;172(3987):1049-1050.
70.	Nikinmaa M. Red cell pH of lamprey ( <i>Lampetra fluviatilis</i> ) is actively regulated. <i>J Comp Physiol B.</i> 1986;156:747-750.

7	<ol> <li>Nikinmaa M, Kunnamo-Ojala T, Railo E. Mechanisms of pH regulation in lamprey (Lampetra fluviatilis) red blood cells. J exp Biol. 1986;122:355-367.</li> </ol>
7	<ol> <li>Nikinmaa M, Railo E. Anion movements across lamprey (<i>Lampetra fluviatilis</i>) red cell membrane. <i>Biochim Biophys Acta</i>. 1987;899:134-136.</li> </ol>
7	3. Ellory JC, Wolowyk MW, Young JD. Hagfish ( <i>Eptatretus stouti</i> ) erythrocytes show minimal chloride transport activity. <i>J exp Biol.</i> 1987;129:377-383.
	4. Tufts BL, Boutilier RG. The absence of rapid chloride/bicarbonate exchange in lamprey erythrocytes: implications for CO <sub>2</sub> transport and ion distributions between plasma and erythrocytes in the blood of <i>Petromyzon marinus</i> . J exp Biol. 1989;144:565-576.
7	<ol> <li>Tufts BL, Boutilier RG. CO<sub>2</sub> transport in agnathan blood: evidence of erythrocyte Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchange limitations. <i>Respir Physiol.</i> 1990;80:335-348.</li> </ol>
7	<ol> <li>Nikinmaa M. Oxygen and carbon dioxide transport in vertebrate erythrocytes: An evolutionary change in the role of membrane transport. <i>J exp Biol.</i> 1997;200:369-380.</li> </ol>
7	7. Tufts BL, Boutilier RG. CO <sub>2</sub> transport properties of the blood of a primitive vertebrate <i>Myxine</i> glutinosa. Exp Biol. 1990;48:341-347.
7	<ol> <li>Martini FH. The ecology of hagfishes. In: J.M. J, Lomholt JP, Weber RE, Malte H, eds. The biology of hagfishes. London: Chapman &amp; Hall; 1998:57-78.</li> </ol>
7	<ol> <li>McKenzie DJ, Hale ME, Domenici P. Locomotion in Primitive Fishes. In: <i>Fish Physiology.</i> Vol 26. Academic Press; 2007:319-380.</li> </ol>
8	<ol> <li>Morrison PR, Gilmour KM, Brauner CJ. 3 - Oxygen and Carbon Dioxide Transport in Elasmobranchs. In: Shadwick RE, Farrell AP, Brauner CJ, eds. <i>Fish Physiology</i>. Vol 34. Academic Press; 2015:127-219.</li> </ol>
8	1. Andersen ME, Olson JS, Gibson QH, Carey FG. Studies on ligand binding to hemoglobins from teleosts and elasmobranchs. <i>J Biol Chem.</i> 1973;248:331-341:331-341.
8	2. Aschauer H, Weber RE, Braunitzer G. The primary structure of the hemoglobin of the dogfish shark ( <i>Squalus acanthias</i> ). Antagonist effects of ATP and urea on oxygen affinity on an elasmobranch hemoglobin. <i>Biol Chem Hoppe-Seyler</i> . 1985;366:589-599.
8	<ol> <li>Musch MW, Davis EM, Goldstein L. Oligomeric forms of skate erythrocyte band 3 - Effect of volume expansion. J Biol Chem. 1994;269(31):19683-19686.</li> </ol>
8	4. Perlman DF, Musch MW, Goldstein L. Band 3 in cell volume regulation in fish erythrocytes. <i>Cell Mol Biol.</i> 1996;42:975-984.
8	5. Yancey PH. 4 - Organic Osmolytes in Elasmobranchs. In: Shadwick RE, Farrell AP, Brauner CJ, eds. Fish Physiology. Vol 34. Academic Press; 2015:221-277.
8	6. Ingermann RL. Vertebrate Hemoglobins. <i>Comprehensive Physiology</i> . 2011:357-408.
	7. Weber RE, Wells RMG, Tougaard S. Antagonistic effect of urea on oxygenation-linked binding of ATP in an elasmobranch hemoglobin. <i>Life Sci.</i> 1983;32:2157-2161.
8	<ol> <li>Carey FG, Teal JM. Mako and porbeagle: warm-bodied sharks. <i>Comp Biochem Physiol.</i> 1969;28:199-204.</li> </ol>
8	<ol> <li>Patterson JC, Sepulveda CA, Bernal D. The vascular morphology and in vivo muscle temperatures of thresher sharks (Alopiidae). J Morphol. 2011;272(11):1353-1364.</li> </ol>
g	<ol> <li>Larsen C, Malte H, Weber RE. ATP-induced reverse temperature effect in isohemoglobins from the endothermic porbeagle shark (Lamna nasus). J Biol Chem. 2003;278(33):30741- 30747.</li> </ol>
g	1. Gilmour KM, Perry SF. Branchial membrane-associated carbonic anhydrase activity maintains CO2 excretion in severely anemic dogfish. <i>American Journal Of Physiology-Regulatory</i> Integrative And Comparative Physiology. 2004;286(6):R1138-R1148.
g	2. Gilmour KM, Bayaa M, Kenney L, McNeill B, Perry SF. Type IV carbonic anhydrase is present in the gills of spiny dogfish (Squalus acanthias). <i>American Journal Of Physiology-Regulatory</i> <i>Integrative And Comparative Physiology</i> . 2007;292(1):R556-R567.

93. Scholander PV, Van Dam L. Secretion of gases against high pressure in the swimbladders of deep sea fishes I. Oxygen dissociation in blood. Biol Bull mar biol Lab Woods Hole. 1954;107:247-259. 94. Brittain T. The Root effect. Comp Biochem Physiol. 1987;86B:473-481. 95. Berenbrink M, Koldkjaer P, Hannah Wright E, Kepp O, Jose da Silva A. Magnitude of the Root effect in red blood cells and haemoglobin solutions of fishes: a tribute to August Krogh. Acta 10 Physiol (Oxf). 2011;202(3):583-592. 11 96. Berenbrink M. Historical reconstructions of evolving physiological complexity: O-2 secretion 12 in the eye and swimbladder of fishes. Journal of Experimental Biology. 2007;210(9):1641-13 1652. 14 97. Jensen FB, Fago A, Weber RE. Hemoglobin structure and function. In: Perry SF, Tufts BL, eds. 15 Fish Respiration. San Diego: Academic Press; 1998:1-40. 16 98. Nikinmaa M, Soivio A. Oxygen dissociation curves and oxygen capacities of blood of a 17 freshwater fish, Salmo gairdneri. Ann Zool Fennici. 1979;16:217-221. 18 19 99. Barlow SL, Metcalfe J, Righton DA, Berenbrink M. Life on the edge: O2 binding in Atlantic cod 20 red blood cells near their southern distribution limit is not sensitive to temperature or 21 haemoglobin genotype. J Exp Biol. 2017;220(Pt 3):414-424. 22 100. Rummer JL, Brauner CJ. Root Effect Haemoglobins in Fish May Greatly Enhance General 23 Oxygen Delivery Relative to Other Vertebrates. *Plos One.* 2015;10(10). 24 101. Rummer JL, Mckenzie DJ, Innocenti A, Supuran CT, Brauner CJ. Root Effect Hemoglobin May 25 Have Evolved to Enhance General Tissue Oxygen Delivery. Science. 2013;340(6138):1327-26 1329. 27 102. Jensen FB. Pronounced influence of Hb-O<sub>2</sub> saturation on red cell pH in tench blood in vivo 28 29 and in vitro. J Exp Zool. 1986;238:119-124. 30 103. Soivio A, Nikinmaa M, Nyholm K, Westman K. The role of gills in the responses of Salmo 31 gairdneri during moderate hypoxia. Comp Biochem Physiol. 1981;70A:133-139. 32 104. Nikinmaa M, Cech JJ, Jr., McEnroe M. Blood oxygen transport in stressed striped bass 33 (Morone saxatilis): role of beta-adrenergic responses. J Comp Physiol B. 1984;154:365-369. 34 105. Harter TS, May AG, Federspiel WJ, Supuran CT, Brauner CJ. The time-course of red blood cell 35 intracellular pH recovery following short-circuiting in relation to venous transit times in 36 rainbow trout, Oncorhynchus mykiss. Am J Physiol Regul Integr Comp Physiol. 2018. 37 106. Nikinmaa M. Effects of adrenaline on red cell volume and concentration gradient of protons 38 39 across the red cell membrane in the rainbow trout, Salmo gairdneri. Mol Physiol. 40 1982;2:287-297. 41 107. Palfrey HC, Greengard P. Hormone-sensitive ion transport systems in erythrocytes as models 42 for epithelial ion pathways. Ann N Y Acad Sci. 1981;372:291-308. 43 108. Kaloyianni M, Rasidaki A. Adrenergic responses of R. ridibunda red cells. J Exp Zool. 44 1996;276:175-185. 45 109. Kristensen K, Koldkjae P, Berenbrink M, Wang T. Oxygen-sensitive regulatory volume 46 47 increase and Na transport in red blood cells from the cane toad, Bufo marinus. Journal of 48 Experimental Biology. 2007;210(13):2290-2299. 49 110. Salama A. The role of cAMP in regulating the beta-adrenergic response of rainbow trout 50 (Oncorhynchus mykiss) red blood cells. Fish Physiol Biochem. 1993;10(6):485-490. 51 111. Salama A, Nikinmaa M. The adrenergic responses of carp (Cyprinus carpio) red cells: effects 52 of Po<sub>2</sub> and pH. J exp Biol. 1988;136:405-416. 53 112. Salama A, Nikinmaa M. Species differences in the adrenergic responses of fish red cells: 54 studies on whitefish, pikeperch, trout and carp. Fish Physiol Biochem. 1989;6:167-173. 55 113. Motais R, Fievet B, Garcia-Romeu F, Thomas S. Na<sup>+</sup>-H<sup>+</sup> exchange and pH regulation in red 56 57 blood cells: role of uncatalyzed H<sub>2</sub>CO<sub>3</sub> dehydration. *Am J Physiol.* 1989;256:C728-C735. 58 59 60

1 2 3

4

5

6

7

8

2	
3	
3 4 5 6 7 8	
5	
6	
7	
, 8	
9	
10	
11 12	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24	
25	
26	
27	
28	
20 21 22 23 24 25 26 27 28 29	
30 31 32 33 34	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
58	
59	
60	

- Nikinmaa M, Tiihonen K, Paajaste M. Adrenergic control of red cell pH in salmonid fish: Roles of the sodium/proton exchange, Jacobs-Stewart cycle and membrane potential. *J exp Biol.* 1990;154:257-271.
   Jensen FB. Hydrogen ion equilibria in fish haemoglobins. *J exp Biol.* 1989;143:225-234:225-234.
   Gibson JS, Cossins AR, Ellory JC. Oxygen-sensitive membrane transporters in vertebrate red cells. *Journal of Experimental Biology.* 2000;203(9):1395-1407.
   Virkki LV, Salama A, Nikinmaa M, Begulation of ion transport across lamprey (*Lampetra*)
- Virkki LV, Salama A, Nikinmaa M. Regulation of ion transport across lamprey (*Lampetra fluviatilis*) erythrocyte membrane by oxygen tension. *J exp Biol.* 1998;201:1927-1937.
   Dependent M, Valkel G, Heisler M, Nikinmaa MA, Q, dependent Kt fluence in tracet and black
- 118. Berenbrink M, Volkel S, Heisler N, Nikinmaa M.  $O_2$ -dependent K<sup>+</sup> fluxes in trout red blood cells: the nature of  $O_2$  sensing revealed by the  $O_2$  affinity, cooperativity and pH dependence of transport. *Journal Of Physiology-London*. 2000;526(1):69-80.
- 119. Berenbrink M, Volkel S, Koldkjaer P, Heisler N, Nikinmaa M. Two different oxygen sensors regulate oxygen-sensitive K+ transport in crucian carp red blood cells. *Journal Of Physiology-London*. 2006;575(1):37-48.
- 120. Nikinmaa M, Bogdanova A, Lecklin T. Oxygen dependency of the adrenergic Na/H exchange in rainbow trout erythrocytes is diminished by a hydroxyl radical scavenger. *Acta Physiologica Scandinavica*. 2003;178(2):149-154.
- 121. Bogdanova A, Berenbrink M, Nikinmaa M. Oxygen-dependent ion transport in erythrocytes. *Acta Physiologica*. 2009;195(3):305-319.
- 122. Nickerson JG, Dugan SG, Drouin G, Perry SF, Moon TM. Activity of the unique β-adrenergic Na<sup>+</sup>/H<sup>+</sup> exchanger in trout erythrocytes is controlled by a novel  $\beta_{3-AR subtype}$ . Am J Physiol Regul Integr Comp Physiol. 2003.
- 123. Strosberg AD. Structure and function of the beta(3)-adrenergic receptor. *Annual Review of Pharmacology and Toxicology*. 1997;37:421-450.
- 124. Nikinmaa M. beta(3)-Adrenergic receptors studies on rainbow trout reveal ancient evolutionary origins and functions distinct from the thermogenic response. *American Journal Of Physiology-Regulatory Integrative And Comparative Physiology*. 2003;285(3):R515-R516.
- 125. Nikinmaa M, Boutilier RG. Adrenergic control of red cell pH, organic phosphate concentrations and haemoglobin function in teleost fish. In: Heisler N, ed. *Mechanisms of systemic regulation: Respiration and circulation.* Berlin: Springer; 1996.
- 126. Gilmour KM, Perry SF. Carbonic anhydrase and acid-base regulation in fish. *Journal of Experimental Biology*. 2009;212(11):1647-1661.
- 127. Randall DJ, Rummer JL, Wilson JM, Wang S, Brauner CJ. A unique mode of tissue oxygenation and the adaptive radiation of teleost fishes. *Journal of Experimental Biology*. 2014;217(8):1205-1214.
- 128. Wang Y, Henry RP, Wright PM, Heigenhauser GJ, Wood CM. Respiratory and metabolic functions of carbonic anhydrase in exercised white muscle of trout. *Am J Physiol.* 1998;275(6):R1766-1779.
- 129. Alderman SL, Harter TS, Wilson JM, Supuran CT, Farrell AP, Brauner CJ. Evidence for a plasma-accessible carbonic anhydrase in the lumen of salmon heart that may enhance oxygen delivery to the myocardium. *Journal of Experimental Biology*. 2016;219(5):719-724.
- 130. Rummer JL, Brauner CJ. Plasma-accessible carbonic anhydrase at the tissue of a teleost fish may greatly enhance oxygen delivery: in vitro evidence in rainbow trout, Oncorhynchus mykiss. *Journal of Experimental Biology.* 2011;214(14):2319-2328.
- 131. Shu JJ, Harter TS, Morrison PR, Brauner CJ. Enhanced hemoglobin-oxygen unloading in migratory salmonids. *Journal of Comparative Physiology B-Biochemical Systemic and Environmental Physiology*. 2018;188(3):409-419.
- 132. Hyde DA, Perry SF. Absence of adrenergic red cell pH and oxygen content regulation in American eel (*Anguilla rostrata*) during hypercapnic acidosis in vivo and in vitro. *J Comp Physiol B.* 1990;159:687-693.

133.	Harter TS, Brauner CJ. 1 - The O2 and CO2 Transport System in Teleosts and the Specialized Mechanisms That Enhance Hb–O2 Unloading to Tissues. In: Gamperl AK, Gillis TE, Farrell AP,
	Brauner CJ, eds. Fish Physiology. Vol 36. Academic Press; 2017:1-106.
134.	Carey FG. Fishes with warm bodies. <i>Sci Amer.</i> 1973;228:36-44.
135.	Carey FG, Teal JM. Regulation of body temperature by the bluefin tuna. <i>Comparative Biochemistry and Physiology.</i> 1969;28(1):205-213.
136.	Fudge DS, Stevens ED. The visceral retia mirabilia of tuna and sharks: an annotated
	translation and discussion of the Eschricht & Müller 1835 paper and related papers <i>Guelph Ichtyology Reviews.</i> 1996;4(1-92).
137.	Carey FG, Teal JM. Heat conservation in tuna fish muscle. <i>Proc Natl Acad Sci U S A.</i> 1966;56(5):1464-1469.
138.	Rossi-Fanelli A, Antonini E. Oxygen equilibrium of haemoglobin from Thunnus thynnus. Nature. 1960;186:895-896.
139.	Carey FG, Gibson QH. Reverse temperature dependence of tuna hemoglobin oxygenation.
	Biochem Biophys Res Commun. 1977;78:1376-1382.
140.	Ikeda-Saito M, Yonetani T, Gibson QH. Oxygen equilibrium studies on hemoglobin from the bluefin tuna ( <i>Thunnus thynnus</i> ). <i>J Mol Biol.</i> 1983;168:673-686.
141.	Wegner NC, Snodgrass OE, Dewar H, Hyde JR. Animal physiology. Whole-body endothermy in a mesopelagic fish, the opah, Lampris guttatus. <i>Science</i> . 2015;348(6236):786-789.
142.	Carey FG. A brain heater in the swordfish. <i>Science</i> . 1982;216(4552):1327-1329.
143.	di Prisco G, Condo SG, Tamburrini M, Giardina B. Oxygen transport in extreme
	environments. Trends in Biochemical Sciences. 1991;16:471-474.
144.	di Prisco G, D'Avino R, Camardella L, Caruso C, Romano M, Rutigliano B. Structure and
	function of hemoglobin in Antarctic fishes and evolutionary implications. <i>Polar Biol.</i> 1990;10:269-274.
145.	Fago A, Wells RMG, Weber RE. Temperature-dependent enthalpy of oxygenation in Antarctic fish hemoglobins. <i>Comp Biochem Physiol B.</i> 1997;118:319-326.
146.	Greaney GS, Hobish MK, Powers DA. The effects of temperature and pH on the binding of ATP to carp ( <i>Cyprinus carpio</i> ) deoxyhemoglobin. <i>J Biol Chem</i> . 1980;255:445-453.
147.	Virkki LV, Nikinmaa M. Activation and physiological role of Na <sup>+</sup> /H <sup>+</sup> exchange in lamprey ( <i>Lampetra fluviatilis</i> ) erythrocytes. <i>J exp Biol.</i> 1994;191:89-105.
148.	Boutilier RG, Ferguson RA, Henry RP, Tufts BL. Exhaustive Exercise in the Sea Lamprey
	(Petromyzon marinus) - Relationship Between Anaerobic Metabolism and Intracellular Acid Base Balance. <i>J exp Biol.</i> 1993;178:71-88:71-88.
149.	Nikinmaa M, Tufts BL, Boutilier RG. Volume and pH regulation in agnathan erythrocytes -
	Comparisons between the hagfish, <i>Myxine glutinosa</i> , and the lampreys, <i>Petromyzon marinus</i> and <i>Lampetra fluviatilis</i> . J Comp Physiol B. 1993;163(7):608-613.
150.	Janvier P. Living Primitive Fishes and Fishes From Deep Time. In: <i>Fish Physiology.</i> Vol 26. Academic Press; 2007:1-51.
151.	Jensen FB. Haemoglobin H+ equilibria in lamprey (Lampetra fluviatilis) and hagfish (Myxine glutinosa). <i>J Exp Biol.</i> 1999;202 (Pt 14):1963-1968.
152.	Potter IC, Gill HS, Renaud CB, Hauocher D. The Taxonomy, Phylogeny, and Distribution of Lampreys. In: Docker MF, ed. <i>Lampreys: Biology, Conservation and Control.</i> Dordrecht,
	Germany: Springer Science+Business Media; 2015.
153.	Bogdanova A, Nikinmaa M. Reactive oxygen species regulate oxygen-sensitive potassium flux in rainbow trout erythrocytes. <i>J Gen Physiol</i> . 2001;117(2):181-190.
154.	Sadrzadeh SM, Graf E, Panter SS, Hallaway PE, Eaton JW. Hemoglobin. A biologic fenton reagent. <i>J Biol Chem.</i> 1984;259(23):14354-14356.
155.	Helms CC, Gladwin MT, Kim-Shapiro DB. Erythrocytes and Vascular Function: Oxygen and Nitric Oxide. <i>Front Physiol.</i> 2018;9:125.

156.	Jensen FB. Nitric oxide formation from nitrite in zebrafish. <i>Journal of Experimental Biology</i> . 2007;210(19):3387-3394.
157.	Jensen FB, Kolind RAH, Jensen NS, Montesanti G, Wang T. Interspecific variation and plasticity in hemoglobin nitrite reductase activity and its correlation with oxygen affinity in vertebrates. <i>Comparative Biochemistry and Physiology a-Molecular &amp; Integrative Physiology.</i> 2017;206:47-53.
158.	Olson KR. Vascular actions of hydrogen sulfide in nonmammalian vertebrates. <i>Antioxidants &amp; Redox Signaling</i> . 2005;7(5-6):804-812.
159.	Olson KR, Whitfield NL, Bearden SE, et al. Hypoxic pulmonary vasodilation: a paradigm shift with a hydrogen sulfide mechanism. <i>American Journal Of Physiology-Regulatory Integrative And Comparative Physiology</i> . 2010;298(1):R51-R60.
160.	Jensen B, Fago A. Reactions of ferric hemoglobin and myoglobin with hydrogen sulfide under physiological conditions. <i>J Inorg Biochem.</i> 2018;182:133-140.
161.	Bianco CL, Savitsky A, Feelisch M, Cortese-Krott MM. Investigations on the role of hemoglobin in sulfide metabolism by intact human red blood cells. <i>Biochem Pharmacol.</i> 2018;149:163-173.
162.	Volkel S, Berenbrink M. Sulphaemoglobin formation in fish: A comparison between the haemoglobin of the sulphide-sensitive rainbow trout ( <i>Oncorhynchus mykiss</i> ) and of the sulphide-tolerant common carp ( <i>Cyprinus carpio</i> ). <i>Journal of Experimental Biology</i> . 2000;203(6):1047-1058.
163.	Volkel S, Berenbrink M, Heisler N, Nikinmaa M. Effects of sulfide on K+ flux pathways in red blood cells of crucian carp and rainbow trout. <i>Fish Physiology and Biochemistry</i> . 2001;24(3):213-223.
164.	Motterlini R, Foresti R. Biological signaling by carbon monoxide and carbon monoxide- releasing molecules. <i>Am J Physiol Cell Physiol.</i> 2017;312(3):C302-C313.
165.	Weber RE. Functional significance and structural basis of multiple hemoglobins with special reference to ectothermic vertebrates. In: Truchot J-P, Lahlou B, eds. <i>Animal nutrition and transport processes 2. Transport, respiration and excretion:comparative and environmental aspects.</i> Basel: Karger; 1990:58-75.
166.	Gotting M, Nikinmaa M. More than hemoglobin - the unexpected diversity of globins in vertebrate red blood cells. <i>Physiol Rep.</i> 2015;3(2).
167.	Burmester T, Hankeln T. What is the function of neuroglobin? <i>Journal of Experimental Biology</i> . 2009;212(10):1423-1428.
168.	Ascenzi P, di Masi A, Leboffe L, et al. Neuroglobin: From structure to function in health and disease. <i>Molecular Aspects of Medicine</i> . 2016;52:1-48.
169.	Blank M, Wollberg J, Gerlach F, et al. A Membrane-Bound Vertebrate Globin. <i>Plos One.</i> 2011;6(9).
170.	Koch J, Burmester T. Membrane-bound globin X protects the cell from reactive oxygen species. <i>Biochemical and Biophysical Research Communications</i> . 2016;469(2):275-280.
171.	Corti P, Xue JM, Tejero J, et al. Globin X is a six-coordinate globin that reduces nitrite to nitric oxide in fish red blood cells. <i>Proceedings of the National Academy of Sciences of the United States of America</i> . 2016;113(30):8538-8543.
172.	Motais R, Garcia-Romeu F, Borgese F. The control of Na <sup>+</sup> /H <sup>+</sup> exchange by molecular oxygen in trout erythrocytes. A possible role of hemoglobin as a transducer. <i>J Gen Physiol.</i> 1987;90:197-207.
173.	Lecklin T, Tuominen A, Nikinmaa M. The adrenergic volume changes of immature and mature rainbow trout ( <i>Oncorhynchus mykiss</i> ) erythrocytes. <i>Journal of Experimental Biology</i> . 2000;203(19):3025-3031.
174.	Koldkjaer P, Pottinger TG, Perry SF, Cossins AR. Seasonality of the red blood cell stress response in rainbow trout (Oncorhynchus mykiss). <i>Journal of Experimental Biology</i> . 2004;207(2):357-367.

175. Brauner CJ, Gilmour KM, Perry SF. Effect of haemoglobin oxygenation on Bohr proton release and CO<sub>2</sub> excretion in the rainbow trout. *Resp Physiol.* 1996;106:65-70.

## **Figure captions**

**Figure 1 a.**  $O_2$  equilibrium curves with  $O_2$  saturation (%) of Hb on *y*-axis and the partial pressure of  $O_2$  (kPa) of the Hb solution on *x*-axis. Often, the  $O_2$  affinity of Hb is given as the  $P_{50}$  value, which is the partial pressure of  $O_2$  at which Hb is 50%  $O_2$ -saturated. The effect of decreasing pH is shown by a right shift of the  $O_2$  equilibrium curve from A to B. The numerical value for the pH effect (Bohr coefficient) is usually given as  $\Delta \log P_{50}$  value/ $\Delta pH$ . **b.** The interaction between  $O_2$ -binding Hb subunits (*n*; sigmoidality) is given by the Hill plot, where the *y*-axis is the logarithm of the ratio of the oxygenated Hb fraction (S) over the deoxygenated Hb fraction (1-S) [log (S/(1-S))] and the *x*-axis is the logarithm of the partial pressure of  $O_2$ ). **1.** gives the log  $P_{50}$  value for Hb in the *R*-state , 2. the slope of the line is *n* (interaction between  $O_2$ -binding Hb subunits), 3. gives the log  $P_{50}$  value for *T*-state Hb, and 4. the overall log  $P_{50}$  value (=the same as the log  $P_{50}$  value derived from OEC).

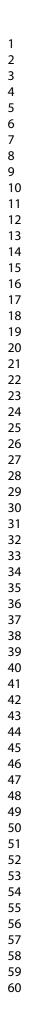
**Figure 2.** Schematic representations of basic patterns of  $CO_2$  excretion in vertebrates. In the agnathans, hagfish (a) and lamprey (b), sharks (c) and teleosts (d). In the agnathans, the red blood cells lack Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchange. In the hagfish, carbonic anhydrase (CA) catalyzes the conversion of HCO<sub>3</sub><sup>-</sup> to CO<sub>2</sub> from both the plasma compartment (via plasma accessible CA; CA-IV) and the erythrocyte. In lamprey, CA-IV is absent and all HCO<sub>3</sub><sup>-</sup> converted to CO<sub>2</sub> is from the erythrocyte and oxygenation of Hb provides H<sup>+</sup> for this reaction through a large Haldane effect. In sharks, the presence of CA-IV permits CO<sub>2</sub> directly from the plasma compartment and erythrocyte Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchange permits plasma HCO<sub>3</sub><sup>-</sup> to enter the erythrocyte for conversion to CO<sub>2</sub>. This pattern represents the general vertebrate pattern of CO<sub>2</sub> excretion in vertebrates and is similar in airbreathers. In teleosts, CA-IV is absent and all CO<sub>2</sub> excreted is through the erythrocyte where a tight coupling of O<sub>2</sub> uptake and CO<sub>2</sub> excretion exists due to the presence of a large Haldane effect.

**Figure 3.** The generation of cooperativity by aggregation/dissociation reactions in lamprey Hbs. Monomers have the highest  $O_2$  affinity, and oxygenation favours dissociation of oligomers to monomers. Dissociation of oligomers to monomers is also favoured by an increase in pH and dilution.

**Figure 4.**  $O_2$  equilibrium curves of rainbow trout **A.** at resting plasma pH (7.65) and **B.** at pH 7.40, where the Root effect has been induced (i.e.  $O_2$  saturation approaches asymptotically a value below full saturation, in this case maximal  $O_2$  saturation is 69%, i.e. Root effect 31%; data from <sup>98</sup>).

**Figure 5.** Arterio-venous pH difference (dpH) in rainbow trout **A.** theoretically, if the only thing affecting blood pH were the difference in  $pK_a$  value between oxy- and deoxyHb (data based on <sup>175</sup>), **B.** in normoxic conditions, and **C.** in moderate hypoxia (40% air saturation) (data for b and c from <sup>103</sup>). A decrease in the negative value of dpH (as compared to A) or an increase in its positive value indicates acid input to blood.

**Figure 6.** Schematic representation on how the presence of Root effect Hb, adrenergic Na<sup>+</sup>/H<sup>+</sup> exchange of erythrocyte membrane, and the absence of carbonic anhydrase in the gills and its presence in the muscle allow to secure O<sub>2</sub> loading at gills, but increase O<sub>2</sub> unloading in muscle of strenuously exercised rainbow trout. In gills, there is no plasma-accessible carbonic anhydrase. Thus, the adrenergically activated Na<sup>+</sup>/H<sup>+</sup> exchange increases erythrocyte pH after the blood has left the muscle. Consequently, the Hb-O<sub>2</sub> affinity increases and O<sub>2</sub> loading in gills remains effective (i.e. Hb reaches close to full saturation) despite plasma acidification. From gills, blood flows to working muscle, which has plasma-accessible carbonic anhydrase. Consequently, erythrocyte pH decreases, even though the adrenergic Na<sup>+</sup>/H<sup>+</sup> exchange of membrane is active. The pH drop can be adequate to induce the Root effect, whereby O<sub>2</sub> is released from Hb and increases the partial pressure of O<sub>2</sub> and speeds up diffusion to O<sub>2</sub>-requiring structures. The changes occur at physiologically relevant time scales.



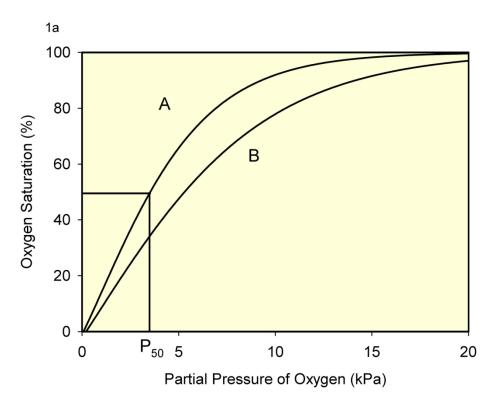


Figure 1 a. O2 equilibrium curves with O2 saturation (%) of Hb on y-axis and the partial pressure of O2 (kPa) of the Hb solution on x-axis. Often, the O2 affinity of Hb is given as the P50 value, which is the partial pressure of O2 at which Hb is 50% O2-saturated. The effect of decreasing pH is shown by a right shift of the O2 equilibrium curve from A to B. The numerical value for the pH effect (Bohr coefficient) is usually given as  $\Delta \log P50 \text{ value}/\Delta pH$ .

161x121mm (300 x 300 DPI)

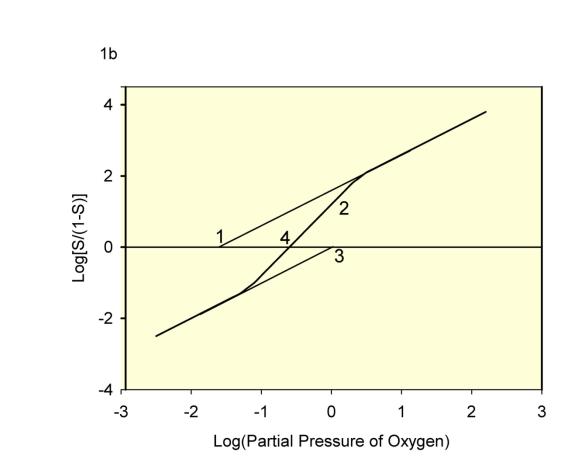
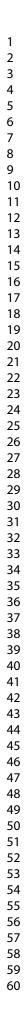


Figure 1b. The interaction between O2-binding Hb subunits (n; sigmoidality) is given by the Hill plot, where the y-axis is the logarithm of the ratio of the oxygenated Hb fraction (S) over the deoxygenated Hb fraction (1-S) [log (S/(1-S))] and the x-axis is the logarithm of the partial pressure of O2). 1. gives the log P50 value for Hb in the R-state , 2. the slope of the line is n (interaction between O2-binding Hb subunits), 3. gives the log P50 value for T-state Hb, and 4. the overall log P50 value (=the same as the log P50 value derived from OEC).

148x121mm (300 x 300 DPI)



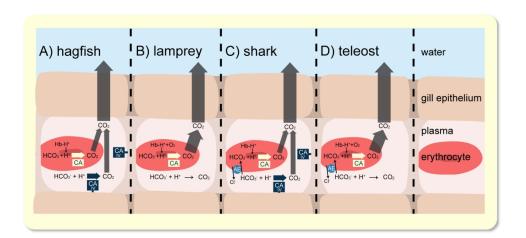
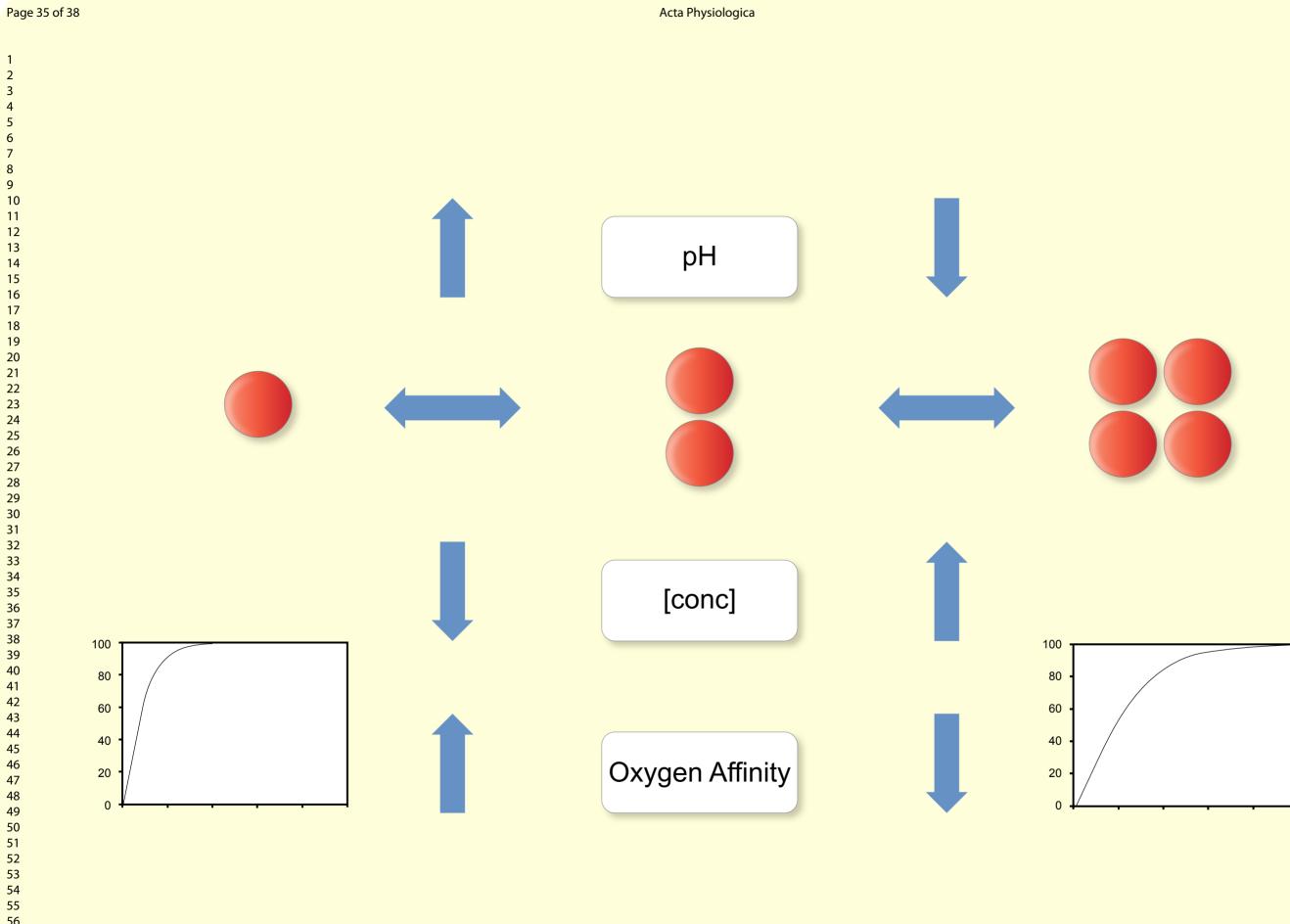


Figure 2. Schematic representations of basic patterns of CO2 excretion in vertebrates. In the agnathans, hagfish (a) and lamprey (b), sharks (c) and teleosts (d). In the agnathans, the red blood cells lack Cl-/HCO3- exchange. In the hagfish, carbonic anhydrase (CA) catalyzes the conversion of HCO3- to CO2 from both the plasma compartment (via plasma accessible CA; CA-IV) and the erythrocyte. In lamprey, CA-IV is absent and all HCO3- converted to CO2 is from the erythrocyte and oxygenation of Hb provides H+ for this reaction through a large Haldane effect. In sharks, the presence of CA-IV permits CO2 directly from the plasma compartment and erythrocyte Cl-/HCO3- exchange permits plasma HCO3- to enter the erythrocyte for conversion to CO2. This pattern represents the general vertebrate pattern of CO2 excretion in vertebrates and is similar in air-breathers. In teleosts, CA-IV is absent and all CO2 excreted is through the erythrocyte where a tight coupling of O2 uptake and CO2 excretion exists due to the presence of a large Haldane effect.

282x211mm (300 x 300 DPI)



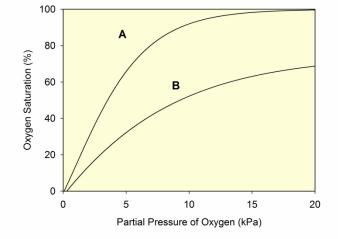
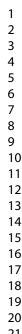


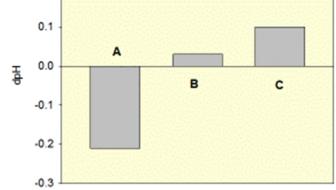
Figure 4. O2 equilibrium curves of rainbow trout A. at resting plasma pH (7.65) and B. at pH 7.40, where the Root effect has been induced (i.e. O2 saturation approaches asymptotically a value below full saturation, in this case maximal O2 saturation is 69%, i.e. Root effect 31%; data from 92).

209x296mm (300 x 300 DPI)

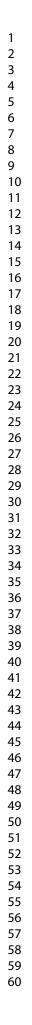


> Figure 5. Arterio-venous pH difference (dpH) in rainbow trout A. theoretically, if the only thing affecting blood pH were the difference in pKa value between oxy- and deoxyHb (data based on 170), B. in normoxic conditions, and C. in moderate hypoxia (40% air saturation) (data for b and c from 97). A decrease in the negative value of dpH (as compared to A) or an increase in its positive value indicates acid input to blood.

> > 190x275mm (96 x 96 DPI)



0.2



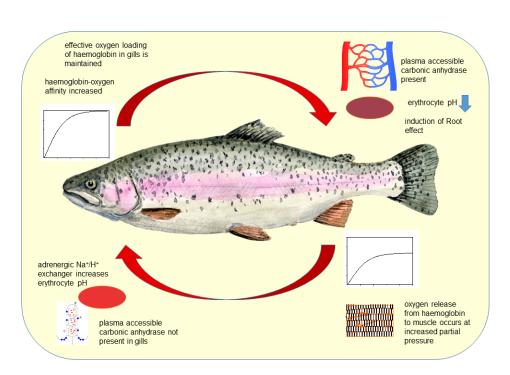


Figure 6. Schematic representation on how the presence of Root effect Hb, adrenergic Na+/H+ exchange of erythrocyte membrane, and the absence of carbonic anhydrase in the gills and its presence in the muscle allow to secure O2 loading at gills, but increase O2 unloading in muscle of strenuously exercised rainbow trout. In gills, there is no plasma-accessible carbonic anhydrase. Thus, the adrenergically activated Na+/H+ exchange increases erythrocyte pH after the blood has left the muscle. Consequently, the Hb-O2 affinity increases and O2 loading in gills remains effective (i.e. Hb reaches close to full saturation) despite plasma acidification. From gills, blood flows to working muscle, which has plasma-accessible carbonic anhydrase. Consequently, erythrocyte pH decreases, even though the adrenergic Na+/H+ exchange of membrane is active. The pH drop can be adequate to induce the Root effect, whereby O2 is released from Hb and increases the partial pressure of O2 and speeds up diffusion to O2-requiring structures. The changes occur at physiologically relevant time scales.

275x190mm (96 x 96 DPI)