

## SIGNALING

# Histone modifiers are oxygen-sensors

## Hypoxia signals directly to chromatin via Histone demethylases to alter gene expression

By Paolo Gallipoli<sup>1, 2, 3, 4</sup> and Brian JP Huntly<sup>1, 2, 3, 4</sup>

1 Approximately 2.6 billion years ago during  
 2 the Proterozoic period, the evolution of pho-  
 3 tosynthesis in cyanobacteria led to the intro-  
 4 duction of the by-product of this reaction, oxy-  
 5 gen, into the Earth's atmosphere (1). This  
 6 great oxidative event heralded the rise of  
 7 multicellular organisms, which are almost  
 8 totally dependent on oxygen as an efficient  
 9 fuel for metabolism and a cofactor in many  
 10 critical physiological enzymatic reactions.  
 11 Central to this adaptation, and to allow cellu-  
 12 lar physiology across a wide range of oxygen  
 13 concentrations (tensions), metazoans have  
 14 evolved the highly conserved hypoxia induc-  
 15 ible factor (HIF) pathway (2). This is im-  
 16 portant for both physiological and patholog-  
 17 ical processes that occur in a hypoxic  
 18 microenvironment, including embryogene-  
 19 sis, stem cell homeostasis, cancer and cardio-  
 20 vascular disease. It has long been observed  
 21 that hypoxia induces histone lysine hyper-  
 22 methylation, a form of epigenetic chromatin  
 23 modification. However, whether this repre-  
 24 sents a direct sensing of oxygen tension or an  
 25 indirect effect, perhaps through the HIF path-  
 26 way, has not been established (4). On page  
 27 XXX and YYY of this issue, Batie *et al.* (5) and  
 28 Chakraborty *et al.* (6) resolve this question,  
 29 demonstrating in different cellular systems  
 30 that the activity of the lysine-specific demeth-  
 31 ylases (KDM), KDM5A and KDM6A, is oxy-  
 32 gen sensitive, identifying them as oxygen  
 33 sensors (5, 6).

34 In ambient normoxic conditions HIF-1 $\alpha$ ,  
 35 the DNA-binding component of the heterodi-  
 36 meric transcription factor complex, HIF, is  
 37 targeted for ubiquitylation and destruction.  
 38 This occurs through hydroxylation on pro-  
 39 line residues in HIF-1 $\alpha$  by the EglN family of  
 40 prolyl hydroxylases (PHDs), which are 2-ox-  
 41 oglutarate and oxygen dependent dioxygen-  
 42 ase enzymes that sense physiological  
 43 changes in oxygen tension, being activated in  
 44 normoxia. However, in hypoxic conditions,  
 45 PHD activity is lost and so HIF-1 $\alpha$  is stabilised  
 46 so it can bind to its partner ARNT (also called  
 47 HIF-1 $\beta$ ). The HIF complex translocates to the  
 48 nucleus and induces hypoxia-specific gene  
 49 expression programmes that mediate alter  
 50 cellular metabolism and survival, through  
 51 binding to specialised hypoxia response ele-  
 52 ments (HRE) in gene promoters. The family  
 53 of 2-oxoglutarate and oxygen dependent di-  
 54 oxygenases is large with more than 60 mem-  
 55 bers (3), and also includes the TET and JmjC

KDMs families of epigenetic regulators.

Using biochemical analysis of recombi-  
 nant proteins, Batie *et al.* and Chakraborty *et al.*  
 add KDM5A and KDM6A to the list of diox-  
 ygenases that have low oxygen affinities ( $K_M$   
 values) comparable to the EglN PHD family.  
 Batie *et al.* then used time course experi-  
 ments to show that histone methylation  
 changes following the induction of hypoxia  
 were rapid and preceded, although predic-  
 tive of subsequent transcriptional events,  
 preceded these changes. Importantly, in cel-  
 lular systems expressing loss-of-function  
 and gain-of-function mutant proteins in the  
 HIF pathway and through documenting the  
 speed of HIF-1 $\alpha$  stabilisation following in-  
 duction of hypoxia, histone methylation  
 changes were found to be independent of  
 HIF and also not dependent on other known  
 hypoxia-inducible inhibitors of KDM activity,  
 such as reactive oxygen species and 2-hy-  
 droxyglutarate. Linking direct histone hyper-  
 methylation to cellular function, Chakraborty  
*et al.* demonstrated that hyper-  
 methylation of lysine 27 of histone H3  
 (H3K27), a histone change that is associated  
 with gene repression, prevented differentia-  
 tion in different cell line model systems. Con-  
 versely, these effects could be antagonised by  
 inhibition of the reciprocal histone methyl-  
 transferase, enhancer of zeste homolog 2  
 (EZH2). Batie *et al.* focussed on histone meth-  
 ylation modifications associated with gene  
 activation, lysine 36 of histone H3 (H3K36)  
 and particularly trimethylation of lysine 4 of  
 histone 3 (H3K4me3). They identified  
 KDM5A as responsible for the hypermethyl-  
 ation of H3K4 in their HeLa, human cervical  
 cancer cell line system and linked H3K4me3  
 to the induction of enhancer (long range pro-  
 moters of transcription) activity and both  
 HIF-dependent and -independent promoter  
 function. Both studies report preliminary  
 data regarding the structural basis of differ-  
 ences in oxygen affinity between these two  
 KDMs.

These two complementary studies fur-  
 ther clarify the regulatory mechanisms of  
 histone demethylases and specifically how  
 these directly, rather than through the HIF  
 pathway or via the influence of metabolic in-  
 termediates coordinate a range of epigenetic  
 alterations, transcriptional outputs and cell  
 fate decisions in response to external envi-

ronmental changes (see the figure). How-  
 ever, these studies raise as many important  
 questions as they answer. The alteration of a  
 large number of histone modifications evi-  
 dent in the multiplexed mass spectrometric  
 assay upon hypoxia and described in the lit-  
 erature (5, 7, 8) suggests that the full identity  
 of oxygen-sensitive JmjC KDMs is not yet  
 known. For example, the increased  
 H3K36me3 in response to hypoxia, is not  
 linked to loss of KDM5A or KDM6A activity  
 and so the responsible KDM needs to be  
 identified. Further studies are therefore war-  
 ranted, with this knowledge not only inform-  
 ing biology but also facilitating elucidation of  
 the structural basis of oxygen affinity in  
 KDMs.

Additionally, the HIF pathway has been  
 implicated in an array of physiological and  
 pathological cellular processes and it is likely  
 that the direct oxygen-sensing KDM path-  
 ways are similarly implicated in a number of  
 these processes. However, the exact nature  
 of the pathways involved and whether they  
 function independently and/or coopera-  
 tively with HIF-mediated transcriptional  
 programmes remains to be determined. Ma-  
 lignancies often develop and/or metastasise  
 to hypoxic environments and activation of  
 the HIF pathway is frequently observed in  
 cancer (9, 10). Moreover, multiple mutations  
 of epigenetic regulators are described in ma-  
 lignancies and both loss-of-function muta-  
 tions in many histone methyltransferases  
 and KDMs (11, 12), potentially mimicking hy-  
 poxia are observed. Moreover, therapeutics  
 that target chromatin modifiers are being  
 evaluated for treating certain cancers in clin-  
 ical trials (13). We speculate that the direct  
 oxygen-sensing KDM pathways are also ab-  
 errant in malignancy and other pathological  
 states, such as cardiovascular disease. Tar-  
 geting the HIF pathway with small molecule  
 inhibitors is currently being explored in can-  
 cer therapy and other non-malignant condi-  
 tions such as renal disease. These studies  
 suggest that oxygen sensing in KDMs might  
 also be specifically therapeutically targeted,  
 if the mechanistic basis for this sensing is de-  
 termined.

Interestingly, Batie *et al.* and Chakraborty  
*et al.* speculate that based on phylogenetic  
 sequence conservation the direct oxygen-  
 sensing KDM pathways may evolutionarily  
 pre-date the HIF pathway. It is likely that

1 both pathways have more recently co-  
2 evolved and function in a coordinated and  
3 temporally defined manner to modulate the  
4 cellular response to low oxygen tensions.  
5 This is suggested by the immediate hyper-  
6 methylation of H3K4 at the promoters of  
7 HIF-target genes. However, further delineat-  
8 ing the interaction between these pathways  
9 and how this might be modulated will be im-  
10 portant to understand. Together, these ob-  
11 servations have profound implications for  
12 our understanding of how microenviron-  
13 mental changes, and specifically oxygen con-  
14 centrations, might affect both physiological  
15 and pathological cell fate decisions and phe-  
16 notypes through direct effects on chromatin  
17 structure.

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#### 44 Hypoxia-mediated alterations in transcription via 45 direct (HDM-mediated) and indirect (HIF- 46 mediated) mechanisms

47 In hypoxic conditions, prolyl hydroxylases (PHDs)  
48 are inactivated, allowing HIF-1 $\alpha$  to dimerize with  
49 ARNT, translocate to the nucleus and activate HIF  
50 target genes. Chakraborty *et al.* and Batie *et al.*  
51 find that the lysine demethylases, KDM6A and  
52 KDM5A, are also direct oxygen sensors that are in-  
53 activated during hypoxia. This allows both imme-  
54 diate hypermethylation of H3K27 (KDM6A target)  
55 and gene repression and hypermethylation of  
56 H3K4 (KDM5A target) and gene activation. Under  
57 normoxic conditions, the HIF-1 $\alpha$  subunit is tar-  
58 geted for destruction through hydroxylation by  
59 PHDs. This allows ubiquitylation by the Von-Hippel  
Lindau tumor suppressor protein (VHL), and sub-  
sequent destruction in the proteasome.