

# 1 Minutes matter: brief hatching asynchrony adversely affects late- 2 hatched hihi nestlings, but not life beyond the nest

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9

## 10 Abstract

11 Size hierarchies are often seen when nestlings hatch asynchronously over a period of  
12 days. Shorter hatch periods are common across passerines however, and while these  
13 may also give rise to asymmetries, their effects are rarely considered. Regardless of  
14 hatch period, the long-term consequences for later-hatched nestlings that survive to  
15 fledge is unknown for wild birds. Here we explored the timing of hatch order in a free-  
16 living population of hihi nestlings (*Notiomystis cincta*) and followed any effects in and  
17 out of the nest. We found that while hatching time from first to last-hatched nestlings  
18 was often less than 24 hours, last-hatched individuals grew more slowly and were  
19 lighter and smaller at fledging than older siblings. Last-hatched nestlings were also less  
20 likely to fledge. These effects were greater in larger broods. Adult body size is  
21 correlated with fledging size in hihi; however, we found no evidence that hatch order  
22 affected longevity post-fledging, or lifetime reproductive success. We then explored if  
23 carotenoid availability might buffer these stressful rearing conditions (through food  
24 supplementation of parents) but found no evidence that increased access to carotenoids  
25 for mothers and/or growing nestlings influenced incubation schedules, or the effects of  
26 hatching late. Together these results suggest that while even a very short hatch period

27 can influence adult phenotype, hatching asynchrony is not maladaptive for hihi: when  
28 last-hatched nestlings survive to fledge they can contribute as much to their mother's  
29 fitness as first-hatched siblings.

30 **Key words:** brood hierarchies, carotenoids, hatching asynchrony, maternal effects,  
31 *Notiomystis cincta*

## 32 **Introduction**

33 Sequential hatching of avian young driven by early onset of incubation (hatching  
34 asynchrony) often establishes size hierarchies within broods (Stokland & Amundsen,  
35 1988; Wiklund, 1985). This can leave younger, smaller chicks at a disadvantage from  
36 sibling competition (Mock & Parker, 1997) if parents feed larger offspring preferentially  
37 (Rodriguez-Girones et al. 2002), but hatching asynchrony can also be an adaptive  
38 strategy if it allows a mother to maximize the overall success of her brood (for example,  
39 through brood reduction to match unpredictable environments (Magrath, 1990;  
40 Stenning, 1996). While the effects of hatching asynchrony on life within the nest are  
41 well-known, the long-term consequences of hatch order on lifespan or reproductive  
42 success are much less understood (Mainwaring, Blount, & Hartley, 2012), particularly  
43 for wild bird populations.

44 It is possible that the availability of specific nutrients may influence the potential  
45 for later-hatched nestlings to catch up with their elder siblings. Carotenoids, a class of  
46 antioxidants synthesised by plants and acquired by birds through their diet, may act as  
47 a buffer to natural stressors due to their ability to boost the immune system (Berthouly,  
48 Cassier, & Richner, 2008). In birds, carotenoids mitigate the effects of stress  
49 experimentally induced by increased sibling competition (Berthouly et al., 2008), and

50 infestation with ectoparasites (Ewen et al., 2009). In both cases, when carotenoid  
51 concentration in eggs was increased by supplementing mothers, nestlings placed at a  
52 disadvantage from an increased brood size (Berthouly et al., 2008), or parasite  
53 infestation (Ewen et al., 2009), fared as well as nestlings from unmanipulated broods  
54 (unsupplemented nestlings fared the worst). As hatching later than siblings can  
55 increase the physiological stress levels of nestlings (Costantini et al., 2006; de Boer,  
56 Eens, Fransen, & Müller, 2015; Eraud, Trouvé, Dano, Chastel, & Faivre, 2008),  
57 carotenoids available in the nestling's diet could therefore counteract the negative  
58 effects of hatching late in a brood hierarchy.

59         Here we investigate whether size hierarchies observed in the hihi (*Notiomystis*  
60 *cincta*) are caused by sequential hatching driven by maternal incubation behavior, and  
61 whether hatching later than siblings has long-term fitness consequences. We expect that  
62 early onset of incubation should result in more asynchronously hatching clutches, and  
63 that last-hatched nestlings should be smaller and grow more slowly than early-hatched  
64 siblings. Body mass at fledging improves a hihi's chances of surviving its first year (M.  
65 Low & Part, 2009). Therefore, if the brood hierarchy order persists throughout the  
66 nestling period, it is likely to have long-term consequences for lifespan and  
67 reproductive success and not just survival to fledging as is most commonly investigated  
68 due to the logistical challenges of tracking individuals throughout their lives.

69         In addition, we supplemented adult hihi with carotenoids during throughout  
70 breeding (including incubation) to test if increased availability of carotenoids to  
71 nestlings (either in eggs or also during nestling provisioning) compensates for any  
72 negative effects of late hatching. Previous research shows that carotenoids are an  
73 important dietary component for hihi nestlings, but that their effects appear to be

74 compensatory: only when challenged by poor rearing conditions (ectoparasites) do  
75 nestlings hatched from carotenoid-rich eggs grow better (Ewen et al., 2009). This may  
76 be particularly important if brood hierarchies arise through factors other than  
77 differences in maternally-driven incubation behaviour, for example, through limitation  
78 of other key egg components (Nager, Monaghan, & Houston, 2000). We therefore expect  
79 increased carotenoid availability to be most effective for nestlings in stressful  
80 conditions: those late in the hatching order.

81

## 82 **Methods**

### 83 **Supplementation experiment and data collection**

84 We studied a breeding population of hihi, a bird endemic to New Zealand and listed by  
85 the IUCN as Vulnerable (BirdLife International, 2013), on Tiritiri Matangi Island, a 220  
86 hectare island ~25 km north of Auckland. This population is part of a conservation  
87 project, with supplementary food (sugar water) and nesting boxes provided across the  
88 island. Re-sighting surveys were conducted each year in February (post-breeding) and  
89 in September (pre-breeding) between February 2005 and February 2015, providing 21  
90 capture occasions (for further details see Thorogood *et al.* 2013). Hihi breed between  
91 October and March, producing clutches of, on average, 4 eggs (this dataset,  $4.21 \pm 0.69$   
92 eggs), which hatch into broods of, on average, 3 nestlings (this dataset,  $3.23 \pm 1.05$   
93 nestlings). Previous work has shown that carotenoid supplementation does not  
94 significantly influence these parameters (Ewen, Thorogood, Karadas, & Cassey, 2008).  
95 All breeding attempts were closely monitored so the parentage of all offspring that  
96 reach fledging age (30 days, blood-sampled at 21 days) could be determined via

97 genotyping and assignment using Colony 2.0 software (Brekke, Ewen, Clucas, & Santure,  
98 2015).

99         Our data were collected during the 2004/05 and 2005/06 breeding seasons  
100 (hereafter Seasons 1 and 2) as part of carotenoid supplementation experiments  
101 designed to investigate effects on maternal egg investment (Ewen, Surai, et al., 2006),  
102 parental provisioning of nestlings (Ewen et al., 2008), and effects of ectoparasites on  
103 nestling health (Ewen et al., 2009). Our supplementation regime differed between years,  
104 with females in 'Season 1' receiving supplementation from nest building to offspring  
105 fledging, whereas in 'Season 2', supplementation was stopped after completion of egg  
106 laying (Table 1). This allowed us to test any effect of carotenoid supplementation at  
107 different times during development. Breeding pairs were allocated to treatment or  
108 control groups once they settled on a nest site. Control pairs were provided with a  
109 supplementary food source (sugar water), while treatment pairs were provided with  
110 sugar water supplemented with carotenoids (lutein and zeaxanthin) at a concentration  
111 of 100 µg/ml (Ewen et al., 2008), in both cases within 10 m of the nest box. Hihi defend  
112 food resources in their territories, and food was provided *ad libitum*, so feeders were  
113 rarely used by either conspecifics or other nectarivorous species, nor did use of feeders  
114 differ among treatment groups (Ewen et al. 2008) Furthermore, supplementation of  
115 female hihi with carotenoids during laying has shown to positively influence yolk  
116 carotenoid concentration (Ewen, Thorogood, Karadas, Pappas, & Surai, 2006), and  
117 supplementation of parents during nestling rearing increases circulating plasma  
118 carotenoid levels of both nestlings and parents (Thorogood et al. 2008, Thorogood et al  
119 2011).

120           Hihi females lay one egg per day, and incubate for 14 days after clutch  
121 completion, but whether they begin incubation during laying (and therefore, whether  
122 they may adaptively adjust hatching synchrony) is unknown. Incubation behavior was  
123 recorded in Season 2 using temperature loggers (Thermochron iButtons® DS1921G,  
124 Maxim), which were inserted into the nest cup and secured around nest material either  
125 the day before or the day the first egg was laid. A second logger was attached to the  
126 inside of the nest box (20 cm from the nest cup) to record ambient temperature. Both  
127 loggers recorded temperature (°C) in 2-minute intervals for up to 7 days. We counted  
128 the onset of incubation from the first night when the nest cup - but not box -  
129 temperature rose above 25°C for more than one hour (Cooper & Mills, 2005; Wang &  
130 Beissinger, 2009). This is 'physiological zero temperature', when embryonic growth  
131 begins (Wilson, 1990). Our measurement therefore recorded incubation effort in nights  
132 until clutch completion.

133           After 13 days of incubation (day final egg laid = day 1) nests were monitored to  
134 determine hatching order of each egg. Nests were visited in the late afternoon (after  
135 4pm) on day 13, and then at no more than two-hourly intervals from dawn until dusk  
136 on day 14. If any eggs remained intact by dusk on day 15 we checked only once more on  
137 day 16 before removing eggs for other analyses (Season 1,  $N = 39$ ; Season 2,  $N = 78$ ;  
138 Thorogood & Ewen 2006): a small proportion of each clutch often fails to hatch due to  
139 unviability (Brekke, Bennett, Wang, Pettorelli, & Ewen, 2010; Thorogood & Ewen,  
140 2006). From these checks the time between hatching events was recorded accurate to  
141 within 120 min.

142           Once nestlings hatched (day 0) they were marked on the tarsus using a  
143 permanent non-toxic marker pen. This identifier was refreshed every two days until

144 nestlings were 21 days old, when birds were given numbered metal rings and plastic  
145 colour ring combinations as part of standard management practice to enable  
146 identification. Nestlings were weighed and the length of their tarsi measured (with  
147 Vernier calipers) throughout the nestling period (Table 1). Measurements were taken  
148 every 3 days until day 24, after which measurements ceased to avoid causing the  
149 premature fledging of offspring. Tarsus length does not change after 21 days so  
150 measurements at day 24 indicate adult tarsus length (Low, 2006).

151         For our analyses of the effects of hatch order (and mitigating effects of  
152 carotenoids), we restricted our dataset to first clutches ( $N = 82$  clutches); hihi  
153 sometimes produce second clutches in a season, but these often fail completely  
154 (Thorogood, Ewen, & Kilner, 2011). We further restricted our dataset to clutches that  
155 hatched at least two nestlings as by definition, broods of 1 cannot hatch  
156 asynchronously. Of these clutches, full data on mass and size were available for 167  
157 nestlings from 64 nests, and full data on growth rate were available for 96 nestlings  
158 from 38 nests. Reported brood sizes represent brood size at hatching.

159

## 160 **Statistical analyses**

### 161 *Onset of incubation and hatching spread*

162 To investigate if hatching asynchrony is influenced by females' incubation behaviour,  
163 and to rule out the possibility that incubation behaviour is a consequence of clutch size  
164 (for example, if females always begin incubating after laying a certain number of eggs),  
165 we tested for relationships between onset of incubation and hatching spread, and the  
166 onset of incubation and clutch size, using Pearson's correlation tests. The sample sizes

167 for these analyses were limited by how many nests we were able to collect data on  
168 incubation behaviour, and hatching spread.

169 To include broods of a range of sizes, we standardized hatch order as first,  
170 second, and last-hatched. Thus, in broods of four and five, the “last-hatched” nestling  
171 was either the fourth or fifth respectively. This selection allows us to compare the  
172 effects of hatching late across brood sizes, as in Badyaev, Hill, & Beck (2003). In broods  
173 of two nestlings, nestlings were coded as first and second-hatched in order to be most  
174 comparable with first and second-hatched offspring from other brood sizes (i.e. there is  
175 at least one nestling between every first and last-hatched nestling). If two nestlings  
176 hatched simultaneously (within same 120 min period between nest checks) they were  
177 given the same (earlier) order. Our dataset included 68 first-hatched nestlings, 49  
178 second-hatched nestlings, and 50 last-hatched nestlings. Hatch order was specified as  
179 an ordinal categorical variable in all models – this allowed us to retain information  
180 about order, without assuming linearity in the time lag between orders. All models  
181 automatically tested for linear and quadratic relationships – all relationships reported  
182 are linear unless stated otherwise, as no significant quadratic relationships were found.

183

#### 184 *Effects of hatch order and carotenoid supplementation on growth*

185 We fitted a standard logistic growth model (Ricklefs, 1968) using the SSlogis function of  
186 the “stats” package in R (R Core Team, 2013) to estimate each individual’s asymptotic  
187 mass (g) and tarsus length (mm), and their growth rates ( $k$ ). Logistic growth models fit  
188 postnatal growth data well in passerines (Starck & Ricklefs, 1998), and have been used  
189 to describe growth before in hihi (Ewen et al. 2009). Nestlings that died before fledging



190 were excluded. Models could not accurately predict growth rates for data from Season 1  
191 due to a lack of measurements before day 10; therefore growth rate analyses included  
192 only nestlings from Season 2 (asymptote analyses included nestlings from both seasons,  
193 as a lack of measurements before day 10 did not affect model asymptote).

194 To determine the interactive effects of carotenoids, sex, and hatch order on  
195 nestling growth (in terms of asymptotic mass and tarsus length, and rate of growth in  
196 both), we set these parameters as dependent variables in linear mixed effects models  
197 constructed using the lme4 package in R (Bates et al. 2008). We included a three-way  
198 interaction term: treatment \* hatch order \* sex. Brood size has been shown to be an  
199 important determinant in the success of late-hatched red-winged blackbird nestlings  
200 (Forbes, Thornton, Glassey, Forbes, & Buckley, 1997), therefore we included a separate  
201 interaction term, hatch order \* brood size. We also included maternal age (years) and  
202 relative time in the season (days since hatching of the first clutch produced in that  
203 season, a proxy for how early or late each nest is relative to other nests in the  
204 population) as covariates, as these have been shown to influence other factors in hihi,  
205 such as hatching failure and nestling survival (M. Low & Part, 2009). Brood identity was  
206 included in all models as a random term to control for multiple individuals from the  
207 same nest. Where data from both Season 1 and Season 2 were used in analyses,  
208 maternal identity and season were also included as random terms to control for  
209 repeated measures, and potential differences between seasons not accounted for by  
210 differences in treatment regime respectively. Any interactions or terms that did not  
211 contribute significantly to model fit were removed using stepwise deletion, by removing  
212 the term of interest from the model, and comparing its fit to the data using chi-squared  
213 tests.

214 To determine the effects of hatch order on nestling survival to fledging, we fit  
215 fledging success of nestlings (0/1) in binomial linear mixed effects models, again using  
216 the lme4 function in R (Bates et al. 2008), and fitting the same interactions, random  
217 terms, and covariates as described above. Sex was not significant in an initial model set  
218 ( $\chi^2_1 = 2.73, P = 0.1$ ), so we repeated the analysis with an expanded dataset in which we  
219 were able to include nestlings that died before being sexed. We report the results of the  
220 second, expanded, analysis. Data were available for 242 nestlings from 76 nests.

221 For all analyses of hatch order and carotenoid treatment effects, we first looked  
222 for differences among our treatment nests between seasons to determine if the timing  
223 of carotenoid supplementation (Table 1) influenced brood size hierarchies and their  
224 effects. If timing of carotenoid availability had no influence, we *a priori* decided to  
225 combine the different supplementation regimes as an overall carotenoid treatment  
226 factor (treatment variable: control/supplementation). If timing of carotenoid  
227 availability (full supplementation in Season 1/laying-only supplementation only in  
228 Season 2) did have an effect in a model, however, we then included this as a 3-level  
229 factor (control/full supplementation/laying-only supplementation). During Season 2,  
230 some nests included in this dataset ( $N = 46$ ) were also used in a mite-removal  
231 experiment (for methodology see Ewen et al., 2009). We therefore also tested for any  
232 interactive effects of hatch order, mite treatment, and carotenoid treatment to  
233 determine whether mite treatment had any effect on the relationships we were  
234 interested in here. An effect of mite treatment was only found in the case of survival to  
235 fledging, so it is only reported for that analysis.

236

237 ***Long term fitness effects of brood hierarchy position***

238 We estimated the effects of hatch order on post-fledging survival using a Cormack-Jolly-  
239 Seber survival analysis in Program MARK (White & Burnham, 1999). Candidate models  
240 could be time-dependent ('time'), sex-dependent ('sex'), and hatch order-dependent  
241 ('hatch order') with three levels differentiating first hatched, second hatched, and last  
242 hatched individuals. Most models were constructed with two age classes with  
243 individuals entering the population as juveniles in each February and transitioning into  
244 an adult age class the subsequent September ('age'). A global model estimated survival  
245 ( $\varphi$ ) according to age, sex, hatch order, and time, while also including time dependence  
246 on detection probability ( $p$ ). Alternative models were then derived by progressive  
247 removal of factors thought least likely to be important based on *a priori* predictions. All  
248 potential alternative models were tested and compared using QAICc (Cooch & White,  
249 2008). Data were restricted to individuals for whom complete information was  
250 available on sex and hatch order ( $N = 140$ ). Global model fit to the data was assessed  
251 using the median c-hat procedure showing a small adjustment was required (c-hat =  
252 1.1).

253 To test whether position in the hatching order influenced individual lifetime  
254 reproductive success, we restricted our dataset to individuals who survived to breeding  
255 age (both sexes can breed in their first year, Ewen et al. 2011), and for which we know  
256 their total reproductive output (i.e. excluding individuals that are still alive and  
257 reproductively active) (30 females, 28 males). For each individual, we calculated the  
258 total number of fledged offspring per year. Social partner identity strongly predicts  
259 reproductive success in female, but not male, hihi (Brekke, Cassey, Ariani, & Ewen,  
260 2013; Brekke et al., 2015). We therefore modeled hatch order effects on lifetime

261 reproductive success in males and females separately and accounted for social partner  
262 identity in our analysis of females. Total number of fledged offspring produced per year  
263 was set as the dependent variable in a mixed model, with hatch order as an independent  
264 variable. We included age (years) as a polynomial covariate, as both male and female  
265 reproductive output is best represented by a bell-shaped curve in this species (M. Low,  
266 Pärt, & Forslund, 2007). Individual identity, nest of origin, mother identity, and season  
267 were set as random terms.

## 268 **Ethical note**

269 Ethical approval for supplementing carotenoids was granted by the Zoological Society  
270 of London Ethics Committee (UK). Permissions to conduct research on Department of  
271 Conservation Estate and to collect samples as detailed above were also granted from the  
272 Auckland Conservancy of the Department of Conservation. These protocols were  
273 derived from standard monitoring protocols used for management of hihi by the  
274 Department of Conservation.

275

## 276 **Results**

277 Our dataset included five nestlings from broods of two, 59 nestlings from broods of  
278 three, 67 nestlings from broods of four, and 36 nestlings from broods of five, but not all  
279 data could be collected from every nestling. Means are reported with standard  
280 deviations, and sample size for each analysis is given.

281

## 282 *Onset of incubation and hatching spread*

283 Across all nests for which we obtained hatch order data ( $N = 82$ ), there was large  
284 variation in the time it took broods to hatch (hatching spread: time in minutes from first  
285 egg hatching to last egg hatching). This ranged from no delay (all eggs hatched within 2  
286 hours between visits) to 28 hours and 10 minutes, with a mean of  $\sim 10$  hours ( $597.7 \pm$   
287  $473.9$  minutes). Larger broods took longer to hatch (Pearson's  $r = 0.36$ ,  $P < 0.01$ ,  $N =$   
288  $81$ ).

289 In Season 2, we recorded incubation effort from 24 nests (14 were from the  
290 carotenoid treated group). Variation among females was great, with some females  
291 commencing incubation only once their clutch was complete ( $N = 4$ ), but others  
292 beginning 1 ( $N = 10$ ), 2 ( $N = 9$ ), or 3 nights ( $N = 1$ ) before their last egg was laid.  
293 However, this was not explained by clutch size (Pearson's  $r: 0.26$ ,  $P = 0.22$ ) or  
294 carotenoid supplementation (Table 2). For a subset of these nests ( $N = 10$ ), we could  
295 correlate onset of incubation with hatching order: more nights of incubation effort  
296 showed a non-significant trend toward a longer spread in hatching from the first to the  
297 last chick (Pearson's  $r: 0.62$ ,  $P = 0.06$ ). Therefore, it is likely that the variation in  
298 hatching spread we detected in our dataset was a consequence of variation in the onset  
299 of mothers' incubation behaviour, but not variation in carotenoids deposited in the eggs.

## 300 *Effects of hatch order and carotenoid supplementation on growth*

301  
302 Hatching later had a large effect on the growth and size of nestlings (Table 3). Hatching  
303 late in the hatching sequence resulted in nestlings that were significantly lighter  
304 (asymptotic mass, Fig 1a) and smaller (asymptotic tarsus length, Fig 1b) than older  
305 siblings at fledging, and grew more slowly (both in terms of mass, and tarsus length)  
306 (Table 3). These effects were particularly pronounced in larger broods, except in the

307 case of tarsus length (hatch order \* brood size  $\chi^2_2 = 4.83, P = 0.09$ ). Male nestlings were  
308 heavier and larger at fledging than female siblings, and grew faster in terms of tarsus  
309 length (Table 3). There was no difference between the sexes in the rate at which they  
310 gained mass (sex  $\chi^2_1 = 0.34, P = 0.24$ ).

311         Regardless of when carotenoids were supplemented, there was no evidence that  
312 any of these hatch order effects were influenced by carotenoid supplementation (Table  
313 3). Nor did carotenoid supplementation independently influence the final mass, or rate  
314 of mass gain, of nestlings, or affect the size of brood hierarchies (all results in Tables 2  
315 and 3). Access to carotenoids did influence tarsus growth rate, however: carotenoid-  
316 treated nestlings of all hatch orders grew more quickly than nestlings from control  
317 nests (Table 3).

### 318 ***Long term fitness effects of brood hierarchy position***

319 Fifty-three out of 242 nestlings died before fledging. Last-hatched nestlings were more  
320 likely to die before fledging than earlier-hatched siblings, and this effect was greater in  
321 larger broods (Table 3). Although not the focus of our study, we detected that mite  
322 treatment also had an additive effect on nestling survival: nestlings that did not have  
323 mites removed were more likely to die in the nest.

324         Once nestlings fledged, however, hatch order no longer influenced survival; the  
325 best models ( $\Delta \text{QAICc} < 2$  of top model) contained only age and sex (Table 4), with older  
326 birds surviving better than first-years, and females living longer than males. We also  
327 found no influence of hatch order on the number of offspring produced by our two  
328 cohorts throughout their lifetime, either in interaction with sex ( $\chi^2_2 = 4.35, P = 0.11$ ), or  
329 independently ( $\chi^2_2 = 0.72, P = 0.7$ ). Hatch order (and carotenoid supplementation)  
330 effects are summarized in Table 5.

331

332 **Discussion**

333 Our study demonstrates that hatching even a short time after siblings can have strong  
334 effects on offspring: later hatched nestlings grew more slowly, and remained smaller at  
335 fledging according to growth models. Last-hatched hihi nestlings were also less likely to  
336 survive to fledge, and these effects of hatching last were strongest in larger broods. As  
337 adult tarsus length does not change after fledging in hihi (Low 2006), the hatch order  
338 effects we detected on body size (as predicted by growth models) are likely to persist  
339 throughout adult life. However, we found no lasting consequences of hatch order on  
340 post-fledging longevity or reproductive success, suggesting that if they survive to fledge,  
341 later hatched offspring contribute as much to parents' fitness as their earlier hatched  
342 (and larger) nest-mates. As the period of time over which eggs hatched was related to  
343 maternal incubation behaviour, together these results are consistent with hatching  
344 asynchrony being adaptive for hihi mothers.

345         During the nestling period at least, hatching later than nest-mates appears to be  
346 more detrimental for young hihi than in other species with comparable or even longer  
347 hatching periods. Tree swallows hatch over a similar period to hihi (28 h on average,  
348 (Clotfelter, Whittingham, & Dunn, 2000), but unlike our study where effects persisted  
349 until fledging, for tree swallows the effects of hatch order on nestling traits have been  
350 shown to disappear by day 12 (Clotfelter et al., 2000) (but see Zach 1982). Even in  
351 species with much longer hatching periods, for example, jackdaws (which hatch over a  
352 number of days: Wingfield Gibbons, 1987), hatch order effects have been shown to  
353 disappear before fledging (Arnold & Griffiths, 2003).

354           Why are brood hierarchies so pronounced in hihi, despite their relatively short  
355 hatching spread? Hihi nestlings may be particularly sensitive to hatch order effects  
356 because of their long nestling period (30 days) relative to other passerines (Roff, Remes,  
357 & Martin, 2005): if earlier hatched nestlings gain a head start, this may exacerbate hatch  
358 order effects over this long time period (Price & Ydenberg, 1995), especially if early  
359 asymmetries in sibling competition persist (Glasse & Forbes, 2002). In addition, food  
360 shortages early in the post-hatching stage can have consequences for growth later on,  
361 even if parental provisioning later increases (Killpack & Karasov, 2012; Lack, 1954).  
362 However, lasting hatch order effects on tarsus size have been found in house finches  
363 (Badyaev et al., 2002), which have a nestling period of around 16 days, so this is not an  
364 entirely satisfactory explanation. More work is needed to investigate whether, for  
365 example, nestling size or sex influences nestling begging, sibling competition, and/or  
366 parental provisioning behaviour, and whether these factors may exacerbate hatch order  
367 effects in this species.

368           If brood hierarchies create stressful growth environments for later-hatched  
369 nestlings, why did carotenoids not mitigate these effects? We know that our  
370 supplementation changed the phenotype of nestlings, as previous analyses of subsets of  
371 these data indicate differences in nestlings' body condition (Ewen et al. 2009) and  
372 tarsus length (Ewen et al. 2008) at fledging, and here we find that carotenoid  
373 supplementation leads to faster growth (at least for tarsi). Other work with hihi has also  
374 shown that increased access to carotenoids influences nestling begging (Ewen et al.  
375 2008, Thorogood et al. 2008), but that this effect is only present when parents do not  
376 provide the carotenoids themselves to their nestlings (Thorogood et al. 2011). Given



377 our lack of effect here, this suggests that parents do not favour feeding later-hatched  
378 nestlings, although this requires further experiments to determine.

379         Alternatively, perhaps we detected no effect of carotenoids because they actually  
380 exacerbated effects of hatch order, instead of mitigating them as we predicted.  
381 Deposition of carotenoids in egg yolk often decreases down the laying order (for  
382 example, lesser black-backed gulls (Blount et al., 2002; Royle, Surai, McCartney, &  
383 Speake, 1999), and barn swallows (Saino et al., 2002) so it may be that mothers skewed  
384 carotenoids to earlier, “more valuable” offspring (Groothuis, Müller, Von Engelhardt,  
385 Carere, & Eising, 2005; Williams, 2012). As we detected no increase in hatch order  
386 effects in our carotenoid-treated group, it seems likely that if carotenoids are implicated  
387 in this relationship mothers retain any extra for themselves and do not boost the yolks  
388 of eggs intended to hatch last. Unfortunately the conservation status of hihi renders it  
389 impossible to destructively sample eggs, or manipulate incubation schedules directly so  
390 we are unable to test this possibility further.

391         Despite strong effects of hatch order on nestling size and mass at fledging, we  
392 found no lasting consequences of this difference on post-fledging longevity or  
393 reproductive success. This is surprising, given that previous analyses have shown that  
394 body mass close to fledging correlates well with survival of females during their first  
395 year (Low & Part, 2009). One possible explanation is that all nestlings in our dataset  
396 were in relatively good condition when they fledged. During our experiment, both our  
397 carotenoid-treated and control nests were supplied with sugar water close to the nest.  
398 Furthermore, the population is provided with sugar water *ad libitum* throughout the  
399 year (Thorogood, Armstrong, Low, Brekke, & Ewen, 2013). Having easy access to food  
400 throughout their lives (beginning with parental provisioning) may have enabled

401 smaller, later hatched individuals to persist in the population regardless of hatch order  
402 or carotenoid availability. It is also unlikely that later-hatched individuals trade  
403 longevity against reproduction, as we detected no effect of hatch order on annual  
404 reproductive success.

405         Finally, we expected male hihi nestlings to be more sensitive to hatching order  
406 effects given that male hihi embryos are more sensitive to the negative effects of  
407 inbreeding (Brekke et al., 2010). Furthermore, male hihi nestlings are larger than  
408 females (Ewen et al., 2009; this study), and sex-specific differences in sensitivity to poor  
409 or harsh environments are common across vertebrates where sexes are dimorphic and  
410 one is more costly to produce or maintain (Anderson *et al.*, 1993; Lindström, 1999).  
411 Hatching order has strong sex-specific effects in certain house finch populations  
412 (Badyaev *et al.*, 2002). Likewise, male house wrens were more negatively affected by  
413 experimentally manipulated hatch orders than their female siblings (Bowers,  
414 Thompson, & Sakaluk, 2015). However, size differences between male and female hihi  
415 did not appear to result in increased male sensitivity to hatch order effects in our study.  
416 Further exploration of whether parents preferentially feed one sex under certain  
417 circumstances, as is seen in eastern bluebirds (Ligon & Hill, 2010) and Arabian babblers  
418 (Ridley & Huyvaert, 2007) would be informative, as such a bias could mask the  
419 sensitivity of males. Alternatively, any male sensitivity may be matched by  
420 disadvantages of hatching late for females, given their already smaller size (Oddie,  
421 2000).

422         The average period over which broods hatched in our dataset was less than 24  
423 hours; so, by the accepted definition, hihi hatch synchronously (Stoleson & Beissinger,  
424 1995). Nevertheless, by investigating hatch order effects in an apparently

425 synchronously hatching species, we have detected implications for hihi in early life that  
426 may have lasting consequences for the adult phenotype. The increased likelihood of  
427 death before fledging for last-hatched offspring is significant, however, we found that  
428 later hatched nestlings that did fledge lived as long and produced as many offspring as  
429 early-hatched nestlings. This suggests that hatching asynchrony is not maladaptive for  
430 hihi; negative effects of hatching late are confined to the nestling period, after which  
431 surviving offspring are equally likely to contribute to their mothers' fitness. Our study  
432 therefore provides valuable insight into the implications and adaptive potential of  
433 hatching asynchrony by broadening the scope under which it has previously been  
434 studied. Closer investigation of hatch order effects in other species with apparent  
435 synchronous hatching may lead to novel insights into what determines when and why  
436 brood size hierarchies arise.

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#### 438 **Data accessibility**

439 Data will be available on Dryad upon acceptance: DOI XXXXXXXX

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631 Table 1. Details of between-year differences in our carotenoid supplementation regime..

Season	Supplementation regime				Incubation monitored	Nestlings weighed from:
	Nest-building	Egg-laying	Incubation	Nestling rearing		
2004/2005 (1)	X	X	X	X	No	Day 10
2005/2006 (2)	X	X			Yes	Day 3

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646 Table 2. Descriptive statistics on all nests from both seasons ( $N = 119$ ).

	Group		Test statistic	<i>P</i>
	Untreated	Carotenoid supplemented		
Total clutches	62	57		
Clutch size (mean $\pm$ SD)	4.26 $\pm$ 0.68	4.16 $\pm$ 0.70	$T = 0.79$	0.43
Average egg mass (per egg mean g $\pm$ SD)	2.95 $\pm$ 0.19 ( $N=104$ )	2.92 $\pm$ 0.26 ( $N=71$ )	$T = 0.97$	0.34
Nights of incubation prior to laying of final egg (mean nights $\pm$ SD)	1.2 $\pm$ 0.92 ( $N = 10$ )	1.4 $\pm$ 0.74 ( $N = 14$ )	$\chi^2 = 0.43$	0.51
Hatch failures	23% (62/264)	22% (53/237)	$\chi^2 = 0.09$	0.76
Average hatching spread (mean mins $\pm$ SD)	571.7 $\pm$ 483.0 ( $N=91$ )	593.06 $\pm$ 477.18 ( $N=90$ )	$T = -0.21$	0.84
Brood size (mean $\pm$ SD)	3.24 $\pm$ 1.10	3.21 $\pm$ 1.01	$T = 0.16$	0.87
Range of nestling masses at fledging (mean coefficient of variation)	15.61	16.58	$\chi^2 = 0.003$	0.95

647 Where appropriate the difference between untreated and supplemented groups are tested statistically;  
 648 tests carried out are indicated with test statistic.  
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668 Table 3. Results of GLMM investigating hatch order and carotenoid treatment effects on male and female  
669 chick size and growth (mass in grams, tarsus length in mm).

		Est.	SE	Z	P
<b>ASYMPTOTIC MASS AND SIZE – 167 nestlings from 64 nests</b>					
1. asymptotic mass	<i>Intercept</i>	36.55	0.59	62.26	
	Hatch order * brood size	-1.92	0.49	-3.88	<0.001
	Sex				<0.000
	Female	0.00			1
Male	6.36	0.61	10.38		
2. asymptotic tarsus length	<i>Intercept</i>	31.34	0.12	253.75	
	Hatch order	-0.43	0.13	-3.38	<0.01
	Sex				<0.001
	Female	0.00			
Male	1.77	0.15	11.62		
<b>GROWTH RATE – 96 nestlings from 38 nests</b>					
3. mass growth rate (N = 96)	<i>Intercept</i>	0.39	0.03	14.45	
	Hatch order * brood size	-0.03	0.01	-3.70	<0.001
4. tarsus growth rate (N = 96)	<i>Intercept</i>	0.33	0.02	19.16	
	Treatment				<0.001
	Control	0.00			
	Laying supplementation	0.01	0.006	2.16	
	Sex				<0.001
Female					
Male	-0.02	0.004	-4.02		
	Hatch order * brood size	-0.01	0.005	-2.74	<0.01
<b>SURVIVAL TO FLEDGING – 242 nestlings from 76 nests</b>					
5. survival to fledging	<i>Intercept</i>	2.22	1.24	1.79	
	Hatch order * Brood size	-2.00	0.55	-3.66	<0.001
	Mite treatment				<0.001
	Mites removed	0.00			
Mites present	-2.66	0.60	-4.42		

670 <sup>1</sup> Removed from models: hatch order\*sex\*treatment (control/laying supp/full supp)  $\chi^2_4 = 6.73$ ,  $P = 0.15$ ; sex\*treatment  
671 (control/laying supp/full supp)  $\chi^2_2 = 0.14$ ,  $P = 0.93$ ; hatch order\*sex  $\chi^2_4 = 0.67$ ,  $P = 0.71$ ; hatch order\* treatment (control/laying  
672 supp/full supp)  $\chi^2_4 = 1.21$ ,  $P = 0.88$ ; maternal age  $\chi^2_1 = 0.06$ ,  $P = 0.81$ ; hatch order\*brood size  $\chi^2_2 = 5.12$ ,  $P = 0.08$ ; brood size  $\chi^2_1 =$   
673  $1.62$ ,  $P = 0.20$ ; date  $\chi^2_1 = 2.85$ ,  $P = 0.09$ ; treatment (control/laying supp/full supp)  $\chi^2_2 = 3.64$ ,  $P = 0.16$ .

674 <sup>2</sup> Removed from models: hatch order\*sex\*treatment (control/laying supp)  $\chi^2_2 = 1.61$ ,  $P = 0.45$ ; sex\*treatment  $\chi^2_2 = 0.61$ ,  $P = 0.43$ ;  
675 hatch order\*treatment (control/laying supp)  $\chi^2_2 = 0.94$ ,  $P = 0.63$ ; hatch order\* sex  $\chi^2_2 = 4.13$ ,  $P = 0.13$ ; maternal age  $\chi^2_1 = 0.01$ ,  $P =$   
676  $0.93$ ; treatment (control/laying supp)  $\chi^2_1 = 0.17$ ,  $P = 0.68$ ; sex  $\chi^2_1 = 0.34$ ,  $P = 0.24$ ; date  $\chi^2_1 = 3.38$ ,  $P = 0.07$ ..

677 <sup>3</sup> Removed from models: hatch order\*sex\*treatment (control/laying supp)  $\chi^2_2 = 2.03$ ,  $P = 0.36$ ; hatch order\*sex  $\chi^2_2 = 0.14$ ,  $P = 0.93$ ;  
678 hatch order\*treatment (control/laying supp)  $\chi^2_2 = 0.3$ ,  $P = 0.86$ ; treatment (control/laying supp)\* sex  $\chi^2_2 = 0.24$ ,  $P = 0.63$ ; hatch  
679 order\*brood size  $\chi^2_2 = 4.83$ ,  $P = 0.09$ ; brood size  $\chi^2_1 = 0.26$ ,  $P = 0.61$ ; date  $\chi^2_1 = 0.21$ ,  $P = 0.65$ ; treatment (control/laying supp)  $\chi^2_1 =$   
680  $1.69$ ,  $P = 0.19$ ; maternal age  $\chi^2_1 = 2.28$ ,  $P = 0.13$ .

681 <sup>4</sup> Removed from models: hatch order\*sex\*treatment (control/laying supp)  $\chi^2_2 = 0.5$ ,  $P = 0.78$ ; hatch order\*treatment (control/laying  
682 supp)  $\chi^2_2 = 0.4$ ,  $P = 0.82$ ; sex\*treatment (control/laying supp)  $\chi^2_2 = 0.38$ ,  $P = 0.54$ ; hatch order\* sex  $\chi^2_2 = 0.88$ ,  $P = 0.65$ ; maternal age  
683  $\chi^2_1 = 0.09$ ,  $P = 0.77$ ; date  $\chi^2_1 = 0.82$ ,  $P = 0.18$ .

684 <sup>5</sup> Removed from models: hatch order\*treatment (control/carotenoid supp.)  $\chi^2_2 = 0.75$ ,  $P = 0.69$ ; date  $\chi^2_1 = 0.05$ ,  $P = 0.83$ ; treatment  
685 (control/ carotenoid supp)  $\chi^2_1 = 0.92$ ,  $P = 0.34$ ; maternal age  $\chi^2_1 = 2.04$ ,  $P = 0.15$ .

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690 Table 4. Results of Cormack-Jolly-Seber survival analysis in Program MARK.

Model	Num. Par	QAICc	$\Delta$ QAICc	Weight	Deviance
$\phi$ (age) p(time)	22	889.08	0.00	0.63	486.56
$\phi$ (age + sex) p(time)	23	890.44	1.36	0.32	485.72
$\phi$ (age + sex + hatch order) p(time)	25	894.17	5.09	0.05	485.03
$\phi$ (age + sex + hatch order + time) p(time)	44	904.16	15.08	0.00	451.08
$\phi$ (sex) p(time)	22	934.59	45.51	0.00	532.07
Null model $\phi(\cdot)$ p(time)	17	925.29	36.21	0.00	533.62

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Table 5. Summary of results showing effects of hatch order and carotenoid supplementation on a) male and b) female nestlings.

		Asymptotic mass	Growth rate (mass)	Asymptotic tarsus length	Growth rate (tarsus)	Survival to fledging	Longevity	Total offspring
a)	Hatch order	-	-	-	-	-	=	=
male	Carotenoids	=	=	=	+	=	=	=
b)	Hatch order	-	-	-	=	=	=	=
female	Carotenoids	=	=	=	=	=	=	=

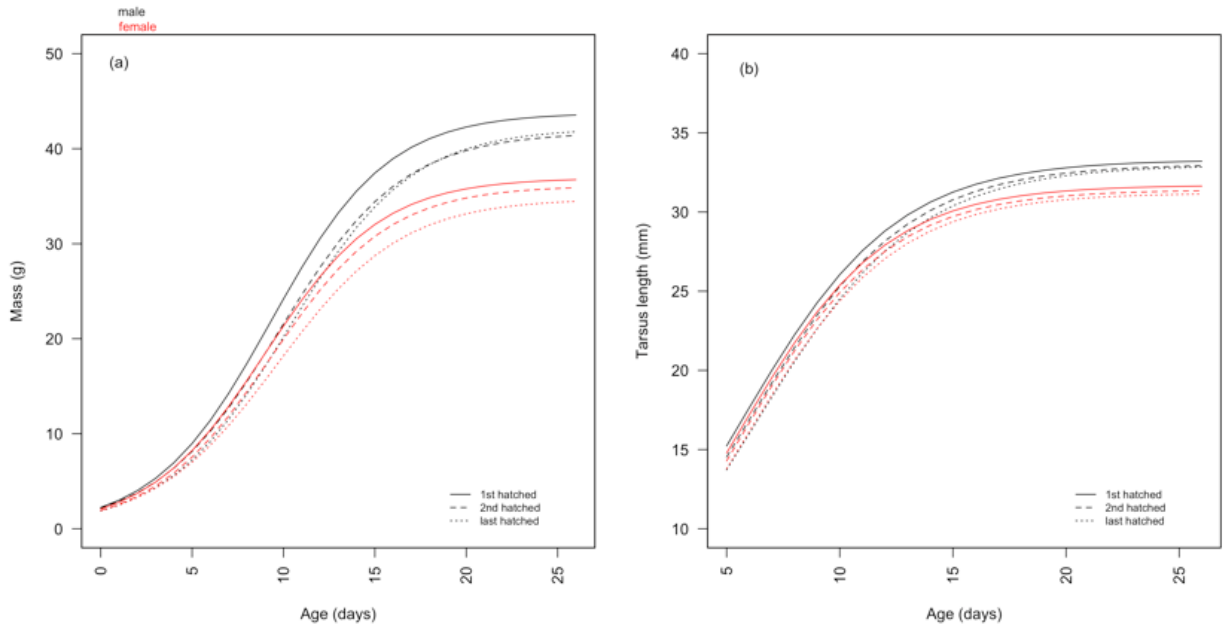
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Negative effects are denoted by “-”, positive effects by “+”, and no effect by “=”.



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751 Fig. 1. Effects of hatch order on (a) mass, and (b) tarsus length. Male nestlings are depicted in black, and female  
752 nestlings are depicted in red.



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