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The effects of aerobic training before and after the induction of Alzheimer's disease on ABCA1 and APOE mRNA expression and the level of soluble Aβ1-42 in the hippocampus of male Wistar rats

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ABSTRACT

Objective(s): The purpose of this study was to investigate the effects of aerobic training before and after the induction of Alzheimer's disease on ABCA1 and APOE mRNA expression and the level of soluble $A\beta1-42$ in the hippocampus of male Wistar rats.

Materials and Methods: Ninety six eight-week-old male Wistar rats were randomly divided into two groups: Training (n=48) and Rest (n=48). After four weeks, each group was randomly divided into two subgroups: intra-hippocampal injection of A β 1-42 (n=24) and DMSO (n=24). Then, each group was again randomly divided into two groups: Training (n=12) and Rest (n=12). After four weeks, each group was again randomly divided into two groups: Behavioral test (n=7) and sacrificed (n=5).

Results: The one-way ANOVA showed a significant increase in the mRNA expression of ABCA1 (P<0.05), a significant decrease in the level of soluble A β 1-42, and no significant difference in the expression of APOE mRNA (P>0.05) in the hippocampus as a result of training. The analysis of the Morris water maze data showed that intra-hippocampal injection of A β 1-42 impaired spatial learning and memory and exercise improved spatial learning (P<0.05) and memory (P<0.05).

Conclusion: Therefore, aerobic training by a significant increase in the mRNA expression of ABCA1, which is the main factors of lipid metabolism in the brain and which is involved in the pathology of Alzheimer's disease, can be consistent with improving cognitive function as an effective way of preventing and improving the symptoms of Alzheimer's disease.

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Introduction

Alzheimer's is a progressive neurodegenerative disease and the most common form of dementia (1). Alzheimer's disease (AD) is associated with the atrophy of the hippocampus and the cerebral cortex (2). It begins with short-term memory impairment and results in the complete decline of cognitive function (3, 4). AD is clinically diagnosed by progressive cognitive im

pairment (5) but pathologically diagnosed by Neurofibrillary Tangles (NFTs) and extracellular amyloid plaques. NFTs include tau hyper-phosphorylated proteins in the brain or the hippocampal cortex (2), and the amyloid plaques are mainly caused by the fibrillation of beta-amyloid peptide (Aβ) (6-8). The level of $A\beta$ in the brain is determined by the dynamic balance between the factors that produce it from the protein of the amyloid precursor (APP), which is a transmembrane protein, and the factors that clear it from the brain. Thus, the deficiency of the balance between the production and the clearance of AB can cause the accumulation and deposition of the Aβ peptide in the brain of AD patients, and the factors that stimulate the cleavage of APP or reduce the clearance of Aβ peptide are considered as the risk factors of AD (9).

Recent research has shown a link between lipid

metabolism and AD and has identified some risk factors of the early onset of AD related to cholesterol metabolism (10-12). Lipid metabolism in the brain is widely differentiated from its peripheral metabolism due to the blood-brain barrier and different lipoproteins. Thus, identifying the mechanism of lipid metabolism in the brain may be a new therapeutic strategy in AD (13). Therefore, some factors are involved in the metabolism of cholesterol and lipids in the brain. Two of these are apolipoprotein E (APOE) and ABCA1 (ATP-binding cassette transporter), whose role in AD pathology, as recent studies consider, is to influence the production and clearance of $A\beta$.

APOE is the most important apolipoprotein in the brain, mainly synthesized by astrocytes and a little by microglia (14). It is a chaperone for A β ; it affects the deposition and clearance of A β (15, 16). APOE increases the proteolytic degradation of A β by NEP (Neprilysin) and IDE (Insulin Degrading Enzyme), especially when it is lipidated (14). On the other hand, the clearance of A β 1-42 is less dependent on the efflux from the bloodbrain barrier, and the role of proteolytic clearance by NEP and IDE is greater (17). Therefore, the levels of APOE and lipidated APOE affect A β metabolism (14, 18).

ABCA1 is a membrane protein that mediates the

cholesterolefflux to low-lipid or lipid-free apolipoproteins. ABCA1 transports the cholesterol to the APOA1 in the periphery and to the APOE in the brain (19, 20). The role of ABCA1 in regulating plasma HDL (high-density lipoprotein) levels in atherosclerosis has been considered. In addition, due to its effects on lipidization and APOE stability, it is involved in AD pathology. ABCA1 increases the degradation of AB by increasing the lipidation of APOE and can cause lower AB deposition and higher clearance of A β (21). On the other hand, APP is embedded in the cell membrane and cleaved by α -secretarase and β-sucrose enzymes that exist in the membrane (22, 23). The increased intracellular or membrane cholesterol can lead to the increasing cleavage of APP and produce Aβ. Therefore, ABCA1 can decrease AB production through the transport of cholesterol to APOE (23). As a result, the increase in the expression of ABCA1 by a decrease in the production of Aβ, and on the other hand, by an increase in the clearance of Aβ, is associated with preventing and treating AD.

Most studies have shown that physical activity and exercise can be effective in improving memory, learning and cognitive function (24-27) and possibly in reducing the risk of dementia (28). In addition, researchers have suggested that the cholesterol metabolism disorder in the brain is involved in the pathogenesis of AD (14), and the polymorphism of genes that regulate the cholesterol metabolism, such as ABCA1 and APOE, increase the risk of AD (23). But the effects of exercise on lipid metabolism in the brain and its role in preventing and improving the symptoms of AD have not been studied. Studies in this regard have been carried out using genetic and pharmaceutical interventions. Both in vivo and in vitro studies have shown that the down-regulation of the ABCA1 expression reduces the APOE in the brain, the cerebrospinal fluid, and plasma, and increases the AB load of the brain (29-33), while the up-regulation of the ABCA1 expression leads to an increase in the lipidation of APOE and reduces the Aβ load (34). Studies have also indicated that the administration of the RXRs (Retinoid X Receptors) and LXRs (Liver X Receptors) agonist increases the ABCA1 and APOE expressions, leads to an increase in the clearance of Aβ by lipidated APOE and causes an improvement in cognitive impairment (34-37). Therefore, because of the strong link between lipid metabolism in the brain and AD and since no studies have been done to investigate the effect of exercise, the purpose of this study was to investigate the effects of aerobic training before and after the induction of AD on ABCA1 and the APOE mRNA expression and the level of soluble Aβ1-42 in the hippocampus of male Wistar rats.

Materials and Methods

Animals

In this experimental study, 96 adult male Wistar rats (8 weeks old, 20 ± 195 g weight) were used. Five rats were housed in a Plexiglas cage in a 12-hr light/dark cycle. The temperature was 22 ± 3 °C and humidity was about 45%. There was no restriction on the animals' access to water and food. After a week of familiarization with the environment, all rats were exposed to a familiarization with the treadmill for a week (10 min, speed: 10 m/min, 5 days). Then, the rats were randomly classified into a training group (48 rats) and a resting group (48 rats).

After four weeks, each group was randomly classified into two subgroups: The Aβ group (24 rats injected with Aβ in the hippocampus) and the sham group (24 rats injected with DMSO in the hippocampus). After seven days, each of these groups was randomly classified into two subgroups: Subgroup 1 (out of 12 rats, which had undergone endurance training for four weeks and 48 hr had passed after their last training session, five were sacrificed and seven took a behavioral test), and Subgroup 2 (out of 12 rats, which had not undergone endurance training and resting for four weeks, five were sacrificed simultaneously with the training group and seven took the behavioral test. All experiments were done following the guidelines of United States National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals and were approved by the Ethics Committee on the use of animals of Ferdowsi University of Mashhad, Mashhad, Iran (code of approval: IR.MUM. FUM.REC.1396.06).

Exercise training protocol

The aerobic exercise protocol was performed on a treadmill with a gradient of zero degrees five days a week for four weeks in both pre- and post-conditioning phases. This protocol was mild to moderate and the speed and duration were gradually increased (Table 1). The exercise-group rats were monitored during the training sessions and were encouraged by a weak electrical shock (0.5 mA intensity) that did not cause stress in the animal (38, 39).

Preparation of A\beta1-42 & stereotaxic

We utilized the intra-hippocampal injection of A\beta1-42 peptide to generate the AD model. To prepare the Aβ1-42 peptide, Aβ1-42 (Abcam, USA) was dissolved in a buffer solution (DMSO 3%) (Sigma Aldrich, USA) at the rate of 5 μ g/ μ l. Then, they were divided into 30 μl per vial and kept at -80 °C. The amyloid solution was incubated for seven days at 37 °C to fibrillate the Aβ1–42 peptides (40). Then, all the animals were anesthetized with the intraperitoneal injection of ketamine (100 mg/kg) and xylazine (10 mg/kg) and they underwent stereotaxic surgery according to the Atlas of Paxinus and Watson: Anterior-posterior (AP), 3.8 mm; mediallateral (ML), ±2.2 mm from the central sagittal line and dorsal-ventral (DV), -2.7 mm to the surface of the skull (41). In the A β groups, the A β 1-42 peptide (1 ul) was injected into the CA1 area of the right and left hippocampus by the Hamilton syringe, and in the sham groups, the DMSO buffer (1 µl) was injected into the CA1 area of each hippocampus.

Tissue biopsies

Twenty-four hours after the last training session,

Table1. Exercise training protocol used in pre- and post-conditioning phases

	Set *time (min)	Speed (m/min)	Days		
Week 1	2*15	10	5		
Week 2	2*15	10	5		
Week 3	3*15	15	5		
Week 4	4*15	15	5		

Table 2. Oligonucleotide primer sequences and real-time PCR amplification parameters temperature

Gene	Forward and reverse primer sequences (5' \rightarrow 3')	Melting temperature (Tm)	Product size (bp)		
ABCA ₁	F: TCATGTACCCAGCGTCCTTT	82.74°C	96		
APOE	R: CCACACTACCATTGATGCCG F: ATCTGTCACCTCCTGCTCT	79.46°C	116		
GAPDH	R: CTTTTCCTTCCGCTGACTGG F: AAG TTC AAC GGC ACA GTC AAG G	82.75°C	121		
	R: CAT ACT CAG CAC CAGCAT CAC C				

the rats were anesthetized by intraperitoneal injection of ketamine (100 mg/kg) and xylazine (10 mg/kg). Then, the animals were decapitated and the brain was removed from the skull bone. Subsequently, the hippocampal tissue was separated from the rest of the brain tissue, frozen with liquid nitrogen and stored at -80 °C for future analysis.

RNA extraction, cDNA synthesis, and real-time PCR

The total RNA was extracted from the right hippocampus tissue homogenized in TRIzol solution (1 ml) according to the manufacturer's instructions. The concentration of RNA was determined by the Nanodrop spectrophotometer and 1000 ng RNA was used to synthesize cDNA. The cDNA was synthesized by a high-capacity cDNA reverse transcription kit (Applied Biosystems) following the instructions of the cDNA synthesis kits. Real-time PCR was performed using the RealQ Plus 2x Master Mix Green (AMPLIQON) and using 1 μl cDNA, 10 μl Master Mix, 1 μl of each of the forward and reverse primers, and 7 μ l RNase-free water. All PCR reactions were performed in duplicate. The polymerase chain reaction was performed in the following thermal conditions: denaturation at 95 °C for 10 min, followed by 40 cycles of 95 °C for 15 sec, and annealing at 60 °C for 30 sec. The oligonucleotide primer sequences for ABCA1, APOE, and GAPDH (as an internal control) genes are listed in Table 2. The mRNA expression relative values were analyzed by the $2^{-\Delta\Delta Ct}$ method (42).

ELISA

Left hippocampus tissue of rats was homogenized with 1 ml of PBS solution and kept at -20 °C for one night. After applying two freeze-thaw cycles to break down the cell membranes, the homogenized tissue was centrifuged at 5,000 rpm for five min and the supernatant was extracted by the sampler. Subsequently, the hippocampal samples were used to determine the levels of soluble A β 1-42 using the ELISA kit (CSB-E10786r; Cusabio Biotech, China), based on its manufacturer's protocol.

Behavioral test

The Morris water maze test was used to measure memory and learning. We used a dark circular pool with a diameter of 150 cm and a depth of 60 cm. The pool was filled with water (21 $^{\circ}$ C \pm 2) to a depth of 30 cm. A hidden platform with a 10 cm diameter was located at the center of the southeast (SE) quadrant, about 2 cm below the water surface. The escape latency and the traveled distance and time in each quadrant were recorded by a video camera placed above the pool and was analyzed using the Ethovision Video Tracking System. The Morris

water maze protocol was as follows:

A) Learning stage: At this stage, the rats were trained for four consecutive days, each day in four separate trials, to find the hidden platform. At the start of each trial, each rat was placed on the platform for 15 to 20 sec. Then, the animal was randomly placed in the water from one of the four starting points (north, south, east, and west). The animal was allowed to swim and find the hidden platform located in the SE quadrant. If the rat could not find the platform within 60 sec, it would be guided by hand. Once it found the platform, the animal was allowed to remain on it for 20 sec. The escape latency and traveled distance were measured and recorded in each trial.

B) Probe trial: A day after the learning stage, the spatial memory of the animals was evaluated in the probe trial. In this stage, the rats were evaluated in a 60-second test in which the Plexiglas platform was removed from the water. The time spent in the target quadrant was recorded (43).

Statistical analysis

To determine the normality of the distribution, we used the Shapiro-Wilk test. Also, Levine's test was used to check the homogeneity of the variances. The Multivariate analysis of variance (MANOVA) and oneway ANOVA with Tukey's *post hoc* test were used for the statistical analysis of the data at the significant level ($P \le 0.05$). All results are expressed as mean \pm SEM.

Results

Analysis of biochemical parameters

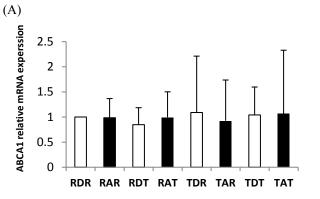
In this research, 40 male Wistar rats were sacrificed and the APOE and ABCA1 expression and the level of soluble Aβ1-42 in the hippocampus were measured in eight groups. Figures 1 and 2 show changes in the level of soluble Aβ1-42, and APOE and ABCA1 expression in the hippocampus of the eight groups. The results of the MANOVA test showed a significant difference between the eight groups with respect to these three dependent variables: Wilks $\wedge = 0.020$, F (21, 86.7) = 12.070, P = 0.000, Eta = 0.730, indicating that aerobic training has a significant effect on the APOE and ABCA1 expressions and the level of soluble A β 1-42 in the hippocampus of rats (P<0.01), and 73% of the variability of these three variables was influenced by this training program. Table 3 shows the results by a one-way ANOVA for comparing APOE and ABCA1 expressions, and the level of soluble Aβ1-42 in the hippocampus of the rats of these eight groups.

The results of Table 3 show that aerobic training can significantly change the ABCA1 expression (P<0.05) in the hippocampus of rats. However, changes in the

Table 3. The APOE and ABCA1 expressions and the level of soluble $A\beta1-42$ in the hippocampus of male Wistar rats in the eight groups. Each column is for each group including five rats

-		RDR	RAR	RDT	RAT	TDR	TAR	TDT	TAT	F	P	Eta
ABCA1	Mean	1	1.2059	6.3204	4.2941	3.5871	2.8837	7.2283	5.9344	2.576	0.032	0.73
	SD	0	1.2509	4.1496	3.0281	3.5228	0.7554	4.8307	4.4343			
APOE	Mean	1	0.9847	0.8470	0.9944	1.0891	0.9243	1.0409	1.0668	0.075	1.000	0.012
	SD	0	0.3819	0.3383	0.5066	1.1225	0.8117	0.5557	1.2624			
Αβ1-42	Mean	17.2060	46.0800	15.5000	31.7600	17.2820	35.2200	15.7300	26.2980	134.4	0.000	0.967
	SD	2.8055	1.7922	1.2549	2.7553	2.4055	2.8083	0.8303	1.7919			

Rest-DMSO-Rest (RDR), Rest-A β -Rest (RAR), Rest-DMSO-Training (RDT), Rest-A β -Training (RAT), Training-DMSO-Rest (TDR), Training-A β -Rest (TAR), Training-DMSO - Training (TDT), Training-A β - Training (TAT). Data expressed as Mean \pm SD



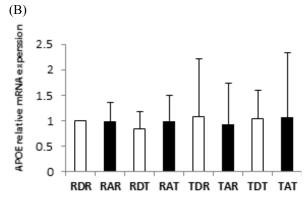


Figure 1. Real-Time PCR of hippocampus ABCA1 (A) and APOE (B) relative mRNA expression in eight groups .Rest-DMSO-Rest (RDR), Rest-Aβ-Rest (RAR), Rest-DMSO-Training (RDT), Rest-Aβ-Training (RAT), Training-DMSO-Rest (TDR), Training-Aβ-Rest (TAR), Training-DMSO - Training (TDT), Training-Aβ- Training (TAT)

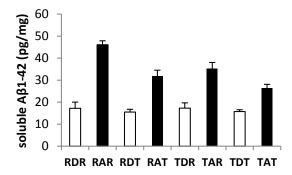


Figure 2. The level of soluble Aβ1-42 in the hippocampus in eight groups. Rest-DMSO-Rest (RDR), Rest-Aβ-Rest (RAR), Rest-DMSO-Training (RDT), Rest-Aβ-Training (RAT), Training-DMSO-Rest (TDR), Training- Aβ-Rest (TAR), Training- DMSO - Training (TDT), Training- Aβ-Training (TAT)

expression of the APOE gene were non-significant (P>0.05). The results also showed that aerobic training can significantly change the level of soluble A β 1-42 in the hippocampus of rats (P<0.01).

The results of the *post hoc* test showed a significant difference in the ABCA1 expression in RAR-TAT (P= 0.028), RDR-RDT (P=0.014), and RDR-TDT (P=0.005). In the A β groups, pre-conditioning, post-conditioning, and pre-post conditioning increased the ABCA1 expression, but a significant increase was observed only between training before and after A β injection and resting before and after A β injection. In the sham groups, pre-post conditioning showed a significant increase in the ABCA1 expression compared to resting before and after the DMSO injection. Therefore, the highest increase in ABCA1 expression was observed in the groups that trained before and after the injection, and the exercise before or after the injection alone could not significantly increase the ABCA1 expression.

The results of the *post hoc* test showed a significant difference in the level of soluble A β 1-42 in the RAR-TAT (P=0.000), TAR-TAT (P=0.000), and RAT-TAT (P=0.000) groups. According to these results, the level of soluble A β 1-42 in the TAT group was significantly lower than the other groups. Also, there was no significant difference between the groups that trained before or after the injection. These results indicated that aerobic training can reduce the level of soluble A β 1-42 and that compared to pre- or post-conditioning, pre-post-conditioning has a greater effect on reducing the level of soluble A β 1-42 in the hippocampus.

Analysis of behavioral test

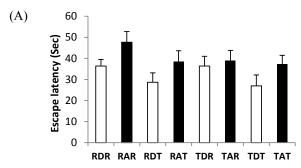
To measure the cognitive function, 56 male Wistar rats participated in the behavioral test. Figures 3 shows the escape latency, the traveled distance, and the time in the target quadrant (probe trial) of male Wistar rats in the eight groups. The results of the MANOVA test showed a significant difference between the eight groups in behavioral variables: Wilks \land = 0.092, F (21, 132.6) = 8.178, P=0.000, Eta = 0.548, which indicates that aerobic training has a significant effect on the learning and memory of rats (P<0.01), and 55% of the variability of these three variables was influenced by aerobic training in this study. Table 4 shows the results of a one-way ANOVA for comparing the behavioral variables in rats between the eight groups.

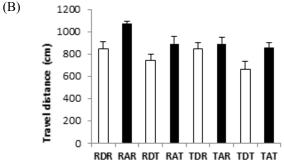
The results of Table 4 show that aerobic training caused a significant decrease in the escape latency (P<0.01) and the traveled distance (P<0.01) and a significant increase in the time spent in the target quadrant (P<0.01) in the trained groups. These results indicate the effect of the training program on the cognitive function in both the sham and A β groups.

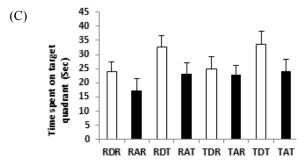
Table 4. The escape latency, traveled distance, and time spent on the target quadrant (probe trial) of male Wistar rats in the eight groups. Each column is for each group including seven rats

		RDR	RAR	RDT	RAT	TDR	TAR	TDT	TAT	F	P	Eta
Travel	Mean	864.99	1069.5	738.00	891.39	842.77	891.29	658.85	854.74	30.32	0.000	0.816
distance	SD	63.74	26.94	56.09	64.59	60.27	59.14	72.81	45.78			
Escape	Mean	36.36	47.67	28.66	38.38	36.37	39.04	26.95	37.35	13.50	0.000	0.663
latency	SD	3.14	5.06	4.47	5.25	4.61	4.75	5.20	4.10			
Probe	Mean	24.01	17.00	32.57	23.31	25.03	22.91	33.52	24.11	12.75	0.000	0.650
	SD	3.35	4.43	4.02	3.66	4.08	3.15	4.84	4.07			

Data expressed as Mean±SD







The results of the *post hoc* test showed a significant difference in the escape latency, the traveled distance, and the time spent in the target quadrant between the sham and the A β groups. The comparison of scores between RAR-RDR, RAT-RDT, TAR-TDR, and TAT-TDT (P<0.01) showed that in all groups, the rats in the A β group showed the worse scores for all three variables. These results indicate cognitive impairment in the rats that were exposed to the induction of AD.

The results of the *post hoc* test showed a significant difference in the escape latency, the traveled distance, and the time spent in the target quadrant between the TAT, TAR, and RAT groups with the RAR group (P<0.01). However, there was no significant difference between TAT, TAR, and RAT groups in the three behavioral variables (P>0.05). These results indicate that pre-conditioning, post-conditioning, and prepost conditioning play an effective role in improving cognitive function in rats with AD.

In addition, the results of the *post hoc* test showed a significant difference in the escape latency, the traveled distance, and the time spent in the target quadrants between the TDT and RDT groups with the RDR group (P<0.01). There was no significant difference between the TDT and RDT groups (P>0.05), indicating no significant difference between four and eight weeks of aerobic training. Therefore, in this study, aerobic training improved the cognitive function in the rats with no AD.

Figure 3. The escape latency (A), the travel distance (B) and the time spent on the target quadrant (C) in Morris water maze in eight groups .. Rest-DMSO-Rest (RDR), Rest- $A\beta$ -Rest (RAR), Rest-DMSO-Training (RDT), Rest- $A\beta$ -Training (RAT), Training-DMSO-Rest (TDR), Training- $A\beta$ -Rest (TAR), Training-DMSO-Training (TDT), Training- $A\beta$ -Training (TAT)

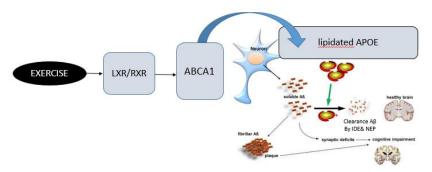


Figure 4. The proposed mechanism through which exercise can improve the clearance of A β . Exercise likely as an LXR/RXR agonist increased mRNA expression ABCA1 and subsequently, leading to an increase in lipidation of APOE. The lipidated APOE facilitates the proteolytic degradation of A β by NEP and IDE and promotes the clearance of A β

Discussion

The results of this study showed that aerobic training before and after the AD induction increased the ABCA1 expression in the hippocampus of rats in both the sham and Aβ groups. There was no significant difference in APOE expression between the eight groups. The expression of ABCA1 and APOE is regulated by nuclear receptors, the peroxisome proliferator-activated receptor γ (PPAR-γ), and the liver X receptors (LXRs) (21, 44). In the metabolic pathway, PPAR-γ results in the expression of LXR and ultimately its target genes ABCA1 and APOE (45-49). Thus, increasing the expression of the ABCA1 gene through the LXR and PPAR-γ agonists results in the synthesis of APOE-containing HDL particles. As a result, the lipidated APOE facilitates the intracellular and extracellular proteolytic degradation of Aβ by NEP and IDE and promotes the clearance of Aβ

Several studies have been conducted on the role of ABCA1 and APOE in AD pathology and most of these have examined the effects of LXR and PPAR-y agonists and genetic interventions (e.g. genetic overexpression or deletion of ABCA1). The results of these studies have shown that the effects of RXR and LXR agonists increase the expression of ABCA1 and APOE in mice and promote the clearance of sAB and improve cognitive impairment (35-37). The genetic overexpression of ABCA1 in transgenic mice showed a decrease in the soluble AB and AB plagues (34), and the deletion of ABCA1 increases the soluble and insoluble A\beta levels and leads to the progress of the pathology of Aβ plaques in AD mice (32). Other studies in APP transgenic rats have shown that the deletion of ABCA1 leads to increased AB deposition and a significant decrease in the total APOE in the brain (29, 31). Therefore, in this study, aerobic exercise before and after AD induction, possibly similar to LXR and PPAR-y agonists, increased the expression of ABCA1 in both the Aβ and the sham groups. Thus, aerobic exercise can be used as an effective alternative method to drugs and genetic interventions and will be considered in improving the pathology of AD.

On the other hand, studies have shown that exercise can promote the clearance of Aβ and decrease the level of soluble A β in Alzheimer's transgenic mice (51-53). The results of the study indicate that the clearance of AB increases via mechanisms like degrading enzymes (54), neural mechanisms (55), and neural growth factors (51). The results of drug and genetic interventions also show a decrease in the level of soluble $A\beta$ along with an increase in the expression of ABCA1. The study results showed that the expression of ABCA1 increased and the level of soluble Aβ1-42 decreased in the hippocampus of rats in the training group compared to the resting group. Therefore, aerobic training before and after the induction of AD probably by increasing the expression of ABCA1 leads to a decrease in the level of soluble Aβ1-42 in the hippocampus of rats. The proposed mechanism of these changes can be explained by an increase in the level of lipidated APOE and a facilitation of the degradation of AB (Figure 4). The better cognitive function and the lower level of soluble Aβ1-42 in the hippocampus, coupled with an increase in the expression of ABCA1, expresses the effects of aerobic training before and after AD induction on the disease pathology.

Most studies indicate that exercise and physical activity improve cognitive function (24, 25). Research has shown the effect of physical activity on CNS adaptations, especially in the hippocampus. Exercise and physical activity lead to neurogenesis, as well as changes in the synaptic plasticity in the hippocampus, which improves cognitive functions, such as learning and memory (26, 27, 56). The results of the behavioral tests in this study showed that the rats of Aß group had a significantly worse performance than the sham group in the memory and learning tests. These results indicated cognitive impairment and AD induction in the Aβ group. In addition, the effects of exercise in both $A\beta$ and sham groups indicated an improvement in the cognitive function as a result of aerobic exercise. Therefore, the results of this study showed that aerobic training before and after the induction of AD, with the effect on the metabolism of lipid in the brain and increasing the expression of the ABCA1 and the clearance of Aβ1-42 in the hippocampus, could improve the cognitive function

Most studies have shown that the APOE genotype is the strongest genetic risk factor for AD (12, 57). In humans, there are three isoforms: APOE2, APOE3, and APOE4 (58). APOE4 increases the risk of AD (57, 59), while APOE2 and APOE3 are associated with reducing the risk (60). APOE2, especially, inhibits A β deposition and plays a protective role (14, 61). The results of research indicate that the APOE4 affinity to complex with A β is lower than APOE2 and APOE3 (62).

Conclusion

APOE4 is thus considered to be the strongest risk factor for AD because APOE4 increases the A β deposition in amyloid plaques and inhibits the clearance of A β . In addition, the lipidation of APOE increases the affinity for A β , and any method that can increase the level of lipidated APOE can be effective in preventing and treating AD. According to the importance of ABCA1 and APOE in the pathology of AD, and according to the outcomes of this study, aerobic exercise before and after the induction of AD can be used as an alternative to genetic and pharmaceutical interventions. It can have beneficial effects on the prevention and reduction of the symptoms of AD.

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Conflicts of Interest

The Authors declares that there is no conflict of interest.

References

- 1. De Souza LC, Sarazin M, Goetz C, Dubois B. Clinical investigations in primary care. Front Neurol Neurosci. 2009;24:1-11.
- 2. Braak H, Braak E. Neuropathological stageing of Alzheimer-related changes. Acta Neuropathol. 1991;82:239-259.
- 3. Pimplikar SW, Nixon RA, Robakis NK, Shen J, Tsai LH. Amyloid-independent mechanisms in Alzheimer's disease

- pathogenesis. J Neurosci. 2010;30:14946-14954.
- 4. Wenk GL. Neuropathologic changes in Alzheimer's disease: potential targets for treatment. J Clin Psychiatry. 2006;3:3-7.
- 5. 2013 Alzheimer's disease facts and figures. Alzheimers Dement. 2013;9:208-245.
- 6. Selkoe DJ. Alzheimer's disease: genes, proteins, and therapy. Physiol Rev. 2001;8:741-766.
- 7. Hardy J. A hundred years of Alzheimer's disease research. Neuron. 2006;52:3-13.
- 8. Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. Science. 2002;297:353-356.
- 9. Allinquant B, Clamagirand C, Potier MC. Role of cholesterol metabolism in the pathogenesis of Alzheimer's disease. Curr Opin Clin Nutr Metab Care. 2014;17:319-323.
- 10. Reitz C. Dyslipidemia and the risk of Alzheimer's disease. Curr Atheroscler Rep. 2013;15:307-318.
- 11. Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, *et al.* Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. Nat Genet. 2013;45:1452-1458.
- 12. Dorszewska J, Prendecki M, Oczkowska A, Dezor M, Kozubski W. Molecular basis of familial and sporadic Alzheimer's Disease. Curr Alzheimer Res. 2016;13:952-963.
- 13. Kim J, Yoon H, Horie T, Burchett JM, Restivo JL, Rotllan N, *et al.* microRNA-33 regulates apoE lipidation and amyloid-beta metabolism in the brain. J Neurosci. 2015;35:14717-14726.
- 14. Kim J, Basak JM, Holtzman DM. The role of apolipoprotein E in Alzheimer's disease. Neuron. 2009;63:287-303.
- 15. Fan J, Donkin J, Wellington C. Greasing the wheels of Abeta clearance in Alzheimer's disease: the role of lipids and apolipoprotein E. Biofactors. 2009;35:239-248.
- 16. Holtzman DM, Herz J, Bu G. Apolipoprotein E and apolipoprotein E receptors: normal biology and roles in Alzheimer disease. Cold Spring Harb Perspect Med. 2012:2:a006312.
- 17. Deane R, Wu Z, Sagare A, Davis J, Du Yan S, Hamm K, *et al.* LRP/amyloid beta-peptide interaction mediates differential brain efflux of Abeta isoforms. Neuron. 2004;43:333-344.
- 18. Bien-Ly N, Gillespie AK, Walker D, Yoon SY, Huang Y. Reducing human apolipoprotein E levels attenuates age-dependent Abeta accumulation in mutant human amyloid precursor protein transgenic mice. J Neurosci. 2012;32:4803-4811.
- 19. Stukas S, Robert J, Wellington CL. High-density lipoproteins and cerebrovascular integrity in Alzheimer's disease. Cell Metab. 2014;19:574-591.
- 20. Hayden MR, Clee SM, Brooks-Wilson A, Genest J, Attie A, Kastelein JJ. Cholesterol efflux regulatory protein, Tangier disease and familial high-density lipoprotein deficiency. Curr Opin Lipidol. 2000;11:117-122.
- 21. Koldamova R, Fitz NF, Lefterov I. ATP-binding cassette transporter A1: from metabolism to neurodegeneration. Neurobio Dis. 2014;72:13-21.
- 22. O'Brien RJ, Wong PC. Amyloid precursor protein processing and Alzheimer's disease. Annu Rev Neurosci. 2011;34:185-204.
- 23. Maulik M, Westaway D, Jhamandas JH, Kar S. Role of cholesterol in APP metabolism and its significance in Alzheimer's disease pathogenesis. Mol Neurobiol. 2013;47:37-63.
- 24. Lin TW, Chen SJ, Huang TY, Chang CY, Chuang JI, Wu FS, *et al.* Different types of exercise induce differential effects on neuronal adaptations and memory performance. Neurobiol Learn Mem. 2012;97:140-147.
- 25. Thomas AG, Dennis A, Bandettini PA, Johansen-Berg H. The effects of aerobic activity on brain structure. Front Psychol. 2012:3:86-97.
- 26. Cotman CW, Berchtold NC. Exercise: a behavioral intervention

- to enhance brain health and plasticity. Trends Neurosci. 2002:25:295-301.
- 27. Garcia PC, Real CC, Ferreira AF, Alouche SR, Britto LR, Pires RS. Different protocols of physical exercise produce different effects on synaptic and structural proteins in motor areas of the rat brain. Brain Res. 2012;1456:36-48.
- 28. Rockwood K, Middleton L. Physical activity and the maintenance of cognitive function. Alzheimers Dement. 2007;3:38-44.
- 29. Hirsch-Reinshagen V, Maia LF, Burgess BL, Blain JF, Naus KE, McIsaac SA, *et al.* The absence of ABCA1 decreases soluble apoE levels but does not diminish amyloid deposition in two murine models of Alzheimer disease. J Biol Chem.. 2005;280:43243-43256.
- 30. Hirsch-Reinshagen V, Zhou S, Burgess BL, Bernier L, McIsaac SA, Chan JY, *et al.* Deficiency of ABCA1 impairs apolipoprotein E metabolism in brain. J Biol Chem. 2004;279:41197-41207.
- 31. Koldamova R, Staufenbiel M, Lefterov I. Lack of ABCA1 considerably decreases brain ApoE level and increases amyloid deposition in APP23 mice. J Biol Chem. 2005;280:43224-43235.
- 32. Wahrle SE, Jiang H, Parsadanian M, Hartman RE, Bales KR, Paul SM, *et al.* Deletion of abca1 increases Abeta deposition in the PDAPP transgenic mouse model of Alzheimer disease. J Biol Chem. 2005;280:43236-43242.
- 33. Wahrle SE, Jiang H, Parsadanian M, Legleiter J, Han X, Fryer JD, *et al.* ABCA1 is required for normal central nervous system ApoE levels and for lipidation of astrocyte-secreted apoE. J Biol Chem. 2004;279:40987-40993.
- 34. Wahrle SE, Jiang H, Parsadanian M, Kim J, Li A, Knoten A, *et al.* Overexpression of ABCA1 reduces amyloid deposition in the PDAPP mouse model of Alzheimer disease. J Clin Invest. 2008;118:671-682.
- 35. Corona AW, Kodoma N, Casali BT, Landreth GE. ABCA1 is necessary for bexarotene-mediated clearance of soluble amyloid beta from the hippocampus of APP/PS1 mice. J Neuroimmune Pharmacol. 2016;11:61-72.
- 36. Terwel D, Steffensen KR, Verghese PB, Kummer MP, Gustafsson JA, Holtzman DM, *et al.* Critical role of astroglial apolipoprotein E and liver X receptor-alpha expression for microglial Abeta phagocytosis. J neuroscience. 2011;31:7049-7059.
- 37. Donkin JJ, Stukas S, Hirsch-Reinshagen V, Namjoshi D, Wilkinson A, May S, *et al.* ATP-binding cassette transporter A1 mediates the beneficial effects of the liver X receptor agonist GW3965 on object recognition memory and amyloid burden in amyloid precursor protein/presenilin 1 mice. J Biol Chem. 2010;285:34144-34154.
- 38. Dao AT, Zagaar MA, Alkadhi KA. Moderate treadmill exercise protects synaptic plasticity of the dentate gyrus and related signaling cascade in a rat model of Alzheimer's disease. Mol Neurobiol. 2015;52:1067-1076.
- 39. Zagaar M, Alhaider I, Dao A, Levine A, Alkarawi A, Alzubaidy M, *et al*. The beneficial effects of regular exercise on cognition in REM sleep deprivation: behavioral, electrophysiological and molecular evidence. Neurobiol Dis. 2012;45:1153-1162.
- 40. Doost Mohammadpour J, Hosseinmardi N, Janahmadi M, Fathollahi Y, Motamedi F, Rohampour K. Non-selective NSAIDs improve the amyloid-beta-mediated suppression of memory and synaptic plasticity. Pharmacol Biochem Behav. 2015;132:33-41.
- 41. Paxinos G $\,$ WC. The rat brain stereotaxic co-ordinates. Massachusetts. 2007.
- 42. Rao X, Huang X, Zhou Z, Lin X. An improvement of the 2[^](-delta delta CT) method for quantitative real-time polymerase chain reaction data analysis. Biol Trace Elem Res. 2013;3:71-85. 43. Tahmasebi Boroujeni S, Naghdi N, Shahbazi M, Farrokhi A, Bagherzadeh F, Kazemnejad A, *et al.* The effect of severe



- zinc deficiency and zinc supplement on spatial learning and memory. Biol Trace Elem Res. 2009;130:48-61.
- 44. Sandoval-Hernandez AG, Buitrago L, Moreno H, Cardona-Gomez GP, Arboleda G. Role of liver X receptor in AD pathophysiology. PloS one. 2015;10:e0145467.
- 45.Mandrekar-Colucci S, Karlo JC, Landreth GE. Mechanisms underlying the rapid peroxisome proliferator-activated receptor-γ-mediated amyloid clearance and reversal of cognitive deficits in a murine model of Alzheimer's disease. J Neurosci. 2012;32:10117-10128.
- 46. Chawla A, Boisvert WA, Lee CH, Laffitte BA, Barak Y, Joseph SB, *et al.* A PPAR gamma-LXR-ABCA1 pathway in macrophages is involved in cholesterol efflux and atherogenesis. Mol cell. 2001;7:161-171.
- 47. Yue L, Mazzone T. Peroxisome proliferator-activated receptor {gamma} stimulation of adipocyte ApoE gene transcription mediated by the liver receptor X pathway. J Biol Chem. 2009;284:10453-10461.
- 48. Seo JB, Moon HM, Kim WS, Lee YS, Jeong HW, Yoo EJ, *et al.* Activated liver X receptors stimulate adipocyte differentiation through induction of peroxisome proliferator-activated receptor y Expression. Mol Cell Biol. 2004;24:3430-3444.
- 49. Tall AR. Cholesterol efflux pathways and other potential mechanisms involved in the athero-protective effect of high density lipoproteins. J Intern Med. 2008;263:256-273.
- 50. Jiang Q, Lee CY, Mandrekar S, Wilkinson B, Cramer P, Zelcer N, *et al.* ApoE promotes the proteolytic degradation of Abeta. Neuron. 2008;58:681-693.
- 51. Lin TW, Shih YH, Chen SJ, Lien CH, Chang CY, Huang TY, *et al.* Running exercise delays neurodegeneration in amygdala and hippocampus of Alzheimer's disease (APP/PS1) transgenic mice. Neurobiol Learn Mem. 2015;118:189-197.
- 52. Nichol KE, Poon WW, Parachikova AI, Cribbs DH, Glabe CG, Cotman CW. Exercise alters the immune profile in Tg2576 Alzheimer mice toward a response coincident with improved cognitive performance and decreased amyloid. J Neuroinflammation. 2008;5:13-27.
- 53. Adlard PA, James SA, Bush AI, Masters CL. Beta-amyloid as

- a molecular therapeutic target in Alzheimer's disease. Drugs Today (Barc). 2009;45:293-304.
- 54. Lazarov O, Robinson J, Tang YP, Hairston IS, Korade-Mirnics Z, Lee VM, *et al.* Environmental enrichment reduces Abeta levels and amyloid deposition in transgenic mice. Cell. 2005;120:701-713.
- 55. Adlard PA, Perreau VM, Pop V, Cotman CW. Voluntary exercise decreases amyloid load in a transgenic model of Alzheimer's disease. J Neurosci. 2005;25:4217-4221.
- 56. Uysal N, Tugyan K, Kayatekin BM, Acikgoz O, Bagriyanik HA, Gonenc S, *et al.* The effects of regular aerobic exercise in adolescent period on hippocampal neuron density, apoptosis and spatial memory. Neurosci Lett. 2005;383:241-245.
- 57. Hori Y, Hashimoto T, Nomoto H, Hyman BT, Iwatsubo T. Role of Apolipoprotein E in beta-Amyloidogenesis: Isoform-specific effects on protofibril to fibril conversion of Abeta in vitro and brain Abeta deposition in vivo. J Biol Chem. 2015;290:15163-15174.
- 58. Strittmatter WJ, Roses AD. Apolipoprotein E and Alzheimer's disease. Annu Rev Neurosci. 1996;19:53-77.
- 59. Mahley RW, Weisgraber KH, Huang Y. Apolipoprotein E4: a causative factor and therapeutic target in neuropathology, including Alzheimer's disease. Proc Natl Acad Sci U S A. 2006;103:5644-5651.
- 60. Roses AD, Saunders AM, Corder EH, Pericak-Vance MA, Han SH, Einstein G, *et al.* Influence of the susceptibility genes apolipoprotein E-epsilon 4 and apolipoprotein E-epsilon 2 on the rate of disease expressivity of late-onset Alzheimer's disease. Arzneimittelforschung 1995;45:413-417.
- 61. Corder EH, Saunders AM, Risch NJ, Strittmatter WJ, Schmechel DE, Gaskell PC, *et al.* Protective effect of apolipoprotein E type 2 allele for late onset Alzheimer disease. Nat genet 1994;7:180-184.
- 62. Tokuda T, Calero M, Matsubara E, Vidal R, Kumar A, Permanne B, *et al.* Lipidation of apolipoprotein E influences its isoform-specific interaction with Alzheimer's amyloid beta peptides. Biochem J 2000;348:359-365.