#### **DE GRUYTER**

## Michaela Zauner\*, Marco Stampanoni and Peter Niemz

# Failure and failure mechanisms of wood during longitudinal compression monitored by synchrotron micro-computed tomography

**Abstract:** The lasting plastic deformation of the cellular elements of beech, fir, and spruce wood under uniaxial compression has been investigated by the combination of *in situ* loading and synchrotron micro-computed tomography. The deformation of singular elements embedded in the tissue and their influence on the deformation lines and surrounding tissue was examined by tomographic reconstructions. An automatic observation of the failure area was applied in the case of the softwoods, which permitted the determination of the densification degree. The development of failure lines differed from the expected pattern by often showing branching. However, the 3D observations confirmed the effects known from 2D examinations in many cases.

**Keywords:** compression, European beech, *in situ* loading, Norway spruce, silver fir, synchrotron radiation microcomputed tomography (SRµCT)

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## Introduction

The deformation of wood with its complex microstructure depends on the properties of its elements. In softwoods, the tube-shaped tracheids transport water and mechanically support the structure in the longitudinal (L) direction. In hardwoods, vessels with large lumens transport water, whereas the thick-walled fibers are the main mechanically stabilizing elements. The kind and amount of the cells and their density lead to differences in the behavior during mechanical load.

Figure 1 illustrates the lasting deformation of cellular elements during L compression (Kucera and Bariska 1982; Cote and Hanna 1983; Reiterer and Stanzl-Tschegg 2001). In the case of thicker-walled elements, the translation of the upper end against the lower end of the structural element leads to buckling (a; Cote and Hanna 1983). However, in the case of thin-walled elements, the upper and lower ends remain aligned in the L direction and the cell wall bulges. Kucera and Bariska (1982) coined the term "telescopic shortening" for this mechanism (b). Other mechanisms may also be observed due to the constrictions and connections of the cells in the surrounding tissue. Additionally, the delamination of the cells (c) can also occur (Mattheck and Kubler 1995). Overall, failure lines are relatively constant for compression in the fiber direction with an angle of  $45^{\circ}$ - $60^{\circ}$  to the L axis (Kucera and Bariska 1982; Cote and Hanna 1983).

To observe failure mechanisms, a variety of nondestructive imaging methods are available (Butterfield and Meylan 1980), such as electron microscopy (EM; Tabarsa and Chui 2000; Sippola and Fruhmann 2002; Fruhmann et al. 2003; Muller et al. 2003; Vasic and Stanzi-Tschegg 2006), light microscopy (LM; Futo 1969; Farruggia and Perre 2000; Dill-Langer et al. 2002), and X-ray micro-computed tomography (X-ray µCT). Both EM and LM necessitate cutting the sample or isolating elements (Eder et al. 2008) due to the low in-depth resolution in the opaque wooden material; thus, the possibilities are limited for the monitoring of the development and evolution of damage mechanisms. This does not apply to X-ray µCT or synchrotron  $\mu$ CT (SR $\mu$ CT), where the complete 3D sample can be recorded with high quality. X-ray µCT has a higher availability and has been used for measuring shrinkage in situ (Taylor et al. 2013), vascular system (Peng et al. 2014), and wax filling (Scholz et al. 2010). SRµCT as an expensive research instrument has a lower availability, but it provides higher recording speeds and higher quality.

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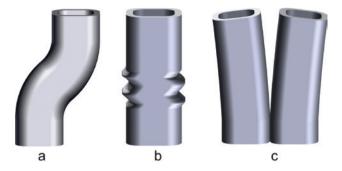
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**Figure 1:** Deformation mechanism during compressive load: (a) buckling, (b) telescopic shortening, and (c) delamination.

Despite the limited access to this instrument, SRµCT has been applied multiple times for the tracing of the vessel network (Hass et al. 2010) or the determination of fiberboard density (Walther and Thoemen 2009).

All methods have to be combined with *in situ* loading to observe the origin and development of lasting visible plastic deformations, which was also done in the case of SR $\mu$ CT for mechanical loading tests (Forsberg et al. 2008; Zauner et al. 2012) and for the observation of the effects of water sorption and desorption (Derome et al. 2011; Patera et al. 2013; Rafsanjani et al. 2013).

The aim of the present study was the *in situ* observation of the compression behavior of European beech (*Fagus sylvatica* L.), white fir (*Abies alba* Mill.), and Norway spruce (*Picea abies* [L.] Karst) wood in the L direction by means of SRµCT. The expectation was that the resulting 3D images may lead to new qualitative and quantitative information concerning the single elements of wood embedded in the surrounding tissue.

## Materials and methods

The samples with a rotationally symmetric hyperboloid shape were made from adult wood without knots or failures (Zauner et al. 2012). As previously described, the method causes small deformations of the first two to four cell walls on the surface, but the cells are not damaged inside. The specimens were produced from white fir (Abies alba Mill.), Norway spruce (Picea abies [L.] Karst.), and European beech (Fagus sylvatica L.). The anatomical differences between fir and spruce are small; spruce may develop nontraumatic resin canals and has heterocellular wood rays with tracheids as bordering elements. A steel negative was used for the production of the hyperboloid samples with a height of 8 mm, a circular cavity of 3 mm radius, and a resulting smallest diameter of 1.35 mm. Three fir, three spruce, and four beech samples were loaded and recorded until failure. A modified version of the setup introduced by Zauner et al. (2012) was used, which allows the measurement of forces and an automatized control of loading and tomographic imaging. The loading procedure was as follows: preload is applied to fix the sample and enable a

tomography of good quality. After the initial tomography, the sample is loaded through moving the crossbeam. After waiting a predetermined time (which increased with the load and was up to 5 min for higher loads) to lessen displacements due to movement, the tomography of the changed state was acquired. This was repeated at least until the compressive strength was reached. The loading velocity was 0.1 mm s<sup>-1</sup>, with a preload of 5 N. The climatic conditions in the beamline chamber were 23°C and 63% relative humidity, and the samples were at equilibrium moisture content.

The SR $\mu$ CT device was used at the Tomographic Microscopy and Coherent Radiology Experiments (TOMCAT) beamline [located at Paul Scherrer Institute (PSI), Villigen, Switzerland; Stampanoni et al. 2006]. Previous experiments at this beamline without loading were described by Trtik et al. (2007) and Mannes et al. (2010). Beam energy: 10 keV; two magnifications with the 4× objective having a digital resolution of 1.85×1.85  $\mu$ m<sup>2</sup> and the 10× objective having a digital resolution of 0.75×0.75  $\mu$ m<sup>2</sup>. The effective lateral resolution was in the first case 4  $\mu$ m, with a resolution of 30 line pairs per millimeter (lp mm<sup>-1</sup>), determined through modulated transfer function (MTF) in the original image. The corresponding data for the 10× objective are 2.3  $\mu$ m with 60 lp mm<sup>-1</sup> (MTF). To obtain the tomographic data, 2001 projections were recorded over 180°, with 50 dark-field and 200 flat images. The exposure time for the 4× objective amounted to 130 and 190 ms for the 10× objective.

After the reconstruction of the 3D tomographic data with the algorithms provided onsite (Marone and Stampanoni 2012), the information was filtered and cleaned. First, the projections were rotated to fit the wood-based coordinate system, and noise in the image with  $4\times$  magnification was removed with a simple threshold. A Markov random field and Gibbs energy approach (measuring the probability of possible configurations) was applied to the images recorded with the  $10\times$  objective, taking the connection of pixels to the neighborhood into account. An iterative (conditional) mode based on that of Besag (1986) and Demirkaya et al. (2009) was better than the simple threshold method (Otsu 1979). A universal outcome for all methods was the identification of the connected regions and the elimination of unconnected noise patterns.

To count pixels representing material and to obtain the area of the sample, repeated morphological operations (e.g., dilate) were executed and the resulting largest region of the material was selected. This allowed an automated identification of the sample. All material pixels were then counted and displayed in relation to the area as "pixel-density."

## **Results and discussion**

All samples were at least loaded until the compressive strength was reached and one or two tomographies were recorded thereafter. The stress-strain curves of three exemplary samples are presented in Figure 2, whereas the matching physical properties and compressive strengths are listed in Table 1. Beech 1 and fir 2 were recorded with the  $4\times$  objective, whereas the other samples were tested with the  $10\times$  objective. The smaller field of view for the  $10\times$  objective resulted in a small portion of the sample surface not being recorded. Expectedly, the observed structure

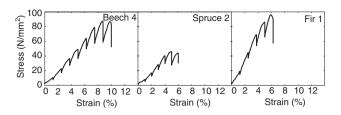


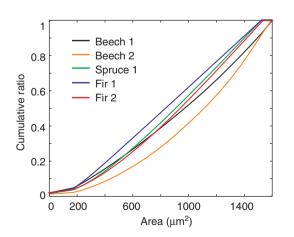
Figure 2: Stress-strain curves of beech 4, spruce 2, and fir 1.

 
 Table 1: Mass, smallest cross-section, and compressive strength of the five samples in Figure 2.

	Beech				Fir
Property	1	4	Spruce 2	1	2
Density (kg m <sup>-3</sup> )	780	846	462	682	538
Area (mm²)	1.94	1.86	1.71	1.62	1.93
Compressive strength (N mm <sup>-2</sup> )	67.7	87.6	33.2	96.4	43.2

and density influences the strength. An example for this is that the strength of fir 1 surpassed that of the beech samples because of the large percentage of latewood (LW) to transition wood. Contrary to beech 4, the stress-strain curve of spruce 2 shows a significant change of the slope between the fourth step and the fifth step.

To be able to compare the porosity of the samples, the area of more than 1000 lumen slices was determined for every specimen listed in Table 1 (Figure 3). The vessels have a large mean area (1600  $\mu$ m<sup>2</sup> for beech 1 and 1500  $\mu$ m<sup>2</sup> for beech 4) compared to the tracheids in fir 2 and spruce 1 with 530 and 440  $\mu$ m<sup>2</sup>, respectively. As expected, fir 1 mainly built from LW has the lowest mean area with 255  $\mu$ m<sup>2</sup>.



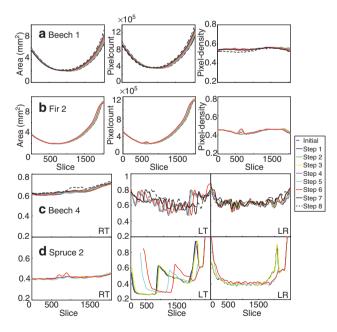
**Figure 3:** Cumulative percentage of distribution of lumen size for vessels in beech and tracheids in softwood.

#### Failure area

To display the change of the material during increasing load, the amount of pixels identified as cell wall substance and the area of the cross-section were calculated for every slice. Figure 4a demonstrates this for beech 1-RT slices. A shortening and upward movement of the visible part of the sample is seen, whereas a compression cannot be directly observed. For softwoods, a compression is visible in the curve modification (Figure 4b). This is especially true for the pixel density. Therefore, an automatic identification of the approximate height of the compressive failure line is possible for softwoods but not for beech.

In Figure 4c and d, the amount of pixels representing material is shown for the RT, LT, and LR plane. LT and LR were calculated around the failure line. Due to the smoothing of the curves and inconsistencies through cleaning process, the values at the beginning and end of the curve are not taken into account. For beech, a change in the LT plane is obvious. The sample is translated and a part of the curve evened out. For the softwood sample, the growth ring is visible. Overall, the differences in the LR slices are found, and for softwood, the slipping of the sample is visible.

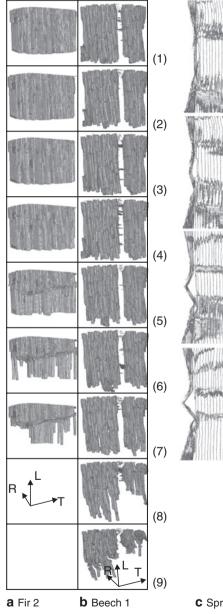
Further, a combination of image processing methods can be used to find a failure line. If the vessels or tracheids are heavily deformed, the lumen is minimized in



**Figure 4:** Area, pixel count, and pixel density (pixel count per area) of beech 1-RT slices and fir 2-RT slices; pixel-density of beech 4 and spruce 2 for sections of the sample in the RT (left row), LT (middle), and LR (right) planes.

Slash-dot line represents the final tomography.

the deformed region. This can be identified through image processing. The disadvantages of this method are that only the part below or above the failure line is identified. Additionally, due to faulty image cleaning tracheids may be closed prematurely and may therefore interrupt the failure line. This procedure was applied to all tomographies of fir 2 and beech 1, resulting in Figure 5. A main



influencing factor in the vessel deformation is the behavior of the thick-walled fibers. Those fibers buckle and deform the vessel structure. Due to the large lumen, the vessels of the hardwood sample stay open for a long time despite deformation. The failure starts to develop relatively late, between the 7<sup>th</sup> picture and the 8<sup>th</sup> picture. Overall, the failure area has an inclination of about  $20^{\circ}-40^{\circ}$  in the T direction and no inclination in the R direction. In contrast, no consistent inclination can be identified in softwood. Rather, a slightly stepwise orientation of the failure is visible starting with Figure 5a.

## Longitudinally oriented structures and failure origin

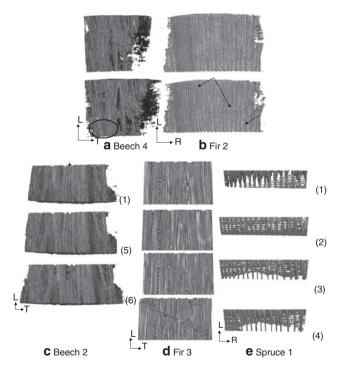
(1)

(2)

(3)

(4)

The behavior of cells depicted in Figure 1 is shown in Figure 6b for fir 2, where multiple failure lines develop. Differences between LW (left side) and earlywood (EW; right side) are found. The failure areas are clearly connected to



C Spruce 3

Figure 5: Identification of failure lines through the isolation of the tissue surrounding the vessels or tracheids running from top to bottom for fir 2 (a) and beech 1 (b).

1, initial tomography; 2–9, tomographies after the first to eighth loading steps, with increasing compressive load. (c) Deformation during compression in an LT slice of a softwood sample (spruce 3) with a resin canal and growth ring.

Figure 6: (a and b) Deformation due to compression and the possible origin shown for the last tomography (top) without visible deformation and the first one (bottom) showing deformation (beech 4 and fir 2). (c) Deformation of beech 2. (1) is the initial tomography, the following the tomographies after the fourth and fifth loading steps. (d and e) Examples of the deformation of singular wood rays in softwoods: fir 3 LW and spruce 1 EW. (1) Initial tomography, (2) last tomography before visible deformation, (3) first deformation, and (4) subsequent tomography.

the surface and originate in regions with cut tracheids (e.g., on the right side). The number and magnitude of deformation of failure lines decrease from the surface to the center of the sample. Similarly, the failure area in beech (left side) is connected to the surface. Due to reconstruction inaccuracies, artifacts are found especially close to the surfaces. The indicated vessel, a thin-walled element, is forced into buckling by the thick-walled fibers surrounding it.

The circles in Figure 5c indicate where EW tracheids are deformed by the connected LW tracheids. The resin canals behave as thin-walled elements (they are built by epithelial cells). The first singular failure line develops in the EW zone (c2) and does not cross the resin canal. During the next loading step, a second failure line, likely connected to the growth-ring border, develops and spans over the resin canal (the upper circle) into the LW region. The resin canal is deformed similar to telescopic shortening. The initial failure line does not develop further, whereas the second failure line further increases in strength, especially surrounding the resin canal. One side of the resin canal shows a stronger deformation probably due to the lower resistance of the thinner cell walls in that region.

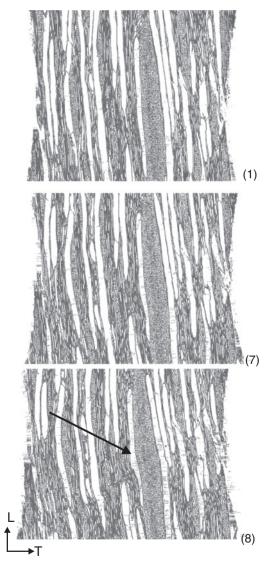
Another detail is also illustrated in Figure 5c. The initial failure area connected to the damaged surface is often not the dominating one when the sample reaches the compressive strength. Instead, a secondary failure area develops later on, especially in structures with a non-uniform distribution of cellular material. With increasing deformations, the originally separated failure lines finally fuse, resulting in a stronger deformation of the areas between them.

Figure 6c shows the deformation of vessels in beech. The arrow indicates a vessel that is deformed during increasing load. In the initial tomography, the cell walls between two vessels are wrinkling. Shortly before the fibers buckle (in state 6), the already existing wrinkles increase in magnitude and the cell material between breaks. Further increasing the load leads to a more pronounced manifestation of this observation.

### Wood rays

Thin wood rays with a thickness of one cell are found in all tested species. The behavior of the rays in softwood is exemplarily shown for EW spruce 1 and LW fir 3 in Figure 6d and e. The original structure is depicted in the initial tomography (1), and the last tomography before visible deformations is illustrated in (2). No visible deformations or cracking were found. After further loading, the wood rays are deformed (3), but the failure line is not influenced. Wood ray cells buckle and are compressed, which indicates that this element does not have a strengthening effect in the L direction. After increasing the load, already deformed cells are further compressed and dislocated. If the failure line crosses the wood ray in the center, the upper and lower parts translate against each other and only adjacent cells are deformed (Figure 6d and e, 4). Only the L-oriented walls buckle. The behavior of wood rays in beech is similar (see Figure 6b). In both LW in softwoods and beech, a uniform failure line with an angle of  $20^{\circ}$ – $40^{\circ}$  of LW is found.

Broad rays seem to significantly dampen the failure lines. This is illustrated through Figure 7. In the time between Figure 7 (7 and 8), the tissue buckles and the



**Figure 7:** Deformation of broad wood rays in beech 1: (1) initial tomography, (7) after the sixth loading step, and (8) after the seventh loading step.

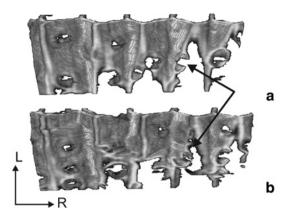


Figure 8: Deformation of bordered pits of spruce 2.

broad wood ray is deformed. Only the vessel adjacent to the ray on the opposing side of the original failure line shows slight buckling. Due to the low quality of the tomographic reconstruction of the broad wood rays and therefore missing information about the cell walls, no detailed interpretation is possible.

Bordered pits found along the tracheids in softwoods are both deformed by crossing failure lines or circumvented by the failure line. An example is presented in Figure 8. After loading the sample, some pits are deformed where a failure line has crossed them.

## Conclusion

In the case of SR $\mu$ CT of softwoods, the height of the failure area and an estimation of the shape can be determined through an automated approach of material identification. This results in good estimates, especially for mixed EW and LW structures. Due to the different failure mechanisms encountered in beech and LW in the two softwoods, this approach is not optimal for those structures. An approach of displaying the vessel structure or tracheids leads to a better estimate of the failure line.

Often, a secondary failure line develops, whereas the original failure does not further magnify. The failure line for LW in the softwoods and beech is relatively constant with an angle of  $20^{\circ}-40^{\circ}$  to the horizontal and does not show deviating tendencies. In contrast, other structures in softwood did not show a continuous failure line. Singular wood rays do not contribute to the strength but are equally deformed with the surrounding tissue. Bordered pits are not exclusively circumvented as was observed in previous tensile load experiments but may be deformed in the failure areas.

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