



APPLICATION NOTE

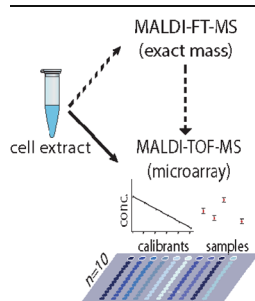
Quantification of Saquinavir from Lysates of Peripheral Blood Mononuclear Cells Using Microarrays and Standard MALDI-TOF-MS

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Abstract. Drug monitoring is usually performed by liquid chromatography coupled with optical detection or electrospray ionization mass spectrometry. More recently, matrix-assisted laser desorption/ionization (MALDI) in combination with triple quadrupole or Fourier-transform (FT) mass analyzers has also been reported to allow accurate quantification. Here, we present a strategy that employs standard MALDI time-of-flight (TOF) mass spectrometry (MS) for the sensitive and accurate quantification of saquinavir from an extract of blood peripheral mononuclear cells. Unambiguous identification of saquinavir in the mass spectra was possible because of using internal mass calibration and by an overall low chemical noise in the low mass range. Exact mass determination of the constant background peaks of the cell extract, which were

used for recalibration, was performed by an initial MALDI-FT-MS analysis. Fast and multiplexed sample analysis was enabled by microarray technology, which provided 10 replicates in the lower nL range for each sample in parallel lanes on a chip. In order to validate the method, we employed various statistical tests, such as confidence intervals for linear regressions, three quality control samples, and inverse confidence limits of the estimated concentration ratios.

Keywords: Quantitative MALDI-MS, Saquinavir, Cell lysate analysis, Peripheral blood mononuclear cells, Microarrays for mass spectrometry (MAMS), Antiretroviral drugs

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Introduction

The limiting factors in the analysis of antiretroviral drugs are the minute amounts of blood peripheral mononuclear cells (PBMCs) from patients that are available, as well as the complex sample matrix from cell lysates. Therefore, highly sensitive as well as selective detection methods are required. “Gold standards” rely on “high performance” liquid chromatography (HPLC) in combination with UV detection or electrospray mass spectrometry (ESI-MS) [1–3]. However, sample throughput is limited because of the long cycle times of chromatographic runs. By contrast, using MALDI-MS, the analysis of one sample (i.e., one spot) merely takes a few seconds and, therefore, high-throughput analysis is possible. MALDI-MS also is less

demanding with respect to sample preparation or purification and allows preferred ionization of selected classes of compounds depending on the used MALDI matrix. On the other hand, MALDI-MS is usually not the method of choice for quantitative applications [4–6]. Nevertheless, in recent years improvements in sample preparation by introducing modified sample targets or protocols, have efficiently addressed this problem. Furthermore, MALDI instrumentation using triple quadrupole and FT-MS analyzers is increasingly used for quantitative applications and has also been mentioned in clinical drug monitoring [7–15].

Here, we report on a method to quantify saquinavir from a cell extract using standard MALDI-TOF-MS. We performed an initial high resolution mass scan of the cell background using a MALDI-FT-MS. Sample analysis was carried out by MALDI-TOF-MS, where spectra were recalibrated using background peaks from reserpine and the cell extract. The overall noise level (e.g., obtained by matrix clusters or fragments) was found to be quite low, although α -cyano-4-hydroxycinnamic acid (CHCA) was used as the MALDI matrix (Supplementary Information Figure 1).

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Importantly, the method presented relies on the assistance of microarray plates, which feature multiple lanes of small hydrophilic spots. The plates support the analysis of a large number of replicates, which in turn increases confidence. To evaluate the performance of our method, we plotted the 95% confidence interval for the calibration series above and below the curve fitted by linear regression. The correlation coefficient (r^2) was evaluated by plotting all data points measured for the calibration, instead of using average values.

Experimental

Chemicals

Alpha-cyano-4-hydroxycinnamic acid (CHCA), saquinavir, atazanavir, indinavir, nelfinavir, and reserpine were purchased from Sigma Aldrich (Buchs, Switzerland).

Microarrays and Sliding Device

Microarray targets (MAMS, Microarrays for Mass Spectrometry, Supplementary Information Figure 2) were produced as previously described. In brief, the targets were made of ITO-coated microscope glass slides with an omniphobic polysilazane coating on top. Lanes of microarray spots with a diameter of 400 or 500 μm and a cavity of 35 μm were prepared by picosecond laser ablation [4, 16]. ITO slides were mounted onto a standard AB SCIEX sample target carrier using a metal mask that was prepared in-house. The sliding device was produced in-house and is made of stainless steel. It consists of an approximately 10 cm long edge and supporting bars on a guiding rail. The edge is positioned sufficiently low so as to contact the series of droplets that are located in the deposition spots. Then, the slider drags the droplets from the deposition spot over the lanes. Each time the droplet passes over a hydrophilic spot, a nanoliter aliquot is retained, thus generating multiple replicates from one droplet. In total, many replicates can be produced by one sliding motion (the microarray target used herein generated 12 replicates per sample).

Cell Extract

PBMCs were purchased from ZenBio and lysed as described by van Kampen et al. [11]. In brief, 1 million cells were lysed in 100 μL of 50% ethanol. Lysis was performed without agitation over night at 5°C. Cells were centrifuged at 14,000 g at 5°C for 10 min, and the supernatant was collected and brought to 20% EtOH in H₂O. The cell lysate was further purified using OasisHLB (Waters, London, UK) extraction columns. The eluate from the extraction columns was dried using a speed-vac, redissolved in 10 μL spike solution (50% acetonitrile in H₂O), internal standard was added, and the samples were mixed with MALDI matrix solution at a ratio of 1 to 2 (v/v).

Sample Aliquoting

A mixture of 4.5 μL was pipetted onto the “deposition reservoir” of each lane and aliquoted using a metal slider as described in the

materials and methods section as well as in Pabst et al. 2013 [4]. CHCA was used as a MALDI matrix at a concentration of 4 mg per mL in 60% acetonitrile and 40% H₂O, (v/v) with 0.1% TFA. The slider and microarray target were cooled to 4°C for at least 15 min before aliquoting. This was found to be beneficial to prevent the samples of the nanoliter volumes from drying too fast. Aliquoting was carried out at the laboratory bench, maintaining a constant speed of the slider.

MALDI Mass Spectrometry

MALDI mass spectra were recorded on a commercial instrument (AB SCIEX TOF/TOF™ 5800 System, Darmstadt, Germany). For MALDI-MS analysis, the whole spot was scanned with a circular scan pattern of 120 individual positions and 20 shots per position (2400 shots in total per spot) at a repetition rate of 1000 Hz, detected in positive reflectron mode. Laser energy was set to 3800 (arbitrary units, as defined by the AB SCIEX software). Spectra were averaged and relative signal areas calculated by dividing the signal area of saquinavir by that of reserpine. The concentration ratios were then used for generating the calibration curve. Internal mass calibration of the MALDI-MS instrument was carried out using peaks from reserpine and constant background signals. Exact masses were obtained by measuring one reference sample with internal calibration on a Bruker Solarix, as described in Steinhoff et al. [17]. All 12 replicates of each data point were measured. The first and last spots of each row were used for performing MS/MS experiments and for tuning the MALDI-TOF-MS instrument.

Internal Standards and Generation of Calibration Curves

As an internal standard for saquinavir, we used reserpine; for atazanavir and nelfinavir, we used indinavir, and for indinavir, nelfinavir was chosen. Five-point calibration curves of standard solutions (in 50% acetonitrile) were acquired for all antiretroviral drugs between concentrations of 15.6 and 2000 ng per mL.

Saquinavir was also quantified from an extract of blood peripheral mononuclear cells, using reserpine as an internal standard. The estimation of the concentration ratios of the quality control samples for saquinavir was carried out using an external calibration method. Ten μL of standard solutions with concentrations of 15.6, 125, 500, 1500, 3000, and 4000 ng/mL with a constant concentration of 750 ng/mL reserpine were spiked to a cell extract of 1×10^6 cells. If we assume a volume of 0.4 pL per cell [18], these concentrations would translate into PBMC concentrations of 0.39, 3.125, 12.5, 37.5, 75, and 100 ng/mL. As estimated in Pabst et al. 2013 [4], each array spot contains approximately 10 nL. Therefore, the lowest concentration (15.6 ng per mL) equals an absolute amount of approximately 250 attomol saquinavir per array spot.

Data Analysis

Data extraction from MALDI-TOF analysis results was carried out using the analysis tool of the TOF/TOFseries Explorer

Table 1. Relative signal intensities of saquinavir for three quality control samples

A_{rel}	Estimated conc.	St. dev. (RSD %)	Nominal conc.	Accuracy	95% Inverse	Mass accuracy (ppm)	Signal-to-noise ratio
$(A_{\text{Saq}}/A_{\text{Res}})$	ng/mL	($n=10$)	ng/mL	% error	confidence limits	saquinavir [M+H] ⁺ =671.4	saquinavir [M+H] ⁺ =671.4
2.010	53.28	2.4 (4.54%)	50.0	6.5	0.97	2.57	3166
1.027	27.25	0.84 (3.1%)	25.0	9.0	0.97	2.97	2094
0.046	1.29	0.04 (3.29%)	1.5	17.1	1.03	9.8	485

Estimated concentrations were within 6.5% and 17.1% relative error of the nominal values. Standard deviations of the repeated measurements are relatively low and confidence limits of the estimated sample concentration appear narrow. Mass accuracy was found within 10 ppm even for the lowest concentration

software from AB SCIEX. Calculation of the 95% confidence interval, r^2 , curve of the linear regression, and inverse confidence limits for quality control samples were all done using Microsoft Excel. All points (i.e., the whole data cloud) were plotted for data analysis as opposed to averaging all points for a given concentration. The formulae for inverse confidence limits were taken from Szajli et al. [19] and Eisenhart et al. [20]. Outlier tests were performed using a Q-test, for which up to one data point was allowed to be treated as an outlier per 10 data points.

Results and Discussion

In a first set of experiments, we evaluated the sensitivity and usefulness of our system for the analysis of selected antiretroviral drugs (saquinavir, atazanavir, indinavir, and nelfinavir) using CHCA (Supplementary Information Figure 3). By using the microarray, we obtained quite low levels of chemical noise in the low m/z range, which was vital for the unambiguous assignment of analyte signals that are close to the limit of quantification (Fig. 1, Supplementary Information Figure 1). We were able to obtain calibration curves with low standard deviations and r^2 values greater than 0.98 for all antiretroviral drugs (Supplementary Information Figure 3).

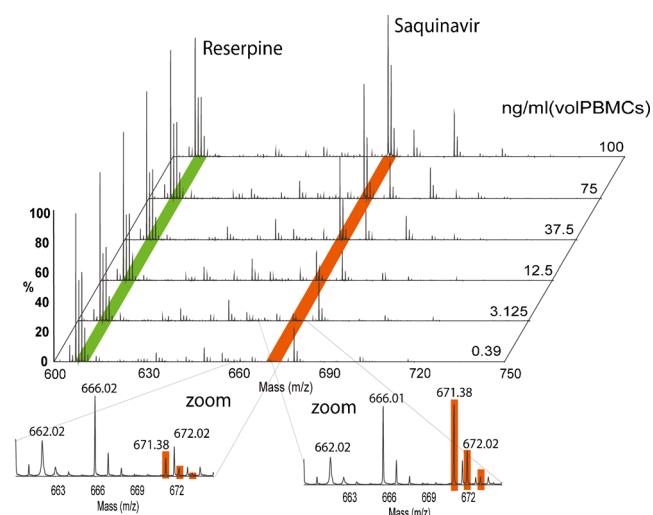


Figure 1. Mass spectra series from 0.39 ng/mL and 100 ng/mL (volume PBMCs) plotted. A zoom of the two lowest concentrations is presented below. Saquinavir is highlighted with orange bars. Reserpine, which was used as internal standard, is highlighted with a green bar

We finally evaluated the practicability of employing standard MALDI-TOF-MS for quantifying saquinavir in the complex matrix of a cell extract. In a single pre-scan, we determined the accurate masses of the constant background peaks by MALDI-FT-MS once and chose four peaks close to the mass of saquinavir $[(M+H)^+=679.1]$ for further internal calibration of the MALDI-TOF-MS spectra (Supplementary Information Figure 4). Therefore, it was not necessary to add other internal standards, which might have resulted in a loss of sensitivity because of competitive ionization. Saquinavir was identified within a 10 ppm mass window, even at the lowest concentration (Supplementary Information Table 1). Mass resolution for the MALDI-TOF-MS instrument was found to be about 10,000 (FWHM). Moreover, the identity of the analytes was confirmed by additional MS/MS experiments.

Average standard deviations, mass accuracy, and signal-to-noise ratios for the data points of the calibration curve can be found in Supplementary Information Table 1). Alongside the calibration curve, we analyzed three quality-control samples for saquinavir. For this, we obtained relative standard deviations for the replicate measurements of 4.5%, 3.1%, and 3.3% for the lowest concentration (1.5 ng/mL). The accuracy for the independent quality control samples were further found to be

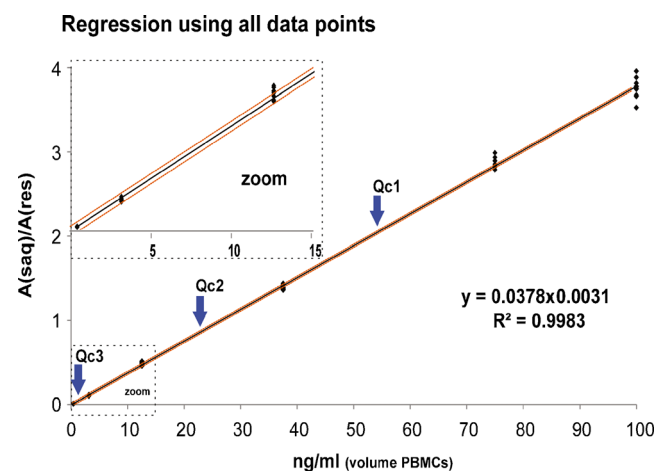


Figure 2. Calibration curve obtained after plotting the whole data series for saquinavir spiked to extracts of PBMCs ($n = 10$ per data point). The series was linear between 0.39 ng/mL and 100 ng/mL (volume of PBMCs expressed in mL); 95% confidence intervals for the curve of best fit are outlined as orange lines. The r^2 value was calculated considering the whole data series in the plot ($n = 60$) in graph. Points for quality control samples (1–3) are marked with blue arrows

6.5% (53.28 ± 2.4 ng/mL), 9% (27.25 ± 0.84 ng/mL), and 17% (1.29 ± 0.04 ng/mL) (Table 1 and Supplementary Information Figure 5). As expected for a confident analysis, the confidence intervals of the curve of best fit and calculated 95% inverse confidence limits according, to Szajli et al. [19] and Eisenhart et al. [20], appeared very narrow (Fig. 2, Supplementary Information Figure 6). We attribute the high data quality to the large number of replicates ($n=60$ for the calibration curve, plus 10 for the quality control sample), and the overall low standard deviations of the MALDI-MS analysis (on average 3.9%, Supplementary Information Table 1). In the theoretical case of using a much lower number of replicates (e.g., $n=18$ for the calibration curve if three replicates per data point are acquired), both confidence limits would increase drastically, even if standard deviations and the correlation of the regression curve might be acceptable. Confidence limits, therefore, allow a reliable decision to be made on the quality of the results of the analysis [19]. The whole chip was aliquoted within a few seconds (six calibration points and three quality control samples). Furthermore, we were able to acquire 10 replicates in approximately 1.5 min, which is much faster than what can be reached by methods involving chromatographic separations.

Conclusions

By combining a suitable sample preparation with microarray technology and internal mass calibration, we demonstrated that accurate, precise, and confident quantification of antiretroviral drugs can be achieved by using standard MALDI-TOF-MS. A single pre-scan of the accurate masses as well as a low background signal noise in the lower mass range allowed ambiguous peak assignment and quantification from a complex matrix (Figs. 1 and 2).

Acknowledgments

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M.P., S.F., R.K., K.J., and R.Z. are developing a commercial product based on the microarray for mass spectrometry technology.

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