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Dendritic Cell Cancer Therapy: Vaccinating the Right Patient at the Right Time

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Immune checkpoint inhibitors propelled the field of oncology with clinical responses in many different tumor types. Superior overall survival over chemotherapy has been reported in various metastatic cancers. Furthermore, prolonged disease-free and overall survival have been reported in the adjuvant treatment of stage III melanoma. Unfortunately, a substantial portion of patients do not obtain a durable response. Therefore, additional strategies for the treatment of cancer are still warranted. One of the numerous options is dendritic cell vaccination, which employs the central role of dendritic cells in activating the innate and adaptive immune system. Over the years, dendritic cell vaccination was shown to be able to induce an immunologic response, to increase the number of tumor infiltrating lymphocytes and to provide overall survival benefit for at least a selection of patients in phase II studies. However, with the success of immune checkpoint inhibition in several malignancies and considering the plethora of other treatment modalities being developed, it is of utmost importance to delineate the position of dendritic cell therapy in the treatment landscape of cancer. In this review, we address some key questions regarding the integration of dendritic cell vaccination in future cancer treatment paradigms.

Keywords: dendritic cell, vaccination, immunotherapy, checkpoint inhibitor, cancer, adjuvant

INTRODUCTION

Since William Coley made his early contributions to the study of cancer immunotherapy in the 1890s, harnessing the capabilities of the immune system to eliminate cancer cells remained a long-sought dream (1). In the last decade, efforts to realize this dream were finally rewarded with the introduction of immune checkpoint inhibitors (ICI). ICI showed the feasibility of immunotherapy and revolutionized the treatment of cancer. The success of ICI spurred a considerable amount of research activity into the field of immunotherapy. Despite its resounding success, ICI still have two important limitations: they are associated with significant (immune-related) toxicity and a portion of patients does not respond (2–7). Immunotherapy however, encompasses more than ICI alone. Dendritic cell (DC) vaccination is an alternative form of immunotherapy and is a prime candidate to enrich the treatment possibilities for cancer. Considering the fact that the field of immunotherapy is a fast-moving field, it is of utmost importance to delineate the position of DC vaccines in the therapeutic landscape of cancer. In this review, we will explore some important questions regarding this position, with the focus on four malignancies (glioblastoma, melanoma, prostate cancer, and renal cell carcinoma) in which phase III trials with DC vaccines have been performed or are ongoing.

The Evolving Field of Immune Checkpoint Inhibition

Currently, the clinical application of immunotherapy is mainly defined by ICI. ICI target immune checkpoint molecules such as CTLA-4, PD-L1, and PD-1. These molecules have immune response inhibiting functions and are involved in the prevention of autoimmunity and the maintenance of peripheral tolerance. It is well known that tumor cells are able to upregulate the expression of checkpoint molecules, leading to anergy of cytotoxic T-cells in the tumor microenvironment. CTLA-4, PD-L1, and PD-1 have distinct functions; CTLA-4 exerts its inhibitory functions on the initial T-cell activation whereas PD-1 and PD-L1 have roles in the inhibition of the effector functions of T-cells (8, 9). ICI antagonize these molecules and thereby aim to augment the anti-cancer immune response.

In 2010, ipilimumab (a monoclonal antibody targeting CTLA-4) was the first immunotherapeutic agent providing clinical benefit in cancer patients, extending median overall survival (OS) to 10 months (compared to 6.4 months for the control group receiving a gp100 peptide vaccine) in metastatic melanoma (3). With an overall response rate (ORR) of ~10–20%, ipilimumab was a great improvement compared to the standard of care at the time, but it still offers clinical benefit in only a portion of melanoma patients (10, 11). However, in a substantial portion of responding patients, clinical benefit is durable (5). In 2014, two monoclonal antibodies (pembrolizumab and nivolumab) targeting the PD-1 pathway were also approved for the treatment of metastatic melanoma. Compared to ipilimumab, anti-PD-1 inhibition achieves a higher ORR of ~40% (4, 5, 12, 13).

After these landmark studies, research into ICI accelerated. With the addition of PD-L1 targeting agents avelumab, atezolimumab, and durvalumab, the field of ICI now encompasses six FDA and EMA-approved monoclonal antibodies (mAb) (14–16). Most of these ICI are approved for the treatment of multiple malignancies (Table 1). The number of approved indications of these mAb is likely to grow as they are currently tested in a large number of additional malignancies (17).

Besides PD-1, PD-L1 and CTLA-4, other checkpoint molecules (such as TIM-3 and LAG-3) have shown to inhibit the anti-cancer immune response (18). Several mAb targeting these alternative checkpoint molecules are in various stages of clinical investigation. Therefore, it is expected that the number of clinically available mAb will be further expanded (17). In addition to the treatment of metastatic disease, research is moving toward the application of ICI in the adjuvant treatment of cancer. For example, adjuvant ipilimumab, nivolumab, and pembrolizumab after surgically resected stage III melanoma recently have shown to improve progression-free survival (PFS) and in case of adjuvant ipilimumab, an prolonged OS was seen (19–21).

ICI come with a different toxicity profile compared to other anti-cancer therapeutics, caused by specific immune-related side effects. Monotherapy with anti-PD-1 mAb and anti-CTLA-4 mAb are associated with 10–16% and 30–40% grade 3 or 4 adverse events, respectively (3, 5, 6, 11, 22). In contrast, DC

TABLE 1 | Indications of the six currently approved monoclonal antibodies in the treatment of cancer (as of May 2018).

Monoclonal antibody	Target	FDA/EMA-approved indications
Ipilimumab	CTLA-4	Melanoma
Nivolumab	PD-1	Melanoma, NSCLC, RCC, urothelial carcinoma, MSI-high/dMMR CRC, HCC, Hodgkin's lymphoma, HNSCC
Pembrolizumab	PD-1	Melanoma, NSCLC, HNSCC, urothelial carcinoma, Hodgkin's lymphoma, MSI-high cancer, gastric/gastroesophageal cancer
Avelumab	PD-L1	Merkel cell carcinoma, urothelial carcinoma
Atezolimumab	PD-L1	Urothelial carcinoma, NSCLC
Durvalumab	PD-L1	Urothelial carcinoma, NSCLC
Combined treatment with ipilimumab and nivolumab	CTLA-4/PD-1	Melanoma, RCC

CRC, colorectal cancer; CTLA-4, cytotoxic T-lymphocyte-associated protein 4; HCC, hepatocellular carcinoma; HNSCC, head and neck squamous cell carcinoma; dMMR, DNA mismatch repair deficiency; MSI, microsatellite instability; NSCLC, non-small-cell lung carcinoma; PD-1, programmed cell death protein; PD-L1, programmed death-ligand 1; RCC, renal cell carcinoma.

vaccination is associated with little toxicity as grade 3 or 4 adverse events are very uncommon (23–25). In addition, the application of DC vaccination might further improve response rates on ICI.

Dendritic Cell Vaccination

Since their discovery by Steinman in 1973, it became clear that DC are antigen-presenting cells crucial in activating the adaptive immune system (26). DC are spread throughout the body, constantly monitoring their surroundings for antigens and danger signals. Once stimulated by an activating stimulus, they undergo maturation and migrate to lymphoid organs where they activate several effector cells of the immune system, primarily T-cells and B-cells (27).

Through this process, DC are vital for immunosurveillance. Immunosurveillance signifies the crucial role of the immune system in the detection and elimination of both pathogens and cancer cells. However, the development of malignancy is an indolent process in its early stages, therefore, immunosurveillance occasionally fails. At an early stage, tumors sometimes silence an initiated immune response or fail to express the “danger signals” necessary for the activation of the immune system. When the process of immunosurveillance fails, one of the hurdles for the outgrowth of cancer cells is omitted. DC vaccination aims to correct this failure by reversing the ignorance of the immune system to malignant cells. To achieve this, DC are stimulated *ex vivo* with danger signals and loaded with tumor-specific antigen(s) on their major histocompatibility complex molecules with the intent of activating antigen-specific T-cells which selectively eliminate antigen-bearing cancer cells (Figure 1). The majority of research groups, including our own, employ treatment schemes with multiple administrations of DC vaccine to induce immunological memory (28).

DC vaccines are produced following some basic principles (Figure 2). Natural circulating DC or monocytes are isolated

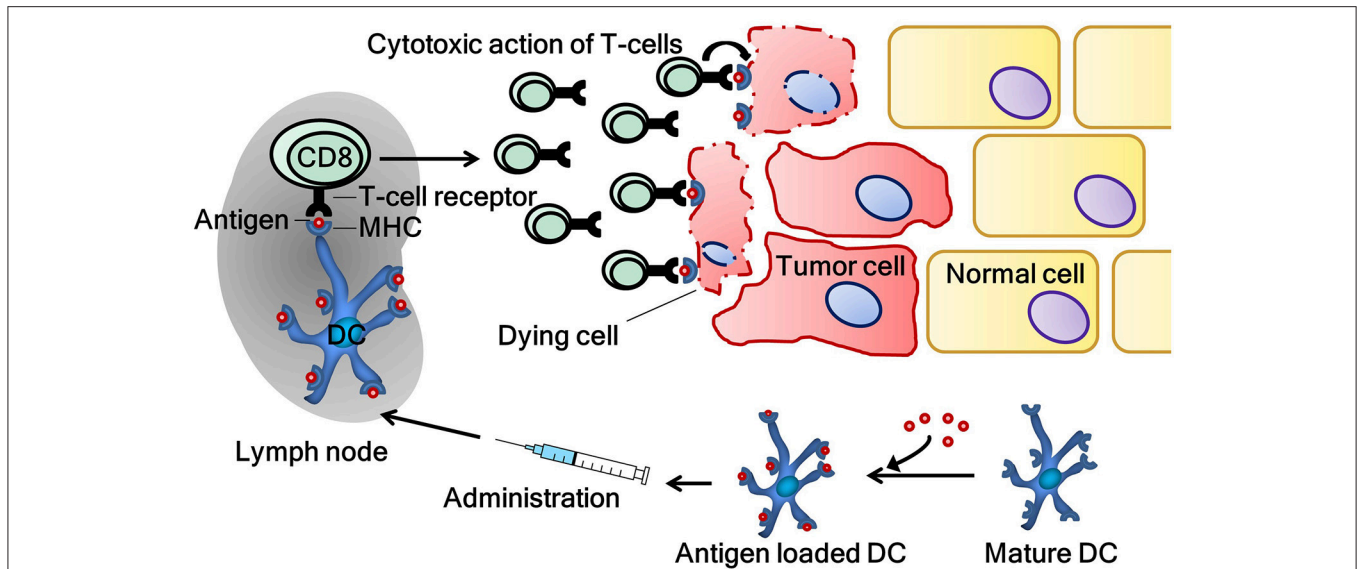


FIGURE 1 | The induction of a tumor-specific immune response by dendritic cell vaccination. Tumor antigen-specific T-cells are activated by dendritic cells, which are *ex vivo* loaded with tumor antigen(s). Activated T-cells subsequently patrol the body in search of their respective antigen. When their target is found, T-cells exert their cytotoxic functions on cancer cells. CD8, cluster of differentiation 8 (cytotoxic T-cell); DC, dendritic cell; MHC, major histocompatibility complex.

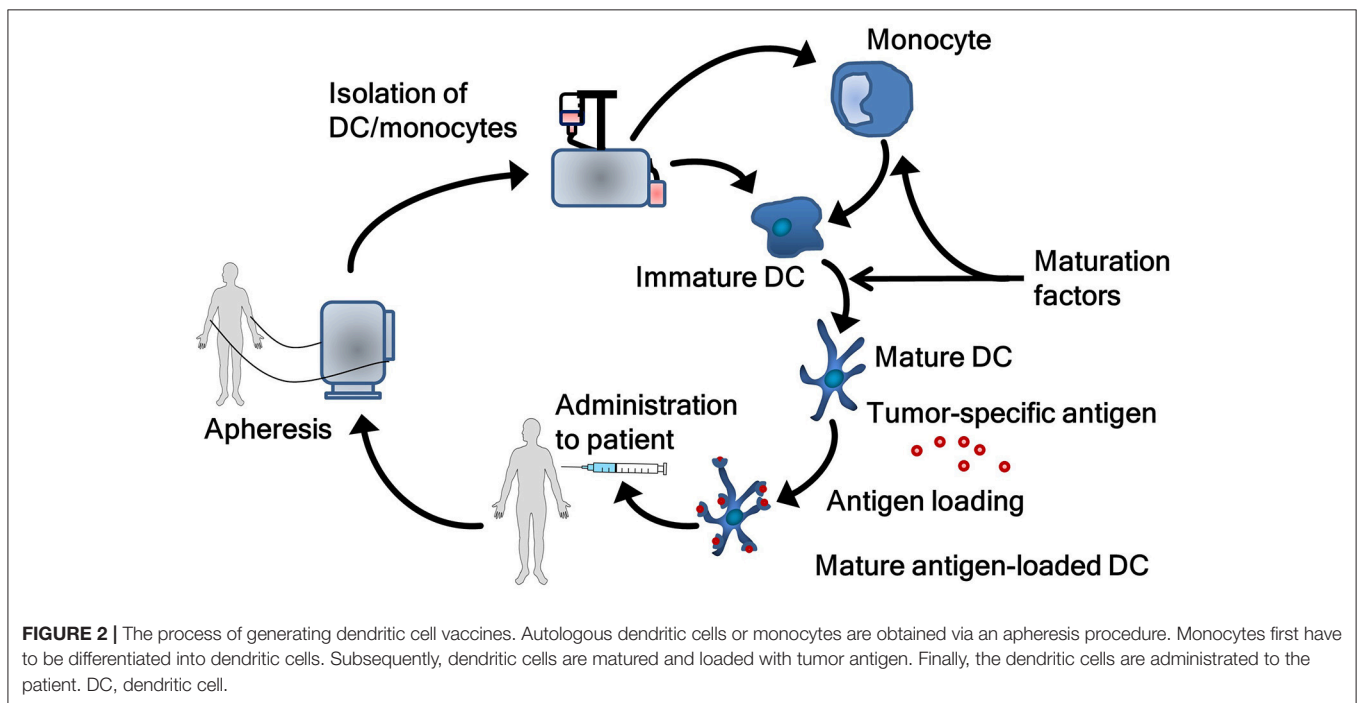


FIGURE 2 | The process of generating dendritic cell vaccines. Autologous dendritic cells or monocytes are obtained via an apheresis procedure. Monocytes first have to be differentiated into dendritic cells. Subsequently, dendritic cells are matured and loaded with tumor antigen. Finally, the dendritic cells are administered to the patient. DC, dendritic cell.

from autologous peripheral blood mononuclear cells obtained by apheresis. In case of monocytes, *ex vivo* differentiation into DC are required. Both natural circulating DC and monocyte-derived DC are matured as this is essential for effective T-cell activation. Maturation is associated with functional and morphological changes in DC. Following maturation, DC show enhanced expression of major histocompatibility complexes I and II, co-stimulatory molecules and increased capability of cytokine

production. These processes are vital, as not or incompletely matured DC can induce tolerance rather than immunity (29). During the process of vaccine manufacturing, DC are loaded with relevant tumor antigen(s) to induce a tumor-specific immune response in the patient. As with the other steps in the process of manufacturing DC, several methods to load DC with antigen exist (30). After quality control, vaccines are administered to the patient.

Despite these basic principles, protocols describing the specific details of DC vaccination manufacturing in trials vary widely. Differences in these protocols cover all aspects of DC vaccination including culture methods, the usage of DC subsets, maturation methods, antigen loading techniques, used antigens and the route of administration. Especially, the subset of DC used, the method of maturation and the choice of antigen(s) are subject of intense research. For example, several groups, including our own, use natural circulating DC instead of monocyte-derived DC. Natural circulating DC do not require extensive culturing which is believed to retain their functionality. Different maturation techniques are also being explored, such as the use of toll-like receptor ligands or electroporation with mRNA-encoding proteins that induce DC maturation (31, 32). Another exciting recent development is the use of neoantigens, which are newly, formed antigens generated from tumor-specific mutated genes, for loading on DC (33). Finally, a more recent development is the recognition that DC, in addition to immune-activating properties, can acquire effector functions (so called killer-DC) following triggering with several differentiating and maturing agents such as interferon (IFN) or lipopolysaccharide (34). Despite these developments, addressing the differences in the generation and production of DC vaccines extensively is beyond the scope of this review.

Regardless of the precise protocol employed, DC vaccination is associated with a very favorable toxicity profile. The majority of side effects reported in various clinical trials were short-lived grade 1 or 2 adverse events, consisting of self-limiting flu like symptoms, fever and local injection site reactions. Treatment-related grade 3 or 4 adverse events following DC vaccination as standalone therapy are uncommon (23, 24).

The goal of DC vaccination is to kill tumor cells by the generation of functional antigen-specific T-cells (23). Despite the challenges associated with measuring the immunological effect of DC vaccination, immunological endpoints are reported in a substantial portion of phase I/II clinical DC vaccination trials using various methods. Several studies even report the generation of antigen-specific T-cells to be positively correlated with survival, strengthening the belief that DC vaccination can result in clinical benefit (25, 35, 36).

Besides the generation of T-cells, intense research is ongoing to find biomarkers, not only for DC vaccination but for immunotherapy in general. Considering ICI treatment, research into predictive biomarkers has revealed several biomarkers predictive for response on ICI (such as mutational burden, PD-L1 expression, and others) (37, 38). Similarly, an example of a predictive biomarker prior to the start of therapy correlated with clinical outcome after DC vaccination is the immune landscape of tumors (39). Up until now, however, biomarkers cannot reliably guide treatment decisions in the clinic for neither ICI nor other forms of immunotherapy, probably owing to the fact that a functional immune response is a complex and multi-step process (40).

The Role of ICI and DC Vaccination in Metastatic Disease

Response rates to DC vaccination vary among cancer types with most studies showing response rates between 10 and 15%

(24). Most clinical studies concerning DC vaccination were performed in patients with metastatic disease. Although head-to-head comparisons are not available, ICI achieve superior clinical benefit compared to DC vaccination in most malignancies. In particular for metastatic melanoma and metastatic renal cell carcinoma (RCC), ICI compare favorably in terms of response rates (approximate ORR on anti-PD-1 mAb in RCC: 25%; in melanoma: 40 and 58% when combined with anti-CTLA-4 mAb) (4, 10, 11, 41). ORR in RCC and melanoma patients after treatment with DC vaccines is less, 12 and 9%, respectively (24). Even more important, whereas overall survival benefit for patients with metastatic RCC and metastatic melanoma after ICI treatment is well established, the OS gain for these patients after DC vaccination is less clear (3, 11, 24, 41).

The immunotherapeutic landscape of metastatic castration-resistant prostate cancer (mCRPC) is very different from that of metastatic RCC and metastatic melanoma. Two phase III trials investigating ipilimumab showed, both in pre-docetaxel and post-docetaxel setting, no improvement in OS compared to their control groups (42, 43). Pembrolizumab has shown clinical activity in patients with any type of cancer bearing DNA mismatch repair deficiency (dMMR) and/or microsatellite instability. Individual reports of clinical benefit on anti-PD-1 mAb for patients with dMMR prostate cancer do exist. Unfortunately, dMMR is present in only about 5% of mCRPC patients (44–47). Similar to patients with dMMR, ICI possibly provide benefit in other subgroups of mCRPC patients. For example, nivolumab combined with ipilimumab was tested on patients with an ARV7 mutation which predisposes for a more aggressive form of prostate cancer. In this study, 4 out of 15 patients showed clinical benefit (47). In addition, pembrolizumab has shown some efficacy in a group of patients who progressed after enzalutamide treatment. In a trial of 20 patients, 11 had a partial response or stable disease (45). These patients might be more susceptible to PD-1 antibodies, as PD-1 was shown to be upregulated on DC in patients progressing after enzalutamide (46). After the failure of ipilimumab in prostate cancer patients, a delay in designing new studies with ICI occurred. Currently, ~35 clinical studies with ICI are enlisted for prostate cancer, usually as combination therapies.

Notably, sipuleucel-T gained approval for the treatment of asymptomatic or minimal symptomatic mCRPC. Sipuleucel-T is manufactured from autologous mononuclear cells obtained via apheresis. These cells are incubated with PA2024, a fusion protein of the tumor antigen prostatic acid phosphatase (PAP) and granulocyte-macrophage colony-stimulating factor (GM-CSF). As DC are not specifically isolated from the apheresis product and the end product contains a variety of cells, sipuleucel-T should strictly speaking not be regarded as a pure DC vaccine. Despite this, sipuleucel-T is generally addressed as a DC based-vaccine and is considered to be the first DC-based therapy approved by the FDA. The approval of sipuleucel-T followed the results of a phase III trial including 512 mCRPC patients. The median survival was prolonged with 4 months compared to placebo (48). Another smaller phase III study confirmed these favorable results (49).

Initial enthusiasm about sipuleucel-T has somewhat subsided in recent years since labor intensive production resulted in

a highly priced cellular product (around \$125,000). At the moment, sipuleucel-T is only available in the USA as market authorization was not granted by the EMA. Recently, a Chinese conglomerate (Sanpower) acquired Dendreon (producer of sipuleucel-T) for over \$800 million with the intention to extend the market to Asia. Sipuleucel-T enhanced immune responses toward its antigen (PAP/PA2024). A PAP/PA2024-specific immune response (which is defined as the generation of antigen-specific antibodies, antigen-specific T-cell activation and/or antigen-specific T-cell proliferation) was seen in 79% of patients. The immune responses correlated with OS and could be beneficial for the response on subsequent or concomitant immunotherapeutics, a paradigm which will be detailed in the final chapter of this review (50).

In conclusion, in metastatic malignancies such as non-small-cell lung cancer, melanoma, urothelial cancer and RCC, where ICI are particularly effective, it is unlikely DC vaccination will gain a role as monotherapy in widespread metastatic disease due to its less established clinical benefit.

Rationale for DC Vaccination in the Adjuvant Treatment of Cancer

Besides the application of anti-cancer therapeutics in the treatment of metastatic disease, the adjuvant treatment of patients after surgery of local disease is also common practice in oncology. Surgical resection with curative intent aims to excise all tumor burden. However, depending on the type of malignancy, occult residual disease remains in a variable portion of patients and can eventually lead to relapse (51). Adjuvant treatment aims to kill cancer cells, thereby reducing the chance of relapse. With advancing knowledge of the interaction between the immune system and cancer, it becomes increasingly clear that higher tumor load is associated with higher tumor-induced immune suppression. For example, regulatory T-cells (Treg) and myeloid derived suppressor cells (MDSC) attracted by tumor cells induce anergy in T-cells (52). Moreover, several soluble factors secreted by tumor cells, such as TGF- β , IL-10 and VEGF, are recognized to suppress infiltrated effector T-cells (53–55). Also, tumors are able to upregulate indoleamine 2,3-dioxygenase (IDO) which converts tryptophan to kynurenine, inhibiting effector T-cells through a mechanism not completely understood (56). Tumor load-associated immune suppression is generally regarded as the underlying cause of the low clinical response to DC vaccination in metastatic disease (57). Indeed, in our group we detected antigen-specific T-cells in 71% of melanoma patients following adjuvant DC vaccination compared to 23% following vaccination in the metastatic setting (58, 59). In the adjuvant setting, the possibly remaining occult disease represents a low tumor burden, and hence less immune suppression (Figure 3). Therefore, DC vaccination may be more successful in the adjuvant compared to the metastatic setting.

There are some additional arguments to consider DC vaccination as an adjuvant treatment option. Besides efficacy, a low toxicity profile is an important hallmark of any adjuvant treatment as a substantial portion of cancer patients

receiving adjuvant treatment would not endure a relapse even without this adjuvant therapy. As noted before, DC vaccination is associated with little toxicity, not only compared to chemotherapy but also compared to ICI. In addition, besides a direct clinical benefit for patients, adjuvant DC vaccination might also prove to be beneficial in improving response to subsequent treatment in case of relapse. In theory, tumor-specific T-cells induced by adjuvant DC vaccination might result in an increased tumor-specific immune response when ICI are given at a later moment in the metastatic setting. Indeed, this effect has been observed retrospectively with administration of ipilimumab in patients with relapse after adjuvant DC vaccination for stage III melanoma (60). In addition to ipilimumab, a similar effect was also seen retrospectively in glioblastoma (GBM) patients receiving chemotherapy after DC vaccination (61). These additive effects should be considered when integrating DC vaccines in the therapeutic landscape of cancer. Considering these arguments, the next part will focus on data obtained with DC vaccines in the adjuvant setting.

Adjuvant DC Vaccination in Glioblastoma

Adjuvant DC vaccination has been studied in GBM. In contrast to most malignancies, distant metastases seldom occur in GBM (62). Nonetheless, GBM represents a lethal disease, with patients having a median survival of ~15 months (63). GBM is commonly treated with maximally safe surgery and adjuvant temozolamide (TMZ) in conjunction with radiotherapy, the so-called Stupp protocol (64). However, even with extensive treatment, residual disease invariably remains, and recurrence is certain. This results from the infiltrative growth and lack of a distinct border between normal brain tissue and tumor. Therefore, DC vaccination in the adjuvant setting after surgery in GBM is different from for example adjuvant DC vaccination in RCC and melanoma in which complete disease control after surgery is possible. In this review, we consider DC vaccination to be adjuvant when it is integrated in treatment protocols after maximally safe surgery in newly diagnosed GBM.

Historically, the central nervous system is considered an immune-privileged site, casting doubt whether GBM could be susceptible to immunotherapy. However, in recent years it has become increasingly clear the central nervous system is subject to active immunosurveillance even with an intact blood-brain barrier (65). Albeit not yet vigorously explored, the research into the treatment of GBM with ICI has not yet resulted in proof of efficacy. Nivolumab is the ICI furthest in clinical development, a phase III trial comparing nivolumab to bevacizumab for the first recurrence after radiotherapy and TMZ is currently ongoing (NCT02017717). Final results are not yet reported in a peer-reviewed journal, but presented results revealed that the primary end-point was not met (median OS in recurrent disease: 9.8 months with nivolumab vs. 10.0 months with bevacizumab) (66). Individual reports of response on anti-PD-1 mAb monotherapy do exist, although these are isolated cases concerning tumors with high mutational load (67–69). With these results in mind and the fact that mutational load and number of tumor infiltrating lymphocytes in GBM are generally low, it is doubtful whether

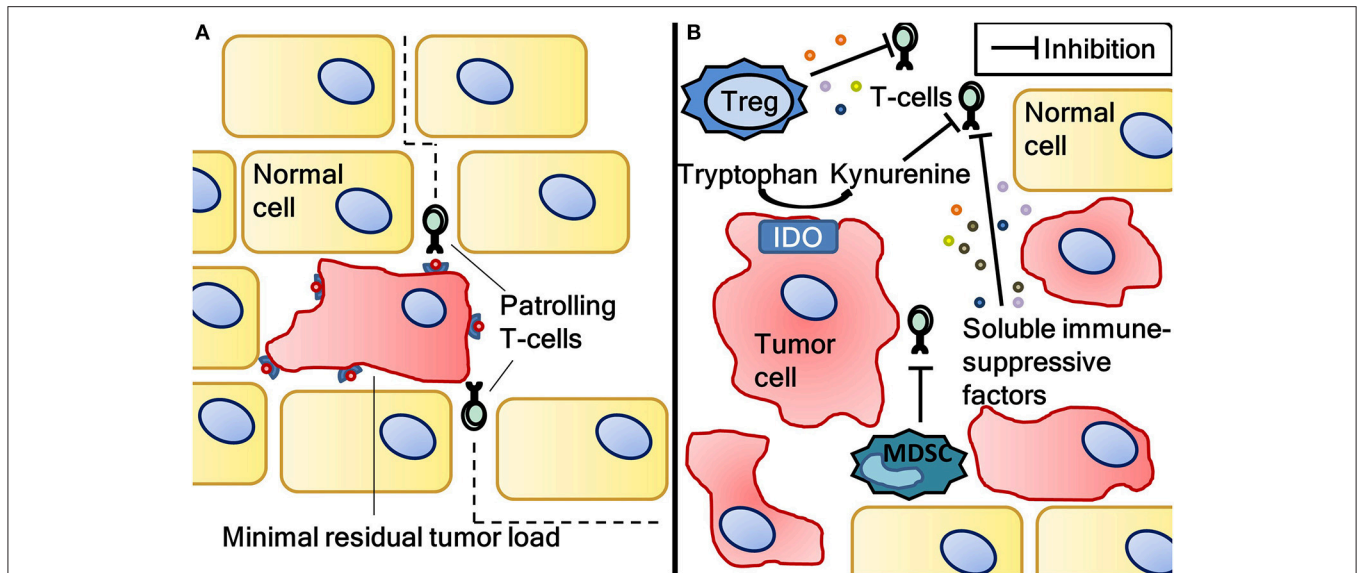


FIGURE 3 | The difference in tumor load-associated immune suppression between minimal residual disease and a situation with high tumor load. Antigen-specific T-cells induced by dendritic cell vaccination eliminating minimal residual disease after surgical resection of cancer **(A)**. Minimal residual disease is associated with less immune suppression as opposed to a situation with more tumor load **(B)**. Tumor load-associated immune suppression is caused by (among other factors) regulatory T-cells, myeloid derived suppressor cells, soluble immune suppressive factors (such as IL-10, TGFβ and VEGF) and indoleamine 2,3-dioxygenase activity. Vaccination-induced T-cells can be rendered anergic by this immune suppression, resulting in inferior clinical results. Therefore, dendritic cell vaccination might be more effective in the adjuvant setting. IDO, indoleamine 2,3-dioxygenase; MDSC, myeloid derived suppressor cells; Treg, regulatory T-cells.

ICI as monotherapy have promise as a future treatment option (70, 71).

Next to monotherapy with ICI, ICI combined with other standard treatment modalities is being investigated in phase III trials. For example, CheckMate 498 (comparing TMZ and radiotherapy to nivolumab and radiotherapy) and the CheckMate 548 (comparing radiotherapy, TMZ, and nivolumab to radiotherapy, TMZ and placebo), both involving nivolumab, are currently ongoing. Similar phase I and II trials combining pembrolizumab or ipilimumab with TMZ and radiotherapy are being performed. Results on such integration of ICI in standard treatment strategies are not yet reported.

Considering DC vaccination studies concerning GBM, DC-based therapy is often integrated into the standard adjuvant treatment for GBM. As of now, the only available phase III trial data involving DC vaccines in GBM are the very recently published interim results of an ongoing clinical study involving a vaccine called DCVax[®]-L (see also **Table 2**) (72). DCVax[®]-L is a vaccine manufactured from autologous DC loaded with tumor lysate derived from autologous GBM cells. Unblinded data on 331 patients with newly diagnosed GBM was presented. After surgery, patients were randomized (2:1) to receive either DCVax[®]-L incorporated into standard of care (TMZ and radiotherapy) or standard of care alone. Due to the study design, which enabled crossover from the standard of care to the vaccination arm upon progression, a total of 86% of patients received vaccination at the time of interim analysis. The authors compare the median OS of 23.1 months for the entire study population with OS data from comparable patients in different

TABLE 2 | Active phase III clinical trials concerning dendritic cell vaccination as adjuvant treatment in various malignancies (as of May 2018).

Disease	Vaccine formulation	Status	Identifier
Melanoma (stage III)	Natural dendritic cell subsets loaded with melanoma-specific peptides	Recruiting	NCT02993315
Uveal melanoma (high risk)	Dendritic cells loaded with autologous tumor RNA	Recruiting	NCT01983748
Glioblastoma (newly diagnosed)	DCVax [®] -L: dendritic cells loaded with tumor lysate	Active, not recruiting	NCT00045968

trials (which have a reported median OS of 15–17 months), from this comparison they suggest a clinical benefit from their vaccine. The definite results on clinical outcome, including PFS data, are eagerly awaited.

Previously, the favorable toxicity profile of DC vaccination was shown in several phase I/II studies showing the safety of adjuvant DC vaccination in GBM (73–78). Important to consider is that in these studies, DC vaccination was often combined with chemotherapy and/or radiotherapy, this combination had little added toxicity compared to chemotherapy and/or radiotherapy without DC vaccination. Despite not being designed for the purpose of assessing clinical outcome, these studies reported favorable median OS compared to their respective control groups ranging from 15 up to 41 months (74, 75, 77, 78). Furthermore, a positive correlation was shown between survival and presence of an immune response after vaccination (61).

Clinical outcome as primary endpoint was reported in several phase II studies. One of the largest studies completed to date involving DC vaccination in GBM, was performed by Ardon et al. and included 77 patients with newly diagnosed GBM (79). There was no control group, all patients received adjuvant DC vaccination integrated in standard treatment with TMZ and radiotherapy after complete resection of their GBM. The study reported favorable median OS of 18.3 months compared to the 14.6 months achieved in the landmark study by Stupp et al (64).

In conclusion, preliminary results on ICI in GBM make it very doubtful monotherapy with ICI will ever gain traction for this indication, results of large trials concerning ICI combined with chemoradiotherapy are pending. For DC vaccination in combination with chemoradiotherapy in GBM, occasionally favorable clinical outcomes have been reported. Due to strict inclusion criteria of these studies, the results are hard to interpret and compare with existing literature. Therefore, these result warrant further research with randomized phase III trials and additional data from the DCVax[®]-L trial are awaited.

Adjuvant DC Vaccination in RCC and Melanoma

Besides GBM, both RCC and melanoma in certain stages also exhibit high recurrence rates after surgery. For melanoma, the risk of relapse is particularly high when the disease has metastasized to regional lymph nodes (stage III). Melanoma with lymph node metastasis has a 5-year survival rate ranging from 40% (stage IIIC) to 78% (stage IIIA) (80). In RCC, recurrence of disease following surgery is also common, resulting in a declining survival rate with increasing stage (81).

Melanoma and RCC are similar in the sense that both tumors are very chemo-resistant and that their adjuvant treatment strategy in the pre-ICI era was mainly based on cytokine treatment with IL-2 and IFN- α (82, 83). In both cancers, IL-2 and IFN- α provide little clinical benefit and are associated with high toxicity. For melanoma, ipilimumab showed clinical activity in the adjuvant setting with a 5-year recurrence-free survival rate of 41% compared to 30% in the placebo group (hazard ratio for recurrence or death, 0.76; $p < 0.001$). Importantly, 5-year distant metastasis-free survival rate was also improved with 48% compared to 39% (hazard ratio for death or distant metastasis, 0.76; $p = 0.002$) (21). Although these results show efficacy, the application of adjuvant ipilimumab is opposed by its significant toxicity (~40% of patients experience immune-related grade 3 or 4 adverse events) (21, 84). In addition, both nivolumab and pembrolizumab have shown to increase PFS in the adjuvant setting for melanoma (19, 20). Adjuvant nivolumab was tested against ipilimumab in completely resected stage IIIB, IIIC and IV melanoma. In this study adjuvant nivolumab improved the 1-year PFS rate to 72.3% compared to 61.6% in ipilimumab-treated patients. Similarly, adjuvant pembrolizumab was compared to placebo in stage IIIA, IIIB and IIIC melanoma. The 1-year PFS rates were 75% and 61%, respectively. Despite pending OS

data, both the FDA and EMA recently granted approval for adjuvant nivolumab and are considering approval for adjuvant pembrolizumab.

For RCC, adjuvant treatment is also available. Adjuvant sunitinib, a tyrosine kinase inhibitor, for RCC has gained approval by the FDA based on improved PFS (6.8 months vs. 5.6 months for placebo; hazard ratio for recurrence, 0.76; $p = 0.03$). However, utility is limited due to high toxicity and lack of OS gain (85). Based on these considerations, the EMA has, in contrast to the FDA, adopted a negative opinion for the adjuvant application of sunitinib. In contrast to melanoma, for RCC no results on adjuvant ICI have been reported. However, several adjuvant clinical trials are ongoing, including the combination of ipilimumab and nivolumab (NCT03138512); atezolizumab (NCT03024996); pembrolizumab (NCT03142334) and nivolumab (NCT03055013) (82).

In both melanoma and RCC, DC vaccination has also been investigated as adjuvant treatment. Retrospective analysis from our group showed clinical benefit in stage III melanoma patients adjuvantly treated with monocyte-based DC vaccination compared to matched controls. In this study, OS for 78 patients treated with DC vaccines doubled compared to the 209 controls (63.6 months vs. 31.0 months; hazard ratio 0.59; $p = 0.018$) (58). Markowicz et al. have shown similar results in a prospective study concerning a peptide-loaded DC vaccine. In 22 vaccinated patients the study achieved a 3-year OS of 68% compared to 26% in the 22 patients of the matched historical control group ($p = 0.029$). The primary endpoint however, 3-year PFS rate, was not significantly improved probably due to the small number of patients (vaccinated patients: 41%; controls 15%; $p = 0.108$) (86). No phase III trials currently have been completed on adjuvant DC for melanoma. However, our group is currently conducting a trial which involves the employment of natural circulating DC vaccines in patients with stage IIIB or stage IIIC melanoma (NCT02993315) (Table 2).

In RCC, research on DC vaccination is mainly focused on metastatic disease and little data regarding adjuvant DC vaccination is available. However, a phase III trial was performed with adjuvant DC vaccination in various stages of disease. Patients vaccinated with DC loaded with tumor lysate in combination with cytokine-induced killer cells were compared to patients treated with IFN- α . Mainly due to a very heterogeneous study population, no definitive conclusions could be drawn. However, the study showed significant PFS and OS benefit suggesting that further research on adjuvant DC vaccination in RCC is warranted (87).

Currently, too little data is available to claim that DC vaccination is effective in the adjuvant setting. Yet, the above presented data, show favorable clinical results and consistently confirm the limited toxicity in a variety of cancers. More robust prove of efficacy may be under way as several phase III trials on adjuvant DC vaccination are currently being performed (Table 2). Whether DC vaccination acquires a definitive role in the adjuvant treatment of cancer will also be dependent on the results of ongoing phase III trials assessing other adjuvant treatments, including trials with ICI (88).

TABLE 3 | Ongoing clinical trials concerning dendritic cell vaccination in combination with clinically approved immune checkpoint inhibitors (ipilimumab, nivolumab, pembrolizumab, avelumab, atezolizumab, and durvalumab) in solid tumors.

Immune checkpoint inhibition (target molecule)	Vaccine formulation	Malignancy	Status	NCT-identifier
Combined Ipilimumab and Nivolumab (CTLA-4/PD-1)	DC with the insertion of the p53 gene	SCLC	Recruiting	NCT03406715
Nivolumab (PD-1)	DC loaded with CMV pp65 mRNA	Recurrent brain tumors	Active, not recruiting	NCT02529072
Nivolumab (PD-1)	DC loaded with NY-ESO-1 peptide	NY-ESO-1 ⁺ solid tumors	Recruiting	NCT02775292
Nivolumab (PD-1)	DC loaded with autologous tumor lysate	Recurrent glioblastoma	Not yet recruiting	NCT03014804
Pembrolizumab (PD-1)	DC loaded with peptide	Melanoma	Recruiting	NCT03092453
Pembrolizumab (PD-1)	DC-CIK	Solid tumors, NSCLC, Mesothelioma	Recruiting	NCT03190811 NCT03360630 NCT03393858
Nivolumab or Pembrolizumab (PD-1)	DC-CIK	Refractory solid tumors	Recruiting	NCT02886897
Avelumab (PD-L1)	DC/AML fusion vaccine	Colorectal cancer	Not yet recruiting	NCT03152565

CIK, cytokine induced natural killer cells; CMV, cytomegalovirus; CTLA-4, cytotoxic T-lymphocyte-associated protein 4; DC, dendritic cells; NSCLC, non-small-cell lung cancer; SCLC, small-cell lung cancer; PD-1, programmed death-1; PD-L1, programmed death ligand 1.

The Combination of DC Vaccination and Other Modalities for the Treatment of Metastatic Disease

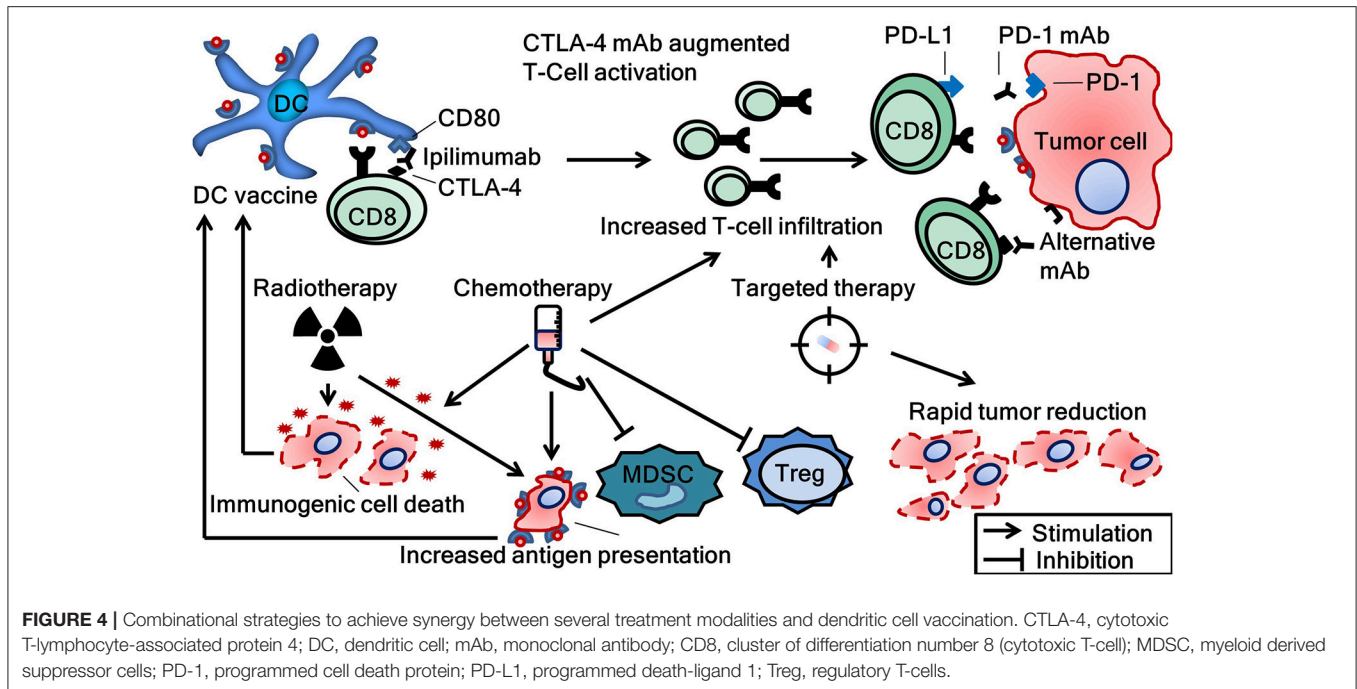
As noted before, the clinical benefit of monotherapy DC vaccination for patients with metastatic disease is probably limited. However, the ultimate role for vaccines may lie in the combination with other modalities. The generation of a cellular immune response upon DC vaccination is commonly reported and may potentiate the effect of other anti-cancer therapeutics (23). Conversely, tumor reduction caused by chemotherapy, radiation therapy or targeted therapy can alleviate tumor-induced immune suppression which hinders efficacy of DC vaccination. However, possible synergies involve more than the mere reduction of tumor load as modalities other than immunotherapy also exhibit immunogenic effects on tumors (Figure 4). For example, although chemotherapeutics are associated with lymphodepletion, positive immune modulatory effects are described, including the induction of immunogenic cell death and depletion of Treg and MDSC (89–92). In addition, radiotherapy and different forms of targeted therapy are known to have immunostimulatory properties, i.e., enhanced T-cell infiltration and killing capacity (93–96). Clinical studies combining DC vaccination with chemotherapy, radiotherapy, and/or targeted therapy have been performed. Without extensive elaboration on these studies, the safety of combining DC vaccination with these modalities is confirmed in phase I trials (97–100). Furthermore, ample data exist suggesting efficacy (101, 102). Besides these treatment modalities, the combination of DC vaccination with other forms of immunotherapy intervening in additional steps of the cancer immunity cycle may be of particular interest as it is thought to result in more additive immunogenic effects. For example, it would be very interesting to explore the combination of DC vaccination with chimeric antigen receptor (CAR) T-cell therapy, oncolytic viruses, or other investigational immunotherapies. Here, we will discuss the combination of DC vaccination

with the most successful immunotherapeutic agents to date, ICI.

Both ICI and DC vaccination exert their effects primarily through the modulation of the immune system and do so on different steps in the cancer immunity cycle. For response on ICI, tumor-specific T-cells have to be present in the tumor microenvironment, the generation of which may be aided with DC vaccination (103). As introduced before, a higher number of tumor-infiltrating lymphocytes are associated with a better response on ICI. In this respect, especially in tumors with low mutational burden, the addition of DC vaccines could prove to be beneficial (104).

Conversely, T-cells induced by DC vaccination are often hindered by the immune suppressive milieu of tumors. ICI might aid the effector functions of these T-cells by reducing inhibition through PD-1 signaling or by enhancing T-cell activation through the modulation of CTLA-4. The idea that tumor-specific T-cells activated by DC vaccination can be further stimulated with ICI is also supported by pre-clinical data. For example, upregulation of PD-1 on T-cells derived from the blood of vaccinated patients has been shown *in vitro* (105). Subsequent blockade of these upregulated PD-1 molecules could augment T-cell function. In addition, ICI exert several immune augmenting effects besides the direct antagonism of PD-1 and CTLA-4. For example, Treg depletion by anti-PD-1 mAb was shown in a mouse model (106).

In contrast to preclinical data, clinical data on combined treatment with ICI and DC vaccination in humans is scarce. In 2009, Ribas et al. reported safety of combining tremelimumab (CTLA-4 mAb) and DC vaccination in melanoma patients (107). Despite the trial was not designed to assess clinical outcome, 4 out of 16 patients (25%) achieved an objective clinical response. The authors state that clinical benefit was at the higher end of what can be expected from monotherapy tremelimumab. In addition, Wilgenhof et al. showed a promising ORR of 38% in 39 metastatic melanoma patients treated with the combination of ipilimumab



and DC vaccination (108). In 36% of patients grade 3 or 4 adverse events were seen, which is comparable with rates seen in large clinical trials with monotherapy ipilimumab (5, 84). This suggests little added toxicity from the addition of DC vaccines to ICI.

Considering its lower toxicity and better response rates compared to anti-CTLA-4 mAb, anti-PD-1 mAb might be more suitable combinational partners for DC vaccines. As of now, no data is published on the combined anti-PD-1 mAb and DC vaccination. However, several clinical trials investigating combinations of DC vaccination with clinically approved ICI are currently being performed (Table 3).

Besides currently approved ICI, DC vaccination can also be combined with ICI targeting alternative immune checkpoints (not yet clinically approved mAb). Currently, mAb targeting LAG-3 and TIM-3 are in various stages of clinical development as monotherapy and might be good candidates for combination. LAG-3 mAb for example, were shown to reduce expansion of Treg (109). TIM-3 was shown to be present in conjunction with PD-1 on dysfunctional T-cells after vaccination, suggesting they might form a target for mAb in addition to anti-PD-1 (110). Finally, the combination of multiple ICI and DC vaccination might be a promising strategy, albeit requiring careful considerations concerning the related toxicities (111).

Despite several ongoing clinical trials, an important aspect of combinational strategies, the timing of administration, might be under-investigated. In theory, it would seem logical to first administer DC vaccines to generate tumor-specific T-cells and consequently release immune suppression with anti-PD-1 mAb. Conversely, the timing of administering DC vaccines and ipilimumab may be more complex as both ipilimumab and these vaccines exert their functions in the priming phase of T-cells. Indeed, in a pre-clinical prostate cancer model optimal response

on ipilimumab was shown when given on the same day as vaccination (112). Whether the timing of anti-PD-1 mAb and DC vaccination is equally important is not known and forms an interesting subject for further research.

In conclusion, combinational strategies for the treatment of cancer incorporating DC vaccination are a promising field of research. Considering the favorable results on the combination of DC vaccination and anti-CTLA-4 mAb, the results on the currently ongoing combinational clinical trials with anti-PD-1 and anti-PD-L1 mAb are eagerly awaited.

CONCLUSION

Immunotherapy for the treatment of cancer is a fast-moving field. It is important to determine the relative position of DC vaccination to other treatments in this rapidly evolving landscape. Ideally, patients can be selected based on biomarkers predictive for response to therapy. Currently, no predictive biomarkers for DC vaccine response are applied in the clinic to guide treatment decisions but the immune landscape of the tumor might hold promise. Also, few clinically useful predictive biomarkers for ICI are known. With the success of ICI and the lesser clinical benefit of DC vaccination in metastatic disease, it becomes increasingly clear that the future of DC vaccination in extensive metastatic disease as standalone treatment is probably limited. However, the immune-inducing properties of DC vaccination makes it a prime candidate for combination with other anti-cancer modalities, especially ICI. The currently ongoing research on DC vaccination combined with ICI such as anti-PD-1 mAb has to determine whether this combination has a future perspective. The theoretical basis

and the promising clinical data on anti-CTLA-4 mAb combined with DC vaccination does imply this perspective exists. With its highly favorable toxicity profile, another application of DC vaccination might lie in the adjuvant setting. Furthermore, DC vaccination as monotherapy may be more effective in adjuvant setting compared to its application in metastatic setting.

Consequently, for DC vaccination to gain a definitive role in the therapeutic landscape of cancer, research should be focused on well-designed trials in the adjuvant setting, combinational strategies, and patient selection.

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Bristol-Myers, Dendreon, Squibb, and Sanofi. WG participated in *ad hoc* consultancy for Aglaia Biomedical Ventures; WG received research grants from Bayer; Astellas and Janssen-Cilag.

AUTHOR CONTRIBUTIONS

WvW, KB, and IdV conception and design; WvW, KB, MB, GS, IdV, and WG writing, review, and/or revision of the manuscript.

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