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Review Series

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Senescent cells: a therapeutic target for cardiovascular disease

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Cellular senescence, a major tumor-suppressive cell fate, has emerged from humble beginnings as an in vitro phenomenon into recognition as a fundamental mechanism of aging. In the process, senescent cells have attracted attention as a therapeutic target for age-related diseases, including cardiovascular disease (CVD), the leading cause of morbidity and mortality in the elderly. Given the aging global population and the inadequacy of current medical management, attenuating the health care burden of CVD would be transformative to clinical practice. Here, we review the evidence that cellular senescence drives CVD in a bimodal fashion by both priming the aged cardiovascular system for disease and driving established disease forward. Hence, the growing field of senotherapy (neutralizing senescent cells for therapeutic benefit) is poised to contribute to both prevention and treatment of CVD.

Introduction

During aging, the human mortality rate rises exponentially as a result of a loss of normal organ functions, including tissue maintenance and repair capacity. Stressors that young tissue may tolerate because of excess functional reserve become insurmountable in the elderly, leading to age-related diseases (1). Age-related cardiovascular disease (CVD) is of particular clinical concern as the global population ages, with 1 billion individuals projected to be over 65 years old by 2030. Indeed, 38% of people ages 40 to 60 and 83% of those over 85 have CVD (2). The primary diseases included under the umbrella of CVD are chronic heart failure (HF), hyper-tension, and coronary artery disease (CAD; atherosclerosis). Although hypertension and atherosclerosis can be pharmacologically managed, therapy-resistant subpopulations exist for both these diseases (3, 4), and the long-term prognosis for HF remains poor (5). Given the growing patient population and the inadequacy of current medical management, there is strong incentive to identify new therapeutic targets to treat CVD or, more optimally, prevent it.

One mechanism behind age-related CVD may be cellular senescence, a stress-response process by which damaged cells exit the cell cycle permanently and produce a proinflammatory senescence-associated secretory phenotype (SASP) (1). The cellular senescence program serves an important tumor-suppressive function by growth-arresting preneoplastic cells. Normally, SASP factors target senescent cells (SNCs) for removal by the immune system, clearing adjacent, nonsenescent preneoplastic cells in the process and therefore providing tumor-suppressive benefits that are supe-

rior to apoptosis (6). However, long-term persistence of SNCs and their secretome drives cancer, atherosclerosis (7), and other major age-related diseases, in addition to aging itself (8–10). The solidifying link between SNCs, cardiovascular aging, and CVD depends on four lines of evidence. First, stressors that drive senescence, such as telomere shortening and oxidative stress, rise with aging and CVD in cardiovascular tissue (11–13). Second, SNC biomarkers are present at the same sites (14, 15). Third, prevention of the SNC fate by genetic elimination of senescence effectors can blunt age-related loss of tissue function (16). Finally, and most compellingly, SNC-killing approaches block the effects of aging on cardiovascular performance (10), attenuate CVD (7), and may ultimately serve as therapies (17).

Based on these lines of evidence, here we propose a bimodal model for cellular senescence in CVD. Initially, primary SNCs accumulate in cardiovascular tissue with age, decreasing organ functional reserve and predisposing to age-related CVD. Then, as CVD develops, heightened cellular stress within the diseased tissue microenvironment produces a second wave of disease-associated SNCs (secondary SNCs). Secondary SNCs go on to exacerbate preexisting CVD. Because SNCs predispose to CVD as well as participate in pathophysiology, we posit that SNCs could be targeted both preventatively and therapeutically in CVD.

Senescence-driven aging is a tradeoff for tumor suppression

Hayflick and Moorhead showed in 1961 that human fibroblasts in vitro can divide a limited number of times before entering a permanent, stable growth arrest (18). This finite proliferative potential, now called replicative senescence, was swiftly proposed as a cell-autonomous mechanism for declining tissue repair capacity in aging (19). The primary molecular event responsible for this “senescence timer” in human cells is now known to be telomere attrition (Figure 1 and ref. 20). Owing to the nature of RNA-templated DNA replication, approximately 50 to 100 base pairs

Conflict of interest: JMVD is a cofounder of Unity Biotechnology, a company developing senolytic medicines, including small molecules that selectively eliminate senescent cells. JMVD, HL, and BGC are co-inventors on patent applications licensed to or filed by Unity Biotechnology (15/455,630 and 62/057,825).

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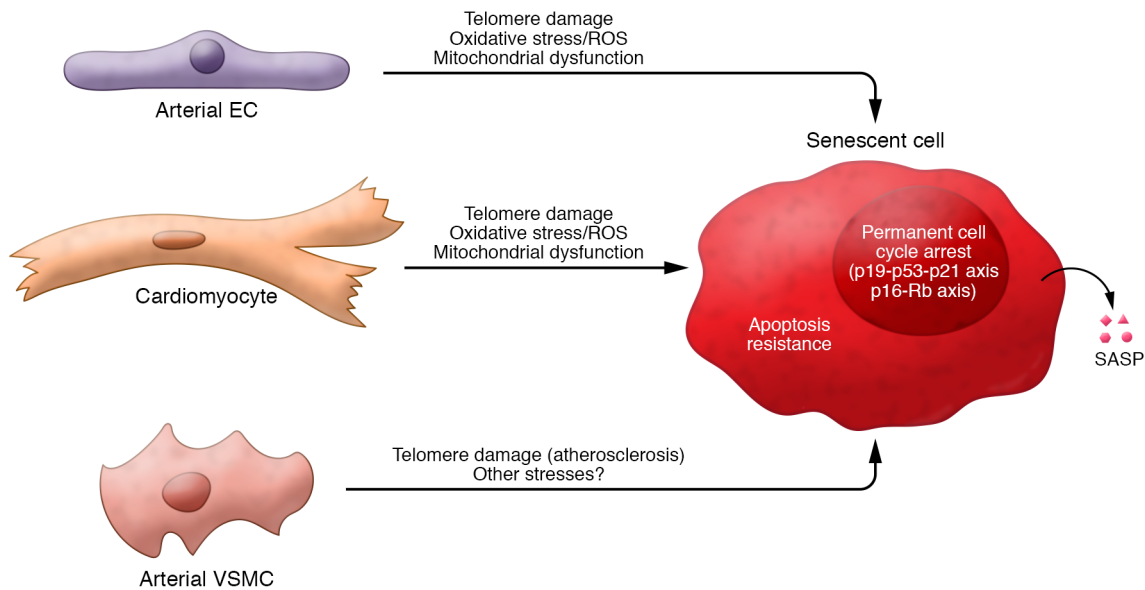


Figure 1. Key properties of senescent cells. In response to various types of stressors (generally irreparable macromolecular damage), replication-competent cardiovascular cells undergo senescence. The 3 major hallmarks of SNCs are a permanent cell cycle arrest, mediated by signaling through the p19^{Arf}-p53-p21^{Cip/Waf} and p16^{Ink4a}-Rb axes; apoptosis resistance, achieved by upregulation of pro-survival factors; and acquisition of the senescence-associated secretory phenotype (SASP), a bioactive secretome containing cytokines, growth factors, proteases, and other signaling molecules.

are lost from the ends of chromosomes with each cell division (21). Telomeres, comprising DNA hexanucleotide repeats complexed with protein, are protective caps terminating each chromosome that prevent this attrition from reaching coding sequences (22). More importantly, telomeres prevent DNA damage response (DDR) machinery from recognizing chromosome free ends as double-strand breaks (DSBs) and attempting erroneous repair, which could result in illegitimate recombination or chromosome fusion events. Fused chromosomes may break during mitosis, with repeated fusion/breakage cycles resulting in chromosomal instability, aneuploidy, and neoplastic transformation (23). Therefore, when telomeres become critically short, the DDR is activated and induces senescence to safeguard against replication with an unstable karyotype.

Elucidation of the molecular mechanisms behind replicative senescence has produced a framework for understanding senescence induction and maintenance. First, some form of molecular damage occurs and is recognized by a damage sensor, such as the apical ATM/R kinases in the case of DSBs (Figure 1). In addition to telomere erosion, other stressors that induce DSBs, such as oncogenic Ras expression (24, 25) and ionizing radiation (26), can also activate ATM/R to trigger senescence. Second, the cell engages effectors of a reversible cell-cycle arrest to halt proliferation and attempt repair. For example, Ras stabilizes p53 to upregulate the cyclin-dependent kinase 2 (CDK2) inhibitor p21^{Cip/Waf}, thereby blocking Rb phosphorylation and preventing S-phase entry. With Ras overexpression, p53 stabilization involves both p53 phosphorylation by ATM/R and, in mice, upregulation of p19^{Arf}, which blocks the ubiquitin ligase MDM2 from degrading p53. Third, should repair prove impossible, the cell enters senescence and permanently exits the cell cycle. Permanent cell-cycle withdrawal requires expression of additional CDK inhibitors (CDKis), most notably the CDK4/6

inhibitor p16^{Ink4a} (27). Finally, the senescence state is reinforced through heterochromatinization of cell-cycle genes (28, 29) and the establishment of the SASP, which includes factors such as IL-6 that help maintain the cell-cycle arrest (30–33).

SNCs *in vitro* are known to produce numerous inflammatory cytokines, chemoattractant factors, proteases, and signaling molecules as part of the SASP (Figure 1 and refs. 30, 31). The SASP is a p16^{Ink4a}-independent SNC property that can be either DDR-dependent or -independent, but appears to rely on proinflammatory signaling through NF- κ B, mTOR, and p38/MAPK (34–36). The SASP *in vivo* has diverse functions and, depending on biological context, can be beneficial or detrimental. For example, transient SNCs arising during wounding secrete PDGF-AA to hasten wound closure and are rapidly cleared (37). In contrast, age-associated SNCs linger, driving tissue inflammation, dysfunction, and carcinogenesis presumably via the SASP (8–10, 38).

Cellular senescence drives cardiovascular aging

Major age-related changes in the cardiovascular system. Normal aging produces multiple alterations in the cardiovascular system that predispose to CVD. Crucially, arterial and cardiac health are tightly interrelated; it is therefore likely that interventions that enhance cardiac or arterial healthspan individually would have widespread benefits for CVD risk.

Aging results in arterial dysfunction at a structural and molecular level. In the arterial wall (media), elastic fiber degradation and enhanced collagen synthesis by vascular smooth muscle cells (VSMCs) result in arterial stiffening (39, 40). Increased arterial stiffness results in higher systolic and lower diastolic pressure (41), producing widened pulse pressure hypertension (42) that, together with aortic root dilation (43), increases cardiac afterload (44). When combined with hypertension, reduced elastic fiber content can predispose to aneurysms (45–47). Endothelial dysfunction

with resultant decreased endothelial nitric oxide synthase (eNOS) activity contributes to these changes. Low NO tone promotes VSMC proliferation and collagen production (48, 49) and can also lead to vasoconstriction, predisposing to angina pectoris and ischemic heart injury (50, 51). Dysfunctional endothelial cells also secrete monocyte chemoattractant protein-1 (MCP-1) and present the leukocyte receptors VCAM1 and ICAM1 on their surfaces, thereby recruiting circulating monocytes and initiating atherosclerosis (52). Upregulated NADPH oxidase (NOX) activity in dysfunctional endothelial cells can also shift eNOS toward production of superoxide anion, a reactive oxygen species (ROS), rather than NO, leading to inflammation and further dysfunction in a vicious cycle (53–55).

The normal aging heart employs several mechanisms to maintain resting systolic function and ejection fraction despite increased afterload (56). Structurally, the left ventricle (LV) thickens to normalize tension across the LV wall but does not increase in mass (48), instead shortening across the long axis. The cardiac cycle changes, shifting from early to late diastolic filling and prolonging isovolumic relaxation time (57). Atria-dependent diastolic filling is enhanced by left atria enlargement and increased contractility (58, 59). Systole is extended as a result of prolonged cardiomyocyte contraction, with a decrease in maximum heart rate driven by impaired SERCA2-dependent Ca^{2+} reuptake (60–62), attenuated responsiveness to β -adrenergic signaling, and increased $\omega 6/\omega 3$ polyunsaturated fatty acid ratio in cardiomyocyte membranes (41).

Although the aforementioned cardiac changes are adaptive and preserve systolic function, this remodeling predisposes to stress-induced pathology. Left atrial dilation can cause atrial fibrillation and result in thrombosis (63). Reduced Ca^{2+} handling ability predisposes to cardiomyocyte death and further cellular hypertrophy. On this background of an aged heart with limited functional reserve, damage following myocardial infarction (MI) can produce HF (64). Crucially, despite adaptive cardiac changes, cardiovascular dynamics are still abnormal. For example, late systolic peak pressure, increased pulse wave velocity, and widened pulse pressure hypertension disrupt laminar flow that would normally trigger protective eNOS expression, resulting in endothelial dysfunction, atherosclerosis, and CAD (65).

Prosenescence stressors increase in aging heart and vasculature. Despite being a largely nondividing cell type, cardiomyocytes from aged rats (66) and mice (67, 68) show telomere shortening, as does healthy human heart tissue (69). Telomeric DNA is particularly susceptible to DSB-inducing oxidative stress (70) and accumulates DNA-damage foci with age (71), potentially explaining telomere dysfunction in noncycling cardiomyocytes. However, cell-cycle reentry of a small fraction of cardiomyocytes has been reported (72). Aged cardiomyocytes also show increased oxidative damage of mitochondrial lipids (73), mitochondrial dysfunction (74, 75), and ROS. Supporting a causal role for ROS in cardiac aging, old mice expressing mitochondrial-targeted catalase, which degrades H_2O_2 into water and O_2 , show longer lifespan, decreased LV hypertrophy, and reduced diastolic dysfunction (76, 77). Additional studies report significant age-related telomere shortening and reduced telomerase activity in human cardiac stem cells, a cKit-positive, CD45-negative cell type that replaces cardiomyocytes throughout life (78, 79).

Prosenescence stressors also rise in aged blood vessels. Telomere shortening occurs with normal aging in human large arteries (12, 13) and drives p53-dependent expression of the CDK1 p21 (80) in endothelial cells (11). Intriguingly, circulating bone marrow-derived early endothelial progenitor cells (EPCs) exhibit age-dependent telomere shortening (81) and less telomerase activity (82). Aged human (83) and murine (84, 85) arterial endothelial cells as well as EPCs (86, 87) also show heightened levels of ROS partially due to activation of NADPH oxidase, which uncouples eNOS (83, 88, 89). Further study is needed to clarify whether these age-related changes to early EPC properties directly or indirectly affect endothelial homeostasis in vivo. Although human VSMCs undergo telomere shortening during atherosclerosis (15), this has not been demonstrated with normal aging. Collectively, these results suggest that prosenescence stressors, particularly telomere attrition and oxidative stress, are present throughout the aging cardiovascular system in the absence of frank age-related disease.

SNCs accumulate in the aging cardiovascular system. Reliably detecting SNCs in vivo is a major ongoing challenge to investigating their biological significance to the aging process. Currently, detection depends on a combination of SNC biomarkers, as any marker in isolation is prone to false positives. These include high expression of the CDK1s p16^{Ink4a} and p21^{Cip/Waf1}; lysosomal hydrolase activity detectable at pH 6.0, known as senescence-associated β -galactosidase (SA β -gal) (90); and expression of SASP-associated inflammatory, proteolytic, and signaling factors. Based on these markers, SNCs have been shown to accumulate in numerous tissues with age in humans, nonhuman primates (91), and rodents (10), including heart and arteries.

Aged murine hearts show increased expression of p16^{Ink4a} (10), the p53-stabilizing protein p19^{Arf} (10, 91), and inflammatory SASP factors (10). Cardiac stem cells have also been shown to upregulate p16^{Ink4a} and p21^{Cip/Waf1}, potentially restricting their ability to produce new cardiomyocytes during aging and HF (78, 79). Surprisingly, a large fraction of postmitotic cardiomyocytes (about 85%) in old hearts (72–104 years) were shown to be p16^{Ink4a}-positive, versus approximately 30% in younger controls (19–49 years) (79). This high baseline may indicate that p16^{Ink4a} expression is a normal differentiation step in cardiomyocyte commitment, as has been reported for macrophages (92), or that senescence may be a relatively early event. Consistent with this interpretation, young murine hearts contain a mixed pool of p16^{Ink4a}-positive, large cardiomyocytes with short telomeres and p16^{Ink4a}-negative, small cardiomyocytes (potentially replication-competent) with long telomeres (93). Low-frequency SA β -gal-positive cardiomyocytes have also been reported in aged mice (94) and rats (95).

These observations suggest that senescence in progenitor cell compartments could restrict cardiomyocyte replacement. However, the baseline ability of the adult human heart to renew cardiomyocytes, and how this changes with age, are controversial. Most studies report yearly replacement rates around 1% for young adult mice (96) and humans (97, 98). New cardiomyocytes reportedly arise from cardiac stem cells residing in the heart (99) and bone marrow-derived progenitors (100), as well as, strikingly, mature cardiomyocytes (101, 102). Therefore, unequivocally determining how senescence rates in these compartments change as a function of age is paramount.

Visceral and serous pericardial epithelial cells, which belong to the trilayered, fibrous sac sheathing the heart, also show an age-related increase in SA β -gal activity and express p16^{ink4a} (10). Whereas the consequence of senescence in cardiac stem cells and cardiomyocytes is unknown, removal of p16^{ink4a}-positive pericardial cells during normal aging in mice blunted cardiomyocyte hypertrophy, normalized cardiac stress tolerance, and reduced expression of inflammatory factors in the heart (10). This result supports the existence of prohomeostatic signaling from the pericardium to the ventricle that is disrupted by SNC accumulation, although the effect of SNCs at other sites cannot be ruled out.

Senescence also occurs with aging in both major arterial cell types, endothelium (103, 104) and VSMCs (105–107). Endothelial NO produced by eNOS normally maintains VSMCs in a nondividing, contractile state and suppresses thrombogenic and inflammatory signaling in endothelium. This system's activity declines with age, partially because of heightened angiotensin II production (108). Intriguingly, NO production is reportedly decreased in senescent endothelial cells (109, 110). Reciprocally, NO activates telomerase (111) to prevent endothelial cell senescence, thereby decreasing presentation of the proatherogenic, leukocyte-binding receptors VCAM1 and ICAM1 (112). Similarly, NO production may inhibit VSMC senescence. Aged arterial VSMCs show elevated expression of p16^{ink4a} and SA β -gal activity (105–107), potentially due to aberrant cell-cycle reentry that results in polyploidization, greater levels of the NADPH oxidase NOX4, and ROS production. Additionally, aging raises circulating levels of angiotensin II, which activates the Ras signaling cascade, a well-known driver of oncogene-induced senescence (113), to produce VSMC senescence (114, 115) and enhance NADPH oxidase expression and ROS production (108). The SASP factors MMP2 and IL-6 are also increased in aged VSMCs (116) as well as proatherogenic VCAM1, although whether these gene expression changes are restricted to senescent VSMCs is unknown.

Neutralizing SNCs prevents age-related cardiovascular impairment. Approaches now exist to block the senescence program or kill SNCs to study their contribution to the aging process. Early evidence that SNCs promote aging came from a mouse model of accelerated aging (progeria) caused by insufficiency of the mitotic checkpoint protein BubR1. BubR1-insufficient mice show several aging phenotypes, including loss of muscle mass (sarcopenia), adipose atrophy, and cataract formation. Dysfunctional skeletal muscle, fat, and eyes in BubR1-insufficient mice prematurely accumulate high levels of SNCs. When SNC formation was blunted by genetic deletion of the senescence effector p16^{ink4a}, development of these degenerative phenotypes was blocked (16). However, this strategy could not distinguish between cell-autonomous deleterious effects of senescence, such as inhibited cycling of tissue-repairing progenitor cells, and paracrine effects of SNCs, such as the SASP.

To discriminate between these possibilities, an SNC suicide transgenic mouse (INK-ATTAC mouse), which permits INK-linked apoptosis through targeted activation of caspase, was developed (38). Briefly, this transgenic uses a 2.5-kbp minimal fragment of the p16^{ink4a} promoter to drive expression of a GFP reporter and an FKBP/caspase-8 fusion protein in p16^{ink4a}-positive SNCs. When mice are treated with an FKBP cross-linking ligand, AP20187, caspase-8 dimerizes and activates to trigger apopto-

sis of p16^{ink4a}-positive cells. Lifelong removal of p16^{ink4a}-positive SNCs in BubR1-progeroid mice using INK-ATTAC profoundly attenuated sarcopenia, fat loss, and cataract formation (38). However, progeroid mice incompletely recapitulate the complex tissue changes associated with normal aging. Furthermore, BubR1-progeroid mice show several cardiovascular phenotypes, including arrhythmia, aortic VSMC loss, and intolerance to isoproterenol stress that are not corrected by SNC killing via INK-ATTAC in this model of aging (117).

To test the contribution of SNCs to normal aging, including cardiovascular aging, transgene-mediated SNC killing was repeated in naturally aged mice. Wild-type INK-ATTAC mice were aged to 12 months (approximately middle-aged) before receiving twice-weekly administration of AP20187 or vehicle to remove SNCs (10). SNC clearance resulted in a sex- and strain-independent increase in median lifespan with corresponding reduction of SNC markers in several vital organs, including heart and kidney. At 18 months, SA β -gal-positive SNCs were identified by transmission electron microscopy in the visceral and serous pericardium of the heart as well as the renal proximal tubule epithelium. AP20187 treatment removed these SNCs, thereby blunting age-related cardiomyocyte hypertrophy and glomerulosclerosis. In the heart, SNC clearance increased tolerance to a lethal dose of isoproterenol, a β -adrenergic agonist, and reduced development of diastolic dysfunction when mice were challenged with sublethal, chronic isoproterenol (10). In the kidney, SNC removal blunted activity of the renin-angiotensin-aldosterone system (RAAS), reducing glomerulosclerosis and blood urea nitrogen, a surrogate marker of kidney function. Although these results are consistent with local action of pericardial and renal SNCs, INK-ATTAC also clears SNCs at other distal sites, such as adipose and skeletal muscle. Tissue-specific SNC clearance models will be necessary to disentangle global versus local effects of SNCs on cardiovascular function.

Senescent cells drive age-related cardiovascular diseases

Although SNCs accumulate with normal aging in cardiovascular tissues, SNCs arise locally in comparably larger numbers as a result of cellular stressors experienced during age-related diseases. We term these “secondary SNCs,” in contrast to primary SNCs that arise with basal aging. Primary and secondary SNCs may be different cell types or arise due to different stressors, thereby possessing distinct properties including unique SASPs. To illustrate the bimodal role of SNCs in age-related CVD, we will explore two major age-related disease processes: atherosclerosis and HF.

Atherosclerosis. During atherogenesis, an aberrant lipid profile drives oxLDL retention in the subendothelial space. This prompts circulating monocytes to invade, where they differentiate, take up oxLDL, and convert to foam cell macrophages (118). These foam cells secrete cytokines and chemokines such as IL-1 α , TNF- α , and MCP-1 to drive further immune cell recruitment and form an atherosclerotic plaque (118, 119). CAD and deaths from atherosclerosis complications (MI, ischemic heart disease, and thromboembolic stroke) rise dramatically with age but can also occur earlier given appropriate stressors such as diabetes, meta-

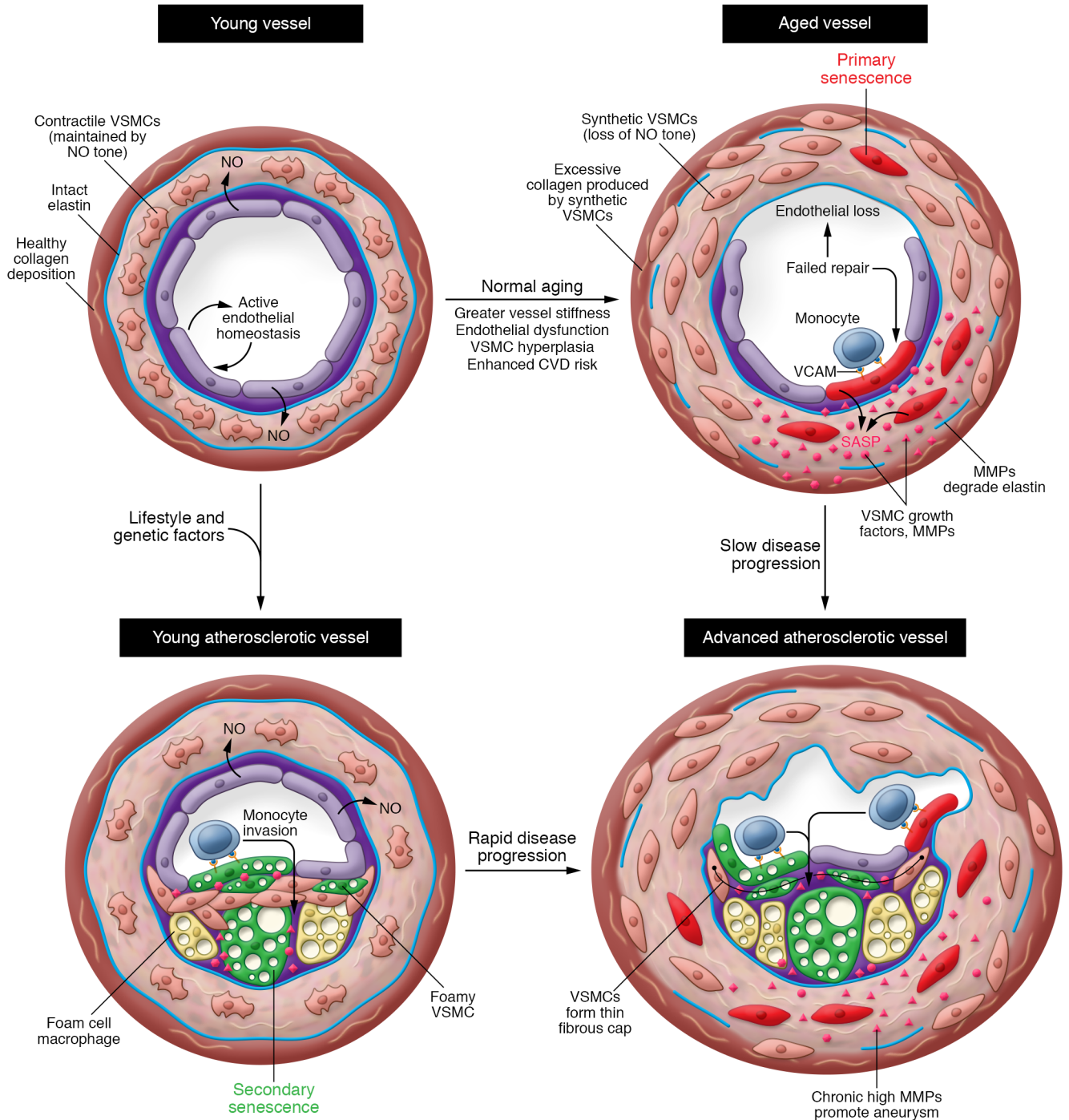


Figure 2. Cellular senescence is a cause and consequence of atherosclerosis. Both primary (age-derived) SNCs and secondary (atherosclerosis-derived) SNCs may promote atherosclerosis. In young arteries (top left), senescence burden is low. Vessel homeostasis is achieved by endothelium-derived NO acting on VSMCs, promoting vasodilation, quiescence, contractile phenotype, and endothelial health. During normal aging (top right), endothelial cells and VSMCs undergo primary senescence (red cells). Loss of NO tone may arise due to failure of endothelial cell homeostasis, leading to endothelial dysfunction and senescence. Excessive VSMC proliferation in aging vessels may also drive VSMC senescence, further exacerbating VSMC hyperplasia via SASP-mediated growth factor release. Whatever the causal chain, loss of NO tone converts VSMC to the synthetic phenotype, prompting VSMC hyperplasia, medial thickening, collagen overproduction, and synthesis of elastin-degrading metalloproteases (MMPs), collectively producing vessel stiffening, widened pulse pressure, hypertension, and increased cardiac afterload. Vessel stiffening interrupts normal shear stress signals that suppress VCAM expression, with proatherogenic effects. Absence of risk factors in late life may produce late-onset cardiovascular disease (CVD; bottom right). Alternatively, early-life CVD risk factors (e.g., diabetes, hypertension, metabolic syndrome, and dyslipidemia) promote aberrant oxi-LDL accumulation in the subendothelial space, initially producing secondary senescent foam cell macrophages. At later stages of disease, secondary senescent foamy endothelium and VSMCs also arise (green cells). This faster trajectory results from intense, proatherogenic stress and results in early CVD (bottom left). The relative contribution of primary and secondary SNCs to atheroprotection probably varies with life history and personal risk factors, but both types likely contribute to key features of clinical disease, including thinning of the fibrous cap as a result of VSMC growth arrest and SASP-derived collagenases and elastases; enhanced lesion growth due to expression of VCAM and ICAM; and lesion-associated aneurysm resulting from medial elastin degradation.

bolic syndrome, and an abnormal circulating lipid profile (120). One interpretation of these facts is that the aging cardiovascular system becomes abnormally sensitive to proatherogenic stressors. Alternatively, the high rate of atherosclerosis in the elderly may represent the inevitable result of slow progression of subclinical disease over decades, and therefore atherosclerosis may not be a “true” age-related disease (i.e., one in which biological age rather than length of disease progression is a causal factor).

Although this question is largely unresolved, proatherogenic mechanisms are active in primary SNCs of normally aging arteries (Figure 2, top right). Dysfunctional and senescent endothelial cells present VCAM1 and ICAM1, which bind circulating monocytes to drive their invasion (112). Senescent endothelial cells *in vitro* also show apoptosis susceptibility and impaired tight junction formation, both of which could enhance oxLDL retention in the subendothelium (121, 122). EPC senescence may contribute in part to endothelial cell dysfunction through failed repair. IL-6 and MCP-1 produced by aged VSMCs may contribute to the atherogenic milieu (123, 124).

Secondary SNCs also arise during atherogenesis, thereby driving lesion expansion and deleterious remodeling by producing an inflammatory, proteolytic environment (refs. 7, 13–15, 125, and Figure 2, bottom left). Early “fatty streak” lesions contain p16^{Ink4a}-positive, SA β -gal-positive foam cell macrophages and can be largely ablated in the *Ldlr*^{-/-} model of atherosclerosis using transgenic or pharmacological approaches to kill SNCs (7). This preventative benefit of SNC killing extends to intermediate and advanced lesions, which contain senescent foamy endothelium and VSMCs in addition to senescent foam cell macrophages. Mechanistically, SNC removal reduced expression of the atherogenic cytokines IL-1 α and TNF- α , as well as MCP-1 (7). Importantly, SNC removal blocked thinning of the fibrous cap, a VSMC- and extracellular matrix-rich structure that contains the plaque to prevent rupture, clotting cascade engagement, and thromboembolism (126). A cap-stabilizing effect of senolysis is consistent with direct production of elastase and collagenase by SNCs as part of the SASP. Indeed, a study using GFP expression in *Ldlr*^{-/-} INK-ATTAC mice to flow-sort lesional SNCs showed that these cells express higher levels of the elastase MMP12, the collagenase MMP13, IL-1 α , and MCP-1 than their nonsenescent counterparts (7). Age-related atherosclerosis likely features contributions from primary and secondary SNCs, and occurs earlier or later in life depending on whether proatherogenic stressors or aging make the larger contribution, respectively (Figure 2, bottom right).

By contrast, the proliferative senescence arrest may initially restrict atherosclerotic lesion development. In murine models of high-fat diet-induced atherogenesis, deficiency of any of the senescence effector genes p21 (127), p27^{Kip} (128), and p53 (129) resulted in greater lesion burden than controls. Similarly, human genome-wide association studies have shown that polymorphisms in the senescence-modulating 9p21.3 locus increase the risk of MI and stroke (130, 131). This locus contains several senescence effectors, including the *CDKN2a* gene, which codes for p19^{Arf} and p16^{Ink4a}, and the *CDKN2b* gene, encoding p15^{Ink4b}. 9p21.3 also contains *CDKN2b-AS1*, encoding the long noncoding RNA ANRIL, whose expression directs inhibitory methylation of *CDKN2a/b* (132, 133). Presence of the CAD 9p21.3 risk locus increases ANRIL expression to inhibit

senescence and promote proliferation in lesional VSMCs (134–136) and macrophages (134). This dichotomy may be analogous to the role of senescence in cancer, where senescence initially disarms preneoplastic cells but persistent SASP production promotes future carcinogenesis (8, 9, 137).

Heart failure. Although aging itself does not cause HF, the aging heart is predisposed to develop HF for multiple reasons. Adaptations that the aging heart undergoes to preserve resting systolic function, such as LV thickening (138), cardiomyocyte hypertrophy (139), and decreased response to β -adrenergic stimulation (140), deplete the functional reserve that can be deployed to deal with disease or heightened demand (41). Additionally, repair processes such as cardiomyocyte replacement are impaired (141), and others, such as fibrosis, can become exaggerated and maladaptive. Finally, further injury from ischemic disease, such as CAD, increases with age. Given that an increasing proportion of elderly HF patients maintain normal systolic function (HF with preserved ejection fraction; HFpEF), a difficult-to-treat subtype of the disease, the need for new preventative and therapeutic strategies is paramount (5).

Cellular senescence drives these three aspects of HF predisposition in the aging mouse heart (Figure 3, A and B). For example, old but not young mice show increased LV mass and diastolic dysfunction when challenged with repeated, low-dose isoproterenol, a β -adrenergic agonist (10). In contrast, old mice cleared of SNCs via INK-ATTAC do not show such dysfunction (10). SNC killing also attenuates other HF risk factors, including diet-induced atherogenesis (7) and age-related cardiomyocyte hypertrophy (10), potentially by normalizing homeostatic signaling from pericardium to myocardium (Figure 3A).

SNCs also accumulate during HF itself (Figure 3C). Aged human hearts with dilated cardiomyopathy show greater numbers of p16^{Ink4a}-positive, cKit-positive progenitor cells and cardiomyocytes with short telomeres than age-matched controls (142). Although cell division was increased in both cell types during HF and cardiomyocytes become hypertrophic, this could not match the rate of cell death or compensate for reduced function, resulting in HF (142). Similarly, human cardiac stem cells (hCSCs) from failing hearts showed increased p16^{Ink4a} and inflammatory factor expression (78). Supporting a causal role for senescence in HF, telomerase-deficient mice show enhanced cardiomyocyte death and hypertrophy leading to HF (143). Similarly, senescence-accelerated mice (SAM), a naturally progeroid mouse strain, develop HFpEF with LV hypertrophy, atrial dilation, and fibrosis when fed a high-fat/high-salt diet that drives endothelial cell senescence (144).

In addition to perhaps removing cardiomyocytes with proliferative potential from the cycling pool via cell-cycle arrest, senescence in HF may also drive signaling changes that predispose to cardiomyocyte death or dysfunction. For example, RAAS hyperactivity occurs in HF, driving aberrant ROS production and cardiomyocyte death (145–147). Age-related senescence in the kidney enhances expression of angiotensin-converting enzyme (ACE) and increases levels of the angiotensin II receptor, driving glomerulosclerosis (10). It is plausible that SNCs in the aged or failing heart also aberrantly elevate RAAS activity, but this is currently unknown. Given that ACE inhibitors are a mainstay of HF therapy (148) and RAAS hyperactivity is a key driver of age-associated

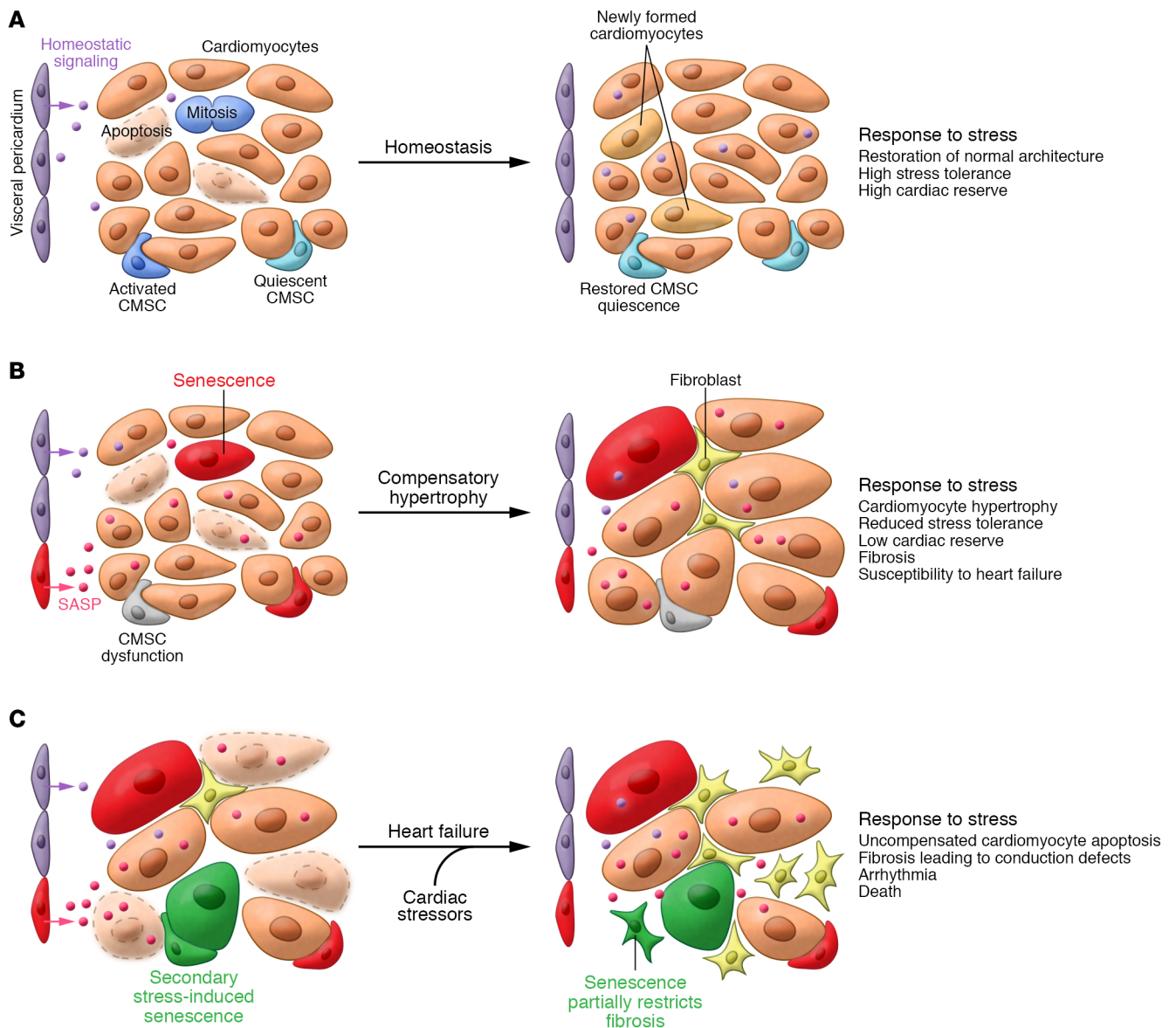


Figure 3. Senescent cells drive heart failure. (A) In young hearts, myocardium homeostasis, normal tissue function, and excess cardiac reserve are maintained through de novo formation of cardiomyocytes from resident cardiomyocyte stem cells (CMSCs) or division of incompletely differentiated, small cardiomyocytes. New cardiomyocyte formation matches a relatively low rate of cardiomyocyte apoptosis (purple cells). Appropriate CMSC function, including return to quiescence and suppression of cardiomyocyte death, may be maintained by homeostatic signals arising from the pericardium. (B) In disease-free, aging hearts, a declining ability to replace apoptotic cardiomyocytes due to progenitor dysfunction/senescence conspires with an increasing rate of cardiomyocyte death to produce compensatory hypertrophy. In the resting state and in the absence of further CVD, this adaptive change partially preserves heart function but predisposes to heart failure upon further stress. Interestingly, the INK-ATTAC SNC-killing transgenic strategy efficiently removes SA β -gal-positive pericardial SNCs from aged mice and prevents both cardiomyocyte hypertrophy and declining β -adrenergic stress tolerance. Based on this, we propose the existence of pericardium-derived prohomeostatic signaling that is disrupted during aging by pericardial SASP factors, although similar disruptive signaling may arise from senescent cardiomyocytes, CMSCs, or other cell types in the myocardium. (C) When challenged with cardiovascular stressors, such as ischemia in coronary artery disease, diabetes, or hypertension, the aged myocardium exceeds its functional reserve and decompensates. Apoptotic cardiomyocytes are not replaced, hypertrophy no longer preserves function, and excessive fibrosis leads to conduction defects, arrhythmia, and HF. To some extent, fibroblast senescence restricts fibrosis, but the long-term presence of these and other SNC types in the failing heart is suspected to be deleterious. It is currently unclear whether SNC removal can improve established heart failure or merely blunts development.

cardiovascular inflammation (149), investigating the link between senescence and the RAAS is a high priority.

HF patients often also show increased myocardial fibrosis, which interrupts normal propagation of electrical signals and drives arrhythmias (Figure 3C and ref. 150). Interestingly, the senescence cell-cycle arrest can halt expansion of fibrosis (151,

152) (Figure 3C). Mice lacking the senescence effector p16^{ink4a} or p53 showed greater fibrosis and greater diastolic dysfunction when challenged with pressure overload aortic banding than controls with an intact senescence program (153). Similarly, in post-MI ventricle, arrhythmic atria, and pressure-overloaded ventricle, p16^{ink4a}-positive and SA β -gal-positive cells are detected in

fibrotic regions (153–155). In atrial fibrillation (AF) patients, higher rates of AF recurrence and fibrosis were found in individuals with lower SNC numbers (154). Although restriction of proliferation is a plausible mechanism for the antifibrotic effect, expression of matrix-degrading collagenases or other tissue remodeling factors as part of the SASP of senescent myofibroblasts may also restrict fibrosis (37). It is unclear whether eliminating senescent fibroblasts from the failing heart would have a beneficial or a deleterious effect.

Senotherapy's potential for treating or preventing CVD

SNCs are an attractive target for preventing and treating CVD for several key reasons. First, SNCs accumulate slowly throughout the lifespan, are nondividing, and, in relatively small numbers, “poison” the tissue microenvironment, leading to organ dysfunction and heightened susceptibility to age-related disease. These characteristics suggest that SNC-killing therapies (senolytics) could be administered at infrequent intervals. Importantly, unlike cytotoxic anticancer chemotherapy, selection of drug-resistant clones is unlikely to produce therapy resistance during senolysis.

Second, the beneficial functions of SNCs are temporally restricted, allowing senotherapy's timing to selectively neutralize deleterious SNCs. For example, the senescence cell-cycle arrest assists in organ patterning (156, 157) and placental syncytiotrophoblast formation (158) during embryogenesis, which would be irrelevant for treating age-related CVD. SNC secretion of PDGF-AA (37) has also been reported to assist in wound closure. In most cases of trauma, this would simply require suspending senolytic therapy temporarily. However, in nonhealing chronic ulceration, a common malady in diabetic patients with peripheral artery disease (159), use of senolytics may be contraindicated (although whether SNCs exist in chronic wounds or have a similar function as in normal wound closure is unknown). Finally, by permitting cell-cycle arrest and then removing SNCs, senolysis may preserve select beneficial functions of SNCs, such as restriction of fibrosis in HF and tumor suppression. Accordingly, removing SNCs in old mice using the transgenic INK-ATTAC approach does not lead to greater fibrosis or cancer incidence (10).

Third, and finally, the first generation of SNC-killing molecules have been described (see Childs et al., ref. 17, for an extensive overview). Existing senolytics work by targeting prosurvival adaptations that are engaged in SNCs but not active in replication-competent or terminally differentiated cells. For example, SNCs generally resist apoptosis despite macromolecular damage, in part by upregulating the antiapoptotic proteins BCL-2, BCL-XL, and BCL-W (160–162). Inhibition of BCL-family members using navitoclax (ABT-263) licenses BAX and BAD assembly at the mitochondrial outer membrane, forming the mitochondrial transmembrane pore and releasing cytochrome *c* to trigger the apoptotic caspase cascade (163). Navitoclax has proven able to remove SNCs during atherogenesis to blunt further lesion development (7) and kill hematopoietic SNCs to rejuvenate aged or irradiated marrow (164). The related BCL-W/BCL-XL inhibitor venetoclax also clears irradiation-induced lung SNCs and skin SNCs produced by p19^{Arf} overexpression (162). Other molecules that reportedly have senolytic properties include a peptide FOXO4 mimetic, which titrates endogenous FOXO4 away from p53, triggering cytosolic

retention of p53 and SNC death (165); HDAC inhibitors (166); HSP90 inhibitors (167); piperlongumine (168); and dasatinib and quercetin (169). However, more work is needed to independently confirm the senolytic properties of these molecules and determine to what extent their healthspan benefits derive from SNC killing. For example, dasatinib and quercetin do not kill senescent IMR-90 (human fetal lung) fibroblasts (165) or reduce p16^{Ink4a} expression in aged aortas (170), but nonetheless improve vasomotor function in aged mice (170).

The major challenges currently facing therapeutic exploitation of senolysis are senolytic selectivity and potency. Local administration of senolytic drugs may help resolve both of these concerns by achieving high local drug concentrations at the site of pathology. This has been demonstrated recently for the senolytic UBX101, which reduced senescent chondrocyte load and improved disease in a murine injury-induced arthritis model following intra-articular injection (171). Similar local administration of senolytics may be of use in treating CVD. For example, sirolimus-eluting stents are frequently used during coronary artery stenting in CAD to prevent VSMC proliferation, immune cell infiltration, and vessel restenosis (172). A senolytic-eluting stent could provide the antiatherogenic benefits of systemic navitoclax administration without as many concerns about off-target effects. Alternatively, a senolytic-eluting “patch” could be applied directly to the heart surface to remove SNCs following MI.

Conclusion and outlook

In the last two decades, SNCs have emerged as bona fide drivers of aging and age-related CVD. The evidence supporting this consists of detection of SNC biomarkers in diseased tissues; alteration of disease pathophysiology following genetic disruption of the senescence program; disruption of disease course via transgene-mediated SNC killing; and modification of pathology by administration of senolytics. Collectively, these advances suggest that SNCs are a viable target for senotherapy to treat and possibly prevent CVD.

Despite our growing understanding of in vivo SNC biology, major knowledge gaps continue to hamper the translational development of SNCs as a therapeutic target. Crucially, a consensus on universal SNC biomarkers is still lacking (reviewed by Sharpless et al., ref. 173). The current gold standard is to use a panel of markers, such as high p16^{Ink4a}, SASP expression, and SA β -gal activity, to detect SNCs. However, this approach risks false negatives arising from genuine SNCs that do not express canonical markers, and false positives due to biological use of these proteins outside of senescence. For example, some speculate that macrophages recruited to SNCs as part of immune surveillance make up a large portion of p16^{Ink4a}-positivity and SA β -gal signal in aged tissue without being senescent per se (174–176).

Refining SNC biomarkers depends on building a comparative understanding of the in vivo biological properties of SNCs. In and of itself, this is a major knowledge gap. For example, how does the SASP change with SNC type, with prosenescence stress, and over time? Do all in vivo SNCs share the same susceptibility to a given senolytic, or do some SNCs favor different apoptosis-resistance mechanisms? Finding answers to these simple-to-frame questions is currently hindered by lack of tools to extract and purify in vivo

SNCs across tissues before analysis by high-throughput transcriptomic or proteomic means. While the INK-ATTAC reporter mouse has been used in several studies to separate p16^{Ink4a}-positive and -negative cell fractions, it has not been successfully applied to produce an “atlas” of SNCs.

The next generation of SNC reporter mice will hopefully surmount the major technical challenges: low SNC number in vivo, age-related autofluorescence, and the need to rapidly acquire and process isolated cells to avoid artifactual changes to cell properties. This tool should also be compatible with means of identifying cell type, such as coimmunofluorescence staining or electron microscopy, and, if possible, be inducible such that SNCs can be

tracked over time. Achieving the goal of a comparative “atlas” of SNCs across aged and diseased tissues will both clarify which SNC biomarkers are most universal and uncover new biology.

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