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Original article

Clostridium difficile outbreak caused by NAP1/BI/027 strain and non-027 strains in a Mexican hospital



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ABSTRACT

Background: *Clostridium difficile* infections caused by the NAP1/B1/027 strain are more severe, difficult to treat, and frequently associated with relapses.

Methods: A case-control study was designed to examine a *C. difficile* infection (CDI) outbreak over a 12-month period in a Mexican hospital. The diagnosis of toxigenic CDI was confirmed by real-time polymerase chain reaction, PCR (Cepheid Xpert *C. difficile*/Epi).

Results: During the study period, 288 adult patients were evaluated and 79 (27.4%) patients had confirmed CDI (PCR positive). *C. difficile* strain NAP1/B1/027 was identified in 31 (39%) of the patients with confirmed CDI (240 controls were included). Significant risk factors for CDI included any underlying disease ($p < 0.001$), prior hospitalization ($p < 0.001$), and antibiotic ($p < 0.050$) or steroid ($p < 0.001$) use. Laboratory abnormalities included leukocytosis ($p < 0.001$) and low serum albumin levels ($p < 0.002$). Attributable mortality was 5%. Relapses occurred in 10% of patients. Risk factors for *C. difficile* NAP1/B1/027 strain infections included prior use of quinolones ($p < 0.03$).

Risk factors for CDI caused by non-027 strains included chronic cardiac disease ($p < 0.05$), chronic renal disease ($p < 0.009$), and elevated serum creatinine levels ($p < 0.003$). Deaths and relapses were most frequent in the 027 group (10% and 19%, respectively).

Conclusions: *C. difficile* NAP1/B1/027 strain and non-027 strains are established pathogens in our hospital. Accordingly, surveillance of *C. difficile* infections is now part of our nosocomial prevention program.

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Introduction

Clostridium difficile infections (CDI) are the leading worldwide cause of healthcare-associated diarrhea and in some countries CDI surpass all other healthcare-associated infections (HCAI).¹ A recent prevalence survey of HCAI conducted across 183 hospitals determined that *C. difficile* was the most frequently reported infectious agent, responsible for 12.1% of all HCAI.¹

In the United States of America (USA) during 2011, 15,461 CDI cases were reported with 24.2% of cases having an onset during hospitalization. Incident CDI cases were estimated to be >450,000 with an estimated >29,000 deaths.² However, the emergence of the *C. difficile* NAP1/B1/027 strain in 2000 changed the morbidity and mortality rates associated with CDI.^{3,4}

Since 2004, the role of other emergent *C. difficile* strains causing human disease has expanded. These strains are derived from 39 different ribotypes and some *C. difficile* strains have been found to be toxin A-negative but toxin B-positive,⁵ and 027 strain was the second most common isolate responsible for CDI.⁶ Ribotype 078 was reported to have an increased prevalence,⁷ and ribotype 244 seems to cause more severe disease with higher mortality rates than rates associated with ribotype 027.^{8,9} The prevalence of other ribotypes now appears to surpass that of 027, including ribotypes 037, 018, and 078.¹⁰⁻¹²

Epidemiologic research of CDI resulting from infections with diverse *C. difficile* strains, including strain NAP1/B1/027 in developing countries, is expanding and includes data regarding hospital epidemiology, clonal spread, and dissemination across the respective countries.¹³⁻¹⁶

The present study reports on a 12-month evaluation of a CDI outbreak caused by different *C. difficile* strains including the NAP1/B1/027 strain.

Methods

Setting, study design, and study population

The outbreak described in this report occurred at the Hospital Civil de Guadalajara Fray Antonio Alcalde, an 899-bed tertiary care teaching hospital located in the city of Guadalajara, the second largest city in Mexico.

This was a case-control study of adult patients with hospital-onset CDI presenting between December 2013 and December 2014. During the study period 288 adult patients were evaluated and all patients had diarrhea defined as the passage of ≥ 3 unformed stools (Bristol scale type 5-7) within 24 or 48 h after admission.¹⁷ Case patients were defined as those with a first episode of nosocomial CDI.

Clostridium difficile toxin identification

Starting in April 2014, all stool samples were tested for *C. difficile* toxins using real-time polymerase chain reaction (PCR) (Cepheid Xpert *C. difficile*/Epi, Cepheid, Sunnyvale CA) to identify toxin-producing *C. difficile* strains, including strain NAP1/B1/027. Prior to the availability of PCR-based diagnostic

approaches all diarrhea specimens were tested by enzyme immunoassay (Meridian Bioscience, Cincinnati, OH, USA). All positive specimens were saved for future testing. All stool specimens were stored at 4°C for five days, and then frozen at -70°C. After PCR retesting, only positive samples were included in the final analysis.

Control patients

Patients without diarrhea or a positive CDI test were selected at the same time and ward that CDI patients were identified. Control patients were randomly selected across the study period. Control patients were matched to case patients at a 3:1 ratio.

Definitions

Previous hospitalization was defined as a hospital stay six weeks prior to the onset of diarrhea. Recent antibiotic therapy and steroid use were defined as exposure to these medicines six weeks prior to diarrhea onset.

Clinical severity score assessment and outcome

Patients were clinically evaluated for disease severity using the SHEA/IDSA definitions of mild, moderate, or severe disease. Serum creatinine levels were included in the definition of severe disease.¹⁸ In addition, age >60 years, fever >38.3°C, and a WBC count >15,000 were used to further define clinically severe disease.¹⁹ Patients with >2 findings were considered to have severe disease.

A poor outcome was defined as death within 14 days after CDI diagnosis. Favorable outcome was defined by survival 14 days after CDI diagnosis. Relapse was defined as a second episode of diarrhea after adequate response to therapy.

Therapy and follow-up

Therapy for CDI was administered for 10 days after an adequate response to treatment was achieved (defined as a 50% reduction of loose stools after 24 h of therapy, continuous reduction after 48 h of treatment, and no diarrhea after 72 h of treatment). All patients discharged were followed via telephone every 30 days.

Statistical analysis

The data generated were coded, entered, validated, and analyzed using the Statistical Package for Social Science (SPSS), version 22.0. Univariate analyses were used to describe significant variables among cases and controls and among individuals infected with strain 027 and individuals infected with non-027 strains. P-values were calculated using the Chi-squared test or the Fisher's exact test for categorical variables and the Student's t-test or Wilcoxon rank-sum test for continuous variables. A p-value ≤ 0.05 was considered statistically significant. Multivariate analysis: logistic regression analysis was carried out considering CDI as dependent variable and clinical and demographic data as independent variables.

Table 1 – Characteristics of CDI patients and controls, severity, outcomes, relapses, and clinical aspects of patients infected with 027 and non-027 strains.

Parameters	CDI patients (n = 79) n (%)	Controls (n = 240) n (%)	p-value	027 strain (n = 31) n (%)	Non-027 strain (n = 48) n (%)	p-value
Age (y)						
18–30	23 (29.1)	62 (25.8)	0.28	7 (23)	16 (34)	0.22
31–50	24 (30.4)	88 (36.7)	0.15	11 (35)	13 (27)	0.29
51–65	21 (26.6)	53 (22.1)	0.20	10 (32)	12 (25)	0.32
>65	11 (13.9)	37 (15.4)	0.38	3 (10)	7 (14)	0.39
Gender						
Male	51 (64.5)	148 (61.7)	0.32	21 (67.7)	30 (62.5)	0.40
Female	28 (35.5)	92 (38.3)	0.32	10 (32.3)	18 (37.5)	0.40
Underlying disease						
Any	73 (92.4)	170 (70.8)	<0.001	27 (87.1)	46 (95.6)	0.15
Malignancy	11 (13.9)	18 (7.5)	0.142	6 (19.3)	5 (10.4)	0.21
Diabetes mellitus	20 (25.3)	51 (21.3)	0.27	5 (16.1)	15 (31.3)	0.10
Chronic cardiac disease	22 (27.8)	49 (20.4)	0.112	5 (16.1)	17 (35.4)	0.05
Chronic hepatic disease	2 (2.5)	5 (2.0)	0.55	0	2 (4.2)	0.36
Previous episode of pneumonia	16 (20.2)	16 (6.7)	0.005	5 (16.1)	11 (22.9)	0.33
Chronic renal disease	23 (29.1)	43 (17.9)	0.027	4 (12.9)	19 (39.6)	0.009
Healthcare-associated exposure						
Prior hospitalization	43 (54.4)	57 (23.8)	<0.001	20 (64.5)	23 (47.9)	0.11
Prior surgery	44 (55.7)	117 (48.8)	0.173	17 (62.9)	27 (56.3)	0.54
Prior antibiotics						
Any	58 (73.4)	150 (62.5)	0.050	23 (74.2)	35 (72.9)	0.55
Betalactams	43 (54.4)	108 (45)	0.155	17 (54.8)	30 (62.5)	0.32
Quinolones	16 (20.3)	31 (12.9)	0.142	9 (39.13)	5 (10.4)	0.03
Clindamycin	10 (12.7)	38 (15.8)	0.313	4 (17.39)	6 (12.5)	0.60
Prior use of acid suppressing medication						
Proton pump inhibitors	67 (84.8)	192 (80.0)	0.21	25 (80.65)	42 (87.5)	0.30
H ₂ blocker	5 (6.3)	18 (7.5)	0.47	1 (3.23)	4 (8.3)	0.34
Prior use of steroids	17 (21.5)	16(6.7)	<0.001	8 (25.8)	9 (18.8)	0.31
White blood cells count >12,000/mm ³	37 (46.8)	63 (26.3)	<0.001	15 (48.4)	22 (45.8)	0.50
Serum creatinine ≥1.5 mg/dl	22 (27.8)	53 (22.1)	0.20	3 (9.7)	19 (39.6)	0.003
Serum albumin <3 g dl	46 (58.2)	91 (37.9)	<0.002	18 (58.1)	28 (58.3)	0.58
Initial Clinical Severity Score ≥2	69 (87.3)	–	–	28 (90)	41 (85)	0.39
Outcome						
Poor/death	4 (5)	–	–	3 (10)	1 (2)	0.16
Good/cured	75 (95)	–	–	28 (90)	47 (98)	0.16
Relapses	8 (10)	–	–	6 (19)	2 (4)	0.03

Results

Study population

The age range of CDI patients and controls were similar (Table 1). Patients >65 years of age were the minority in both groups (Table 1). There was no gender difference between cases and controls; however, males were more frequently affected with CDI than females (Table 1). The presence of any underlying disease was an important risk factor for acquiring CDI, especially a previous episode of pneumonia or the presence of chronic renal disease (Table 1). Additional risk factors associated with CDI included prior hospitalization, antibiotic or steroid use, and elevated white blood cell

counts (>12,000/mm³) combined with low serum albumin levels (<3 g/dl) (Table 1). Four (5%) patients died in the CDI group and 8 (10%) relapsed (Table 1). The incidence of CDI was 1.7 per 1000 discharges.

NAP1/B1/027 infections

Strain NAP1/B1/027 was identified in 31 (39%) patients with CDI. There were some differences between CDI resulting from infections with strain 027 and non-027 strains. The presence of chronic cardiac disease and chronic renal disease were found to be significantly more frequent in the non-027 group (Table 1). Although both groups had a similar initial severity score, more deaths and relapses were associated with strain

Table 2 – Multivariate logistic regression analysis of risk factors for CDI.

Risk factor	OR	CI 95%	p-value
White blood cells count >12,000/mm ³	2.541	1.414–4.567	0.002
Prior hospitalization	4.029	2.240–7.246	0.001
Serum Albumin <3 g dl	2.026	1.138–3.608	0.016
Previous episode of pneumonia	4.251	1.848–9.779	0.001
Prior use of steroids	5.077	2.203–11.698	0.001

027 infections (Table 1). Additional risk factors associated with 027 and non-027 infections were prior use of quinolone and abnormal serum creatinine level (>1.5 mg/dl), respectively (Table 1).

Logistic regression analysis included the significant risk factors for CDI prior use of steroids, a previous episode of pneumonia, and prior hospitalization (Table 2). Also identified abnormal white blood cell count and low serum albumin levels as independent risk factors for acquiring CDI (Table 2).

Discussion

The CDI outbreak described in this report occurred following introduction of the *C. difficile* 027 strain into our hospital by a patient diagnosed in December 2013. This individual had had multiple healthcare contacts in the USA (including several due to diarrhea) prior to being admitted to our neurosurgical ward. Introduction of *C. difficile* into a hospital will usually develop into an outbreak and previous studies have documented outbreaks following detection of strain 027.^{3,4,20}

Other Latin American countries from Central and South America have now described the presence and dissemination of *C. difficile*.^{15,21} The *C. difficile* dissemination pattern seen in Mexico was similar to that described in hospitals in the USA and Canada, but different from that of the European Union where *C. difficile* 027 is not yet as prevalent. The appearance, establishment, and dissemination of *C. difficile* in Mexico seemed to occur in large referral hospitals where a high percentage of patients admitted had multiple risk factors for CDI.^{14,22}

The presence of a serious underlying disease is a frequent risk factor for the development of CDI²³ and the presence of any underlying illness (particularly a previous episode of pneumonia) was a significant risk factor in patients compared to controls.

In our population, community acquired pneumonia was diagnosed most frequently in older patients with comorbidities including chronic obstructive pulmonary disease. These patients typically had multiple previous healthcare exposures including prior hospitalizations allowing for a greater probability of acquiring *C. difficile*. Because current guidelines of our hospital recommend administration of quinolones as empiric treatment for pneumonia²⁴ this patient group had been exposed to this drug.

A frequent risk factor in our study included patients with chronic renal disease. Since our hospital is a regional center for the diagnosis and care of patients in need of renal replacement therapy or a renal transplant it is responsible for a large

population that is affected by this underlying illness.²⁵ Similar to patients with other chronic diseases, patients with chronic renal disease have multiple healthcare contacts (including dialysis) and have multiple previous antibiotic exposures due to empiric or definitive treatment of different infectious diseases complications, including peritonitis resulting from peritoneal dialysis. Concomitant administration of steroids occurs frequently when patients have serious comorbidities and need assistance in the treatment of various complications.

The clinical features found in our CDI patients included an increased white blood cell count, elevated serum creatinine and reduced serum albumin levels. These findings are the basis for most clinical prediction rules used today.^{18,19,26-29} Patients presenting with severe CDI typically were older and had an increased number of bowel movements, had a history of systemic antibiotic use, and presented with fever, abdominal distention, abnormal respiratory rate, abnormal level of C-reactive protein, prior episodes of CDI, increased white blood cell count, elevated serum creatinine level, and low serum albumin level.^{18,19,26-29} Using these clinical prediction rules most of our patients had severe CDI.

The use of clinical prediction rules in CDI are also used to determine individuals at risk of having poor outcomes or a relapse.^{28,30-34} The most prominent factor predictive of a poor outcome or relapse among CDI patients described in this report was infection with strain 027³⁵ and chronic renal disease.

After eliminating confounders, independent risk factors for CDI included prior use of steroids, previous episodes of pneumonia, and prior hospitalizations (Table 2). The epidemiology of *C. difficile* infections is constantly changing and probably explains some of the differences found in our study compared with previous observations made in Mexico.^{14,22,36}

The diagnosis of CDI was primarily carried out using a commercial PCR kit, a test that has high sensitivity and specificity but with several limitations, including the inability of this test to identify emergent *C. difficile* variants.^{9,37,38}

The choice for initial empirical therapy in our study consisted of metronidazole. Other therapeutic choices included administration of oral vancomycin as opposed to intravenous administration combined with either intravenous metronidazole or intravenous tigecycline based on individual response to oral metronidazole.^{39,40}

In an effort to control the outbreak our intervention program focused on identifying CDI cases as quickly as possible, providing early treatment, isolating CDI cases in a dedicated ward, and restricting all quinolone use.⁴¹⁻⁴³ The presence of a disease such as CDI that is transmitted via oral-fecal contamination prompted us to reevaluate patient hand washing practices prior to each meal or the intake of oral medication, in addition to assessing hand washing practices of the staff assigned to help feed patients. This resulted in the implementation of an aggressive patient hand washing campaign.

All patients discharged after an episode of CDI received careful instructions on how to proceed should a relapse occur.⁴⁴ The instructions included a description of some of the symptoms that may present during a relapse, where to get medical attention, and what to inform healthcare personnel on arrival to clinics.

The present study had several limitations including the lack of *C. difficile* cultures to enable typing, limited use of a computed tomography scan for abdominal radiographic imaging prior to colonoscopy, colonoscopy for diagnosis of CDI, follow-up PCR testing was used only in select patients,⁴⁵ and no autopsies were performed.

In conclusion, the control of CDI in our hospital now represents a constant challenge. The control of CDI in a hospital like ours should include a tailored strategy designed to identify cases of CDI as rapidly as possible. This study represents the first description of an extended CDI outbreak caused by diverse *C. difficile* strains including the NAP1/B1/027 strain in a Mexican hospital.

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Ethical approval

This study was performed with the approval of the Hospital Civil de Guadalajara Fray Antonio Alcalde Institutional Review Board. Written informed consent approved by the Ethics Committee was obtained from all patients.

Conflicts of interest

The authors declare no conflicts of interest.

Appendix A.

Members for the Hospital Civil de Guadalajara, Fray Antonio Alcalde *Clostridium difficile* Team were: Leon-Garnica G, Castillo-Mondragon A, Camacho-Rubio JR, Rodriguez-Nuñez AJ, Mendoza-Mujica C, Heredia-Cervantes J, Mata-Esteban RA, Llamas-Alonso J, Lucio-Figueroa JO, Macias-Hernandez KZ, Macías-Bolaños DJ, Cardenas-Lara FJ, Fernandez-Ramirez A (Instituto de Patología Infecciosa y Experimental, Centro Universitario Ciencias de la Salud, Universidad de Guadalajara), Vazquez-León M, Gomez-Quiroz P. (Infectious Diseases Unit Attendings), Eduardo Ortigosa-Medrano, Gomez-Gomez K, Vargas-Garcia LF (Infectious Diseases Unit Fellows) Gutierrez-Martinez ES, Garcia-Reyes MG, Zamora-Morales S, Casillas-Pacheco MA, Martinez-Cardona L, Magaña-Ibarra S, Tello G. (Epidemiology) Rodriguez-Chagollan JJ, Anguiano-Gaytan G, Atilano-Duran MCG, Llanos-Perez E, Gomez-Quiroz A, Zavala M (Microbiology).

REFERENCES

- Magill SS, Edwards JR, Bamberg W, et al. Multistate point-prevalence survey of health care-associated infections. *N Engl J Med.* 2014;370:1198–208.
- Lessa FC, Mu Y, Bamberg WM, et al. Burden of *Clostridium difficile* infection in the United States. *N Engl J Med.* 2015;372:825–34.
- Loo VG, Poirier L, Miller MA, et al. A predominantly clonal multi-institutional outbreak of *Clostridium difficile*-associated diarrhea with high morbidity and mortality. *N Engl J Med.* 2005;353:2442–9.
- McDonald LC, Killgore GE, Thompson A, et al. An epidemic, toxin gene-variant strain of *Clostridium difficile*. *N Engl J Med.* 2005;353:2433–41.
- van den Berg RJ, Claas EC, Oiyb DH, et al. Characterization of toxin A-negative, toxin B-positive *Clostridium difficile* isolates from outbreaks in different countries by amplified fragment length polymorphism and PCR ribotyping. *J Clin Microbiol.* 2004;42:1035–41.
- Martin H, Willey B, Low DE, et al. Characterization of *Clostridium difficile* strains isolated from patients in Ontario, Canada, from 2004 to 2006. *J Clin Microbiol.* 2008;46:2999–3004.
- Goorhuis A, Bakker D, Corver J, et al. Emergence of *Clostridium difficile* infection due to a new hypervirulent strain, polymerase chain reaction ribotype 078. *Clin Infect Dis.* 2008;47:1162–70.
- Baldan R, Cavallerio P, Tuscano A, et al. First report of hypervirulent strains polymerase chain reaction ribotypes 027 and 078 causing severe *Clostridium difficile* infection in Italy. *Clin Infect Dis.* 2010;50:126–7.
- Lim SK, Stuart RL, Mackin KE, et al. Emergence of a ribotype 244 strain of *Clostridium difficile* associated with severe disease and related to the epidemic ribotype 027 strain. *Clin Infect Dis.* 2014;58:1723–30.
- Du P, Cao B, Wang J, et al. Sequence variation in *tcdA* and *tcdB* of *Clostridium difficile*: ST37 with truncated *tcdA* is a potential epidemic strain in China. *J Clin Microbiol.* 2014;52:3264–70.
- Ngamskulrungrroj P, Sanmee S, Pusathit P, et al. Molecular epidemiology of *Clostridium difficile* infection in a large teaching hospital in Thailand. *PLOS ONE.* 2015;10:e0127026.
- Baldan R, Trovato A, Bianchini V, et al. A successful epidemic genotype: *Clostridium difficile* PCR ribotype 018. *J Clin Microbiol.* 2015;53:2575–80.
- Lopardo G, Morfin-Otero R, Moran V 2nd, et al. Epidemiology of *Clostridium difficile*: a hospital-based descriptive study in Argentina and Mexico. *Braz J Infect Dis.* 2015;19:8–14.
- Camacho-Ortiz A, Lopez-Barrera D, Hernandez-Garcia R, et al. First report of *Clostridium difficile* NAP1/027 in a Mexican Hospital. *PLOS ONE.* 2015;10:e0122627.
- Aguayo C, Flores R, Levesque S, et al. Rapid spread of *Clostridium difficile* NAP1/027/ST1 in Chile confirms the emergence of the epidemic strain in Latin America. *Epidemiol Infect.* 2015;1–5.
- Quesada-Gomez C, Lopez-Urena D, Acuna-Amador L, et al. Emergence of an outbreak-associated *Clostridium difficile* variant with increased virulence. *J Clin Microbiol.* 2015;53:1216–26.
- Caroff DA, Edelstein PH, Hamilton K, Pegues DA, Program CDCPE. The Bristol stool scale and its relationship to *Clostridium difficile* infection. *J Clin Microbiol.* 2014;52:3437–9.
- Cohen SH, Gerding DN, Johnson S, et al. Clinical practice guidelines for *Clostridium difficile* infection in adults: 2010 update by the society for healthcare epidemiology of America (SHEA) and the infectious diseases society of America (IDSA). *Infect Control Hosp Epidemiol.* 2010;31:431–55.
- Zar FA, Bakkanagari SR, Moorthi KM, Davis MB. A comparison of vancomycin and metronidazole for the treatment of *Clostridium difficile*-associated diarrhea, stratified by disease severity. *Clin Infect Dis.* 2007;45:302–7.
- Johnson S, Samore MH, Farrow KA, et al. Epidemics of diarrhea caused by a clindamycin-resistant strain of *Clostridium difficile* in four hospitals. *N Engl J Med.* 1999;341:1645–51.

21. Lopez-Urena D, Quesada-Gomez C, Miranda E, Fonseca M, Rodriguez-Cavallini E. Spread of epidemic *Clostridium difficile* NAP1/027 in Latin America: case reports in Panama. *J Med Microbiol.* 2014;63:322-4.
22. Camacho-Ortiz A, Galindo-Fraga A, Rancel-Cordero A, et al. Factors associated with *Clostridium difficile* disease in a tertiary-care medical institution in Mexico: a case-control study. *Rev Invest Clin.* 2009;61:371-7.
23. Leffler DA, Lamont JT. *Clostridium difficile* infection. *N Engl J Med.* 2015;372:1539-48.
24. Mandell LA, Wunderink RG, Anzueto A, et al. Infectious Diseases Society of America/American Thoracic Society consensus guidelines on the management of community-acquired pneumonia in adults. *Clin Infect Dis.* 2007;44 Suppl. 2:S27-72.
25. Thongprayoon C, Cheungpasitporn W, Phatharacharukul P, Mahaparn P, Bruminhent J. High mortality risk in chronic kidney disease and end stage kidney disease patients with *Clostridium difficile* infection: a systematic review and meta-analysis. *J Nat Sci.* 2015:1.
26. Fujitani S, George WL, Murthy AR. Comparison of clinical severity score indices for *Clostridium difficile* infection. *Infect Control Hosp Epidemiol.* 2011;32:220-8.
27. Miller MA, Louie T, Mullane K, et al. Derivation and validation of a simple clinical bedside score (ATLAS) for *Clostridium difficile* infection which predicts response to therapy. *BMC Infect Dis.* 2013;13:148.
28. Butt E, Foster JA, Keedwell E, et al. Derivation and validation of a simple, accurate and robust prediction rule for risk of mortality in patients with *Clostridium difficile* infection. *BMC Infect Dis.* 2013;13:316.
29. Na X, Martin AJ, Sethi S, et al. A multi-center prospective derivation and validation of a clinical prediction tool for severe *Clostridium difficile* infection. *PloS one.* 2015;10:e0123405.
30. Miller M, Gravel D, Mulvey M, et al. Health care-associated *Clostridium difficile* infection in Canada: patient age and infecting strain type are highly predictive of severe outcome and mortality. *Clin Infect Dis.* 2010;50:194-201.
31. Eyre DW, Walker AS, Wyllie D, et al. Predictors of first recurrence of *Clostridium difficile* infection: implications for initial management. *Clin Infect Dis.* 2012;55 Suppl. 2: S77-87.
32. Petrella LA, Sambol SP, Cheknis A, et al. Decreased cure and increased recurrence rates for *Clostridium difficile* infection caused by the epidemic *C. difficile* BI strain. *Clin Infect Dis.* 2012;55:351-7.
33. See I, Mu Y, Cohen J, et al. NAP1 strain type predicts outcomes from *Clostridium difficile* infection. *Clin Infect Dis.* 2014;58:1394-400.
34. Rao K, Míćic D, Natarajan M, et al. *Clostridium difficile* Ribotype 027: relationship to age, detectability of toxins A or B in stool with rapid testing, severe infection, and mortality. *Clin Infect Dis.* 2015;61:233-41.
35. Scardina T, Labuszewski L, Pacheco SM, Adams W, Schreckenberger P, Johnson S. *Clostridium difficile* infection (CDI) severity and outcome among patients infected with the NAP1/BI/027 strain in a non-epidemic setting. *Infect Control Hosp Epidemiol.* 2015;36:280-6.
36. Freeman J, Bauer MP, Baines SD, et al. The changing epidemiology of *Clostridium difficile* infections. *Clin Microbiol Rev.* 2010;23:529-49.
37. Gilbreath JJ, Verma P, Abbott AN, Butler-Wu SM. Comparison of the Verigene *Clostridium difficile*, Simplex A *C. difficile* Universal Direct, BD MAX Cdiff, and Xpert *C. difficile* assays for the detection of toxigenic *C. difficile*. *Diagn Microbiol Infect Dis.* 2014;80:13-8.
38. Schroeder LF, Robilotti E, Peterson LR, Banaei N, Dowdy DW. Economic evaluation of laboratory testing strategies for hospital-associated *Clostridium difficile* infection. *J Clin Microbiol.* 2014;52:489-96.
39. Abou Chakra CN, Pepin J, Sirard S, Valiquette L. Risk factors for recurrence, complications and mortality in *Clostridium difficile* infection: a systematic review. *PLOS One.* 2014;9:e98400.
40. Bagdasarian N, Rao K, Malani PN. Diagnosis and treatment of *Clostridium difficile* in adults: a systematic review. *JAMA.* 2015;313:398-408.
41. Muto CA, Blank MK, Marsh JW, et al. Control of an outbreak of infection with the hypervirulent *Clostridium difficile* BI strain in a university hospital using a comprehensive bundle approach. *Clin Infect Dis.* 2007;45:1266-73.
42. ABBETT SK, YOKOE DS, LIPSITZ SR, et al. Proposed checklist of hospital interventions to decrease the incidence of healthcare-associated *Clostridium difficile* infection. *Infect Control Hosp Epidemiol.* 2009;30:1062-9.
43. Dubberke ER, Carling P, Carrico R, et al. Strategies to prevent *Clostridium difficile* infections in acute care hospitals: 2014 Update. *Infect Control Hosp Epidemiol.* 2014;35:628-45.
44. D'Agostino RB Sr, Collins SH, Pencina KM, Kean Y, Gorbach S. Risk estimation for recurrent *Clostridium difficile* infection based on clinical factors. *Clin Infect Dis.* 2014;58:1386-93.
45. Dubberke ER, Reske KA, Seiler S, Hink T, Kwon JH, Burnham CA. Risk factors for acquisition and loss of *Clostridium difficile* colonization in hospitalized patients. *Antimicrob Agents Chemother.* 2015;59:4533-43.