SCIENCE MEETS LIFE

MSqRob: analysis of label-free proteomics data in an R/Shiny environment

Ludger Goeminne

Promotors: Lieven Clement Kris Gevaert

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11/01/2017

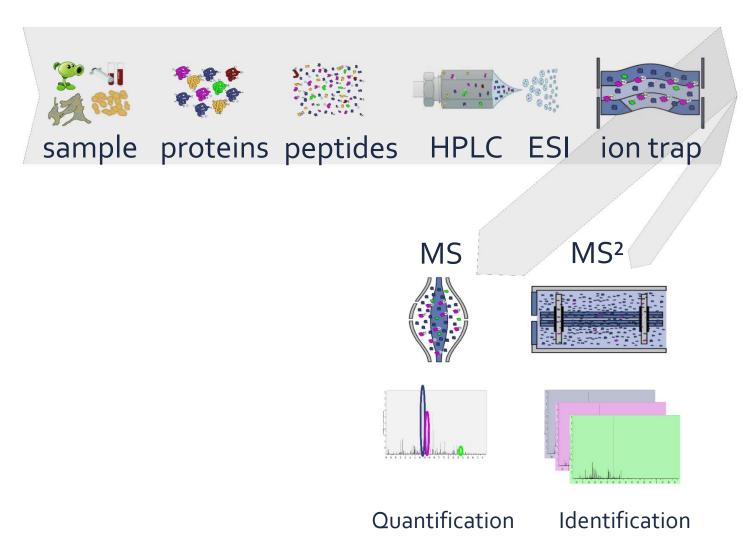




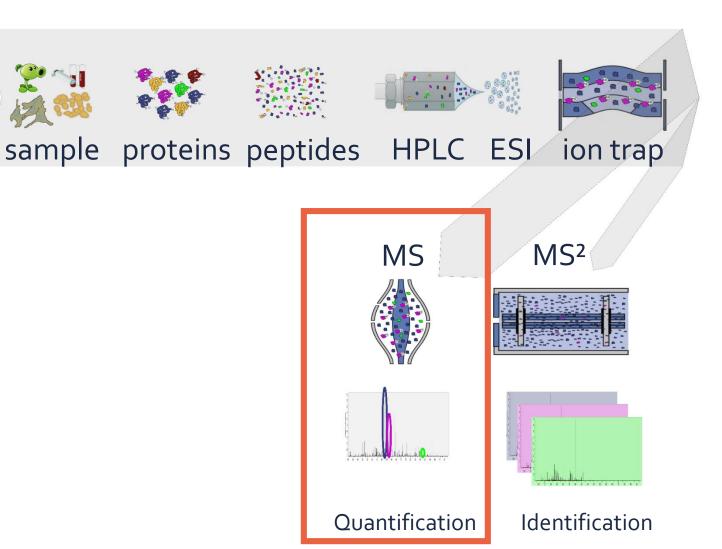
How often have you been stuck in the data analysis part?



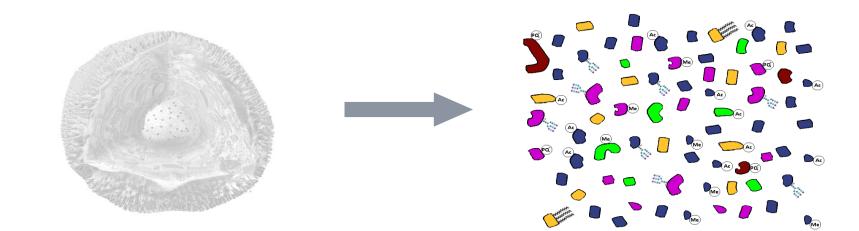
MS-based proteomics identifies many thousands of peptides



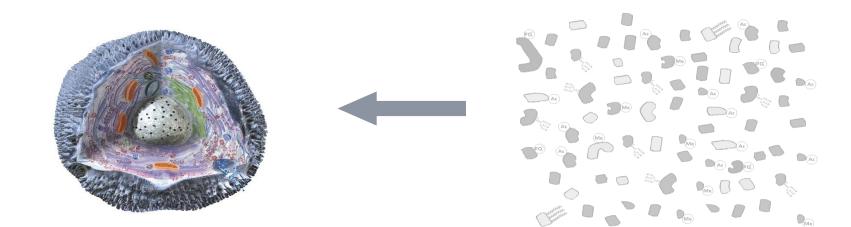
MS-based proteomics identifies many thousands of peptides



MS-based proteomics identifies peptides...



But we need protein-level information



Statistics and proteomics join forces

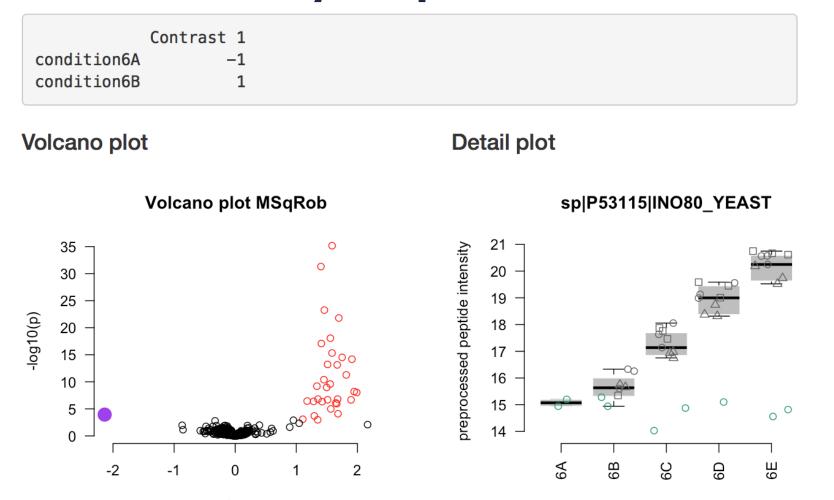


Lieven Clement: statistics



Kris Gevaert: proteomics

MSqRob can solve your data-analysis problems



Analysis of label-free proteomics data with MSqRob

Performance: why it works so well

Features: how you can use MSqRob

Analysis of label-free proteomics data with MSqRob

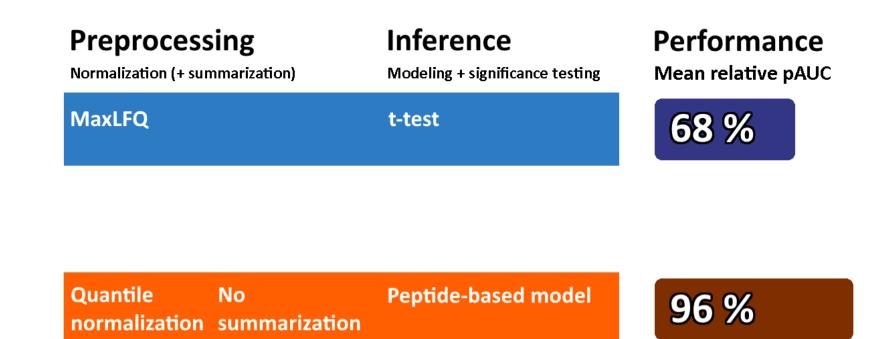
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Features: how you can use MSqRob

Peptide-based models work better than summarization-based approaches

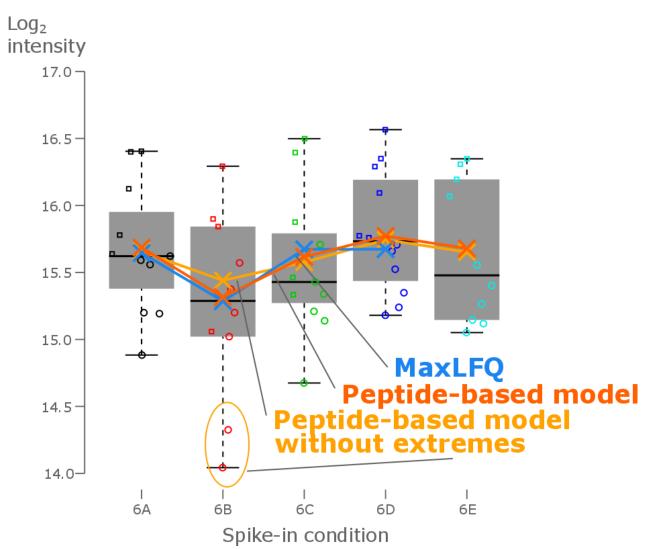


Identification



(Goeminne *et al.*, 2015, JPR)

Existing methods suffer from overfitting, unstable variances and outliers



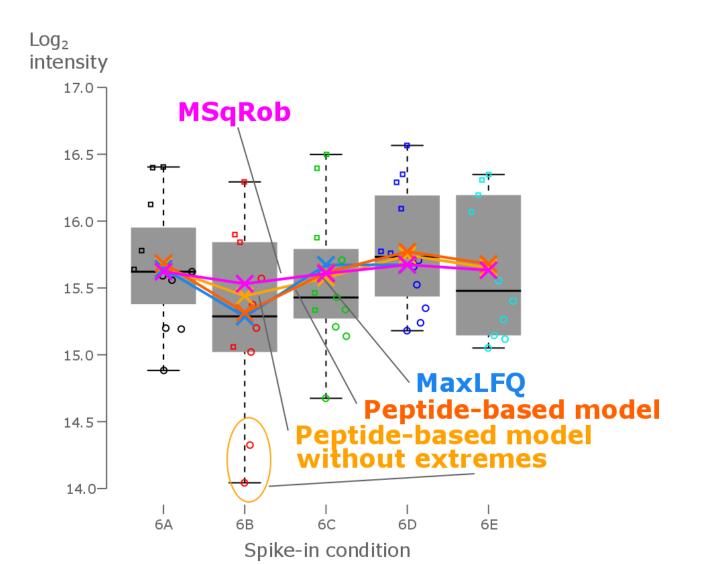
MSqRob adds 3 improvements to peptide-based models

Shrinkage estimation

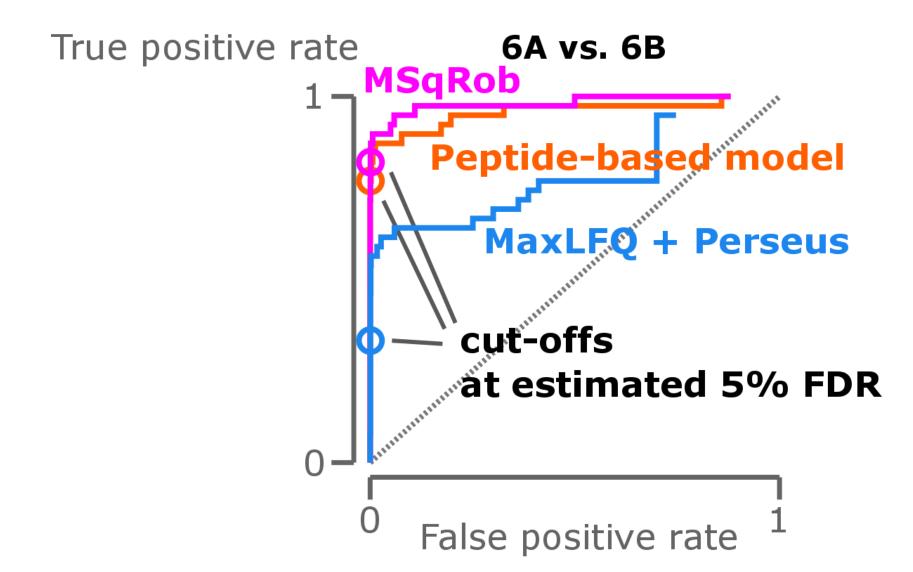
Borrowing information across proteins

Weighing down outliers

MSqRob provides more stable estimates



MSqRob outperforms other methods



Analysis of label-free proteomics data with MSqRob

Performance: why it works so well

Features: how you can use MSqRob

MSqRob handles data in a natural way

Fixed effects: genotype,treatment

Random effects: peptide, biological repeat, mass spec run

=> MSqRob can handle **complex designs**

Import from MaxQuant's peptides.txt







Project Name

project

Specify the location where your output will be saved

Browse... /Users/Igoeminn/Doc Upload complete

Specify the location of your experimental annotation file



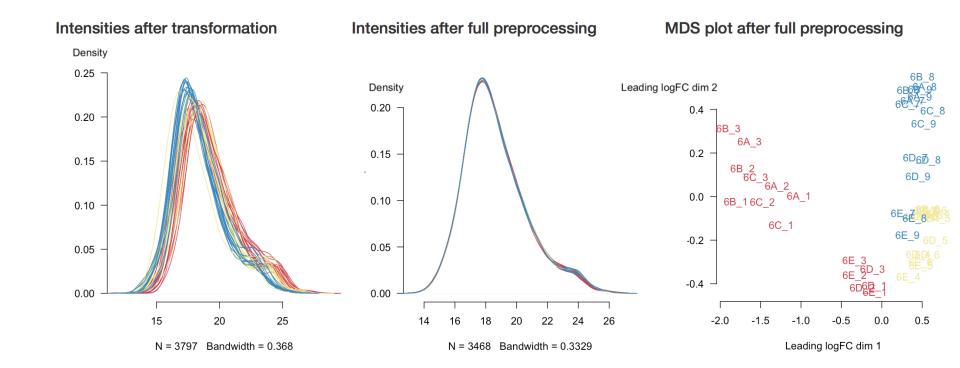
Specify the location of your peptides.txt file



Add an annotation file

A1		\cdot : \times \checkmark f_x	run
	Α	В	С
1	run	condition	lab
2	6A_1	6A	LTQ-Orbitrap_86
3	6A_2	6A	LTQ-Orbitrap_86
4	6A_3	6A	LTQ-Orbitrap_86
5	6A_4	6A	LTQ-OrbitrapO_65
6	6A_5	6A	LTQ-OrbitrapO_65
7	6A_6	6A	LTQ-OrbitrapO_65
8	6A_7	6A	LTQ-OrbitrapW_56
9	6A_8	6A	LTQ-OrbitrapW_56
10	6A_9	6A	LTQ-OrbitrapW_56
11	6B_1	6B	LTQ-Orbitrap_86
12	6B_2	6B	LTQ-Orbitrap_86
13	6B_3	6B	LTQ-Orbitrap_86
14	6B_4	6B	LTQ-OrbitrapO_65
15	6B_5	6B	LTQ-OrbitrapO_65
16	6B_6	6B	LTQ-OrbitrapO_65
17	6B_7	6B	LTQ-OrbitrapW_56
18	6B_8	6B	LTQ-OrbitrapW_56
19	6B_9	6B	LTQ-OrbitrapW_56
20	6C_1	6C	LTQ-Orbitrap_86
21	6C_2	6C	LTQ-Orbitrap_86
22	6C_3	6C	LTQ-Orbitrap_86
23	6C_4	6C	LTQ-OrbitrapO_65
24	6C_5	6C	LTQ-OrbitrapO_65
25	6C_6	6C	LTQ-OrbitrapO_65
26	6C_7	6C	LTQ-OrbitrapW_56
27	6C_8	6C	LTQ-OrbitrapW_56
28	6C_9	6C	LTQ-OrbitrapW_56

Preprocess your data



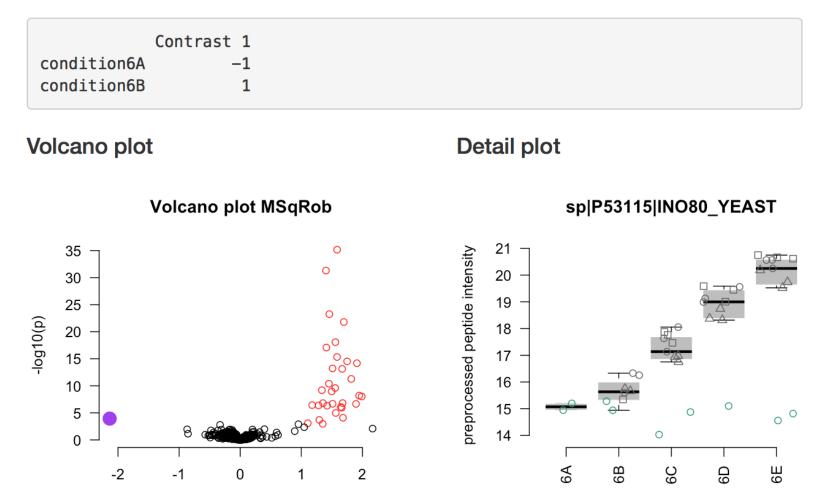
Select fixed and random effects

Select fixed effects				
condition	lab			
elect rando	om effects			

Test the appropriate research hypotheses

condition6A	
-1	٢
condition6B	
1	٢
condition6C	
0	٢
condition6D	
0	٢
condition6E	
0	٢
labLTQ-Orbitrap_86	
0	٢
labLTQ-OrbitrapO_65	
0	٢
labLTQ-OrbitrapW_56	
0	٢

Inspect the results graphically



estimate

Export the results to Excel

A	В	L	D	E	F	G	Н		J
Uniprot ID	Gene names	Protein names	Log2 fold change	se	df	Tval	pval	FDR	significant
CONTR.		er an er an de state	1.792590082	0.218879783	514.657895	8.1898385	2.079E-15	6.56682E-12	TRUI
March 1	40	Alian Martin Contractor	-4.930024672	0.363901606	31.01100511	-13.547686	1.444E-14	2.27994E-11	TRUI
and the second se		and the states and the state of the	0.711208822	0.09672183	159.1498915	7.3531365	9.62E-12	1.01271E-08	TRUI
and the second se		and the second	0.929529407	0.127592166	125.9804888	7.2851605	3.089E-11	2.43899E-08	TRUI
	201	a second state of the second state of the	0.472814156	0.083519485	296.228924	5.661124	3.553E-08	2.24381E-05	TRUI
and a second	1	and the second sec	0.551675066	0.133293731	199.7756981	4.1387923	5.145E-05	0.027080517	TRUI
TAMAT			0.210339963	0.053593612	811.0413003	3.9247208	9.42E-05	0.042496228	TRUI
and a		W. Lat.	0.630422645	0.167280773	90.14706836	3.7686498	0.0002926	0.115514005	FALSI
		and the second second second	0.291723958	0.086473092	94.09547413	3.3735807	0.0010787	0.378520358	FALSI
		Harry sales and sales and	0.288652204	0.08955748	223.0215215	3.2230943	0.001458	0.460424432	FALSI
			0.770263588	0.249837936	189.1221162	3.083053	0.0023554	0.593851302	FALS
		and the second s	0.411683714	0.134979257	313.3658685	3.0499776	0.0024839	0.593851302	FALS
S YOM		Change and the Ter of the	-0.890390597	0.253699582	17.41830988	-3.5096258	0.0026066	0.593851302	FALS
	N.		0.235832447	0.077786908	315.5706436	3.0317756	0.0026327	0.593851302	FALS
an a		- 20.	-0.305719733	0.107404642	207.8966242	-2.8464294	0.0048642	0.963224232	FALS
and the second se		a second s	0.265058666	0.093912922	833.8952564	2.8223876	0.0048802	0.963224232	FALS
and the second s			0.376058415	0.134944021	454.4736408	2.7867735	0.0055465	1	FALS
			-0.337146273	0.120273478	104.1174501	-2.8031639	0.0060369	1	FALS
and a set of the set o		the two restances we are an	0.2901366	0.108279012	84.11443528	2.6795276	0.0088677	1	FALS
		A GARLE TRADING	-0.417028694	0.155807686	83.56480618	-2.6765605	0.0089505	1	FALS
111		and a state of the	0.127083543	0.048999423	484.0927075	2.5935722	0.0097861	1	FALS
and the second sec	and the second se	Constant and the second state	0.160771955	0.062727998	238.1932984	2.5630015	0.0109929	1	FALSI
		and a second	-0.244473317	0.095897536	151.802168	-2.549318	0.0117852	1	FALSI
		and the second	0.484253154	0.189173715	71.87076044	2.5598332	0.0125755	1	FALSI
5		41.1	0.323971336	0.126270242	60.29591579	2.5656982	0.0128003	1	FALSI
	ar.	Allen and and an allen a substant	0.236609776	0.094416929	241.7741862	2.5060101	0.0128683	1	FALSI
	•		1.991348988	0.666375754	10.05944338	2.9883275	0.0135279	1	FALS
- 144.5	*	And	0.182496243	0.073579922	232.4366824	2.4802451	0.0138387	1	FALS

Download MSqRob from GitHub

https://github.com/ludgergoeminne/MSqRob

Goeminne, L.J.E., Gevaert, K. and Clement, L. **Peptide-level robust regression improves estimation, sensitivity and specificity in datadependent quantitative label-free shotgun proteomics**. Molecular and Cellular Proteomics 15(2), pp 567-668.

Analysis of label-free proteomics data with MSqRob

Performance: why it works so well

Features: how you can use MSqRob

How often have you been stuck in the data analysis part?



MSqRob can solve your problem



MSqRob





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MSqRob: analysis of label-free proteomics data in an R/Shiny environment



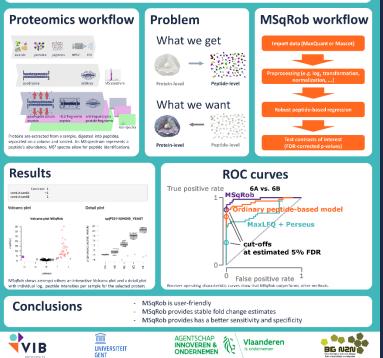
Analyzing repeated measures designs in label-free proteomics with MSqRob (MCP 2016 15(2):657-68.)



In MS-based proteomics, proteins are not completely covered and peptides that are identified in one sample are often missing in other samples. Common workflows adopt software tools that have graphical user interfaces, but are often based on less sensitive protein level abundance values and/or provide inefficient or even inappropriate statistical inference.

MSqRob is an R package that accounts for peptide-specific effects as well as differences in the number of peptide identifications. It copes with overfitting, unstable variances and outliers by three modular extensions: (1) ridge regression, (2) empirical Bayes variance estimation and (3) M-estimation. MSqRob provides state-of-the-art statistical inference for labelfree proteomics experiments with simple and complex designs: MSqRob can cope with multifactorial, block, repeated measures and time series designs, which cannot be analyzed properly in existing proteomics data analysis software. The Shiny graphical user interface for MSqRob is very user-friendly and requires no statistical programming experience. Goeminne, L.I.E., Gevaert, K. and Clement, L. Molecular and Cellular Proteomics 15(2), pp 567-668.

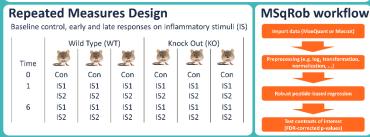
Download MSqRob: https://github.com/ludgergoeminne/MSqRob



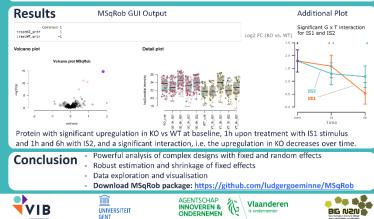
¹Modela Bioicechnology Center, VB, Ghent, Belgium; ²Oppartment of Biochemistry; ¹Department of Applied Mathematics, Computer Science and Statistics; ^{Bioinformatics institute Ghent, Biol ⁴/₂PN, Ghent University, Belgium; ²Contact: Ileven.clement@UGent.be Background: In repeated measures designs different observations are obtained on the same experimental unit (EU), which increases statistical power for within subject treatment effects because the betweensubject variability can be eliminated from the estimation. Data of the same EU, however, are typically}

Lieven Clement^(3,4,*), Ludger Goeminne^(1,2,3,4), Emmy Van Quickelberghe^(1,2,4) & Kris Gevaert^(1,2,4)

more similar than data between EUs. Most existing workflows cannot address experiments with complex designs and correlation, resulting in a power loss when assessing treatment effects within EU (e.g. compound effects) and improper error rate control for effects between EU (e.g. KO vs WT).



Model $\log_2 |$ Intensity =G×IS×T+(1|mouse)+(1|run)+(1|Peptide)+ ε Log₂ peptide intensity modeled by genotype (G). Inflammatory Stimulus (IS) & time (T) main effects & interactions + random effects for mouse, run and peptide to address correlation. Normal error.



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Lieven Clement Kris Gevaert





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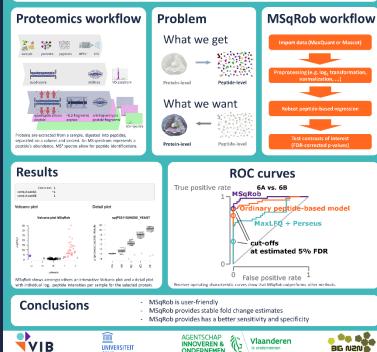


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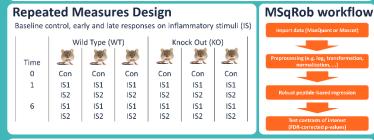
Download MSqRob: https://github.com/ludgergoeminne/MSqRob





Lieven Clement^(3,4,*), Ludger Goeminne^(1,2,3,4), Emmy Van Quickelberghe^(1,2,4) & Kris Gevaert^(1,2,4) ¹Medical Biotechnology Center, VIB, Chent, Belgium; ¹Cepartment of Biochemistry; ¹Department of Applied Mathematics, Computer Science and Statistics; ¹Bioinformatics institutes 6hent, Biol - 8470; 6hert University Legium; ¹Constant: Ileven Clement@Ucentbe

Background: In repeated measures designs different observations are obtained on the same experimental unit (EU), which increases statistical power for within subject treatment effects because the betweensubject variability can be eliminated from the estimation. Data of the same EU, however, are typically more similar than data between EUs. Most existing workflows cannot address experiments with complex designs and correlation, resulting in a power loss when assessing treatment effects within EU (e.g. compound effects) and improper error rate control for effects between EU (e.g. KO vs WT).



Model $\log_2 |\text{ntensity} = G \times IS \times T + (1 | \text{mouse}) + (1 | \text{run}) + (1 | \text{Peptide}) + \varepsilon$ $\log_2 \text{ peptide intensity modeled by genotype (G), Inflammatory Stimulus (IS) & time (T) main effects & interactions + random effects for mouse, run and peptide to address correlation. Normal error.$

