academicJournals

Vol. 4(7) pp. 98-104, November 2013 DOI: 10.5897/IJBMBR2013.0154 ISSN 2141-2154 ©2013 Academic Journals http://www.academicjournals.org/IJBMBR

International Journal for Biotechnology and Molecular Biology Research

Full Length Research Paper

Effects of harvesting stage and storage duration on postharvest quality and shelf life of sweet bell pepper (Capsicum annuum L.) varieties under passive refrigeration system

Dargie Tsegay*, Bizuayehu Tesfaye, Ali Mohammed, Haddis Yirga and Andnet Bayleyegn

*Montpellier SupAgro_Centre International d'études Supérieures en Sciences Agronomiques, 2 Place Pierre Viala, 34060 Cedex 2, Montpellier, FRANCE.

Accepted 29 October, 2013

A laboratory experiment was carried out to determine the effects of harvesting stages (0, 25, 50, 75 and 100% fruit colourations) and storage durations (0, 1, 2, 3 and 4 weeks) on physicochemical quality and shelf life of sweet pepper varieties (Telmo-Red and Velez-Yellow) under passive refrigeration system (PRS). The aim of the study was to identify the optimum stage of maturity at harvest and storage period under PRS that can ensure better quality and longer shelf life of two greenhouse sweet pepper varieties. The experiment was arranged in 2 x 5 x 5 factorial combinations in complete randomized design (CRD) with three replications. Thirty (30) fruits of sweet pepper were packed in card-board boxes for each treatment and stored under PRS optimum storage conditions. Fruits were assessed for weight loss percentage, fruit firmness, total soluble solids, titratable acidity, postharvest decay percentage and shelf life. Total soluble solids were increased; whereas fruit firmness decreased with increasing harvesting stages. Weight loss percentage, postharvest decay percentage and shelf life increased; while fruit firmness decreased with increasing storage periods. Telmo variety showed significantly better postharvest quality and storability potential than Velez variety.

Key words: Harvesting stage, postharvest, passive refrigeration system, sweet bell pepper.

INTRODUCTION

Sweet bell pepper (*Capsicum annuum* L.) is one of the most commercially important horticultural crops grown in tropical and sub-tropical regions of the world. From the nutritional point of view, peppers are generally considered as a balanced source of most of essential nutrients, high content of vitamins, important antioxidants, rich in flavonoids and phytochemicals (Maria et al., 2010). Sweet peppers are currently the object of much attention due to possible links to prevention of certain types of

cardiovascular diseases, atherosclerosis, cancer, haemorrhage, delaying of ageing process, avoiding cholesterol, improving physical resistance and increasing appetite (Marin et al., 2004).

Growing and marketing of fresh produce is complicated by high postharvest losses which are estimated to reach as high as 25-35% of the produced volume for vegetables (Agonafir, 1991). Sweet peppers like other vegetables are quite perishable, about 28.6 and 38.7% postharvest losses were reported during the dry and wet seasons, respectively (Tunde-Akintunde et al., 2005). Optimum temperature and relative humidity can be achieved using passive refrigeration system (PRS) cooling machine, which is a very efficient technique to store and transport products. The system works without ventilation thus assuring shelf life which is better than the active refrigeration system equipment. The thermal autonomy allows the storage and transport without use of power during operations (Nomos, 2008).

However, there is no available scientific literature regarding the effect of harvesting stages and storage durations on retaining the postharvest physicochemical quality properties of sweet bell pepper varieties under passive refrigeration system storage condition. The main objective of the present study was to evaluate the effect of harvesting at different maturity stages and storing in PRS, on shelf life and quality of sweet bell pepper varieties.

MATERIALS AND METHODS

Experimental design and treatments

The treatments were comprised of two varieties of sweet pepper (Telmo and Velez) picked at five harvesting stages (0, 25, 50, 75, 100% colourations) and stored for five storage durations (0, 1, 2, 3, and four weeks) under PRS. The treatments were combined in CRD factorial experiment, resulting in a total of 50 treatment combinations (2x5x5) with three replications and 150 total observations (2x5x5x3). Each treatment consisted of 30 fruits packed in standard card board boxes for storage under PRS.

Experimental procedures

Fruits of two sweet pepper varieties with similar size (160 g) and shape (bell shaped) were harvested from Hawassa Jittu Horticulture PLC greenhouse. Maturity stages of fruits were determined by fruit colouration guide and days from anthesis. Fruits were harvested manually with care to minimize mechanical injuries. After harvest, fruits were immediately transported using standard plastic crates to packing house within 10 min and held at 10°C pre-cooling room overnight. Fruits with bruises, sign of infection or those different from the group were discarded from the samples. Fruits were washed with tap water, surface dried with soft cloth and subdivided, sorted, and weighed in the packinghouse; thereafter stored under PRS (model DS-TP-001-03) on three shelves as replication. Samples were taken to food technology laboratory for quality analysis. The treatments were tested at test room environmental conditions (20°C temperature and 70% relative humidity) combined with 24 h lighting to assess the shelf life of fruits after removing from the PRS.

Data collection

Weight loss percentage (WLP)

Five sweet pepper fruits were weighed at day zero and in each storage duration using sensitive balance. The difference between initial and final weight of fruits was considered as total weight loss during storage interval and expressed as percentage (AOAC, 2007):

$$WLP = \frac{Initial - Final\ Weight}{Initial\ Weght} \times 100\%$$

Fruit firmness

Firmness of three fruits was measured using a computer-controlled automatic fruit texture analyzer (model: *TA-LEVEL-05*) according to Manolopoulou et al. (2010). The firmness measurement was carried out using a cylindrical stainless steel probe of 2 mm in diameter. Puncture tests were taken from the two opposite equatorial sides of the same fruit.

Total soluble solids (TSS)

Juice of sweet pepper fruits was extracted from three fruits in a blender as described by Antoniali et al. (2007). The homogenized sample was filtered using funnel with filter paper in a beaker. The filtrate was taken for TSS determination using digital refractometer (model: RFM-860, Japan) in °Brix by placing a few drops of clear juice on the prism surface.

Titratable acidity (TA)

10 ml of juice was extracted from three fruits and then homogenized and filtered using funnel with filter paper in a beaker. The TA was measured using NaOH (0.1 N) as a standardized titration solution. When the end point of titration was reached at pH 8.2, the amount of NaOH used on the burette was read off and recorded to calculate TA:

$$TA = \frac{\text{Titre} \times 0.1\text{N NaOH} \times 0.67}{1000} \times 100\%$$

Postharvest decay percentage (PDP)

Fruits were visually evaluated for symptoms of decay at the end of each storage interval based on the method prescribed by El-Mougy et al. (2012). Samples having symptoms of chilling injury and of diseases were counted. Pathogens causing decay were not identified.

$$PDP = \frac{Number of Decayed Fruits}{Number of Total Fruits} \times 100\%$$

Shelf life

Shelf life of fruits was evaluated by counting the number of days required to attain fruits remaining still acceptable for marketing as described by Rao et al. (2011). It was decided based on the appearance and spoilage of fruits. When 50% of fruits showed symptoms of shrinkage or spoilage due to pathogens and chilling injury, lot of fruits was considered to have reached end of shelf life.

Statistical analysis

Data were subjected to ANOVA using SAS software version 9.

Harvesting stage (%)		Mean				
	0	1	2	3	4	-
0	0.00 ⁿ	2.67 ^k	3.61 ^{hi}	4.60 ^{ef}	6.01 ^b	3.38
25	0.00 ⁿ	1.39 ^m	2.25 ^l	3.30 ^{ij}	4.54 ^{fg}	2.30
50	0.00^{n}	2.03 ^l	3.00 ^j	3.89 ^h	4.88 ^{de}	2.70
75	0.00 ⁿ	2.28 ¹	3.30 ^{ij}	4.27 ^g	5.47 ^c	3.06
100	0.00 ⁿ	3.28 ^j	4.23 ^g	5.16 ^{cd}	6.50 ^a	3.83
Mean	0.00	2.33	3.35	4.24	5.48	
LSD _(0.05)		0.33				
CV (%)		9.29				

Table 1. Interaction effect of harvesting stage and storage duration on mean weight loss percentage of sweet pepper fruits under passive refrigeration system.

Means within a column followed by same letter(s) are not significantly different at 5% LSD test.

Verification of significant differences was done using LSD test at 5% probability level.

RESULTS AND DISCUSSION

Weight loss percentage

The interaction effect of harvesting stage and storage duration on mean weight loss percentage (WLP) of sweet pepper fruits was highly significant (P<0.001); while all other interaction effects were non-significant (P>0.05). At one week of storage, mean WLP of fruits harvested at 0, 25, 50, 75 and 100% colouration stages were 2.67, 1.39, 2.03, 2.28 and 3.28%, respectively; similar trends were observed at other storage times (Table 1). Mean WLP of fruits harvested at full green stage were 0.00, 2.67, 3.61, 4.60 and 6.01 at 0, 1, 2, 3 and 4 weeks of storage, respectively; the same results were apparent at other harvesting stages (Table 1).

The highest and lowest WLP were recorded for combinations of harvested at completely ripened stage and four weeks storage as well as harvested at 25% colouration stage and one week storage under PRS, respectively (Table 1).

Across all storage periods, the WLP of sweet pepper fruits harvested at completely ripened and full green stages were significantly higher than fruits harvested at intermediate stages (Table 1). This is in agreement with the findings of Moneruzzaman et al. (2009) who observed a higher WLP in fruits harvested at early matured stage than intermediate stages. This might be due to poorly developed waxy layer and cuticle on the surface of green pepper fruits as supported by Melaku et al. (2006). The high WLP in completely ripened fruits could be due to changes in permeability of cell membranes, making them more sensitive to the loss of water as confirmed by

Antoniali et al. (2007).

Fruit firmness

The main effects of variety, harvesting stage and storage duration on mean firmness of fruits were highly significant (P<0.001); while all interaction effects were non-significant (P>0.05). The highest fruit firmness of 36.06 N was recorded for variety Telmo-Red whereas the lowest value (30.97N) was recorded for Velez-Yellow variety (Table 2). The mean firmness of fruits harvested at 0, 25, 50, 75 and 100% colouration stages were 38.41, 36.33, 33.60, 31.06 and 28.17 N, respectively. The maximum and minimum fruit firmnesses were recorded at full green and completely ripened harvesting stages, respectively (Table 2). The mean fruit firmness of sweet peppers stored for 0. 1, 2, 3 and 4 weeks under PRS were 35.75, 34.73, 33.35, 32.58 and 31.16 N, respectively. The highest and lowest values were recorded at four weeks and zero week storage periods, respectively (Table 2).

Telmo-Red variety was 14.12% firmer than Velez-Yellow variety (Table 2). This finding is in agreement with results of Lahay et al. (2013) who reported that the value of fruit firmness varied in magnitude between varieties of tomato fruits. The observed variation might be due to genetic or environmental factors as confirmed by Beckles (2012). Ilic et al. (2012) disclosed that the higher pericarp thickness of a variety, the better is the firmness of fruit.

Fruit firmness decreased with increase in harvesting stages (Table 2). The present result is in coherence with the findings of Zhou et al. (2011) who found a decrease in fruit firmness with increasing harvesting stages. The apparent decline in fruit firmness with age might be due to cell wall softening directly influencing the levels of fruit firmness. This is in line with the work of Rao et al. (2011) who found that cell wall softening is due to the activity

Table 2.	Effect	of	variety,	harvesting	stage	and	storage	duration	on	mean	fruit
firmness a	and tota	l s	oluble so	lids under p	assive	refrig	eration s	ystem.			

Variety	Fruit firmness (N)	Total soluble solids (°Brix)
Variety	Mean	Mean
Telmo-Red	36.06 ^a	7.22 ^a
Velez-Yellow	30.97 ^b	6.56 ^b
LSD _(0.05)	0.52	0.10
Harvesting stage (%)	Mean	Mean
0	38.41 ^a	5.36 ^e
25	36.33 ^b	6.40 ^d
50	33.60 ^c	7.02 ^c
75	31.06 ^d	7.63 ^b
100	28.17 ^e	8.03 ^a
LSD _(0.05)	0.82	0.16
Storage duration (Weeks)	Mean	Mean
0	35.75 ^a	6.48 ^e
1	34.73 ^b	6.88 ^c
2	33.35 ^c	7.35 ^a
3	32.58 ^d	7.07 ^b
4	31.16 ^e	6.66 ^d
LSD _(0.05)	0.82	0.16
CV (%)	4.76	4.60

Means within a column followed by the same letter(s) are not significantly different at 5% LSD test.

of softening enzymes such as pectin methylesterase.

The mean fruit firmness progressively decreased with increase in storage time (Table 2). This result is consistence with reports of Lahay et al. (2013) who found a reduction in firmness of fruits during prolonged storage periods. This could be due to high respiration rate and weight loss as supported by Cantwell et al. (2009).

Total soluble solids

The main effects of variety, harvesting stage and storage duration on mean total soluble solids (TSS) were highly significant (P<0.001); while all interaction effects were non-significant (P>0.05). The maximum TSS of 7.22 °Brix was recorded for Telmo-Red variety whereas the lowest (6.56 °Brix) was recorded for Velez-Yellow variety (Table 2). The mean TSS content of fruits harvested at 0, 25, 50, 75 and 100% colouration stages were 5.36, 6.40, 7.02. 7.63 and 8.03 °Brix, respectively. The maximum and minimum TSS contents were recorded at completely ripened and full green harvesting stages, respectively (Table 2). The mean TSS of fruits stored for 0, 1, 2, 3 and 4 weeks under PRS were 6.48, 6.88, 7.35, 7.07 and 6.66 °Brix, respectively. The highest and lowest TSS values were recorded at two weeks and zero week storage periods, respectively (Table 2).

The maximum TSS content was recorded in Telmo-

Red variety which showed 0.66 °Brix higher than Velez-Yellow variety (Table 2). This is in agreement with the results of Bernardo et al. (2008) who reported that the value of TSS varied in magnitude between varieties of sweet pepper fruits. The observed TSS variation between varieties might be due to genetic or environmental factors as confirmed by Beckles (2012).

The level of TSS content progressively increased with increase in harvesting stage (Table 2). The Mean TSS in completely ripened fruits was 2.67 °Brix higher than those harvested at full green stage (Table 2). The TSS content in this study is in line with reports of Antoniali ° (2007) who found minimum and maximum TSS values in yellow sweet pepper fruits assessed at full green and completely ripened maturity stages, respectively. The increment in TSS might be due to disassociation of some molecules and structural enzymes in soluble compounds, which directly influence the levels of TSS.

TSS content was increased during the first two weeks storage under PRS followed by a decreasing trend with increase in storage duration (Table 2). This result is in agreement with reports of Rao et al. (2011) who found an increase in TSS as fruits were stored for short period followed by a decreasing trend during prolonged storage periods. The increment in TSS for stored fruits was probably due to increase of respiration and metabolic activity. In this regard, Ali et al. (2011) found that the higher respiration rate increases the synthesis and use of

Table 3. Interaction effect of variety and harvesting stage on mean titratable acidity of sweet pepper fruits stored under passive refrigeration system.

Titratable acidity (%)									
Variety	Harvesting stage (%)								
	0	25	50	75	100	Mean			
Telmo-Red	0.56 ^c	0.62 ^b	0.69 ^a	0.51 ^d	0.39 ^g	0.55			
Velez-Yellow	0.43 ^f	0.45 ^e	0.51 ^d	0.36 ^g	0.29 ^h	0.41			
Mean	0.49	0.54	0.60	0.43	0.34				
LSD _(0.05)			0.03						
CV (%)			7.58						

Means within a column followed by same letter(s) are not significantly different at 5% LSD test.

metabolites result in higher TSS due to the higher change from carbohydrates to sugars.

Titratable acidity

The interaction effect of variety and harvesting stage on mean titratable acidity (TA) was highly significant (P<0.001); while all other interaction effects were non-significant (P>0.05). For Telmo-Red variety, mean TA of fruits harvested at 0, 25, 50, 75 and 100% colouration stages were 0.56, 0.62, 0.69, 0.51 and 0.39%, respectively; while for Velez-Yellow variety, TA of fruits harvested at 0, 25, 50, 75 and 100% colouration stages were 0.43, 0.45, 0.51, 0.36 and 0.29%, respectively (Table 3).

TA values of fruits harvested at full green stage were 0.56 and 0.43% for Telmo-Red and Velez-Yellow varieties, respectively; the same results were apparent at other harvesting stages (Table 3). The highest and lowest TA values were recorded at combinations of Telmo-Red variety and harvested at 50% colouration as well as Velez-Yellow variety and harvested at completely ripened stage, respectively (Table 3).

For both varieties, the TA values of fruits harvested at 50 and 25% colouration stages were significantly higher than fruits harvested at other stages. There was an increasing trend in TA value until fruits attained their half ripening stage and thereafter decreased with increasing harvesting stages for both varieties (Table 3).

The results are in coherence with reports of Anthon et al. (2011) who found that TA of tomato fruits was increased with maturity stages and reached the peak at half ripening stage and thereafter started to decrease. The increment in TA value might be due to the presence of pectin methylesterase enzyme activity; while the reduction in TA of fruits harvested after half ripening stage could be due to high respiration rate and reduction in organic acids as supported by Anthon and Barrette (2012).

Postharvest decay percentage

The three-way interaction effect of variety, harvesting stage and storage duration on mean postharvest decay percentage of fruits under PRS was highly significant (P<0.001). At zero and one week storage periods, all fruits of both varieties were free from any postharvest decay across all harvesting stages. At two weeks of storage, mean PDP of Telmo-Red variety harvested at 0, 25, 50, 75 and 100% colouration stages were 1.63, 0.00, 0.20, 0.90 and 2.33%, respectively; similar trends were observed at three and four weeks under passive refrigeration system (Table 4). Postharvest decay percentages of Telmo-Red fruits harvested at full green stage were 1.63, 4.45 and 5.45 at 2, 3, and 4 weeks of storage, respectively; the same results were apparent at other harvesting stages (Table 4). Similarly, at two weeks of storage, mean postharvest decay percentage of Velez-Yellow variety harvested at 0, 25, 50, 75 and 100% colouration stages were 2.78, 1.16, 1.96, 2.44 and 3.35%, respectively; similar trends were observed at three and four weeks under passive refrigeration system (Table 4).

Postharvest decay percentage of Velez-Yellow sweet pepper fruits harvested at full green stage were 2.78, 5.89 and 7.20% at 2, 3, and 4 weeks, respectively; the same results were apparent at other harvesting stages (Table 4). Starting from two weeks storage period, the highest and lowest postharvest decay percentage were recorded at combinations of Velez-Yellow variety harvested at completely ripened stage and four weeks storage as well as Telmo-Red variety harvested at 25% colouration and two weeks storage under Passive Refrigeration System, respectively (Table 4).

Starting from two weeks of storage, PDP of both varieties harvested at all maturity stages was increased with increasing storage periods (Table 4). Starting from two weeks of storage, fruits of both varieties harvested at completely ripened and full green stages had significantly higher PDP than the other harvesting stages; however it was significantly lower for Telmo-Red variety (Table 4). The present findings are in conformity with reports of Ciccarese et al. (2013) who found that PDP in fruits harvested at completely ripened stage and stored for longer period of time was always higher than fruits harvesting at intermediate stages and stored for less time. Bayoumi (2008) concluded that the higher PDP in late harvesting stage of fruits was due to higher rate of respiration, more skin permeability for water loss and high susceptibility to decay. Moneruzzaman et al. (2009) also determined that fruit PDP increases when fruits are harvested at early matured stage due to poorly developed fruit cuticular wax layer. The increment in PDP during prolonged period of time could be due to the influence of high respiration rate, fruit senescence and enzymatic degradation of fruits' cell wall (Ciccarese et al., 2013).

Table 4. Interaction effect of variety, harvesting stage and storage duration on postharvest decay percentage of sweet pepper fruits stored under passive refrigeration system.

		Postharvest decay (%) Storage duration (weeks)						
Variety	Harvesting stage (%)							
		0	1	2	3	4	Mean	
	0	0.00 ^v	0.00 ^v	1.63 ^{rs}	4.45 ^g	5.45 ^e	2.31	
	25	0.00°	0.00°	0.00^{v}	1.39 st	2.21 ^{op}	0.72	
Telmo-Red	50	0.00°	0.00°	0.20^{uv}	1.85 ^{qr}	3.23 ^{ij}	1.06	
	75	0.00°	0.00°	0.90^{u}	2.07 ^{opq}	3.45 ⁱ	1.28	
	100	0.00°	0.00°	2.33^{mo}	4.77 ^f	7.30 ^b	2.88	
	Mean	0	0	1.01	2.91	4.33		
	0	0.00 ^v	0.00 ^v	2.78 ^{kl}	5.89 ^d	7.20 ^b	3.17	
	25	0.00°	0.00°	1.16 ^{tu}	2.57 ^{lm}	2.94 ^{jk}	1.33	
Velez-Yellow	50	0.00°	0.00°	1.96 ^{pq}	2.87^{k}	3.89 ^h	1.74	
	75	0.00°	0.00°	2.44 ^{mn}	3.27 ⁱ	4.45 ^g	2.03	
	100	0.00°	0.00°	3.35 ⁱ	6.49 ^c	8.38 ^a	3.64	
	Mean	0.00	0.00	2.34	4.22	5.37		
LSD _(0.05)				0.29				
CV (%)				8.95				

Means within a column followed by same letter(s) are not significantly different at 5% LSD test.

Table 5. Interaction effect of harvesting stage and storage duration on mean shelf life of sweet pepper fruits stored under passive refrigeration system.

			Shelf life (days)			
Harvesting stage (%)			Storage duration (weeks)			
	0	1	2	3	4	Mean
0	11.17 ^r	14.00 ^{no}	19.85 ^l	26.32 ^h	30.17 ^{de}	20.30
25	14.00 ^{no}	16.17 ^m	24.84 ⁱ	31.00 ^d	36.00 ^a	24.40
50	13.34 ^p	15.83 ^m	23.31 ^j	29.50 ^{ef}	34.00 ^b	23.20
75	12.33 ^q	14.52 ⁿ	21.50 ^k	28.00 ^g	33.00^{c}	21.87
100	9.67 ^s	13.00 ^{pq}	19.00 ^l	24.82 ⁱ	29.00 ^f	19.10
Mean	12.10	14.70	21.70	27.93	32.43	
LSD _(0.05)			0.99			
CV (%)			3.97			

Means within a column followed by same letter(s) are not significantly different at 5% LSD test.

Shelf life

The interaction effect of harvesting stage and storage duration on mean overall shelf life (shelf life under PRS plus after being transferred to room temperature) of sweet pepper fruits was highly significant (P<0.001); while all other interaction effects were non-significant (P>0.05). At zero week of storage, mean shelf life of fruits harvested at 0, 25, 50, 75 and 100% colouration were 11.17, 14.00, 13.34, 12.33 and 9.67 days, respectively; similar trends were observed at other storage periods (Table 5). Mean shelf life of fruits harvested at full green

stage were 11.17, 14.00, 19.85, 26.32 and 30.17 days stored for 0, 1, 2, 3 and 4 weeks under PRS, respectively; the same results were apparent at other harvesting stages (Table 5). The maximum and minimum overall shelf lives were recorded at combinations of harvested at 25% colouration stage and four weeks storage under PRS as well as harvested at completely ripened stage and zero week storage under PRS, respectively (Table 5).

Across all storage periods, the shelf life of fruits harvested at 25 and 50% colourations were significantly higher than fruits harvested at full green and late harvesting stages

stages (Table 5). The present results are in line with the findings of Dilmacunal et al. (2011) who observed that tomato fruits harvested at breaker stage had a better storability potential under cold storage than the unripe and full red fruits. This could be due to the high weight loss percentage and respiration rate of completely ripened fruits and lack of a well developed fruit cuticular wax layer at full green stage which in turn might have resulted in lower shelf life. Moreover, the increasing trend in overall shelf life of fruits during prolonged storage period might be due to the presence of the new, modern and innovative passive refrigeration system storage equipment. This reality is supported by Shen et al. (2013) who found that refrigeration is used to reduce spoilage and extend the shelf life of fresh fruit by slowing down the metabolism and reducing fruit deterioration.

Conclusion

The postharvest quality and shelf life of sweet pepper fruits was affected by varieties, harvesting stage and storage duration. TSS content was increased while fruit firmness decreased with increasing harvesting stages. Weight loss percentage, postharvest decay and overall shelf life were found to increase; whereas fruit firmness declined correspondingly with increasing storage periods. The present results showed that Telmo-Red variety harvested at 25 and 50% harvesting stages and stored under Passive Refrigeration System storage condition could maintain better postharvest quality and extend their shelf life for more than one month.

REFERENCES

- Agonafir Y (1991). Economics of horticultural production in Ethiopia. International symposium on horticultural economics in developing countries. Acta Hortic. 270:15-19.
- Ali A, Muhammad M, Sijam K, Siddiqui Y (2011). Effect of chitosan coating on the physicochemical characteristics of Eksotika II papaya (*Carica papaya* L.) fruit during cold storage. J. Food Chem. 124:620-625
- Anthon E, Barrette M (2012). Pectin Methylesterase activity and other factors affecting pH and titratable acidity in processing tomatoes. J. Food Chem. 132:915-920.
- Anthon E, Lestrange M, Barrett M (2011). Changes in pH, acids, sugars and other quality parameters during extended vine holding of ripe processing tomatoes. J. Sci. Food Agric. 93:98-109.
- Antoniali S, Paulo M, Ana-Maria M, Rogerio T, Juliana S (2007). Physicochemical characterization of 'zarco hs' yellow bell pepper for different ripeness stages. J. Sci. Food Agric. 64:19-22.
- Association of Official Analytical Chemists (AOAC) (2007). Official Methods of Analysis of the Association of Official Analytical Chemists International. *In:Horwitz, W. (Ed.), 17th ed., AOAC Press, Arlington, VA. USA.*
- Bayoumi Y (2008). Improvement of postharvest keeping quality of white

- pepper fruits (Capsicum annuum L.) by hydrogen peroxide treatment under storage conditions. Acta Biologica Szegediensis, 52:7-15.
- Beckles M (2012). Factors affecting the postharvest soluble solids and sugar content of tomato (*Solanum lycopersicum L.*) fruit. J. Postharvest Biol. Technol. 63:129-140.
- Bernardo A, Martinez S, Alvarez M, Fernandez A, Lopez A (2008). The composition of two Spanish pepper varieties in different ripening stages. J. Food Qual. 31:701-716.
- Cantwell M, Nie X, Hong G (2009). Impact of storage conditions on grape tomato quality. ISHS Postharvest Symposium, 6th ed. Antalya, Turkev.
- Ciccarese A, Stellacci M, Gentilesco G, Rubino P (2013). Effectiveness of pre- and post-veraison calcium applications to control decay and maintain table grape fruit quality during storage. J. Postharvest Biol. Technol. 75:135-141.
- Dilmacunal T, Koyuncu A, Bayindir D (2011). Effects of Several Postharvest Treatments on Shelf Life Quality of Bunch Tomatoes. J. Not Bot. Hortic Agrobo 39(2):209-213.
- El-Mougy S, Abdel-Kader M, Aly H (2012). Effect of a new chemical formula on postharvest decay incidence in citrus Fruit. J. Plant Protect. Res. 52(1):156-164.
- Ilic S, Trajkovic R, Pavlovic R, Alkalai-Tuvia S, Perzelan Y, Fallik E (2012). Effect of heat treatment and individual shrink packaging on quality and nutritional value of bell pepper stored at suboptimal temperature. Int. J. Food Sci.Technol. 47:83-90.
- Lahay M, Devauxa F, Poole M, Seymour B, Causse M (2013). Pericarp tissue microstructure and cell wall polysaccharide chemistry are differently affected in lines of tomato with contrasted firmness. J. Postharvest Bio. Technol. 76:83-90.
- Manolopoulou H, Xanthopoulos G, Douros N, Lambrinos G (2010). Modified atmosphere packaging storage of green bell peppers: Quality criteria. J. Bio-Syst. Eng.106:535-543.
- Maria S, Zapata P, Castillo S, Guillen F, Martinez-Romero D 2010.

 Antioxidant and nutritive constituents during sweet pepper development and ripening are enhanced by nitrophenolate treatments. J. Food Chem.118:497-503.
- Marin A, Ferreres F, Tomas-Barberan F, Gil M (2004). Characterization and quantitation of antioxidant constituents of sweet pepper (*Capsicum annuum* L.). J. Agric. Food Chem. 52:3861-3869.
- Melaku K, ElkindY, Leikin-Frenkel A, Lurie S, Fallik E (2006). The relationship between water loss, lipid content, membrane integrity and LOX activity in ripe pepper fruit after storage. J. Postharvest Biol. Technol. 42:248-255.
- Moneruzzaman M, Hossain S, Sani W, Saifuddin M, Alenazi M (2009). Effect of harvesting and storage conditions on the post harvest quality of tomato (*Lycopersicon esculentum* Mill) cv. Roma VF. Australian J. Crop Sci. 3(2):113-121.
- Nomos (2008). From farms to markets. Providing know-how and finance. *International conference on sharing innovative agribusiness solutions*. Cairo, Egypt.
- Rao R, Gol B, Shah K (2011). Effect of postharvest treatments and storage temperatures on the quality and shelf life of sweet pepper. Sci. Hortic.132:18-26.
- Shen Y, Sun Y, Qiao L, Chen J, Liu D, Ye X (2013). Effect of UV-C treatments on phenolic compounds and antioxidant capacity of minimally processed Satsuma mandarin during refrigerated storage. J. Postharvest Biol.Technol. 76:50-57.
- Tunde-Akintunde T, Afolabi T, Akintunde B (2005). Influence of drying methods on drying of sweet bell pepper (*Capsicum annuum*). J. Food Eng. 68:439-442.
- Zhou R, Li Y, Yan L, Xie J (2011). Effect of edible coatings on enzymes, cell membrane integrity and cell-wall constituents in relation to brittleness and firmness of Huanghua pears (*Pyrus pyrifolia Nakai*) during storage. J. Food Chem. 124:569-575.