Accepted Manuscript

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S1568-9972(19)30115-6
https://doi.org/10.1016/j.autrev.2019.05.012
AUTREV 2335
Autoimmunity Reviews
12 January 2019
18 January 2019

Please cite this article as: K. Rahimi, A. Ahmadi, K. Hassanzadeh, et al., Targeting the balance of T helper cell responses by curcumin in inflammatory and autoimmune states, Autoimmunity Reviews, https://doi.org/10.1016/j.autrev.2019.05.012

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Targeting the balance of T helper cell responses by curcumin in inflammatory and autoimmune states

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Abstract

CD4⁺ T helper (Th) cells are a crucial player in host defense but under certain conditions can contribute to the pathogenesis of inflammatory and autoimmune diseases. Beside the Th1/Th2 subset, several additional Th subsets have been identified, each with a distinctive transcription factor, functional properties, signature cytokine profile, and possible role in the pathophysiology of diseases. These newer Th subsets include Th17, regulatory Th cells (Tregs), and more

recently, Th9, Th22, and follicular T helper cells. Interestingly, imbalance of Th subsets contributes to the immunopathology of several disease states. Therefore, targeting the imbalance of Th subsets and their signature cytokine profiles by a safe, effective and inexpensive nutraceutical agent such as curcumin could be helpful to treat autoimmune and inflammatory diseases. In this study different Th subsets and how the imbalance of these subsets could promote pathology of several diseases has been reviewed. Furthermore, the role of curcumin in this process will be discussed and the impact of targeting Th subsets by curcumin.

Keywords:

T cell; Curcumin; Inflammation; Autoimmunity

Introduction

Immune system mounts an effective and appropriate response towards several pathogens such as viruses, bacteria, fungi, parasites, tumors, allergens, and other immune stimulants [1]. T cells, the cellular base of cell-mediated immunity (CMI), are the main subtypes of lymphocytes [2]. The 2 main subtypes of T lymphocytes are CD4⁺ and CD8⁺ T [2]. T helper (Th) cells are the subpopulations of CD4⁺ T lymphocytes that is crucial in adaptive immune responses [2]. They exert distinct biological functions such as recruiting neutrophils, eosinophils, basophils, and mast cells to sites of stimulations, inducing macrophages to enhanced anti-microbial activity, helping B cells to makes antibodies, and production of chemokines and cytokines for shaping and orchestrating of various immune responses [2]. Activated CD4⁺ Th lymphocytes based on their function, transcription factors and signature cytokines are differentiated and classified into various distinct subsets such as Th1, Th2, Th9, Th17, Th22, T-regulatory (Tregs), and follicular T helper (Tfh) cells [2].

Undoubtedly, the balance between Th cytokine secretion and their responses are necessary for orchestrating the normal immune responses against intracellular and extracellular pathogens, tumor cells, tolerance to autoantigens and alteration in Th1-Th2 and Th17-Treg balance results in several diseases such as allergic and atopic disease, tumors and autoimmune and inflammatory states such as Parkinson's disease, Alzheimer's disease, inflammatory bowel disease (IBD), and

rheumatoid arthritis (RA) [2]. Therefore, tending to rebalance Th imbalance may be a therapeutic way for manipulating immune responses.

Nowadays, 25% of current drugs are made from herbal medicine and also 10% of all medications are produced from microbial sources [3, 4]. Several herbal medications such as curcumin act as an immunomodulator, thus, could be a benefit for changing the immune response [5-8]. Curcumin is used since time immemorial as a dietary spice for the management of inflammatory conditions [9]. Curcumin has shown various biological and pharmacological actions including immunomodulatory, anti-oxidant and anti-inflammatory properties [9]. Based on various in-vitro and in-vivo studies curcumin has shown therapeutic potential in several Th cells-mediated conditions [9].

Therefore, here we review the different Th subsets and how the imbalance of these subsets could promote the pathology of diseases and the potential role of curcumin in modulating these imbalances.

The role of CD4 Th lymphocyte subsets

CD4⁺ T cells have a central role in defending the body against pathogens, however, they could also be pathogenic and driving inflammation. Activated CD4⁺ Th lymphocytes were classified into different subsets including Th1, Th2, Th9, Th17, Th22, Tfh and Tregs cells [10-12]. Th1 cells produce mainly interleukin (IL)-2 and interferon-gamma (IFN- γ). It also secretes a number of cytokines such as granulocyte-macrophage-colony-stimulating factor (GM-CSF), tumor necrosis factor-beta (TNF- β) and lymphotoxin as well as various factors which trigger Th1 differentiation [10-12].

IFN- γ -secreting Th1 cells increases toll-like receptors (TLRs) expression [13], promotes class switching of immunoglobulin G (IgG) and pathogenic autoantibody production [14], increases antigen presentation through major histocompatibility complex (MHC) [15], as well as suppression and induction of over 200 genes whose functional significance remains unknown [15]. Th1 cells also stimulate the bactericidal activity of phagocytes, activate macrophages, increased phagocytosis, and responsible for cell-mediated immunity [15]. IFN- γ , the signature cytokine of the Th1 subset, is a potent pro-inflammatory cytokine which plays crucial roles in

immunopathology [16]. IFN- γ is associated with the immunopathology of different autoimmune states [16].

By contrast, Th2 cells subset produce IL-4, IL-5, IL-6, IL-9, IL-10, IL-13, IL-25, and Amphiregulin [2]. Th2 cells cause antibody production, eosinophil activation and recruitment, and inhibition of various macrophage functions [2, 17]. They also defense host against extracellular parasites including helminths as well as involves in the induction and persistence of various allergic and atopic diseases [2, 17].

The dichotomy of the Th1/Th2 cells axis was broken down after the identification of Th17 and Tregs cells [2]. Th17 cells are characterized by expression of the transcription factor retinoid-related orphan receptor- γ t (ROR γ t) and secrete potent pro-inflammatory cytokines [18]. TGF- β and inflammatory cytokines have pivotal roles in the production of Th17 cells [19]. However, under certain states, these cells are responsible for the central nervous system (CNS) inflammatory demyelination and several inflammatory and autoimmune diseases, including asthma and allergy, RA, SLE, MS, IBD and psoriasis. [19].

The other subset of CD4⁺ T lymphocytes is Tregs cells which is characterized by a CD4⁺, CD25⁺, and Forkhead box P3 (FoxP3⁺) transcription factor (CD4⁺CD25⁺FoxP3⁺) expression [20-22]. Treg cells suppress inflammation and autoimmunity by inhibiting immune responses and autoreactive effector T cells. A reduction in function and/or frequency of these cells contributes to conditions such as enteropathy, polyendocrinopathy, immune-dysregulation and X-linked syndrome (IPEX) [23-25]. Treg and Th17 cells are potentially associated with protection, pathogenesis, and progression of autoimmunity [26, 27]. Tregs have a protective role in autoimmune conditions, whereas Th17 cells enhance the progression of autoimmune diseases [26-28]. The Th cells subpopulation balance is critical for modulation of the immune system. Targeting the Th cells imbalance by safe and effective nutraceuticals may provide clinical efficacy in the conditions such as autoimmune diseases, asthma and allergy, inflammation, and organ transplantation.

An overview of the immunomodulatory properties of curcumin

Interest in the use of nutraceuticals and phytochemicals are growing nowadays so that 25% of current drugs are made from herbal medicines [29, 30]. Curcumin is a polyphenolic compound

which first described in 1910 [29, 30]. Multiple studies have demonstrated that curcumin is safe and effective in humans [30-34]. Curcumin possesses diverse pharmacological functions such as immunomodulatory and anti-inflammatory activities [30-34]. The immunomodulatory activities of curcumin which protect against immune-mediated diseases are due to its interaction with immune system cells [35-44]. Curcumin also interacts with several transcription factors and signal transducers [44, 45]. Curcumin reduces the numbers of neutrophils and eosinophils and increases the lymphocyte and NK cells count [35-44]. Curcumin also increases macrophage migration and infiltration as well as inhibits the activation of NF-κB [44, 45]. Moreover, curcumin reduces M1 macrophage polarization and induces M2 macrophage polarization [44, 45]. The functional properties and multiple pharmacological activities of curcumin on the immune system originate from NF-κB suppression and preventing the nuclear translocation of NF-κB p65 subunit [44, 45]. NF-κB has a crucial role in producing proinflammatory cytokines [44-46]. Therefore, curcumin and other NF-κB inhibitor agents are able to modulate and decreases the production of pro-inflammatory mediators. [45-47]. Curcumin also modulates immune responses by modulating IgG, IgM, and sIgA production.

Therefore, curcumin modulates the immune system by interacting with immune cells and potentially might have a therapeutic role in various inflammatory and autoimmune conditions. We have reviewed the therapeutic effect of curcumin by restoring the balance between various Th subsets.

Effects of curcumin on Th subset balance

There is growing evidence that curcumin regulates the function and proliferation of immune cells (**Fig.1**). Investigation of curcumin on modulating the immune response [37] showed that the development of CTLs and the proliferation of splenic lymphocytes is significantly reduced by curcumin [37]. Curcumin inhibits the activation of NF-kB irreversibly, thereby impairing the expression/production of TNF- α and IL-12 by peritoneal macrophages and IFN- γ and IL-2 by splenic T lymphocytes [37]. Curcumin significantly reduced IL-12 production by heat-killed Listeria monocytogenes (HKL)-stimulated macrophages or lipopolysaccharide (LPS). Curcumin administration inhibited IL-12 production by HKL or LPS mediated macrophages stimulation which leads to Th1 cytokine profile inhibition [48]. This suggests that curcumin suppresses Th1

cytokine production by inhibiting IL-12 production thereby demonstrating its potential in the Th1-mediated disease states.

It is also a powerful inhibitor of T-lymphocytes proliferation (**Fig.1**). Curcumin inhibited IL-2dependent DNA synthesis by mouse $CD4^+$ T-lymphocytes, IL-2 and CD25 expression, demonstrating a suppressive effect on the IL-2 receptor (IL-2R) signaling. In addition, $CD4^+CD25^+$ regulatory T-cells treatment with curcumin down-regulated FoxP3 expression as well as Treg cells suppressor function [49].

Evidence suggests that curcumin shifts the Th1/Th2 balance towards Th2-type response in human peripheral mononuclear cells (PBMCs) [50]. Curcumin supplementation impairs Th1/Th17 cells differentiation during encephalomyelitis whilst induces Th2 cells differentiation. Feeding with 2% curcumin produced an anti-inflammatory phenotype in DCs [51]. These findings demonstrate that curcumin silences Th17/IL-23-mediated immunopathology through enhancing STAT-3 interaction in DCs.

When human $CD4^+$ T cells were stimulated with anti-CD2/CD3/CD28 and treated with curcumin, it inhibited cell proliferation, differentiation, and cytokine production suppressed $CD4^+$ T cell activation (**Fig.1**). In addition, curcumin increased CCR7, L-selectin, and TGF- β expression at the late phase of T cell activation. Moreover, TGF- β is involved in curcumin-mediated late-phase Treg cells generation and the regulation of T cell activation (**Fig.1**). Curcumin trough augmenting CCR7, CD69, TGF- β and L-selectin expression followed by Treg cells generation not only blocks but regulates CD4⁺ T cell activation [52] (**Fig.1**).

CD4⁺CD25⁺ Treg cells have a pivotal role in regulating the immune responses and critical to the maintenance of immunological tolerance to peripheral self-antigens and suppressing deleterious immune responses to the host [27], whereas Th17-cells contribute to the development, induction and progression of auto-inflammatory disorders [53]. Breaking the balance between Treg cells and Th17-cells was favoring the pro-inflammatory Th17 responses and recently implicated in several autoimmune diseases [53] raising the possibility of discovering orally available small molecules which can modulate the function of Th17 cells or the downstream signaling of the IL-17 receptor [54]. Therefore, inhibiting the master regulator of Th17 cells, RORyt, is a potential therapeutic target for the treatment of inflammatory and autoimmune disorders [54]. The role of Th17 cells in the immunopathology of neuroinflammatory disorders such as MS, AD, PD, and

schizophrenia has been clarified recently [55]. The expression of the IL-17A and ROR γ t, a Th17 cell related cytokine profiles, was significantly reduced in curcumin-treated groups. These findings showed that curcumin inhibited the induction, differentiation, development, and progression of Th17 cells in Th17-related disorders [56] (**Fig.1**).

Curcumin supplementation inhibits the suppression of CD4⁺CD25⁺ Treg cells [57]. According to the role of curcumin in modulating Treg cells populations and function, it may be feasible in some cases to use curcumin as an immunotherapeutic agent for Treg-mediated diseases, such as sepsis and tumors. These results collectively indicate that curcumin is an immunomodulatory and immunosuppressive agent which inhibits cell-mediated cytotoxicity (CMC), cell proliferation, and pro-inflammatory cytokines production through inhibiting NF-kB activation and its nuclear translocation (**Fig.1**).

Curcumin on Th balance in autoimmune disease

Autoimmune diseases are mainly caused by the immunological intolerance to self-antigens [27]. Myasthenia gravis (MG) is mainly caused by loss of self-tolerance, which allows the development of autoreactive lymphocytes [58]. The exact mechanisms involved in the failure of self-tolerance in autoimmune diseases have not been fully elucidated, one possible mechanism is deficits in numerical, functional, and migratory properties of Tregs cells [58]. When the safety and efficacy of curcumin in experimental autoimmune myasthenia gravis (EAMG) were investigated, curcumin administration ameliorated the clinical manifestation of EAMG, down-regulated the expression of MHC class II and CD80 and CD86 co-stimulatory molecules, suppressed the levels of pro-inflammatory cytokines and up-regulated the levels of anti-inflammatory cytokines [59]. Curcumin administration shifted the balance of Th1/Th17 toward Th2/Treg and promoted the differentiation of B cells into a subset of B10 cells [59]. Therefore, curcumin effectively ameliorates the clinical score of EAMG and is a potential management option for the treatment of MG.

IPEX syndrome is an autoimmune disease in boys which is lethal and is caused by mutations in the FoxP3 gene scurfin (scurfy) [60, 61]. The product of the Scurfin gene is required for the development of CD4⁺CD25⁺ Treg cells [61]. Currently, there are no effective therapies for IPEX and immunosuppressive therapy in patients is generally ineffective and causes several side effects [60, 61]. Curcumin administration via suppressing the Th1/Th2/Th17 responses in mice

attenuated the scurfy-induced immune disorder, a model of IPEX syndrome [62]. Curcumin administration reduced all of the Th1/Th2/Th17 cell populations and attenuated clinical symptoms of scurfy mice.

A good animal model for MS is an experimental autoimmune encephalomyelitis (EAE). It is a Th1 cell-mediated autoimmune demyelinating disease of the nervous system [63]. IL-12 trough inducing Ag-specific Th1 cells has a crucial role in the pathogenesis of CNS demyelination in EAE model and MS patients. Curcumin exhibits an inhibitory effect on the generation of proinflammatory cytokines by human macrophages and monocytes ages and has inhibited the STAT-4 activation and IL-12 production in the EAE model [44]. Curcumin by reducing the IL-12 production from macrophage/microglial cells and suppressing the neural Ag-specific Th1 cells differentiation significantly reduced the clinical severity and duration of EAE [44]. The EAE mice express higher levels of IFN- γ , IL-12, IL-17, and IL-23 in the CNS and lymphoid organs which significantly decreased following curcumin treatment. Curcumin treatment resulted in a decrease in the secretion of IFN- γ , IL-12, IL-17, and IL-23 in-vitro and in-vivo [64] (Table.1. and .2). Curcumin also up-regulates the CD4⁺CD25⁺ FoxP3⁺ Treg cells populations, peroxisome proliferator-activated receptor γ (PPAR γ) and IL-10 expression in the lymphoid and CNS organs of EAE mice [64] (Fig.1). The Janus kinase-STAT pathway inhibition by curcumin markedly decreased Th1 differentiation and T cell proliferation by inhibition of IL-12 [65-67] (**Fig.1**).

Effects of nanocurcumin supplementation on Th17 cells frequency, transcription factor expression, and Th17-related cytokines expression in healthy controls and patients with relapsing-remitting multiple sclerosis (RRMS) and also from patients whose received placebo at baseline and after 6 months of treatment were investigated [68]. Results showed that Th17 cells population in MS patients when compared with the healthy control group significantly increased. The nanocurcumin group showed a significant reduction in Th17 associated parameters such as Th17 proportions, ROR γ t, and IL-17 expression and also IL-17 secretion, but mRNA expression and protein levels of IL-23 were not significantly changed. While, in the placebo group, there are no significant changes in these parameters. Results imply that the increase in Th17 cells proportion might contribute to the pathogenesis of RRMS and nanocurcumin is able to restore these Th17 cells dysregulated populations in MS patients [68]. These findings showed that

curcumin differentially regulates CD4⁺ T helper cell responses, reduced the duration and clinical severity of EAE by inhibiting IL-12 signaling in T cells and suggest its use in the treatment of MS and other Th1 and Th17-mediated autoimmune conditions [67-70]. Therefore, curcumin by influencing the Th1 and Th17 balance and secretion of pro-inflammatory mediators is a novel potential natural therapeutic agent for MS treatment.

Curcumin as a natural immunosuppressant agent has a bright prospect for preventing periodontitis and RA [71]. Periodontitis is a rising public health problem as well as a risk factor for the development of RA. It is mainly triggered by *Porphyromonas gingivalis* [72]. *P. gingivalis* in humans and animal models is able to induce experimental autoimmune arthritis through its unique bacterial peptidyl arginine deiminase [73, 74]. Curcumin appears to be an effective anti-bacterial agent against *P. gingivalis* biofilm formation and infection [71]. Curcumin also exerts unique immunosuppressant properties through the promotion of Treg cells population and inhibition of Th17 pro-inflammatory responses suppressing autoimmunity [71] (**Fig.1**). Results affirmed that curcumin as a natural product is a potent protective compound for the management of both periodontitis and also RA.

Curcumin has been shown to ameliorate various nicotine-induced chronic inflammatory disorders [75]. The cytokines profile of Th1 and Th2, mRNA and protein expression of cytokines, transcription factors (AP-1), regulatory molecule (P53), growth factors (GM-CSF; TGF- β) in female Wister rats were determined to understand the mechanisms of curcumin action in this area. Data showed that the nicotine through up-regulation and down-regulation of different factors induces Th1/Th2 balance disruption. The study demonstrated that this Th1/Th2 imbalance was safely and effectively restored by curcumin administration [76].

The Th subsets imbalance is implicated in the pathogenesis of SLE [12]. Curcumin was added after stimulating $CD4^+$ T cells from untreated SLE patients and healthy volunteers with Th17 differentiating factors. After curcumin administration Th17 percentages decreased, IL-17A productions reduced, Treg cells populations and subsequently TGF- β 1 productions increased. These findings were not observed in CD4⁺ T cells cultures of individuals without SLE [77]. This shows that curcumin could rebalance Th17/Treg imbalance of SLE patients.

Effects of curcumin on Th subset balance in psoriasis

Psoriasis is an inflammatory disease of the skin which is characterized by keratinocytes hyperproliferation and leukocytes infiltration [78, 79]. Recent evidence showed that several cytokines are involved in their pathogenesis [78-81]. It was suggested that the IL-17A/IL-23 cytokine axis have an important role in the pathophysiology of psoriasis [80, 81]. Curcumin besides its antibacterial and antioxidant activity is also able to decrease the expression of pro-inflammatory cytokines in keratinocytes [82-84]. Results showed that the curcumin gel formulation when used topically effectively suppressed imiquimod (IMQ), -induced psoriasis-like inflammation[83, 84]. In addition, curcumin by inhibiting IL-1 β /IL-6 production indirectly down-regulated IL-17A and IL-23 expression [83, 84]. Thus, curcumin is able to ameliorate inflammation induced in the IMQ-induced psoriasis-like mice model by directly down-regulating pro-inflammatory cytokines production.

Effects of curcumin on Th1/Th2 and Th17/Treg balance in IBD

IBD is a disease of the gastrointestinal tract which mainly divided into two major subgroups: Ulcerative Colitis and Crohn's disease [85, 86]. Ulcerative colitis is an inflammatory disorder of the mucosa of the colon that affects millions of people worldwide and impairs their quality of life. While the etiology of IBD is not completely determined, it appears to be induced by proinflammatory cytokines such as TNF- α [85, 86]. Emerging evidence shows intriguing pharmacological effects of curcumin [44, 45, 47]. These include inhibitory effects on inducible nitric oxide synthase (iNOS), Cyclo-oxygenase (COX)-1 and-2, lipoxygenase (LOX), TNF-a, IFN- γ , and the NF-kB [44]. NF-kB is an essential factor for up-regulation of pro-inflammatory cytokines that increase in inflammatory diseases [44]. Recently, the safety and therapeutic effects of curcumin have been evaluated in various experimental models of IBD [44]. Curcumin changes the cytokine profiles of Th1 cells toward the anti-inflammatory Th2 type [87]. The safety profile and inhibitory effects of curcumin on major pro-inflammatory mediators suggest that it has a promising future for the treatment of patients with IBD [87]. Mitogen-activated protein kinases (MAPKs) and the c-Jun N-terminal kinase (JNK) modulate the transcription and expression of several genes that are involved in the inflammatory process of various chronic inflammatory diseases such as ulcerative colitis [85, 86]. Trinitrobenzene sulphonic acid (TNBS)-induced colitis, an animal model of IBD, has been described as Th1-mediated inflammation. The TNBSinduced colonic injury was also characterized by an increase of the TNF- α and reduction of the

IL-10. Curcumin decreased the expression of Th1-related cytokines and increased the expression of Th2-related cytokines in the mucosa of the colon [88]. Curcumin exerted therapeutic effects on colitis by modulating the Th1/Th2 balance and regulating the shift from Th1 to Th2 responses in TNBS-induced colitis [88]. Curcumin reduced the p38 MAPK activation and decreased COX-2 and iNOS expression. Also, the levels of TNF- α were significantly decreased and the IL-10 production was significantly increased in curcumin-treated rats [89, 90]. CD4⁺ T lymphocytes proliferation was induced and secretion of Th2-related cytokines such as IL-4 and IL-5 were significantly increased by ConA and reduced by curcumin [91] (Fig.1).

In colitis mice, curcumin notably increased CD4⁺CD25⁺FoxP3⁺ Treg cells, while the secretion of TNF- α , IL-2, IL-6, IL-12 p40, IL-17 and IL-21 and the expression of co-stimulatory molecules such as TLR4, CD54, CD205, CD252, CD254, and CD256 on DCs were significantly decreased (**Fig.1**). Curcumin potentially suppresses DCs maturation, induce tolerogenic option, and modulates DCs activation to enhance the inhibitory functions of Treg cells in colonic mucosa of the IBD model [92]. Also, curcumin Nanoparticle administration increased CD4⁺CD25⁺FoxP3⁺ Treg cells expansion in the colonic mucosa of experimental colitis [93]. In conclusion, the inhibition of NF- κ B expression and p38MAPK signaling by curcumin could explain the reduced COX-2, iNOS, and inflammatory cytokines (TNF- α , IL-2, IL-6, IL-12 p40, IL-17 and IL-21) expression in colonic mucosa and reducing the development of chronic experimental colitis. Therefore, in animal models of IBD and in humans, curcumin has demonstrated therapeutic potential.

Effects of curcumin on Th1/Th2 and Th17/Treg balance in asthma and allergy

Asthma and allergy worldwide increased particularly in industrially developed countries over the last few decades. Thus, recent progress has been made to explore novel alternative therapies includes herbal medicines. Curcumin has traditionally been used and recently has been shown to be effective in a treat and ameliorating inflammatory and immune-mediated disorders such as fever, pain, asthma, bronchitis, arthritis, dermatitis, and allergic inflammation [94]. A murine model of latex allergy showed an increase in the Th2 type of immune responses [95]. Latex-sensitized mice showed an increased in serum IgE, latex specific IgG1, IL-4, IL-5, IL-13, eosinophils count, and lungs inflammation [95]. Treatment of Latex-sensitized mice with curcumin showed a significant reduces in Th2-related responses with a simultaneous reduction in

eosinophils count. Inflammation of lung in curcumin-treated mice was markedly reduced and expression of co-stimulatory molecules on macrophages and DCs were significantly decreased [95]. Further, the anti-asthmatic properties of curcumin, through the intranasal route in an ovalbumin (OVA)-challenged asthma mouse model has been evaluated. Exposure to LPS by increasing IgE level, Th2-related cytokine IL-4, and IL-5 expression, and histamine release, exacerbates airway inflammation. administration of curcumin through the intranasal route reduced airway inflammation and exacerbation, whereas dexamethasone, was not as good as curcumin [96].

The effect of curcumin on the lymphocytes of atopic asthmatic patients was investigated. Lymphocytes stimulated with Dermatophagoides farinea in the absence or presence of curcumin. The protein levels of IL-4, IL-5, and GM-CSF in the culture supernatants were determined. Curcumin inhibited Dermatophagoides farinea-induced lymphocyte proliferation and IL-2 production (**Fig.1**). IL-2 administration in the presence of curcumin restored the proliferative capacity of lymphocytes to Dermatophagoides farinea. Moreover, curcumin in a dose-dependent manner inhibited IL-4, IL-5, and GM-CSF production [97].

The protective effects of curcumin on food allergies in OVA and alum immunized mice were investigated. Results showed that curcumin significantly attenuated increased levels of IgE, IgG1, and MCP-1 in OVA-immunized mice. Furthermore, curcumin as an anti-allergic agent showed an immunosuppressive and immunoregulatory effect through modulating Th1/Th2 balance [98]. Also, curcumin strongly suppressed Th17 cells expansion and increased Treg cells populations (**Fig.1**). Curcumin substantially decreased IL-17A level and increased IL-10 level, blocked OVA-induced increases in eosinophils and significantly rebalanced Treg/Th17 imbalance in bronchoalveolar lavage fluid and in the lung [99]. Curcumin administration significantly attenuated the inflammatory conditions in the asthma model by regulating Treg/Th17 balance [99]. These results suggest that curcumin administration as a therapeutic drug may be useful for controlling amelioration of Th2-mediated allergic diseases.

Effects of curcumin on Th1/Th2 and Th17/Treg balance in organ transplantation

DCs and Treg cells are essential factors in the development of organ transplant tolerance. Curcumin arrests DCs maturation and differentiation and induces a tolerogenic phenotype which increases CD4⁺CD25⁺FoxP3⁺ Tregs cells populations and suppressive functions in-vitro and in-

vivo [100] (**Table.1. and .2**). Curcumin-treated DCs demonstrated a significant reduction in costimulatory molecules expressions. Curcumin-treated DCs also decreased their IL-12 mRNA expression and protein secretion. Functionally, the allostimulatory capacity of DCs and intracellular IFN- γ expression in response to T cells stimulation significantly decreased. Hyporesponsiveness of T cell was arising from the generation of CD4⁺CD25^{hi}CD127^{low} FoxP3⁺ Treg cells phenotype that put on suppressive effects on naïve CD4⁺T cells, although the effect was not specific to the antigen. DCs treated with curcumin promoted the development of CD4⁺CD25⁺FoxP3⁺ Tregs and reduced subsequent alloproliferative capacity in mice [100].

The effect of curcumin in the prevention or progression of acute graft-versus-host disease (GVHD) in a murine model were investigated [101] (**Table.1. and .2**). When treated with curcumin the acute GVHD mice showed a reduction in the severity of acute GVHD. While flow cytometric analysis showed that in the curcumin-treated acute GVHD mice the number of CD4⁺CD25⁺FoxP3⁺ Tregs cells populations, as well as the CD8+ Treg cells, were notably increased [101]. Therefore, these results collectively revealed that curcumin by up-regulation of Treg populations percentage and functions has protective effects on acute GVHD.

Curcumin on Th1/Th2 and Th17/Treg balance in cancers

Tumor cells escaping from the immune system are mostly targeted and inhibited by effector and memory T cells function [102-104]. Curcumin, by targeting several molecular molecules and transcription factors, has shown great antitumor activity. Several pieces of evidence have shown the immune-protective potential of this natural polyphenol against tumor cells [105, 106]. Effect of curcumin on prevention of tumor-induced immune dysfunction was examined and results showed that in tumor-bearing hosts, curcumin inhibited loss of T-cell populations, expanded central and effector memory T cell percentage, reversed the Th2-related immune responses and prevented T-cell proliferation inhibition [107]. Moreover, tumors up-regulated Treg cell populations and induce the production of the immunosuppressive cytokines such as TGF- β and IL-10 [108]. Curcumin also enhances CD8⁺ T cells cytotoxicity against tumors by changes in the tumor microenvironment. Moreover, results showed that the accumulation and function of T-cells were increased through the inhibition of different immunosuppressors agents [109]. In patients with colon cancer after curcumin therapy, there was a suppression of the FoxP3 gene expression showing a reversal of bounding of FoxP3 to T-bet which prevented IFN- γ expression

[110]. Collectively, in contrast to other conditions which curcumin increased Treg cells population and suppressed Th1 responses, in cancer microenvironment, curcumin treatment convert Tregs to Th1 cells by increasing IFN- γ production and suppressing FoxP3 expression. Due to the various properties of curcumin in cancer, asthma, allergy, transplantation, and autoimmune disease, understanding the mechanisms of action of curcumin is an intriguing challenge. Collectively, these findings indicate that curcumin could be exploited for the prevention and management of tumor-induced immune system suppression and may have the potential for clinical application.

Conclusion and future perspectives

In brief, curcumin has potent immunosuppressive properties both *in vitro and in-vivo*. Curcumin has the potency to selectively enhance the function and number Tregs as well as suppress macrophage and DC-mediated differentiation of Th1 and Th17. The effects of curcumin on Th9 and Th22 are unclear and need further investigations. Emerging evidence points towards curcumin having a therapeutic potential in Th1-, Th2- and Th17-driven inflammatory as well as autoimmune diseases. However, confirmatory clinical data from randomized controlled trials would be integral to introduce curcumin to the armamentarium of drugs for these diseases.

Conflict of interests: None.

References

- [1] A. Shapouri-Moghaddam, S. Mohammadian, H. Vazini, M. Taghadosi, S.-A. Esmaeili, F. Mardani, B. Seifi, A. Mohammadi, J.T. Afshari and A. Sahebkar, Macrophage plasticity, polarization, and function in health and disease, Journal of cellular physiology 233 (2018) 6425-6440.
- [2] J. Zhu, H. Yamane and W.E. Paul, Differentiation of effector CD4 T cell populations (*), Annual review of immunology 28 (2010) 445-89.
- [3] D.A. Dias, S. Urban and U. Roessner, A historical overview of natural products in drug discovery, Metabolites 2 (2012) 303-336.
- [4] H. Yuan, Q. Ma, L. Ye and G. Piao, The Traditional Medicine and Modern Medicine from Natural Products, Molecules (Basel, Switzerland) 21 (2016).
- [5] G.C. Jagetia and B.B. Aggarwal, "Spicing up" of the immune system by curcumin, Journal of clinical immunology 27 (2007) 19-35.
- [6] V.S. Yadav, K.P. Mishra, D.P. Singh, S. Mehrotra and V.K. Singh, Immunomodulatory effects of curcumin, Immunopharmacology and immunotoxicology 27 (2005) 485-97.

- [7] C. Varalakshmi, A.M. Ali, B.V. Pardhasaradhi, R.M. Srivastava, S. Singh and A. Khar, Immunomodulatory effects of curcumin: in-vivo, International immunopharmacology 8 (2008) 688-700.
- [8] H. Mollazadeh, A.F.G. Cicero, C.N. Blesso, M. Pirro, M. Majeed and A. Sahebkar, Immune modulation by curcumin: The role of interleukin-10, Critical reviews in food science and nutrition (2017) 1-13.
- [9] R.A. Sharma, A.J. Gescher and W.P. Steward, Curcumin: the story so far, European journal of cancer (Oxford, England : 1990) 41 (2005) 1955-68.
- [10] J. Kumagai, K. Hirahara and T. Nakayama, Pathogenic Th cell subsets in chronic inflammatory diseases, Nihon Rinsho Men'eki Gakkai kaishi = Japanese journal of clinical immunology 39 (2016) 114-23.
- [11] S. Romagnani, Th1/Th2 cells, Inflammatory bowel diseases 5 (1999) 285-94.
- [12] R.M. Talaat, S.F. Mohamed, I.H. Bassyouni and A.A. Raouf, Th1/Th2/Th17/Treg cytokine imbalance in systemic lupus erythematosus (SLE) patients: Correlation with disease activity, Cytokine 72 (2015) 146-53.
- [13] D. Bosisio, N. Polentarutti, M. Sironi, S. Bernasconi, K. Miyake, G.R. Webb, M.U. Martin, A. Mantovani and M. Muzio, Stimulation of toll-like receptor 4 expression in human mononuclear phagocytes by interferon-gamma: a molecular basis for priming and synergism with bacterial lipopolysaccharide, Blood 99 (2002) 3427-31.
- [14] S.L. Peng, S.J. Szabo and L.H. Glimcher, T-bet regulates IgG class switching and pathogenic autoantibody production, Proceedings of the National Academy of Sciences of the United States of America 99 (2002) 5545-50.
- [15] U. Boehm, T. Klamp, M. Groot and J.C. Howard, Cellular responses to interferongamma, Annual review of immunology 15 (1997) 749-95.
- [16] I. Raphael, S. Nalawade, T.N. Eagar and T.G. Forsthuber, T cell subsets and their signature cytokines in autoimmune and inflammatory diseases, Cytokine 74 (2015) 5-17.
- [17] J. Zhu and W.E. Paul, CD4 T cells: fates, functions, and faults, Blood 112 (2008) 1557-69.
- [18] J.K. Kolls and A. Linden, Interleukin-17 family members and inflammation, Immunity 21 (2004) 467-76.
- [19] M.S. Maddur, P. Miossec, S.V. Kaveri and J. Bayry, Th17 cells: biology, pathogenesis of autoimmune and inflammatory diseases, and therapeutic strategies, The American journal of pathology 181 (2012) 8-18.
- [20] U. Baron, S. Floess, G. Wieczorek, K. Baumann, A. Grutzkau, J. Dong, A. Thiel, T.J. Boeld, P. Hoffmann, M. Edinger, I. Turbachova, A. Hamann, S. Olek and J. Huehn, DNA demethylation in the human FOXP3 locus discriminates regulatory T cells from activated FOXP3(+) conventional T cells, European journal of immunology 37 (2007) 2378-89.
- [21] J.A. Hill, M. Feuerer, K. Tash, S. Haxhinasto, J. Perez, R. Melamed, D. Mathis and C. Benoist, Foxp3 transcription-factor-dependent and -independent regulation of the regulatory T cell transcriptional signature, Immunity 27 (2007) 786-800.
- [22] M.G. Roncarolo, S. Gregori, M. Battaglia, R. Bacchetta, K. Fleischhauer and M.K. Levings, Interleukin-10-secreting type 1 regulatory T cells in rodents and humans, Immunological reviews 212 (2006) 28-50.
- [23] W. Kleczynska, B. Jakiela, H. Plutecka, M. Milewski, M. Sanak and J. Musial, Imbalance between Th17 and regulatory T-cells in systemic lupus erythematosus, Folia histochemica et cytobiologica 49 (2011) 646-53.

- [24] S. Dolff, M. Bijl, M.G. Huitema, P.C. Limburg, C.G. Kallenberg and W.H. Abdulahad, Disturbed Th1, Th2, Th17 and T(reg) balance in patients with systemic lupus erythematosus, Clinical immunology (Orlando, Fla.) 141 (2011) 197-204.
- [25] A. Kuhn, S. Beissert and P.H. Krammer, CD4(+)CD25 (+) regulatory T cells in human lupus erythematosus, Archives of dermatological research 301 (2009) 71-81.
- [26] J. Yang, M.S. Sundrud, J. Skepner and T. Yamagata, Targeting Th17 cells in autoimmune diseases, Trends in pharmacological sciences 35 (2014) 493-500.
- [27] Y.C. Qiao, Y.H. Pan, W. Ling, F. Tian, Y.L. Chen, X.X. Zhang and H.L. Zhao, The Yin and Yang of regulatory T cell and therapy progress in autoimmune disease, Autoimmunity reviews 16 (2017) 1058-1070.
- [28] F. Jadidi-Niaragh and A. Mirshafiey, The deviated balance between regulatory T cell and Th17 in autoimmunity, Immunopharmacology and immunotoxicology 34 (2012) 727-39.
- [29] S.C. Gupta, S. Patchva, W. Koh and B.B. Aggarwal, Discovery of curcumin, a component of golden spice, and its miraculous biological activities, Clinical and experimental pharmacology & physiology 39 (2012) 283-99.
- [30] H. Hatcher, R. Planalp, J. Cho, F.M. Torti and S.V. Torti, Curcumin: from ancient medicine to current clinical trials, Cellular and molecular life sciences : CMLS 65 (2008) 1631-52.
- [31] A. Storka, B. Vcelar, U. Klickovic, G. Gouya, S. Weisshaar, S. Aschauer, G. Bolger, L. Helson and M. Wolzt, Safety, tolerability and pharmacokinetics of liposomal curcumin in healthy humans, International journal of clinical pharmacology and therapeutics 53 (2015) 54-65.
- [32] N. Chainani-Wu, Safety and anti-inflammatory activity of curcumin: a component of tumeric (Curcuma longa), Journal of alternative and complementary medicine (New York, N.Y.) 9 (2003) 161-8.
- [33] V. Soleimani, A. Sahebkar and H. Hosseinzadeh, Turmeric (Curcuma longa) and its major constituent (curcumin) as nontoxic and safe substances: Review, Phytotherapy research : PTR 32 (2018) 985-995.
- [34] S.C. Gupta, S. Patchva and B.B. Aggarwal, Therapeutic roles of curcumin: lessons learned from clinical trials, The AAPS journal 15 (2013) 195-218.
- [35] S.C. Gautam, X. Gao and S. Dulchavsky, Immunomodulation by curcumin, Advances in experimental medicine and biology 595 (2007) 321-41.
- [36] A.A. Momtazi-Borojeni, S.M. Haftcheshmeh, S.A. Esmaeili, T.P. Johnston, E. Abdollahi and A. Sahebkar, Curcumin: A natural modulator of immune cells in systemic lupus erythematosus, Autoimmunity reviews 17 (2018) 125-135.
- [37] X. Gao, J. Kuo, H. Jiang, D. Deeb, Y. Liu, G. Divine, R.A. Chapman, S.A. Dulchavsky and S.C. Gautam, Immunomodulatory activity of curcumin: suppression of lymphocyte proliferation, development of cell-mediated cytotoxicity, and cytokine production in vitro, Biochemical pharmacology 68 (2004) 51-61.
- [38] D. R Cundell and F. Wilkinson, Curcumin: Powerful immunomodulator from turmeric, Current Immunology Reviews 10 (2014) 122-132.
- [39] A. Ahmed, A. El Fouhil, R. Mohamed, M. Atteya, N. Abdel-Baky, A. AlRoalle and A. Aldahmash, Curcumin ameliorates experimental autoimmune acute myocarditis in rats as evidenced by decrease in thioredoxin immunoreactivity, Folia morphologica 74 (2015) 318-324.

- [40] R.M. Srivastava, S. Singh, S.K. Dubey, K. Misra and A. Khar, Immunomodulatory and therapeutic activity of curcumin, International immunopharmacology 11 (2011) 331-41.
- [41] M.H. Seyedzadeh, Z. Safari, A. Zare, J.G. Navashenaq, G.A. Kardar and M.R. Khorramizadeh, Study of curcumin immunomodulatory effects on reactive astrocyte cell function, International immunopharmacology 22 (2014) 230-235.
- [42] C. Varalakshmi, A.M. Ali, B. Pardhasaradhi, R.M. Srivastava, S. Singh and A. Khar, Immunomodulatory effects of curcumin: in-vivo, International immunopharmacology 8 (2008) 688-700.
- [43] G.C. Jagetia and B.B. Aggarwal, "Spicing up" of the immune system by curcumin, Journal of clinical immunology 27 (2007) 19-35.
- [44] A. Mohammadi, C.N. Blesso, G.E. Barreto, M. Banach, M. Majeed and A. Sahebkar, Macrophage plasticity, polarization and function in response to curcumin, a diet-derived polyphenol, as an immunomodulatory agent, The Journal of nutritional biochemistry 66 (2018) 1-16.
- [45] H. Zhou, C.S. Beevers and S. Huang, The targets of curcumin, Current drug targets 12 (2011) 332-47.
- [46] M. Karimi, H. Mohammadi, M. Hemmatzadeh, A. Mohammadi, H. Rafatpanah and B. Baradaran, Role of the HTLV-1 viral factors in the induction of apoptosis, Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie 85 (2017) 334-347.
- [47] A. Mohammadi, B. Fazeli, M. Taheri, A. Sahebkar, Z. Poursina, V. Vakili, S.Z. Yazdi, Z. Keramati, R. Boostani, I. Hampson and H. Rafatpanah, Modulatory effects of curcumin on apoptosis and cytotoxicity-related molecules in HTLV-1-associated myelopathy/tropical spastic paraparesis (HAM/TSP) patients, Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie 85 (2017) 457-462.
- [48] B.Y. Kang, Y.J. Song, K.M. Kim, Y.K. Choe, S.Y. Hwang and T.S. Kim, Curcumin inhibits Th1 cytokine profile in CD4+ T cells by suppressing interleukin-12 production in macrophages, British journal of pharmacology 128 (1999) 380-4.
- [49] N.A. Forward, D.M. Conrad, M.R. Power Coombs, C.D. Doucette, S.J. Furlong, T.J. Lin and D.W. Hoskin, Curcumin blocks interleukin (IL)-2 signaling in T-lymphocytes by inhibiting IL-2 synthesis, CD25 expression, and IL-2 receptor signaling, Biochemical and biophysical research communications 407 (2011) 801-6.
- [50] E. Maier, K. Kurz, M. Jenny, H. Schennach, F. Ueberall and D. Fuchs, Food preservatives sodium benzoate and propionic acid and colorant curcumin suppress Th1type immune response in vitro, Food and chemical toxicology : an international journal published for the British Industrial Biological Research Association 48 (2010) 1950-6.
- [51] J. Bruck, J. Holstein, I. Glocova, U. Seidel, J. Geisel, T. Kanno, J. Kumagai, N. Mato, S. Sudowe, K. Widmaier, T. Sinnberg, A.S. Yazdi, F.C. Eberle, K. Hirahara, T. Nakayama, M. Rocken and K. Ghoreschi, Nutritional control of IL-23/Th17-mediated autoimmune disease through HO-1/STAT3 activation, Scientific reports 7 (2017) 44482.
- [52] G. Kim, M.S. Jang, Y.M. Son, M.J. Seo, S.Y. Ji, S.H. Han, I.D. Jung, Y.M. Park, H.J. Jung and C.H. Yun, Curcumin inhibits CD4(+) T cell activation, but augments CD69 expression and TGF-beta1-mediated generation of regulatory T cells at late phase, PloS one 8 (2013) e62300.
- [53] P. Fasching, M. Stradner, W. Graninger, C. Dejaco and J. Fessler, Therapeutic Potential of Targeting the Th17/Treg Axis in Autoimmune Disorders, Molecules (Basel, Switzerland) 22 (2017).

- [54] F. Isono, S. Fujita-Sato and S. Ito, Inhibiting RORgammat/Th17 axis for autoimmune disorders, Drug discovery today 19 (2014) 1205-11.
- [55] F. Tahmasebinia and A. Pourgholaminejad, The role of Th17 cells in auto-inflammatory neurological disorders, Progress in neuro-psychopharmacology & biological psychiatry 79 (2017) 408-416.
- [56] G. Zhao, Y. Liu, X. Yi, Y. Wang, S. Qiao, Z. Li, J. Ni and Z. Song, Curcumin inhibiting Th17 cell differentiation by regulating the metabotropic glutamate receptor-4 expression on dendritic cells, International immunopharmacology 46 (2017) 80-86.
- [57] G.J. Zhao, Z.Q. Lu, L.M. Tang, Z.S. Wu, D.W. Wang, J.Y. Zheng and Q.M. Qiu, Curcumin inhibits suppressive capacity of naturally occurring CD4+CD25+ regulatory T cells in mice in vitro, International immunopharmacology 14 (2012) 99-106.
- [58] K.M. Danikowski, S. Jayaraman and B.S. Prabhakar, Regulatory T cells in multiple sclerosis and myasthenia gravis, Journal of neuroinflammation 14 (2017) 117.
- [59] S. Wang, H. Li, M. Zhang, L.T. Yue, C.C. Wang, P. Zhang, Y. Liu and R.S. Duan, Curcumin ameliorates experimental autoimmune myasthenia gravis by diverse immune cells, Neuroscience letters 626 (2016) 25-34.
- [60] L.M. Charbonnier, E. Janssen, J. Chou, T.K. Ohsumi, S. Keles, J.T. Hsu, M.J. Massaad, M. Garcia-Lloret, R. Hanna-Wakim, G. Dbaibo, A.A. Alangari, A. Alsultan, D. Al-Zahrani, R.S. Geha and T.A. Chatila, Regulatory T-cell deficiency and immune dysregulation, polyendocrinopathy, enteropathy, X-linked-like disorder caused by lossof-function mutations in LRBA, The Journal of allergy and clinical immunology 135 (2015) 217-27.
- [61] R.S. Wildin and A. Freitas, IPEX and FOXP3: clinical and research perspectives, Journal of autoimmunity 25 Suppl (2005) 56-62.
- [62] G. Lee, H.S. Chung, K. Lee, H. Lee, M. Kim and H. Bae, Curcumin attenuates the scurfy-induced immune disorder, a model of IPEX syndrome, with inhibiting Th1/Th2/Th17 responses in mice, Phytomedicine : international journal of phytotherapy and phytopharmacology 33 (2017) 1-6.
- [63] C.S. Constantinescu, N. Farooqi, K. O'Brien and B. Gran, Experimental autoimmune encephalomyelitis (EAE) as a model for multiple sclerosis (MS), British journal of pharmacology 164 (2011) 1079-1106.
- [64] S. Kanakasabai, E. Casalini, C.C. Walline, C. Mo, W. Chearwae and J.J. Bright, Differential regulation of CD4(+) T helper cell responses by curcumin in experimental autoimmune encephalomyelitis, The Journal of nutritional biochemistry 23 (2012) 1498-507.
- [65] C. Natarajan and J.J. Bright, Curcumin inhibits experimental allergic encephalomyelitis by blocking IL-12 signaling through Janus kinase-STAT pathway in T lymphocytes, Journal of immunology (Baltimore, Md. : 1950) 168 (2002) 6506-13.
- [66] A.J. Fahey, R. Adrian Robins and C.S. Constantinescu, Curcumin modulation of IFNbeta and IL-12 signalling and cytokine induction in human T cells, Journal of cellular and molecular medicine 11 (2007) 1129-37.
- [67] S. Dolati, M. Ahmadi, L. Aghebti-Maleki, A. Nikmaram, F. Marofi, R. Rikhtegar, H. Ayromlou and M. Yousefi, Nanocurcumin is a potential novel therapy for multiple sclerosis by influencing inflammatory mediators, Pharmacological reports : PR 70 (2018) 1158-1167.

- [68] S. Dolati, M. Ahmadi, R. Rikhtegar, Z. Babaloo, H. Ayromlou, L. Aghebati-Maleki, M. Nouri and M. Yousefi, Changes in Th17 cells function after nanocurcumin use to treat multiple sclerosis, International immunopharmacology 61 (2018) 74-81.
- [69] N. Ghanaatian, N.A. Lashgari, A.H. Abdolghaffari, S.M. Rajaee, Y. Panahi, G.E. Barreto, A.E. Butler and A. Sahebkar, Curcumin as a therapeutic candidate for multiple sclerosis: Molecular mechanisms and targets, Journal of cellular physiology (2018).
- [70] L. Xie, X.K. Li and S. Takahara, Curcumin has bright prospects for the treatment of multiple sclerosis, International immunopharmacology 11 (2011) 323-30.
- [71] E. Asteriou, A. Gkoutzourelas, A. Mavropoulos, C. Katsiari, L.I. Sakkas and D.P. Bogdanos, Curcumin for the Management of Periodontitis and Early ACPA-Positive Rheumatoid Arthritis: Killing Two Birds with One Stone, Nutrients 10 (2018).
- [72] M.T. Leech and P.M. Bartold, The association between rheumatoid arthritis and periodontitis, Best practice & research. Clinical rheumatology 29 (2015) 189-201.
- [73] M. Ebbers, P.M. Lübcke, J. Volzke, K. Kriebel, C. Hieke, R. Engelmann, H. Lang, B. Kreikemeyer and B. Müller-Hilke, Interplay between P. gingivalis, F. nucleatum and A. actinomycetemcomitans in murine alveolar bone loss, arthritis onset and progression, Scientific reports 8 (2018) 15129.
- [74] K.J. Maresz, A. Hellvard, A. Sroka, K. Adamowicz, E. Bielecka, J. Koziel, K. Gawron, D. Mizgalska, K.A. Marcinska, M. Benedyk, K. Pyrc, A.M. Quirke, R. Jonsson, S. Alzabin, P.J. Venables, K.A. Nguyen, P. Mydel and J. Potempa, Porphyromonas gingivalis facilitates the development and progression of destructive arthritis through its unique bacterial peptidylarginine deiminase (PAD), PLoS pathogens 9 (2013) e1003627.
- [75] J. Lee, V. Taneja and R. Vassallo, Cigarette smoking and inflammation: cellular and molecular mechanisms, Journal of dental research 91 (2012) 142-149.
- [76] M. Maiti, K. Chattopadhyay, M. Verma and B. Chattopadhyay, Curcumin protects against nicotine-induced stress during protein malnutrition in female rat through immunomodulation with cellular amelioration, Molecular biology reports 42 (2015) 1623-37.
- [77] K. Handono, M.Z. Pratama, A.T. Endharti and H. Kalim, Treatment of low doses curcumin could modulate Th17/Treg balance specifically on CD4+ T cell cultures of systemic lupus erythematosus patients, Central-European journal of immunology 40 (2015) 461-9.
- [78] M.P. Schon and W.H. Boehncke, Psoriasis, The New England journal of medicine 352 (2005) 1899-912.
- [79] M.A. Lowes, A.M. Bowcock and J.G. Krueger, Pathogenesis and therapy of psoriasis, Nature 445 (2007) 866-73.
- [80] A. Di Cesare, P. Di Meglio and F.O. Nestle, The IL-23/Th17 axis in the immunopathogenesis of psoriasis, The Journal of investigative dermatology 129 (2009) 1339-50.
- [81] L.C. Zaba, J. Fuentes-Duculan, N.J. Eungdamrong, M.V. Abello, I. Novitskaya, K.C. Pierson, J. Gonzalez, J.G. Krueger and M.A. Lowes, Psoriasis is characterized by accumulation of immunostimulatory and Th1/Th17 cell-polarizing myeloid dendritic cells, The Journal of investigative dermatology 129 (2009) 79-88.
- [82] R. Yadav, B. Jee and S.K. Awasthi, Curcumin Suppresses the Production of Proinflammatory Cytokine Interleukin-18 in Lipopolysaccharide Stimulated Murine Macrophage-Like Cells, Indian journal of clinical biochemistry : IJCB 30 (2015) 109-12.

- [83] J. Sun, Y. Zhao and J. Hu, Curcumin inhibits imiquimod-induced psoriasis-like inflammation by inhibiting IL-1beta and IL-6 production in mice, PloS one 8 (2013) e67078.
- [84] J. Sun, Y. Zhao, H. Jin and J. Hu, Curcumin relieves TPA-induced Th1 inflammation in K14-VEGF transgenic mice, International immunopharmacology 25 (2015) 235-41.
- [85] K. Conrad, D. Roggenbuck and M.W. Laass, Diagnosis and classification of ulcerative colitis, Autoimmunity reviews 13 (2014) 463-6.
- [86] I. Ordas, L. Eckmann, M. Talamini, D.C. Baumgart and W.J. Sandborn, Ulcerative colitis, Lancet (London, England) 380 (2012) 1606-19.
- [87] H. Hanai and K. Sugimoto, Curcumin has bright prospects for the treatment of inflammatory bowel disease, Current pharmaceutical design 15 (2009) 2087-94.
- [88] M. Zhang, C.S. Deng, J.J. Zheng and J. Xia, Curcumin regulated shift from Th1 to Th2 in trinitrobenzene sulphonic acid-induced chronic colitis, Acta pharmacologica Sinica 27 (2006) 1071-7.
- [89] L. Camacho-Barquero, I. Villegas, J.M. Sanchez-Calvo, E. Talero, S. Sanchez-Fidalgo, V. Motilva and C. Alarcon de la Lastra, Curcumin, a Curcuma longa constituent, acts on MAPK p38 pathway modulating COX-2 and iNOS expression in chronic experimental colitis, International immunopharmacology 7 (2007) 333-42.
- [90] C.B. Larmonier, J.K. Uno, K.M. Lee, T. Karrasch, D. Laubitz, R. Thurston, M.T. Midura-Kiela, F.K. Ghishan, R.B. Sartor, C. Jobin and P.R. Kiela, Limited effects of dietary curcumin on Th-1 driven colitis in IL-10 deficient mice suggest an IL-10dependent mechanism of protection, American journal of physiology. Gastrointestinal and liver physiology 295 (2008) G1079-91.
- [91] C. Billerey-Larmonier, J.K. Uno, N. Larmonier, A.J. Midura, B. Timmermann, F.K. Ghishan and P.R. Kiela, Protective effects of dietary curcumin in mouse model of chemically induced colitis are strain dependent, Inflammatory bowel diseases 14 (2008) 780-93.
- [92] H.M. Zhao, R. Xu, X.Y. Huang, S.M. Cheng, M.F. Huang, H.Y. Yue, X. Wang, Y. Zou, A.P. Lu and D.Y. Liu, Curcumin improves regulatory T cells in gut-associated lymphoid tissue of colitis mice, World journal of gastroenterology 22 (2016) 5374-83.
- [93] M. Ohno, A. Nishida, Y. Sugitani, K. Nishino, O. Inatomi, M. Sugimoto, M. Kawahara and A. Andoh, Nanoparticle curcumin ameliorates experimental colitis via modulation of gut microbiota and induction of regulatory T cells, PloS one 12 (2017) e0185999.
- [94] J. Sun, F. Chen, C. Braun, Y.Q. Zhou, H. Rittner, Y.K. Tian, X.Y. Cai and D.W. Ye, Role of curcumin in the management of pathological pain, Phytomedicine : international journal of phytotherapy and phytopharmacology 48 (2018) 129-140.
- [95] V.P. Kurup, C.S. Barrios, R. Raju, B.D. Johnson, M.B. Levy and J.N. Fink, Immune response modulation by curcumin in a latex allergy model, Clinical and molecular allergy : CMA 5 (2007) 1.
- [96] A. Kumari, D. Dash and R. Singh, Lipopolysaccharide (LPS) exposure differently affects allergic asthma exacerbations and its amelioration by intranasal curcumin in mice, Cytokine 76 (2015) 334-342.
- [97] T. Kobayashi, S. Hashimoto and T. Horie, Curcumin inhibition of Dermatophagoides farinea-induced interleukin-5 (IL-5) and granulocyte macrophage-colony stimulating factor (GM-CSF) production by lymphocytes from bronchial asthmatics, Biochemical pharmacology 54 (1997) 819-24.

- [98] H.S. Shin, H.J. See, S.Y. Jung, D.W. Choi, D.A. Kwon, M.J. Bae, K.S. Sung and D.H. Shon, Turmeric (Curcuma longa) attenuates food allergy symptoms by regulating type 1/type 2 helper T cells (Th1/Th2) balance in a mouse model of food allergy, Journal of ethnopharmacology 175 (2015) 21-9.
- [99] C. Ma, Z. Ma, Q. Fu and S. Ma, Curcumin attenuates allergic airway inflammation by regulation of CD4+CD25+ regulatory T cells (Tregs)/Th17 balance in ovalbumin-sensitized mice, Fitoterapia 87 (2013) 57-64.
- [100] N.M. Rogers, S. Kireta and P.T. Coates, Curcumin induces maturation-arrested dendritic cells that expand regulatory T cells in vitro and in vivo, Clinical and experimental immunology 162 (2010) 460-73.
- [101] M.J. Park, S.J. Moon, S.H. Lee, E.J. Yang, J.K. Min, S.G. Cho, C.W. Yang, S.H. Park, H.Y. Kim and M.L. Cho, Curcumin attenuates acute graft-versus-host disease severity via in vivo regulations on Th1, Th17 and regulatory T cells, PloS one 8 (2013) e67171.
- [102] H. Yu, M. Kortylewski and D. Pardoll, Crosstalk between cancer and immune cells: role of STAT3 in the tumour microenvironment, Nature reviews. Immunology 7 (2007) 41-51.
- [103] J.E. Talmadge and D.I. Gabrilovich, History of myeloid-derived suppressor cells, Nature reviews. Cancer 13 (2013) 739-52.
- [104] Y. Mao, I. Poschke and R. Kiessling, Tumour-induced immune suppression: role of inflammatory mediators released by myelomonocytic cells, Journal of internal medicine 276 (2014) 154-70.
- [105] J. Rajasingh, H.P. Raikwar, G. Muthian, C. Johnson and J.J. Bright, Curcumin induces growth-arrest and apoptosis in association with the inhibition of constitutively active JAK-STAT pathway in T cell leukemia, Biochemical and biophysical research communications 340 (2006) 359-68.
- [106] C. Zhang, B. Li, X. Zhang, P. Hazarika, B.B. Aggarwal and M. Duvic, Curcumin selectively induces apoptosis in cutaneous T-cell lymphoma cell lines and patients' PBMCs: potential role for STAT-3 and NF-kappaB signaling, The Journal of investigative dermatology 130 (2010) 2110-9.
- [107] S. Bhattacharyya, D. Md Sakib Hossain, S. Mohanty, G. Sankar Sen, S. Chattopadhyay, S. Banerjee, J. Chakraborty, K. Das, D. Sarkar, T. Das and G. Sa, Curcumin reverses T cell-mediated adaptive immune dysfunctions in tumor-bearing hosts, Cellular & molecular immunology 7 (2010) 306-15.
- [108] J.Y. Zou, C.H. Su, H.H. Luo, Y.Y. Lei, B. Zeng, H.S. Zhu and Z.G. Chen, Curcumin converts Foxp3+ regulatory T cells to T helper 1 cells in patients with lung cancer, Journal of cellular biochemistry 119 (2018) 1420-1428.
- [109] Y.F. Chang, H.Y. Chuang, C.H. Hsu, R.S. Liu, S.S. Gambhir and J.J. Hwang, Immunomodulation of curcumin on adoptive therapy with T cell functional imaging in mice, Cancer prevention research (Philadelphia, Pa.) 5 (2012) 444-52.
- [110] B. Xu, L. Yu and L.Z. Zhao, Curcumin up regulates T helper 1 cells in patients with colon cancer, American journal of translational research 9 (2017) 1866-1875.
- [111] C. Csaki, A. Mobasheri and M. Shakibaei, Synergistic chondroprotective effects of curcumin and resveratrol in human articular chondrocytes: inhibition of IL-1beta-induced NF-kappaB-mediated inflammation and apoptosis, Arthritis research & therapy 11 (2009) R165.

- [112] S.W. Oh, J.Y. Cha, J.E. Jung, B.C. Chang, H.J. Kwon, B.R. Lee and D.Y. Kim, Curcumin attenuates allergic airway inflammation and hyper-responsiveness in mice through NF-kappaB inhibition, Journal of ethnopharmacology 136 (2011) 414-21.
- [113] C. Jobin, C.A. Bradham, M.P. Russo, B. Juma, A.S. Narula, D.A. Brenner and R.B. Sartor, Curcumin blocks cytokine-mediated NF-kappa B activation and proinflammatory gene expression by inhibiting inhibitory factor I-kappa B kinase activity, Journal of immunology (Baltimore, Md. : 1950) 163 (1999) 3474-83.
- [114] K.H. Kim, E.N. Lee, J.K. Park, J.R. Lee, J.H. Kim, H.J. Choi, B.S. Kim, H.W. Lee, K.S. Lee and S. Yoon, Curcumin attenuates TNF-alpha-induced expression of intercellular adhesion molecule-1, vascular cell adhesion molecule-1 and proinflammatory cytokines in human endometriotic stromal cells, Phytotherapy research : PTR 26 (2012) 1037-47.
- [115] J. Gertsch, M. Guttinger, J. Heilmann and O. Sticher, Curcumin differentially modulates mRNA profiles in Jurkat T and human peripheral blood mononuclear cells, Bioorganic & medicinal chemistry 11 (2003) 1057-63.
- [116] M. Zheng, Q. Zhang, Y. Joe, B.H. Lee, D.G. Ryu, K.B. Kwon, S.W. Ryter and H.T. Chung, Curcumin induces apoptotic cell death of activated human CD4+ T cells via increasing endoplasmic reticulum stress and mitochondrial dysfunction, International immunopharmacology 15 (2013) 517-23.
- [117] Y. Abe, S. Hashimoto and T. Horie, Curcumin inhibition of inflammatory cytokine production by human peripheral blood monocytes and alveolar macrophages, Pharmacological research 39 (1999) 41-7.
- [118] B. Cipriani, G. Borsellino, H. Knowles, D. Tramonti, F. Cavaliere, G. Bernardi, L. Battistini and C.F. Brosnan, Curcumin inhibits activation of Vgamma9Vdelta2 T cells by phosphoantigens and induces apoptosis involving apoptosis-inducing factor and large scale DNA fragmentation, Journal of immunology (Baltimore, Md. : 1950) 167 (2001) 3454-62.
- [119] F. Antoine, J.C. Simard and D. Girard, Curcumin inhibits agent-induced human neutrophil functions in vitro and lipopolysaccharide-induced neutrophilic infiltration in vivo, International immunopharmacology 17 (2013) 1101-7.
- [120] A.C. Bharti, N. Donato and B.B. Aggarwal, Curcumin (diferuloylmethane) inhibits constitutive and IL-6-inducible STAT3 phosphorylation in human multiple myeloma cells, Journal of immunology (Baltimore, Md. : 1950) 171 (2003) 3863-71.
- [121] S. Shishodia, H.M. Amin, R. Lai and B.B. Aggarwal, Curcumin (diferuloylmethane) inhibits constitutive NF-kappaB activation, induces G1/S arrest, suppresses proliferation, and induces apoptosis in mantle cell lymphoma, Biochemical pharmacology 70 (2005) 700-13.
- [122] G.Y. Kim, K.H. Kim, S.H. Lee, M.S. Yoon, H.J. Lee, D.O. Moon, C.M. Lee, S.C. Ahn, Y.C. Park and Y.M. Park, Curcumin inhibits immunostimulatory function of dendritic cells: MAPKs and translocation of NF-kappa B as potential targets, Journal of immunology (Baltimore, Md. : 1950) 174 (2005) 8116-24.
- [123] H. Hidaka, T. Ishiko, T. Furuhashi, H. Kamohara, S. Suzuki, M. Miyazaki, O. Ikeda, S. Mita, T. Setoguchi and M. Ogawa, Curcumin inhibits interleukin 8 production and enhances interleukin 8 receptor expression on the cell surface:impact on human pancreatic carcinoma cell growth by autocrine regulation, Cancer 95 (2002) 1206-14.

- [124] F. Milano, L. Mari, W. van de Luijtgaarden, K. Parikh, S. Calpe and K.K. Krishnadath, Nano-curcumin inhibits proliferation of esophageal adenocarcinoma cells and enhances the T cell mediated immune response, Frontiers in oncology 3 (2013) 137.
- [125] S.A. Shirley, A.J. Montpetit, R.F. Lockey and S.S. Mohapatra, Curcumin prevents human dendritic cell response to immune stimulants, Biochemical and biophysical research communications 374 (2008) 431-6.
- [126] H.Y. Kim, E.J. Park, E.H. Joe and I. Jou, Curcumin suppresses Janus kinase-STAT inflammatory signaling through activation of Src homology 2 domain-containing tyrosine phosphatase 2 in brain microglia, Journal of immunology (Baltimore, Md. : 1950) 171 (2003) 6072-9.
- [127] J. Epstein, G. Docena, T.T. MacDonald and I.R. Sanderson, Curcumin suppresses p38 mitogen-activated protein kinase activation, reduces IL-1beta and matrix metalloproteinase-3 and enhances IL-10 in the mucosa of children and adults with inflammatory bowel disease, The British journal of nutrition 103 (2010) 824-32.
- [128] C. Kliem, A. Merling, M. Giaisi, R. Kohler, P.H. Krammer and M. Li-Weber, Curcumin suppresses T cell activation by blocking Ca2+ mobilization and nuclear factor of activated T cells (NFAT) activation, The Journal of biological chemistry 287 (2012) 10200-9.
- [129] H. Kamohara, M. Takahashi, T. Ishiko, M. Ogawa and H. Baba, Induction of interleukin-8 (CXCL-8) by tumor necrosis factor-alpha and leukemia inhibitory factor in pancreatic carcinoma cells: Impact of CXCL-8 as an autocrine growth factor, International journal of oncology 31 (2007) 627-32.
- [130] X. Li and X. Liu, Effect of curcumin on immune function of mice, Journal of Huazhong University of Science and Technology. Medical sciences = Hua zhong ke ji da xue xue bao. Yi xue Ying De wen ban = Huazhong ke ji daxue xuebao. Yixue Yingdewen ban 25 (2005) 137-40.
- [131] J.M. Yun, I. Jialal and S. Devaraj, Epigenetic regulation of high glucose-induced proinflammatory cytokine production in monocytes by curcumin, The Journal of nutritional biochemistry 22 (2011) 450-8.
- [132] X. Gao, J. Kuo, H. Jiang, D. Deeb, Y. Liu, G. Divine, R.A. Chapman, S.A. Dulchavsky and S.C. Gautam, Immunomodulatory activity of curcumin: suppression of lymphocyte proliferation, development of cell-mediated cytotoxicity, and cytokine production in vitro, Biochemical pharmacology 68 (2004) 51-61.
- [133] A. Olivera, T.W. Moore, F. Hu, A.P. Brown, A. Sun, D.C. Liotta, J.P. Snyder, Y. Yoon, H. Shim, A.I. Marcus, A.H. Miller and T.W. Pace, Inhibition of the NF-kappaB signaling pathway by the curcumin analog, 3,5-Bis(2-pyridinylmethylidene)-4-piperidone (EF31): anti-inflammatory and anti-cancer properties, International immunopharmacology 12 (2012) 368-77.
- [134] M.M. Chan, Inhibition of tumor necrosis factor by curcumin, a phytochemical, Biochemical pharmacology 49 (1995) 1551-6.
- [135] J.W. Cho, K.S. Lee and C.W. Kim, Curcumin attenuates the expression of IL-1beta, IL-6, and TNF-alpha as well as cyclin E in TNF-alpha-treated HaCaT cells; NF-kappaB and MAPKs as potential upstream targets, International journal of molecular medicine 19 (2007) 469-74.

- [136] Y. Cong, L. Wang, A. Konrad, T. Schoeb and C.O. Elson, Curcumin induces the tolerogenic dendritic cell that promotes differentiation of intestine-protective regulatory T cells, European journal of immunology 39 (2009) 3134-46.
- [137] J. Sun, W. Guo, Y. Ben, J. Jiang, C. Tan, Z. Xu, X. Wang and C. Bai, Preventive effects of curcumin and dexamethasone on lung transplantation-associated lung injury in rats, Critical care medicine 36 (2008) 1205-13.
- [138] A. Sahebkar, A.F.G. Cicero, L.E. Simental-Mendia, B.B. Aggarwal and S.C. Gupta, Curcumin downregulates human tumor necrosis factor-alpha levels: A systematic review and meta-analysis ofrandomized controlled trials, Pharmacological research 107 (2016) 234-242.
- [139] S.R. Kinney, L. Carlson, J. Ser-Dolansky, C. Thompson, S. Shah, A. Gambrah, W. Xing, S.S. Schneider and C.B. Mathias, Curcumin Ingestion Inhibits Mastocytosis and Suppresses Intestinal Anaphylaxis in a Murine Model of Food Allergy, PloS one 10 (2015) e0132467.
- [140] A. Gulcubuk, K. Altunatmaz, K. Sonmez, D. Haktanir-Yatkin, H. Uzun, A. Gurel and S. Aydin, Effects of curcumin on tumour necrosis factor-alpha and interleukin-6 in the late phase of experimental acute pancreatitis, Journal of veterinary medicine. A, Physiology, pathology, clinical medicine 53 (2006) 49-54.
- [141] A. Ukil, S. Maity, S. Karmakar, N. Datta, J.R. Vedasiromoni and P.K. Das, Curcumin, the major component of food flavour turmeric, reduces mucosal injury in trinitrobenzene sulphonic acid-induced colitis, British journal of pharmacology 139 (2003) 209-18.
- [142] S. Bhaumik, M.D. Jyothi and A. Khar, Differential modulation of nitric oxide production by curcumin in host macrophages and NK cells, FEBS letters 483 (2000) 78-82.
- [143] W. Kim, Y.Y. Fan, R. Smith, B. Patil, G.K. Jayaprakasha, D.N. McMurray and R.S. Chapkin, Dietary curcumin and limonin suppress CD4+ T-cell proliferation and interleukin-2 production in mice, The Journal of nutrition 139 (2009) 1042-8.
- [144] M.F. El-Azab, F.M. Attia and A.M. El-Mowafy, Novel role of curcumin combined with bone marrow transplantation in reversing experimental diabetes: Effects on pancreatic islet regeneration, oxidative stress, and inflammatory cytokines, European journal of pharmacology 658 (2011) 41-8.
- [145] H. Lee, H. Kim, G. Lee, H.S. Chung and H. Bae, Curcumin attenuates lupus nephritis upon interaction with regulatory T cells in New Zealand Black/White mice, The British journal of nutrition 110 (2013) 69-76.
- [146] J.P. Gaddipati, S.V. Sundar, J. Calemine, P. Seth, G.S. Sidhu and R.K. Maheshwari, Differential regulation of cytokines and transcription factors in liver by curcumin following hemorrhage/resuscitation, Shock (Augusta, Ga.) 19 (2003) 150-6.

Cell lines	Curcumin dose	Duration	Effects	Reference
T cells	50 mg/kg (low-dose group) or 100 mg/kg curcumin	11 to 45day	Down-regulated the levels of pro-inflammatory cytokines (IL-17, IFN- α and TNF- α) and up-regulated the levels of the anti-inflammatory cytokine IL-10	[1]

Table 1. In-vitro studies that have evaluated the effects of curcumin on Th balance.

	(high-dose group)			
Human chondrocytes	50 μM curcumin	4 & 24 hrs	Resveratrol suppressed NF- κ B-regulated gene products involved in inflammation (cyclooxygenase- 2, matrix metalloproteinase (MMP)-3, MMP-9, vascular endothelial growth factor), inhibited apoptosis (Bcl-2, Bcl-xL, and TNF- α receptor- associated factor 1) and prevented activation of caspase-3. IL-1 β -induced NF- κ B activation	[2]
T cell	2.5, 5 and 10 μm	0 to 4 days	Regulation of T helper 1 (Th1) and Treg	[3]
Raw264.7 cells	200 mg/kg		Inhibition of NF-kb activation in the asthmatic lung tissue	[4]
T cells	1% curcumin	92.5 days	Decreased all of the Th1/Th2/Th17 cell populations	[5]
Intestinal epithelial cells	(0–150 mm)	45 min	Inhibited IL-1b-mediated ICAM-1 and IL-8 gene expression in IEC-6, HT-29, and Caco-2 cells. Cytokine-induced NF-kb DNA binding activity	[6]
T-lymphocyte	5, 10 and 20 μm	120 h	Inhibits IL-2 signaling by reducing available IL-2 and high-affinity IL-2R, as well as interfering with IL-2R signaling	[7]
Stromalcellsisolatedfromwomenwithendometriosis	1, 10, 30 or 50 mm	24 hrs	Inhibited TNF-a-induced secretion of IL-6, IL-8 and MCP-1.	[8]
Human peripheral blood mononuclear cells	1.5 and 13.6 mm		Modulates the expression profile of Th1 cells	[9]
T cells	0, 5, 10 and 20 μm	0 to 6 hours	Expression of the anti-apoptotic protein Bcl-2.	[10]
T cells	0, 2.5, 5, 10, and 25 µg/ml	30 min	Induced a dose-dependent decrease in JAK and STAT phosphorylation	[11]
Lymphocytes	Cr, 10 PM	3 days	Curcumin inhibited IL-5, GM-CSF, and IL-4 production in a concentration-dependent manner.	[12]
Monocytes and alveolar macrophages	0.5m 5 m 1 and 10 μm	24 hrs	The inhibitory effect on the production of IL-8, MIP- 1a, MCP-1, IL-1b, and TNF-a by PMA- or LPS- stimulated monocytes and alveolar macrophages	[13]
γδ T cells	1 μm 10 μm 30 and 75 μm	16, 24 hrs	Inhibited isopentenyl pyrophosphate-induced release of the chemokines macrophage inflammatory protein- 1α and -1β and RANTES. Curcumin also blocked isopentenyl pyrophosphate-induced activation of NF-kb and AP-1.	[14]
Neutrophilic	25, 50 and 75 μm	6 hrs	Curcumin was found to inhibit LPS-induced cytokine production, includingmip-1 α , MIP-1 β , IL-6, IL-8 (CXCL-8) and GRO- α .	[15]
Myeloma cells	10 μm and 100 μm	30 min, 8 hrs	Curcumin was a potent inhibitor of STAT3 phosphorylation, and this plays a role in the suppression of MM proliferation	[16]
Human mantle cell lymphoma	10, 25, 50 and 100 μm	8 hrs.	Downregulated constitutive active NF-kb and inhibited the constitutively active IkB kinase (IKK), and phosphorylation of IkB and p65. Curcumin also	[17]

			inhibited constitutive activation of Akt, needed for IKK activation.	
Dendritic cells	0, 1, 5 10 and 25 μm	45 min	Induction of Th1 responses and a normal cell- mediated immune response.	[18]
SUIT-2	0 μm, 10 μm, 50 μm, and 100 μm	2 hrs	<i>IL-8</i> was inhibited NF-kb activity was reduced	[19]
EAC cell lines, OE33, and OE19	50mm nano- curcumin	48 hrs	Decreased the secretion of pro-inflammatory cytokines from <i>in vitro</i> activated T cells	[20]
CD4+ T cells	0.5, 1.0, 2.5, 5.0 mg	48 hrs	Inhibition of IL-12 production by macrophages stimulated in vitro with either LPS or HKL, leading to the inhibition of Th1 cytokine profile (decreased IFN- γ and increased IL-4 production) in CD4+ T cells. These findings suggest that curcumin may inhibit Th1 cytokine profile in CD4+ T cells by suppressing IL-12	[21]
T cells	20 µg	18 hrs	Decreases IL-12-induced STAT4 phosphorylation, IFN- γ production, and IL-12 R β 1 and β 2 expressions.	[22]
CD 4+ monocytes	20μm or 30μm		Cytokine and chemokine expression and reducing both migration and endocytosis	[23]
CD8 cytotoxic T lymphocytes Mice	10 mm	24 hrs	Inhibited the suppressive activity of Treg cells by downregulating the production of TGF-b and IL-10 in these cells	[24]
T-cell lymphoma cell lines	5–20 mm	48 hours	Curcumin selectively induces apoptosis in association with the downregulation of STAT-3 and NF-kb signaling pathways in CTCL cells	[25]
Human peripheral blood mononuclear Cells	With dose of 0.01 0.1, 1, 5 & 10 µm for 48 hours	48 hrs.	Shift the Th1–Th2-type immune balance towards Th2-type immunity	[26]
BV2 cell line	5 and 10 µm		Inhibited the phosphorylation of STAT1 and 3 as well as JAK1 and 2 in microglia activated with gangliosides, LPS, or IFN- γ .	[27]
Colonic myofibroblasts	5, 10 and 20 μm	30 min, 24 hrs	Reduced p38 MAPK activation in curcumin-treated mucosal biopsies enhanced IL-10 and reduced IL-1b. We demonstrate dose-dependent suppression of MMP-3 in CMF with curcumin.	[28]
T cell	2.5, 5 & 10 μm	24 hrs.	Curcumin suppresses NF-кВ activation.	[29]
Murine macrophage-like cells RAW264.7	1, 25, 50 μm	12 hr [30]s.	Inhibited the production of pro-inflammatory cytokine IL-18 in E. Coli LPS stimulated murine macrophage-like cells RAW264.7	[30, 31]
Carcinoma cell lines of the Pancreas (bxpc-3, Hs-700T and Hs- 766T, aspc-1, PANC-1, Capan-1 and Capan-2)	0.1, 1, 10 and 100 μm		Curcumin (diferuloylmethane), NF-κB inhibitor	[32]
Murine spleen lymphocytes	0, 3. 125, 6. 25, 12.5, 25, 50, 100, 200/µmol/L	24 hrs	Suppress the expression of NF~B p65	[33]

Human monocytes	1.5-12.5µm	72 hrs	Suppressed NF-kB binding and cytokine Release in THP-1 cells	[34]
Cytotoxic T lymphocytes	12.5–30 mmol/L	8 hrs	Interleukin-2 (IL-2) or alloantigen-induced proliferation of splenic lymphocytes, and development of cytotoxic T lymphocytes is significantly suppressed	[35]
Mouse RAW264.7 macrophages	1–100µm	1 hrs	Inhibition of NF- κ B nuclear translocation as well as the induction of downstream inflammatory mediators including pro-inflammatory cytokine mrna and protein (tumor necrosis factor- α , interleukin-1 β , and interleukin-6).	[36]
Human monocytic macrophage cell line	0.3, 2.5 and 5 μm	30-6- min	Inhibited lipopolysaccharide (LPS)-induced production of TNF and IL-l by a human monocytic macrophage cell line, Mono Mac 6.	[37]
Human keratinocyte cell line HACAT	20 µm	24 hrs	Inhibited the expression of TNF- α -induced IL-1 β , IL- 6, and TNF- α , but not IL-8, in TNF- α -treated HACAT cells as well as the TNF- α -induced cyclin E expression. In addition, curcumin inhibited the activation of MAPKS (JNK, p38 MAPK, and ERK) and NF- κ B in TNF- α -treated HACAT cells.	[38]
Dendritic cells	25 µm	24 hrs	Reduced IL-6 and IL-23	[39]
CD4+ T cell cultures of SLE patients	0.1 and 1 μg/ml	72 hrs	Reduce IL-17A	[40]
Dendritic cells	25 mm		Development of foxp3+ Tregs and reduced subsequent alloproliferative capacity	[41]
Dendritic cells	10 or 50 mm	24 hrs	Curcumin-treated DC induced differentiation of native CD41 T cells into Treg resembling Treg in the intestine, including both CD41CD251 Foxp31 Treg and IL-10- producing Tr1 cells. Such Treg induction required IL-10, TGF-b and retinoic acid produced by curcumin-modulated DC.	[42]
Human CD4+ T cells	0.2 or 2 mg/ml of Curcumin	3 days	Curcumin not merely blocks, but regulates CD2/CD3/CD28-initiated CD4+ T cell activation by augmenting CD69, CCR7, L-selectin and TGF-b1 expression followed by regulatory T cell generation	[43]
CD4+CD25+ T cells	5, 10, 20 μM	36 hours	Suppress IL-2 production Foxp3 expression was also reduced On Tregs after curcumin stimulation	[44]
T-cell	60 mmol/L	24 hrs	CD8 T-cell activation was significantly Increased when cocultured with curcumin-pretreated	[45]

Table 2.	In vivo and	l clinical studio	es that have a	evaluated the	effects of	curcumin on	Th balance.
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Animal	Curcumin dose	Duration	Route	Th1 Th2 Th17 Treg	Reference
Rat	(30 mg·kg-1·d-1, ip) for 15 days	15 days	i.p	Decreased the expression of Th1 cytokines increase the expression of Th1 cytokines	(46)
Rat	200 mg/kg	2, 4 & 6 hrs	i.p	Increase in messenger RNA levels and protein content of tumor necrosis factor- α ,	(47)

				interleukin-6, and matrix metalloproteinase- 9	
Rat	50 mg/kg (low- dose group) or 100 mg/kg curcumin (high- dose group)	45 days	i.p	Lowering circulating TNF-α concentration	(48)
Mouse	300 µg curcumin (Sigma) in 1% carboxymethyl cellulose		oral	Inhibited the phosphorylation of the p65 subunit of NF-κB in BMMCs	(49)
Rat	50 or 100 μg	days 0 to 25		Inhibits EAE by blocking IL-12 signaling in T cells and suggest its use in the treatment of MS and other Th1 cell-mediated inflammatory diseases	(50)
Mouse	Curcumin gel (curcumin 1 g, HPC 3 g, azone 1 g, ethanol (96%) 17 g, and distilled water q.s. to 100 g.	10 days		Was capable of impacting the IL-23/IL-17A axis by inhibiting IL-1b/IL-6 and then indirectly down-regulating IL-17A/IL-22 production.	(51)
Mouse	curcumin (0.1– 1%)	Diet		Curcumin demonstrates limited effectiveness on Th-1 mediated colitis in IL- 10-/- mice,	(52)
Rat	100 mg/kg	6 days	intragastric	Reduce serum TNF- α and IL-6 levels	(53)
Rat	80 mg/kg body weight/day	21 days	oral	Th1/Th2 balance through upregulation and downregulation of different factors	(54)
Rat	30 mg	15 days	i,p	Increased proportion of IFN- γ /IL-4 in splenocytes and circulation. Dex and Cur+Dex decreased the expression of Th1 cytokines but could not increase the expression of Th2 cytokines and the proportion of IFN- γ /IL-4. Cur exerted therapeutic effects on colitis by regulating the shift from Th1 to Th2	(46)
Mouse	curcumin gel	12 days		Relieving TPA-induced inflammation by directly down-regulating IFN _γ production	(55)
Human	10 μm	30 days		After the curcumin therapy, the Forkhead box protein (Foxp) 3 positive Treg frequency was markedly reduced, the frequency of Th1 cells was significantly increased in Cca patients.	(30)
Rat	50–100 mg/kg/day	2 week	Oral	Curcumin significantly attenuated the damage and caused substantial reductions of the rise in MPO activity and tumor necrosis factor alpha (TNF)- α . Also, curcumin was able to reduce nitrites colonic levels and induced downregulation of COX-2 and iNOS expression, and a reduction in the activation of p38 MAPK;	(56)

Mouse	50, 100 and 300 mg/kg	10 days		Curcumin also reduced the levels of nitric oxide (NO) and O2-associated with the favorable expression of Th1 and Th2	(57)
				cytokines and inducible NO synthase. These findings suggest that curcumin or diferuloylmethane, a major component of the food flavor turmeric, exerts beneficial effects in experimental colitis and may, therefore, be useful in the treatment of IBD.	
Murine	360mg/dose	3 or 4 times/day for three months		The inhibitory effects of curcumin on major inflammatory mechanisms like COX-2, LOX, TNF- α , IFN- γ , NF-kB and its unrivaled safety profile suggest that it has bright prospects in the treatment of IBD	(58)
Rat	20 mg	20 days	i.p	downregulation of Th1 cytokine response and NO production by macrophages, and their upregulation in NK cells	(59)
Rat	1% Cur or 0.02%	2 week	diet	suppressed (P, 0.05) NF-kB p65 nuclear translocation in activated CD41 T-cells.	(60)
Muse	10 mM; 100 μl/mouse	28 days	IP	up-regulated activities of the antioxidant enzymes, and suppressed serum levels of TNF- α and IL-1 β .	(61)
Mouse	250 μg		intragastric	suppression of Th2 responses as evidenced by reduced IL-4 and IL-13 production and depletion of eosinophils in the lungs, and attenuation of lung inflammation through expression of molecules such as TSLP, MMP-9, and OAT.	(62)
Mouse	5 and 10 mg/kg	9th to 14th day	i.n	Th2 cytokine, IL-5 is responsible for eosinophil and neutrophil recruitments to the lungs and TNF-a release.	(63)
male BALB/c and SJL/J mice	2 %	7 days	diet	Secretion of IL-4 and IL-5 by CD4+ lymphocytes of BALB/c mice but not SJL/J mice was significantly augmented	(64)
Mice	3 mg/ kg or 30 mg/kg	16 days	oral	maintaining Th1/Th2 balance	(65)
Rat	(50–100 mg/kg/day)	2 week	Oral gavage	downregulation of COX-2 and iNOS expression, and a reduction in the activation of p38 MAPK	(56)
Mouse	(50 mg/kg, 100 mg/kg, 200 mg/kg).	28 days	ip	Th17 cells and significantly increased Treg cells.	(66)
Mouse	100 µg	14 days		IL-12 and IL-23 that decreased after treatment with curcumin.	(67)
Mouse	2%	22 days	diet	enhanced STAT3 phosphorylation and suppressed expression of <i>Il12b</i> and <i>Il23a</i> .	(68)
Mouse	1%	18 and 32 week age	diet	decreased the proteinuria level and serum levels of IgG1, IgG2a and anti-dsDNA IgG antibodies in NZB/W F1 female mice	(69)
Rat	40 mg/kg	7 days	gavage	Curcumin pretreatment significantly inhibited the expression of IL-1 β , IL-2, and	(70)

				IL-6.	
Mouse	7mg/mL	5 days	ip	CD8 T-cell activation was significantly increased when co-cultured with curcumin pretreated	(45)
Mouse	Nanoparticle curcumin	7 days		suppressed mucosal mRNA expression of TNF- α , IL-1 β , IL-6, CXCL1 and CXCL2 in colonic epithelial tissues.	(71)

Figure 1. Effect of curcumin on T helper cells differentiation, survival, proliferation, polarization, and functions. Curcumin has potent immunosuppressive and immunomodulatory effects on T helper cells in-vitro and in-vivo. Curcumin has the potency to selectively enhance Tregs percentage and function and suppress macrophages and DCs maturation-mediated Th1 and Th17 differentiation. Therefore, curcumin may be an effective therapeutic agent for Th1-, Th2-, and Th17-driven inflammatory and autoimmune diseases and its use may thus be useful as a good strategy to treat inflammatory and autoimmune diseases. The effects of curcumin on Th9 and Th22 in unclear and need investigations.

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