



OPEN ACCESS

Edited by:

Daniela Ceccarelli,
Wageningen Bioveterinary Research
(WBVR), Netherlands

Reviewed by:

Jean-Yves Madec,
Agence Nationale de Sécurité
Sanitaire de l'Alimentation, de
l'Environnement et du Travail (ANSES),
France
Ákos Tóth,
National Public Health Institute (OKI),
Hungary

***Correspondence:**

Nicole Roschanski
nicole.roschanski@fu-berlin.de

†Present Address:

Nicole Roschanski,
Institute of Microbiology and
Epizootics, Freie Universität Berlin,
Berlin, Germany
Sebastian Guenther,
Institute of Pharmacy, University of
Greifswald, Greifswald, Germany
Beatriz Guerra,
Biological Hazards and Contaminants,
European Food Safety Authority,
Parma, Italy

Specialty section:

This article was submitted to
Antimicrobials, Resistance and
Chemotherapy,
a section of the journal
Frontiers in Microbiology

Received: 10 January 2018

Accepted: 08 March 2018

Published: 27 March 2018

Citation:

Roschanski N, Fischer J,
Falgenhauer L, Pietsch M,
Guenther S, Kreienbrock L,
Chakraborty T, Pfeifer Y, Guerra B and
Roesler UH (2018) Retrospective
Analysis of Bacterial Cultures Sampled
in German Chicken-Fattening Farms
During the Years 2011–2012 Revealed
Additional VIM-1
Carbapenemase-Producing
Escherichia coli and a Serologically
Rough *Salmonella enterica* Serovar
Infantis. *Front. Microbiol.* 9:538.
doi: 10.3389/fmicb.2018.00538

Retrospective Analysis of Bacterial Cultures Sampled in German Chicken-Fattening Farms During the Years 2011–2012 Revealed Additional VIM-1 Carbapenemase-Producing *Escherichia coli* and a Serologically Rough *Salmonella enterica* Serovar Infantis

Nicole Roschanski^{1*†}, Jennie Fischer², Linda Falgenhauer³, Michael Pietsch⁴, Sebastian Guenther^{1†}, Lothar Kreienbrock⁵, Trinad Chakraborty³, Yvonne Pfeifer⁴, Beatriz Guerra^{2†} and Uwe H. Roesler¹

¹ Institute for Animal Hygiene and Environmental Health, Freie Universität Berlin, Berlin, Germany, ² Department for Biological Safety, Federal Institute for Risk Assessment, Berlin, Germany, ³ German Center for Infection Research, Institute of Medical Microbiology, Justus Liebig University Giessen, Partner Site Giessen-Marburg-Langen, Giessen, Germany, ⁴ FG13 Nosocomial Pathogens and Antibiotic Resistance, Robert Koch Institute, Wernigerode, Germany, ⁵ Epidemiology and Information Processing and WHO Collaborating Center for Research and Training for Health at the Human-Animal-Environment Interface, Institute for Biometry, University of Veterinary Medicine Hannover, Hannover, Germany

Carbapenems are last-resort antibiotics used in human medicine. The increased detection of carbapenem-resistant Enterobacteriaceae (CRE) is therefore worrying. In 2011 we reported the first livestock-associated VIM-1-producing *Salmonella* (*S.*) *enterica* serovar Infantis (R3) isolate from dust, sampled in a German chicken fattening farm. Due to this observation we retrospectively investigated more than 536 stored bacterial cultures, isolated from 45 chicken fattening farms during the years 2011 and 2012. After a non-selective overnight incubation, the bacteria were transferred to selective media. *Escherichia* (*E.*) *coli* and *Salmonella* growing on these media were further investigated, including antibiotic susceptibility testing, carbapenemase gene screening and whole genome sequencing (WGS). In total, four CRE were found in three out of 45 investigated farms: Besides R3, one additional *Salmonella* (G-336-1a) as well as two *E. coli* isolates (G-336-2, G-268-2). All but G-268-2 harbored the *bla*_{VIM-1} gene. *Salmonella* isolates R3 and G-336-1 were closely related although derived from two different farms. All three *bla*_{VIM-1}-encoding isolates possessed identical plasmids and the *bla*_{VIM-1}-containing transposon showed mobility at least *in vitro*. In isolate G-268-2, the AmpC beta-lactamase gene *bla*_{CMY-2} but no known carbapenemase gene was identified. However, a transfer of the phenotypic resistance was possible. Furthermore, G-268-2 contained the *mcr-1* gene, combining phenotypical carbapenem- as well as colistin resistance in one isolate. Carbapenem-resistant Enterobacteriaceae have been

found in three out of 45 investigated chicken flocks. This finding is alarming and emphasizes the importance of intervention strategies to contain the environmental spread of resistant bacteria in animals and humans.

Keywords: antimicrobial resistance, plasmids, livestock, carbapenems, VIM-1 producing Enterobacteriaceae, Germany, broiler

INTRODUCTION

Carbapenem resistance increased in many countries during the last years, causing serious problems in the public health system (Wilson, 2017). As carbapenems serve as one of the last remaining options for the treatment of serious infections caused by multidrug-resistant Gram-negative bacteria (e.g., Enterobacteriaceae, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*), the recent situation is alarming. In terms of Gram-negative bacteria, mainly the expression of carbapenemases leads to a decreased carbapenem-susceptibility. Most frequently detected genes are *bla*_{VIM}, *bla*_{IMP}, *bla*_{NDM} (class B β -lactamases), *bla*_{KPC} (class A β -lactamases) and *bla*_{OXA-48-like} (class D β -lactamases) (Nordmann et al., 2012). Their location on mobile genetic elements though contributes to a successful spread of these resistance genes (Nordmann et al., 2012; Carattoli, 2013). Within the last couple of years, the occurrence of carbapenemase-producing bacteria relocated from clinical settings. Carbapenemase-producing bacteria have been isolated from the environment (Walsh et al., 2011; Zurfluh et al., 2013), wild-birds (Fischer et al., 2013b), seafood (Rubin et al., 2014; Morrison and Rubin, 2015; Roschanski et al., 2017b), companion- and food-producing animals all over the world (Stolle et al., 2013; Mollenkopf et al., 2016; Ewers et al., 2017; Fischer et al., 2017; He et al., 2017). In terms of food-producing animals, beside pigs and cattle another focus was chicken and chicken meat. In 2011, the first VIM-1 producing *Salmonella enterica* subsp. *enterica* serovar Infantis (*S. Infantis*)—isolate R3—was isolated on a German chicken fattening farm (Fischer et al., 2013a). This isolate was closely related to other *S. Infantis* isolates detected in tree pig fattening farms in the same year (Fischer et al., 2017), based on identical macrorestriction patterns and the presence of a *bla*_{VIM-1}-carrying IncHI2 plasmid of 300 kb. Therein, the carbapenemase gene was located within a class 1 integron embedded in a Tn21 homolog (Falgenhauer et al., 2017). Since 2015, additional publications described the finding of carbapenemase-producing bacteria in chicken or retail chicken meat. Carbapenemase-producing Enterobacteriaceae (CPE) were detected in retail chicken meat and in broiler farms in Egypt (Abdallah et al., 2015), and quite recently, the prevalence of NDM and Mcr-1 in Chinese poultry production as well as VIM-positive *Pseudomonas* species in Chinese chicken and their surrounding has been described (Wang et al., 2017; Zhang et al., 2017). To follow-up on the study of Fischer et al. (Fischer et al., 2013a), 536 stored bacterial cultures, isolated from 45 chicken-fattening farms as well as 125 stored single colony cultures derived from the previously *bla*_{VIM-1}-positive chicken farm 1, were retrospectively investigated for the

presence of carbapenem-resistant *E. coli* as well as *Salmonella* isolates.

MATERIALS AND METHODS

Bacterial Cultures and Screening for Carbapenem-Resistant Isolates Primary Mixed Bacterial Cultures

In total, 536 primary bacterial cultures, isolated from pooled feces, pooled dust as well as boot swab samples were retrospectively investigated. The samples were initially taken in terms of the first period of the national research project RESET (www.reset-verbund.de), carried out during the years 2011–2013, and focused on screening for ESBL and AmpC-producing Enterobacteriaceae in different habitats (Laube et al., 2013; Hering et al., 2016). Therein, an overall number of 45 chicken fattening farms distributed throughout Germany have been investigated. Briefly, the samples were taken from each farm, incubated non-selectively in LB broth (Luria/ Miller), (Carl Roth, Karlsruhe, Germany), followed by a selective cultivation on MacConkey agar plates containing 1 mg/L cefotaxime (293 cultures – 55%) or Endo-agar containing 2 mg/L enrofloxacin (243 cultures – 45%). Mixed sets of bacteria, able to grow on these plates (primary cultures), were taken from the plates and stored in Cryobank™ (MAST Diagnostica, Reinfeld, Germany) at –80°C (Laube et al., 2013; Hering et al., 2016). For our retrospective analyses, the stored bacteria were re-cultured non-selectively in LB broth at 37°C, overnight. Each culture was spread on selective MacConkey agar plates (MacConkey agar No 3; OXOID, Hampshire, UK) containing 0.125 mg/L meropenem, (Sigma Aldrich, Seelze, Germany). *E. coli* and *Salmonella* colonies were isolated and species were confirmed using MALDI-TOF mass spectrometry (MALDI Microflex®LT and Biotyper® database, Bruker Daltronics, Bremen, Germany). Per sample and species, one colony was picked and subsequently spread on chromID™-CARBA agar plates (bioMérieux, Nürtingen, Germany). Thereon grown colonies (one per sample and species) were further investigated for the presence of carbapenemase genes (*bla*_{VIM}, *bla*_{KPC}, *bla*_{NDM}, *bla*_{OXA-48}, and *bla*_{GES}) using real-time PCR (Roschanski et al., 2017a). The presence of *bla*_{IMP} was checked in a conventional PCR format (van der Zee et al., 2014).

Single Colonies Derived From Chicken-Farm 1

In a separate investigation, performed in the department of biological safety of the Federal Institute for Risk Assessment, 120 stored single colony cultures (111 *E. coli*, 5 *Pseudomonas*, 4 *Acinetobacter*), derived from the previously VIM-positive tested

chicken farm 1 (*S. Infantis* R3), were screened for the presence of further *bla*_{VIM-1}-encoding isolates. The cultures derived from one of the seven investigated chicken fattening farms which were sampled in terms of a longitudinal study (three samplings per fattening period) conducted by the FU Berlin (Laube et al., 2013).

The cultures were taken from the -80°C stock and non-selectively re-cultured. On the following day the overnight cultures were transferred to LB broth containing 1 mg/L imipenem (1:500). In case of bacterial growth, cultures were diluted 1:10 and used as template for amplification of the *bla*_{VIM-1} gene by PCR. Cultures tested positive by PCR were subsequently spread on selective agar plates for single colony isolation and their *bla*_{VIM-1} confirmation by PCR as previously described (Fischer et al., 2017).

Antimicrobial Susceptibility Testing and Blue-Carba Assay

Minimal inhibitory concentrations for the wildtypes as well as the transformants were determined by using the VITEK-2[®] compact system and the AST-card N248 (bioMérieux, Nuertingen, Germany). The Blue-Carba assay for confirmation of carbapenemase activity was performed in two technical replicates as previously described (Pires et al., 2013).

Genotypic Investigation of Wildtype and Transformant Isolates

Plasmids were isolated using the NucleoBond Xtra Midi kit (Macherey-Nagel, Dueren, Germany) and transferred into *E. coli* NEB[®]10-beta or NEB[®]5-alpha (NEB, Frankfurt a. M., Germany) by electroporation (2.5 kV). WGS was performed for the wildtype strains (*Salmonella* R3 and G-336-1a; *E. coli* G-336-2 and G-268-2) as well as the G-336-1a- and G-336-2-transformants T_{G-336-1a}, T_{G-336-2_VIM}, T_{G-336-2_CMY+VIM}, T_{G-336-2_CMY} using MiSeq (Illumina) (Borowiak et al., 2017). In addition, the *E. coli* recipient strain NEB[®]10-beta was sequenced. The raw-data were *de novo* assembled using SPAdes (Bankevich et al., 2012). An additional assembly was performed for G-268-2 by A5-miseq (v. 0.0.9 beta; default parameters) using trimmed raw reads (Trimmomatic: v. 0.0.9; default parameters except maxinfo 15:0.5) (Bolger et al., 2014; Coil et al., 2015). Resistance genes, virulence genes, plasmid incompatibility groups as well as multilocus sequence types were identified using the Web-tools ResFinder (Zankari et al., 2012), PlasmidFinder (Carattoli et al., 2014) and MLST 1.8 (Larsen et al., 2012). In addition, the Resistance Gene Identifier (RGI) was used (Jia et al., 2017). The genetic relationship of the two *S. Infantis* isolates R3 and G-336-1a was determined by using the CSI Phylogeny-1.4 Server (Kaas et al., 2014). Therefore the raw reads were uploaded and mapped against a *S. Infantis* reference sequence (LN649235.1), (Olasz et al., 2015). Parameters were set as follows: Minimum depth at SNP position: 10, relative depth at SNP position: 10, minimum distance between SNPs (prune): 10, minimum SNP quality: 30, minimum read mapping quality: 25 as well as minimum Z-score: 1.96.

A prediction of Inc RNA folding was performed using the RNAfold WebServer (<http://rna.tbi.univie.ac.at/cgi-bin/RNAWebSuite/RNAfold.cgi>) (Gruber et al., 2008). Furthermore, a subsequent plasmid comparison was performed using the

BLAST Ring Image Generator (BRIG) (Alikhan et al., 2011). For this purpose contigs containing plasmid sequences were plotted against two already published *bla*_{VIM-1} containing IncHI2-plasmid sequences derived from a *S. Infantis* as well as an *E. coli* isolated on a German pig fattening farm (pRH-R27, LN555650.1 and pRH-R178 HG530658.1) (Falgenhauer et al., 2017). Whole genome data of all isolates have been deposited in the European Nucleotide Archive (<https://www.ebi.ac.uk/ena>) of the European Bioinformatics Institute (EMBL-EBI); accession numbers R3 (ERS2154041), G-336-1a (ERS2101552), G-336-2 (ERS2101553), G-268-2 (1969-10-8; ERS2101554), NEB[®]10-beta (ERS2101551), T_{G-336-1a} (FU11995; ERS2101550), T_{G-336-2_VIM} (FU12739; ERS2101547), T_{G-336-2_CMY+VIM} (FU12738; ERS2101548), T_{G-336-2_CMY} (FU11994; ERS2101549).

The phenotypic carbapenem resistance of G-268-2 as well as its transformant T_{G-268-2} was further characterized by an additional screening for the presence of the outer-membrane protein genes *ompC* and *ompF*. Therefore, the whole genome reads were mapped against respective *E. coli* K-12 MG1655 (NC_000913.3) reference sequences. In addition, a PCR-based screening was performed, using the primer pairs Ec_OmpC-fwd – ATGAAAGTTAAAGTACTGTCCCTCC, Ec_OmpC-rev. – TTAGAAGTGGTAAACCAGACCCA (1,150 bp), Ec_OmpF-fwd. – ATGATGAAGCGCAATATTCTGG, Ec_OmpF-rev. – TTAGAAGTGGTAAACGATACCCACA (1,190 bp) as well as primers described by Lartigue et al. (2007).

Classical Bacterial Strain Typing

Classification of the *E. coli* isolates into one of the eight described phylogenetic groups was done in accordance to the protocol of Clermont et al. (2013). Serotyping of *Salmonella* isolates was performed in the German National Reference Laboratory for the Analysis and Testing of Zoonoses (NRL Salmonella - BfR, Berlin) according to the White-Kauffmann-Le Minor scheme (Grimont and Weill, 2007). Genetic relatedness of the *Salmonella* isolates was analyzed using XbaI-restriction of bacterial DNA and subsequent pulsed-field gel electrophoresis (PFGE) (Ribot et al., 2006). PFGE was conducted using the CHEF-DR III system (Bio-Rad Laboratories GmbH, Munich, Germany) using a 1.1% agarose gel (Biozyme LE GP agarose; Biozym Scientific GmbH, Hessisch Oldendorf, Germany). The following conditions were used: Initial switch time 5 s, final switch time 50 s at a gradient of 5.6 V/cm and an included angle of 120 V. The run time was 21 h at a system temperature of 14°C. For plasmid characterization, S1-nuclease restriction and PFGE (Guerra et al., 2004) was performed using the following running conditions: 1-25 s, 17 h, 6 V/cm, 120 V.

RESULTS

Occurrence of Carbapenem-Resistant Isolates Within German Chicken Fattening Farms

In 2011 the first VIM-1-producing *S. Infantis* (R3) was isolated from dust sampled on a German chicken fattening farm. The primary mixed bacterial culture from this chicken farm was included in this study and served as an internal identification

control for the applied isolation procedure of CPE. Besides R3, one additional *Salmonella* isolate, serologically typed as subspecies I (rough phenotype, G-336-1a) as well as two *E. coli* isolates (G-336-2, G-268-2) were isolated from the selective agar plates. The real-time PCR-based screening of the new isolates indicated the presence of the *bla*_{VIM-1} gene in G-336-1a and G-336-2. Both of them were found in one dust sample from a chicken fattening farm in South Germany. Apparently, there was no regional connection to the previously described chicken fattening farm (isolate R3) which was located in the eastern part of Germany. G-268-2 was isolated from a boot swab sample originated from a third chicken fattening farm located in East Germany. For this *E. coli* isolate none of the six investigated carbapenemase genes (*bla*_{VIM}, *bla*_{KPC}, *bla*_{NDM}, *bla*_{OXA-48}, *bla*_{GES}, and *bla*_{IMP}) was detected.

Detailed Characterization of *bla*_{VIM-1} Containing *Salmonella* and *E. coli* Isolates

An overview of the whole genome results is provided in Table 1. These data confirm the relatedness of both *Salmonella* isolates (R3 and G-336-1). They belonged to the multilocus sequence type ST32 and both, PFGE as well as SNP analysis, showed only small differences of 2 bands (data not shown) and 8 SNPs, respectively. These data reveal that G-336-1 just as R3, genotypically belongs to the serovar Infantis. The *bla*_{VIM-1}-encoding plasmids (IncHI2; 300 kb) derived from the two *S. Infantis* isolates (R3, G-336-1a) as well as the *E. coli* (G-336-2) were compared with the IncHI2 plasmid sequences of *E. coli* R178 as well as *S. Infantis* R27, both isolated in a German pig-fattening farm (Falgenhauer et al., 2017). An identity of 100% was detected between the plasmids of R3, G-336-1a and R27 (*Salmonella*) as well as

the *bla*_{VIM-1} containing plasmid of *E. coli* G-336-2 (Figure 1). In all four plasmids the *bla*_{VIM-1} gene was part of a class 1 integron accompanied by the genes *aacA4* and *aadA1* in its variable region. As described previously, the integron was inserted in a Tn21 homolog (Falgenhauer et al., 2017). On the same plasmid, the AmpC gene *bla*_{ACC-1} was detected. S1-PFGE indicated that *E. coli* isolate G-336-2 (sequence type ST131) contained two additional plasmids, one of them an IncI1 plasmid, carrying the AmpC gene *bla*_{CMY-2} (Figure 2). Proofed by WGS, it was possible to show that in course of *in vitro* cultivation and transformation experiments, one *E. coli* transformant was received, which contained the *bla*_{VIM-1}-encoding transposon integrated into the *pilU* gene of the *bla*_{CMY-2}-encoding IncI1 plasmid (Figure 2). This indicates that at least *in vitro* the transposon is highly mobile and self-transmissible.

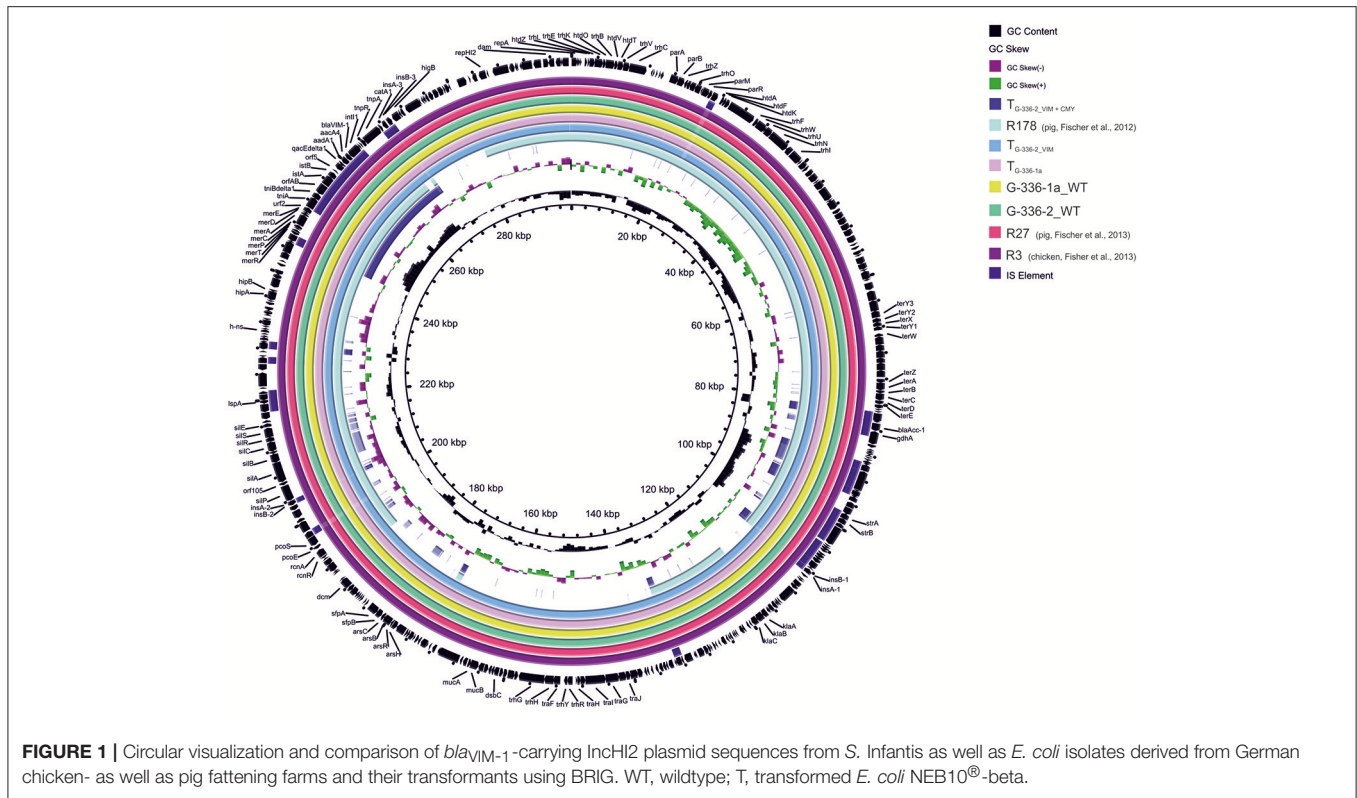
The Carbapenem Resistance of the *E. coli* Isolate G-268-2 Remains Unclear

In case of the second *E. coli* isolate G-268-2 (ST-354) neither ResFinder (Zankari et al., 2012) nor RGI (Jia et al., 2017) indicated the presence of a known carbapenemase gene. Data assembly using two different algorithms (SPAdes and A5-miseq) provided identical results. Besides several other antibiotic resistance genes, this isolate contained the colistin resistance gene *mcr-1* encoded on an IncX4 plasmid (Table 1). Resistance to colistin (MIC = 8mg/L) was confirmed for G-268-2 and its transformant T_{G-268-2_mcr-1} (Table 2). Moreover, the increased MIC for meropenem (4 mg/L) and imipenem (MIC = 8 mg/L) in another G-268-2 transformant indicated a transferrable carbapenem resistance. While the plasmid-encoded colistin resistance was transferred separately (IncX4 plasmid),

TABLE 1 | Characteristics of carbapenem-resistant *E. coli* and *S. Infantis* isolates including their respective transformants derived from whole genome data analyses.

Wildtype-isolate/ transformants	Farm no.	Species [MLST]	Resistance genes	Plasmids [pMLST]
R3*	1	<i>Salmonella</i> Infantis [ST-32]	<i>aac(6')/lb-cr</i> -like, <i>aacA4</i> -like, <i>aadA1</i> , <i>bla</i> _{ACC-1} , <i>bla</i>_{VIM-1} , <i>catA1</i> -like, <i>ere(A)</i> -like, <i>strA</i> , <i>strB</i> , <i>sul1</i> -like	IncHI2 [ST-1]
G-336-1a	78	<i>Salmonella</i> subspecies I [ST-32]	<i>aac(6')/lb-cr</i> -like, <i>aacA4</i> -like, <i>aadA1</i> , <i>bla</i> _{ACC-1} , <i>bla</i>_{VIM-1} , <i>catA1</i> -like, <i>strA</i> , <i>strB</i> , <i>sul1</i> -like	IncHI2 [ST-1]
T _{G-336-1a}	–	<i>E. coli</i> [ST-1060]	<i>aac(6')/lb-cr</i> -like, <i>aacA4</i> -like, <i>aadA1</i> , <i>bla</i> _{ACC-1} , <i>bla</i>_{VIM-1} , <i>catA1</i> -like, <i>strA</i> , <i>strB</i> , <i>sul1</i> -like	IncHI2 [ST-1]
G-336-2	78	<i>E. coli</i> [ST-131]	<i>aac(6')/lb-cr</i> -like, <i>aacA4</i> -like, <i>aadA1</i> , <i>bla</i> _{ACC-1} , <i>bla</i> _{CMY-2} , <i>bla</i>_{VIM-1} , <i>catA1</i> -like, <i>strA</i> , <i>strB</i> , <i>sul1</i> -like	IncI1 [ST-12], IncHI2 [ST-1], IncF [F18:A6:B1]
T _{G-336-2_VIM}	–	<i>E. coli</i> [ST-1060]	<i>aac(6')/lb-cr</i> -like, <i>aacA4</i> -like, <i>aadA1</i> , <i>bla</i> _{ACC-1} , <i>bla</i>_{VIM-1} , <i>catA1</i> -like, <i>strA</i> , <i>strB</i> , <i>sul1</i> -like	IncHI2 [ST-1]
T _{G-336-2_VIM+CMY}	–	<i>E. coli</i> [ST-1060]	<i>aac(6')/lb-cr</i> -like, <i>aacA4</i> -like, <i>aadA1</i> , <i>bla</i> _{CMY-2} , <i>bla</i>_{VIM-1} , <i>catA1</i> -like, <i>sul1</i> ,	IncI1 [ST-12]
T _{G336-2_CMY}	–	<i>E. coli</i> [ST-1060]	<i>bla</i> _{CMY-2}	IncI1 [ST-12]
G-268-2	54	<i>E. coli</i> [ST-354]	<i>aadA1</i> , <i>aadA2</i> , <i>aph(3')</i> -Ia-like, <i>bla</i> _{CMY-2} , <i>bla</i> _{TEM-1B} , <i>cmiA1</i> -like, <i>dfrA17</i> , <i>dfrA8</i> , <i>mcr-1</i> , <i>strA</i> , <i>strB</i> , <i>sul2</i> , <i>sul3</i> , <i>tet(B)</i>	ColRNAI, IncX1, p0111, IncX4, IncQ1, IncB/O/K/Z, IncF [F-:A1:B1]
RECIPIENT				
NEB10-beta		<i>E. coli</i> [ST-1060]	None	None

*Previously identified by Fischer et al. (2013a); carbapenemase genes in bold print.



the reduced carbapenem susceptibility was detected in the transformant containing the AmpC- β -lactamase encoding gene *bla*_{CMY-2} located on an IncB/O/K/Z-plasmid (Table 2). A sequence comparison of the 83,592 kb contig containing the *bla*_{CMY-2} gene of G-268-2 with available plasmid sequences in the GenBank database exhibited 99.98% nucleotide sequence identity (97.3% coverage) to the IncK2 plasmid pDV45 (KR905384) from an *E. coli* isolate from poultry meat (Seiffert et al., 2017). A minor difference was observed in the organization within the shuffleon region of *pilV*.

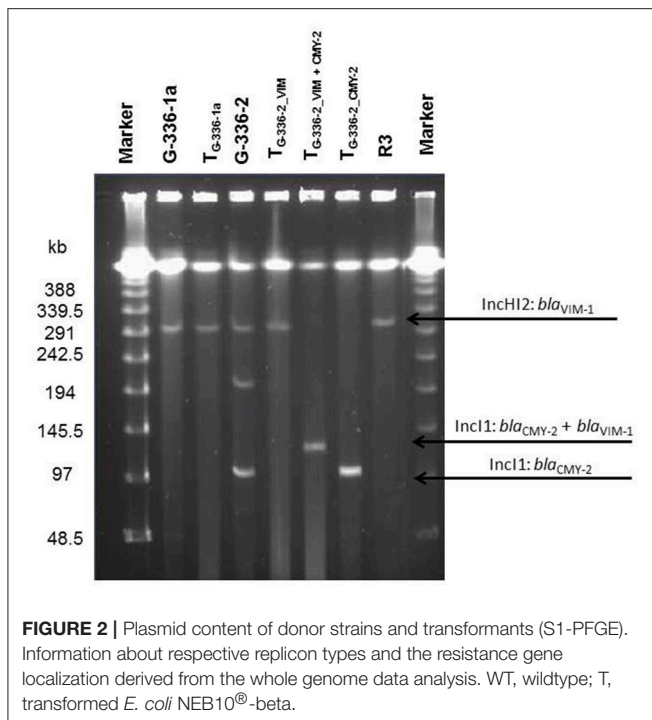
The *bla*_{CMY-2} gene itself was located within a mobile genetic element consisting of *ISEcp1* – *bla*_{CMY-2} – *blc* and *sugE1* as described by Seiffert et al. (2017).

As previously the correlation between carbapenem resistance and the combination of elevated CMY-2-production and porin deficiency has been shown (Goessens et al., 2013; van Bortel et al., 2017), the whole genome data of the G-268-2 wildtype isolate were additionally checked for mutations in associated regions. Neither the *bla*_{CMY-2} gene nor its promoter region showed any kind of modification. However, the average read coverage of the *bla*_{CMY-2} containing contig was 8.5-fold higher than the coverage of chromosomal contigs. Analysis of the plasmid copy number controlling antisense *incRNAI* upstream of *repA* showed a nucleotide substitution comparing to the *incRNAI* of plasmid pDV45 (KR905384) (Seiffert et al., 2017). Analysis by RNA folding prediction software (RNAfold WebServer) indicated that this nucleotide substitution might have an impact on the folding structure of the antisense *incRNA* (Supplementary Figure 1).

Mapping of the whole genome reads of G-268-2 against the *E. coli* K12 porin genes *ompF* as well as *ompC* exhibited a deletion of 19 nucleotides at nucleotide position 249 in the *ompC* gene, resulting in a frameshift and a premature stop codon at nucleotide position 285. *OmpF* showed 48 silent nucleotide substitutions and 22 nucleotide substitutions leading to amino acid substitutions and exhibited a deletion of 15 nucleotides in comparison to the *ompF* gene of *E. coli* K12 MG1655 (NC_000913.3). Subsequently performed PCR and Sanger sequencing of the *ompF*- and *ompC*-products supported this observation in both genes. As a control, the mapping of the whole genome reads of the recipient strain NEB10[®]-beta against both K12 *omp*-genes was performed and both of them were unaltered according to the reference. In contrast to G-268-2, 100% accordance to the *ompF* and *ompC* reference sequences (*E. coli* K12 MG1655) was detected for the transformant T_{G-268-2}. Moreover, the additionally performed Blue-Carba assay clearly indicated imipenem-hydrolyzing activity in case of the investigated wild-type isolate G-268-2 as well as its transformant T_{G-268-2}_{CMY-2} (Supplementary Figure 2).

Investigation of Additional Isolates Derived From the Initially VIM-1-Positive Tested Chicken Farm 1 Did Not Possess Additional Carbapenemase-Producing Isolates

The retrospective analysis of 125 bacterial isolates derived from three different samplings in chicken farm 1, revealed no additional VIM-1-producing isolates. Therefore, the *S. Infantis*



(R3) containing dust sample, which was included in analysis as a positive control remained the only *bla*_{VIM-1} positive sample in the investigated stable.

DISCUSSION

Although the here described study is based on a retrospective investigation of bacterial isolates sampled in the years 2011 and 2012, the finding of additional carbapenemase-producing Enterobacteriaceae within German chicken farms is worrying. Aware that in the same timeframe also *bla*_{VIM-1} positive *E. coli* and/or *S. Infantis* have been isolated from three German pig fattening farms (Fischer et al., 2017), it becomes obvious that the entry of carbapenemase-producing bacteria into livestock farms got on the way some years ago. Although three out of 45 chicken fattening farms (6.6%), harboring carbapenem-resistant bacteria have been identified in 2011/12 it has to be considered that this study was merely based on pooled feces, boot swabs as well as dust samples. Therefore, the received results provide just a brief overview of the situation within the investigated years and as a previously performed reinvestigation on a VIM-1-positive pig-farm has shown, changes within the resistance situation could occur over the years (Roschanski et al., 2016). However, newer publications indicate that the trend of finding carbapenemase-producing bacteria in livestock as well as food did not stop. Three recently published manuscripts described the finding of additional VIM-1-positive isolates in Germany: *E. coli* derived from the colon contents of slaughter pigs (Irrgang et al., 2017), as well as two *S. Infantis* isolated from minced pork meat and a sick piglet (Borowiak et al., 2017). All of the—so far—in Germany detected *bla*_{VIM-1} encoding

livestock associated *Salmonella* isolates belonged to sequence type ST32 and possessed a highly related plasmid (size 300kb, **Figure 1**). The latter *S. Infantis*, isolated in 2015 and 2016, showed a major homology to the previously detected *S. Infantis* (ST32) isolates R25 and R27 (pig farms) as well as R3 and G-336-1a (chicken farms). Therefore, the here described data as well as previous findings within the pig-production chain suggested a broad circularization in livestock animals (pig- as well as poultry). As in Germany the treatment of livestock with carbapenems is not licensed, a co-selection process over the years seemed to be most likely to explain the consistent re-occurrence of highly similar isolates or plasmids over the years. Moreover an additional report depicted the finding of a *bla*_{VIM-1} positive *E. coli* from a venus clam derived from a Berlin retail market (Roschanski et al., 2017b). However, compared to the *E. coli* isolates derived from livestock in this case neither a strain- nor a plasmid homology has been detected.

Though, not only in Germany an increased finding of carbapenemase producers in livestock or food has been described. *Klebsiella* containing *bla*_{NDM} have been detected in Egypt chicken retail meat and 35% of investigated Egyptian broilers were carrying NDM-, KPC- and or OXA-48- producing *Klebsiella pneumoniae* (Hamza et al., 2016). In 2017, the number of reports was even higher: VIM-positive *Pseudomonas* species were isolated from chicken and their surroundings in China (Zhang et al., 2017). Moreover, in China *bla*_{NDM-5} positive *K. pneumoniae* have been detected in dairy cows, while *bla*_{NDM}-producing *E. coli* have been identified in piglets housed in India (Pruthivishree et al., 2017). Furthermore, in the USA *bla*_{IMP-27} containing Enterobacteriaceae were recovered from the environment of a swine farrow-to-finish operation (Mollenkopf et al., 2016). This increase within the last 6 years is alarming and the development of intervention strategies are urgently needed to curtail a further spread of these bacteria. However, beside the bacteria themselves also mobile genetic elements like plasmids or transposons play an important role for the spread of carbapenemase genes. Like here described, farm 78 harbored an *E. coli* as well as a *S. Infantis* (serological typed as *S. subspecies I* with a rough phenotype), carrying the ca. 300 bp IncHI2 plasmid encoding the *bla*_{VIM-1} gene which was previously described for different *S. Infantis* isolates (Fischer et al., 2013a; Borowiak et al., 2017; Falgenhauer et al., 2017). Moreover, our *in-vitro* experiments demonstrated that the Tn21-like transposon harboring the *bla*_{VIM-1} gene was able to change its localization from the ca. 300 kb IncHI2 plasmid to a much smaller (ca. 100 kb) IncI1 plasmid. If this event happens in the farm surrounding, it might contribute to the spread of this carbapenemase gene even more efficiently. The exclusive finding of the VIM-1-producing isolates in dust samples of the two farms might be due to the high survival rates of *Salmonella* species in dust or dried manure. In 2015, viable *Salmonella* were detected for up to 291 days in manure dust with 5% moisture (Oni et al., 2015). The fact that the *bla*_{VIM-1}-containing isolates did not spread throughout the 2011 and 2012 investigated stables can be explained when the antimicrobial treatment of the two respective flocks is taken into account: While in farm

TABLE 2 | Antibiotic susceptibilities of wildtype strains and transformants.

Farm/region	Wildtype isolate/ transformants (†)	Species - ST	PIP	PTZ	CTX	CAZ	FEP	ATM	IMI	MEM	AMK	GEN	TBM	CIP	TGC	FOS	CST	SXT	
1/east	R3 (Fischer et al., 2013a)	<i>Salmonella</i> Infantis- ST32	≥128	≥128	≥64	≥64	8	≤1	8	≥16	≤2	2	8	0.5	≤0.5	≤16	≤0.5	≥320	
54/east	G-268-2	<i>E. coli</i> - ST354	≥128	≥128	≥64	≥64	8	≥64	≥16	8	4	≤1	2	≥4	≤0.5	≤16	8	≥320	
	T _{G268-2} _CMY	<i>E. coli</i> - ST1060	≥128	≥128	≥64	≥64	16	≥64	8	4	4	≤1	≤1	≤0.25	≤0.5	≤16	≤0.5	≤20	
	T _{G268-2} _Mcr-1	<i>E. coli</i> - n.d.	≤4	≤4	≤1	≤1	≤1	≤1	≤0.25	≤0.25	≤2	≤1	≤1	≤0.25	≤0.5	≤16	8	≤20	
78/south	G-336-1a	<i>Salmonella</i> subspecies I- ST32	64	64	32	≥64	≥64	≤1	≥16	disc.	≤2	≤1	2	0.5	≤0.5	≤16	≤0.5	≤20	
	T _{G336-1a}	<i>E. coli</i> - ST1060	≥128	≥128	≥64	≥64	16	≤1	8	8	≤2	≤1	4	0.5	≤0.5	≤16	≤0.5	≤20	
78/ south	G-336-2	<i>E. coli</i> - ST131	≥128	≥128	≥64	≥64	16	16	≥16	≥16	≤2	≤1	8	1	≤0.5	≤16	≤0.5	≤20	
	T _{G336-2} _VIM	<i>E. coli</i> - ST1060	≥128	≥128	≥64	≥64	16	≤1	4	disc.	≤2	≤1	8	0.5	≤0.5	≤16	≤0.5	≤20	
	T _{G-336-2} _VIM+CMY	<i>E. coli</i> - ST1060	≥128	≥128	≥64	≥64	≥64	≥64	8	≥16	≤2	4	≥16	0.5	≤0.5	≤16	≤0.5	≤20	
	T _{G336-2} _CMY	<i>E. coli</i> - ST1060	≥128	≥128	≥64	≥64	2	≥64	0.5	≤0.25	≤2	≤1	≤1	≤0.25	≤0.5	≤16	≤0.5	≤20	
RECIPIENTS																			
	NEB10-beta	<i>E. coli</i> - ST1060	≤4	≤4	≤1	≤1	≤1	≤1	≤0.25	≤0.25	≤2	≤1	≤1	≤0.25	≤0.5	≤16	≤0.5	≤20	
	NEB5-alpha	<i>E. coli</i> - n.d.	≤4	≤4	≤1	≤1	≤1	≤1	≤0.25	≤0.25	≤2	≤1	≤1	≤0.25	≤0.5	≤16	≤0.5	≤20	

PIP, piperacillin; PTZ, piperacillin-tazobactam; CTX, cefotaxime; CAZ, ceftazidime; FEP, cefepime; ATM, aztreonam; IMI, imipenem; MEM, meropenem; AMK, amikacin; GEN, gentamicin; TBM, tobramycin; CIP, ciprofloxacin; TGC, tigecycline; FOS, fosfomicin; CST, colistin; SXT, trimethoprim-sulfamethoxazole.
disc., run discontinued; n.d., not determined; ST, sequence type.

1 the animals remained untreated during the whole fattening period, in farm 78 an oral treatment with colistin-sulfate was performed for 4 days. As in both cases, no selection pressure was present to favor the spread of the carbapenemase gene-carrying plasmid; this might be a possible explanation for the rare finding of positive tested isolates. However, as the survival rate of *Salmonella* seemed to be pretty high within manure dust (Oni et al., 2015), follow-up investigations of the respective stables would have been desirable.

In addition, the presence of a transferable carbapenem resistance has been shown for the *E. coli* isolate G-268-2 and the assumed presence of a carbapenemase was supported by the Blue-Carba assay (Pires et al., 2013), indicating an imipenem-hydrolyzing activity (**Supplementary Figure 2**), however, no known carbapenemase gene has been detected within the whole genome data. However, Mammeri et al. reported a higher overall catalytic activity of CMY-2 for imipenem compared to the ones of other pAmpCs (Mammeri et al., 2010). Moreover, several publications depicted the combination of elevated *bla*_{CMY-2} expression caused by an increased plasmid copy number in combination with the lack of the outer membrane proteins OmpC and OmpF as a main reason for detected carbapenem resistance without the presence of a carbapenemase gene in *E. coli* isolates (Chia et al., 2009; Mammeri et al., 2010; Goessens et al., 2013; van Bortel et al., 2017). In 2012 Kurpiel et al. reported that point mutations in the *inc* antisense RNA gene can be associated with an increased plasmid copy number and subsequently higher expression of *bla*_{CMY-2} (Kurpiel and Hanson, 2012). The observed nucleotide substitution in the *inc* RNA gene of plasmid pG-268-2_CMY-2, however, might be associated with a change in the RNA folding of the *Inc* antisense RNA and hence led to an increased copy number of this plasmid due to inhibited pseudoknot formation (**Supplementary Figure 1**). Additional studies will be necessary to check this hypothesis *in-situ*.

Regarding the outer membrane composition, a loss of OmpC was found in isolate G-268-2, while the amount of mutations within the *ompF* gene sequence in the same way suggested a malfunctioned outer membrane protein F.

However, as mentioned previously, the phenotypic carbapenem resistance of G-268-2 (MIC_{IMI} = ≥ 16 mg/L; MIC_{MEM} = 8 mg/L) was transferrable to the *E. coli* recipient strain NEB10[®]-beta (MIC_{IMI} = 8 mg/L; MIC_{MEM} = 4 mg/L) and the *ompF* as well as *ompC* sequence data of the transformant strain (T_{G-268-2}) showed 100% accordance to the reference sequences of *E. coli* K12. Furthermore, the G-268-2 wildtype and its transformant have shown the ability to hydrolyze imipenem in the Blue-Carba assay. Due to the fact that on the one hand site T_{G-268-2} did not possess *ompC* as well as *ompF* mutations and in addition the results of the Blue-Carba assay showed its ability to hydrolyze imipenem, the final explanation for the detected carbapenem resistance of the *E. coli* isolate G-268-2 remains unclear. Further analyses addressing the question if an overexpression of CMY-2 or the occurrence of a new carbapenemase gene might have contributed this observation have to be checked in the future.

Taken together, the knowledge that carbapenem-resistant *Salmonella* as well as *E. coli* isolates can be found within German livestock-farms is alarming and it becomes even worse when these isolates acquired plasmids containing the colistin resistance gene *mcr-1*. Once more, it depicts the importance of comprehensive as well as harmonized monitoring programs in Germany as well as abroad. Beyond this, the implementation of proper intervention strategies to prevent a further dissemination of multidrug-resistant bacteria as well as the spread of their mobile genetic elements within and between animals and humans are urgently needed.

AUTHOR CONTRIBUTIONS

UR and LK: performed the design of the initial studies; NR, JF, BG, YP: designed the experiments; NR, JF: performed the laboratory work; LF, TC, JF, MP: performed whole genome sequencing including the subsequent data analyses; SG: performed MIC determination; BG, YP: provided scientific support regarding the data evaluation; NR: evaluated the final data and wrote the manuscript. All authors have read and approved the final draft of the article.

FUNDING

This work was partly funded by German Federal Ministry of Education and Research [funding number 01KI1013C/01KI1313C (to UR), 01KI1013B/01KI1313B (to BG), 01KI1013A (to LK), 01KI1013G/01KI1313G (to TC) as well

as 01KI1313F (to MP and YP)] as part of the research consortium RESET (<http://www.reset-verbund.de/>).

ACKNOWLEDGMENTS

We would like to thank the technicians Maja Thieck, Heike Rose, Diana Steinke, and Christina Gerstmann for excellent technical assistance. Special thanks to Henriette Laube and Johanna Hering for sampling work within the farms. The team of the NRL *Salmonella* is gratefully acknowledged for its help regarding the serotyping of the *Salmonella* isolates. We deeply thank Beatrice Baumann for excellent laboratory assistance in sequencing of isolates and Carlus Deneke for his support processing WGS data.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmicb.2018.00538/full#supplementary-material>

Supplementary Figure 1 | (A) Sequence alignment of the incRNA1 of IncB (I), IncK2-pDV45 (II) and pG-268-2 (III). The IncB incRNA1 reference sequence derived from Siemerling et al. (1993), the IncK2-pDV45 incRNA1 reference sequence from Seiffert et al. (2017). **(B)** Predicted RNA folding of the incRNA1 of IncB (I), IncK2-pDV45 (II) and pG-268-2 (III) by RNAfold web server.

Supplementary Figure 2 | Results of the Blue-Carba assay obtained from *E. coli* isolate G-268-2. Beside the wildtype strain also its transformants containing the IncB/O/K/Z-plasmid (*bla_{CMY-2}*) and the IncX4-plasmid (*mcr-1*) were tested. As positive control (PC) the *bla_{VIM-1}* containing isolate R178 was used. NTC, no template control.

REFERENCES

- Abdallah, H. M., Reuland, E. A., Wintermans, B. B., Al Naiemi, N., Koek, A., Abdelwahab, A. M., et al. (2015). Extended-spectrum beta-lactamases and/or carbapenemase-producing enterobacteriaceae isolated from retail chicken meat in zagazig, Egypt. *PLoS ONE* 10:e0136052. doi: 10.1371/journal.pone.0136052
- Alikhan, N. F., Petty, N. K., Ben Zakour, N. L., and Beatson, S. A. (2011). BLAST Ring Image Generator (BRIG): simple prokaryote genome comparisons. *BMC Genomics* 12:402. doi: 10.1186/1471-2164-12-402
- Bankevich, A., Nurk, S., Antipov, D., Gurevich, A. A., Dvorkin, M., Kulikov, A. S., et al. (2012). SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J. Comput. Biol.* 19, 455–477. doi: 10.1089/cmb.2012.0021
- Bolger, A. M., Lohse, M., and Usadel, B. (2014). Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30, 2114–2120. doi: 10.1093/bioinformatics/btu170
- Borowiak, M., Szabo, I., Baumann, B., Junker, E., Hammerl, J. A., Kaesbohrer, A., et al. (2017). VIM-1-producing *Salmonella* Infantis isolated from swine and minced pork meat in Germany. *J. Antimicrob. Chemother.* 72, 2131–2133. doi: 10.1093/jac/dkx101
- Carattoli, A. (2013). Plasmids and the spread of resistance. *Int. J. Med. Microbiol.* 303, 298–304. doi: 10.1016/j.ijmm.2013.02.001
- Carattoli, A., Zankari, E., Garcia-Fernandez, A., Larsen, M. V., Lund, O., Villa, L., et al. (2014). *In silico* detection and typing of plasmids using plasmidfinder and plasmid multilocus sequence typing. *Antimicrobial Agents Chemother.* 58, 3895–3903. doi: 10.1128/AAC.02412-14
- Chia, J. H., Siu, L. K., Su, L. H., Lin, H. S., Kuo, A. J., Lee, M. H., et al. (2009). Emergence of carbapenem-resistant *Escherichia coli* in Taiwan: resistance due to combined CMY-2 production and porin deficiency. *J. Chemother.* 21, 621–626. doi: 10.1179/joc.2009.21.6.621
- Clermont, O., Christenson, J. K., Denamur, E., and Gordon, D. M. (2013). The Clermont *Escherichia coli* phylo-typing method revisited: improvement of specificity and detection of new phylo-groups. *Environ. Microbiol. Rep.* 5, 58–65. doi: 10.1111/1758-2229.12019
- Coil, D., Jospin, G., and Darling, A. E. (2015). A5-miseq: an updated pipeline to assemble microbial genomes from Illumina MiSeq data. *Bioinformatics* 31, 587–589. doi: 10.1093/bioinformatics/btu661
- Ewers, C., Klotz, P., Leidner, U., Stamm, I., Prenger-Berninghoff, E., Gottig, S., et al. (2017). OXA-23 and ISAb1-OXA-66 class D beta-lactamases in *Acinetobacter baumannii* isolates from companion animals. *Int. J. Antimicrob. Agents* 49, 37–44. doi: 10.1016/j.ijantimicag.2016.09.033
- Falgenhauer, L., Ghosh, H., Guerra, B., Yao, Y., Fritzenwanker, M., Fischer, J., et al. (2017). Comparative genome analysis of IncHI2 VIM-1 carbapenemase-encoding plasmids of *Escherichia coli* and *Salmonella enterica* isolated from a livestock farm in Germany. *Vet. Microbiol.* 200, 114–117. doi: 10.1016/j.vetmic.2015.09.001
- Fischer, J., Rodriguez, I., Schmogger, S., Friese, A., Roesler, U., Helmuth, R., et al. (2013a). *Salmonella enterica* subsp. enterica producing VIM-1 carbapenemase isolated from livestock farms. *J. Antimicrob. Chemother.* 68, 478–480. doi: 10.1093/jac/dks393

- Fischer, J., San Jose, M., Roschanski, N., Schmogger, S., Baumann, B., Irrgang, A., et al. (2017). Spread and persistence of VIM-1 Carbapenemase-producing Enterobacteriaceae in three German swine farms in 2011 and 2012. *Vet. Microbiol.* 200, 118–123. doi: 10.1016/j.vetmic.2016.04.026
- Fischer, J., Schmogger, S., Jahn, S., Helmuth, R., and Guerra, B. (2013b). NDM-1 carbapenemase-producing *Salmonella enterica* subsp. *enterica* serovar Corvallis isolated from a wild bird in Germany. *J. Antimicrob. Chemother.* 68, 2954–2956. doi: 10.1093/jac/dkt260
- Goessens, W. H. F., van der Bij, A. K., van Boxtel, R., Pitout, J. D. D., van Ulsen, P., Melles, D. C., et al. (2013). Antibiotic trapping by plasmid-encoded CMY-2 beta-lactamase combined with reduced outer membrane permeability as a mechanism of carbapenem resistance in *Escherichia coli*. *Antimicrob. Agents Chemother.* 57, 3941–3949. doi: 10.1128/AAC.02459-12
- Gruber, A. R., Lorenz, R., Bernhart, S. H., Neubock, R., and Hofacker, I. L. (2008). The Vienna RNA website. *Nucleic Acids Res.* 36, W70–W74. doi: 10.1093/nar/gkn188
- Guerra, B., Junker, E., Miko, A., Helmuth, R., and Mendoza, M. C. (2004). Characterization and localization of drug resistance determinants in multidrug-resistant, integron-carrying *Salmonella enterica* serotype Typhimurium strains. *Microb. Drug Resist.* 10, 83–91. doi: 10.1089/1076629041310136
- Hamza, E., Dorgham, S. M., and Hamza, D. A. (2016). Carbapenemase-producing *Klebsiella pneumoniae* in broiler poultry farming in Egypt. *J. Glob. Antimicrob. Resist.* 7, 8–10. doi: 10.1016/j.jgar.2016.06.004
- He, T., Wang, Y., Sun, L., Pang, M., Zhang, L., and Wang, R. (2017). Occurrence and characterization of blaNDM-5-positive *Klebsiella pneumoniae* isolates from dairy cows in Jiangsu, China. *J. Antimicrob. Chemother.* 72, 90–94. doi: 10.1093/jac/dkw357
- Hering, J., Fromke, C., von Munchhausen, C., Hartmann, M., Schneider, B., Friese, A., et al. (2016). Cefotaxime-resistant *Escherichia coli* in broiler farms—A cross-sectional investigation in Germany. *Prev. Vet. Med.* 125, 154–157. doi: 10.1016/j.prevetmed.2016.01.003
- Irrgang, A., Fischer, J., Grobbel, M., Schmogger, S., Skladnikiewicz-Ziemer, T., Thomas, K., et al. (2017). Recurrent detection of VIM-1-producing *Escherichia coli* clone in German pig production. *J. Antimicrob. Chemother.* 72, 944–946. doi: 10.1093/jac/dkw479
- Jia, B., Raphenya, A. R., Alcock, B., Waglechner, N., Guo, P., Tsang, K. K., et al. (2017). CARD 2017: expansion and model-centric curation of the comprehensive antibiotic resistance database. *Nucleic Acids Res.* 45, D566–D573. doi: 10.1093/nar/gkw1004
- Kaas, R. S., Leekitcharoenphon, P., Aarestrup, F. M., and Lund, O. (2014). Solving the problem of comparing whole bacterial genomes across different sequencing platforms. *PLoS ONE* 9:e104984. doi: 10.1371/journal.pone.0104984
- Kurpiel, P. M., and Hanson, N. D. (2012). Point mutations in the inc antisense RNA gene are associated with increased plasmid copy number, expression of blaCMY-2 and resistance to piperacillin/tazobactam in *Escherichia coli*. *J. Antimicrob. Chemother.* 67, 339–345. doi: 10.1093/jac/dkr479
- Larsen, M. V., Cosentino, S., Rasmussen, S., Friis, C., Hasman, H., Marvig, R. L., et al. (2012). Multilocus sequence typing of total-genome-sequenced bacteria. *J. Clin. Microbiol.* 50, 1355–1361. doi: 10.1128/JCM.06094-11
- Lartigue, M. F., Poirel, L., Poyart, C., Reglier-Poupet, H., and Nordmann, P. (2007). Ertapenem resistance of *Escherichia coli*. *Emerg. Infect. Dis.* 13, 315–317. doi: 10.3201/eid1302.060747
- Laube, H., Friese, A., von Salviati, C., Guerra, B., Kasbohrer, A., Kreienbrock, L., et al. (2013). Longitudinal monitoring of Esbl/Ampc-Producing *Escherichia coli* in German broiler chicken fattening farms. *Appl. Environ. Microbiol.* 79, 4815–4820. doi: 10.1128/AEM.00856-13
- Mammeri, H., Guillon, H., Eb, F., and Nordmann, P. (2010). Phenotypic and biochemical comparison of the carbapenem-hydrolyzing activities of five plasmid-borne AmpC beta-lactamases. *Antimicrob. Agents Chemother.* 54, 4556–4560. doi: 10.1128/AAC.01762-09
- Mollenkopf, D. F., Stull, J. W., Mathys, D. A., Bowman, A. S., Feicht, S. M., Grooters, S. V., et al. (2016). Carbapenemase-producing Enterobacteriaceae recovered from the environment of a swine farrow-to-finish operation in the United States. *Antimicrob. Agents Chemother.* 61:aac.02348-16. doi: 10.1128/AAC.01298-16
- Morrison, B. J., and Rubin, J. E. (2015). Carbapenemase producing bacteria in the food supply escaping detection. *PLoS ONE* 10:e0126717. doi: 10.1371/journal.pone.0126717
- Nordmann, P., Dortet, L., and Poirel, L. (2012). Carbapenem resistance in Enterobacteriaceae: here is the storm! *Trends Mol. Med.* 18, 263–272. doi: 10.1016/j.molmed.2012.03.003
- Olasz, F., Nagy, T., Szabo, M., Kiss, J., Szmolka, A., Barta, E., et al. (2015). Genome sequences of three *Salmonella enterica* subsp. *enterica* Serovar Infantis strains from healthy broiler chicks in Hungary and in the United Kingdom. *Genome announc.* 3:e01468-14. doi: 10.1128/genomeA.01468-14
- Oni, R. A., Sharma, M., and Buchanan, R. L. (2015). Survival of *Salmonella enterica* in dried Turkey manure and persistence on spinach leaves. *J. Food Prot.* 78, 1791–1799. doi: 10.4315/0362-028X.JFP-15-047
- Grimont, P. A. D., and Weill, F.-X. (2007). *Antigenic Formulae of the Salmonella Serovars WHO Collaborating Centre for Reference and Research on Salmonella, 9th Edn.* Paris: Institut Pasteur.
- Pires, J., Novais, A., and Peixe, L. (2013). Blue-carba, an easy biochemical test for detection of diverse carbapenemase producers directly from bacterial cultures. *J. Clin. Microbiol.* 51, 4281–4283. doi: 10.1128/JCM.01634-13
- Pruthivishree, B. S., Vinodh Kumar, O. R., Sinha, D. K., Malik, Y. P. S., Dubal, Z. B., Desingu, P. A., et al. (2017). Spatial molecular epidemiology of carbapenem-resistant and New Delhi metallo beta-lactamase (blaNDM)-producing *Escherichia coli* in the piglets of organized farms in India. *J. Appl. Microbiol.* 122, 1537–1546. doi: 10.1111/jam.13455
- Ribot, E. M., Fair, M. A., Gautom, R., Cameron, D. N., Hunter, S. B., Swaminathan, B., et al. (2006). Standardization of pulsed-field gel electrophoresis protocols for the subtyping of *Escherichia coli* O157:H7, *Salmonella*, and *Shigella* for PulseNet. *Foodborne Pathog. Dis.* 3, 59–67. doi: 10.1089/fpd.2006.3.59
- Roschanski, N., Friese, A., C., von Salviati-Claudius, Hering, J., Kasbohrer, A., Kreienbrock, L., et al. (2017a). Prevalence of carbapenemase producing Enterobacteriaceae isolated from German pig-fattening farms during the years 2011–2013. *Vet. Microbiol.* 200, 124–129. doi: 10.1016/j.vetmic.2015.11.030
- Roschanski, N., Friese, A., Thieck, M., and Roesler, U. (2016). Follow-up investigation of the first VIM-1 positive pig farm in Germany - How is the situation 4 years after the first detection? *Clin. Microbiol. Infect.* 22, 951–953. doi: 10.1016/j.cmi.2016.08.018
- Roschanski, N., Guenther, S., Vu, T. T. T., Fischer, J., Semmler, T., Roesler, U., et al. (2017b). VIM-1 carbapenemase-producing *Escherichia coli* isolated from retail seafood, Germany 2016. *Euro Surveill.* 22, 17:32. doi: 10.2807/1560-7917.ES.2017.22.43.17-00032
- Rubin, J. E., Ekanayake, S., and Fernando, C. (2014). Carbapenemase-producing organism in food, 2014. *Emerg. Infect. Dis.* 20, 1264–1265. doi: 10.3201/eid2007.140534
- Seiffert, S. N., Carattoli, A., Schwendener, S., Collaud, A., Endimiani, A., and Perreten, V. (2017). Plasmids carrying blaCMY-2/4 in *Escherichia coli* from poultry, poultry meat, and humans belong to a novel IncK subgroup designated IncK2. *Front. Microbiol.* 8:407. doi: 10.3389/fmicb.2017.00407
- Siemering, K. R., Praszker, J., and Pittard, A. J. (1993). Interaction between the antisense and target RNAs involved in the regulation of IncB plasmid replication. *J. Bacteriol.* 175, 2895–2906.
- Stolle, I., Prenger-Berninghoff, E., Stamm, I., Scheufen, S., Hassdenteufel, E., Guenther, S., et al. (2013). Emergence of OXA-48 carbapenemase-producing *Escherichia coli* and *Klebsiella pneumoniae* in dogs. *J. Antimicrob. Chemother.* 68, 2802–2808. doi: 10.1093/jac/dkt259
- van Boxtel, R., Wattel, A. A., Arenas, J., Goessens, W. H., and Tommassen, J. (2017). Acquisition of carbapenem resistance by Plasmid-Encoded-AmpC-Expressing *Escherichia coli*. *Antimicrob. Agents Chemother.* 61:e01413-16. doi: 10.1128/AAC.01413-16
- Walsh, T. R., Weeks, J., Livermore, D. M., and Toleman, M. A. (2011). Dissemination of NDM-1 positive bacteria in the New Delhi environment and its implications for human health: an environmental point prevalence study. *Lancet Infect. Dis.* 11, 355–362. doi: 10.1016/S1473-3099(11)70059-7
- Wang, Y., Zhang, R., Li, J., Wu, Z., Yin, W., Schwarz, S., et al. (2017). Comprehensive resistome analysis reveals the prevalence of NDM and MCR-1 in Chinese poultry production. *Nat. microbiol.* 2:16260. doi: 10.1038/nmicrobiol.2016.260

- Wilson, A. P. R. (2017). Sparing carbapenem usage. *J. Antimicrob. Chemother.* 72, 2410–2417 doi: 10.1093/jac/dkx181
- Zankari, E., Hasman, H., Cosentino, S., Vestergaard, M., Rasmussen, S., Lund, O., et al. (2012). Identification of acquired antimicrobial resistance genes. *J. Antimicrob. Chemother.* 67, 2640–2644. doi: 10.1093/jac/dks261
- van der Zee, A., Roorda, L., Bosman, G., Fluit, A. C., Hermans, M., Smits, P. H., et al. (2014). Multi-centre evaluation of real-time multiplex PCR for detection of carbapenemase genes OXA-48, VIM, IMP, NDM and KPC. *BMC Infect. Dis.* 14:27. doi: 10.1186/1471-2334-14-27
- Zhang, R., Liu, Z., Li, J., Lei, L., Yin, W., Li, M., et al. (2017). Presence of VIM-Positive pseudomonas species in chickens and their surrounding environment. *Antimicrob. Agents Chemother.* 61:e00167-17. doi: 10.1128/AAC.00167-17
- Zurfluh, K., Hachler, H., Nuesch-Inderbinen, M., and Stephan, R. (2013). Characteristics of extended-spectrum beta-lactamase- and carbapenemase-producing Enterobacteriaceae isolates from rivers and lakes in Switzerland. *Appl. Environ. Microbiol.* 79, 3021–3026. doi: 10.1128/AEM.00054-13

Disclaimer: BG is employed with the European Food Safety Authority (EFSA) in its BIOCONTAM Unit that provides scientific and administrative support to EFSA's scientific activities. The positions and opinions presented in this article are those of the authors alone and are not intended to represent the views or scientific works of EFSA.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Roschanski, Fischer, Falgenhauer, Pietsch, Guenther, Kreienbrock, Chakraborty, Pfeifer, Guerra and Roesler. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.