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# Cytomegaloviruses inhibit Bak- and Bax-mediated apoptosis with two separate viral proteins

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*Apoptosis of infected cells can limit virus replication and serves as an innate defense mechanism against viral infections. Consequently, viruses delay apoptosis by expressing antiapoptotic proteins, many of which structurally resemble the cellular antiapoptotic protein Bcl-2. Like Bcl-2, the viral analogs inhibit apoptosis by preventing activation and/or oligomerization of the proapoptotic mitochondrial proteins Bax and Bak. Here we show that cytomegaloviruses (CMVs) have adopted a different strategy. They encode two separate mitochondrial proteins that lack obvious sequence similarities to Bcl-2-family proteins and specifically counteract either Bax or Bak. We identified a small mitochondrion-localized protein encoded by the murine CMV open reading frame (ORF) m41.1, which functions as a viral inhibitor of Bak oligomerization (vIBO). It blocks Bak-mediated cytochrome c release and Bak-dependent induction of apoptosis. It protects cells from cell death-inducing stimuli together with the previously identified Bax-specific inhibitor viral mitochondria-localized inhibitor of apoptosis (vMIA) (encoded by ORF m38.5). Similar vIBO proteins are encoded by CMVs of rats, and possibly by other CMVs as well. These results suggest a non-redundant function of Bax and Bak during viral infection, and a benefit for CMVs derived from the ability to inhibit Bak and Bax separately with two viral proteins.*

Programmed cell death (PCD) or apoptosis has numerous functions in a multicellular organism. It is important during development, for maintenance of tissue homeostasis, for elimination of damaged or transformed cells, and as a first line of defense against infectious agents. The cellular suicide program can be particularly effective as an antiviral defense mechanism, since viruses depend on the host cell for replication.<sup>1</sup> Viral dissemination can be severely impaired if an infected cell executes apoptosis before the viral replication cycle is completed. The importance of apoptosis is underscored by the fact that many viruses have evolved genes encoding antiapoptotic proteins.<sup>2</sup> Viral infection and replication exert different kinds of stress on the host cell: depletion of nutrients, impairment of cellular protein biosynthesis, induction of the unfolded protein response (also called endoplasmic reticulum stress), and a DNA-damage response. All kinds of stress can initiate signaling pathways culminating in apoptosis.<sup>2</sup> Caspases, a group of aspartate-specific cysteine proteases, are pivotal signal transducers within apoptotic signaling cascades, whose activation is regulated by proteins of the Bcl-2 family.<sup>3</sup> This family consists of pro- and antiapoptotic members, which differ in the number of Bcl-2 homology (BH) domains. According to the current model, antiapoptotic Bcl-2-family proteins prevent caspase activation by preserving mitochondrial integrity.<sup>3</sup> The activity of the antiapoptotic Bcl-2 proteins is antagonized by the proapoptotic multidomain proteins Bax and Bak, and the so-called BH3-only proteins, which possess only the third out of four BH domains. In response to apoptotic stimuli, the BH3-only proteins translocate to mitochondria where they counteract the function of pro-survival Bcl-2 proteins and activate the proapoptotic proteins Bax and Bak.<sup>4</sup> The monomeric proteins undergo a conformational change, oligomerize, and increase the permeability of the mitochondrial outer membrane.<sup>5</sup> This leads to release of cytochrome c (cyt c) into the cytosol and subsequent activation of caspases. The release of other mitochondrial proteins such as apoptosis-inducing factor (AIF), Smac/DIABLO, Htr2A/Omi, and endonuclease-G also contributes to the execution of PCD.<sup>4</sup> Earlier studies have shown that Bak and Bax function in a largely redundant manner during development<sup>6</sup> and in response to various apoptosis inducers.<sup>5</sup> However, more recent results indicated that the two proteins also fulfill some non-redundant functions.<sup>7–10</sup> A number of viruses, particularly adenoviruses and  $\gamma$ -herpesviruses, express proteins homologous to the cellular antiapoptotic proteins Bcl-2 and Bcl-xL.<sup>2,11</sup> Poxviruses also express apoptosis inhibitors that resemble Bcl-xL in their three-dimensional structure, even though their amino-acid sequences are not homologous.<sup>12,13</sup> Like Bcl-xL, these viral

proteins inhibit both Bax- and Bak-mediated cell death. A different strategy to inhibit apoptosis at the mitochondrial checkpoint is pursued by cytomegaloviruses (CMVs). Human CMV encodes a viral mitochondrion-localized inhibitor of apoptosis (vMIA, encoded by open reading frame (ORF) UL37x1) that interacts with Bax and specifically inhibits Bax-mediated cell death.<sup>14–16</sup> The primary structure of human CMV (HCMV) vMIA has no obvious similarities to Bcl-2-family proteins,<sup>14</sup> but an *in silico* structural analysis predicted a fold similar to Bcl-xL.<sup>17</sup> Murine CMV (MCMV) also expresses a vMIA protein, which is encoded at an analogous position within the viral genome by ORF m38.5, but displays little sequence similarity to HCMV vMIA.<sup>18,19</sup> The MCMV m38.5 protein inhibits Bax- but not Bak-mediated apoptosis,<sup>20–22</sup> and also induces mitochondrial fragmentation in the absence of Bak (i.e., in bak<sup>-/-</sup> cells).<sup>23</sup> Moreover, the observation that MCMV-infected cells are protected from Bax- and Bak-mediated apoptosis suggested that the virus encodes an additional, Bak-specific antiapoptotic protein.<sup>20</sup> In the present study, we identified the product of MCMV ORF m41.1 as a Bak-specific inhibitor of apoptosis. The m41.1 protein localizes to mitochondria and prevents Bak oligomerization, cytochrome c release, and execution of PCD. In conjunction with the m38.5/vMIA protein, it protects infected cells from apoptosis induced by cytotoxic drugs or the viral infection itself. Moreover, structurally and functionally similar proteins are encoded by rat CMVs. The unusual ability of CMVs to block Bax and Bak with two separate proteins suggests that it might be advantageous for these viruses to modulate Bax- and Bak-dependent pathways separately. The identification of these two viral inhibitors should provide new insights into the differential importance of the two proapoptotic Bcl-2 family members in viral infection-induced cell death.

## Results

**Identification of the protein expressed by m41.1.** We identified ORF m41 as a region of the MCMV genome encoding an antiapoptotic protein in a previous study.<sup>24</sup> Deletion of the m41 ORF from the MCMV genome resulted in premature death of infected cells that could be reduced by addition of broad-spectrum caspase inhibitors. The m41 protein was shown to localize to the Golgi apparatus, but its mechanism of action has remained undefined. A recent computational re-annotation of the MCMV genome revealed the presence of a second ORF embedded within m41 in a different reading frame.<sup>18</sup> This ORF was named m41.1 and has the potential to code for a 57-amino-acid protein (Figure 1a and c). The putative start codon is located 10 nucleotides (nt) downstream of the m41 translational start. A predominant transcript of approximately 0.6 kb was identified by Northern blot analysis (Figure 1b). The 5' and 3' ends of the mRNA transcripts were determined by rapid amplification of cDNA ends (RACE). The major transcript starts at nt position 54 227 of the MCMV genome and terminates at position 53 680, 93-nt downstream of the m41 ORF and 21-nt downstream of a canonical AATAAA polyadenylation signal. Several larger transcripts of low abundance (Figure 1b) were also cloned and sequenced. All of them used a splice acceptor site at position 54 217 (Figure 1a), but none of them had the capacity to encode an N-terminally extended m41 or m41.1 protein (data not shown). Hence we concluded that the m41 and m41.1 gene products must be translated from the same 0.6-kb mRNA transcript by using the first or one of the following start codons.

We have previously shown that the m41 ORF is expressed during MCMV infection and translated into a protein of 138 amino acids with an apparent molecular mass of approximately 20 kDa.<sup>24</sup> To determine whether an m41.1 protein is also synthesized during MCMV infection, we inserted an HA epitope tag sequence and a selectable marker (kan) at the 3' end of the m41.1 ORF (Figure 1c). This procedure leaves the transcriptional and translational start sites of m41.1 unchanged, and is, therefore, unlikely to affect the expression pattern of m41.1. We detected the HA-tagged m41.1 protein in lysates of infected 10.1 fibroblasts by Western blotting starting about 5 hpi (Figure 1d). Further experiments showed that m41.1 was expressed in the presence of a DNA-synthesis inhibitor, but not after release from the cytoheximide block (Supplementary Figure S1A). Hence m41.1 can be classified as an early gene product. The protein had an apparent molecular mass of approximately 7 kDa (not shown), which corresponds to its predicted mass. **Functional dissection of the m41 locus.** In order to determine whether m41, m41.1, or both gene products are responsible for the previously described antiapoptotic activity, we constructed mutant viruses lacking m41.1, m41, or both ORFs (Figure 1c). In the Dm41 mutant, the m41 ORF was replaced with a zeocin-resistance gene. To construct an m41.1-knockout mutant (m41.1ko), we introduced three silent mutations into the m41 ORF, which eliminated the three potential ATG start codons of m41.1 (Figure 1a). The m41ko virus was constructed by inserting the m41.1 ORF at an ectopic position into the genome of the Dm41 mutant. In a similar way, two genes encoding well-characterized mitochondrial apoptosis inhibitors were inserted: the cellular Bcl-xL and the myxoma virus Bcl-2 analog M11L.<sup>12,25</sup> The integrity of the constructed mutants was determined by restriction-fragment length analysis and by immunoblotting (Supplementary Figure S1B–D). As our previous work has shown that the Dm41 mutant induced

apoptosis of infected cells,<sup>24</sup> we first analyzed how the newly constructed mutants affected the viability of infected cells. Murine 10.1 fibroblasts were infected with the mutant viruses at a high multiplicity of infection (MOI), and cell viability was determined using an MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay. Fibroblasts infected with Dm41 or m41.1ko showed clearly reduced viability at 72 hpi (Figure 2a). By contrast, the viability of cells infected with m41ko or Dm41/M11L was similar to those infected with the wild-type (wt) virus, suggesting that the m41 protein is not required to inhibit premature PCD in fibroblasts. We also infected RAW264.7 macrophages, which are more sensitive to virus-induced PCD. At 18 hpi their viability was reduced after infection with wt MCMV and even more with Dm41, m41ko, or m41.1ko (Figure 2b and Supplementary Figure S2). Similar results were obtained with IC-21 macrophages (not shown). We also determined nuclear DNA fragmentation as a sign of apoptosis in infected fibroblasts and macrophages using a terminal deoxynucleotidyltransferase-mediated dUTP-biotin nick-end-labeling (TUNEL) assay (Figure 2c and d). However, this assay was difficult to employ because infected cells do not die synchronously, and dead cells eventually disintegrate and detach, making them unavailable for analysis. Even though the TUNEL assay considerably underestimates the true percentage of dead cells, the results were basically concordant with the results of the cell viability assay (Figure 2a and b). A strong activation of caspase-3 was detected in fibroblasts and macrophages infected with the Dm41 or the m41.1ko mutant (Figure 2e and f). By contrast, caspase-3 activation was largely suppressed in cells infected with m41.1- or M11L-expressing MCMVs. Taken together these results suggested that m41.1 is a potent inhibitor of infection-induced apoptosis. Under the conditions tested here, m41 was only a weak cell death suppressor, whose activity became apparent primarily in macrophages. We then infected fibroblasts and macrophages with the different viruses and analyzed viral replication after infection with low MOI. Multistep replication kinetics in 10.1 fibroblasts and RAW264.7 macrophages indicated that viral replication in fibroblasts is largely unaffected by the absence of m41 and/or m41.1 (Figure 2g), but is severely impaired in macrophages in the absence of m41.1 (Figure 2h). The m41.1 protein localizes to mitochondria and protects infected cells from drug-induced apoptosis. The observation that the loss of m41.1 could be compensated by inserting the Bcl-xL or the M11L gene into the MCMV genome (Figure 2) suggested that m41.1 might function in a similar way and inhibit apoptosis at the mitochondrial checkpoint. To determine the localization of the m41.1 protein, we transfected cells with a plasmid expressing m41.1 with a C-terminal HA tag and analyzed its localization by immunofluorescence. As shown in Figure 3a, the m41.1 protein colocalized with the mitochondrial marker Hsp60 and with the mitochondrion-specific dye, MitoTracker, but did not colocalize with an endoplasmic reticulum (ER) marker. The m41.1 protein also displayed mitochondrial localization in MCMV-infected cells (Figure 3b). Next we tested whether m41.1 protects infected cells from exogenous proapoptotic stimuli. To this end, fibroblasts were infected with wt and mutant MCMVs, and treated with the drugs staurosporine (STS) or actinomycin-D (ActD), both of which activate the mitochondrial apoptosis pathway.<sup>5</sup> Indeed, deletion of m41.1 sensitized infected cells to STS and ActD (Figure 4a). We also treated infected cells with STS and tested whether m41.1 was required to inhibit mitochondrial cytochrome c release. The results in Figure 4b show that cytochrome c release was inhibited in the presence of m41.1. The lack of m41.1 was compensated by expression of Bcl-xL or M11L. m41.1 specifically inhibits Bak-mediated cell death and synergizes with m38.5. Recently, we and others have shown that MCMV expresses a mitochondrial protein that specifically blocks Bax-mediated cell death.<sup>20,21,23</sup> We further showed that Bax-knockout cells infected with an m38.5-deficient MCMV mutant (Dm38.5) were resistant to STS-induced apoptosis, suggesting that Bak-mediated apoptosis was inhibited by a different MCMV protein.<sup>20</sup> To test whether m41.1 inhibits apoptosis in a Bak-specific manner, we infected fibroblasts deficient for Bax, Bak, or both proteins with different MCMV mutants and analyzed their sensitivity to STS. For comparison, we also included the previously described Dm38.5 mutant and constructed a double-knockout (dko) mutant lacking both m41.1 and m38.5. The results shown in Figure 5a clearly demonstrated that m41.1 was required to inhibit Bak-mediated cell death, whereas m38.5 was required to block Bax-mediated cell death. In the absence of both m41.1 and m38.5, infected cells were sensitive to Bax- and Bak-mediated apoptosis. Similar results were obtained without STS treatment, when the infection was allowed to proceed for a longer period of time (Figure 5b). This indicated that the viral infection itself leads to an induction of Bak- and Bax-mediated cell death. The use of viral deletion mutants allowed us to determine that m41.1 is required for inhibition of Bak-dependent apoptosis. However, it was not clear from these results whether m41.1 by itself is sufficient to block Bak-dependent apoptosis. Therefore, we expressed m41.1 in Bakp/Bax<sup>-</sup> and Bak<sup>-</sup>/Baxp fibroblasts by retroviral transduction and analyzed the sensitivity of the cells to drug-induced apoptosis. Indeed, m41.1 provided protection against STS- or ActD-induced apoptosis in Bak-only but not in Bax-only cells (Figure 5c–e). By contrast, Bcl-xL inhibited drug-induced apoptosis in both cell types, consistent with its known ability to inhibit Bak- and Bax-mediated cell death.<sup>26,27</sup> In NIH-3T3 cells (which express both Bax and Bak), transfection of an

m41.1 expression plasmid was not sufficient to inhibit apoptosis induced by Fas stimulation. Transfection of plasmids encoding Bax inhibitors of MCMV (m38.5) or HCMV (UL37x1) did also not protect from Fas-induced cell death. However, when m41.1 was coexpressed with m38.5 or UL37x1, Fas-induced apoptosis was inhibited to a similar extent as with Bcl-xL (Figure 5f). These results confirmed that m41.1 and m38.5 synergize to inhibit Bak- and Bax-mediated cell death. m41.1 interacts with Bak and inhibits Bak oligomerization. During activation of the mitochondrial apoptosis pathway, Bax and Bak undergo a conformational change leading to exposure of their N-terminus. In this open (activated) conformation, Bak and Bax molecules can oligomerize within the mitochondrial outer membrane.<sup>4</sup> This leads to increased mitochondrial membrane permeability and release of proapoptotic factors such as cyt c. As the previous experiments had shown that m41.1 blocks cyt c release, we wondered whether the preceding activation and oligomerization of Bak were also inhibited. Bak activation can be assessed by immunofluorescence analysis using an antibody directed against the N-terminus of Bak28 or against the Flag epitope if an N-terminally Flagtagged Bak is used. As shown in Figure 6a, MCMV infection induced Bak activation regardless of the presence or absence of m41.1. However, m41.1 itself is not responsible for Bak activation, because Bak is activated in cells infected with the m41.1ko virus, and m41.1 expression from a plasmid vector does not induce the active conformation. Hence, m41.1 does not prevent Bak activation, and neither does Bcl-xL (Figure 6a). It is known that Bcl-xL and Mcl-1 interact with Bak in its open (active) conformation,<sup>29</sup> whereas VDAC2 interacts with the closed conformation.<sup>30</sup> Thus, m41.1 might inhibit Bak in a similar way as Bcl-xL. In support of this hypothesis, we found that m41.1 colocalized with activated Flag-Bak in MCMV-infected cells (Figure 6b). To test whether m41.1 interacts with Bak, we transfected HEK 293 cells with plasmids encoding Flag-Bak and HAtagged m41.1 or m41, respectively. As shown in Figure 7a, m41.1 co-immunoprecipitated with Bak and vice versa, but m41 did not. This indicated a direct or indirect interaction of m41.1 and Bak. To determine the influence of m41.1 on Bak oligomerization, we infected fibroblasts expressing Bak (but not Bax) with wt and mutant MCMVs, stimulated the mitochondrial apoptosis pathway with STS, and then stabilized oligomeric complexes by adding the crosslinker 10,60-bismaleimidohexane (BMH). As shown in Figure 7b, Bak oligomerization was inhibited by MCMV only if the virus expressed m41.1 or Bcl-xL. Similarly, m41.1 inhibited Bak oligomerization in MCMV-infected cells expressing Bak and Bax (Supplementary Figure S3), suggesting that m41.1 inhibits both homo- and heterooligomerization of Bak. STS-induced Bak oligomerization was also inhibited by m41.1 or Bcl-xL alone in transduced cells (Figure 7c). Hence we concluded that the m41.1 protein functions as a viral inhibitor of Bak oligomerization (vIBO). Evolutionary conservation of m41.1. HCMV expresses a Bax-specific mitochondrial inhibitor of apoptosis (vMIA) that is conserved in primate CMVs.<sup>31</sup> Rodent CMVs also express vMIA proteins, but there is no apparent sequence similarity between primate and rodent vMIAs.<sup>19</sup> When we searched all available CMV sequences for the presence of putative m41.1 homologs, we identified ORFs encoding highly similar proteins in two rat CMVs, the Maastricht and the English isolates (Figure 8a). The ORFs are located at analogous positions within the viral genomes and were named r41.1 and e41.1, respectively. By contrast, we did not detect ORFs similar to m41.1 in human or other primate CMVs. To test whether sequence conservation correlated with functional conservation, we analyzed the function of the e41.1 protein of the rat CMV (RCMV) English isolate. Indeed, the e41.1 protein localized to mitochondria (Figure 8b) and inhibited ActD-induced apoptosis in Bakp/Bax\_ cells (Figure 8c), suggesting that the function of m41.1/vIBO is conserved among rodent CMVs.

## Discussion

In this study, we identified a new viral antiapoptotic protein that inhibits apoptosis induced by cytotoxic drugs or by the viral infection itself in a Bak-dependent manner. We further showed that this Bak-specific inhibitor synergizes with a previously identified Bax-specific inhibitor to protect infected cells from apoptosis-inducing stimuli. The m41.1 protein is an unusual apoptosis inhibitor as it differs in many ways from known viral Bcl-2-like proteins: (i) With 57 amino acids and a molecular mass of approximately 7 kDa, the m41.1 protein is much smaller than cellular and viral Bcl-2 analogs; (ii) the m41.1 protein shows no recognizable sequence similarity to known antiapoptotic proteins and lacks BH domains characteristic of the cellular and viral Bcl-2-family proteins; and (iii) it is the first viral protein that specifically inhibits Bak- but not Bax-mediated cell death. The small size of the m41.1 protein initially suggested that it could be an essential component of a larger viral protein complex acting at the mitochondrial checkpoint. Additional mitochondrial proteins of unknown function have recently been discovered in MCMV,<sup>32</sup> and these might be part of such a complex. However, our subsequent results showed that the m41.1 protein is sufficient for inhibiting Bak oligomerization

(Figure 7c) and Bak-mediated cell death (Figure 5c and d). Although we cannot exclude a role of the other MCMV mitochondrial proteins (e.g., for stabilizing or reinforcing the activity of m41.1 or m38.5), these proteins are clearly not essential for the functions of m41.1/vIBo and m38.5/vMIA. It might seem surprising that m41.1-negative MCMVs do not replicate in macrophages, although they grow with only minimal defects in fibroblasts (Figure 2g and h). The most likely reason for this is that macrophages are much more sensitive than fibroblasts to cell death induced by infection with m41.1-negative viruses. In macrophages, cell death occurs 15–24 h after infection with high MOI, but in fibroblasts only after 48–72 h (Figure 2a–d). As the replication cycle of MCMV takes approximately 24 h, the early onset of apoptosis in macrophages should massively impair virus progeny production. An increased sensitivity to infection-induced cell death and/or reduced viral replication in macrophages has also been observed with MCMVs lacking other cell-death suppressors such as m38.5,22 M36,26 or M45,33 suggesting that macrophages are generally more sensitive to infection-induced cell death. Although widely used, the MTT assay has occasionally been criticized for relying on mitochondrial respiration as a measure of cell viability. This point was worth considering, as it has been shown that HCMV stabilizes mitochondrial respiration.<sup>33</sup> Therefore, it is important to be sure that the MTT assay really measures the viability of MCMV-infected cells and does not merely reflect changes in mitochondrial respiration. We have obtained results very similar to the MTT assay results using a conventional Trypan blue exclusion assay (data not shown) and have also published similar results to those for the Dm41 mutant using a neutral red inclusion assay.<sup>24</sup> Moreover, the results of the MTT assay were consistent with the results of two apoptosis assays (TUNEL and caspase-3 activation; Figure 2). Thus we concluded that the MTT assay is an adequate means to quantify cell viability in this study. Mammalian cells express at least six antiapoptotic Bcl-2-family proteins: Bcl-2, Bcl-xL, Bcl-w, Bcl-B, Bfl-1/A1, and Mcl-1. Four of them interact with and inhibit both Bax- and Bak-, but Bcl-B predominantly blocks Bax-, and Mcl-1 predominantly blocks Bak-mediated apoptosis.<sup>27</sup> This finding has lent further support to the hypothesis that the numerous Bcl-2 proteins exist because they serve at least some nonoverlapping functions and can be regulated independently in different cells and tissues.<sup>11</sup> If correct, this hypothesis would imply that viral proteins with Bcl-2-like functions could also differentially modulate the proapoptotic proteins Bax and Bak. The present study demonstrates that this is being done by MCMV and related viruses. The evolutionary origin of m41.1/vIBo remains enigmatic. The protein shows no apparent sequence homology to any cellular gene and is, therefore, unlikely to be a gene captured from the host-cell genome, as it is assumed to be the case for the Bcl-2 homologs of other viruses. It is unclear why CMVs have evolved their own antiapoptotic proteins instead of capturing them. However, CMVs have also taken their own path for inhibition of caspase-8 (a.k.a. FLICE): The UL36 and M36 proteins of HCMV and MCMV inhibit caspase-8 activation,<sup>26,35</sup> but share no homology with the cellular FLICE inhibitory proteins (cFLIPs) and the vFLIPs found in g-herpesviruses and poxviruses.<sup>36</sup> Orthologs of m41.1/vIBo are encoded by two RCMVs (Figure 8), and these viruses also encode orthologs of the Bax antagonist m38.5.20 Hence, the principle of blocking Bax- and Bak-mediated cell death with two separate viral proteins seems to be conserved in these related viruses. HCMV expresses vMIA from ORF UL37x1,<sup>14</sup> which is located at a genomic position analogous to m38.5, but shows no obvious sequence similarity to the MCMV gene. Orthologs of HCMV vMIA were also found in a number of primate CMVs.<sup>31</sup> It has been shown that HCMV vMIA interacts with Bax and inhibits Bax- but not Bak-mediated cell death.<sup>15,16,21</sup> This suggests that human and primate CMVs might also express a yet to be identified Bak-specific inhibitor. Alternatively, it has been argued that Bax is dominant over Bak in human cells (but not in mouse cells), and that a Bax-specific inhibitor might suffice to prevent mitochondrial outer membrane permeabilization.<sup>15</sup> This would explain why vMIA blocks apoptosis efficiently in human cells expressing Bax and Bak. However, another study challenged the Bax specificity of HCMV vMIA by presenting evidence that vMIA also interacts with Bak.<sup>37</sup> Future studies will elucidate the similarities and differences in the way how primate and rodent CMVs inhibit Bax and Bak. Particularly the rodent CMVs provide an excellent model to study the role of Bax and Bak during viral infection and will help to reveal non-redundant functions of these two cellular apoptosis mediators.

## Materials and Methods

*Cells and viruses.* IC-21 (ATCC TIB-186) and RAW264.7 (ATCC TIB-71) macrophages were grown in RPMI 1640 or DMEM supplemented with 10% FCS, and 10mM HEPES. Bak<sub>+/+</sub> and Bax<sub>+/+</sub> mouse fibroblast cell lines<sup>29</sup> were provided by Georg Häcker (Technical University Munich, Germany), with permission from David Huang (WEHI, Melbourne, Australia). NIH-3T3 cells, 10.1 fibroblasts and HEK 293 cells were cultured as previously described.<sup>38</sup> Wt and recombinant MCMVs were propagated in 10.1 mouse fibroblasts as described,<sup>38</sup> and viral titers were determined using the TCID<sub>50</sub> (median tissue culture infective dose) method.

*Plasmids and retroviral vectors.* ORFs m41.1 and e41.1 were PCR amplified and cloned in pcDNA3 (Invitrogen, Karlsruhe, Germany) with and without a C-terminal HA tag. The murine bak gene was excised from pBabe-Bak-IRES-GFP39 (kindly provided by Wei-Xing Zong, SUNY, Stony Brook, NY, USA) and inserted into pcDNA3-Flag. The Flag-tagged bak gene was subsequently inserted into pMSCVpuro (Clontech). A codon-optimized version of the myxoma virus M11L gene with an N-terminal Flag tag was synthesized by GeneArt (Regensburg, Germany). The m41.1 coding sequence and FlagM11L were inserted into pReplacer essentially as described.<sup>38</sup> Retroviral vector plasmids pRetroGFP, pRetro-Bcl-XL, and pRetro-m38.5 have been described,<sup>20</sup> and pRetro-m41, pRetro-m41.1, and pRetroe41.1 were constructed analogously. To exclude m41.1 protein expression from the pRetro-m41 vector, the three potential m41.1 ATG start codons were changed to ACG. Retroviruses were generated by transfecting retroviral vector plasmids into the Phoenix Amphi packaging cell line as described.<sup>24</sup> Filtered supernatants were used for transduction of bak<sub>+/+</sub> or bax<sub>+/+</sub> fibroblasts. Transduced fibroblasts were maintained as bulk cultures without selection. Fibroblasts stably expressing Flagtagged Bak were made by transducing bax<sub>+/+</sub>/bak<sub>+/+</sub> or baxp<sub>+/+</sub>/bak<sub>+/+</sub> fibroblasts with MSCVpuro-Flag-Bak and selection with 4 mg/ml puromycin.

*Recombinant MCMVs.* All recombinant MCMVs are based on a GFPexpressing MCMV, except for MCMV-m41.1HA, which does not express GFP. The mutant viral genomes were constructed using the bacterial artificial chromosome (BAC) technology as described previously.<sup>24</sup> An HA epitope sequence and a kan gene flanked by FLP recognition target (FRT) sequences were inserted at the 30 end of the m41.1 ORF. MCMVDm41 has been described in reference.<sup>24</sup> The Dm41/m41.1, Dm41/Bcl-XL, and Dm41/M11L were obtained by inserting the indicated genes into the m02–m06 region of MCMVDm41 using the pReplacer-based recombination system.<sup>38</sup> An m41 coding sequence with the three potential ATG codons of m41.1 mutated to ACG was re-inserted into MCMVDm41 with an FRT–kan–FRT cassette for selection. MCMV-m41.1ko was obtained after removal of the kan gene with FLP recombinase. Recombinant MCMV BACs were analyzed by restriction digest and by sequencing of the mutated site(s) (data not shown). BACs were transfected into 10.1 fibroblasts to reconstitute recombinant viruses. MCMV virion DNA was prepared essentially as described.<sup>40</sup>

*RNA analyses.* For Northern blot analysis, total cellular RNA was isolated with Trizol reagent (Invitrogen) and further purified with an RNeasy kit (Qiagen, Hilden, Germany) and subsequent DNase-I (Roche, Mannheim, Germany) digestion. A 1-mg weight of total RNA per sample was denatured, separated on a 1.2% denaturing agarose gel, and blotted onto a positively charged nylon membrane (Schleicher & Schuell, Dassel, Germany). The RNA was crosslinked to the membrane using an UV crosslinker (Peqlab, Erlangen, Germany). Digoxigeninlabeled m41 RNA probes were synthesized with a DIG RNA Labeling kit (Roche) using linearized pcDNA–m4124 as template for in vitro transcription with T7 or SP6 polymerase (Fermentas, St. Leon-Rot, Germany). Hybridization, washing, and detection of the DIG-labeled probe were performed according to the manufacturer's protocol (Roche). Rapid amplification of 50 and 30 cDNA ends was performed with a SMART RACE kit (Clontech, Mountain View, CA, USA). Amplification products were cloned in pGEM-T Easy (Promega, Mannheim, Germany) and sequenced. For reverse transcriptase (RT)-PCR analysis of gene expression, total cellular RNA was extracted and DNase-treated as described.<sup>41</sup> C-myc transcripts were PCR amplified with published primers,<sup>41</sup> and m41.1 transcripts with primers binding to the first and last 20 nt of the m41.1 ORF.

*Cell viability and apoptosis assays.* STS and ActD were purchased from Sigma (Munich, Germany) and dissolved in DMSO at 500 mM and 100 mg/ml concentrations, respectively. To induce cell death, MCMV-infected 10.1 cells were treated with 200 nM STS or 250 ng/ml ActD. Infected, knockout MEFs were treated with 450 nM STS. Retrovirus-transduced cells were treated 24 h after transduction with 425 nM STS or 2 mg/ml ActD for 48 h. NIH-3T3 cells grown in 48-well dishes were transfected with 0.7 mg plasmid DNA using Superfect (Qiagen). Twenty-five hours after transfection, cells were treated with 0.5 mg/ml anti-Fas antibody (BD Pharmingen, San Diego, CA, USA) and 2.5 mg/ml

cycloheximide for 45 h. Cell viability was measured using an MTT assay as described.<sup>38</sup> Absorbance at 570 nm was determined photometrically. Mean values and standard deviations of at least four parallel experiments are shown. To analyze nuclear DNA fragmentation, 10.1 fibroblasts were grown and infected on coverslips. Transduced 10.1 cells were treated for 25 h with 600 nM STS. Infected RAW264.7 were harvested from culture dishes and sedimented onto glass slides using Cytospin (Thermo Scientific, Waltham, MA, USA). Cells were fixed with 3% paraformaldehyde and stained with a terminal TUNEL assay kit (Roche) containing tetramethylrhodamine-coupled dUTP. Nuclei were counterstained with 40,60-diamidino-2-phenylindole (DAPI). The TUNEL assay using transduced cells was performed in triplicate and more than 100 nuclei were evaluated per sample. For infected fibroblasts and macrophages, an average of 500 and 280 nuclei, respectively, in five or more randomly selected visual fields were counted. To measure cyt c release, 106 cells were mock-infected or infected at an MOI of 3. After 15 h, cells were treated for 5 h with 450 nM STS. Cells were harvested and separated into a mitochondria-enriched heavy membrane fraction and a supernatant fraction as described elsewhere.<sup>30</sup>

**Bak oligomerization assay.** Fibroblasts were infected at an MOI of 5 for 15 h and treated with 450 nM STS for 5 h. Bak oligomerization was determined as described elsewhere.<sup>42</sup> Briefly, the mitochondria-enriched fraction was resuspended in HIM buffer (200mM mannitol, 70mM sucrose, 10mM HEPESKOH, 1mM EGTA, pH 7.5) and freshly prepared 10, 60-bismaleimido-hexane (from TCI Europe, Zwijndrecht, Belgium) in DMSO was added to obtain a 2-mM final concentration. BMH-mediated crosslinking was allowed to proceed for 30 min at room temperature. Bak oligomerization in transduced cells was induced by 15 h incubation with 450 nM STS.

**Immunoprecipitation and immunoblotting.** For immunoprecipitation experiments, 1.5\_106 HEK 293 cells were co-transfected by calcium phosphate precipitation with pcDNAFlag-Bak and pcDNAm41.1HA, pcDNAm41HA, or a control plasmid. After 25 h cells were lysed in a buffer containing 150mM NaCl, 50mM Tris-HCl (pH 7.5), 0.5% Triton-X-100, and a protease inhibitor cocktail (Roche). Flag- or HA-tagged proteins were precipitated at 4 °C overnight with anti-Flag M2 (Sigma) or an anti-HA (Sigma) antibodies bound to protein-G or protein-A sepharose, respectively. Precipitates were washed six times with washing buffer (100mM NaCl, 20mM Tris (pH 7.5), 100mM EDTA, 0.05% Tween-20) and eluted with boiling sample buffer. Cell lysates were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis and transferred to nitrocellulose or PVDF membranes by semi-dry blotting following standard protocols. Anti-HA 16B12 (Covance), anti-b-actin Ac74 (Sigma), anti-a-tubulin (Santa Cruz Biotechnology, Santa Cruz, CA, USA), anti-Bak (Sigma), anti-Bax (Santa Cruz), anti-caspase-3 8G10 (Cell Signaling), and anti-cyt c (Santa Cruz) antibodies were purchased from suppliers as indicated. Antibodies against MCMV IE1 and E1 were provided by Stipan Jonjic (University of Rijeka, Croatia) and antibodies against M44 and glycoprotein B were a gift from Lambert Loh (University of Saskatchewan, Canada). Analysis of immediate-early, early, and late gene expression was performed as described previously.<sup>20</sup> A sequence encoding amino acids 1–116 of m41 was cloned in pMAL-C2 (New England Biolabs, Frankfurt, Germany) that expresses maltose-binding protein (MBP). The MBP fusion protein was expressed in Escherichia coli and purified using an amylose resin column (New England Biolabs). Mice were immunized with the fusion protein and monoclonal antibodies were produced according to standard procedures. Note that the m41-specific 2A6 antibody does not recognize m41.1, since the amino-acid sequences of the two proteins are unrelated (Figure 1a).

**Immunofluorescence.** Cells were seeded on coverslips at least 24 h prior to infection or transfection. Mitochondria were stained with 100 nM MitoTracker Red CMXRos (Invitrogen) for 20 min at 37 °C. Cells were washed with PBS and fixed with 3% paraformaldehyde for 20 min at 4 °C, neutralized with 50mM ammonium chloride, permeabilized with 0.3% Triton X-100, and blocked with 0.2% gelatin. Proteins of interest were detected with primary antibodies against HA (3F10; Roche), Flag (M2; Sigma), Hsp60 (clone 24; BD Pharmingen), PDI (1D3; Stressgen, Ann Arbor, MI, USA), or the Bak N-terminus (TC-102; Calbiochem, Nottingham, UK), and secondary antibodies coupled to AlexaFluor568 or AlexaFluor488 (Invitrogen), respectively. Confocal laser-scanning microscopy was performed using a Zeiss LSM510 META microscope.

Database accession numbers. Sequences of m41.1, r41.1, and e41.1 have been deposited at GenBank under the accession numbers FJ477245, FJ477244, and FJ477243, respectively.



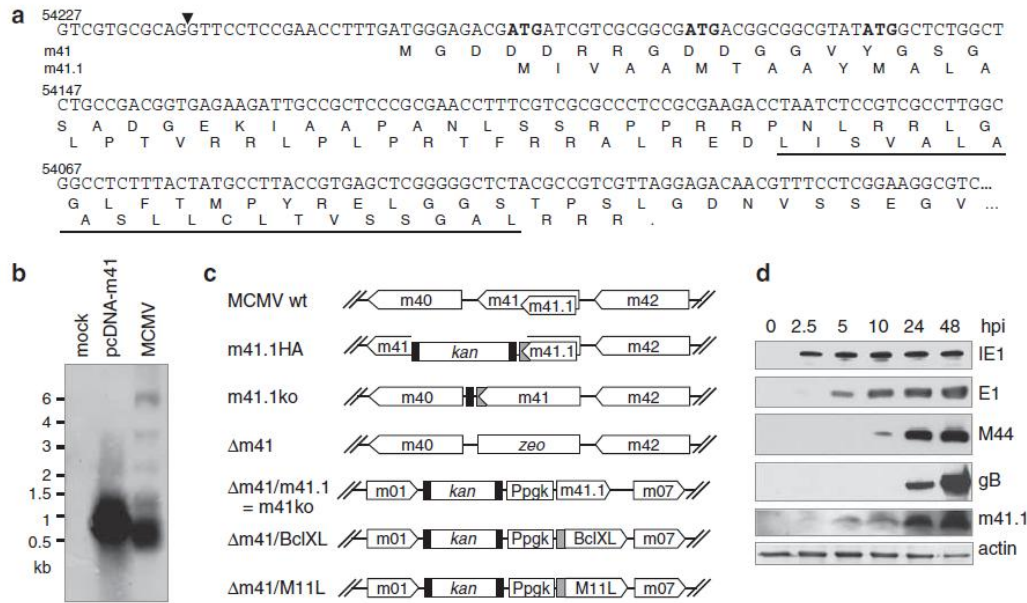
## Acknowledgements

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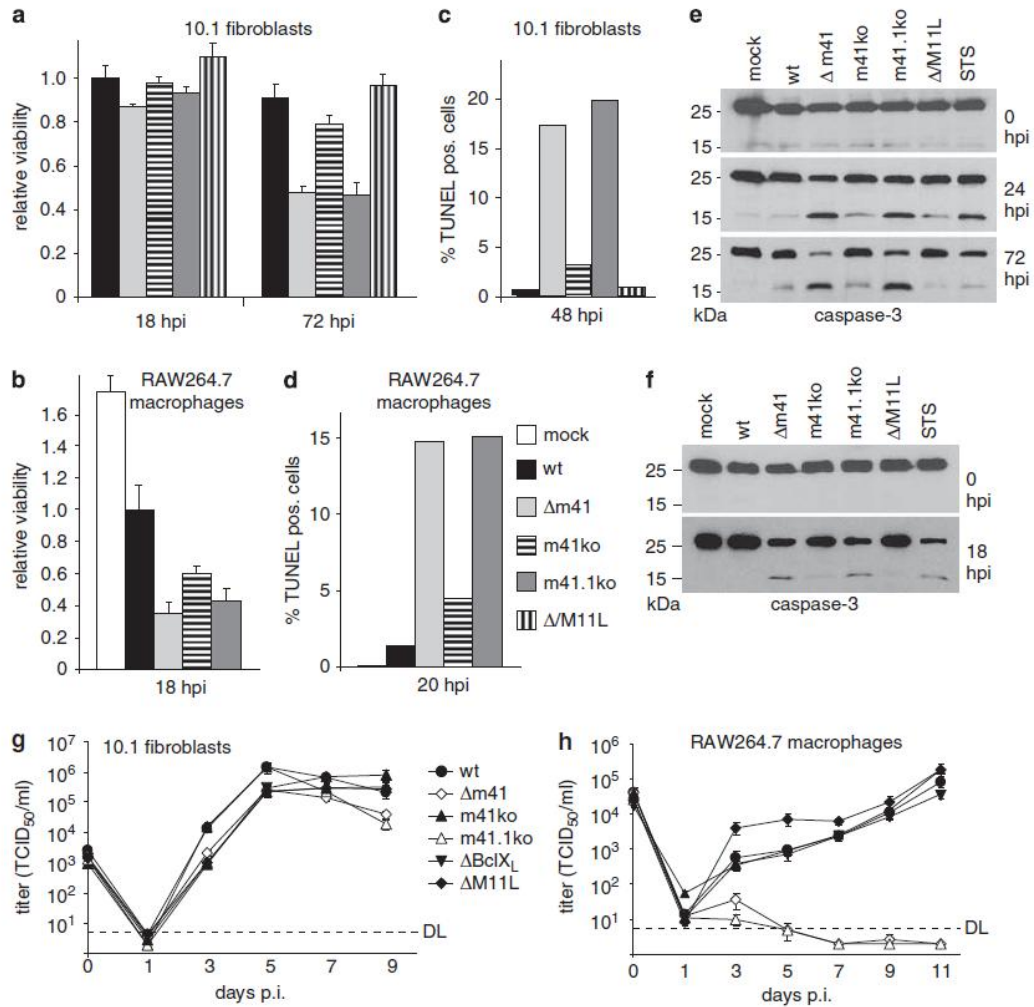
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## Figures

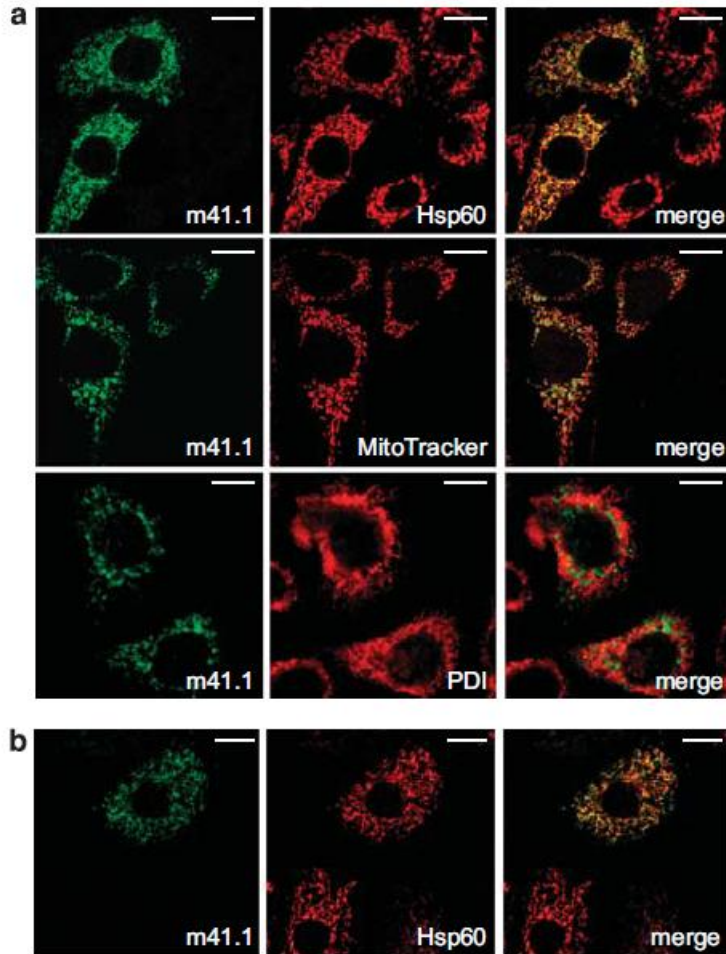


**Figure 1** Transcriptional analysis and mutagenesis of the m41 locus. (a) The major transcript starts at nt position 54 227 and is cleaved and polyadenylated after nt position 53 680. Two protein products encoded in different reading frames are translated from this transcript: the 138-amino-acid m41 (upper translation) and the 57-aminoacid m41.1 (lower translation) proteins. Three possible m41.1 start codons are shown in bold and the predicted transmembrane domain is underlined. Note that only the first 70 amino acids of m41 are shown. (b) Northern blot analysis of m41 transcripts. In addition to the major transcript, a few larger, low-abundance transcripts are detectable, which are spliced transcripts initiating upstream of m41. They use the splice acceptor site indicated by an arrowhead in panel a. Transcripts of similar size are detected in cells transfected with pcDNA-m41. (c) Schematic view of the m41 locus within the MCMV genome. The m41.1 ORF was tagged within the viral genome with an HA epitope tag sequence (grey box), resulting in disruption of the m41 ORF. In the previously described Dm41 mutant, the m41 ORF was replaced with a zeocin-resistance marker.<sup>24</sup> The m41.1ko mutant was obtained by re-inserting an HA-tagged version of m41, in which the m41.1 ATGs had been mutated to ACG. Based on Dm41, three additional mutants were constructed by inserting ORFs m41.1, HA-Bcl-xL, or FlagM11L driven by a phosphoglycerate kinase promoter (Ppgk) into the nonessential m02-m06 region of the MCMV genome. (d) The HA-tagged m41.1 protein was detected in cells infected with MCMV-m41.1HA by Western blot starting 5 hpi. The immediate-early-1 protein (IE1), the early-1 protein (E1), the M44 protein, and glycoprotein-B (gB) are shown as representatives of the immediate-early, early, early-late, and late kinetic classes of MCMV proteins

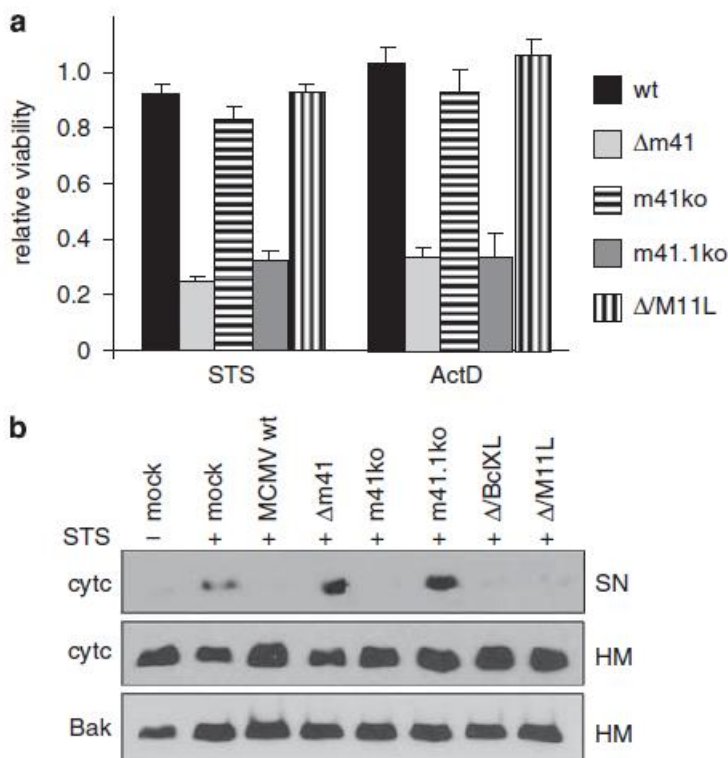


**Figure 2** Phenotypic characterization of the mutant viruses. (a) 10.1 fibroblasts were infected at an MOI of 5 with wt or mutant MCMVs. Cell viability was measured using an MTT assay and values are shown relative to the viability of wt MCMV-infected cells. (b) RAW264.7 macrophages were infected at an MOI of 10 and cell viability was measured as above. (c, d) Nuclear DNA fragmentation was measured in infected fibroblasts and macrophages by TUNEL assay. (e, f) Caspase-3 activation was determined in the same cells by Western blotting using an anti-caspase-3 antibody. The upper band represents the inactive procaspase-3 and the lower band represents the activated (cleaved) form. STS-treated cells are used as positive control. Note that the effect of STS has waned at 72 hpi. (g, h) To determine the replication capacities of the mutant viruses, 10.1 fibroblasts and RAW264.7 macrophages were infected at an MOI of 0.05 and 0.5 respectively. Viral replication was determined by titration on different days post-infection. DL, detection limit

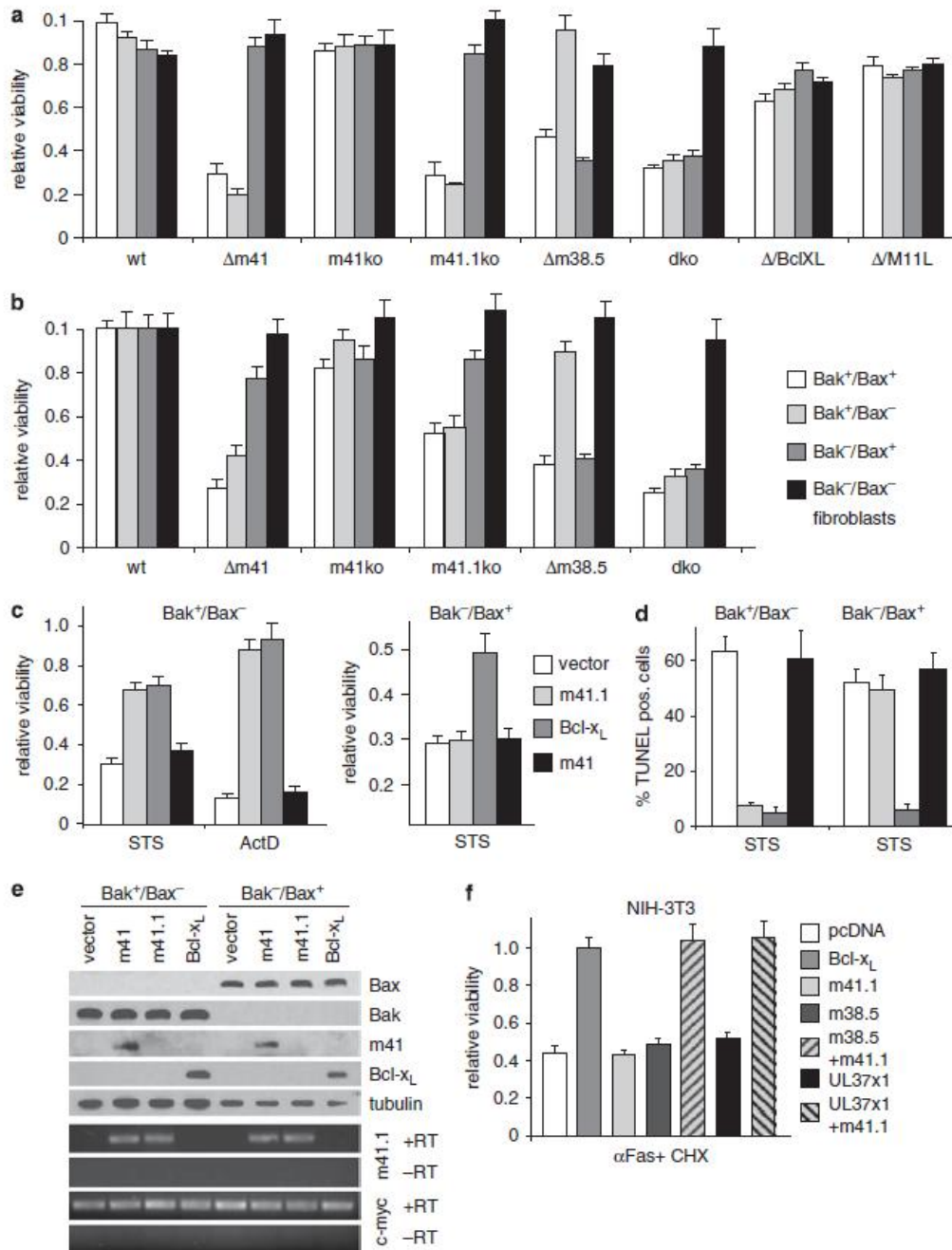
**Figure 3** Fluorescence images showing mitochondrial localization of m41.1. (a) NIH-3T3 cells were transfected with a plasmid expressing HA-tagged m41.1. The protein was detected by immunofluorescence using an anti-HA antibody. The mitochondrial Hsp60 protein and the ER marker PDI were detected with specific antibodies. The red fluorescent MitoTracker dye was used to stain mitochondria. (b) Colocalization of m41.1 and Hsp60 was also detected in NIH-3T3 cells 10 hpi with MCMV-m41.1HA. Bar, 10 mm



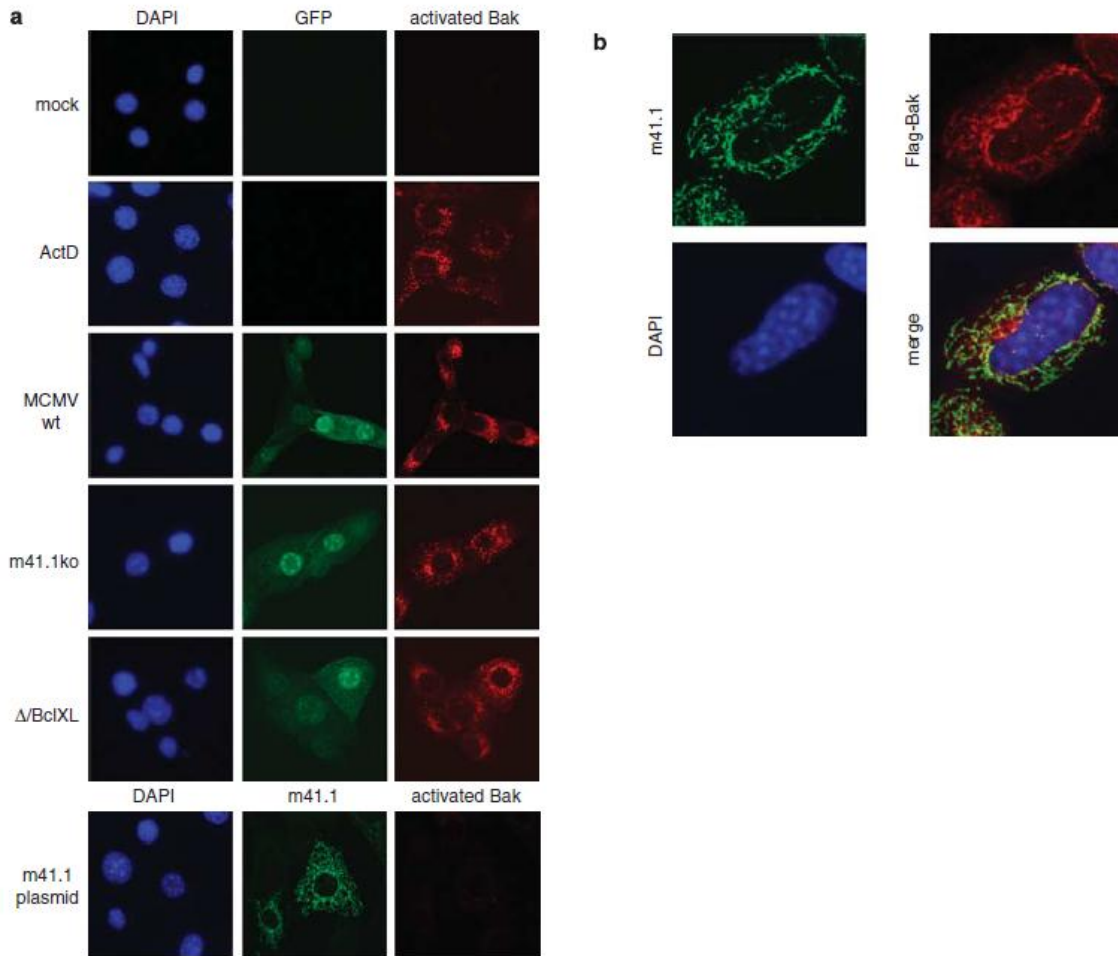
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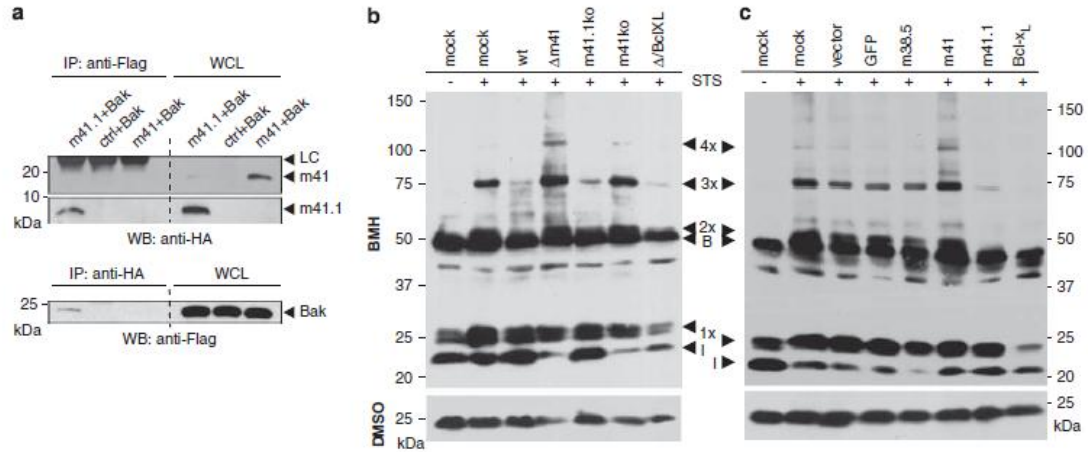
**Figure 4** m41.1 is required to protect MCMV-infected cells from STS- or ActD-induced apoptosis. (a) Infected 10.1 fibroblasts were treated 6 hpi with STS or ActD to induce apoptosis. Cell viability was determined 18 h later by MTT assay and is shown as relative viability compared with infected cells treated with the solvent DMSO. (b) Infected 10.1 fibroblasts were treated with STS and presence of cytochrome c in the heavy membrane (HM) and supernatant (SN) fractions was determined by immunoblotting. Bak was used as a loading control for the HM fraction



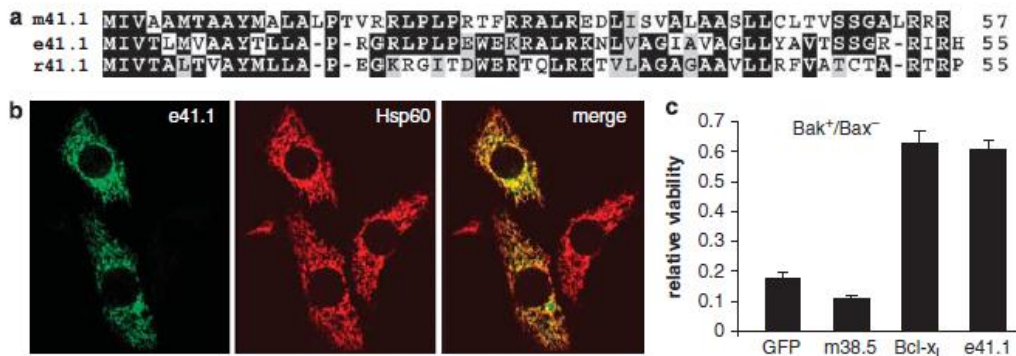
**Figure 5** Bak-specific inhibition of apoptosis by m41.1 and synergy with the Bax-specific inhibitor vMIA. (a) Wt fibroblasts and fibroblasts lacking Bax and/or Bak were infected at an MOI of 5 with wt and mutant MCMVs as indicated. Six hpi cells were treated with STS for 24 h. Cell viability relative to DMSO-treated control cells was determined by MTT assay. (b) Cells were infected as described in panel a, but were not treated with STS. Cell viability at 70 hpi is shown relative to wt MCMV-infected cells. (c) Fibroblasts expressing only Bak or only Bax were transduced with retroviral vectors encoding m41, m41.1, or Bcl-xL. Apoptosis was induced by treatment with STS or ActD. Cell viability was determined 48 h later and is shown relative to the DMSO-treated control cells. (d) Cells were transduced and treated with STS as in panel c. Nuclear-DNA fragmentation was measured after 25 h by TUNEL assay. (e) The presence of the relevant proteins in the cells used in panel d was determined by immunoblotting using antibodies specific to Bax, Bak, and m41. HA-tagged Bcl-xL was detected with anti-HA antibody and tubulin served as loading control. Pilot experiments had indicated that the C-terminal HA tag reduces the antiapoptotic activity of m41.1, and, therefore, vectors expressing untagged m41.1 were used in all cell death assays. Expression of the untagged m41.1 and c-myc (as control) was determined by RT-PCR. Note that the m41.1 sequence is also PCR-amplified from m41 transcripts, as m41.1 lies within m41. However, the m41.1 ATGs were mutated to ACG in the m41 expression vector, and, therefore, m41.1 is not translated from the m41 transcript. (f) NIH-3T3 fibroblasts were transfected with plasmids expressing the indicated proteins and treated with an anti-Fas antibody and cycloheximide (CHX) to induce apoptosis. Cell viability was determined by MTT assay and is shown relative to cells transfected with a Bcl-xL expression plasmid



**Figure 6** Bak activation by MCMV infection. (a) NIH-3T3 cells were infected with wt or mutant MCMVs, all of which express GFP. Activated Bak was detected by immunofluorescence using an antibody directed against the Bak N-terminus. Bak is inactive in mock-infected cells and in cells transfected with pcDNA-m41.1HA. Activated Bak is detected after treatment with 10 mg/ml ActD for 10 h. (b) Cells stably expressing Flag-Bak were infected with MCMV-m41.1HA. Cells were fixed 10 hpi, stained with anti-HA and anti-Flag antibodies, and analyzed by confocal microscopy



**Figure 7** Interaction with Bak and inhibition of Bak oligomerization by m41.1. (a) HEK 293 cells were co-transfected with plasmids expressing Flag-tagged Bak and HA-tagged m41.1, m41, or a control (ctrl) plasmid. Whole-cell lysates (WCL) were subjected to immunoprecipitation (IP) using either an anti-HA or an anti-Flag antibody. The co-immunoprecipitated proteins were detected by Western blotting (WB). LC indicates the antibody light chain. (b) Fibroblasts stably expressing Flag-Bak (but not Bax) were infected with wt and mutant MCMVs, and treated with STS to enhance Bak oligomerization. Cell lysates were treated with DMSO or the crosslinking agent BMH for 30 min, separated by gel electrophoresis, and probed with an anti-Flag antibody. Mono-, di-, tri-, and tetrameric complexes are indicated with arrow heads (1\_ to 4\_). I indicates a faster migrating, internally crosslinked Bak isoform. B probably represents Bak crosslinked to a different cellular protein. (c) Fibroblasts expressing Flag-Bak were transduced with retroviruses encoding the indicated proteins and treated with STS. Bak oligomerization was determined as described above



**Figure 8** Evolutionary conservation of m41.1. (a) ClustalW amino-acid sequence alignment of MCMV protein m41.1, rat CMV English isolate protein e41.1, and rat CMV Maastricht isolate protein r41.1. Identical and similar amino acids are shaded in black and gray, respectively. (b) NIH-3T3 cells were transfected with a plasmid expressing HA-tagged e41.1. Colocalization of e41.1 with the mitochondrial protein Hsp60 was determined by confocal microscopy as in Figure 3. (c) Bak<sup>+</sup>/Bax<sup>-</sup> fibroblasts were transduced with retroviral vectors expressing the indicated proteins and treated with ActD for 48 h as in Figure 5c. Bcl-xL and e41.1 protected from ActD-induced cell death, but m38.5 did not