Atlas of Genetics and Cytogenetics in Oncology and Haematology

brought to you by **CORE** provided by I-Revues

INIST-CNRS

OPEN ACCESS JOURNAL

Leukaemia Section

Short Communication

Classical Hodgkin lymphoma

Samir Dalia, Luis Miguel Juarez Salcedo

Department of Pathology Centro di Riferimento Oncologico Aviano (CRO), Istituto Nazionale Tumori, IRCCS, Aviano, Italy; acarbone@cro.it (AC); Department of Diagnostic Pathology and Laboratory Medicine, Fondazione IRCCS Istituto Nazionale dei Tumori, Milano, Italy; annunziata.gloghini@istitutotumori.mi.it (AG)

Published in Atlas Database: April 2016

Online updated version : http://AtlasGeneticsOncology.org/Anomalies/ClassicHodgkinID1569.html Printable original version : http://documents.irevues.inist.fr/bitstream/handle/2042/68128/04-2016-ClassicHodgkinID1569.pdf DOI: 10.4267/2042/68128

This work is licensed under a Creative Commons Attribution-Noncommercial-No Derivative Works 2.0 France Licence. © 2016 Atlas of Genetics and Cytogenetics in Oncology and Haematology

Abstract

Hodgkin lymphoma (HL) was one of the earliest cancers to be cured with multiagent chemotherapy even before its biology was understood.

Over the past 50 years, a relevant progress has been made toward our understanding of HL pathology, cell biology and treatment options. Histologic classification of HL evolved through different systems, starting from the modern histologic classifications by Jackson and Parker in 1944 and Lukes and Collins in 1966, to the 2008 World Health Organization (WHO) classification. Classical HL involves a clonal expansion of neoplastic B lymphocytes, though a small subset of cases may derive from T cells. Cure rates approaching 80% have been achieved in patients undergoing chemoradiotherapy, qualifying cHL as a chemosensitive disease.

Keywords: Hodgkin lymphoma; classical Hodgkin lymphoma; microenvironment; clinics, pathology; genetics; EBV infection

Identity

Other names: Hodgkin lymphoma, classical Hodgkin disease, Hodgkin disease

Clinics and pathology

Disease

Hodgkin lymphoma(HL) has been classified into classical HL (cHL), which accounts for 95% of all

HL cases, and the less common nodular lymphocyte predominant HL (NLPHL), which is considered to be a separate entity (Stein et al., 2008; Poppema et al., 2008).

Classical HL is a distinct neoplastic entity with typical clinical, epidemiological, pathological, genetic, and virological features. It accounts for approximately 10% of all malignant lymphomas.

Phenotype/cell stem origin

Cell origin: Hodgkin and Reed-Sternberg (HRS) cells, the tumour cells of cHL, derive from preapoptotic crippled germinal center (GC) B cells. In fact, molecular features of HRS cells in cHL demonstrate that they are derived from GC B cells that have acquired disadvantageous immunoglobulin variable chain gene mutations and normally would have undergone apoptosis (Kuppers et al., 2012).

As shown in gene expression profiling (GEP) studies, HRS cells have lost the expression of most B-cell genes and acquired expression of genes that are typical for other types of immune cells (Greaves and Gribben 2012; Steidl et al. 2012; Tiacci et al., 2012).

Phenotype: Phenotypically, HRS cells of cHL are consistently positive for CD30, CD15, CD40, and IRF4/MUM1 (Stein et al., 2008).

Expression of molecular markers in cHL include (Younes et al., 2014)

- B-cell markers (CD20 and CD79) usually negative - GC B-cell markers (BCL6 and AID) usually negative

- Plasma cell markers (MUM1/IRF4) usually positive

- Molecules involved in Ag presentation (MHC class II, CD40, CD80, CD86) positive

A surfaceoma study by TMA analysis indicated that gamma-glutamyltranspeptidase 1 is a potential additional marker for differential diagnosis of cHL versus non Hodgkin lymphoma (Hofmann et al., 2015).

Cellular components of the cHL microenvironment express molecules involved in cancer cell growth and survival (such as CD30L or CD40L), and in immune escape (programmed death 1 (PD-1). For example, CD30L+ eosinophils and mast cells, and proliferation-inducing ligand (APRIL)+ neutrophils, are consistently admixed to HRS cells, whereas CD40L-expressing CD4+ T lymphocytes rosette HRS cells. A fraction of infiltrating CD4+ T cells are regulatory T (Treg) cells. Treg cells and PD-1+ T cells also interact with HRS cells (Aldinucci et al., 2010; Liu et al., 2014; Carbone et al., 2015).

Epidemiology

Classical HL is the most common cancer in patients under 20 years (adolescents and younger adults). The first peak of incidence can be observed in patients under 35 years of age, whereas a second incidence peak can be observed in the elderly (Hjalgrim et al, 2008; Stein et al., 2008).

Cytology

Binucleated and multinucleated HRS cells are giant cells wirh bi- or multinucleation and huge nucleoli. These cells and their mononuclear variant, the socalled Hodgkin cells, are pathognomonic for cHL identification.

Pathology

HRS cells reside in an inflammatory cell microenvironment.

Based on the characteristics of the HRS cells (lacunar cells, multinucleated giant cells, pseudosarcomatous cells) and of the reactive infiltrate, four histologic subtypes have been distinguished: lymphocyte-rich cHL (LRCHL), nodular sclerosis (NS) cHL, mixed cellularity (MC) cHL, and lymphocyte depletion (LD) cHL. Most cHL can be classified as NS or MC subtypes. The remaining LRCHL and LD subtypes are uncommon. LRCHL cases display histological and clinical features intermediate between those of cHL and NLPHL (Poppema et al., 2008; Stein et al., 2008; Swerdlow et al., 2016).

In cHL, microenvironmental cell types include Tand B-reactive lymphocytes, eosinophils, granulocytes, histiocytes/macrophages, plasma cells, mast cells. In addition, a great number of fibroblast-like cells and fibrosis are frequently found (Aldinucci et al., 2010).

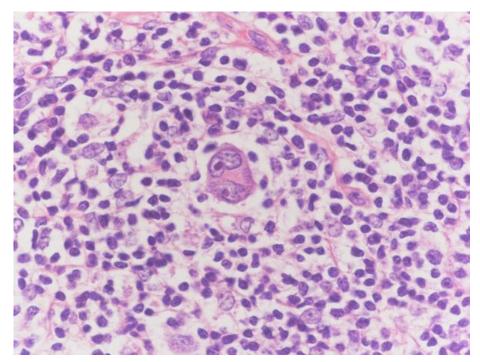


Figure 1. A multinucleated giant cell, the so called Reed-Sternberg cell.

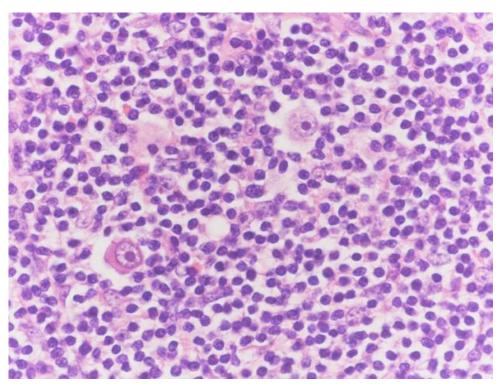


Figure 2. Mononucleated giant cells, the so called Hodgkin cells

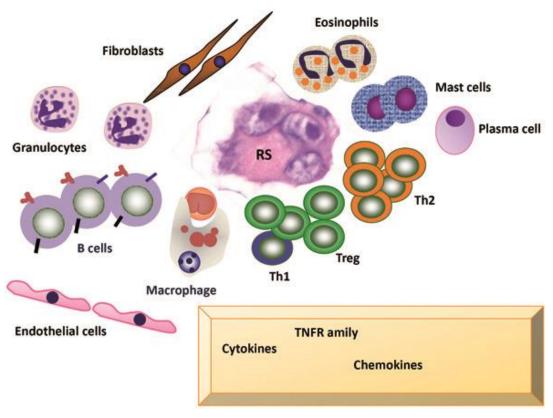


Figure 3. The schema shows a Reed-Sternberg within its cell microenvironment.

Other features

EBV infection

The immunophenotypic and genetic features of HRS cells are identical in the different histologic subtypes of cHL. Conversely, the association with EBV shows differences. EBV is found in HRS cells preferentially in cases of MC and LD cHL, and less frequently in NS and LRCHL. Notably, EBV is found in HRS cells in nearly all cases of cHL occurring in patients infected with HIV (Younes et al., 2014; Dolcetti et al., 2016).

The virologic characteristics of cHL vary according to the immunocompetence status of the host and cHL subtype (IARC, 2012) as follows:

cHL of the general population

- NS cHL, usually EBV negative
- MC cHL, usually EBV positive
- LRCHL, variably EBV positive
- LD cHL, variably EBV positive

Immunodeficiency-associated cHL

- HIV-associated cHL, EBV positive
- Post-transplant cHL, EBV positive
- Iatrogenic (methotrexate), variaby EBV positive

Treatment

Cure rates approaching 80% have been achieved in patients undergoing chemo-radiotherapy, qualifying cHL as a chemosensitive disease (Santoro et al., 1987, Canellos et al., 2014). However, 25% to 30% of these patients show either primary refractoriness to chemotherapy, early disease relapse or late disease relapse (Canellos et al., 2014; Carbone et al., 2015).

Prognosis

The implementation of novel agents for the treatment of multi-relapsed cHL patients has improved the outcome of these patients and will significantly impact the history of multi-relapsed cHL in the near future when the results of combination studies become available. For example, the synergistic effect of Dehydroxymethylepoxyquinomicin (DHMEQ) with three chemotherapeutic drugs widely used in cHL treatment, doxorubicin, gemcitabine and cisplatin, has recently been demonstrated (Locatelli et al., 2014).

Genetics

Recurrent genetic alterations have been identified in HRS cells of cHL. These lesions affecting members of the NF-kappaB or JAK/STAT signalling pathways include inactivating mutation in NFKBIA (10-20% of cases), NFKBIE (10%), TNFAIP3 (40%), SOCS1 (40%), genomic gains of RELA (30%) and JAK2 (30%) and rare BCL3

translocations. TNFAIP3 mutations are found in Epstein-Barr virus-negative cases of cHL. Mutations

have been found in the tumour suppressor genes FAS (CD95) and TP53.

Further genomic imbalances, identified by comparative genomic hybridization studies include gains of IKBKB, CD40 and MAP3K14 that are regulators of NF-kappaB signaling (Küppers and Re, 2007; Hartmann et al., 2008; Steidl et al., 2010; Küppers 2011; Küppers et al., 2012; Pasqualucci and Dalla Favera, 2014).

Interestingly, HRS cells show aberrant somatic hypermutation of several proto-oncogenes (PIM1, RHOH (TTF), MYC, PAX5) in a considerable fraction of cases (Küppers et al., 2012; Pasqualucci and Dalla Favera, 2014).

Cytogenetics

HRS cells are clonal with variable modal chromosome numbers as indicated from direct chromosome analysis and DNA measurements and shown by the detection of clonal immunoglobulin V gene rearrangements in single HRS cells.

The modes are about twice as frequently in the triploid-tetraploid as neardiploid region. Translocations involving the immunoglobulin loci have been found in about 20% of cHL; deletions and duplications, common in other types of tumour, have also been described in cHL.

Diploid as well as aneuploid metaphases are commonly found in chromosome studies, both direct and after culturing. Using FISH 1-12% of "normal" nuclei in cHL exhibit abnormalities, especially trisomies for various chromosomes (Atkin, 1998; Jensen et al., 1998; Hartmann et al., 2008; Schmitz et al., 2009; Steidl et al., 2010; Küppers 2011).

References

A Review of Human Carcinogens. Part B: Biological Agents IARC Monograph on the Evaluation of Carcinogenic Risk to Humans. Vol. 100. IARC, Lyon, France, 2012.

Küppers R. Hodgkin lymphoma Atlas Genet Cytogenet Oncol Haematol. 2011; 15(6): 527-528.

Pasqualucci L, Dalla Favera R.. Molecular Biology of Lymphomas. De Vita, Hellman, and Rosenberg's Cancer: Principles Practice of Oncology. 10th ed. De Vita VTJ, Lawrence TS, Rosemberg SA, (eds). Wolters Kluwer Health/Lippincott Williams & Wilkins, 2014; 1511-1525.

Younes A, Carbone A, Johnson P, Dabaja B, Ansell S, Kuruvilla L.. Hodgkin's lymphoma. De Vita VTJ, Lawrrence TS, Rosemberg SA (eds). De Vita, Hellman, and Rosenberg's Cancer: Principles Practice of Oncology: Wolters Kluwer Health/Lippincott Williams & Wilkins; 2014.

Aldinucci D, Gloghini A, Pinto A, De Filippi R, Carbone A. The classical Hodgkin's lymphoma microenvironment and its role in promoting tumour growth and immune escape J Pathol 2010 Jul;221(3):248-63

Atkin NB. Cytogenetics of Hodgkin's disease Cytogenet Cell Genet 1998;80(1-4):23-7

Canellos GP, Rosenberg SA, Friedberg JW, Lister TA, Devita VT. Treatment of Hodgkin lymphoma: a 50-year perspective J Clin Oncol 2014 Jan 20;32(3):163-8

Carbone A, Gloghini A, Castagna L, Santoro A, Carlo-Stella C. Primary refractory and early-relapsed Hodgkin's lymphoma: strategies for therapeutic targeting based on the tumour microenvironment J Pathol 2015 Sep;237(1):4-13

Dolcetti R, Gloghini A, Caruso A, Carbone A. A lymphomagenic role for HIV beyond immune suppression? Blood 2016 Mar 17;127(11):1403-9 doi: 10

Greaves P, Gribben JG.. Lymphoid neoplasia. Lasercapturing the essence of Hodgkin lymphoma. Blood 2012; 120(23): 4451-4452

Hartmann S, Martin-Subero JI, Gesk S, Hüsken J, Giefing M, Nagel I, Riemke J, Chott A, Klapper W, Parrens M, Merlio JP, Küppers R, Bräuninger A, Siebert R, Hansmann ML. Detection of genomic imbalances in microdissected Hodgkin and Reed-Sternberg cells of classical Hodgkin's lymphoma by array-based comparative genomic hybridization Haematologica 2008 Sep;93(9):1318-26

Hjalgrim H, Engels EA. Infectious aetiology of Hodgkin and non-Hodgkin lymphomas: a review of the epidemiological evidence J Intern Med 2008 Dec;264(6):537-48

Hofmann A, Thiesler T, Gerrits B, Behnke S, Sobotzki N, Omasits U, Bausch-Fluck D, Bock T, Aebersold R, Moch H, Tinguely M, Wollscheid B. Surfaceome of classical Hodgkin and non-Hodgkin lymphoma Proteomics Clin Appl 2015 Aug;9(7-8):661-70

Jansen MP, Hopman AH, Haesevoets AM, Gennotte IA, Bot FJ, Arends JW, Ramaekers FC, Schouten HC. Chromosomal abnormalities in Hodgkin's disease are not restricted to Hodgkin/Reed-Sternberg cells J Pathol 1998 Jun;185(2):145-52

Küppers R, Engert A, Hansmann ML. Hodgkin lymphoma J Clin Invest 2012 Oct;122(10):3439-47

Ku?ppers R, Re D.. Nature of Reed-Sternberg and L H Cells, and their Molecular Biology in Hodgkin Lymphoma. Hodgkin Lymphoma. Hoppe RT, Mauch PM, Armitage JO, et al.,(eds). Lippincott Williams & Wilkins, 2007; 74 - 88

Liu Y, Sattarzadeh A, Diepstra A, Visser L, van den Berg A. The microenvironment in classical Hodgkin lymphoma: an actively shaped and essential tumor component Semin Cancer Biol 2014 Feb;24:15-22

Locatelli SL, Cleris L, Stirparo GG, Tartari S, Saba E, Pierdominici M, Malorni W, Carbone A, Anichini A, Carlo-Stella C. BIM upregulation and ROS-dependent necroptosis mediate the antitumor effects of the HDACi Givinostat and Sorafenib in Hodgkin lymphoma cell line xenografts Leukemia 2014 Sep;28(9):1861-71

Poppema S, Delsol G, Pileri SA, Stein H.. Nodular lymphocyte predominant Hodgkin lymphoma. Swerdlow

SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, Thiele H, Vardiman JW (eds.) World Health Organization Classification of Tumours, Pathology and Genetics of Tumours of Haematopoietic and Lymphoid Tissues, Lyon: IARC Press, 2008: 323-325

Santoro A, Bonadonna G, Valagussa P, Zucali R, Viviani S, Villani F, Pagnoni AM, Bonfante V, Musumeci R, Crippa F, et al. Long-term results of combined chemotherapyradiotherapy approach in Hodgkin's disease: superiority of ABVD plus radiotherapy versus MOPP plus radiotherapy J Clin Oncol 1987 Jan;5(1):27-37

Schmitz R, Hansmann ML, Bohle V, Martin-Subero JI, Hartmann S, Mechtersheimer G, Klapper W, Vater I, Giefing M, Gesk S, Stanelle J, Siebert R, Küppers R. TNFAIP3 (A20) is a tumor suppressor gene in Hodgkin lymphoma and primary mediastinal B cell lymphoma J Exp Med 2009 May 11;206(5):981-9

Steidl C, Connors JM, Gascoyne RD. Molecular pathogenesis of Hodgkin's lymphoma: increasing evidence of the importance of the microenvironment J Clin Oncol 2011 May 10;29(14):1812-26

Steidl C, Diepstra A, Lee T, Chan FC, Farinha P, Tan K, Telenius A, Barclay L, Shah SP, Connors JM, van den Berg A, Gascoyne RD. Gene expression profiling of microdissected Hodgkin Reed-Sternberg cells correlates with treatment outcome in classical Hodgkin lymphoma Blood 2012 Oct 25;120(17):3530-40

Steidl C, Telenius A, Shah SP, Farinha P, Barclay L, Boyle M, Connors JM, Horsman DE, Gascoyne RD. Genome-wide copy number analysis of Hodgkin Reed-Sternberg cells identifies recurrent imbalances with correlations to treatment outcome Blood 2010 Jul 22;116(3):418-27

Stein H, Delsol G, Pileri SA, Weiss LM, Poppema S, Jaffe ES.. Classical Hodgkin lymphoma, introduction. Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, Thiele H, Vardiman JW (eds.) World Health Organization Classification of Tumours, Pathology and Genetics of Tumours of Haematopoietic and Lymphoid Tissues, Lyon: IARC Press, 200

Swerdlow SH, Campo E, Pileri SA, Harris NL, Stein H, Siebert R, Advani R, Ghielmini M, Salles GA, Zelenetz AD, Jaffe ES. The 2016 revision of the World Health Organization classification of lymphoid neoplasms Blood 2016 May 19;127(20):2375-90

Tiacci E, Döring C, Brune V, van Noesel CJ, Klapper W, Mechtersheimer G, Falini B, Küppers R, Hansmann ML. Analyzing primary Hodgkin and Reed-Sternberg cells to capture the molecular and cellular pathogenesis of classical Hodgkin lymphoma Blood 2012 Nov 29;120(23):4609-20

This article should be referenced as such:

Carbone A, Gloghini A. Classical Hodgkin lymphoma. Atlas Genet Cytogenet Oncol Haematol. 2017; 21(1):9-13.